

# Metabolism, Metabolomics, and Inflammation in Posttraumatic Stress Disorder

Synthia H. Mellon, Aarti Gautam, Rasha Hammamieh, Marti Jett, and Owen M. Wolkowitz

## ABSTRACT

Posttraumatic stress disorder (PTSD) is defined by classic psychological manifestations, although among the characteristics are significantly increased rates of serious somatic comorbidities, such as cardiovascular disease, immune dysfunction, and metabolic syndrome. In this review, we assess the evidence for disturbances that may contribute to somatic pathology in inflammation, metabolic syndrome, and circulating metabolites (implicating mitochondrial dysfunction) in individuals with PTSD and in animal models simulating features of PTSD. The clinical and preclinical data highlight probable interrelated features of PTSD pathophysiology, including a proinflammatory milieu, metabolomic changes (implicating mitochondrial and other processes), and metabolic dysregulation. These data suggest that PTSD may be a systemic illness, or that it at least has systemic manifestations, and the behavioral manifestations are those most easily discerned. Whether somatic pathology precedes the development of PTSD (and thus may be a risk factor) or follows the development of PTSD (as a result of either shared pathophysiologies or lifestyle adaptations), comorbid PTSD and somatic illness is a potent combination placing affected individuals at increased physical as well as mental health risk. We conclude with directions for future research and novel treatment approaches based on these abnormalities.

**Keywords:** Animal models, Inflammation, Metabolic syndrome, Metabolomics, Neuroinflammation, PTSD

<https://doi.org/10.1016/j.biopsych.2018.02.007>

Posttraumatic stress disorder (PTSD) is highly prevalent, with an estimate of prevalence among adult Americans to be 6.8% by the National Comorbidity Survey Replication (1). Rates vary greatly by characteristics of the individual and of the trauma, with rates generally higher after exposure to intentional, personally directed trauma as opposed to unintentional, non-personally directed trauma (2). Lifetime PTSD prevalence rates following combat trauma may be especially high, ranging from 10.1% to 30.9% in United States veterans (Vietnam and subsequent conflicts) (3–5). PTSD is precipitated by experiencing or witnessing actual or threatened death, serious injury, or violence, and it has manifestations that include reexperiencing, avoidance, negative thoughts or moods associated with the traumatic event, and hyperarousal (6). In addition to these traditional symptoms, individuals with PTSD, on average, have a substantially higher medical burden, with increased rates of cardiovascular disease, metabolic syndrome (MetS), diabetes, autoimmune diseases, and early mortality, suggesting widespread physical concomitants of PTSD (7–10). In addition to lifestyle-related factors (e.g., decreased physical activity, obesity, tobacco and substance use, medications) (11), certain processes intrinsic to PTSD pathophysiology have been proposed as contributing to somatic disease risk in PTSD, such as accelerated biological aging, sympathetic and glucocorticoid dysregulation, metabolic changes, inflammation, and others (12–17). Thus, PTSD might be considered to be either a systemic condition or one with significant systemic pathologies,

rather than solely a mental illness or a brain disorder (9,12,13,18–21). This article is not intended to be an exhaustive review of the biology of PTSD. Rather, it is a selective overview of certain aspects of PTSD that have been under-studied, namely: 1) inflammation; 2) metabolic dysregulation; and 3) changes in circulating metabolites (especially those implicating mitochondrial dysfunction) that may play a role, not just in the medical disease burden but also in the core psychological symptoms of PTSD. Investigating these processes may reveal interconnected networks or pathways leading to the pathologies in PTSD or following in its wake (19,22–24). An important aspect of this article is the comparison of clinical and animal data for each of the processes we highlight, the latter being useful for mechanistic studies but limited by replicating only selected features of human PTSD. Methodological differences in the clinical studies include the severity and nature of the trauma (e.g., combat vs. civilian trauma, multiple vs. single exposure, interpersonal or intentional trauma vs. witnessed or nonpersonally directed trauma, psychological state, social support and coping abilities of the affected individuals, psychiatric and somatic comorbidities, recency of the trauma relative to the time of testing and source of recruitment of the study sample). Of particular importance in the preclinical studies are the ecological validity of the trauma, single versus multiple (or multimodal) trauma exposures and time of testing relative to the trauma exposure. Discussion of these and other methodological aspects are available for the interested reader

(25,26). We conclude our review with a discussion of possible linkages between metabolomic, metabolic, and inflammatory abnormalities in this illness and with suggestions for novel treatments (Table 1).

## INFLAMMATION

### Clinical Studies

Human studies of PTSD have consistently found pronounced immune alterations, including increased concentrations of inflammatory cytokines and imbalances in immune cell proportions (27–32); these may increase medical morbidity and contribute to core symptoms of PTSD itself (29). In a recent meta-analysis of 20 studies (30), concentrations of interleukin-1 $\beta$  (IL-1 $\beta$ ), IL-6, and interferon gamma (IFN $\gamma$ ) were significantly elevated in PTSD individuals compared with those of control participants. These remained significantly elevated after excluding individuals with comorbid major depressive disorder (MDD), although one study found elevated overnight serum IL-6 levels in individuals with PTSD plus comorbid MDD compared with levels in either control participants or those with PTSD alone (33). When only unmedicated participants were evaluated, these same cytokines plus tumor necrosis factor  $\alpha$  (TNF $\alpha$ ) were found to be significantly elevated in individuals with PTSD (34).

In one of largest studies exclusively in men exposed to combat trauma, those with PTSD showed a significantly elevated composite “proinflammatory score” comprising IL-1 $\beta$ ,

IL-6, TNF $\alpha$ , IFN $\gamma$ , and C-reactive protein (CRP) levels compared with scores in those without PTSD (19). The individual cytokines whose levels significantly differed between groups included TNF $\alpha$  and IFN $\gamma$ , with a trend for IL-6. The proinflammatory score remained significantly higher in individuals with PTSD after controlling for early-life trauma, MDD and its severity, body mass index, ethnicity, education, asthma and/or allergies, time since combat, potentially confounding inflammatory illnesses, and medications. Significant immune activation in PTSD was replicated by the same investigators in a separate group of combat trauma-exposed men (18).

The increase in inflammatory cytokines in PTSD is likely of clinical significance, since chronic inflammation can negatively affect cardiovascular and other aspects of physical health (7) and since individuals with PTSD are significantly more likely to suffer autoimmune disorders compared with individuals with other psychiatric diagnoses (9). Immune mediators, such as IL-1, IL-6, and TNF $\alpha$ , are able to cross the blood-brain barrier (35), and overproduction of proinflammatory cytokines can activate brain microglia (36,37). Nonetheless, the relationship of peripheral markers of inflammation to neuroinflammation is not clear (28,30,38,39). A few small studies have examined cerebrospinal fluid levels of cytokines in PTSD and have yielded conflicting results (40,41).

The underlying causes of immune activation in PTSD are not understood but may represent “sterile inflammation”; in other words, they may be related to diminished glucocorticoid levels (and/or altered glucocorticoid receptor sensitivity) and

**Table 1. Hypothesized Druggable Targets in PTSD**

Target and/or Process	Drug Mechanism of Action	Examples	References
Metabolism; Glucose and Insulin Regulation	Insulin and insulin sensitizers	Insulin (e.g., intranasal); metformin	(116,118,119,123–127)
	PPAR agonists	Thiazolidinediones; PPAR/PGC-1 $\alpha$ activators	(128)
	Adiponectin upregulation	PPAR agonists; angiotensin receptor type I blockers; ACE inhibitors; cannabinoid receptor antagonists; thiazolidinediones; omega-3 fatty acids	(129)
Inflammation	Anti-inflammatories	Cortisol; TNF $\alpha$ antagonists	(42,52,53,130,131)
Mitochondrial Biogenesis and Energetics	PPAR $\gamma$ coactivator 1	Bezafibrate/fenofibrate; rosiglitazone/pioglitazone	(116,118,119,124,125,127,132–134)
	AICAR		(127)
	AMP kinase-activated protein kinase		(127)
	Sirtuins (SIRT 1 activator)	Quercetin; resveratrol; SRT1720	(127)
	Mitochondrial antioxidants	Coenzyme Q10	(133)
	Enhance ATP production	Creatine; lipoic acid; carnitine	(127)
	Trigger the NRF2 antioxidant response element	Oleanolic acid derivatives	(127)

Current pharmacologic treatment of PTSD is inadequate. The disordered processes presented in this article, if verified, would suggest novel therapeutics, such as insulin sensitizers, lipid regulators, mitochondrial biogenesis and/or function enhancers, and anti-inflammatories. Only the biochemical targets reviewed in this article are listed here. Many other potential targets exist. Trials with such agents could provide proof of concepts, especially if they are analyzed in conjunction with indices of target engagement. None of these classes of drugs is yet approved for treating PTSD, and their use would be investigational at this point. Current first-line approaches for most of these targets and/or processes involve aerobic exercise and caloric restriction and, to some extent, selective serotonin reuptake inhibitors. In any event, treatment of PTSD should involve improving physical as well as mental health; therefore, even if the approaches suggested here fail to ameliorate the core psychological symptoms of PTSD, their amelioration of physical disease and disease risk would be salutary.

ACE, angiotensin-converting enzyme; AICAR, 5-amino-imidazole-4-carboxamide ribonucleotide; AMP, adenosine monophosphate; ATP, adenosine triphosphate; NRF2, nuclear factor erythroid 2-related factor 2; PGC, peroxisome proliferator-activated receptor  $\gamma$  coactivator; PPAR, peroxisome proliferator-activated receptor; PTSD, posttraumatic stress disorder; TNF $\alpha$ , tumor necrosis factor  $\alpha$ .

## Metabolism, Metabolomics, and Inflammation in PTSD

increased sympatho-medullary-adrenal activity as well as increased visceral adiposity, although this hypothesis remains to be adequately tested (29,42). It is also possible that the immune activation is related to microbial antigens (the gut microbiome), but further studies are needed (43–45). Longitudinal studies could assess directions of causality and determine whether the proinflammatory state follows the development of PTSD or whether it represents a preexisting risk factor for developing PTSD (29,42). One longitudinal study, the Marine Resiliency Study, showed that preexisting concentrations of CRP were directly correlated with the occurrence and severity of PTSD 3 months after a 7-month military deployment (adjusted for PTSD severity, trauma exposure, etc.) (46). In a study of civilian PTSD from orthopedic injury, elevated levels of IL-6, IL-8, and transforming growth factor  $\beta$  during hospitalization predicted the development of PTSD 1 month later (47). These data raise the possibility that immune dysregulation predisposes individuals to PTSD, although others disagree (20). Inflammation and PTSD could be either reciprocally related or indirectly rather than directly related, and connected via common mechanisms (28,29,42,48). Several studies suggest genetic and epigenetic mechanisms underlie aspects of the proinflammatory milieu in PTSD (49–51).

In summary, immune activation, along with possible imbalances in immune cell types, are among the most replicable biological findings in PTSD. Almost all clinical studies have assessed blood-based markers of inflammation; these may or may not be relevant to brain inflammatory activity. However, peripheral immune activation could contribute to the somatic illnesses seen in PTSD, although definitive routes or even directions of causality have not been proven. Results of studies predicting PTSD treatment response by baseline immune activation or by treatment-associated changes in immune activation have been inconsistent (28,30). Surprisingly few clinical studies have investigated whether primary treatment of immune abnormalities would improve PTSD symptoms (42,52,53). The one notable exception is treatment with hydrocortisone, which showed a therapeutic effect (52,53); however, mechanistic interpretation is difficult. In light of emerging evidence that immune blockade may benefit certain patients with MDD (namely, those with baseline evidence of immune activation) (54), immunosuppressant trials should be a research priority in selected patients with PTSD who show immune activation.

### Preclinical Studies

Multiple animal models simulating features of PTSD have been developed and are discussed in the [Supplement](#) and in recent reviews (55–57). Inflammatory responses in animal models were seen in specific brain regions and throughout the system. A predator-exposure rat study found increased proinflammatory cytokines in the hippocampus, amygdala, and prefrontal cortex, with a concomitant reduction in anti-inflammatory cytokines (58,59). Similarly, a stress-enhanced fear-learning model showed increased hippocampal IL-1 $\beta$  concentrations, and the learning decrement was prevented by blocking central IL-1 $\beta$  signaling after the stress (60). In a predator scent–stress mouse model, activation of the pathway of the nuclear factor  $\kappa$  light-chain enhancer of activated B cells promoted anxiety, and inhibition

of this pathway reduced both IL-1 $\beta$  concentrations and anxiety levels (61).

Molecular investigations found neuroinflammation to relate to behavioral manifestations of simulated PTSD in rodents (62). That study showed proinflammatory mediators (TNF $\alpha$ , CRP, IL-6, IFN $\gamma$ , IL-1 $\beta$ , and cysteine-cysteine chemokine receptor type 2) upregulated in brain and spleen immediately and up to 4 weeks after stress withdrawal. These cytokines inhibited neurogenesis (62). Similarly, upregulation of haptoglobin, myeloperoxidase, and serum amyloid P-component in plasma samples indicated that there was inflammation resulting from aggressor-exposure stress (63). This indication was strengthened by the finding of elevated inflammation in liver and heart (64) immediately after aggressor exposure; this inflammation in the heart paralleled transcriptomic and histopathologic data, indicating cardiac susceptibility (64), which may have relevance for cardiovascular disease associated with human PTSD. Importantly, the kinetics of increased inflammatory responses have not been studied thoroughly, so it is unclear whether this response is sustained, and if so, for how long.

Animal studies have investigated the impact of anti-inflammatory therapies to alter PTSD-like features (65–70). In a rat model of psychogenic stress with elevated cytokine levels, treatment with minocycline, an anti-inflammatory, anti-apoptotic, and neuroprotective tetracycline agent, reduced levels of the cytokines IL-1, IL-6 and TNF $\alpha$  in the hippocampus, frontal cortex, and hypothalamus and reduced anxious behaviors (69). In another model with increased inflammation, ibuprofen not only decreased hippocampal expression of proinflammatory mediators TNF $\alpha$ , IL-1 $\beta$ , and brain-derived neurotrophic factor but also alleviated anxiety symptoms (68). A mouse foot-shock fear-conditioning study used treatment with cyclooxygenase-2 inhibitors, reducing a variety of stress-induced behavioral pathologies (65). Selective serotonin reuptake inhibitors, including fluoxetine, are considered first-line medication treatments for human PTSD. Using the foot-shock fear-conditioning mouse model (67), administration of fluoxetine improved PTSD symptoms while concurrently inhibiting stress-induced inflammatory gene expression (65,71).

Thus, the animal data, like the human data, support an inflammatory component of PTSD both systemically and locally in the brain, both on characterization of inflammatory mediator production and their inhibition.

## METABOLIC SYNDROME

### Clinical Studies

PTSD also is associated with a significantly elevated risk for MetS and for its individual components of obesity, insulin resistance and/or elevated fasting glucose, hypertension, and dyslipidemia (8,12,72). The presence of MetS is highly prognostic of future cardiovascular events and could contribute to the increased morbidity and mortality seen in PTSD (73). Markers of systemic inflammation have been proposed to be included in the definition of MetS, since increased CRP and IL-6 levels are correlated with individual components of MetS and they confer additional health risks beyond those ascribed to MetS alone (74,75). This interrelationship may have causal

elements, because inflammation can lead to obesity and insulin resistance (74), and since increased adiposity can lead to increased production of inflammatory cytokines including TNF $\alpha$  and IL-6. Supporting this interrelationship, Marsland *et al.* (75) studied inflammatory and MetS markers in 645 community volunteers aged 30 to 54 years (48% male, 82% European American, 18% African American), and found, using structural equation modeling, that a higher order common factor of MetS variables (especially adiposity) was significantly and positively correlated with inflammation (elevated CRP and IL-6).

One meta-analysis compared MetS prevalence in PTSD with that in participants from the general population and found an almost doubled risk for MetS with PTSD (relative risk, 1.82; 95% confidence interval, 1.72–1.92) (72); the pooled MetS prevalence for the PTSD group was 38.7%; abdominal obesity, 49.3%; hyperglycemia, 36.1%; hypertriglyceridemia, 45.9%; lowered high-density lipoprotein cholesterol, 46.4%; and hypertension, 76.9%. The prevalence of MetS in PTSD was independent of geographical region or population of participants (combat vs. noncombat PTSD, men vs. women, with vs. without comorbid MDD). A second meta-analysis (76) showed the pooled odds ratio (95% confidence interval) for MetS in PTSD compared with healthy control participants was 1.37 (1.03–1.82).

In a recent study of combat-related trauma in men (8) (with PTSD,  $n = 82$ ; without PTSD,  $n = 82$ ), the prevalence of MetS was significantly higher in individuals with PTSD (18.8% vs. 1.3%,  $p < .0005$ ). The participants with PTSD showed significantly elevated homeostatic model assessment–estimated insulin resistance, fasting glucose concentration, and fasting insulin concentration, even adjusting for body mass index. These differences also remained significant after adjusting for tobacco use, comorbid MDD, and antidepressant use.

Thus, PTSD is associated with a significantly increased incidence of MetS and of its components, which may ultimately relate to the higher disease risk and mortality (12). Apart from contributing to somatic illness risk, MetS may also contribute to core psychiatric symptoms of PTSD, since increased fasting glucose levels and insulin resistance may be associated with damaging effects in the central nervous system (77,78).

While the direction of causality between PTSD and MetS, if any, is not known, Wolf *et al.* (79) examined MetS and PTSD in a longitudinal study and found that PTSD severity predicted subsequent increases in MetS after 2.5 years, but MetS did not predict subsequent PTSD. The causes of increased glucose concentration, insulin resistance, obesity, dyslipidemia, and hypertension found in PTSD are unclear, although increased inflammation, hypothalamic-pituitary-adrenal axis and sympathetic nervous system dysregulation, mitochondrial impairment, and metabolically active hormones (e.g., neuropeptide Y, leptin, adiponectin), as well as lifestyle changes (11,78,80), are possible (12,81–84). For example, Blessing *et al.* (8) found that insulin resistance, a hallmark of MetS, was directly correlated with pulse and the inflammatory marker CRP, although not with IL-6 or TNF $\alpha$ , suggesting a relationship for immune and sympathetic regulation. Polygenic risk for obesity (85) as well as early-life adversity (86) may also play a role in the interaction between MetS and PTSD.

In sum, PTSD is characterized by increased rates of MetS and of its components. When MetS is comorbid with PTSD, it may increase somatic illness comorbidity and possibly affect brain function.

### Preclinical Studies

As with human studies, metabolic dysregulation of lipids was observed in several preclinical studies. Rats exposed to repeated python aggression showed delayed (6 weeks) decreases in “good” high-density lipoprotein cholesterol and sharp increases in serum triglycerides (a risk for cardiovascular disease) (87). In the resident-intruder model, mice exhibited features of MetS with weight gain (88), lipid dysregulation, and indicators of insulin resistance (63) plus activation of hormone-sensitive lipases leading to mobilization of lipids from adipose tissue, while carbohydrate and amino acid mobilization were suppressed. This was supported by increased activity in the liver, with the upregulation of lipid metabolism, fatty acid uptake, and lipogenesis. Human PTSD patients showed significantly reduced fatty acids and few differences in other lipid classes (metabolomics analysis) (18,19,23; Mellon *et al.*, Ph.D., unpublished data, December 2017), with increased levels of total cholesterol, triglycerides, and low-density lipoprotein cholesterol and decreased high-density lipoprotein cholesterol levels (8).

Chronic psychosocial stress in mice also induced lipid dysregulation (89) and intrahepatic accumulation of triglycerides and indicators of MetS (90,91). In aggressor-exposed mice, changes associated with metabolic disorders also were observed by profiling transcripts in blood, brain, and spleen (62) and metabolites in plasma (63). The detection of 2-hydroxybutyrate, an indicator of insulin resistance and impaired glucose regulation, was significantly greater in the stressed mice, suggesting potential changes in insulin function (76,92).

Consistent with the human data (79), the animal literature also suggests that PTSD may predispose individuals to MetS (89,93–95), but data suggesting that MetS predisposes individuals to PTSD are scant (96). In sum, both human and animal data suggest a relationship between PTSD and MetS. The mechanisms (and causality, if any) remain unknown.

### METABOLOMIC ANALYSIS IN PTSD

Many psychiatric and somatic diseases disrupt metabolism, resulting in long-lasting metabolic signatures for a particular disease. Metabolomics studies have the advantage of probing a very large number of metabolites, but personal lifestyle differences may add noise. Metabolomics data are most convincing when 1) identified metabolites interrelate in metabolic pathways and 2) results are replicated in separate samples of participants.

### Clinical Studies

The first clinical study applying unbiased metabolomics using serum from PTSD cases showed glycerophospholipids and endocannabinoid signaling as potential pathologic pathways in PTSD (97). The Department of Defense-funded Systems Biology of PTSD study is the only other study utilizing an unbiased metabolomics analysis in PTSD (18,19,23; Mellon *et al.*, Ph.D., unpublished data, December 2017), and it included 164 combat trauma-exposed men: 82 PTSD cases and 82 control

## Metabolism, Metabolomics, and Inflammation in PTSD

participants. Data for these men were discussed previously regarding inflammatory markers (18,19) and MetS (8). Most of the metabolomics differences were found in energy-related pathways and in dysregulation of carbohydrate, lipid, and amino acid production and utilization. PTSD cases had increased plasma glucose levels and alterations in downstream glucose metabolites, shuttling toward much less efficient nonaerobic metabolism (extramitochondrial) versus aerobic (mitochondrial tricarboxylic acid [TCA] cycle-related) glycolytic pathways, which results in lactic acid buildup and inefficient energy production. Importantly, three separate intermediates of the TCA pathway (lactate, pyruvate, and citrate) showed coordinated changes in the PTSD participants, suggesting decreased entry of pyruvate into the mitochondrial TCA cycle. Also, PTSD cases had reduced plasma concentrations of many essential and nonessential fatty acids, including linoleate, linolenate, eicosapentaenoate, docosapentaenoate, and docosahexaenoate, as well as saturated and unsaturated fatty acids. The reduced abundance of some of these omega-3 fatty acids (docosapentaenoate, docosahexaenoate, and eicosapentaenoate) may contribute to insulin resistance and increased cytokine production as well as to cardiovascular disease (98) and reduced neuroprotective capacity (99). While the overall metabolic profile in PTSD pointed to inflammation, reduced energy utilization, and possibly mitochondrial dysfunction, the differences observed in levels of essential fatty acids may result from altered nutrient absorption or diet, issues with the gut microbiome, or differences in hepatic handling and metabolism of fatty acids.

Apart from metabolomic evidence, preclinical (reviewed below) and clinical studies have also suggested mitochondrial dysfunction in PTSD. Studies of human blood and postmortem brain samples (15,16) showed large numbers of dysregulated genes associated with mitochondrial function, many of which correlated significantly with the severity of PTSD symptoms. Among these, two common dysregulated pathways were found in PTSD: fatty acid metabolism ( $p = .0027$ ) and peroxisome proliferator-activated receptors (PPARs) ( $p = .006$ ). Additionally, mitochondrial DNA copy number, which is related to mitochondrial biogenesis, was positively correlated with positive affect ratings and was low in participants with combat-related PTSD (23). Mitochondrial DNA copy number is normally tightly regulated (100); hence, differences in the mitochondrial DNA copy number in PTSD participants may reflect dysregulation of this cellular process.

In sum, preliminary data from humans suggest the involvement of mitochondrial dysfunction in PTSD pathophysiology. The causes of possible mitochondrial dysfunction in PTSD are unknown but could be related to genetic or epigenetic factors, oxidative stress, cell damage responses, or premature cell aging, among others (101,102). The potential sequelae of mitochondrial dysfunction are extensive and include diminished fuel utilization and energy synthesis (e.g., anaerobic compared with aerobic respiration), altered glucose and lipid utilization, increased oxidative stress, cellular senescence and/or apoptosis (101–104), and others. It is possible that mitochondrial function may be a key target to prevent or reverse certain neurobehavioral and physiological aspects of PTSD (15,101,105,106) or may be directly related to immune activation and MetS in PTSD, as discussed below.

**Preclinical Studies**

A brain microdialysis metabolomics study in live mice enabled *in vivo* investigation of metabolites (at baseline) as predictors of subsequent sensitivity or resilience to PTSD-like behaviors in a foot-shock model (107). At day 2 after a stress event, behavioral symptoms (hyperarousal) in shocked mice were predicted by the enrichment of the TCA cycle and glyoxylate and dicarboxylate metabolism in the medial prefrontal cortex prior to foot shock. Another study also showed increased TCA cycle activity in synaptosomes of high-anxiety mice (108), suggesting that there may be inherent differences in functional synapses, which are enriched with mitochondria in control mice and high-anxiety mice.

The same investigators used proteomic and metabolomic analysis of specific brain regions in the foot-shock model to assess stress-induced dysregulated metabolic pathways after foot shock and to assess metabolic pathways that are responsive to treatment (fluoxetine). The stress led to decreases in TCA cycle pathway enzyme abundance and metabolites in the nucleus accumbens and the anterior cingulate cortex (66). Interestingly, fluoxetine treatment (12 hours after foot shock) prevented alterations of the TCA cycle in the nucleus accumbens and anterior cingulate cortex and decreased conditioned fear responses (66).

In a rat model of single, prolonged stress (109), ultrastructural examination of hippocampal neurons showed differential cellular organelle damage. Increased cytochrome oxidase release (mitochondrial) and enlarged and/or swollen mitochondrial structures with vacuolar and crest degeneration suggested mitochondrial damage.

Mice exposed to prolonged inescapable tail-shock stress showed induction of hippocampal apoptosis, suggesting involvement of mitochondrial pathways (16,110–113). Studies also found that 34 mitochondrial-focused genes were upregulated in the amygdala of stressed rats (16). As with humans, fatty acid metabolism and PPARs were among the 10 pathways found to be dysregulated. Finally, mitochondrial dysfunction, and the effects of risperidone and paroxetine on mitochondrial function, were studied in a rat model (114). Both drugs ameliorated stress-induced behavioral symptoms; risperidone ameliorated stress-induced increases in brain mitochondrial enzyme activities, and both drugs reduced stress-induced brain apoptosis, suggesting that both apoptosis and mitochondrial dysfunction may contribute to PTSD-like behavioral symptoms.

In summary, metabolomic changes in PTSD are understudied, especially in large clinical samples. Nonetheless, clinical and preclinical studies to date suggest that PTSD involves some aspects of mitochondrial dysfunction, and these may be related to the other abnormalities.

**ARE INFLAMMATION, MetS, AND MITOCHONDRIAL DYSFUNCTION INTERRELATED IN PTSD?**

In this review, we have highlighted inflammation, MetS, and metabolomic changes, especially those involving mitochondrial function, as potential individual pathologies in PTSD. However, these and other pathological features may be interrelated, although there is insufficient evidence to posit causal relationships. For example, mitochondrial dysfunction can lead

to reduced fatty acid metabolism (beta oxidation) and increased lipid accumulation in muscle and liver tissue, resulting in increased diacylglycerol, ceramide, and acylcarnitine accumulation and increased reactive oxygen species levels, all of which can lead to insulin resistance and further mitochondrial damage. Mitochondrial dysfunction can also result in increased inflammation via reactive oxygen species; reactive oxygen species trigger inflammasome (nucleotide-binding domain and leucine-rich repeat containing protein 3) activation, resulting in increased cytokine (e.g., IL1 $\beta$  and IL-18) levels. Nucleotide-binding domain and leucine-rich repeat containing protein 3 activation reciprocally regulates glucose and lipid metabolism. Central adiposity (and macrophages accumulating in adipose tissue) generates inflammatory cytokines (e.g., TNF) and inflammatory adipokines, leading to an inflammatory state seen with obesity and insulin resistance [see (22,115–120)].

Apart from these interrelationships, a “mitochondrial allostatic load” model has also been proposed to connect these perturbations (104). In this model, metabolic dysregulation and chronically elevated glucose concentration, as seen in PTSD, damage mitochondria and mitochondrial DNA, generating byproducts that promote systemic inflammation, alter gene expression and accelerate cellular aging. Lastly, mitochondrial dysfunction can affect cellular responses, suggesting “metabolic checkpoints” (121) or “cell danger responses” (101) that connect metabolism with mitochondrial function (121). In all of these models, mitochondrial dysfunction may be a “central link” between inflammation, oxidative stress, and metabolism, as suggested by Kusminski and Scherer (118).

A “systems biology” approach, assessing multiple features and levels of analysis in the same individuals, holds the greatest promise for delineating interlinked pathology, hopefully delineating biologically informed phenotypes, subgrouping, and diagnoses (122).

## CONCLUSIONS

An accumulation of evidence suggests that PTSD has significant somatic manifestations and may, in fact, have aspects of a systemic illness or of an illness with significant systemic comorbidities. Any model of PTSD should, therefore, account not only for psychological symptoms but also for the physical morbidity and premature mortality seen in this illness. We have reviewed several, but far from all, systemic pathologies that may accompany PTSD, although it is not known how well these peripheral pathologies are reflected in the brain. Whether somatic pathology precedes the development of PTSD (and thus, may be a risk factor) or follows the development of PTSD (as a result of either shared pathophysiology or of lifestyle factors), comorbid PTSD and somatic illness place affected individuals at increased health risk. Identification of novel mechanism-based treatment targets in PTSD holds promise for relieving both the psychological and somatic symptoms in this prevalent disorder.

To the extent the processes reviewed here participate in the pathophysiology of PTSD, new therapeutic opportunities, based on specific pathological targets, may exist. Certain such possibilities are listed in Table 1, but very few of these interventions have yet been tested. Even if the psychological

symptoms of PTSD do not respond to these novel types of interventions, somatic health might improve and novel biochemical targets might be clarified.

## ACKNOWLEDGMENTS AND DISCLOSURES

This work was supported by the United States Army Medical Research and Materiel Command (Grant No. 09284002 to MJ) and the United States Army Medical Research and Materiel Command Military Operational Medicine Research Program/Defense Health Agency/Congressional Special Interests (Grant No. 190040 to MJ).

This work was funded by the Department of Defense (Grant Nos. W81XWH-11-2-0223 and W81XWH-10-1-0021 to SHM and OMW and Grant Nos. VB4, T869, and MM3 to MJ).

We acknowledge the United States Army Research Office for their support. We also acknowledge Dr. Charles Marmar and his research group at New York University Medical Center and Dr. Rachel Yehuda and her research group at Mount Sinai School of Medicine and the James J. Peters Veterans Affairs Medical Center (Bronx, NY) for their tremendous efforts in sample collections and clinical assessments as well as all of the collaborators in the Systems Biology of PTSD research study. We thank Dr. Julia Scheerer for her editorial comments.

All authors report no biomedical financial interests or potential conflicts of interest.

The views, opinions, and findings contained in this report are those of the authors and should not be construed as official Department of the Army position, policy, or decision, unless so designated by other official documentation.

Citations of commercial organizations or trade names in this report do not constitute an official Department of the Army endorsement or approval of the products or services of these organizations.

This research complied with the Animal Welfare Act, implementing Animal Welfare Regulations and the Public Health Service Policy on Humane Care and Use of Laboratory Animals, and it adhered to the principles noted in the Guide for the Care and Use of Laboratory Animals (National Research Council, 2011).

## ARTICLE INFORMATION

From the Department of Obstetrics, Gynecology and Reproductive Sciences (SHM) and Department of Psychiatry (OMW), University of California—San Francisco, San Francisco, California, and Integrative Systems Biology (AG, RH, MJ), United States Army Medical Research and Materiel Command, United States Army Center for Environmental Health Research, Fort Detrick, Frederick, Maryland.

Address correspondence to Marti Jett, Ph.D., Chief Scientist, United States Army Center for Environmental Health Research, 568 Doughten Drive, Fort Detrick, MD 21702; E-mail: [marti.jett-tilton.civ@mail.mil](mailto:marti.jett-tilton.civ@mail.mil).

Received Sep 6, 2017; revised Feb 8, 2018; accepted Feb 14, 2018.

Supplementary material cited in this article is available online at <https://doi.org/10.1016/j.biopsych.2018.02.007>.

## REFERENCES

1. Kessler RC, Berglund P, Demler O, Jin R, Merikangas KR, Walters EE (2005): Lifetime prevalence and age-of-onset distributions of DSM-IV disorders in the National Comorbidity Survey Replication. *Arch Gen Psychiatry* 62:593–602.
2. Santiago PN, Ursano RJ, Gray CL, Pynoos RS, Spiegel D, Lewis-Fernandez R, *et al.* (2013): A systematic review of PTSD prevalence and trajectories in DSM-5 defined trauma exposed populations: intentional and non-intentional traumatic events. *PLoS One* 8: e59236.
3. Kang HK, Natelson BH, Mahan CM, Lee KY, Murphy FM (2003): Post-traumatic stress disorder and chronic fatigue syndrome-like illness among Gulf War veterans: A population-based survey of 30,000 veterans. *Am J Epidemiol* 157:141–148.
4. Kulka RA, Schlenger WA, Fairbanks JA, Hough RL, Jordan BK, Marmar CR (1990): Trauma and the Vietnam War Generation: Report

## Metabolism, Metabolomics, and Inflammation in PTSD

- of Findings from the National Vietnam Veterans Readjustment Study. New York, NY: Brunner/Mazel.
5. Tanielian T, Jaycox LE (2008): Invisible Wounds of War: Psychological and Cognitive Injuries, Their Consequences, and Services to Assist Recovery. Santa Monica, CA: RAND Corporation.
  6. American Psychiatric Association (2013): Diagnostic and Statistical Manual of Mental Disorders, 5th ed. Arlington, VA: American Psychiatric Publishing.
  7. Levine AB, Levine LM, Levine TB (2014): Posttraumatic stress disorder and cardiometabolic disease. *Cardiology* 127:1–19.
  8. Blessing E, Reus VI, Mellon SH, Wolkowitz OM, Flory JD, Bierer LM, *et al.* (2018): Biological predictors of insulin resistance associated with posttraumatic stress disorder in young military veterans. *Psychoneuroendocrinology* 82:91–97.
  9. O'Donovan A, Cohen BE, Seal KH, Bertenthal D, Margaretten M, Nishimi K, *et al.* (2015): Elevated risk for autoimmune disorders in Iraq and Afghanistan veterans with posttraumatic stress disorder. *Biol Psychiatry* 77:365–374.
  10. Pacella ML, Hruska B, Delahanty DL (2013): The physical health consequences of PTSD and PTSD symptoms: A meta-analytic review. *J Anxiety Disord* 27:33–46.
  11. Zen AL, Whooley MA, Zhao S, Cohen BE (2012): Post-traumatic stress disorder is associated with poor health behaviors: Findings from the Heart and Soul study. *Health Psychol* 31:194–201.
  12. Michopoulos V, Vester A, Neigh G (2016): Posttraumatic stress disorder: A metabolic disorder in disguise? *Exp Neurol* 284:220–229.
  13. Lohr JB, Palmer BW, Eidt CA, Aailaboyina S, Mausbach BT, Wolkowitz OM, *et al.* (2015): Is post-traumatic stress disorder associated with premature senescence? A review of the literature. *Am J Geriatr Psychiatry* 23:709–725.
  14. Michopoulos V, Norrholm SD, Jovanovic T (2015): Diagnostic biomarkers for posttraumatic stress disorder: promising horizons from translational neuroscience research. *Biol Psychiatry* 78:344–353.
  15. Su YA, Wu J, Zhang L, Zhang Q, Su DM, He P, *et al.* (2008): Dysregulated mitochondrial genes and networks with drug targets in postmortem brain of patients with posttraumatic stress disorder (PTSD) revealed by human mitochondria-focused cDNA microarrays. *Int J Biol Sci* 4:223–235.
  16. Zhang L, Li H, Hu X, Benedek DM, Fullerton CS, Forsten RD, *et al.* (2015): Mitochondria-focused gene expression profile reveals common pathways and CPT1B dysregulation in both rodent stress model and human subjects with PTSD. *Transl Psychiatry* 5:e580.
  17. Lindqvist D, Epel ES, Mellon SH, Penninx BW, Revesz D, Verhoeven JE, *et al.* (2015): Psychiatric disorders and leukocyte telomere length: Underlying mechanisms linking mental illness with cellular aging. *Neurosci Biobehav Rev* 55:333–364.
  18. Lindqvist D, Mellon SH, Dhabhar FS, Yehuda R, Marlene Grenon S, Flory JD, *et al.* (2017): Increased pro-inflammatory milieu in combat related PTSD—a new cohort replication study. *Brain Behav Immun* 59:260–264.
  19. Lindqvist D, Wolkowitz OM, Mellon S, Yehuda R, Flory JD, Henn-Haase C, *et al.* (2014): Proinflammatory milieu in combat-related PTSD is independent of depression and early life stress. *Brain Behav Immun* 42:81–88.
  20. O'Donovan A, Ahmadian AJ, Neylan TC, Paucult MA, Edmondson D, Cohen BE (2017): Current posttraumatic stress disorder and exaggerated threat sensitivity associated with elevated inflammation in the Mind Your Heart Study. *Brain Behav Immun* 60:198–205.
  21. McFarlane AC, Lawrence-Wood E, Van Hooff M, Malhi GS, Yehuda R (2017): The need to take a staging approach to the biological mechanisms of PTSD and its treatment. *Curr Psychiatry Rep* 19:10.
  22. Tschopp J (2011): Mitochondria: Sovereign of inflammation? *Eur J Immunol* 41:1196–1202.
  23. Bersani FS, Morley C, Lindqvist D, Epel ES, Picard M, Yehuda R, *et al.* (2016): Mitochondrial DNA copy number is reduced in male combat veterans with PTSD. *Prog Neuropsychopharmacol Biol Psychiatry* 64:10–17.
  24. Hammamieh R, Chakraborty N, Gautam A, Muhie S, Yang R, Donohue D, *et al.* (2017): Whole-genome DNA methylation status associated with clinical PTSD. *Transl Psychiatry* 7:e1169.
  25. Cohen H, Matar MA, Zohar J (2014): Maintaining the clinical relevance of animal models in translational studies of post-traumatic stress disorder. *ILAR J* 55:233–245.
  26. Friedman MJ, Keane TM, Resick PA (2014): Handbook of PTSD: Science and Practice, 2nd ed. New York, NY: Guilford Press.
  27. Wang Z, Mandel H, Levingston CA, Young MR (2016): An exploratory approach demonstrating immune skewing and a loss of coordination among cytokines in plasma and saliva of veterans with combat-related PTSD. *Hum Immunol* 77:652–657.
  28. Wang Z, Young MR (2016): PTSD, a disorder with an immunological component. *Front Immunol* 7:219.
  29. Pace TW, Heim CM (2011): A short review on the psychoneuroimmunology of posttraumatic stress disorder: from risk factors to medical comorbidities. *Brain Behav Immun* 25:6–13.
  30. Wang Z, Caughron B, Young MRI (2017): Posttraumatic stress disorder: An immunological disorder? *Front Psychiatry* 8:222.
  31. Lindqvist D, Mellon SH, Dhabhar FS, Yehuda R, Grenon SM, Flory JD, *et al.* (2017): Increased circulating blood cell counts in combat-related PTSD: Associations with inflammation and PTSD severity. *Psychiatry Res* 258:330–336.
  32. Bersani FS, Wolkowitz OM, Milush JM, Sinclair E, Eppling L, Aschbacher K, *et al.* (2016): A population of atypical CD56(-)CD16(+) natural killer cells is expanded in PTSD and is associated with symptom severity. *Brain Behav Immun* 56:264–270.
  33. Gill J, Luckenbaugh D, Charney D, Vythilingam M (2010): Sustained elevation of serum interleukin-6 and relative insensitivity to hydrocortisone differentiates posttraumatic stress disorder with and without depression. *Biol Psychiatry* 68:999–1006.
  34. Passos IC, Vasconcelos-Moreno MP, Costa LG, Kunz M, Brietzke E, Quevedo J, *et al.* (2015): Inflammatory markers in post-traumatic stress disorder: A systematic review, meta-analysis, and meta-regression. *Lancet Psychiatry* 2:1002–1012.
  35. Banks WA, Kastin AJ, Broadwell RD (1995): Passage of cytokines across the blood-brain barrier. *Neuroimmunomodulation* 2:241–248.
  36. Liu B, Hong JS (2003): Role of microglia in inflammation-mediated neurodegenerative diseases: Mechanisms and strategies for therapeutic intervention. *J Pharmacol Exp Ther* 304:1–7.
  37. de Pablos RM, Herrera AJ, Espinosa-Oliva AM, Sarmiento M, Munoz MF, Machado A, *et al.* (2014): Chronic stress enhances microglia activation and exacerbates death of nigral dopaminergic neurons under conditions of inflammation. *J Neuroinflammation* 11:34.
  38. Song C, Phillips AG, Leonard BE, Horrobin DF (2004): Ethyl-eicosapentaenoic acid ingestion prevents corticosterone-mediated memory impairment induced by central administration of interleukin-1beta in rats. *Mol Psychiatry* 9:630–638.
  39. Johansen JP, Cain CK, Ostroff LE, LeDoux JE (2011): Molecular mechanisms of fear learning and memory. *Cell* 147:509–524.
  40. Baker DG, Ekhtator NN, Kasckow JW, Hill KK, Zoumakis E, Dashevsky BA, *et al.* (2001): Plasma and cerebrospinal fluid interleukin-6 concentrations in posttraumatic stress disorder. *Neuroimmunomodulation* 9:209–217.
  41. Bonne O, Gill JM, Luckenbaugh DA, Collins C, Owens MJ, Alesci S, *et al.* (2011): Corticotropin-releasing factor, interleukin-6, brain-derived neurotrophic factor, insulin-like growth factor-1, and substance P in the cerebrospinal fluid of civilians with posttraumatic stress disorder before and after treatment with paroxetine. *J Clin Psychiatry* 72:1124–1128.
  42. Olf M, van Zuiden M (2017): Neuroendocrine and neuroimmune markers in PTSD: pre-, peri- and post-trauma glucocorticoid and inflammatory dysregulation. *Curr Opin Psychol* 14:132–137.
  43. Brenner LA, Stearns-Yoder KA, Hoffberg AS, Penzenik ME, Starosta AJ, Hernandez TD, *et al.* (2017): Growing literature but limited evidence: A systematic review regarding prebiotic and probiotic interventions for those with traumatic brain injury and/or posttraumatic stress disorder. *Brain Behav Immun* 65:57–67.

44. Gocan AG, Bachg D, Schindler AE, Rohr UD (2012): Balancing steroid hormone cascade in treatment-resistant veteran soldiers with PTSD using a fermented soy product (FSWW08): A pilot study. *Horm Mol Biol Clin Investig* 10:301–314.
45. Hemmings SMJ, Malan-Muller S, van den Heuvel LL, Demmitt BA, Stanislawski MA, Smith DG, *et al.* (2017): The microbiome in post-traumatic stress disorder and trauma-exposed controls: An exploratory study. *Psychosom Med* 79:936–946.
46. Eraly SA, Nievergelt CM, Maihofer AX, Barkauskas DA, Biswas N, Agorastos A, *et al.* (2014): Assessment of plasma C-reactive protein as a biomarker of posttraumatic stress disorder risk. *JAMA Psychiatry* 71:423–431.
47. Cohen M, Meir T, Klein E, Volpin G, Assaf M, Pollack S (2011): Cytokine levels as potential biomarkers for predicting the development of posttraumatic stress symptoms in casualties of accidents. *Int J Psychiatry Med* 42:117–131.
48. Lopresti AL, Drummond PD (2013): Obesity and psychiatric disorders: Commonalities in dysregulated biological pathways and their implications for treatment. *Prog Neuropsychopharmacol Biol Psychiatry* 45:92–99.
49. Rosen RL, Levy-Carrick N, Reibman J, Xu N, Shao Y, Liu M, *et al.* (2017): Elevated C-reactive protein and posttraumatic stress pathology among survivors of the 9/11 World Trade Center attacks. *J Psychiatr Res* 89:14–21.
50. Michopoulos V, Rothbaum AO, Jovanovic T, Almlil LM, Bradley B, Rothbaum BO, *et al.* (2015): Association of CRP genetic variation and CRP level with elevated PTSD symptoms and physiological responses in a civilian population with high levels of trauma. *Am J Psychiatry* 172:353–362.
51. Miller MW, Maniates H, Wolf EJ, Logue MW, Schichman SA, Stone A, *et al.* (2018): CRP polymorphisms and DNA methylation of the AIM2 gene influence associations between trauma exposure, PTSD, and C-reactive protein. *Brain Behav Immun* 67:194–202.
52. Amos T, Stein DJ, Ipser JC (2014): Pharmacological interventions for preventing post-traumatic stress disorder (PTSD). *Cochrane Database Syst Rev* CD006239.
53. Birur B, Math SB, Fargason RE (2017): A review of psychopharmacological interventions post-disaster to prevent psychiatric sequelae. *Psychopharmacol Bull* 47:8–26.
54. Raison CL, Rutherford RE, Woolwine BJ, Shuo C, Schettler P, Drake DF, *et al.* (2013): A randomized controlled trial of the tumor necrosis factor antagonist infliximab for treatment-resistant depression: The role of baseline inflammatory biomarkers. *JAMA Psychiatry* 70:31–41.
55. Borghans B, Homberg JR (2015): Animal models for posttraumatic stress disorder: An overview of what is used in research. *World J Psychiatry* 5:387–396.
56. Schöner J, Heinz A, Endres M, Gertz K, Kronenberg G (2017): Post-traumatic stress disorder and beyond: an overview of rodent stress models. *J Cell Mol Med* 21:2248–2256.
57. Deslauriers J, Toth M, Der-Avakian A, Risbrough VB (2017): Current status of animal models of posttraumatic stress disorder: Behavioral and biological phenotypes, and future challenges in improving translation [published online ahead of print Nov 20]. *Biol Psychiatry*.
58. Wilson CB, Ebenezer PJ, McLaughlin LD, Francis J (2014): Predator exposure/psychosocial stress animal model of post-traumatic stress disorder modulates neurotransmitters in the rat hippocampus and prefrontal cortex. *PLoS One* 9:e89104.
59. Deslauriers J, Powell S, Risbrough VB (2017): Immune signaling mechanisms of PTSD risk and symptom development: insights from animal models. *Curr Opin Behav Sci* 14:123–132.
60. Jones ME, Lebonville CL, Barrus D, Lysle DT (2015): The role of brain interleukin-1 in stress-enhanced fear learning. *Neuropsychopharmacology* 40:1289–1296.
61. Zimmerman G, Shaltiel G, Barbash S, Cohen J, Gasho CJ, Shenhar-Tsarfaty S, *et al.* (2012): Post-traumatic anxiety associates with failure of the innate immune receptor TLR9 to evade the pro-inflammatory NFkappaB pathway. *Transl Psychiatry* 2:e78.
62. Muhie S, Gautam A, Chakraborty N, Hoke A, Meyerhoff J, Hammamieh R, *et al.* (2017): Molecular indicators of stress-induced neuroinflammation in a mouse model simulating features of post-traumatic stress disorder. *Transl Psychiatry* 7:e1135.
63. Gautam A, D'Arpa P, Donohue DE, Muhie S, Chakraborty N, Luke BT, *et al.* (2015): Acute and chronic plasma metabolomic and liver transcriptomic stress effects in a mouse model with features of post-traumatic stress disorder. *PLoS One* 10:e0117092.
64. Cho JH, Lee I, Hammamieh R, Wang K, Baxter D, Scherler K, *et al.* (2014): Molecular evidence of stress-induced acute heart injury in a mouse model simulating posttraumatic stress disorder. *Proc Natl Acad Sci U S A* 111:3188–3193.
65. Gamble-George JC, Baldi R, Halladay L, Kocharian A, Hartley N, Silva CG, *et al.* (2016): Cyclooxygenase-2 inhibition reduces stress-induced affective pathology. *Elife* 5.
66. Kao CY, He Z, Henes K, Asara JM, Webhofer C, Filiou MD, *et al.* (2016): Fluoxetine treatment rescues energy metabolism pathway alterations in a posttraumatic stress disorder mouse model. *Mol Neuropsychiatry* 2:46–59.
67. Kao CY, He Z, Zannas AS, Hahn O, Kuhne C, Reichel JM, *et al.* (2016): Fluoxetine treatment prevents the inflammatory response in a mouse model of posttraumatic stress disorder. *J Psychiatr Res* 76:74–83.
68. Lee B, Sur B, Yeom M, Shim I, Lee H, Hahm DH (2016): Effects of systemic administration of ibuprofen on stress response in a rat model of post-traumatic stress disorder. *Korean J Physiol Pharmacol* 20:357–366.
69. Levkovitz Y, Fenchel D, Kaplan Z, Zohar J, Cohen H (2015): Early post-stressor intervention with minocycline, a second-generation tetracycline, attenuates post-traumatic stress response in an animal model of PTSD. *Eur Neuropsychopharmacol* 25:124–132.
70. Deslauriers J, van Wijngaarde M, Geyer MA, Powell S, Risbrough VB (2017): Effects of LPS-induced immune activation prior to trauma exposure on PTSD-like symptoms in mice. *Behav Brain Res* 323:117–123.
71. Ebenezer PJ, Wilson CB, Wilson LD, Nair AR, Francis J (2016): The anti-inflammatory effects of blueberries in an animal model of post-traumatic stress disorder (PTSD). *PLoS One* 11:e0160923.
72. Rosenbaum S, Stubbs B, Ward PB, Steel Z, Lederman O, Vancampfort D (2015): The prevalence and risk of metabolic syndrome and its components among people with posttraumatic stress disorder: A systematic review and meta-analysis. *Metabolism* 64:926–933.
73. Wolf EJ, Schnurr PP (2016): PTSD-related cardiovascular disease and accelerated cellular aging. *Psychiatr Ann* 46:527–532.
74. Bastard JP, Maachi M, Lagathu C, Kim MJ, Caron M, Vidal H, *et al.* (2006): Recent advances in the relationship between obesity, inflammation, and insulin resistance. *Eur Cytokine Netw* 17:4–12.
75. Marsland AL, McCaffery JM, Muldoon MF, Manuck SB (2010): Systemic inflammation and the metabolic syndrome among middle-aged community volunteers. *Metabolism* 59:1801–1808.
76. Bartoli F, Carra G, Crocamo C, Carretta D, Clerici M (2013): Metabolic syndrome in people suffering from posttraumatic stress disorder: A systematic review and meta-analysis. *Metab Syndr Relat Disord* 11:301–308.
77. Heni M, Kullmann S, Preissl H, Fritsche A, Haring HU (2015): Impaired insulin action in the human brain: Causes and metabolic consequences. *Nat Rev Endocrinol* 11:701–711.
78. Reagan LP, Grillo CA, Piroli GG (2008): The As and Ds of stress: Metabolic, morphological and behavioral consequences. *Eur J Pharmacol* 585:64–75.
79. Wolf EJ, Bovin MJ, Green JD, Mitchell KS, Stoop TB, Barretto KM, *et al.* (2016): Longitudinal associations between post-traumatic stress disorder and metabolic syndrome severity. *Psychol Med* 46:2215–2226.
80. Gavrieli A, Farr OM, Davis CR, Crowell JA, Mantzoros CS (2015): Early life adversity and/or posttraumatic stress disorder severity are associated with poor diet quality, including consumption of trans fatty acids, and fewer hours of resting or sleeping in a US middle-



## Metabolism, Metabolomics, and Inflammation in PTSD

- aged population: A cross-sectional and prospective study. *Metabolism* 64:1597–1610.
81. Gregor MF, Hotamisligil GS (2011): Inflammatory mechanisms in obesity. *Annu Rev Immunol* 29:415–445.
  82. Koenen KC, Sumner JA, Gilsanz P, Glymour MM, Ratanatharathorn A, Rimm EB, *et al.* (2017): Post-traumatic stress disorder and cardiometabolic disease: Improving causal inference to inform practice. *Psychol Med* 47:209–225.
  83. Zhang D, Wang X, Wang B, Garza JC, Fang X, Wang J, *et al.* (2017): Adiponectin regulates contextual fear extinction and intrinsic excitability of dentate gyrus granule neurons through AdipoR2 receptors. *Mol Psychiatry* 22:1044–1055.
  84. Jha SK, Jha NK, Kumar D, Ambasta RK, Kumar P (2017): Linking mitochondrial dysfunction, metabolic syndrome and stress signaling in neurodegeneration. *Biochim Biophys Acta* 1863:1132–1146.
  85. Wolf EJ, Miller DR, Logue MW, Sumner J, Stoop TB, Leritz EC, *et al.* (2017): Contributions of polygenic risk for obesity to PTSD-related metabolic syndrome and cortical thickness. *Brain Behav Immun* 65:328–336.
  86. Farr OM, Ko BJ, Joung KE, Zaichenko L, Usher N, Tsoukas M, *et al.* (2015): Posttraumatic stress disorder, alone or additively with early life adversity, is associated with obesity and cardiometabolic risk. *Nutr Metab Cardiovasc Dis* 25:479–488.
  87. Tsikunov SG, Klyueva NN, Kusov AG, Vinogradova TV, Klimenko VM, Denisenko AD (2006): Changes in the lipid composition of blood plasma and liver in rats induced by severe psychic trauma. *Bull Exp Biol Med* 141:636–638.
  88. Hammamieh R, Chakraborty N, De Lima TC, Meyerhoff J, Gautam A, Muhie S, *et al.* (2012): Murine model of repeated exposures to conspecific trained aggressors simulates features of post-traumatic stress disorder. *Behav Brain Res* 235:55–66.
  89. Chuang JC, Cui H, Mason BL, Mahgoub M, Bookout AL, Yu HG, *et al.* (2010): Chronic social defeat stress disrupts regulation of lipid synthesis. *J Lipid Res* 51:1344–1353.
  90. Czech B, Neumann ID, Müller M, Reber SO, Hellerbrand C (2013): Effect of chronic psychosocial stress on nonalcoholic steatohepatitis in mice. *Int J Clin Exp Pathol* 6:1585–1593.
  91. Sanghez V, Razzoli M, Carobbio S, Campbell M, McCallum J, Cero C, *et al.* (2013): Psychosocial stress induces hyperphagia and exacerbates diet-induced insulin resistance and the manifestations of the metabolic syndrome. *Psychoneuroendocrinology* 38:2933–2942.
  92. Edmondson D, von Kanel R (2017): Post-traumatic stress disorder and cardiovascular disease. *Lancet Psychiatry* 4:320–329.
  93. Johannessen KB, Berntsen D (2013): Losing the symptoms: Weight loss and decrease in posttraumatic stress disorder symptoms. *J Clin Psychol* 69:655–660.
  94. Jia M, Meng F, Smerin SE, Xing G, Zhang L, Su DM, *et al.* (2012): Biomarkers in an animal model for revealing neural, hematologic, and behavioral correlates of PTSD. *J Vis Exp* e3361.
  95. Cohen H, Kozlovsky N, Savion N, Matar MA, Loewenthal U, Loewenthal N, *et al.* (2009): An association between stress-induced disruption of the hypothalamic-pituitary-adrenal axis and disordered glucose metabolism in an animal model of post-traumatic stress disorder. *J Neuroendocrinol* 21:898–909.
  96. Kalyan-Masih P, Vega-Torres JD, Miles C, Haddad E, Rainsbury S, Baghchechi M, *et al.* (2016): Western high-fat diet consumption during adolescence increases susceptibility to traumatic stress while selectively disrupting hippocampal and ventricular volumes. *eNeuro* 3.
  97. Karabatsiakos A, Hamuni G, Wilker S, Kolassa S, Renu D, Kadereit S, *et al.* (2015): Metabolite profiling in posttraumatic stress disorder. *J Mol Psychiatry* 3:2.
  98. Fielding BA (2017): Omega-3 index as a prognosis tool in cardiovascular disease. *Curr Opin Clin Nutr Metab Care* 20:360–365.
  99. Dyllal SC (2015): Long-chain omega-3 fatty acids and the brain: A review of the independent and shared effects of EPA, DPA and DHA. *Front Aging Neurosci* 7:52.
  100. Clay Montier LL, Deng JJ, Bai Y (2009): Number matters: Control of mammalian mitochondrial DNA copy number. *J Genet Genomics* 36:125–131.
  101. Naviaux RK (2014): Metabolic features of the cell danger response. *Mitochondrion* 16:7–17.
  102. Picca A, Lezza AMS, Leeuwenburgh C, Pesce V, Calvani R, Landi F, *et al.* (2017): Fueling inflamm-aging through mitochondrial dysfunction: Mechanisms and molecular targets. *Int J Mol Sci* 18:e933.
  103. Sahin E, Depinho RA (2010): Linking functional decline of telomeres, mitochondria and stem cells during ageing. *Nature* 464:520–528.
  104. Picard M, Juster RP, McEwen BS (2014): Mitochondrial allostatic load puts the 'gluc' back in glucocorticoids. *Nat Rev Endocrinol* 10:303–310.
  105. Schiavone S, Trabace L (2016): Pharmacological targeting of redox regulation systems as new therapeutic approach for psychiatric disorders: A literature overview. *Pharmacol Res* 107:195–204.
  106. Zhang L, Zhou R, Li X, Ursano RJ, Li H (2006): Stress-induced change of mitochondria membrane potential regulated by genomic and non-genomic GR signaling: A possible mechanism for hippocampus atrophy in PTSD. *Med Hypotheses* 66:1205–1208.
  107. Kao CY, Anderzhanova E, Asara JM, Wotjak CT, Turck CW (2015): NextGen Brain microdialysis: Applying modern metabolomics technology to the analysis of extracellular fluid in the central nervous system. *Mol Neuropsychiatry* 1:60–67.
  108. Filiou MD, Zhang Y, Teplytska L, Reckow S, Gormanns P, Maccarrone G, *et al.* (2011): Proteomics and metabolomics analysis of a trait anxiety mouse model reveals divergent mitochondrial pathways. *Biol Psychiatry* 70:1074–1082.
  109. Wan J, Liu D, Zhang J, Shi Y, Han F (2016): Single-prolonged stress induce different change in the cell organelle of the hippocampal cells: A study of ultrastructure. *Acta Histochem* 118:10–19.
  110. Li XM, Han F, Liu DJ, Shi YX (2010): Single-prolonged stress induced mitochondrial-dependent apoptosis in hippocampus in the rat model of post-traumatic stress disorder. *J Chem Neuroanat* 40:248–255.
  111. Liu D, Xiao B, Han F, Wang E, Shi Y (2012): Single-prolonged stress induces apoptosis in dorsal raphe nucleus in the rat model of post-traumatic stress disorder. *BMC Psychiatry* 12:211.
  112. Xiao B, Yu B, Wang HT, Han F, Shi YX (2011): Single-prolonged stress induces apoptosis by activating cytochrome C/caspase-9 pathway in a rat model of post-traumatic stress disorder. *Cell Mol Neurobiol* 31:37–43.
  113. Xing G, Barry ES, Benford B, Grunberg NE, Li H, Watson WD, *et al.* (2013): Impact of repeated stress on traumatic brain injury-induced mitochondrial electron transport chain expression and behavioral responses in rats. *Front Neurol* 4:196.
  114. Garabadu D, Ahmad A, Krishnamurthy S (2015): Risperidone attenuates modified stress-re-stress paradigm-induced mitochondrial dysfunction and apoptosis in rats exhibiting post-traumatic stress disorder-like symptoms. *J Mol Neurosci* 56:299–312.
  115. Elmquist JK, Scherer PE (2012): The cover. *Neuroendocrine and endocrine pathways of obesity*. *JAMA* 308:1070–1071.
  116. Hernandez-Aguilera A, Rull A, Rodriguez-Gallego E, Riera-Borrull M, Luciano-Mateo F, Camps J, *et al.* (2013): Mitochondrial dysfunction: A basic mechanism in inflammation-related non-communicable diseases and therapeutic opportunities. *Mediators Inflamm* 2013:135698.
  117. Koves TR, Ussher JR, Noland RC, Slentz D, Mosedale M, Ilkayeva O, *et al.* (2008): Mitochondrial overload and incomplete fatty acid oxidation contribute to skeletal muscle insulin resistance. *Cell Metab* 7:45–56.
  118. Kusminski CM, Scherer PE (2012): Mitochondrial dysfunction in white adipose tissue. *Trends Endocrinol Metab* 23:435–443.
  119. Montgomery MK, Turner N (2015): Mitochondrial dysfunction and insulin resistance: An update. *Endocr Connect* 4:R1–R15.
  120. Muoio DM, Newgard CB (2008): Mechanisms of disease: Molecular and metabolic mechanisms of insulin resistance and beta-cell failure in type 2 diabetes. *Nat Rev Mol Cell Biol* 9:193–205.
  121. Green DR, Galluzzi L, Kroemer G (2014): Cell biology. Metabolic control of cell death. *Science* 345:1250–1256.
  122. Thakur GS, Daigle BJ Jr, Dean KR, Zhang Y, Rodriguez-Fernandez M, Hammamieh R, *et al.* (2015): Systems biology

- approach to understanding post-traumatic stress disorder. *Mol Biosyst* 11:980–993.
123. Friedrich N (2012): Metabolomics in diabetes research. *J Endocrinol* 215:29–42.
  124. Swarbrick MM, Havel PJ (2008): Physiological, pharmacological, and nutritional regulation of circulating adiponectin concentrations in humans. *Metab Syndr Relat Disord* 6:87–102.
  125. Hotamisligil GS (2006): Inflammation and metabolic disorders. *Nature* 444:860–867.
  126. Frey WH 2nd (2013): Intranasal insulin to treat and protect against posttraumatic stress disorder. *J Nerv Ment Dis* 201:638–639.
  127. Valero T (2014): Mitochondrial biogenesis: Pharmacological approaches. *Curr Pharm Des* 20:5507–5509.
  128. Rolland B, Deguil J, Jardri R, Cottencin O, Thomas P, Bordet R (2013): Therapeutic prospects of PPARs in psychiatric disorders: A comprehensive review. *Curr Drug Targets* 14:724–732.
  129. Fisman EZ, Tenenbaum A (2014): Adiponectin: A manifold therapeutic target for metabolic syndrome, diabetes, and coronary disease? *Cardiovasc Diabetol* 13:103.
  130. Miller AH, Haroon E, Felger JC (2017): Therapeutic Implications of Brain-Immune Interactions: Treatment in Translation. *Neuropsychopharmacology* 42:334–359.
  131. Yehuda R (2009): Status of glucocorticoid alterations in post-traumatic stress disorder. *Ann N Y Acad Sci* 1179:56–69.
  132. Parikh S, Saneto R, Falk MJ, Anselm I, Cohen BH, Haas R, *et al.* (2009): A modern approach to the treatment of mitochondrial disease. *Curr Treat Options Neurol* 11:414–430.
  133. Kanabus M, Heales SJ, Rahman S (2014): Development of pharmacological strategies for mitochondrial disorders. *Br J Pharmacol* 171:1798–1817.
  134. Friedrich MJ (2012): Studies probe mechanisms that have a role in obesity-associated morbidities. *JAMA* 308:1077–1079.