# **UC Davis**

# **UC Davis Previously Published Works**

## **Title**

Draft Genome Sequence of Botrytis cinerea BcDW1, Inoculum for Noble Rot of Grape Berries

#### **Permalink**

https://escholarship.org/uc/item/698971xb

### **Journal**

Microbiology Resource Announcements, 1(3)

#### **ISSN**

2576-098X

#### **Authors**

Blanco-Ulate, Barbara Allen, Greg Powell, Ann LT et al.

#### **Publication Date**

2013-06-27

#### DOI

10.1128/genomea.00252-13

Peer reviewed



# Draft Genome Sequence of *Botrytis cinerea* BcDW1, Inoculum for Noble Rot of Grape Berries

Barbara Blanco-Ulate, a,b Greg Allen, Ann L. T. Powell, Dario Cantua

Department of Viticulture and Enology, University of California—Davis, Davis, California, USA<sup>a</sup>; Department of Plant Sciences, University of California—Davis, Davis, California, USA<sup>b</sup>; Dolce Winery, Oakville, California, USA<sup>c</sup>

Botrytized wines are produced from grape berries infected by *Botrytis cinerea* under specific environmental conditions. Here, we report the draft genome sequence of *B. cinerea* BcDW1, a strain isolated from Sémillon grapes in Napa Valley in 1992 that is used with the intent to induce noble rot for botrytized wine production.

Received 1 April 2013 Accepted 8 April 2013 Published 23 May 2013

Citation Blanco-Ulate B, Allen G, Powell ALT, Cantu D. 2013. Draft genome sequence of *Botrytis cinerea* BcDW1, inoculum for noble rot of grape berries. Genome Announc 1(3):e00252-13. doi:10.1128/genomeA.00252-13.

Copyright © 2013 Blanco-Ulate et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license. Address correspondence to Dario Cantu, dacantu@ucdavis.edu.

**Potrytis cinerea** (teleomorph, *Botryotinia fuckeliana*) is a necrotrophic plant pathogen that is particularly aggressive on ripe fruit (1). Under specific environmental conditions, infections of wine grape berries by *B. cinerea* cause noble rot, characterized by physicochemical modifications of infected berries that contribute to the unique properties of the botrytized wines (2). Changes attributed to noble rot include dehydration as result of the actions of fungal cell wall-degrading enzymes, oxidation of phenolic compounds by fungal laccases, and changes in the aroma profile caused by the breakdown of fruit esters and the synthesis of fungal volatiles (2–4).

B. cinerea isolate BcDW1 was recovered in 1992 from grape berries (Vitis vinifera cv. "Sémillon") in Napa, California, and this isolate has been used as source of inoculum to induce noble rot in the production of Dolce Wine (Oakville, CA). DNA from an axenic BcDW1 culture was extracted using a modified cetyltrimethylammonium bromide (CTAB) method (5), and 6.9 Gb of Illumina HiSeq 2000 sequence reads was generated to achieve > 100 $\times$ coverage of the genome. After quality trimming (Q > 30), 99.78% of the reads were assembled into a nuclear genome composed of 453 scaffolds and a total length of 42.1 Mb (N<sub>50</sub>, 194 kb; L<sub>50</sub>, 69; gaps, 58 kb; G+C, 42%; median coverage, 125.3 $\times$ ). The complete mitochondrial genome was assembled into a single contig of 84 kb. De novo assembly was done using CLC Genomic Workbench v6.0 with the parameters optimized to achieve the highest completeness of the gene space estimated using Core Eukaryotic Genes Mapping Approach (CEGMA) (6). The BcDW1 genome was estimated to be >98% complete by mapping 248 low-copy core eukaryotic genes conserved across eukaryotes (6). Fiftythree million BcDW1 reads were uniquely aligned to the B05.10 (7) and T4 (8) scaffolds using Novoalign (v2.08.02, Novocraft), and 162,882 (4.0 single-nucleotide polymorphisms [SNPs]/kb) and 162,464 (3.9 SNPs/kb) SNP variants were identified by comparisons to the B05.10 and T4 genomes, respectively (Freebayes v0.9.9 [9]). A total of 5,620 nonsynonymous substitutions and 46 early stop codons were identified.

After masking repeats with RepeatMasker (10), gene models

were identified with Augustus (11) using B05.10 transcripts (7), which were training sets for ab initio gene prediction and for evidence-based gene finding. As result, 11,073 complete gene models were identified. We found a large set of candidate secreted proteins (SignalP v4.0 [12]) that are involved in plant tissue penetration and decomposition, including 165 glycoside hydrolases, 44 carbohydrate esterases, and 10 polysaccharide lyases (CAZy database [13]). The most abundant CAZy families identified among these secreted proteins were 19 polygalacturonases (GH28), 15 xyloglucanases (GH16), 10 cutinases (CE5), and 9 pectin/pectate lyases (PL1 and PL3). We also detected other secreted proteins that we predict are relevant for noble rot, such as 3 laccases (2) and 9 carboxylesterases (14). The BcDW1 draft genome sequence will be useful for comparative studies as the genome sequences of more B. cinerea isolates become available, and it will contribute to elucidating the genetic bases of host specialization and the commercially relevant roles of B. cinerea.

**Nucleotide sequence accession numbers.** This Whole-Genome Shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. AORW000000000. The version described in this paper is the first version, accession no. AORW01000000.

#### **ACKNOWLEDGMENTS**

This work was partially supported by funding to D.C. from the College of Agricultural and Environmental Sciences (University of California—Davis) and to A.L.T.P. from the NSF (IOS0957264). Support to B.B.U. was provided by the Consejo Nacional de Ciencia y Tecnología (Ministerio de Ciencia y Tecnología, Costa Rica).

We thank Henriette O'Geen (UC Davis Genome Center) and Abraham Morales for technical assistance.

#### **REFERENCES**

- Cantu D, Blanco-Ulate B, Yang L, Labavitch JM, Bennett AB, Powell AL. 2009. Ripening-regulated susceptibility of tomato fruit to *Botrytis cinerea* requires NOR but not RIN or ethylene. Plant Physiol. 150: 1434–1449.
- Magyar I. 2011. Botrytized wines, p 147–206. In Ronald SJ (ed), Advances in food and nutrition research, vol 63. Academic Press, San Diego, CA.

- 3. Sarrazin E, Dubourdieu D, Darriet P. 2007. Characterization of keyaroma compounds of botrytized wines, influence of grape botrytization. Food Chem. 103:536–545.
- Schouten A, Wagemakers L, Stefanato FL, van der Kaaij RM, van Kan JA. 2002. Resveratrol acts as a natural profungicide and induces selfintoxication by a specific laccase. Mol. Microbiol. 43:883–894.
- Möller EM, Bahnweg G, Sandermann H, Geiger HH. 1992. A simple and efficient protocol for isolation of high molecular weight DNA from filamentous fungi, fruit bodies, and infected plant tissues. Nucleic Acids Res. 20:6115–6116.
- Parra G, Bradnam K, Ning Z, Keane T, Korf I. 2009. Assessing the gene space in draft genomes. Nucleic Acids Res. 37:289–297.
- 7. Staats M, van Kan JA. 2012. Genome update of *Botrytis cinerea* strains B05.10 and T4. Eukaryot. Cell 11:1413–1414.
- 8. Amselem J, Cuomo CA, van Kan JA, Viaud M, Benito EP, Couloux A, Coutinho PM, de Vries RP, Dyer PS, Fillinger S, Fournier E, Gout L, Hahn M, Kohn L, Lapalu N, Plummer KM, Pradier JM, Quévillon E, Sharon A, Simon A, ten Have A, Tudzynski B, Tudzynski P, Wincker P, Andrew M, Anthouard V, Beever RE, Beffa R, Benoit I, Bouzid O, Brault B, Chen Z, Choquer M, Collémare J, Cotton P, Danchin EG, Da Silva C, Gautier A, Giraud C, Giraud T, Gonzalez C, Grossetete S,

- Güldener U, Henrissat B, Howlett BJ, Kodira C, Kretschmer M, Lappartient A, Leroch M, Levis C, et al. 2011. Genomic analysis of the necrotrophic fungal pathogens *Sclerotinia sclerotiorum* and *Botrytis cinerea*. PLoS Genet. 7:e1002230.
- Marth GT, Korf I, Yandell MD, Yeh RT, Gu Z, Zakeri H, Stitziel NO, Hillier L, Kwok PY, Gish WR. 1999. A general approach to singlenucleotide polymorphism discovery. Nat. Genet. 23:452–456.
- 10 Smit AFA, Hubley R, Green P. 2003. RepeatMasker. http://www.repeatmasker.org.
- 11. Stanke M, Diekhans M, Baertsch R, Haussler D. 2008. Using native and syntenically mapped cDNA alignments to improve *de novo* gene finding. Bioinformatics 24:637–644.
- Petersen TN, Brunak S, von Heijne G, Nielsen H. 2011. SignalP 4.0: discriminating signal peptides from transmembrane regions. Nat. Methods 8:785–786.
- 13. Cantarel BL, Coutinho PM, Rancurel C, Bernard T, Lombard V, Henrissat B. 2009. The carbohydrate-active enzymes database (CAZy): an expert resource for glycogenomics. Nucleic Acids Res. 37:D233–D238.
- Sumby KM, Grbin PR, Jiranek V. 2010. Microbial modulation of aromatic esters in wine: current knowledge and future prospects. Food Chem. 121:1-16