UCLA

UCLA Previously Published Works

Title

Role of endoplasmic reticulum stress signalling in diabetic endothelial dysfunction and atherosclerosis

Permalink

https://escholarship.org/uc/item/6b5424tg

Journal

Diabetes and Vascular Disease Research, 14(1)

ISSN

1479-1641

Authors

Dong, Yunzhou Fernandes, Conrad Liu, Yanjun et al.

Publication Date

2017

DOI

10.1177/1479164116666762

Peer reviewed

Published in final edited form as:

Diab Vasc Dis Res. 2017 January; 14(1): 14-23. doi:10.1177/1479164116666762.

Role of endoplasmic reticulum stress signalling in diabetic endothelial dysfunction and atherosclerosis

Yunzhou Dong¹, Conrad Fernandes², Yanjun Liu³, Yong Wu³, Hao Wu¹, Megan L Brophy¹, Lin Deng⁴, Kai Song¹, Aiyun Wen¹, Scott Wong¹, Daoguang Yan⁵, Rheal Towner⁶, and Hong Chen¹

¹Vascular Biology Program, Department of Surgery, Boston Children's Hospital, Harvard Medical School, Boston, MA, USA

²Department of Biosciences, Rice University, Houston, TX, USA

³Department of Internal Medicine, Charles R. Drew University of Medicine and Science, University of California–Los Angeles School of Medicine, Los Angeles, CA, USA

⁴Department of Biological Chemistry & Molecular Pharmacology, Harvard Medical School, Boston, MA, USA

⁵Department of Biology, Jinan University, Guangzhou, China

⁶Advanced Magnetic Resonance Center, Oklahoma Medical Research Foundation, Oklahoma, OK, USA

Abstract

It is well established that diabetes mellitus accelerates atherosclerotic vascular disease. Endothelial injury has been proposed to be the initial event in the pathogenesis of atherosclerosis. Endothelium not only acts as a semi-selective barrier but also serves physiological and metabolic functions. Diabetes or high glucose in circulation triggers a series of intracellular responses and organ damage such as endothelial dysfunction and apoptosis. One such response is high glucose-induced chronic endoplasmic reticulum stress in the endothelium. The unfolded protein response is an acute reaction that enables cells to overcome endoplasmic reticulum stress. However, when chronically persistent, endoplasmic reticulum stress response could ultimately lead to endothelial dysfunction and atherosclerosis. Herein, we discuss the scientific advances in understanding endoplasmic reticulum stress-induced endothelial dysfunction, the pathogenesis of diabetes-accelerated atherosclerosis and endoplasmic reticulum stress as a potential target in therapies for diabetic atherosclerosis.

Keywords

Diabetes mellitus; endoplasmic reticulum stress; endothelial dysfunction; atherosclerosis

Reprints and permissions: sagepub.co.uk/journalsPermissions.nav

Corresponding author: Hong Chen, Vascular Biology Program, Department of Surgery, Boston Children's Hospital, Harvard Medical School, Boston, MA 02115, USA. hong.chen@childrens.harvard.edu.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship and/or publication of this article.

Introduction

Diabetes mellitus is a disease where high blood sugar levels occur from decreased insulin signalling, either by a lack of insulin secretion in Type I diabetes or insulin resistance in Type II diabetes.¹ Diabetes is a complex group of diseases with a variety of causes including cardiovascular risk factors, such as obesity, hypertension, hypertriglyceridimia, genetic alterations, and lifestyle factors, such as smoking. Further complications from the disease can often result in chronic kidney failure, brain damage, retinal blood vessel damage and other cardiovascular diseases.

Atherosclerosis is marked by the appearance of plaques consisting of leukocytes, lipids, smooth muscle cells and low-density lipoprotein (LDL) cholesterol in the arterial walls.⁴ When the condition worsens over time, it leads to ischemia-causing cerebrovascular accidents and myocardial infarctions in diabetic patients.⁴ Along with coronary artery disease and congestive heart failures,⁷ diabetic-induced atherosclerosis has been linked to endothelial dysfunction (ED) – a pathological imbalance between vasoconstriction and vasodilation in the endothelium.⁸ Understanding the molecular mechanisms behind diabetic atherosclerosis is therefore of great clinical significance.

Healthy endothelium serves as a moderator producing autocrine and paracrine signals⁹ to decide between vasodilation and vasoconstriction contractions in blood arteries. Nitric oxide (NO), formerly endothelium-derived relaxing factor,¹⁰ bradykinin and prostacyclin are all potent vasodilators,¹⁰ while thromboxanes,⁸ serotonin,⁸ endothelin¹⁰ and angiotensin II¹⁰ are vasoconstrictors involved in maintaining this equilibrium. In ED, a disorder of the endothelium lining the blood vessels, a decrease in bioavailability of NO^{8,11} and an increase in vasoconstrictors¹¹ disrupt the equilibrium by increasing thrombus formation,⁸ platelet aggregation,⁸ chemokine production,⁹ leukocyte adherence⁹ and smooth muscle cell proliferation;^{8,9} all of which contribute to atherosclerosis. The accumulation of cardiovascular risk factor superoxide, asymmetric dimethylarginine (ADMA, an endogenous NO inhibitor),^{7,10} angiotensin II,^{7,10} hyperhomocysteine (HHcy)^{7,12} and impaired insulin signalling reduces endothelial nitric oxide synthase (eNOS) activity and levels of NO,⁷ which are both major determinants of ED.¹³

ED has been suggested to predict the onset of atherosclerosis and is therefore considered to be an early marker for the disease. Experiments using constriction of blood vessels in response to acetylcholine to test endothelial response have shown that ED occurs before and throughout the progression of atherosclerosis. Al,15 Similarly, plaque growth from ED is also part of the mechanism for the progression of the disease. A dysfunctional endothelium promotes factors (mentioned above) such as migration of smooth muscle cells, chemokine and cytokine secretions (that promote plaque formation) and the development of cardiovascular diseases. Even in the absence of inhibitors of smooth muscle cell proliferation like heparin, a blood thinner anticoagulant, one study found that administration of NO decreases the migration of vascular smooth muscle cells.

Endoplasmic reticulum stress and its role in atherosclerosis

The rough endoplasmic reticulum (RER) is the organelle in the cell responsible for the production and correct folding of proteins destined for the endomembrane system and secretion. Pathological stimuli disrupts the endoplasmic reticulum (ER) homeostasis and causes misfolded or unfolded proteins to accumulate in the ER until properly folded or destroyed. 18 In general, when the ER is overwhelmed by the accumulation of misfolded or unfolded proteins, a condition known as ER stress, the unfolded protein response (UPR) is then activated, and the expression of chaperones is upregulated to help fold ER proteins. ¹⁹ The UPR happens through the activation of evolutionally conserved signalling pathways involving inositol-requiring protein-1 (IRE-1), activating transcription factor 6 (ATF6) and protein kinase RNA-like ER kinase (PERK), where acute-responsible chaperone proteins are synthesized.^{20,21} If not chronic, this response is adaptive and helps the cell recover to normal health.²² However, if the UPR is sustained, apoptosis is induced through a pathway involving the transcription factor C/EBPa-homologous protein (CHOP) that induces DR5 expression.²³ This sustained ER stress is implicated in many disease processes and has been linked to Type II diabetes, ²⁴ cardiovascular disease, ²⁵ Alzheimer's disease, ²⁶ as well as atherosclerosis. ^{25,27} In one study, administration of ER stress inhibitors in hypertensive mouse models improved endothelial function, indicating that ER stress may have been the cause of ED.²⁸ Later, Lenna et al.²⁹ linked ER stress to dysfunctional eNOS in endothelial cells (ECs), and Galan et al.³⁰ showed that ER stress causes ED through oxidative stress. A link between ER stress and atherosclerosis was also established in atheroma ECs and macrophages.³¹ Because ER stress is linked to ED and poor endothelium health is linked to atherosclerosis, the mechanisms that cause ER stress in diabetic conditions could be of great therapeutic potential. In this review, we will focus on the progress made in understanding molecular mechanisms involving reactive oxygen species (ROS)/reactive nitrogen species (RNS), mitochondrial dysfunction, intracellular calcium homeostasis, free fatty acids (FFAs), advanced glycation end products (AGEs), inflammation and insulin resistance. Therapeutic potentials of targeting ER stress signalling pathways will also be examined. For mechanisms not discussed in this review, such as abnormal disposal of unwanted proteins in ER, defects in autophagy, inhibition of the proteasome and flaws in ER-Golgi trafficking, readers should refer to other reviews. 32–38

ROS and RNS

High glucose in diabetes stresses the electron transport chain in which electrons are directly passed from coenzyme Q to oxygen in generating superoxides or ROS.³⁹ As reviewed earlier, ED is linked to a reduction in NO bioavailability, either caused by reduced production or increased degradation of NO. One of the mechanisms for degradation of NO comes from its reaction with ROS⁴⁰ to generate peroxynitrite – a RNS.⁴¹ This reaction occurs faster than reactions that normally remove ROS [which are catalysed by superoxide dismutase (SOD)]⁴² and produces elevated levels of ROS, a key downregulator of NO bioavailability. During chronic ER stress, the UPR generates ROS,⁴³ which suggests that ER stress is connected to ED and the decrease in NO concentration. The generation of ROS partly comes from ER oxidoreductin 1 protein (Ero1p), an oxidative thiol enzyme that produces H_2O_2 when it oxides protein disulphide isomerase (PDI) to fold proteins in the

UPR. 44 When non-chronic, the UPR actually reduces ROS by upregulating antioxidant pathways, 45 however, but only when the response is overwhelmed by ER stress. In fact, ROS can even cause ER stress, as one study showed that the UPR triggered by TNFa actually required ROS to be present. 45 This indicates that in some cases, the UPR is activated by ROS, where ROS also activates the NF- κ B pathway 46 (which will be discussed later as a mechanism for ER stress response).

In addition to ER stress causing ED from producing peroxynitrite (through the reaction of ROS with NO), peroxynitrite itself can lead to ER stress. When ECs were treated with Sin-1, a peroxynitrite generator, or with uric acid (which reacts with peroxynitrite), the expression of chaperones characteristic of ER stress were induced with the former but not with the latter, ⁴⁷ indicating that peroxynitrite (a RNS) is capable of causing ER stress. ⁴⁷ Furthermore, peroxynitrite has a more direct effect on endothelial function through decreasing production of NO from eNOS. ⁴² Oxidized LDL can then decrease eNOS activity by reducing the dimer/monomer ratio and the total amount via calpain-mediated, ROS-dependent and Ca²⁺-dependent manners. ⁴⁸ Although NO is important for endothelial health as a RNS, excessive amounts of the compound can cause a disruption of ER chaperones that promote ER stress, ⁴⁹ nitrosylate Akt, and inhibit insulin signalling in contributing to diabetes, where the effect can be further amplified in the presence of ROS. ⁵⁰ In summary, both ROS and RNS can cause and be caused by ER stress, linking them to ED and atherosclerosis in diabetic patients.

The mitochondria's electron transport chain normally functions to produce adenosine triphosphate (ATP) for the cell. However, the mitochondria generate ROS as a by-product and can induce apoptosis through the activation of the caspase system by cytochrome C,⁵¹ which is normally a component of the electron transport chain. Constant leakage of Ca²⁺ through inositol 1,4,5-trisphosphate receptor (IP3R) is required to generate ATP in the mitochondria, and when this pathway is blocked, ATP production decreases substantially.⁵² During the early stages of ER stress, Ca²⁺ leakage from the ER and subsequent uptake by the mitochondria increase the rate of energy production⁵³ and help the cell in responding to ER stress. In chronic ER stress, small amount of cytochrome C is solubilized (partially by ROS oxidation of cardiolipin⁵⁴) and leak from mitochondrial pores through apoptotic stimuli.⁵⁴ The small amount of cytochrome C binds to IP3R and causes a release of Ca²⁺, which then activates a positive feedback loop to release more cytochrome C^{54,55} to induce apoptosis (see further discussion in 'Intracellular calcium homeostasis').

Mitochondrial dysfunction is common in diabetic patients, and it has been suggested to be a cause of insulin resistance and impaired insulin secretion⁵⁶ having multiple effects on insulin signalling. It has been well established that mitochondrial dysfunction and ROS production accelerate diabetes, and because these two conditions could also be caused by the high glucose and fatty acid states of obesity-induced diabetes,⁵⁷ it demonstrates that there is a reciprocal causality between the two. This mitochondrial dysfunction has been linked to ED in humans^{58,59} and also to ER stress through its activation of c-Jun N-terminal kinase (JNK)⁶⁰ with dependency on Ca²⁺. Win et al.⁶⁰ reported that activation of JNK by ER stress correlated with increased ROS production and inhibited mitochondrial respiration. Additionally, mitochondrial dysfunction has been shown to induce inflammation by

activating redox-sensitive inflammatory pathways (since dysfunctional mitochondria generate high levels of ROS) or by activating the inflammosome. ⁶¹ Mitochondrial dysfunction directly promotes insulin resistance, diabetes and ED by generating ROS and activating inflammatory pathways that stress the ER and induce apoptosis.

Intracellular calcium homeostasis

Intracellular calcium homeostasis is critical to the maintenance of normal physiological functions of the cell. The process is mainly modulated through plasma membrane calcium pumps and ER-resident calcium pumps and receptors. Mitochondrial Ca²⁺ uniporters, Na-Ca²⁺ and H-Ca²⁺ exchangers also play an important role in intracellular calcium homeostasis. The ER normally serves as the major Ca²⁺ reservoir to maintain intracellular Ca²⁺ homeostasis. ⁶² It accomplishes this mainly through the sarcoplasmic/endoplasmic reticulum Ca²⁺ ATPase (SERCA) and ER-resident receptors such as IP3R. SERCA utilizes ATP hydrolysis to actively transport Ca^{2+} into the ER, 63 and oxidation of SERCA at a specific thiol group of cysteine increases atherosclerosis.⁶⁴ AMP-activated kinase alpha 2 (AMPKa2)-knockout mice exhibit augmented ER stress by changing the redox status via SERCA oxidation that disrupts the intracellular calcium homeostasis. 65 On the one hand, ox-LDL (the mediator for atherosclerosis) decreases SERCA activity by oxidizing with it and disturbing intracellular calcium balance, leading to EC ER stress. 66 On the other hand, Ca²⁺ is released from the ER during certain processes by Ca²⁺ release channels such as the ryanodine receptors (RyRs), ⁶⁷ the IP3R⁶⁷ and translocon (TLC). ⁶⁸ However, Ca²⁺ also plays a role in maintaining the environment of the ER,⁶³ where the balance of Ca²⁺ in the ER is disrupted and released into the cell, causing major problems in both the mitochondria⁶⁹ and ER.⁷⁰ In the mitochondria, excessive Ca²⁺ can lead to increased production of ROS and induce ED, linking Ca²⁺ leakage from ER to vascular complications.⁶⁹ Disruption of Ca²⁺ homeostasis in the ER can further lead to ER stress and activation of the UPR, 70 providing another link between intracellular Ca²⁺ and ED. This over-release of Ca²⁺ can then induce apoptosis in the cell.⁷¹

As mentioned earlier, when there are exceeding unfolded proteins in the ER, its stress is reduced when the cell activates the UPR. One of the mechanisms the cell uses to signal ER stress involves Ca²⁺, which binds to chaperones such as calreticulin.⁷² These and other chaperones are unable to keep up with the protein load during ER stress, where they appear to signal Ca²⁺ leak channels in causing release of Ca²⁺ into the cytosol. Hammadi et al.⁶⁸ found such a relationship between TLC and the ER chaperone glucose-regulated protein 78 or BiP (GRP78). The release of Ca²⁺ then leads to further misfolding of proteins similar to when Ca²⁺ homeostasis is disrupted, resulting in the dysfunction of these chaperones and protein misfolding.⁷² When protein misfolding activates the UPR, a vicious cycle eventually ends in apoptosis where more Ca²⁺ are released.³⁵ However, with an abundance of chaperones around, they can signal the release channels to close. This is evidenced when increased concentrations of BiP closes Sec61, preventing ER Ca²⁺ leakage through the Sec61 complex by binding to Sec61a.⁷³ Loss of IP3R causes ER stress-induced apoptosis and brain damage,⁷⁴ and inhibiting SERCA activity also augments ER stress and apoptosis.⁷⁵

Calcium also interacts with other pathways of ER stress as mentioned before. Both Ca^{2+} release and ROS production were necessary to stimulate NF- κ B activation in cells under a condition of ER stress. ⁷⁶ Ca^{2+} is also released in response to treatment with FFAs, which is another way in which FFAs cause ER stress. ^{77,78} In summary, intracellular calcium disruption causes ER stress and ED.

FFAs and AGEs

Obese individuals tend to have higher levels of FFAs, which are thought to play a role in the development of insulin resistance and diabetes. While not all individuals with diabetes have elevated FFAs, many individuals with Type II diabetes often do. High concentrations of saturated FFAs are a contributing factor to ED. These saturated fatty acids appear to cause ER stress, while unsaturated fatty acids do not have the same effect. When β -cells were treated for an extended time with palmitate, a saturated fatty acid, ER stress was induced, and the UPR was activated via a Ca²⁺-dependent pathway. Another study found that palmitate-induced ER stress decreased when the lipids were packed into triglycerides via overexpression of PPAR- γ and ACSL-1, while lipases increased ER stress.

Including the ER, FFAs are packaged into phospholipids that are incorporated into the cell's membranes. RE-1 and PERK can sense the lipid composition of the ER membrane, and when the degree of saturation of the membrane's fatty acids become excessive, the UPR will be activated. This occurs even when the protein-sensing domains of IRE-1 and PERK are non-functional. Taken together, these results indicate a mechanism for the induction of ER stress through FFAs. High levels of saturated fatty acids increase the saturation of the ER membrane (which causes the UPR to activate). In obese and diabetic patients, this stress is chronic, causing apoptosis, ED and therefore atherosclerosis.

High glucose state of diabetes also causes an increase in the formation of AGEs.⁸⁶ These molecules are proteins, nucleic acids and lipids with abnormal glycosylations.⁸⁶ AGEs are linked to ED through binding with receptors for AGE (RAGEs) and activating responses that inhibit eNOS downstream.⁸⁷ Linden et al. found that patients with diabetic nephropathy have higher AGE levels correlated with higher levels of ADMA and impaired endothelial function, further linking ED to AGE.⁸⁸ Like FFAs, AGEs have been linked to and are speculated to cause ER stress in ECs⁸⁹, suggesting yet another link between ER stress and ED.

In the same study with ADMA, AGEs were found to increase ROS production, 88 contributing to ER stress and the activation of the UPR. When AGEs increase ROS production, NF- κ B is activated and TNF α is produced, causing a further increase in superoxide production. $^{46,90-92}$ Furthermore, AGEs have also been suggested to phosphorylate eNOS and reduce the production of NO. 93 Through these mechanisms, AGEs cause ED, and ER stress is implicated in this process.

Inflammation

It is clear that obesity promotes states of both chronic low-grade inflammation and insulin resistance. ⁹⁴ However, even in the absence of obesity, infusion of animals with

proinflammatory cytokines or lipids can cause insulin resistance. 94,95 Elevated levels of TNFa, 96 interleukin-6 (IL-6) 97 and IL-1 β^{97} are often seen in patients with diabetes. The adipose tissue of obese patients produced higher levels of TNFa as assessed by mRNA levels, and these patients also exhibited hyperinsulinemia, indicative of diabetes. 96 TNFa promotes ROS production 91 and inflammation. 98 Similarly, an enzyme-linked immunosorbent assay (ELISA) assay of samples from the EPIC-Potsdam study revealed that patients with elevated levels of IL-6 and IL-1 β together had an increased risk of Type II diabetes. 97 Interestingly, having IL-1 β or IL-6 levels elevated alone did not lead to a significant increase in risk, indicating that the interplay of cytokines is important for the development of diabetes. 97

In turn, the high glucose state of diabetes promotes inflammation. Cytokines (such as TNFa and IL-1 β) and receptors IL-1R or tumour necrosis factor receptor (TNFR) are produced by monocytes, ⁹⁰ often working together in a positive feedback loop in obesity-induced diabetes. Proinflammatory cytokines produced by adipose tissue recruit monocytes and other leukocytes, and these cells in turn secrete more cytokines, suggesting an interaction between adipocytes and immune cells. Additionally, the NF- κ B pathway (which activates and produces these cytokines⁹⁰) is also activated in monocytes, ⁹⁹ and the high glucose state causes the monocytes in diabetic patients to exhibit increased expression of toll-like receptor 2 (TLR2) and toll-like receptor 4 (TLR4)¹⁰⁰ (which then activates these inflammatory pathways through their recognition of heat-shock protein 60 (HSP60)¹⁰¹). These results are significant because TNFa, IL-1 and NF- κ B signalling pathways are critical to promote cardiovascular complications. ^{100,102}

The role of proinflammatory cytokines in atherosclerosis has been well established. 103-106 The mechanism for proinflammatory cytokines affecting endothelial function again involves ER stress. ER stress has been shown to induce the NF-κB pathway through its activation of IRE-1 in myocytes, ¹⁰⁷ leading to inflammatory responses. CHOP, a transcription factor during chronic stress, induces cell death and inflammatory responses after saturated FFA exposure by activating NF-κB through a pathway involving IRAK2 expression, resulting in secretion of cytokines IL-8 and TNFa. 108 The UPR has also been shown to selectively increase levels of IL-1α in hepatocytes ¹⁰⁹ and IL-1β in macrophages ¹¹⁰ even though translation is normally inhibited during the response. Interestingly, upregulation of IL-1β has been shown to be independent of the UPR, since ER stress activates the NLRP3 (nucleotidebinding domain and leucine-rich repeat containing proteins) inflammasome¹¹¹ in providing a more direct link between it and inflammation. Akerfeldt et al. 112 demonstrated that cytokine-induced β-cell death is independent of ER stress signalling in INS-1 cells. This data further suggest a complexity between ER stress-induced and proinflammatory cytokineinduced apoptosis in Type I diabetes. To summarize, chronic ER stress may upregulate the expression of proinflammatory cytokines in diabetes or obesity. High level of proinflammatory cytokines in circulation is one of the key mediators for ER stress-induced ED¹¹³ in atherosclerosis.

Insulin resistance

Type II diabetes is characterized by insensitivity to insulin that keeps blood glucose level elevated. ED has been linked to this insulin insensitivity, since normal insulin signalling is important in the production of NO and in the process of vasodilation. ¹¹⁴ In normal physiological functions, insulin signalling activates the kinase Akt (also known as protein kinase B), which phosphorylates many downstream targets with varying effects. One of these targets is eNOS, ¹¹⁵ where Akt causes an increase in the production of NO from the EC, which is important for vascular health. As a consequence of the disrupted insulin signalling, the activation of eNOS cannot occur, and the bioavailability of NO decreases, explaining a link between insulin and ED.

Extensive studies have demonstrated that insulin resistance is one of the key factors for ED in vitro and in vivo. ^{116–119} Furthermore, high level of insulin in circulation under the conditions of insulin resistance has been shown to upregulate ER stress and the UPR in humans. ¹²⁰ Conversely, ER stress is implicated as a potential disrupter of insulin signalling itself, likely through JNK-interacting protein 1 (JIP-1). ⁹⁴ As recently suggested by Zhang et al., ¹²¹ ER stress potentiates insulin resistance through PERK-mediated FOXO phosphorylation and has also been shown to induce phosphorylation of rictor, a crucial phosphorylation site on mTORC-2, which inactivates and prevents the activation of Akt, ¹²² a key molecule in insulin signalling. Taken together, insulin resistance leads to ER stress and ED. In turn, ER stress feedback augments insulin resistance in providing another link between ER stress and ED.

Interplay among different mechanisms and pathways

As discussed above, diabetic-induced ER stress mechanisms for ED and atherosclerosis are complex. These pathways not only interact with ER stress signalling but also affect each other. For example, ROS/RNS promotes ER stress, while ER stress also induces oxidative species production. ¹²³ Insulin resistance can induce ER stress, while chronic ER stress also adversely increases insulin resistance. ¹²⁴ Similarly, ER stress enhances inflammation, ¹²⁵ while chronic inflammation further promotes ER stress. ^{124,126} Because chronic ER stress in diabetic patients can lead to ED and apoptosis in atherosclerosis, ¹²⁷ reducing or controlling one or several of the pathways to restore endothelial function would be beneficial to halt diabetic atherosclerosis.

Therapeutic potential of ER stress as target in diabetic atherosclerosis

ER stress has been proposed to be a potential therapeutic target in different diseases. ¹²⁸ Indeed, targeting ER stress has shown promise. ^{27,129–132} GRP78 and CHOP have been suggested to be important targets to reduce ER stress. ^{133,134} Chemical ER chaperones have also been tested in animal models. ¹³⁵ For example, recent studies have shown that chemical chaperones 4-PBA (phenylbutyrate acid) and TUDCA (tauroursodeoxycholic acid) reduce ER stress and restore glucose homeostasis in a mouse model with Type 2 diabetes ^{135,136} and cardiovascular disease. ^{65,137} Erbay et al. ¹³⁸ have shown that ER modification from chemical chaperones in macrophages and adipocytes has therapeutic efficacy against atherosclerosis

in mouse models. We have previously reported that deletion of AMPKα2 significantly increases the level of ER stress and reduces ATPase (SERCA) activity in ECs due to thiol group oxidations and intracellular calcium imbalances. Furthermore, administration of an ER stress alleviator can significantly reverse the progression of atherosclerosis in an AMPKα2-deficient mouse model. AICAR and metformin attenuate atherosclerosis by AMPK activation and reduction in ER stress. All these results suggest that ER stress is a promising target to treat diabetic atherosclerosis.

Closing remarks

Although the link between ER stress, ED and atherosclerosis has been established, diabetic-induced atherosclerotic mechanisms remain elusive. Future studies need to address several critical questions: (1) the molecular mechanisms underlying ED and the new pathways yet to be identified, (2) the interaction between endothelium and immune cells in pathological conditions, (3) identification of the main targeting molecules in ER stress signalling, (4) the possibility of combinational therapy by targeting multiple pathways in ER stress signalling and (5) the development of efficient technologies to specifically deliver candidate drugs to the atheroma.

Acknowledgments

Funding

This work was supported in part by NIH grants R01HL-093242, R01 HL118676, R01HL-130845, P20 RR018758; Established Investigator Award and a National Scientific Development Grant from the American Heart Association (0835544N); grant W81XWH-11-1-00226 from the Department of Defense; grant HR09-116 from the Oklahoma Center for Advanced Science and Technology (OCAST) to H.C.; AHA-SDG grant 12SDG8760002 from the American Heart Association; and OCAST grants AR11-043, HR14-056 to Y.D.; NIH grant SC1DK104821 to Y.L.; AHA fellowship 15PRE21400010 and NIH fellowship 1F31HL127982-01 to M.L.B.

References

- 1. ADA. Diagnosis and classification of diabetes mellitus. Diabetes Care. 2008; 31(Suppl. 1):S55–S60. [PubMed: 18165338]
- Epstein M, Sowers JR. Diabetes mellitus and hypertension. Hypertension. 1992; 19:403–418.
 [PubMed: 1568757]
- 3. Fisher WR. Hypertriglyceridemia in diabetes. An approach to management. J Fla Med Assoc. 1991; 78:747–750. [PubMed: 1765754]
- 4. Ross R. Atherosclerosis an inflammatory disease. N Engl J Med. 1999; 340:115–126. [PubMed: 9887164]
- Will JC, Galuska DA, Ford ES, et al. Cigarette smoking and diabetes mellitus: evidence of a positive association from a large prospective cohort study. Int J Epidemiol. 2001; 30:540–546. [PubMed: 11416080]
- 6. Forbes JM, Cooper ME. Mechanisms of diabetic complications. Physiol Rev. 2013; 93:137–188. [PubMed: 23303908]
- Endemann DH, Schiffrin EL. Endothelial dysfunction. J Am Soc Nephrol. 2004; 15:1983–1992.
 [PubMed: 15284284]
- 8. Yang Z, Ming XF. Recent advances in understanding endothelial dysfunction in atherosclerosis. Clin Med Res. 2006; 4:53–65. [PubMed: 16595793]
- 9. Favero G, Paganelli C, Buffoli B, et al. Endothelium and its alterations in cardiovascular diseases: life style intervention. Biomed Res Int. 2014; 2014:801896. [PubMed: 24719887]

 Davignon J, Ganz P. Role of endothelial dysfunction in atherosclerosis. Circulation. 2004; 109:III27–III32. [PubMed: 15198963]

- 11. Mudau M, Genis A, Lochner A, et al. Endothelial dysfunction: the early predictor of atherosclerosis. Cardiovasc J Afr. 2012; 23:222–231. [PubMed: 22614668]
- 12. Cheng Z, Jiang X, Pansuria M, et al. Hyperhomocysteinemia and hyperglycemia induce and potentiate endothelial dys-function via μ -calpain activation. Diabetes. 2015; 64:947–959. [PubMed: 25352635]
- 13. Versari D, Daghini E, Virdis A, et al. Endothelial dysfunction as a target for prevention of cardiovascular disease. Diabetes Care. 2009; 32(Suppl. 2):S314–S321. [PubMed: 19875572]
- 14. Reddy KG, Nair RN, Sheehan HM, et al. Evidence that selective endothelial dysfunction may occur in the absence of angiographic or ultrasound atherosclerosis in patients with risk factors for atherosclerosis. J Am Coll Cardiol. 1994; 23:833–843. [PubMed: 8106687]
- Ludmer PL, Selwyn AP, Shook TL, et al. Paradoxical vasoconstriction induced by acetylcholine in atherosclerotic coronary arteries. N Engl J Med. 1986; 315:1046–1051. [PubMed: 3093861]
- Esper RJ, Nordaby RA, Vilarino JO, et al. Endothelial dysfunction: a comprehensive appraisal. Cardiovasc Diabetol. 2006; 5:4. [PubMed: 16504104]
- 17. Sarkar R, Meinberg EG, Stanley JC, et al. Nitric oxide reversibly inhibits the migration of cultured vascular smooth muscle cells. Circ Res. 1996; 78:225–230. [PubMed: 8575065]
- Klausner RD, Sitia R. Protein degradation in the endoplasmic reticulum. Cell. 1990; 62:611–614.
 [PubMed: 2201450]
- 19. Shamu CE, Cox JS, Walter P. The unfolded-protein-response pathway in yeast. Trends Cell Biol. 1994; 4:56–60. [PubMed: 14731868]
- 20. Zhang K, Kaufman RJ. From endoplasmic-reticulum stress to the inflammatory response. Nature. 2008; 454:455–462. [PubMed: 18650916]
- Xu C, Bailly-Maitre B, Reed JC. Endoplasmic reticulum stress: cell life and death decisions. J Clin Invest. 2005; 115:2656–2664. [PubMed: 16200199]
- 22. Hotamisligil GS. Endoplasmic reticulum stress and atherosclerosis. Nat Med. 2010; 16:396–399. [PubMed: 20376052]
- 23. Yamaguchi H, Wang HG. CHOP is involved in endoplasmic reticulum stress-induced apoptosis by enhancing dr5 expression in human carcinoma cells. J Biol Chem. 2004; 279:45495–45502. [PubMed: 15322075]
- 24. Ozcan U, Cao Q, Yilmaz E, et al. Endoplasmic reticulum stress links obesity, insulin action, and type 2 diabetes. Science. 2004; 306:457–461. [PubMed: 15486293]
- 25. Civelek M, Manduchi E, Riley RJ, et al. Chronic endoplasmic reticulum stress activates unfolded protein response in arterial endothelium in regions of susceptibility to atherosclerosis. Circ Res. 2009; 105:453–461. [PubMed: 19661457]
- 26. Lindholm D, Wootz H, Korhonen L. ER stress and neurodegenerative diseases. Cell Death Differ. 2006; 13:385–392. [PubMed: 16397584]
- Chistiakov DA, Sobenin IA, Orekhov AN, et al. Role of endoplasmic reticulum stress in atherosclerosis and diabetic macrovascular complications. Biomed Res Int. 2014; 2014:610140. [PubMed: 25061609]
- 28. Kassan M, Galan M, Partyka M, et al. Endoplasmic reticulum stress is involved in cardiac damage and vascular endothelial dysfunction in hypertensive mice. Arterioscler Thromb Vasc Biol. 2012; 32:1652–1661. [PubMed: 22539597]
- Lenna S, Townsend DM, Tan FK, et al. Hla-b35 upregulates endothelin-1 and downregulates endothelial nitric oxide synthase via endoplasmic reticulum stress response in endothelial cells. J Immunol. 2010; 184:4654–4661. [PubMed: 20335527]
- 30. Galan M, Kassan M, Kadowitz PJ, et al. Mechanism of endoplasmic reticulum stress-induced vascular endothelial dysfunction. Biochim Biophys Acta. 2014; 1843:1063–1075. [PubMed: 24576409]
- 31. Tabas I. The role of endoplasmic reticulum stress in the progression of atherosclerosis. Circ Res. 2010; 107:839–850. [PubMed: 20884885]

32. Chambers JE, Marciniak SJ. Cellular mechanisms of endoplasmic reticulum stress signaling in health and disease. 2. Protein misfolding and ER stress. Am J Physiol Cell Physiol. 2014; 307:C657–C670. [PubMed: 24944205]

- 33. Senft D, Ronai ZA. UPR, autophagy, and mitochondria crosstalk underlies the ER stress response. Trends Biochem Sci. 2015; 40:141–148. [PubMed: 25656104]
- 34. Rashid HO, Yadav RK, Kim HR, et al. ER stress: autophagy induction, inhibition and selection. Autophagy. 2015; 11:1956–1977. [PubMed: 26389781]
- 35. Hoyer-Hansen M, Jaattela M. Connecting endoplasmic reticulum stress to autophagy by unfolded protein response and calcium. Cell Death Differ. 2007; 14:1576–1582. [PubMed: 17612585]
- 36. Benbrook DM, Long A. Integration of autophagy, proteasomal degradation, unfolded protein response and apoptosis. Exp Oncol. 2012; 34:286–297. [PubMed: 23070014]
- 37. Taxis C, Vogel F, Wolf DH. ER-Golgi traffic is a prerequisite for efficient ER degradation. Mol Biol Cell. 2002; 13:1806–1818. [PubMed: 12058050]
- 38. Guerriero CJ, Brodsky JL. The delicate balance between secreted protein folding and endoplasmic reticulum-associated degradation in human physiology. Physiol Rev. 2012; 92:537–576. [PubMed: 22535891]
- 39. Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circ Res. 2010; 107:1058–1070. [PubMed: 21030723]
- 40. Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. Circ Res. 2000; 87:840–844. [PubMed: 11073878]
- 41. Beckman JS, Koppenol WH. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. Am J Physiol. 1996; 271:C1424–C1437. [PubMed: 8944624]
- 42. Xu J, Zou MH. Molecular insights and therapeutic targets for diabetic endothelial dysfunction. Circulation. 2009; 120:1266–1286. [PubMed: 19786641]
- 43. Santos CX, Tanaka LY, Wosniak J, et al. Mechanisms and implications of reactive oxygen species generation during the unfolded protein response: roles of endoplasmic reticulum oxidoreductases, mitochondrial electron transport, and NADPH oxidase. Antioxid Redox Signal. 2009; 11:2409– 2427. [PubMed: 19388824]
- 44. Gross E, Sevier CS, Heldman N, et al. Generating disulfides enzymatically: reaction products and electron acceptors of the endoplasmic reticulum thiol oxidase Ero1p. Proc Natl Acad Sci U S A. 2006; 103:299–304. [PubMed: 16407158]
- 45. Xue X, Piao JH, Nakajima A, et al. Tumor necrosis factor alpha (TNFalpha) induces the unfolded protein response (UPR) in a reactive oxygen species (ROS)-dependent fashion, and the UPR counteracts ROS accumulation by TNFalpha. J Biol Chem. 2005; 280:33917–33925. [PubMed: 16107336]
- 46. Ungvari Z, Orosz Z, Labinskyy N, et al. Increased mitochondrial H2O2 production promotes endothelial NF-kappaB activation in aged rat arteries. Am J Physiol Heart Circ Physiol. 2007; 293:H37–H47. [PubMed: 17416599]
- 47. Dickhout JG, Hossain GS, Pozza LM, et al. Peroxynitrite causes endoplasmic reticulum stress and apoptosis in human vascular endothelium: implications in athero-genesis. Arterioscler Thromb Vasc Biol. 2005; 25:2623–2629. [PubMed: 16210571]
- 48. Dong Y, Wu Y, Wu M, et al. Activation of protease cal-pain by oxidized and glycated LDL increases the degradation of endothelial nitric oxide synthase. J Cell Mol Med. 2009; 13:2899–2910. [PubMed: 18624772]
- 49. Uehara T, Nakamura T, Yao D, et al. S-nitrosylated protein-disulphide isomerase links protein misfolding to neurodegeneration. Nature. 2006; 441:513–517. [PubMed: 16724068]
- 50. Yasukawa T, Tokunaga E, Ota H, et al. S-nitrosylation-dependent inactivation of Akt/protein kinase B in insulin resistance. J Biol Chem. 2005; 280:7511–7518. [PubMed: 15632167]
- Green DR, Reed JC. Mitochondria and apoptosis. Science. 1998; 281:1309–1312. [PubMed: 9721092]
- 52. Bravo R, Gutierrez T, Paredes F, et al. Endoplasmic reticulum: ER stress regulates mitochondrial bioenergetics. Int J Biochem Cell Biol. 2012; 44:16–20. [PubMed: 22064245]

53. Bravo R, Vicencio JM, Parra V, et al. Increased ER-mitochondrial coupling promotes mitochondrial respiration and bioenergetics during early phases of ER stress. J Cell Sci. 2011; 124:2143–2152. [PubMed: 21628424]

- 54. Ow YP, Green DR, Hao Z, et al. Cytochrome c: functions beyond respiration. Nat Rev Mol Cell Biol. 2008; 9:532–542. [PubMed: 18568041]
- 55. Boehning D, Patterson RL, Sedaghat L, et al. Cytochrome c binds to inositol (1,4,5) trisphosphate receptors, amplifying calcium-dependent apoptosis. Nat Cell Biol. 2003; 5:1051–1061. [PubMed: 14608362]
- Lowell BB, Shulman GI. Mitochondrial dysfunction and type 2 diabetes. Science. 2005; 307:384–387. [PubMed: 15662004]
- 57. Gao CL, Zhu C, Zhao YP, et al. Mitochondrial dysfunction is induced by high levels of glucose and free fatty acids in 3T3-L1 adipocytes. Mol Cell Endocrinol. 2010; 320:25–33. [PubMed: 20144685]
- 58. Kizhakekuttu TJ, Wang J, Dharmashankar K, et al. Adverse alterations in mitochondrial function contribute to type 2 diabetes mellitus-related endothelial dysfunction in humans. Arterioscler Thromb Vasc Biol. 2012; 32:2531–2539. [PubMed: 22879582]
- Shenouda SM, Widlansky ME, Chen K, et al. Altered mitochondrial dynamics contributes to endothelial dys-function in diabetes mellitus. Circulation. 2011; 124:444–453. [PubMed: 21747057]
- 60. Win S, Than TA, Fernandez-Checa JC, et al. JNK interaction with Sab mediates ER stress induced inhibition of mitochondrial respiration and cell death. Cell Death Dis. 2014; 5:e989. [PubMed: 24407242]
- 61. Lopez-Armada MJ, Riveiro-Naveira RR, Vaamonde-Garcia C, et al. Mitochondrial dysfunction and the inflammatory response. Mitochondrion. 2013; 13:106–118. [PubMed: 23333405]
- 62. Verkhratsky A, Toescu EC. Endoplasmic reticulum Ca(2+) homeostasis and neuronal death. J Cell Mol Med. 2003; 7:351–361. [PubMed: 14754504]
- 63. Wuytack F, Raeymaekers L, Missiaen L. Molecular physiology of the SERCA and SPCA pumps. Cell Calcium. 2002; 32:279–305. [PubMed: 12543090]
- 64. Adachi T, Weisbrod RM, Pimentel DR, et al. S-glutathiolation by peroxynitrite activates SERCA during arterial relaxation by nitric oxide. Nat Med. 2004; 10:1200–1207. [PubMed: 15489859]
- 65. Dong Y, Zhang M, Liang B, et al. Reduction of AMP-activated protein kinase alpha2 increases endoplasmic reticulum stress and atherosclerosis in vivo. Circulation. 2010; 121:792–803. [PubMed: 20124121]
- 66. Dong Y, Zhang M, Wang S, et al. Activation of AMP-activated protein kinase inhibits oxidized LDL-triggered endoplasmic reticulum stress in vivo. Diabetes. 2010; 59:1386–1396. [PubMed: 20299472]
- 67. Marks AR. Calcium channels expressed in vascular smooth muscle. Circulation. 1992; 86:III61–III67. [PubMed: 1330365]
- 68. Hammadi M, Oulidi A, Gackiere F, et al. Modulation of ER stress and apoptosis by endoplasmic reticulum calcium leak via translocon during unfolded protein response: involvement of GRP78. FASEB J. 2013; 27:1600–1609. [PubMed: 23322163]
- 69. Hong Q, Qi K, Feng Z, et al. Hyperuricemia induces endothelial dysfunction via mitochondrial Na +/Ca2+ exchanger-mediated mitochondrial calcium overload. Cell Calcium. 2012; 51:402–410. [PubMed: 22361139]
- 70. Kaufman RJ. Orchestrating the unfolded protein response in health and disease. J Clin Invest. 2002; 110:1389–1398. [PubMed: 12438434]
- 71. Orrenius S, Zhivotovsky B, Nicotera P. Regulation of cell death: the calcium-apoptosis link. Nat Rev Mol Cell Biol. 2003; 4:552–565. [PubMed: 12838338]
- 72. Gelebart P, Opas M, Michalak M. Calreticulin, a Ca2+-binding chaperone of the endoplasmic reticulum. Int J Biochem Cell Biol. 2005; 37:260–266. [PubMed: 15474971]
- 73. Schauble N, Lang S, Jung M, et al. BiP-mediated closing of the Sec61 channel limits Ca2+ leakage from the ER. Embo J. 2012; 31:3282–3296. [PubMed: 22796945]
- 74. Higo T, Hamada K, Hisatsune C, et al. Mechanism of ER stress-induced brain damage by IP(3) receptor. Neuron. 2010; 68:865–878. [PubMed: 21145001]

75. Luciani DS, Gwiazda KS, Yang TL, et al. Roles of IP3R and RyR Ca2+ channels in endoplasmic reticulum stress and beta-cell death. Diabetes. 2009; 58:422–432. [PubMed: 19033399]

- 76. Pahl HL, Baeuerle PA. Activation of NF-kappa B by ER stress requires both Ca2+ and reactive oxygen intermediates as messengers. FEBS Lett. 1996; 392:129–136. [PubMed: 8772190]
- 77. Cui W, Ma J, Wang X, et al. Free fatty acid induces endoplasmic reticulum stress and apoptosis of β -cells by Ca²⁺/calpain-2 pathways. PLoS ONE. 2013; 8:e59921. [PubMed: 23527285]
- 78. Egnatchik RA, Leamy AK, Jacobson DA, et al. ER calcium release promotes mitochondrial dysfunction and hepatic cell lipotoxicity in response to palmitate overload. Mol Metab. 2014; 3:544–553. [PubMed: 25061559]
- Boden G, Shulman GI. Free fatty acids in obesity and type 2 diabetes: defining their role in the development of insulin resistance and beta-cell dysfunction. Eur J Clin Invest. 2002; 32(Suppl. 3): 14–23. [PubMed: 12028371]
- 80. Li H, Bao Y, Zhang X, et al. Free fatty acids induce endothelial dysfunction and activate protein kinase C and nuclear factor-κB pathway in rat aorta. Int J Cardiol. 2011; 152:218–224. [PubMed: 20692055]
- 81. Wei Y, Wang D, Topczewski F, et al. Saturated fatty acids induce endoplasmic reticulum stress and apoptosis independently of ceramide in liver cells. Am J Physiol Endocrinol Metab. 2006; 291:E275–E281. [PubMed: 16492686]
- Salvado L, Coll T, Gomez-Foix AM, et al. Oleate prevents saturated-fatty-acid-induced ER stress, inflammation and insulin resistance in skeletal muscle cells through an AMPK-dependent mechanism. Diabetologia. 2013; 56:1372–1382. [PubMed: 23460021]
- 83. Bosma M, Dapito DH, Drosatos-Tampakaki Z, et al. Sequestration of fatty acids in triglycerides prevents endoplasmic reticulum stress in an in vitro model of cardiomyocyte lipotoxicity. Biochim Biophys Acta. 2014; 1841:1648–1655. [PubMed: 25251292]
- 84. Leamy AK, Egnatchik RA, Young JD. Molecular mechanisms and the role of saturated fatty acids in the progression of non-alcoholic fatty liver disease. Prog Lipid Res. 2013; 52:165–174. [PubMed: 23178552]
- 85. Volmer R, Van der Ploeg K, Ron D. Membrane lipid saturation activates endoplasmic reticulum unfolded protein response transducers through their transmembrane domains. Proc Natl Acad Sci U S A. 2013; 110:4628–4633. [PubMed: 23487760]
- 86. Bierhaus A, Hofmann MA, Ziegler R, et al. AGEs and their interaction with AGE-receptors in vascular disease and diabetes mellitus. I. The AGE concept. Cardiovasc Res. 1998; 37:586–600. [PubMed: 9659442]
- 87. Linden E, Cai W, He JC, et al. Endothelial dysfunction in patients with chronic kidney disease results from advanced glycation end products (AGE)-mediated inhibition of endothelial nitric oxide synthase through rage activation. Clin J Am Soc Nephrol. 2008; 3:691–698. [PubMed: 18256374]
- 88. Ando R, Ueda S, Yamagishi S, et al. Involvement of advanced glycation end product-induced asymmetric dimethylarginine generation in endothelial dysfunction. Diab Vasc Dis Res. 2013; 10:436–441. [PubMed: 23766377]
- 89. Adamopoulos C, Farmaki E, Spilioti E, et al. Advanced glycation end-products induce endoplasmic reticulum stress in human aortic endothelial cells. Clin Chem Lab Med. 2014; 52:151–160. [PubMed: 23454718]
- 90. Shanmugam N, Reddy MA, Guha M, et al. High glucose-induced expression of proinflammatory cytokine and chemokine genes in monocytic cells. Diabetes. 2003; 52:1256–1264. [PubMed: 12716761]
- 91. Kim JJ, Lee SB, Park JK, et al. TNF-alpha-induced ROS production triggering apoptosis is directly linked to Romo1 and Bcl-X(L). Cell Death Differ. 2010; 17:1420–1434. [PubMed: 20203691]
- Roberge S, Roussel J, Andersson DC, et al. TNF-α-mediated caspase-8 activation induces ROS production and TRPM2 activation in adult ventricular myocytes. Cardiovasc Res. 2014; 103:90– 99. [PubMed: 24802330]
- 93. Xu B, Chibber R, Ruggiero D, et al. Impairment of vascular endothelial nitric oxide synthase activity by advanced glycation end products. FASEB J. 2003; 17:1289–1291. [PubMed: 12738813]

94. Wellen KE, Hotamisligil GS. Inflammation, stress, and diabetes. J Clin Invest. 2005; 115:1111–1119. [PubMed: 15864338]

- 95. Yu C, Chen Y, Cline GW, et al. Mechanism by which fatty acids inhibit insulin activation of insulin receptor substrate-1 (IRS-1)-associated phosphatidylinositol 3-kinase activity in muscle. J Biol Chem. 2002; 277:50230–50236. [PubMed: 12006582]
- 96. Hotamisligil GS, Arner P, Caro JF, et al. Increased adi-pose tissue expression of tumor necrosis factor-alpha in human obesity and insulin resistance. J Clin Invest. 1995; 95:2409–2415. [PubMed: 7738205]
- 97. Spranger J, Kroke A, Mohlig M, et al. Inflammatory cytokines and the risk to develop type 2 diabetes: results of the prospective population-based European Prospective Investigation into Cancer and Nutrition (EPIC)-Potsdam Study. Diabetes. 2003; 52:812–817. [PubMed: 12606524]
- 98. Popa C, Netea MG, van Riel PL, et al. The role of TNF-alpha in chronic inflammatory conditions, intermediary metabolism, and cardiovascular risk. J Lipid Res. 2007; 48:751–762. [PubMed: 17202130]
- 99. Blackwell TS, Christman JW. The role of nuclear factor-kappa B in cytokine gene regulation. Am J Respir Cell Mol Biol. 1997; 17:3–9. [PubMed: 9224203]
- 100. Dasu MR, Devaraj S, Park S, et al. Increased toll-like receptor (TLR) activation and TLR ligands in recently diagnosed type 2 diabetic subjects. Diabetes Care. 2010; 33:861–868. [PubMed: 20067962]
- 101. Vabulas RM, Ahmad-Nejad P, da Costa C, et al. Endocytosed HSP60s use toll-like receptor 2 (TLR2) and TLR4 to activate the toll/interleukin-1 receptor signaling pathway in innate immune cells. J Biol Chem. 2001; 276:31332–31339. [PubMed: 11402040]
- 102. Bhagat K, Vallance P. Inflammatory cytokines impair endothelium-dependent dilatation in human veins in vivo. Circulation. 1997; 96:3042–3047. [PubMed: 9386173]
- 103. Ait-Oufella H, Taleb S, Mallat Z, et al. Recent advances on the role of cytokines in atherosclerosis. Arterioscler Thromb Vasc Biol. 2011; 31:969–979. [PubMed: 21508343]
- 104. Kleemann R, Zadelaar S, Kooistra T. Cytokines and atherosclerosis: a comprehensive review of studies in mice. Cardiovasc Res. 2008; 79:360–376. [PubMed: 18487233]
- 105. Ramji DP, Davies TS. Cytokines in atherosclerosis: key players in all stages of disease and promising therapeutic targets. Cytokine Growth Factor Rev. 2015; 26:673–685. [PubMed: 26005197]
- 106. Tedgui A, Mallat Z. Cytokines in atherosclerosis: pathogenic and regulatory pathways. Physiol Rev. 2006; 86:515–581. [PubMed: 16601268]
- 107. Zha X, Yue Y, Dong N, et al. Endoplasmic reticulum stress aggravates viral myocarditis by raising inflammation through the IRE1-associated NF-κB pathway. Can J Cardiol. 2015; 31:1032–1040. [PubMed: 26111668]
- 108. Willy JA, Young SK, Stevens JL, et al. CHOP links endoplasmic reticulum stress to NF-κB activation in the pathogenesis of nonalcoholic steatohepatitis. Mol Biol Cell. 2015; 26:2190–2204. [PubMed: 25904325]
- 109. Kandel-Kfir M, Almog T, Shaish A, et al. Interleukin-1α deficiency attenuates endoplasmic reticulum stress-induced liver damage and CHOP expression in mice. J Hepatol. 2015; 63:926– 933. [PubMed: 26022690]
- 110. Shenderov K, Riteau N, Yip R, et al. Cutting edge: endoplasmic reticulum stress licenses macrophages to produce mature IL-1 β in response to TLR4 stimulation through a caspase-8- and TRIF-dependent pathway. J Immunol. 2014; 192:2029–2033. [PubMed: 24489101]
- 111. Menu P, Mayor A, Zhou R, et al. ER stress activates the NLRP3 inflammasome via an UPR-independent pathway. Cell Death Dis. 2012; 3:e261. [PubMed: 22278288]
- 112. Akerfeldt MC, Howes J, Chan JY, et al. Cytokine-induced beta-cell death is independent of endoplasmic reticulum stress signaling. Diabetes. 2008; 57:3034–3044. [PubMed: 18591394]
- 113. Zhang C. The role of inflammatory cytokines in endothelial dysfunction. Basic Res Cardiol. 2008; 103:398–406. [PubMed: 18600364]
- 114. Kim JA, Montagnani M, Koh KK, et al. Reciprocal relationships between insulin resistance and endothelial dys-function: molecular and pathophysiological mechanisms. Circulation. 2006; 113:1888–1904. [PubMed: 16618833]

115. Dimmeler S, Fleming I, Fisslthaler B, et al. Activation of nitric oxide synthase in endothelial cells by Akt-dependent phosphorylation. Nature. 1999; 399:601–605. [PubMed: 10376603]

- 116. Cersosimo E, DeFronzo RA. Insulin resistance and endothelial dysfunction: the road map to cardiovascular diseases. Diabetes Metab Res Rev. 2006; 22:423–436. [PubMed: 16506274]
- 117. Muniyappa R, Sowers JR. Role of insulin resistance in endothelial dysfunction. Rev Endocr Metab Disord. 2013; 14:5–12. [PubMed: 23306778]
- 118. Caballero AE. Endothelial dysfunction in obesity and insulin resistance: a road to diabetes and heart disease. Obes Res. 2003; 11:1278–1289. [PubMed: 14627747]
- 119. Duncan ER, Crossey PA, Walker S, et al. Effect of endothelium-specific insulin resistance on endothelial function in vivo. Diabetes. 2008; 57:3307–3314. [PubMed: 18835939]
- 120. Boden G, Cheung P, Salehi S, et al. Insulin regulates the unfolded protein response in human adipose tissue. Diabetes. 2014; 63:912–922. [PubMed: 24130338]
- 121. Zhang W, Hietakangas V, Wee S, et al. ER stress potentiates insulin resistance through PERK-mediated FOXO phosphorylation. Genes Dev. 2013; 27:441–449. [PubMed: 23431056]
- 122. Chen CH, Shaikenov T, Peterson TR, et al. ER stress inhibits mTORC2 and Akt signaling through GSK-3β-mediated phosphorylation of rictor. Sci Signal. 2011; 4:ra10. [PubMed: 21343617]
- 123. Malhotra JD, Kaufman RJ. Endoplasmic reticulum stress and oxidative stress: a vicious cycle or a double-edged sword? Antioxid Redox Signal. 2007; 9:2277–2293. [PubMed: 17979528]
- 124. Hummasti S, Hotamisligil GS. Endoplasmic reticulum stress and inflammation in obesity and diabetes. Circ Res. 2010; 107:579–591. [PubMed: 20814028]
- 125. Hotamisligil GS. Endoplasmic reticulum stress and the inflammatory basis of metabolic disease. Cell. 2010; 140:900–917. [PubMed: 20303879]
- 126. Kolattukudy PE, Niu J. Inflammation, endoplasmic reticulum stress, autophagy, and the monocyte chemoattractant protein-1/CCR2 pathway. Circ Res. 2012; 110:174–189. [PubMed: 22223213]
- 127. Scull CM, Tabas I. Mechanisms of ER stress-induced apoptosis in atherosclerosis. Arterioscler Thromb Vasc Biol. 2011; 31:2792–2797. [PubMed: 22096099]
- 128. Hetz C, Chevet E, Harding HP. Targeting the unfolded protein response in disease. Nat Rev Drug Discov. 2013; 12:703–719. [PubMed: 23989796]
- 129. Fu HY, Minamino T, Tsukamoto O, et al. Overexpression of endoplasmic reticulum-resident chaperone attenuates cardiomyocyte death induced by proteasome inhibition. Cardiovasc Res. 2008; 79:600–610. [PubMed: 18508854]
- Kudo T, Kanemoto S, Hara H, et al. A molecular chaperone inducer protects neurons from ER stress. Cell Death Differ. 2008; 15:364–375. [PubMed: 18049481]
- 131. Wang G, Yang ZQ, Zhang K. Endoplasmic reticulum stress response in cancer: molecular mechanism and therapeutic potential. Am J Transl Res. 2010; 2:65–74. [PubMed: 20182583]
- 132. Cao SS, Kaufman RJ. Targeting endoplasmic reticulum stress in metabolic disease. Expert Opin Ther Targets. 2013; 17:437–448. [PubMed: 23324104]
- 133. Thorp E, Li G, Seimon TA, et al. Reduced apoptosis and plaque necrosis in advanced atherosclerotic lesions of Apoe-/- and Ldlr-/- mice lacking CHOP. Cell Metab. 2009; 9:474–481. [PubMed: 19416717]
- 134. Myoishi M, Hao H, Minamino T, et al. Increased endoplasmic reticulum stress in atherosclerotic plaques associated with acute coronary syndrome. Circulation. 2007; 116:1226–1233. [PubMed: 17709641]
- 135. Minamino T, Komuro I, Kitakaze M. Endoplasmic reticulum stress as a therapeutic target in cardiovascular disease. Circ Res. 2010; 107:1071–1082. [PubMed: 21030724]
- 136. Ozcan U, Yilmaz E, Ozcan L, et al. Chemical chaperones reduce ER stress and restore glucose homeostasis in a mouse model of type 2 diabetes. Science. 2006; 313:1137–1140. [PubMed: 16931765]
- 137. Liang B, Wang S, Wang Q, et al. Aberrant endoplasmic reticulum stress in vascular smooth muscle increases vascular contractility and blood pressure in mice deficient of AMP-activated protein kinase-α2 in vivo. Arterioscler Thromb Vasc Biol. 2013; 33:595–604. [PubMed: 23288166]

138. Erbay E, Babaev VR, Mayers JR, et al. Reducing endoplasmic reticulum stress through a macrophage lipid chaperone alleviates atherosclerosis. Nat Med. 2009; 15:1383–1391. [PubMed: 19966778]