

UC San Diego

UC San Diego Previously Published Works

Title

Presence of ApoE epsilon 4 Allele Associated with Thinner Frontal Cortex in Middle Age

Permalink

<https://escholarship.org/uc/item/67v1s7qx>

Journal

HANDBOOK OF IMAGING THE ALZHEIMER BRAIN, 2

ISSN

2210-5727

Authors

Fennema-Notestine, Christine

Panizzon, Matthew S

Thompson, Wesley R

et al.

Publication Date

2011

DOI

10.3233/978-1-60750-793-2-77

Peer reviewed

Presence of ApoE ϵ 4 Allele Associated with Thinner Frontal Cortex in Middle Age

Christine Fennema-Notestine^{a,b,*}, Matthew S. Panizzon^a, Wesley R. Thompson^a, Chi-Hua Chen^a, Lisa T. Eyler^{a,c}, Bruce Fischl^{d,e,f}, Carol E. Franz^a, Michael D. Grant^g, Amy J. Jak^{a,c}, Terry L. Jernigan^{a,b,h,i}, Michael J. Lyons^g, Michael C. Neale^j, Larry J. Seidman^g, Ming T. Tsuang^{a,k,l}, Hong Xian^m, Anders M. Dale^{b,n} and William S. Kremen^{a,c,k}

^aDepartment of Psychiatry, University of California, San Diego, La Jolla, CA, USA

^bDepartment of Radiology, University of California, San Diego, La Jolla, CA, USA

^cVeterans' Administration San Diego Healthcare System, San Diego, CA, USA

^dDepartment of Radiology, Massachusetts General Hospital, Boston, MA, USA

^eHarvard Medical School, Boston, MA, USA

^fComputer Science and AI Lab, Massachusetts Institute of Technology, Cambridge, MA, USA

^gDepartment of Psychology, Boston University, Boston, MA, USA

^hDepartment of Cognitive Science, University of California, San Diego, La Jolla, CA, USA

ⁱCenter for Human Development, University of California, San Diego, La Jolla, CA, USA

^jVirginia Institute for Psychiatric and Behavioral Genetics, Virginia Commonwealth University School of Medicine, Richmond, VA, USA

^kCenter for Behavioral Genomics, University of California, San Diego, La Jolla, CA, USA

^lHarvard Institute of Psychiatric Epidemiology and Genetics, Harvard Medical School and School of Public Health, Boston, MA, USA

^mDepartment of Psychiatry, Washington University School of Medicine, St. Louis, MO, USA

ⁿDepartment of Neurosciences, University of California, San Diego, La Jolla, CA, USA

Abstract. The presence of an ApoE ϵ 4 allele (ϵ 4+) increases the risk of developing Alzheimer's disease (AD). Previous studies support an adverse relationship between ϵ 4+ status and brain structure and function in mild cognitive impairment and AD; in contrast, the presence of an ϵ 2 allele may be protective. Whether these findings reflect disease-related effects or pre-existing endophenotypes, however, remains unclear. The present study examined the influence of ApoE allele status on brain structure solely during middle-age in a large, national sample. Participants were 482 men, ages 51–59, from the Vietnam Era Twin Study of Aging (VETSA). T1-weighted images were used in volumetric segmentation and cortical surface reconstruction methods to measure regional volume and thickness. Primary linear mixed effects models predicted structural measures with ApoE status (ϵ 3/3, ϵ 2/3, ϵ 3/4) and control variables for effects of site, non-independence of twin data, age, and average cranial vault or cortical thickness. Relative to the ϵ 3/3 group, the ϵ 3/4 group demonstrated significantly thinner cortex in superior frontal and left rostral and right caudal midfrontal regions; there were no significant effects of ϵ 4 status on any temporal lobe measures.

*Correspondence to: Christine Fennema-Notestine, Ph.D. UCSD School of Medicine 9500 Gilman Dr #0738 La Jolla, CA 92093-0738 Tel.: 858 246 0605; Fax: 858 246 0556; E-mail: Fennema@UCSD.edu.

The $\epsilon 2/3$ group demonstrated significantly thicker right parahippocampal cortex relative to the $\epsilon 3/3$ group. The ApoE $\epsilon 4$ allele may influence cortical thickness in frontal areas, which are later developing regions thought to be more susceptible to the natural aging process. Previous conflicting findings for mesial temporal regions may be driven by the inclusion of older individuals, who may evidence preclinical manifestations of disease, and by unexamined moderators of $\epsilon 4$ -related effects. The presence of the $\epsilon 2$ allele was related to thicker cortex, supporting a protective role. Ongoing follow-up of the VETSA sample may shed light on the potential for age- and disease-related mediation of the influence of ApoE allele status.

Keywords: Magnetic resonance imaging, cerebral cortex, brain, frontal lobe, apolipoproteins E, apolipoprotein E2, apolipoprotein E3, apolipoprotein E4, genetic association studies, aging

INTRODUCTION

The ApoE $\epsilon 4$ allele is studied within imaging genetics as the most common polymorphism associated with late-onset Alzheimer's disease (AD) [1–4]. ApoE is thought to play a role in lipoprotein transport and cell maintenance and repair, including amyloid clearance, and is bound to senile plaques and neurofibrillary tangles [5–7]. Of the three alleles ($\epsilon 2$, $\epsilon 3$, $\epsilon 4$), the $\epsilon 3/3$ pairing is the most common phenotype in the U.S. population ($\sim 60\%$), while the presence of $\epsilon 2$ and $\epsilon 4$ alleles is less frequent [8]. There is an increased prevalence of the $\epsilon 4$ allele in disease populations relative to healthy controls [1, 4, 9–12], and individuals carrying at least one $\epsilon 4$ allele ($\epsilon 4+$) are at an increased risk for developing AD [13–15]. In contrast, the presence of an $\epsilon 2$ allele may impart protection from AD-related neurodegeneration [8, 15–21].

In combination with the risk conferred by ApoE allele status, neuroimaging biomarkers may improve the identification of individuals at risk for AD and the potential for successful intervention in the earliest stages. Studies in AD and mild cognitive impairment (MCI) often demonstrate more significant mesial temporal lobe (MTL) atrophy in $\epsilon 4+$ individuals relative to non-carriers [22–29]. In a positron emission tomography (PET) study using a marker of amyloid and tau proteins (FDDNP), $\epsilon 4+$ MCI demonstrated abnormally high binding in the MTL [30]. Neuropathological studies of $\epsilon 4$ carriers support earlier and greater amyloid deposition in AD, as well as in MCI and in older healthy individuals [20]. Further evidence of an earlier and faster rate of cognitive decline also has been demonstrated in MCI and AD $\epsilon 4+$ individuals [15]. These and other studies support strong disease-related effects within $\epsilon 4+$ MCI and AD individuals.

Studying individuals earlier in life, prior to the development of MCI or AD, is critical to understand-

ing the influence of ApoE allele status. PET studies have shown glucose metabolism reductions in $\epsilon 4+$ late-middle-aged individuals with a positive family history for AD [31, 32] and an accelerated rate of decline in regional cerebral blood flow for $\epsilon 4$ carriers [33]. The affected areas overlap with AD-related regions supporting the potential for a pre-symptomatic endophenotype. Of particular interest, however, a recent PET FDDNP study [30] found higher amyloid and tau binding in frontal areas for $\epsilon 4+$ relative to $\epsilon 4$ -healthy individuals, in contrast to an increased temporal lobe binding in the $\epsilon 4+$ MCI group [30]. Structural neuroimaging studies also have been somewhat inconsistent, with reports of smaller MTL structures, including the hippocampus [34–36] and entorhinal cortex [37, 38], in $\epsilon 4+$ carriers, alongside other reports of no significant $\epsilon 4$ -related effect in these or other areas [24, 37, 39]. Studies beyond the MTL that have included younger-old and middle-aged individuals are varied, reporting thickening of small cortical areas [40], thinning in medial orbitofrontal areas [24], and lower gray matter density in small anterior frontal and temporal regions [36]. Several reports, however, have suggested that such effects may be driven by older individuals in the samples, rather than reflecting an early $\epsilon 4$ -related endophenotype [35, 41].

Fewer studies have examined the potential protective influence of the $\epsilon 2$ allele, particularly in healthy individuals, in part due to the lower prevalence of this allele in the U.S. population. Previous work has provided neuropathological evidence for less cortical amyloid and fewer plaques and neurofibrillary tangles in $\epsilon 2$ carriers ($\epsilon 2+$) [16–18, but see 21]. In addition, $\epsilon 2$ carriers may evidence a reduced rate of cognitive decline [8, 15, 19–21] and fewer are diagnosed with AD [8]. Neuroimaging corroboration for such a protective effect is rarer. A recent study of older individuals reported larger cortical gray matter volume and smaller

ventricles in MCI and AD but found no significant effect related to the $\epsilon 2$ allele in healthy older individuals; the sample sizes for $\epsilon 2$ carriers, however, were quite small across all groups studied [42]. A study of adolescents suggested a tendency for thicker mesial temporal and medial orbitofrontal cortex in a larger $\epsilon 2+$ group [38]. An investigation of the $\epsilon 2$ allele in a large community sample may provide complementary insight into the potentially opposing influences of ApoE $\epsilon 4$ and $\epsilon 2$ alleles.

The present study examined the influence of ApoE allele status on brain structure solely during middle age in a national sample from the Vietnam Era Twin Study of Aging (VETSA). This cohort captures individuals in their 6th decade of life likely prior to the onset of AD or other age-related complications [43]. We examined *a priori* AD-related regions of interest (ROIs) as well as regions expected to be influenced by normal aging, which tend to follow an anterior–posterior gradient, exhibiting the greatest rates of decline in frontal areas [44, 45]. Relative to $\epsilon 3/3$ carriers, we expected the $\epsilon 3/4$ group to show the smallest and thinnest MTL areas, most affected in AD, and we also proposed that this group would demonstrate thinner frontal cortex, associated with normal aging. In contrast, the $\epsilon 2/3$ group may evidence larger, thicker MTL areas, supporting a potential protective effect. Continuous surface maps were also generated to explore the extent of effects without the constraints of predefined boundaries.

MATERIALS AND METHODS

Participants

Data were obtained in the first wave of VETSA, a longitudinal study of cognitive and brain aging beginning in midlife [46]. Participants were randomly sampled from over 3,300 Vietnam Era Twin (VET) Registry twin pairs with the constraint that they were in their 50s at the time of recruitment into VETSA. The VET Registry is a nationally distributed sample of male-male twin pairs who served in the U.S. military sometime between 1965 and 1975; descriptions of the composition and method of ascertainment have been reported elsewhere [47]. Importantly, these are Vietnam era, not necessarily Vietnam, veterans; the large majority did not serve in combat. In comparison to census data, VETSA participants are similar in demographic and health characteristics to American men

in their age range [48]. Aside from standard exclusion criterion for MRI studies (e.g., metal in the body), there were no additional eligibility requirements for selection into the MRI component.

Participants traveled either to Boston University or the University of California, San Diego (UCSD) for a series of physical, psychosocial, and neurocognitive assessments. Informed consent was obtained from all participants prior to data collection, and the scanning protocol was approved of by the Institutional Review Boards at UCSD, Boston University, and the Massachusetts General Hospital (MGH).

A subset of the 1237 VETSA participants underwent structural MRI, and the present non-twin analyses include data from 482 participants for whom neuroimaging data and APOE genotyping were adequate and available. The dataset included 205 twin pairs (119 monozygotic and 86 dizygotic twin pairs) and 72 unpaired individuals with an average age of 55.7 years ($sd = 2.6$; range 51–59). Participants in this MRI study were similar to the larger VETSA sample with respect to education (mean=13.8; $sd=2.1$), ethnicity (85.7% Caucasian), employment (75% employed full-time), and self-reported health status.

ApoE genotype was determined from blood samples using established methods [49, 50]. All genotypes were independently determined twice by laboratory personnel at the VA Puget Sound Healthcare System who were blind to the initial genotype and the identity of the co-twin. Of the 482 participants, 2 (0.4%) possessed a $\epsilon 2/2$ genotype, 67 (13.9%) $\epsilon 2/3$, 18 (3.7%) $\epsilon 2/4$, 288 (59.8%) $\epsilon 3/3$, 94 (19.5%) $\epsilon 3/4$, and 13 (2.7%) $\epsilon 4/4$ (Table 1). These rates are roughly equivalent to those found in the general population [14, 51]. Because the proportion of individuals with $\epsilon 2/2$, $\epsilon 2/4$, and $\epsilon 4/4$ pairings were small, these cases were not included in the primary models, however, a secondary overall analyses comparing $\epsilon 4+$ and non- $\epsilon 4$ carriers was completed using all available data.

Participants studied for the primary model were classified as $\epsilon 2/3$, $\epsilon 3/3$, or $\epsilon 3/4$ (see Table 1). These groups did not differ on age ($F=1.4$, $p>.05$). General cognitive ability was assessed by the Armed Forces Qualification Test (AFQT), a well-validated test that also was given to VETSA participants in early adulthood [52]. The mean for the entire sample was 63.1 ($sd=20.8$); this AFQT score is slightly above the mean and would be comparable to an average IQ of approximately 105. The mean across the three primary groups was 63.2 ($sd=20.6$) and the means did not differ between these groups (Table 1; $F<1.0$, $p>.05$).

Table 1

Participant characteristics. For all ApoE allele pairings, sample size, mean age in years, and mean Armed Forces Qualifications Test (AFQT) score are provided, along with the standard deviation (sd) and *range*. The primary model considered the first three groups ($\epsilon 2/3$, $\epsilon 3/3$, $\epsilon 3/4$) with sufficient power to examine influence on brain structure

	$\epsilon 2/3$	$\epsilon 3/3$	$\epsilon 3/4$	$\epsilon 2/2$	$\epsilon 2/4$	$\epsilon 4/4$
<i>n</i>	67	288	94	2	18	13
<i>age</i>	56.2 (2.5) 52–59	55.7 (2.6) 51–59	55.5 (2.8) 51–59	55.0 (4.2) 52–58	55.1 (2.5) 52–58	56.2 (2.6) 51–58
<i>AFQT (%)</i>	63.2 (22.7) 15–97	62.8 (19.8) 4–95	64.4 (21.5) 15–98	71.5 (27.6) 52–91	55.0 (24.4) 14–94	70.9 (18.9) 28–89

MR Image Acquisition

As described previously [53], images were acquired on 1.5 Tesla scanners (255 at UCSD; 227 at MGH). Sagittal T1-weighted MPRAGE sequences were employed with TI=1000ms, TE=3.31ms, TR=2730ms, flip angle=7degrees, slice thickness = 1.33 mm, voxel size $1.3 \times 1.0 \times 1.3$ mm. Raw DICOM MRI scans (two T1 volumes per case) were transferred to MGH for image processing. These raw data were reviewed for quality, registered, and averaged to improve signal-to-noise.

Image processing

As described elsewhere [53], we employed volumetric segmentation [54] and cortical surface reconstruction [55–57] methods based on the publicly available FreeSurfer software package (<http://surfer.nmr.mgh.harvard.edu/fswiki>; Version 3.0.1b). The 3D whole-brain segmentation procedure [54] uses a probabilistic atlas and applies a Bayesian classification rule to assign a neuroanatomical label to each voxel. The atlas consists of a manually-derived training set created by the Center for Morphometric Analysis (<http://www.cma.mgh.harvard.edu/>) from 20 unrelated, randomly selected VETSA participants. Use of this study-specific atlas produced more accurate measurements than more commonly used atlases [53]. Estimated total cranial vault (eTIV) volume was calculated to control for differences in head size for volumetric measures. Based on Buckner et al. [58], FreeSurfer provides an eTIV volume derived from the atlas scaling factor on the basis of the transformation of the full brain mask into atlas space. Although this estimate is not a direct volume, this eTIV measure has been shown to correlate well with other cranial vault volumes incorporating T2-weighted information, including manual tracings in controls and individuals with Alzheimer's Disease ($r=0.93$) [58] and multi-channel tissue segmentations [as in 44] in older controls and

individuals with Alzheimer's disease ($r=0.87$) [59]. The primary volumetric ROI was the hippocampus; exploratory ROIs included amygdala, caudate nucleus, putamen, nucleus accumbens, and thalamus.

The cortical surface was reconstructed to measure thickness at each surface location, or vertex [described in 53, 55, 56]. The explicit reconstruction of the cortical surface requires inhomogeneity corrections, creation of a normalized intensity image, and removal of non-brain. The resulting surface is covered with a polygonal tessellation and smoothed to reduce metric distortions. The gray/white boundary surface is deformed outwards to obtain a representation of the pial surface; the surface model is manually reviewed and edited for technical accuracy in alignment with standard, objective editing rules. Each individual surface is non-rigidly aligned to an atlas in a spherical surface-based coordinate system and divided into distinct ROIs [57], with each vertex assigned a neuroanatomical label [60], to estimate average thickness in each ROI. Primary cortical thickness ROIs included mesial temporal (entorhinal, parahippocampal); lateral temporal (inferior, middle, and superior temporal); and frontal (caudal and rostral middle; superior; inferior; orbitofrontal) cortex (Fig. 1). Exploratory ROIs included superior and inferior parietal, supramarginal, lingual, fusiform, cingulate, and precuneus cortex. Cortical thickness was also estimated over continuous maps on the surface with no predefined regional boundaries as described in Statistical Analysis; smoothing of volumes was done prior to the vertex-wise analyses using a 30 mm FWHM Gaussian kernel.

Statistical analysis

Although the study participants were twins, all analyses in this article are non-twin analyses. Derived ROI values (thickness in mm or volume in mm^3) were submitted to linear mixed effects models with fixed effects of site, ApoE allele status ($\epsilon 2/3$ and $\epsilon 3/4$ were compared to $\epsilon 3/3$), and age. Importantly, site was included

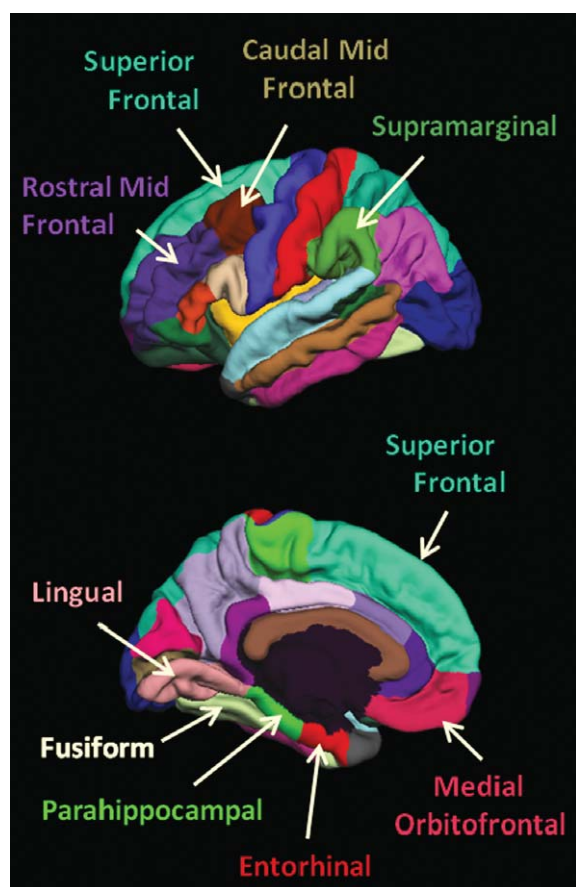


Fig. 1. Cortical region of interest parcellation (30). Primary ROIs include superior frontal (teal), rostral (purple) and caudal (brown) mid frontal, parahippocampal (green), entorhinal (red), and medial orbitofrontal (rose) cortex. Additional exploratory ROIs include fusiform (yellow), supramarginal (olive green), and lingual (pink) gyrus. Top row: lateral views; bottom row: medial views.

in the model to control for effects related to differences in scanner hardware, known to differentially influence morphometric measures of volume and thickness [e.g., 59, 61–63]. Because twins within pairs are not independent observations, it is necessary to adjust for this non-independence when performing non-twin analyses in a twin sample. Therefore, the “family ID” of each member of a twin pair was entered as a random effect in the model. Doing so adjusts the degrees of freedom and makes it more difficult to attain statistical significance. Finally, to adjust for individual differences in overall head size or thickness of the cortical ribbon, an additional fixed effect was included in each model: eTIV for volumetric measures and average cortical thickness for thickness measures.

Planned comparisons included ROIs implicated in AD: hippocampus, entorhinal cortex, parahippocampal cortex, and lateral temporal gyri; and regions susceptible to the effects of normal aging: superior frontal gyrus, middle frontal gyrus (rostral and caudal), inferior frontal (pars opercularis, pars orbitalis and pars triangularis), and orbitofrontal cortex (medial and lateral). Planned comparisons were limited to these predefined ROIs driven by prior work, and we employed an alpha level of 0.05. Effect sizes were calculated by ROI using Cohen’s d and were based on estimated marginal means resulting from the full model. In general, a Cohen’s d of 0.2–0.3 is considered a small, 0.5 a medium, and 0.8 a large effect size.

In a secondary analysis, the same model was modified to utilize the entire cohort of 482 participants to compare $\epsilon 4+$ ($n=125$) and non- $\epsilon 4$ ($n=357$) carriers, as has been done in some previous studies. That is, the variables for ApoE allele status were replaced by ApoE $\epsilon 4$ allele status. Given the small sample of homozygous $\epsilon 4/4$ genotype, a dose effect of $\epsilon 4$ (0, 1, or 2 alleles) was not examined due to insufficient power.

To further explore the statistical findings based on our *a priori* ROI analyses, the same model was implemented at each vertex, or point on the cortical surface, resulting in a continuous surface map of cortical thickness without the predefined constraints of ROI boundaries. The resulting map is exploratory in nature and provides guidance for future studies.

RESULTS

There was no significant effect of ApoE allele status on eTIV or average cortical thickness (all $t \leq 1.0$, $p > 0.05$). Relative to the $\epsilon 3/3$ group, the $\epsilon 3/4$ group demonstrated significantly thinner cortex in bilateral superior frontal, left rostral midfrontal, and right caudal midfrontal regions (Table 2; Fig. 2). Although the right rostral midfrontal and left caudal midfrontal ROIs tended to be thinner, these effects were not significant (Table 2). No temporal areas or any other frontal regions were significantly related to $\epsilon 4$ status. Analysis of the entorhinal cortex did not reveal any significant influence of $\epsilon 4$ status, although the variability of thickness in this area was larger than in other ROIs (Table 2). Exploratory cortical analyses suggested thicker fusiform cortex in the $\epsilon 3/4$ group (Table 2, bottom). Primary volumetric analyses did not reveal any significant effect of $\epsilon 4$ status on the hippocampus (Table 2). Exploratory volumetric analyses

Table 2

Effect of ApoE allele status by Regions of Interest. Controlling for other variables in the model, the estimated marginal mean (in mm for cortical thickness and in mm^3 for volumetric ROIs) and standard deviation (sd) are reported by ROI for each primary ApoE allele group. Based on results of the full statistical model, within which the $\epsilon 3/4$ and $\epsilon 2/3$ groups were compared to the $\epsilon 3/3$ group, the resultant t -value, level of significance, and the associated effect size (Cohen's d) are reported. Negative t -values reflect an effect thinner or smaller than the $\epsilon 3/3$ group; positive t -values reflect thicker or larger effects relative to the $\epsilon 3/3$ group

Region of Interest	Hemi-sphere	$\epsilon 3/3$ mean (sd)	$\epsilon 3/4$ mean (sd)	t value	d	$\epsilon 2/3$ mean (sd)	t value	d
Superior Frontal Gyrus	right	2.204 (0.080)	2.179 (0.087)	-2.80**	-0.30	2.202 (0.085)	<1 ^{ns}	-0.02
	left	2.195 (0.084)	2.173 (0.091)	-2.42*	-0.26	2.189 (0.089)	<1 ^{ns}	-0.07
Rostral Mid Frontal Gyrus	right	1.819 (0.080)	1.803 (0.090)	-1.73 ^{ns}	-0.19	1.820 (0.088)	<1 ^{ns}	0.01
	left	1.851 (0.075)	1.833 (0.085)	-1.95*	-0.21	1.859 (0.083)	<1 ^{ns}	0.11
Caudal Mid Frontal Gyrus	right	2.052 (0.121)	2.018 (0.133)	-2.49*	-0.27	2.023 (0.129)	-1.80 ^{ns}	-0.23
	left	2.042 (0.112)	2.028 (0.126)	-1.12 ^{ns}	-0.12	2.029 (0.122)	<1 ^{ns}	-0.11
Entorhinal Cortex	right	2.800 (0.385)	2.883 (0.426)	+1.89 ^{ns}	0.20	2.816 (0.415)	<1 ^{ns}	0.04
	left	2.553 (0.336)	2.574 (0.378)	<1 ^{ns}	-0.06	2.628 (0.367)	+1.68 ^{ns}	0.21
Hippocampal Volume	right	4216 (444)	4168 (479)	<1 ^{ns}	-0.11	4289 (466)	+1.28 ^{ns}	0.16
	left	3988 (399)	3954 (430)	<1 ^{ns}	-0.08	4067 (419)	+1.54 ^{ns}	0.19
Parahippocampal Gyrus	right	1.901 (0.242)	1.922 (0.266)	<1 ^{ns}	-0.09	1.967 (0.259)	+2.10*	0.27
	left	1.900 (0.260)	1.894 (0.288)	<1 ^{ns}	-0.02	1.916 (0.280)	<1 ^{ns}	0.06
Medial Orbitofrontal	right	1.847 (0.159)	1.838 (0.175)	<1 ^{ns}	-0.06	1.851 (0.170)	<1 ^{ns}	0.02
	left	1.849 (0.156)	1.838 (0.175)	<1 ^{ns}	-0.06	1.882 (0.170)	+1.59 ^{ns}	0.20
Fusiform Gyrus	right	2.011 (0.101)	2.035 (0.114)	+2.04*	0.22	2.000 (0.110)	<1 ^{ns}	-0.11
	left	1.975 (0.106)	2.000 (0.119)	+1.88 ^{ns}	0.20	1.968 (0.115)	<1 ^{ns}	-0.06
Putamen Volume	right	5002 (558)	4846 (582)	-2.60*	-0.27	5010 (567)	<1 ^{ns}	0.01
	left	4942 (582)	4788 (598)	-2.50*	-0.26	4868 (582)	-1.04 ^{ns}	-0.13
Supramarginal Gyrus	right	2.085 (0.110)	2.089 (0.124)	<1 ^{ns}	0.04	2.054 (0.120)	-2.08*	-0.26
	left	2.071 (0.099)	2.076 (0.112)	<1 ^{ns}	0.05	2.071 (0.109)	<1 ^{ns}	0.00
Lingual Gyrus	right	1.703 (0.093)	1.715 (0.104)	+1.12 ^{ns}	0.12	1.700 (0.101)	<1 ^{ns}	-0.03
	left	1.654 (0.095)	1.655 (0.105)	<1 ^{ns}	0.01	1.630 (0.102)	-1.97*	-0.25

** $p < 0.01$, * $p < 0.05$, ns = $p > 0.05$

suggested a significantly smaller putamen volume in $\epsilon 4$ carriers (Table 2, bottom).

In the secondary ROI analyses utilizing the entire 482 datasets, a comparison of all $\epsilon 4+$ with all non- $\epsilon 4$ carriers provided similar results. Given the significant pattern of effects in the ROI analyses, we reviewed the influence of $\epsilon 4$ allele status on the continuous cortical surface map to explore effects without the predefined constraints of ROI boundaries. Continuous maps of the cortical surface supported a broad distribution of thinner lateral and mesial superior frontal, and thicker fusiform cortex in the $\epsilon 4+$ relative to the non- $\epsilon 4$ group (Fig. 3). In the context of Fig. 1, Fig. 3 shows that some regional effects (e.g., left middle frontal area) fall across the confines of predefined ROIs. The thinnest areas (in orange/yellow) may lie across the intersection of a number of ROIs, to include the more lateral, inferior extent of the caudal midfrontal gyrus, posterior rostral midfrontal gyrus, posterior inferior frontal gyrus, and inferior portions of the pre-central gyrus. With respect to the unexpectedly thicker regions in the $\epsilon 4+$ group suggested by exploratory ROI analyses, posterior regions may be more broadly affected. This map provides guidance for future studies.

Relative to the $\epsilon 3/3$ group, our investigation of the $\epsilon 2/3$ group demonstrated significantly thicker right parahippocampal cortex (Table 2; Fig. 2) and non-significant tendencies towards thicker left entorhinal cortex, left medial orbitofrontal cortex, and hippocampal volumes (Table 2). Exploratory analyses suggested significantly thinner right supramarginal and left lingual gyri (Table 2, bottom).

DISCUSSION

This study of a large, community-dwelling sample provides a comprehensive view of the influence of ApoE allele status on brain structure in men. Few previous studies have captured such a broad description particularly within a solely middle-aged sample. The findings suggest that carriers of the $\epsilon 4$ allele on average have thinner frontal cortices in middle age, without direct evidence of any significant $\epsilon 4$ effect on MTL regions commonly affected in AD. These frontal effects were widespread, although the effect sizes were small, suggesting that studies with smaller sample sizes may not have sufficient power to reliably detect such

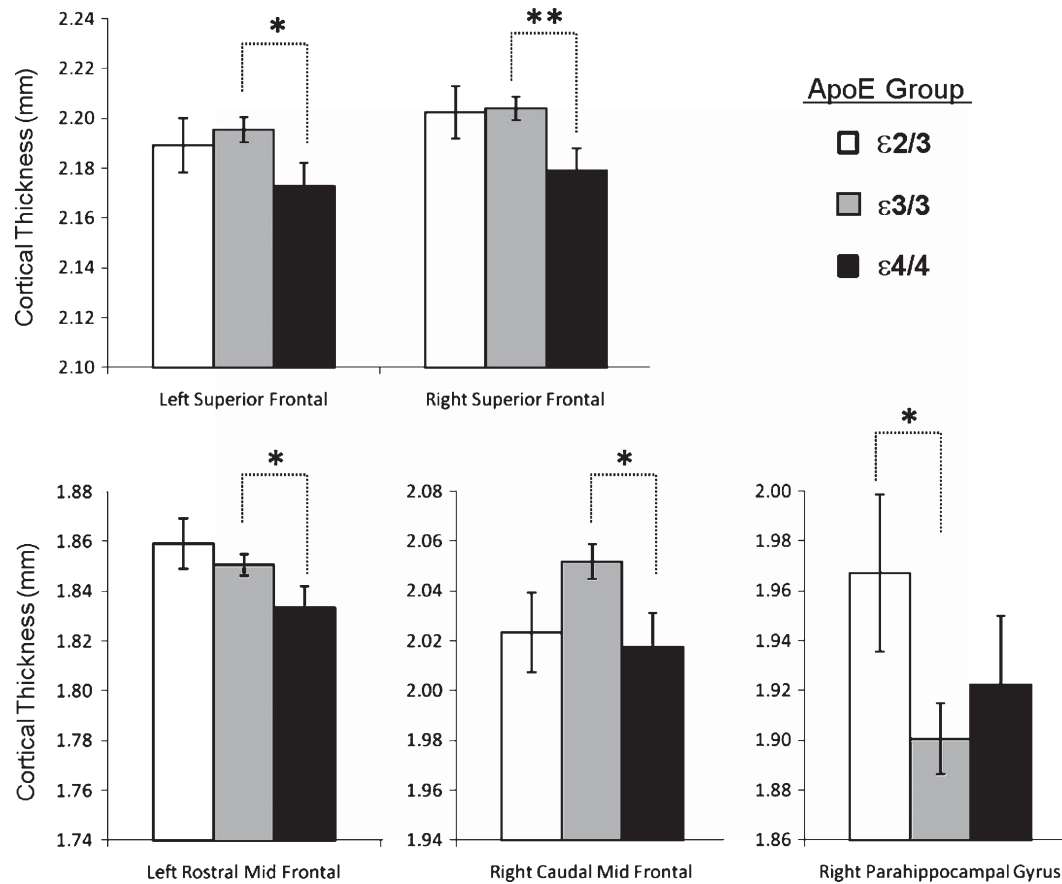


Fig. 2. Cortical thickness in ROIs with significant effects of ApoE allele status. Estimated marginal means (with standard error bars), controlling for all variables in the full model, are shown for cortical thickness (mm) in frontal and temporal ROIs that demonstrated a significant effect of ApoE allele status. Significance levels are reported in Table 2 and denoted in the graph with ** for $p < .01$ and * for $p < .05$.

effects. Exploratory analyses also suggested thicker fusiform cortex in $\epsilon 4+$ carriers, in line with a previous study [40]. Potential protective effects of the $\epsilon 2/3$ genotype were supported in part by thicker left parahippocampal cortex and broader MTL and medial orbitofrontal tendencies towards thicker cortex, relative to the $\epsilon 3/3$ group.

Our findings of $\epsilon 4$ -related differences in superior and middle frontal cortical thickness are relatively unique and of interest in the context of normal aging. One cross-sectional study including middle-aged and older individuals suggested accelerated age-related thinning in $\epsilon 4$ carriers in the superior medial frontal gyrus; however, the majority of the participants were over the age of 60, limiting the inference of effects in middle age [40]. Within Shaw et al.'s study of children and adolescents [38], there were potential $\epsilon 4$ status effects in frontal regions, with continuous maps showing small areas of thinner orbitofrontal cortex in the

$\epsilon 4+$ group. While the present study does not show significant orbitofrontal ROI effects, the continuous surface maps (Fig. 3) further explore patterns without the predefined constraints of ROI boundaries, which may be relatively arbitrary with respect to the underlying cellular, functional, or developmental aspects of the brain. The maps support widespread frontal effects, and the potential influence on frontal cortex development into the adult age range may inform these regional differences.

The $\epsilon 4$ -related effects on frontal cortical thickness are bolstered by findings from other modalities and disorders. Amyloid and tau binding PET studies in healthy individuals suggest that binding is higher for $\epsilon 4+$ carriers in frontal areas, as opposed to commonly reported increased temporal lobe binding in $\epsilon 4+$ MCI individuals [30]. In addition, $\epsilon 4$ status may influence dendritic density and complexity in the cortex [64], and may differentially influence cortical patterns of

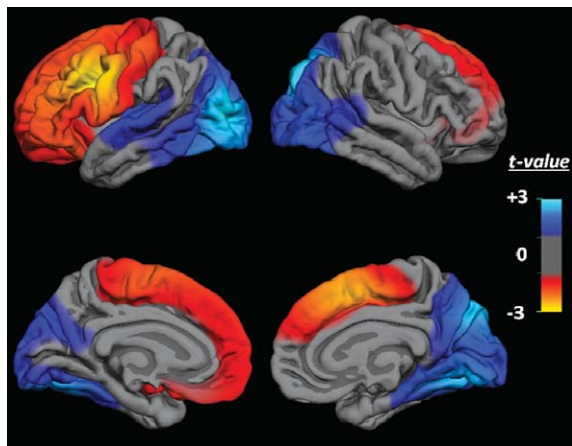


Fig. 3. Continuous surface maps of the estimated ApoE allele status effect on cortical thickness. Using the entire available sample ($n=482$), the t -statistic for the effect of carrying the $\epsilon 4$ allele, from the full statistical model, was applied vertex-wise on the pial surface. The color scale denotes effects for the $\epsilon 4+$ relative to the non- $\epsilon 4$ group as follows: thinner cortex in orange/yellow areas (larger negative t -values) and thicker cortex in areas with bright blue (cyan) (larger positive t -values). Both left (*left column*) and right (*right column*) hemispheres are presented.

thinning based on mediating factors. In a study of AD and frontotemporal dementia, cortical atrophy was greater in both $\epsilon 4+$ subgroups; however, the pattern of thinning in AD represented known neuropathological areas such as the mesial temporal lobe, whereas in frontotemporal dementia, the $\epsilon 4+$ group evidenced greater frontal atrophy [65]. The broad, frontal findings support the relationship between the $\epsilon 4$ allele and increased amyloid deposition in these areas with normal aging, although any progressive nature of such effects must be demonstrated in a longitudinal study, currently underway.

The lack of significant MTL $\epsilon 4$ related effects is not unexpected given conflicting previous reports and may reflect studies including a low proportion of individuals in a preclinical phase of AD and, importantly, other mediating influences on the impact of $\epsilon 4$ status, such as gender and hormones. While substantial support exists for $\epsilon 4$ -related MTL effects in MCI and AD, findings in healthy individuals are inconsistent, even in older adults [23, 37, 39, 40, 66]. There is some evidence suggesting the influence of $\epsilon 4$ status on MTL structures in middle age [34] and in children and adolescents [38]; however, other studies including middle-aged individuals have not found the same effects [35, 40] or have found that effects across a broad age range were driven by individuals over 60 or 65 years of age [35]. Some of

these older individuals may demonstrate poor cognitive performance relative to their non- $\epsilon 4$ counterparts and some may be in the prodromal stages of AD. Indeed, a recent study of cognition suggests that family history of AD and $\epsilon 4$ status may be additive factors, and that, with the removal of individuals known to convert to AD, only individuals with both a positive family history of AD and $\epsilon 4+$ status demonstrate a more rapid cognitive decline [67]. The present sample represents individuals in their 6th decade of life, when few are likely to be affected by dementia, although we do not have data on family history at this time. In contrast, the unique study of children and adolescents ($n=174$ non- $\epsilon 4$, $n=65$ $\epsilon 4+$; 8–21 years) provides support for the thinner left entorhinal cortex for $\epsilon 4+$ individuals [38], although these effects were subtle and the variability in thickness was slightly larger within the $\epsilon 4+$ relative to the non- $\epsilon 4$ group, similar to the present study. These findings together support the hypothesis that additional factors likely mediate the influence of $\epsilon 4$ status on brain structure.

Other studies have demonstrated differences in $\epsilon 4$ -related effects by gender and report potential mediating or moderating factors such as hormones. There may be an interaction between gender and ApoE $\epsilon 4$ status [68] such that, in general, females are more influenced by $\epsilon 4$ status than males. In MCI, female $\epsilon 4+$ carriers have a higher risk of developing AD than men of the same genotype [14]. A neuroimaging study reported that female, but not male, $\epsilon 4+$ carriers had significantly smaller hippocampal volumes relative to non- $\epsilon 4$ individuals; the authors suggested the potential for hormonal mediation of the influence of $\epsilon 4$ status [69]. It is possible, then, that in the present male sample, $\epsilon 4$ -related MTL effects may be reduced and/or obscured by other factors. In fact, a study of VETSA participants revealed a significant interaction between testosterone and $\epsilon 4$ status indicating that $\epsilon 4+$ men who also had low levels of testosterone have smaller hippocampal volumes [70]. A similar interaction between $\epsilon 4$ status and cortisol levels or patterns also has been observed with respect to cognition in older adults [71].

The present study also included a larger sample, relative to published reports [e.g., 42], that allowed for a characterization of the influence of carrying an $\epsilon 2$ allele in middle-aged individuals. In contrast to $\epsilon 4$ status, the $\epsilon 2$ allele appears to have a subtle impact on thickness in MTL and medial orbitofrontal areas. The significantly thicker right parahippocampal cortex and broader tendencies for thicker cortex in these areas lend support to findings in adolescents [38] and corroborate the protec-

tive influence of $\epsilon 2$ demonstrated in neuropathological and cognitive studies [8, 15–21]. Exploratory analyses suggesting thinner right supramarginal and left lingual cortex are intriguing but require further replication.

The unique VETSA cohort provided significant power to examine the influence of ApoE allele status, although the study presents some limitations to generalizability. Because our sample was solely male and largely Caucasian, we cannot be certain of the generalizability of these findings to women or ethnic minorities. Furthermore, although the sample is quite similar in health and demographics to comparably-aged men in the U.S., a minority of them did experience varying amounts of combat exposure 35 years earlier. Thus, concerns might be raised as to the effect of combat exposure or possible posttraumatic stress disorder (PTSD) on the results. As of their mid-40s, 7.7% had a lifetime diagnosis of PTSD, slightly higher than the 5.0% prevalence for men nationally [72]. Importantly, this is unlikely to be create a confound in the present study because previous co-twin control findings indicate that smaller hippocampal volume may be a risk factor for PTSD, rather than a consequence [73]. Another potential limitation of our study is that, with T1-based image processing approaches, it is difficult to distinguish tentorium cerebelli from cortex in some mesial and inferior temporal regions. That is, while we have made every effort to separate cortical gray matter from tentorium, thickness estimates in these regions, such as the entorhinal cortex, may be more variable than in other areas. Such an increase in variability may result in less power to detect significant effects of ApoE allele status on thickness, although we would not expect differential effects across ApoE groups.

CONCLUSION

This study of middle-aged men suggests that the presence of the ApoE $\epsilon 4$ allele may influence cortical thickness in frontal areas, later developing regions thought to be more susceptible to natural aging. In contrast, previous conflicting findings of $\epsilon 4$ effects on MTL regions may be driven by the inclusion of older individuals who may evidence preclinical manifestations of neurodegenerative disease, and by moderators of $\epsilon 4$ -related effects, such as hormone levels. The finding of unexpectedly thicker fusiform cortex in the $\epsilon 4+$ group needs to be explored further and replicated. The examination of the $\epsilon 2$ allele supports a protective role, suggesting tendencies for thicker cor-

tex in some MTL and orbitofrontal areas, although some exploratory areas were thinner. Whether these $\epsilon 2$ and $\epsilon 4$ related findings reflect pre-existing endophenotypes or early neurodegeneration is not clear in these cross-sectional data. Ongoing follow-up studies of the VETSA sample may shed light on the potential for age- and disease-related mediation of the influence of ApoE allele status, as these participants enter the age range within which normal age-related neurodegeneration along with memory decline in $\epsilon 4+$ individuals may accelerate [43, 74].

ACKNOWLEDGMENTS

The VETSA project is supported by National Institutes of Health (NIH) Grants R01 AG018386, R01 AG018384, R01 AG022381, R01 AG022982, and U24 RR021382. The U.S. Department of Veterans Affairs has provided support for the development and maintenance of the Vietnam Era Twin Registry. Additional support for this research was provided in part by NIH grants P41 RR14075, R01 EB006758, and R01 NS052585-01, and the Autism & Dyslexia Project funded by the Ellison Medical Foundation. Dr. A. M. Dale is a founder and holds equity in CorTechs Labs, and also serves on its Scientific Advisory Board. The terms of this arrangement have been reviewed and approved by the University of California, San Diego, in accordance with its conflict of interest policies.

Numerous organizations have provided invaluable assistance, including VA Cooperative Studies Program; Department of Defense; National Personnel Records Center, National Archives and Records Administration; the Internal Revenue Service; National Institutes of Health; National Opinion Research Center; National Research Council, National Academy of Sciences; the Institute for Survey Research, Temple University; Schulman, Ronca, and Bucuvalas, Inc. Most importantly, we gratefully acknowledge the cooperation and participation of the members of the Vietnam Era Twin Registry and their families. Without their contribution this research would not have been possible.

REFERENCES

- [1] Saunders AM, Strittmatter WJ, Schmechel D, George-Hyslop PH, Pericak-Vance MA, Joo SH, Rosi BL, Gusella JF, Crapper-MacLachlan DR, Alberts MJ, Hulette C, Crain B, Goldgaber D, Roses AD (1993) Association of apolipoprotein

- tein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. *Neurology* **43**, 1467-1472.
- [2] Bookheimer S, Burggren A (2009) APOE-4 genotype and neurophysiological vulnerability to Alzheimer's and cognitive aging. *Annu Rev Clin Psychol* **5**, 343-362.
 - [3] Pericak-Vance MA, Bebout JL, Gaskell PC Jr, Yamaoka LH, Hung WY, Alberts MJ, Walker AP, Bartlett RJ, Haynes CA, Welsh KA, Earl NL, Heyman A, Clark CM, Roses AD (1991) Linkage studies in familial Alzheimer disease: evidence for chromosome 19 linkage. *Am J Hum Genet* **48**, 1034-1050.
 - [4] Strittmatter WJ, Saunders AM, Schmechel D, Pericak-Vance M, Enghild J, Salvesen GS, Roses AD (1993) Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc Natl Acad Sci U S A* **90**, 1977-1981.
 - [5] Siest G, Pillot T, Regis-Bailly A, Leininger-Muller B, Steinmetz J, Galteau MM, Visvikis S (1995) Apolipoprotein E: an important gene and protein to follow in laboratory medicine. *Clin Chem* **41**, 1068-1086.
 - [6] Mahley RW (1988) Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science* **240**, 622-630.
 - [7] Schipper H.M. Apolipoprotein E: Implications for AD neurobiology, epidemiology and risk assessment. *Neurobiol Aging*. (In Press)
 - [8] Corder EH, Saunders AM, Risch NJ, Strittmatter WJ, Schmechel DE, Gaskell PC, Rimmler JB, Locke PA, Conneally PM, Schmechel KE, Small GW, Roses AD, Haines JL, Pericak-Vance MA (1994) Protective effect of apolipoprotein E type 2 allele for late onset Alzheimer disease. *Nat Genet* **7**, 180-184.
 - [9] DeCarli C, Reed T, Miller BL, Wolf PA, Swan GE, Carmelli D (1999) Impact of apolipoprotein E epsilon4 and vascular disease on brain morphology in men from the NHLBI twin study. *Stroke* **30**, 1548-1553.
 - [10] Finch CE, Sapolsky RM (1999) The evolution of Alzheimer disease, the reproductive schedule, and apoE isoforms. *Neurobiol Aging* **20**, 407-428.
 - [11] Bleumink GS, van Duijn CM, Kingma JH, Witteman JC, Hofman A, Stricker BH (2004) Apolipoprotein E epsilon4 allele is associated with left ventricular systolic dysfunction. *Am Heart J* **147**, 685-689.
 - [12] Fazekas F, Enzinger C, Ropele S, Schmidt H, Schmidt R, Strasser-Fuchs S (2006) The impact of our genes: consequences of the apolipoprotein E polymorphism in Alzheimer disease and multiple sclerosis. *J Neurol Sci* **245**, 35-39.
 - [13] Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, Roses AD, Haines JL, Pericak-Vance MA (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* **261**, 921-923.
 - [14] Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, Myers RH, Pericak-Vance MA, Risch N, van Duijn CM (1997) Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. *JAMA* **278**, 1349-1356.
 - [15] Martins CA, Oulhaj A, de Jager CA, Williams JH (2005) APOE alleles predict the rate of cognitive decline in Alzheimer disease: a nonlinear model. *Neurology* **65**, 1888-1893.
 - [16] Lippa CF, Smith TW, Saunders AM, Hulette C, Pulaski-Salo D, Roses AD (1997) Apolipoprotein E-epsilon 2 and Alzheimer's disease: genotype influences pathologic phenotype. *Neurology* **48**, 515-519.
 - [17] Tiraboschi P, Hansen LA, Masliah E, Alford M, Thal LJ, Corey-Bloom J (2004) Impact of APOE genotype on neuropathologic and neurochemical markers of Alzheimer disease. *Neurology* **62**, 1977-1983.
 - [18] Oyama F, Shimada H, Oyama R, Ihara Y (1995) Apolipoprotein E genotype, Alzheimer's pathologies and related gene expression in the aged population. *Brain Res Mol Brain Res* **29**, 92-98.
 - [19] Blacker D, Lee H, Muzikansky A, Martin EC, Tanzi R, McArdle JJ, Moss M, Albert M (2007) Neuropsychological measures in normal individuals that predict subsequent cognitive decline. *Arch Neurol* **64**, 862-871.
 - [20] Baum L, Chen L, Ng HK, Pang CP (2000) Apolipoprotein E isoforms in Alzheimer's disease pathology and etiology. *Microsc Res Tech* **50**, 278-281.
 - [21] Berlau DJ, Corrada MM, Head E, Kawas CH (2009) APOE epsilon2 is associated with intact cognition but increased Alzheimer pathology in the oldest old. *Neurology* **72**, 829-834.
 - [22] Juottonen K, Lehtovirta M, Helisalmi S, Riekkinen PJ Sr., Soininen H (1998) Major decrease in the volume of the entorhinal cortex in patients with Alzheimer's disease carrying the apolipoprotein E epsilon4 allele. *J Neurol Neurosurg Psychiatry* **65**, 322-327.
 - [23] Schuff N, Woerner N, Boreta L, Kornfield T, Shaw LM, Trojanowski JQ, Thompson PM, Jack CR Jr., Weiner MW (2009) MRI of hippocampal volume loss in early Alzheimer's disease in relation to ApoE genotype and biomarkers. *Brain* **132**, 1067-1077.
 - [24] Liu Y, Paajanen T, Westman E, Zhang Y, Wahlund LO, Simmons A, Tunnard C, Sobow T, Proitsi P, Powell J, Mecocci P, Tsolaki M, Vellas B, Muehlboeck S, Evans A, Spenger C, Lovestone S, Soininen H (2010) APOE epsilon2 allele is associated with larger regional cortical thicknesses and volumes. *Dement Geriatr Cogn Disord* **30**, 229-237.
 - [25] Wolk DA, Dickerson BC (2010) Apolipoprotein E (APOE) genotype has dissociable effects on memory and attentional-executive network function in Alzheimer's disease. *Proc Natl Acad Sci U S A* **107**, 10256-10261.
 - [26] Farlow MR, He Y, Tekin S, Xu J, Lane R, Charles HC (2004) Impact of APOE in mild cognitive impairment. *Neurology* **63**, 1898-1901.
 - [27] Geroldi C, Pihlajamaki M, Laakso MP, DeCarli C, Beltramello A, Bianchetti A, Soininen H, Trabucchi M, Frisoni GB (1999) APOE-epsilon4 is associated with less frontal and more medial temporal lobe atrophy in AD. *Neurology* **53**, 1825-1832.
 - [28] Filippini N, Rao A, Wetten S, Gibson RA, Borrie M, Guzman D, Kertesz A, Loy-English I, Williams J, Nichols T, Whitcher B, Matthews PM (2009) Anatomically-distinct genetic associations of APOE epsilon4 allele load with regional cortical atrophy in Alzheimer's disease. *Neuroimage* **44**, 724-728.
 - [29] Hua X, Leow AD, Parikshak N, Lee S, Chiang MC, Toga AW, Jack CR Jr., Weiner MW, Thompson PM (2008) Tensor-based morphometry as a neuroimaging biomarker for Alzheimer's disease: an MRI study of 676 AD, MCI, and normal subjects. *Neuroimage* **43**, 458-469.
 - [30] Small GW, Siddarth P, Burggren AC, Kepe V, Ercoli LM, Miller KJ, Lavretsky H, Thompson PM, Cole GM, Huang SC, Phelps ME, Bookheimer SY, Barrio JR (2009) Influence of cognitive status, age, and APOE-4 genetic risk on brain

- FDDNP positron-emission tomography imaging in persons without dementia. *Arch Gen Psychiatry* **66**, 81-87.
- [31] Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S, Thibodeau SN, Osborne D (1996) Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. *N Engl J Med* **334**, 752-758.
- [32] Reiman EM, Chen K, Alexander GE, Caselli RJ, Bandy D, Osborne D, Saunders AM, Hardy J (2005) Correlations between apolipoprotein E epsilon4 gene dose and brain-imaging measurements of regional hypometabolism. *Proc Natl Acad Sci U S A* **102**, 8299-8302.
- [33] Thambisetty M, Beason-Held L, An Y, Kraut MA, Resnick SM (2010) APOE epsilon4 genotype and longitudinal changes in cerebral blood flow in normal aging. *Arch Neurol* **67**, 93-98.
- [34] Plassman BL, Welsh-Bohmer KA, Bigler ED, Johnson SC, Anderson CV, Helms MJ, Saunders AM, Breitner JC (1997) Apolipoprotein E epsilon 4 allele and hippocampal volume in twins with normal cognition. *Neurology* **48**, 985-989.
- [35] Mueller SG, Schuff N, Raptentsetsang S, Elman J, Weiner MW (2008) Selective effect of Apo e4 on CA3 and dentate in normal aging and Alzheimer's disease using high resolution MRI at 4 T. *Neuroimage* **42**, 42-48.
- [36] Wishart HA, Saykin AJ, McAllister TW, Rabin LA, McDonald BC, Flashman LA, Roth RM, Mamourian AC, Tsongalis GJ, Rhodes CH (2006) Regional brain atrophy in cognitively intact adults with a single APOE epsilon4 allele. *Neurology* **67**, 1221-1224.
- [37] Burggren AC, Zeineh MM, Ekstrom AD, Braskie MN, Thompson PM, Small GW, Bookheimer SY (2008) Reduced cortical thickness in hippocampal subregions among cognitively normal apolipoprotein E e4 carriers. *Neuroimage* **41**, 1177-1183.
- [38] Shaw P, Lerch JP, Pruessner JC, Taylor KN, Rose AB, Greenstein D, Clasen L, Evans A, Rapoport JL, Giedd JN (2007) Cortical morphology in children and adolescents with different apolipoprotein E gene polymorphisms: an observational study. *Lancet Neurol* **6**, 494-500.
- [39] Lemaitre H, Crivello F, Dufouil C, Grassiot B, Tzourio C, Alperovitch A, Mazoyer B (2005) No epsilon4 gene dose effect on hippocampal atrophy in a large MRI database of healthy elderly subjects. *Neuroimage* **24**, 1205-1213.
- [40] Espeseth T, Westlye LT, Fjell AM, Walhovd KB, Rootwelt H, Reinvang I (2008) Accelerated age-related cortical thinning in healthy carriers of apolipoprotein E epsilon 4. *Neurobiol Aging* **29**, 329-340.
- [41] Lind J, Larsson A, Persson J, Ingvar M, Nilsson LG, Backman L, Adolfsson R, Cruts M, Sleegers K, Van Broeckhoven C, Nyberg L (2006) Reduced hippocampal volume in non-demented carriers of the apolipoprotein E epsilon4: relation to chronological age and recognition memory. *Neurosci Lett* **396**, 23-27.
- [42] Liu Y, Pajanan T, Westman E, Wahlund LO, Simmons A, Tunnard C, Sobow T, Proitsi P, Powell J, Mecocci P, Tsolaki M, Vellas B, Muehlboeck S, Evans A, Spenger C, Lovestone S, Soininen H (2010) Effect of APOE epsilon4 allele on cortical thicknesses and volumes: the AddNeuroMed study. *J Alzheimers Dis* **21**, 947-966.
- [43] Caselli RJ, Dueck AC, Osborne D, Sabbagh MN, Connor DJ, Ahern GL, Baxter LC, Rapsak SZ, Shi J, Woodruff BK, Locke DE, Snyder CH, Alexander GE, Rademakers R, Reiman EM (2009) Longitudinal modeling of age-related memory decline and the APOE epsilon4 effect. *N Engl J Med* **361**, 255-263.
- [44] Jernigan TL, Archibald SL, Fennema-Notestine C, Gamst AC, Stout JC, Bonner J, Hesselink JR (2001) Effects of age on tissues and regions of the cerebrum and cerebellum. *Neurobiol Aging* **22**, 581-594.
- [45] Thambisetty M, Wan J, Carass A, An Y, Prince JL, Resnick SM (2010) Longitudinal changes in cortical thickness associated with normal aging. *Neuroimage* **52**, 1215-1223.
- [46] Kremen WS, Thompson-Brenner H, Leung YM, Grant MD, Franz CE, Eisen SA, Jacobson KC, Boake C, Lyons MJ (2006) Genes, environment, and time: the Vietnam Era Twin Study of Aging (VETSA). *Twin Res Hum Genet* **9**, 1009-1022.
- [47] Goldberg J, Curran B, Vitek ME, Henderson WG, Boyko EJ (2002) The Vietnam Era Twin Registry. *Twin Res* **5**, 476-481.
- [48] Centers for Disease Control and Prevention. Health data for all ages [online]. Available at: <http://www.cdc.gov/nchs/hdi.htm>. Accessed April 20, 2007
- [49] Emi M, Wu LL, Robertson MA, Myers RL, Hegele RA, Williams RR, White R, Lalouel JM (1988) Genotyping and sequence analysis of apolipoprotein E isoforms. *Genomics* **3**, 373-379.
- [50] Hixson JE, Vernier DT (1990) Restriction isotyping of human apolipoprotein E by gene amplification and cleavage with HhaI. *J Lipid Res* **31**, 545-548.
- [51] Rebeck GW, Perls TT, West HL, Sodhi P, Lipsitz LA, Hyman BT (1994) Reduced apolipoprotein epsilon 4 allele frequency in the oldest old Alzheimer's patients and cognitively normal individuals. *Neurology* **44**, 1513-1516.
- [52] Lyons MJ, York TP, Franz CE, Grant MD, Eaves LJ, Jacobson KC, Schaie KW, Panizzon MS, Boake C, Xian H, Toomey R, Eisen SA, Kremen WS (2009) Genes determine stability and the environment determines change in cognitive ability during 35 years of adulthood. *Psychol Sci* **20**, 1146-1152.
- [53] Kremen WS, Prom-Wormley E, Panizzon MS, Eyster LT, Fischl B, Neale MC, Franz CE, Lyons MJ, Pacheco J, Perry ME, Stevens A, Schmitt JE, Grant MD, Seidman LJ, Thermenos HW, Tsuang MT, Eisen SA, Dale AM, Fennema-Notestine C (2010) Genetic and environmental influences on the size of specific brain regions in midlife: the VETSA MRI study. *Neuroimage* **49**, 1213-1223.
- [54] Fischl B, Salat DH, Busa E, Albert M, Dieterich M, Haselgrove C, van der Kouwe A, Killiany R, Kennedy D, Klaveness S, Montillo A, Makris N, Rosen B, Dale AM (2002) Whole brain segmentation: automated labeling of neuroanatomical structures in the human brain. *Neuron* **33**, 341-355.
- [55] Dale AM, Fischl B, Sereno MI (1999) Cortical surface-based analysis. I. Segmentation and surface reconstruction. *Neuroimage* **9**, 179-194.
- [56] Fischl B, Sereno MI, Dale AM (1999) Cortical surface-based analysis. II: Inflation, flattening, and a surface-based coordinate system. *Neuroimage* **9**, 195-207.
- [57] Fischl B, Van Der Kouwe A, Destrieux C, Halgren E, Segonne F, Salat DH, Busa E, Seidman LJ, Goldstein J, Kennedy D, Caviness V, Makris N, Rosen B, Dale AM (2004) Automatically parcellating the human cerebral cortex. *Cereb Cortex* **14**, 11-22.
- [58] Buckner RL, Head D, Parker J, Fotenos AF, Marcus D, Morris JC, Snyder AZ (2004) A unified approach for morphometric and functional data analysis in young, old, and demented adults using automated atlas-based head size normalization: reliability and validation against manual measurement of total intracranial volume. *Neuroimage* **23**, 724-738.

- [59] Fennema-Notestine C, Gamst AC, Quinn BT, Pacheco J, Jernigan TL, Thal L, Buckner R, Killiany R, Blacker D, Dale AM, Fischl B, Dickerson B, Gollub RL (2007) Feasibility of multi-site clinical structural neuroimaging studies of aging using legacy data. *Neuroinformatics* **5**, 235-245.
- [60] Desikan RS, Segonne F, Fischl B, Quinn BT, Dickerson BC, Blacker D, Buckner RL, Dale AM, Maguire RP, Hyman BT, Albert MS, Killiany RJ (2006) An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *Neuroimage* **31**, 968-980.
- [61] Barnes J, Ridgway GR, Bartlett J, Henley SM, Lehmann M, Hobbs N, Clarkson MJ, MacManus DG, Ourselin S, Fox NC (2010) Head size, age and gender adjustment in MRI studies: a necessary nuisance? *Neuroimage* **53**, 1244-1255.
- [62] Han X, Jovicich J, Salat D, van der Kouwe A, Quinn B, Czanner S, Busa E, Pacheco J, Albert M, Killiany R, Maguire P, Rosas D, Makris N, Dale A, Dickerson B, Fischl B (2006) Reliability of MRI-derived measurements of human cerebral cortical thickness: The effects of field strength, scanner upgrade and manufacturer. *Neuroimage*
- [63] Stonnington CM, Tan G, Kloppel S, Chu C, Draganski B, Jack CR Jr., Chen K, Ashburner J, Frackowiak RS (2008) Interpreting scan data acquired from multiple scanners: a study with Alzheimer's disease. *Neuroimage* **39**, 1180-1185.
- [64] Dumanis SB, Tesoriero JA, Babus LW, Nguyen MT, Trotter JH, Ladu MJ, Weeber EJ, Turner RS, Xu B, Rebeck GW, Hoe HS (2009) ApoE4 decreases spine density and dendritic complexity in cortical neurons in vivo. *J Neurosci* **29**, 15317-15322.
- [65] Agosta F, Vessel KA, Miller BL, Migliaccio R, Bonasera SJ, Filippi M, Boxer AL, Karydas A, Possin KL, Gorno-Tempini ML (2009) Apolipoprotein E epsilon4 is associated with disease-specific effects on brain atrophy in Alzheimer's disease and frontotemporal dementia. *Proc Natl Acad Sci U S A* **106**, 2018-2022.
- [66] Cherbuin N, Leach LS, Christensen H, Anstey KJ (2007) Neuroimaging and APOE genotype: a systematic qualitative review. *Dement Geriatr Cogn Disord* **24**, 348-362.
- [67] Hayden KM, Zandi PP, West NA, Tschanz JT, Norton MC, Corcoran C, Breitner JC, Welsh-Bohmer KA (2009) Effects of family history and apolipoprotein E epsilon4 status on cognitive decline in the absence of Alzheimer dementia: the Cache County Study. *Arch Neurol* **66**, 1378-1383.
- [68] Rao VS, Cupples A, van Duijn CM, Kurz A, Green RC, Chui H, Duara R, Auerbach SA, Volicer L, Wells J, van Broeckhoven C, Growdon JH, Haines JL, Farrer LA (1996) Evidence for major gene inheritance of Alzheimer disease in families of patients with and without apolipoprotein E epsilon 4. *Am J Hum Genet* **59**, 664-675.
- [69] Fleisher A, Grundman M, Jack CR Jr., Petersen RC, Taylor C, Kim HT, Schiller DH, Bagwell V, Sencakova D, Weiner MF, DeCarli C, DeKosky ST, van Dyck CH, Thal LJ (2005) Sex, apolipoprotein E epsilon 4 status, and hippocampal volume in mild cognitive impairment. *Arch Neurol* **62**, 953-957.
- [70] Panizzon MS, Hauger R, Dale AM, Eaves LJ, Eyler LT, Fischl B, Fennema-Notestine C, Franz CE, Grant MD, Jak AJ, Jacobson KC, Lyons MJ, Mendoza SP, Neale MC, Prom-Wormley EC, Seidman LJ, Tsuang MT, Xian H, Kremen WS (2010) Testosterone modifies the effect of APOE genotype on hippocampal volume in middle-aged men. *Neurology* **75**, 874-880.
- [71] Gerritsen L, Comijs HC, Deeg DJ, Penninx BW, Geerlings MI Salivary cortisol, APOE-varepsilon4 allele and cognitive decline in a prospective study of older persons. *Neurobiol Aging* (In Press)
- [72] Kessler RC, Sonnega A, Bromet E, Hughes M, Nelson CB (1995) Posttraumatic stress disorder in the National Comorbidity Survey. *Arch Gen Psychiatry* **52**, 1048-1060.
- [73] Gilbertson MW, Shenton ME, Ciszewski A, Kasai K, Lasko NB, Orr SP, Pitman RK (2002) Smaller hippocampal volume predicts pathologic vulnerability to psychological trauma. *Nat Neurosci* **5**, 1242-1247.
- [74] Jernigan TL, Gamst AC (2005) Changes in volume with age-consistency and interpretation of observed effects. *Neurobiol Aging* **26**, 1271-1274; discussion 1275-1278.