

# UC Davis

## UC Davis Previously Published Works

### Title

Fragile X targeted pharmacotherapy: lessons learned and future directions

### Permalink

<https://escholarship.org/uc/item/6h13d66k>

### Journal

Journal of Neurodevelopmental Disorders, 9(1)

### ISSN

1866-1947

### Authors

Erickson, Craig A  
Davenport, Matthew H  
Schaefer, Tori L  
et al.

### Publication Date

2017-12-01

### DOI

10.1186/s11689-017-9186-9

Peer reviewed

REVIEW

Open Access



# Fragile X targeted pharmacotherapy: lessons learned and future directions

Craig A. Erickson<sup>1,2\*</sup>, Matthew H. Davenport<sup>1,3</sup>, Tori L. Schaefer<sup>1</sup>, Logan K. Wink<sup>1,2</sup>, Ernest V. Pedapati<sup>1,2</sup>, John A. Sweeney<sup>2</sup>, Sarah E. Fitzpatrick<sup>1</sup>, W. Ted Brown<sup>10</sup>, Dejan Budimirovic<sup>11,12</sup>, Randi J. Hagerman<sup>4,5</sup>, David Hessl<sup>4,6</sup>, Walter E. Kaufmann<sup>7,8</sup> and Elizabeth Berry-Kravis<sup>9</sup>

## Abstract

Our understanding of fragile X syndrome (FXS) pathophysiology continues to improve and numerous potential drug targets have been identified. Yet, current prescribing practices are only symptom-based in order to manage difficult behaviors, as no drug to date is approved for the treatment of FXS. Drugs impacting a diversity of targets in the brain have been studied in recent FXS-specific clinical trials. While many drugs have focused on regulation of enhanced glutamatergic or deficient GABAergic neurotransmission, compounds studied have not been limited to these mechanisms. As a single-gene disorder, it was thought that FXS would have consistent drug targets that could be modulated with pharmacotherapy and lead to significant improvement. Unfortunately, despite promising results in FXS animal models, translational drug treatment development in FXS has largely failed. Future success in this field will depend on learning from past challenges to improve clinical trial design, choose appropriate outcome measures and age range choices, and find readily modulated drug targets. Even with many negative placebo-controlled study results, the field continues to move forward exploring both the new mechanistic drug approaches combined with ways to improve trial execution. This review summarizes the known phenotype and pathophysiology of FXS and past clinical trial rationale and results, and discusses current challenges facing the field and lessons from which to learn for future treatment development efforts.

**Keywords:** Fragile X syndrome, Translational treatment, Targeted treatments, Drug development, Genetic disorder

## Background

Fragile X syndrome (FXS) is the most common single gene disorder associated with autism spectrum disorder (ASD) and most common inherited cause of developmental disability. FXS impacts 1 in 4000 males and 1 in 4000–6000 females worldwide [1–5]. FXS results from silencing of the fragile X mental retardation gene (*FMRI*) on the long arm of the X chromosome. Silencing of *FMRI* is nearly always caused by hypermethylation of a cytosine guanine guanine (CGG) trinucleotide repeat expansion ( $\geq 200$  repeats is termed the “full mutation” and causes FXS) in the 5′ untranslated region (UTR) of the *FMRI* gene [6, 7]. *FMRI* inactivation

results in absent or deficient production of fragile X mental retardation protein (FMRP). In all cases, full mutation FXS results from maternal transmission, in which a mother transmits her full mutation allele or her pre-mutation (carrier) allele (55–200 CGG repeats; typical population has fewer than 45 repeats), which undergoes CGG repeat expansion when it transmits to the next generation. As an X-linked disorder, FXS universally impacts affected males, while its presentation is variable in females due to random X inactivation patterns. In FXS, excessive and poorly regulated protein synthesis is pathogenic, which then manifests in myriad ways [8]. Developmental disability, most commonly in the moderate to severe range of cognitive impairment, is universal in males. Common physical and medical features in FXS include increased risk for chronic otitis media, esotropia, hyperextensible joints, high arched palate, low muscle tone, seizures, and macroorchidism with puberty [9, 10]. The neurobehavioral presentation of FXS includes risk

\* Correspondence: craig.erickson@cchmc.org

<sup>1</sup>Division of Child and Adolescent Psychiatry (MLC 4002), Cincinnati Children’s Hospital Medical Center, 3333 Burnet Ave., Cincinnati, OH 45229-3039, USA

<sup>2</sup>Department of Psychiatry, College of Medicine, University of Cincinnati, Cincinnati, OH, USA

Full list of author information is available at the end of the article



for sleep disturbance, aggression, attention deficit hyperactivity disorder (ADHD) symptoms, significant anxiety, sensory hypersensitivities, self-injury, and physical aggression [4, 11, 12]. There is a significant overlap between FXS and ASD, with up to 2 in 3 males with FXS having features consistent with the broader ASD phenotype [3, 5, 13, 14].

FMRP is widely expressed in humans. In the human brain, FMRP is expressed in mature astrocytes and in the dendrites, spines, and soma of mature neurons [15–19]. FMRP is involved in translational repression and thought to selectively bind to about 4% of all mRNAs translated in the brain [17–20], but the impact of deficient FMRP is complex, including an expected increase in translation of many RNA targets. However, protein expression of other mRNA targets of FMRP may be unchanged or even reduced in FXS, thus pointing to poorly understood compensatory or other regulatory mechanisms [21].

The loss of FMRP results in a number of brain effects at macroscopic, microscopic, and molecular levels. Neuroimaging abnormalities noted in youth with FXS include larger temporal lobe white matter, cerebellar gray matter, and caudate nucleus with smaller amygdala compared to controls [22]. At a microscopic level, dendritic abnormalities associated with deficient FMRP include increased spine density with longer, spindly, and immature morphology reported in postmortem human and *Fmr1* knockout (KO) mouse brain tissue [23–25]. Molecular and dendritic abnormalities may result in functional brain deficits, including disruption of synaptic plasticity with enhanced long-term depression (LTD) [26–29] and brain region-specific long-term potentiation (LTP) deficits [30–33]. Not surprisingly, given the large number of proteins for which production is altered by FMRP deficiency, many molecular signaling cascades involved in synaptic plasticity, learning, and memory are known to function abnormally in the *Fmr1* KO mouse and in human cells. Some of these likely dysregulated molecular systems include phosphoinositide 3-kinase (PI3K) [34–36], extracellular signal-related kinase (ERK1/2) [37–39], matrix metalloproteinase 9 (MMP-9) [40, 41], endocannabinoid [42–45], brain-derived neurotrophic factor (BDNF) [46, 47], and mammalian target of rapamycin (mTOR) [48–50]. The broad array of potential molecular targets for pharmacotherapy in FXS is a testament to the broad impact of deficient FMRP, and thus the potential challenge of targeting multiple aspects of molecular dysregulation simultaneously [42–45].

As a single gene disorder with increasing efforts to define and then address the neurobiological underpinning of the disorder, FXS has been the subject of a recent wave of targeted treatment development efforts. Despite significant hopes for translational treatment success, to date no drug has met approval for use specifically in

FXS. Given this, the pharmacotherapy of FXS in clinic continues to be limited to symptomatic treatments of comorbid abnormal behaviors, employing medications such as selective serotonin reuptake inhibitors (SSRIs), stimulants, and second-generation (atypical) antipsychotics [51]. We will focus on efforts to develop FXS-specific pharmacotherapy, including the translational basis for various treatment hypotheses, focused on results of human studies. Finally, we will look in detail at potential reasons for study failure providing future directions for consideration to address such challenges.

### Modulation of glutamate and GABA neurotransmission

Many of the recent targeted clinical trials in FXS have addressed a potential excitatory/inhibitory neurotransmission imbalance associated with the disorder (Table 1). In the FXS brain, there is believed to be an excess of excitatory, glutamatergic signaling coupled with deficiencies in inhibitory,  $\gamma$ -Aminobutyric acid (GABA)-ergic signaling [52]. Several recent human trials in FXS have focused on reducing excitatory glutamatergic neurotransmission. Specific glutamatergic effects probed in humans with FXS have included antagonism of group I metabotropic glutamate receptors (mGluRs), particularly mGluR5 (ClinicalTrials.gov Identifiers: NCT01253629, NCT01357239, NCT01517698, NCT01015430, NCT01750957), the N-methyl-D-aspartate (NMDA) receptor, stimulation of GABA(B) receptors (NCT01325220, NCT00788073, NCT01282268), and modulation of the  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid glutamate receptor (AMPA; NCT00054730).

The mGluR theory of FXS pathophysiology has driven the wave of glutamatergic modulator studies in FXS. The mGluR theory postulates that in FXS, excessive signaling through mGluRs contributes to behavioral, electrophysiological, and molecular dysfunction associated with the disorder [8]. The mGluR theory brought together several key findings, including observations that FMRP represses protein translation at the synapse [53], synaptic protein synthesis can be triggered by activation of mGluRs [16], FMRP deficit leads to increased downstream effects of mGluR signaling [26, 54], and many of these downstream effects are dependent on mRNA translation at the synapse [55–58]. The mGluR theory has been extensively corroborated in FXS preclinical research using 2-methyl-6(phenylethynyl)pyridine (MPEP), a selective mGluR5 antagonist neurotoxic to humans. This work has included many studies of treatment of the *Fmr1* KO mouse with selective mGluR5 antagonists, demonstrating rescue of aberrant AMPAR expression, behavioral deficits, electrophysiological abnormalities, protein expression dysregulation, and altered dendritic spine morphology [29, 59–61]. Further corroboration of

**Table 1** Clinical trials to date in fragile X syndrome by drug and study type

Drug	Mechanism	Open Label/ Chart Review	Phase II	Phase III
Acamprosate	GABA-R agonist	C[98]; C[99]	O	
Arbaclofen	GABA <sub>B</sub> -R agonist		C[102]	P
Basimglurant	mGluR5 antagonist		P	
CX516	AMPA (+) modulator		C[85]	
Donepezil	Anticholinergic	C[132]	C[134]	
Fenobam	mGluR5 antagonist	C[73]		
Ganaxolone	GABA <sub>A</sub> R agonist		O	
Lithium	GSK3 inhibitor	C[114]		
Lovastatin	ERK inhibitor	C[125]		
Mavoglurant <sup>a</sup>	mGluR5 antagonist		C[75], C[76, D 106]	
Memantine	NMDAR antagonist	C[84]		
Metadoxine ER	GABA agonist		P	
Minocycline	MMP9 inhibitor	C[117]	C[117]	
Oxytocin	OXTR agonist		C[143]	
Riluzole	Glutamate agonist	C[93]		
R04917523	mGluR5 antagonist		D	
Trofnetide <sup>b</sup>	IGF-1 mimic		P	

Key: C completed and published, P completed and pending publication, O ongoing clinical trial, www.clinicaltrials.gov, D discontinued and unpublished  
<sup>a</sup>AFQ056  
<sup>b</sup>NNZ-2566

the mGluR theory has come from genetic knockdown studies in the *Fmr1* KO mice, in which reduction in mGluR5 expression normalized protein synthesis, dendritic spine deficits, and aspects of aberrant behavior [62], although a second study using the same model found very few behavioral improvements [63].

Within investigation of excitatory/inhibitory imbalance, enhanced glutamatergic signaling has received the most attention in FXS, but GABAergic deficits have been increasingly recognized as contributing to the hypothesized excitatory/inhibitory imbalance. In the *Fmr1* KO mouse, deficits in GABAergic signaling occur in a variety of brain regions including the hippocampus, striatum, amygdala, and somatosensory cortex [64–67]. Most commonly, preclinical findings of GABAergic deficits have centered on reduction in GABA(A) subunit receptor expression, although GABA synthesis and release may also be affected [64]. In preclinical treatment study in the mouse and fly model of FXS, positive modulation of GABA(A) receptors can rescue some behavioral and neurophysiological alterations [68, 69]. GABA(B) activators also rescued an array of phenotypes including spine morphology, audiogenic seizures, and cellular abnormalities [70, 71]. Overall, the imbalance of glutamatergic

and GABAergic signaling in FXS may represent a complex interplay of abnormalities on both sides of the excitatory/inhibitory neurotransmission balance.

Phenotypic rescue demonstrated in the *Fmr1* KO mouse, first with use of selective mGluR5 antagonists and later with other mechanistic approaches, has driven the extensive clinical trial work in the field since 2008. To date, a total of 22 such studies have been identified through search of literature and other sources; 19/22 (86%) have been registered on www.ClinicalTrials.gov. As expected from FXS neurobiology, the vast majority of the studies have targeted the core excitatory/inhibitory imbalance in the disorder primarily through either mGluR5 antagonists (mavoglurant-AFQ056, NCT01357239, NCT01253629; basimglurant-RO4917523, NCT01517698, NCT01750957) or GABA agonists (arbaclofen-GABA-B agonist, NCT01282268, NCT00788073, NCT01325220; ganaxolone-GABA-A agonist, NCT 01725152). These studies represent the majority of the total (14/22, 64%) and FDA-registered (14/19, 74%) trials. Reflecting that over 2/3 of these trials were phase II, most of them have studied adults and adolescents (i.e., regulations specify that novel drugs need to be tested first in adults, particularly in vulnerable populations). Trials of three specific mGluR5 antagonists—fenobam, mavoglurant (AFQ056), and basimglurant (RO4917523)—in human FXS study have been completed. The first pilot trial involved fenobam [N-(3-chlorophenyl)-N'-(4,5-dihydro-1-methyl-4-oxo-1H-imidazole-2-yl)urea], a nonbenzodiazepine anxiolytic drug and negative allosteric modulator of mGluR5 [72]. An open-label single-dose study was conducted in 6 males and 6 females with FXS in the 18 to 30-year-old range [73]. This pilot study initially evaluated drug safety and pharmacokinetics and explored aspects of sensory gating, attention and inhibition by evaluating prepulse inhibition (PPI) before and after each single dose fenobam treatment. No significant adverse events were noted. Six out of 12 (50%) subjects met predefined response criterion of at least 20% improvement over baseline on PPI at 120 ms. Further studies in FXS were not carried out because the company that manufactured fenobam failed financially. In earlier reports in the general population, fenobam has a challenging tolerability profile, with reports of hallucinations, vertigo, paraesthesias, and insomnia with fenobam use [74].

Mavoglurant is a noncompetitive mGluR5 antagonist developed by Novartis Pharmaceuticals. Three placebo-controlled trials of mavoglurant have been completed in FXS. The first trial was a double-blind, placebo-controlled 20-day treatment period crossover trial including 30 adults with full mutation FXS [75]. While this initial study failed to show any positive drug-associated effect on primary or secondary outcomes, including the

Aberrant Behavior Checklist (ABC) or Clinical Global Impressions (CGI) scales, in the whole study group, a post-hoc subset analysis of seven individuals with complete *FMRI* promoter methylation noted significant drug-associated improvement on a number of behavioral outcome measures. This post-hoc effect may have been driven by an abnormally low placebo response in the small seven-subject subset of patients with complete methylation. This finding could also be attributable to regression to the mean with those with complete methylation with a potentially more significantly impaired phenotype. Two additional large-scale double-blind, placebo-controlled, parallel groups, fixed-dose four-arm (placebo, 25 mg BID, 50 mg BID, and 100 mg BID dosing) studies with open label extension were conducted with mavoglurant in FXS [76]. The trials included 12-week adult (age range 18–45 years) and adolescent (age range 12–17 years) placebo-controlled treatment periods, both utilizing the total score of a FXS-refactored version [77] of the Aberrant Behavior Checklist-Community (ABC-C) [78], termed ABC<sub>FXS</sub>, as a primary outcome. Neither study met significance on the primary endpoint and the sponsor subsequently terminated the open-label extension portion of the studies and discontinued the development program of mavoglurant in FXS (ClinicalTrials.gov Identifiers: NCT01253629, NCT01357239); [76, 79].

A third selective mGluR5 antagonist, basimglurant, was evaluated in a small phase IIa placebo-controlled PK study in adults, a subsequently a larger phase IIb 3-month double-blind, placebo-controlled study in adolescents and adults (age range 14–50 years) (ClinicalTrials.gov Identifier: NCT01517698), and a small phase IIa pharmacokinetic study in youth (age range 5–13 years) with FXS (ClinicalTrials.gov Identifiers: NCT01015430, NCT01750957). Although all the trial results remain unpublished, due to lack of efficacy on a number of behavioral and other outcome measures employed in phase IIb adult/adolescent study, the Roche Group subsequently terminated its program for the development of basimglurant in FXS [80].

Mavoglurant and basimglurant trial results have made it clear that short-term selective mGluR5 antagonism is not associated with significant behavioral improvement in the age ranges studied. Several questions remain to be answered, including what impact this class of drugs may have on very young children and what the impact of prolonged treatment may be on other outcomes such as cognitive or communication metrics. The selective mGluR5 antagonist trials to date have also been limited by the employed outcome measures, with a focus on parent-reported behavioral outcomes that resulted in dramatically enhanced placebo response.

In addition to evidence of mGluR5 dysregulation in FXS, activity at the NMDA glutamate receptor may be anomalous in the disorder, although the overall directionality of dysregulation is unclear, appearing in preclinical models to depend on brain region and developmental stage [26, 81–83]. Memantine (3,5-dimethyladamantan-1-amine), a compound that acts as a NMDA noncompetitive antagonist, is the US Food and Drug Administration (FDA) approved for the management of Alzheimer's disease. In the only published trial of memantine in humans (age range 13–22 years,  $n = 6$ ) with FXS to date, over a mean 34.7 weeks of open-label treatment, 4 subjects showed clinical improvement as rated by the Clinical Global Impression-Improvement subscale (CGI-I), but 2 subjects had to discontinue therapy due to increased irritability with treatment [84]. No specific domains of symptom or behavioral improvement were noted and the authors cautioned about future exploration of this drug given the worsening irritability noted in one third of subjects.

AMPA is an ionotropic glutamate receptor mediating fast synaptic transmission. Modulation of AMPAR activity downstream of mGluR signaling, was initially proposed as a method of restoring excitatory:inhibitory signaling balance in FXS [8]. Level of internalization of AMPARs, which is increased in the *Fmr1* KO mouse, may contribute to alterations in LTD and LTP in FXS since AMPAR signaling is required for maintaining synaptic plasticity. A positive allosteric modulator of AMPAR, CX516, was tested in a 4-week double-blind, placebo-controlled trial in FXS. The trial failed to find a significant treatment-associated improvement in memory, the study's primary measure, or any secondary endpoint including measures of language skills, behavior, and global improvement. The authors hypothesized that CX516 drug effect may have been limited by potential subtherapeutic dosing based on information that became available concurrent to the trial [85]. Despite the negative study findings, the CX516 trial laid the groundwork for the use of many outcome measures in future FXS trials by providing invaluable data regarding which measures may not be as affected by floor, ceiling, or test-retest inconsistency effects.

Beyond drugs solely modulating glutamatergic neurotransmission, several agents with combined glutamate and GABA activity have been studied in FXS. Dysregulation of the ERK intracellular signaling pathway has been implicated in the pathophysiology of FXS and as such has become a potential target of treatment in the disorder. Acting downstream from several cellular receptors including mGluRs, ERK activity is required for normal synaptic plasticity and the regulation of activity dependent protein synthesis [86]. ERK activity has been demonstrated to be upregulated under baseline conditions in the *Fmr1* KO



mouse and in human post-mortem tissue [28, 49, 87]. The kinetics of ERK activation is delayed in FXS, potentially because of baseline hyperactivation. Reduction of phosphorylated (activated) ERK has been linked to rescuing the audiogenic seizure phenotype [87] and the increased hippocampal protein synthesis in the *Fmr1* KO mouse [88].

Riluzole is FDA-approved for the treatment of amyotrophic lateral sclerosis (ALS) and may potentially be useful in the treatment of depression and anxiety [89, 90]. The drug is hypothesized to inhibit glutamate release [91] and potentiate post-synaptic GABA(A) receptor activity [92]. Riluzole was the subject of a small six subject 6-week open-label study in adult males with FXS [93]. Only 1 in 6 subjects (16%) showed a positive clinical response after the brief treatment (100 mg/day). Despite the lack of significant clinical effect, peripheral lymphocytic ERK activation, which is known to be delayed in blood lymphocytes in FXS human and animal models [94], was significantly corrected in all subjects following the 6-week riluzole treatment. This result, potential rescue of molecular dysregulation combined with a lack of clinical response, may be a signal that short-term trials are not allowing the time for molecular change to generate resultant positive change in the clinical phenotype of FXS. It may also be possible that ERK dysregulation is a downstream consequence of a different molecular mechanism and thus an isolated correction of ERK activity could lack significant clinical impact.

Acamprosate is FDA-approved for the maintenance of abstinence in alcohol dependence. Acamprosate is hypothesized to have combined effects on excitatory:inhibitory balance in brain, including potential potentiation of GABA(A) activity [95] and antagonism at mGluR5 [96] and NMDA glutamate receptors [97]. Acamprosate has been the subject of several open-label reports in FXS including an initial report on 3 adults with FXS who received a mean 21.3 weeks of acamprosate treatment [98]. In this study, each adult showed a positive clinical treatment response marked in part by enhancement in language and communication skills. In the first investigation of acamprosate in youth with FXS, over 10 weeks of open-label treatment (mean dose  $1054 \pm 422$  mg/day), 9 of 12 youths were deemed treatment responders with response marked by a score of "much" or "very much improved" on the CGI-I [99]. In addition, overall group improvement was noted on the ABC-C Social Withdrawal subscale (ABC-SW), Social Responsiveness Scale (SRS), and attention deficit hyperactivity disorder rating scale (ADHD-RS). In this report, plasma BDNF was sampled at baseline and after acamprosate treatment. Acamprosate use was associated with increases in peripheral BDNF levels after 10 weeks of treatment. Additionally, plasma amyloid precursor protein (APP)

and APP alpha were reduced with acamprosate use in this trial [100]. Overall, in open-label studies, acamprosate has shown promise to improve the clinical phenotype of FXS and the drug has been associated with two aspects of molecular change that may signal aspects of engagement with the underlying molecular dysregulation that characterizes the disorder. Acamprosate is currently undergoing double-blind, placebo-controlled 10-week study in 48 persons with FXS age 5–22 years (ClinicalTrials.gov Identifier: NCT01911455).

The neuroactive steroid ganaxolone (3 $\alpha$ -hydroxy-3 $\beta$ -methyl analog of allopregnanolone) is a positive allosteric modulator at GABA(A) receptors. Ganaxolone has blocked audiogenic seizures in the *Fmr1* KO mouse [101]. A double-blind, placebo-controlled 6-week treatment period, crossover trial of ganaxolone was recently completed in 6–17 years old with FXS (ClinicalTrials.gov Identifier: NCT01725152), although results have not yet been published or presented. This first in FXS ganaxolone study aimed to determine the safety, tolerability, and efficacy of the drug for the treatment of anxiety and attention deficits in FXS.

Arbaclofen, the active enantiomer of racemic baclofen, is a GABA(B) agonist studied to date in FXS and in idiopathic ASD. Arbaclofen, a presynaptic GABA(B) agonist, is postulated to inhibit glutamatergic release, thus potentially limiting neuronal hyperexcitability associated with FXS. In the *Fmr1* KO mouse, arbaclofen was shown to reduce susceptibility to audiogenic seizures and normalize excessive dendritic spine density and protein synthesis [70]. In a double-blind, placebo-controlled 4-week treatment period crossover trial in 63 persons with FXS age 6–40 years, the drug was well tolerated but not associated with a positive drug effect on the study primary outcome, the ABC-C Irritability subscale (ABC-I) [102]. Other outcomes, including a parent visual analog scale (VAS) of the child's 3 most challenging behaviors reported by caregivers and the ABC Social Avoidance (ABC-SA) subscale, which was developed specifically by re-factoring the ABC-C in the FXS population (ABC<sub>FX</sub>) [77] were improved on arbaclofen in the entire per intent-to-treat (ITT) treatment group. In post-hoc analyses, a more socially impaired subgroup defined by high ABC-C Social Withdrawal (ABC-SW) scores at baseline showed a positive drug-associated treatment effect on many measures including the CGI-Severity (CGI-S), CGI-I, treatment preference, VAS, and ABC<sub>FX</sub> Social Avoidance subscale. Subsequent phase III studies of arbaclofen in FXS across children, adolescents, and adults did not show significant drug-associated improvement on primary outcome measures tested (Berry-Kravis et al. 2016, companion paper in this issue of the Journal). The pediatric study did show arbaclofen-associated broader measures improvement on several key secondary

outcomes and a trend toward significance for arbaclofen on the primary endpoint, but the study was not adequately powered for the primary outcome measure due to early closure for financial reasons. These results combined with parallel negative phase III findings in idiopathic ASD led to the discontinuation of the development of arbaclofen by Seaside Therapeutics, when the company ceased operations.

### Targeted treatment development beyond glutamate and/or GABA modulation

Metadoxine (pyridoxol 1-2-pyrrolidone-5-carboxylate) has been used to treat alcohol intoxication outside of the USA for many years. Metadoxine ER (MDX), which has been shown to increase striatal dopamine levels in murine models [103], is currently being developed by Alcobra Pharmaceuticals for use in adult and pediatric ADHD. MDX is currently in phase III development for adults with ADHD (ClinicalTrials.gov Identifiers: NCT02477748 and NCT02189772). In the *Fmr1* KO mouse, as reported by Alcobra but not available in a peer-reviewed manuscript format, MDX use was associated with improvements in attention, memory, learning, hyperactivity, and sociability, which were associated with molecular normalization of Akt and ERK over-activity (<http://www.alcobra-pharma.com/releasedetail.cfm?ReleaseID=847048>). A 6-week randomized, double-blind, placebo-controlled trial of MDX enrolling 62 persons (57 completed treatment) with FXS, age 14–55 years (mean age: 24 years), was recently completed (ClinicalTrials.gov Identifier: NCT02126995) [104]. MDX use was not associated with significant improvement on the study's primary outcome measure: the inattentive subscale of the ADHD Rating Scale 4th Edition (ADHD-RS-IV). An analysis of secondary outcomes included a positive report of MDX-associated benefit as measured by the Vineland Adaptive Behavior Scale, Second Edition (Vineland-II) Daily Living Skills Domain [76, 104, 105] and the computerized cognitive Test of Attentional Performance for Children (KiTAP) Go-NoGo subscale false reactions ( $p = 0.043$ ). Although improvement on the Vineland-II would be an important functional outcome in FXS, future work including replication is necessary to further understand any potential utility of MDX use in persons with FXS.

Lithium is an effective mood stabilizer, FDA-approved for the treatment of bipolar disorder. The drug has combined effects including inhibition of glycogen synthase kinase-3 beta (GSK-3b). GSK-3b has been shown to be dysregulated in the *Fmr1* KO mouse [106, 107]. In this model of FXS, lithium use has been associated with improvements in hyperactivity, social preference, learning, and aberrant dendritic spine development [108]. Lithium has also been shown to rescue synaptic plasticity, protein

synthesis, and aberrant GSK-3b activity in the *Fmr1* KO mouse [109–112]. Lithium has been evaluated in a 15-person open-label 2-month study in FXS [113]. In this report, lithium use was not associated with significant improvement on the ABC-I, but did show treatment-associated improvement on a number of secondary outcome measures including other ABC-C subscales, the ABC-C total score, the CGI-I, a visual analog scale (VAS) for behavior, the Repeatable Battery for the Assessment of Neuropsychological Status (RBANS) List Learning subtest assessing verbal memory, and ERK activation in lymphocytes. Adverse effects associated with lithium use in this trial included aggression, polydipsia, and enuresis. The side effect profile of lithium including risk of thyroid and renal impairment has limited further development of this compound in FXS.

Minocycline, an FDA-approved antibiotic treatment for acne in youth, is known to have inhibitory effects on MMP-9 activity. MMP-9 activity has been demonstrated to be elevated in the hippocampus of *Fmr1* KO mice [114]. In this mouse model of FXS, minocycline treatment was associated with reduced hyperactivity and improvement in dendritic spine phenotype [115]. In an initial 8-week open-label minocycline trial in 20 persons with FXS age 13 to 35 years, the drug treatment was associated with widespread improvement as captured by the CGI and ABC subscales [116]. A subsequent 12-week treatment period double-blind, placebo-controlled crossover trial in 55 subjects age 3.5 to 16 years with FXS noted drug-associated improvement on the CGI-I, but no group-wide improvement in specific behavioral domains [117]. A post-hoc analysis noted improvement on the VAS specific to anxiety and mood-related concerns. In an electrophysiology analysis of a 12-subject subgroup from the placebo-controlled minocycline trial, minocycline use was associated with improvement in habituation to auditory stimuli as shown by an event-related potentials (ERP) passive auditory oddball paradigm [118]. The authors hypothesized that this electrophysiological improvement in habituation may be related to improvements in hypersensitivity to auditory stimuli noted in humans with FXS following minocycline administration. Dziembowska et al. (2013) demonstrated that MMP-9 blood level was significantly elevated in 20 patients with FXS compared to controls; 6 of the 10 children treated with minocycline for 12 weeks showed a significant lowering of their MMP-9 levels, although their response on the CGI-I did not correlate with the degree of MMP-9 lowering. Although minocycline is recognized primarily for lowering MMP-9, it has multiple other effects including stalling translation, decreasing apoptosis, and working as an antioxidant, all of which may be helpful in FXS [119].

Lovastatin is a compound FDA-approved for the long-term management of familial hyper-cholesterolemia

[120], with demonstrated effects on intracellular signaling. In cultured rat brain neuroblasts, lovastatin was shown to inhibit Ras signaling, an upstream effect that resulted in reduction in ERK activation [121] that supported previous work in fibroblasts [122]. In *Fmr1* KO mice, lovastatin was confirmed to inhibit Ras, reduce increased basal ERK activation, lower protein synthesis to wild type levels, and ameliorate FXS audiogenic seizure susceptibility [123]. Based upon the known safety profile of lovastatin and the aforementioned promising preclinical results, efficacy of lovastatin in FXS was assessed in a 16-person, open-label trial of children and adolescents. Treatment response was assessed using the ABC-C, CGI-I, and Vineland-II. Significant improvement was observed after 4 and 12 weeks of treatment, with the ABC-C, CGI, and Vineland-II scores improving from week 4 to week 12. Excessive ERK activity measured in platelets was reduced by lovastatin in this trial and correlated with behavioral improvement on the ABC-C. There was modest improvement on the CGI-I, but the open-label nature of the trial precludes any strong inferences of efficacy at this stage of development [124]. Furthermore, particular importance should be placed on lipid monitoring in future lovastatin trials since individuals with FXS were reported to have lower levels of low and high density lipoprotein and total cholesterol [125].

A synthetic analog of the naturally occurring active N-terminal tripeptide derived of the insulin-like growth factor 1 (IGF-1), known as NNZ-2566 or trofinetide has been studied in the *Fmr1* KO mouse and in humans with FXS. The drug was initially developed for traumatic brain injury, where trofinetide showed promise with improved recovery, reduction in apoptotic cell death, and reduced neuroinflammation noted in a rat model [126–128]. In the *Fmr1* KO mouse, trofinetide has been reported to rescue learning and memory deficits, normalize dendritic spine morphology, and restore normal ERK signaling [129]. Recently, a double-blind, placebo-controlled 4-week trial of NNZ-2566 in 12 to 45 year old males with FXS was completed. The study used both parent- and clinician-reported outcomes designed to address the entire FXS phenotype, including an FXS Rating Scale and FXS Domain Specific Concerns measure. A composite analysis of five measures from three domains utilized a novel direction of clinical change analysis plan, including a group and individual analyses. Both clinicians and caregivers observed a pattern of consistent improvements in FXS-specific measures and the ABC total score, but only at the higher dose (70 mg/kg b.i.d.;  $p = 0.045$  by permutation testing) ( $p = 0.045$  by permutation testing) (ClinicalTrials.gov Identifier: NCT01894958). Considering similar positive results in a comparable trial of adolescents and adults with Rett syndrome, trofinetide appears to be a promising

drug for neurodevelopmental disorders (ClinicalTrials.gov Identifier: NCT01703533).

Given reports of altered choline levels and cholinergic function in FXS murine models and in human studies, donepezil, an acetylcholinesterase inhibitor FDA-approved for the treatment of Alzheimer's disease, has been initially investigated in FXS. Specifically, *FMRI* has been shown to be highly expressed in cholinergic neurons during the course of normal development [130], and choline levels were shown to be lower in persons with FXS in a small 1H magnetic resonance spectroscopy study [131]. Additionally, dysregulated cholinergic function has also been demonstrated in the subiculum of *Fmr1* KO mice [132]. In humans with FXS, a 9-subject, 6-week open-label trial of donepezil reported good drug tolerability and significant treatment-associated improvement on the ABC-C Hyperactivity and Irritability subscales [131]. Recently, results from a 12-week randomized double-blind, placebo-controlled trial of donepezil (maximum dose 5 mg per day) in 20 boys (mean age  $9.1 \pm 2.6$  years) with full mutation FXS have been reported [133]. In this study, donepezil use was not associated with significant positive change on outcome measures employed including the Stanford-Binet Intelligence Scale, Conners 3 parent ADHD rating scale, or the Childhood Autism Rating Scale (CARS). Researchers at Stanford University recently completed a 12-week randomized double-blind, placebo-controlled, parallel-group study of donepezil (dosed 2.5 to 10 mg per day) in 42 persons with FXS (27 males, 15 females; enrolling both youth and adults under age 65) (ClinicalTrials.gov Identifier: NCT01120626), using the Contingency Naming Test (CNT) as the primary outcome measure. Analyzed results from this project are not yet available.

Sertraline, an SSRI, is known to improve BDNF levels in the CNS and to boost the deficient serotonin levels seen on positron emission tomography (PET) scanning in the brains of children with ASD who are under 5 years old [134, 135]. A randomized controlled trial of low-dose sertraline (2.5 to 5.0 mg) for 6 months in 57 young children ages 2 to 6 years with FXS was recently carried out (Greiss-Hess et al. 2016, *JDBP*, in press). Significant improvements were not shown on the primary outcome measures—the CGI-I and the expressive language subtest on the Mullen Scales of Early Learning (MSEL). However, subjects did demonstrate significant improvement on the Visual Perception subscale, Fine Motor subscale, and composite T score of the MSEL in secondary analyses. In addition, in a post-hoc analysis those children with comorbid FXS and ASD (60% of sample) demonstrated significant improvement on the Expressive Language subscale of the MSEL.

## Discussion

Despite a large number of positive preclinical drug studies in animal models of FXS, to date no approved FXS-



specific drug treatments have been developed. Therefore, the treatment of FXS (mainly behavioral abnormalities) continues to be symptomatic. The numerous trial failures in the last decade could be attributed to a variety of factors; prominently trial design and outcome measures. Nevertheless, several promising areas of translational treatment and strategies to develop such treatments in FXS remain. Success in this field will be predicated in part by learning from past challenges [136, 137].

During the recent phase of significant translational research development in FXS, early failures may have been in part due to attempts to pattern FXS clinical trials after work on the FDA-approved atypical antipsychotics, aripiprazole and risperidone, for targeting irritability (aggression, self-injury, and severe tantrums) in youth with ASD. This approach was likely influenced by advice from regulatory bodies such as the FDA and others in industry, whose experience with neurodevelopmental disability drug approvals was limited and anchored specifically on the use of the ABC-C to measure irritability in registration trials. Earlier on during the development of arbaclofen for FXS, it became clear that irritability may not be the most sensitive or specific metric by which to judge FXS-specific treatments in development. This early reliance on precedents in ASD irritability-focused drug development was also likely driven by the lack of FXS-specific outcome measures. As discussed in detail in another article in this issue, the FXS field has been spending considerable efforts to develop novel and better outcome measures, but to date such instruments are not in regular use. The emphasis on using the ABC-C because it was previously vetted by the FDA may have focused trials on a behavioral irritability, a potentially less marked clinical manifestation in FXS when compared to ASD. Moreover, this focus also likely led to use of the ABC-C total score (combination of all ABC-C subscales' scores) as an outcome, which is not a recommended or empirically supported use of the measure because a subject may have a mix of worsening and improvement in various behavioral aspects evaluated by different ABC-C subscales, thus blurring interpretation of a "total" score.

The FXS clinical trial field developed quickly marked by the need for a multi-site trial infrastructure where none previously existed. Within the last decade, industry rapidly developed molecules for large-scale placebo-controlled study in FXS. The rapid rate of clinical trials developed in FXS may have contributed to some early multi-site trial challenges. Issues such as across-site rating and assessment of enrollment criteria fidelity may have presented challenges to consistent study ratings. For example, in a study of arbaclofen in idiopathic ASD, a result was reported on the Vineland-II for only subjects assessed per protocol with a description of how

many subjects did have the Vineland-II administered as requested. Such a finding may point to how having a more established multi-site trial infrastructure over time may improve study fidelity. Fortunately, developments such as the Fragile X Clinical and Research Consortium by the National Fragile X Foundation and enhanced site training over time has led to establishment of a trial infrastructure prepared to generate the most accurate and consistent trial data possible.

One important concern about the progression of the translational effort in FXS is the reliance on post-hoc analyses of preliminary studies in making critical subsequent clinical trial study design decisions for larger, more pivotal studies examining efficacy. For example, in the first AFQ056 clinical trial, a post-hoc analysis showed significant improvement in just 7 subjects with complete methylation, leading to a very substantial effort to stratify by this variable in a much larger trial. In the earlier arbaclofen trial, a post-hoc analysis showed improvement in the social avoidance subscale of the ABC [102], leading to the decision to focus on social avoidance as the primary outcome in a subsequent trial [Berry-Kravis et al.; companion paper in this journal]. In the metadoxine study, which focused on ADHD symptoms as the primary target, post-hoc analyses showed significant improvement in daily living skills, which may lead to decisions on the target for future study of this compound. It may be quite rational to design future studies based on post-hoc observations, as these adjustments may lead to demonstration of the true benefits of a compound on a different clinical problem or subgroup. However, investigators must also appreciate the risks in moving a focus of research in a new direction that is based on type II error occurring when a large number of post-hoc analyses are completed yielding one or more "false positives". In this regard, it may be useful to consider whether several independent pieces of data suggest a similar clinical benefit, whether the post-hoc results represent clinically meaningful, not just statistically significant changes, whether the changes could be more associated with side effects of the drug rather than true improvement, and whether the clinical changes make sense based on the understanding of the neurobiological and pharmacological mechanisms of the compound under investigation.

Enhanced placebo response rates have also potentially contributed to early failures in FXS-specific treatment development. In ASD, registration trials for risperidone and aripiprazole targeting irritability reported placebo response rates in the range of 12–14% [138, 139]. In FXS, we have seen a much higher rate of placebo responders—as high as 34.6% in the arbaclofen trial—that increased the necessary treatment effect for detecting statistically significant changes [76, 102]. This issue may be driven by

several factors. First, there has been great anticipation among FXS stakeholders that new treatments were on the horizon and would build from positive drug treatment data in FXS animal models. Developments like the mGluR theory of FXS received significant worldwide scientific attention and coverage in the general press, thus potentially enhancing the perception that new FXS-specific treatments being studied would be met with significant and brisk success. To date, attempts at reducing placebo effect, such as single-blind run-in treatment periods used in study of AFQ056 (mavoglurant) have had limited benefit in the field. Recent efforts to use more clinician-anchored measures performed by clinician interview show promise to reduce reliance on single parent-report measures, which may be more prone to enhanced placebo effect. For example, one effort by Neuren Pharmaceuticals focused on assessing group and individual analyses for direction of change in a diverse basket of outcomes, including both parent and clinician report. This kind of analysis is also more likely to assess changes impacting any aspect of the FXS phenotype rather than just a single behavior. As the FXS field tests treatments targeting general mechanisms, it is expected that it will be important to evaluate the entire phenotype since the degree of behavioral manifestations in different domains vary across patients. In this regard, the further development and full validation of a disease-specific outcome measure, such as the Fragile X Syndrome Rating Scale (FXSRS) used in the trofinetide trial, becomes a milestone in the process of creating better instruments for intervention studies in FXS. There is increasing evidence that individual behavioral problems in FXS may appear differently by their co-occurrence with others (e.g., anxiety and ASD) [140, 141]. In this regard, the development of a comprehensive FXS behavioral phenotype measure such as the FXSRS represents an important effort in this area. Objective measures, either performance-based measures of cognition or biomarkers (discussed below) may also be used to more accurately track treatment response with much less concern about placebo response.

There have likely been shortcomings in attempts to match outcome measures in FXS clinical trials to what may be expected from short-term treatment with drugs that rescue synaptic and other cellular pathology in mouse models of FXS. There are no clear correlates between outcomes employed in FXS animal studies and outcomes employed in initial human FXS clinical trials. For example, a drug may correct protein synthesis, dendritic spine morphology, learning, and audiogenic seizure deficits in the *Fmr1* KO mouse. Then, when moved into human studies, outcomes have included parent-report checklists focused on interfering behavior, mood, anxiety, inattention, and adaptive behavior. Thus, it is unclear what type of behavioral change would be expected with a new

treatment that rescues brain connectivity, protein synthesis, and/or neuronal circuit excitability among other features targeted in *Fmr1* KO animal studies. Given this, it will be important for the field to attempt to develop testing paradigms, particularly in rodent FXS models that can be recapitulated in FXS human studies as much as possible. Despite this, and considering that species differences in FMRP and other regulatory proteins (e.g., MeCP2) exist and influence brain connectivity, it is expected that only a few phenotypes and paradigms could be applied in both rodents and humans.

The FXS translational treatment field, like most other similar fields attempting to bring treatments from bench to bedside, has not developed a sense of what age range is most appropriate to enhance chances of treatment success. While efforts in the *Fmr1* KO mouse have demonstrated phenotypic improvement in adult animals, it is possible that initial negative trial results in humans utilizing drugs with marked success in animal models of FXS may be due to missing appropriate developmental windows in the human FXS condition that best respond to treatment. This thinking has led to the concept that the earlier in life treatment is initiated, the better the chance may be of success; this has led to the upcoming study of mavoglurant in toddlers with FXS. Additionally, the duration of treatment necessary to enact a significant change in humans with FXS is not known. While improvement in behavioral and other neurobiological phenomena has occurred rapidly in FXS animal models, it is unclear if change in affected humans may take significantly longer than the short-term trial efforts to date. Longer trials have been inhibited by increased expense and other logistical issues, and by a lack of available natural history data in the field, which would allow for detection of subtle yet potentially meaningful developmental changes over time. There is also a potential that an effective FXS-specific drug treatment would enact improvements in neurobiological parameters that may actually enhance learning over time as a primary readout versus quickly providing behavioral symptomatic relief. If true, it will likely be necessary to pair new treatments with structured training/learning paradigms in order to provide standard didactic methods to a brain that has enhanced capacity to learn (synaptic plasticity) under the influence of a beneficial drug. Such efforts are ongoing in the novel, recently funded trial of mavoglurant in toddlers with FXS, where drug or placebo will be paired with a structured, standardized language-learning paradigm. Finally, extrapolations from genetically homogenous inbred mice raised in controlled settings are inherently difficult to apply to humans who, despite sharing a single gene disorder, have different genetic backgrounds and environmental exposures.

As a single gene cause of developmental disability with increasingly well-understood neurobiology, FXS

has been viewed as a disorder with relative homogeneity in particular in comparison to idiopathic ASD. This led to the idea that it would be easier to find consistent druggable targets in FXS whose modulation is associated with significant symptomatic improvement across a broad section of persons with the disorder compared to treatment development in behaviorally defined, etiologically diverse syndromes. Several factors have eroded the simplicity and accuracy of this approach. As trials have occurred in FXS, it has been clear that significant phenotypical differences exist in enrollees. This has likely led to several post-hoc comparison approaches looking at various subgroups, such as those with significant social withdrawal, ASD, or anxiety. Despite being caused by dysfunctional expression of a single gene, FXS presents with real heterogeneity, including such features as presence or absence of spoken language, comorbid ASD, epilepsy, significant ADHD symptoms, and, while anxiety is one of the most consistent features of the syndrome at least in males, the severity of anxiety may significantly differ between patients. This variability even within just the male *FMRI* full mutation population clearly confounds study development and outcome assessment. This is not to mention the significant variation between the presentation of FXS in males versus females and highly variable presentation within females alone given their random X chromosome inactivation patterns. Because of this variability of presentation, it is critical to find unifying features of the disorder that are readily and reproducibly measured. Thus, efforts to address this variability using quantitative biological or bio-behavioral parameters (i.e. biomarkers) through molecular blood assays, eye tracking technologies, or electrophysiology will be important to the field. These measures will in time need to be correlated with clinical measures to show relevance to daily functioning. Hopefully, these quantitative measures can define subgroups of individuals with consistent biological abnormalities correlated with some clinical measure, or alternatively prove useful in more directly tracking drug effects on functional brain systems. Such work would not only allow for potentially more effective outcome measures, but also will provide opportunity to identify at baseline certain persons with FXS who may best respond to a particular treatment. It is clear that parsing the heterogeneity of even a single gene disorder will likely be essential to future treatment development efforts.

As a protein responsible for translational regulation of hundreds of other proteins throughout the brain, FMRP has wide-reaching impact that likely cannot be constrained to a pathogenicity focused on a single neurotransmitter or other single-pharmacological approaches.

The complex sequelae of deficient FMRP represent a diverse set of known and likely many still unidentified molecular systems set into cellular dysregulation. This complexity may well render single drug approaches ineffective in the disorder. It is quite possible that combined pharmacotherapy approaches targeting a number of molecular or neural systems, all in a state of dysregulation, may be necessary to render significant clinical improvement in humans with FXS. Such approaches may mirror efforts in human immunodeficiency virus (HIV) and oncology work where combined treatments may be required to address disease.

Despite the many challenges facing the FXS translational drug development field, there are many recent developments that portend potential enhancement of future success. These include recent reports from Neuren Pharmaceuticals describing a positive phase II study result with trofinetide in older cohorts (adolescents and adults) with FXS, using a novel outcome measure analysis strategy that addresses the entire phenotype. Ongoing work to quantify EEG abnormalities across humans with FXS and the *Fmr1* KO mouse are underway in an attempt to bridge the translation treatment gap, thus enhancing the ability to predict [142] and monitor treatment response across species. Given that cognitive dysfunction is ubiquitous in FXS, treatments targeting the cognitive phenotype of the disease and development or validation of cognitive outcome measures are also important new directions for research. The FXS clinical trial field has undergone significant growth and development in recent years, thus creating a solid foundation to enhance trial fidelity across many locations. Investigators are using new models such as human induced pluripotent stem cell (iPSC) technology to develop new bench treatment assays that may aid in determining fundamental neuronal deficits in human FXS-derived cells and potentially better replicate pathology germane to the human FXS condition.

## Conclusions

Overall, the significant wave of FXS translational drug treatment development in the last decade has been marked with select improvement in FXS preclinical models that has not yet been extrapolated to human trial breakthroughs. Learning from the lessons of this experience will position the field well to move forward and enhance the opportunity for future success.

## Abbreviations

ABC: Aberrant Behavior Checklist-Community; ABC-I: ABC Irritability; ABC-SA: ABC Social Avoidance; ABC-SW: Aberrant Behavior Checklist Social Withdrawal subscale; ADHD: Attention deficit hyperactivity disorder; ADHD-RS: Attention deficit hyperactivity disorder rating scale; ADHD-RS-IV: ADHD Rating Scale 4th Edition; ALS: Amyotrophic lateral sclerosis; AMPAR:  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid glutamate receptor; APP: Amyloid precursor protein; ASD: Autism spectrum disorder; BDNF: Brain-derived neurotrophic factor; CARS: Childhood autism rating scale; CGG: Cytosine guanine guanine; CNT: Contingency Naming Test; ERK; ERK1/2: Extracellular signal-related kinase; ERPs: Event-related potentials; FDA: Food

and Drug Administration; *FMR1*: Fragile X mental retardation gene; *FMRP*: Fragile X mental retardation protein; *FXS*: Fragile X syndrome; *GABA*:  $\gamma$ -Aminobutyric acid; *GSK-3 $\beta$* : Glycogen synthase kinase-3 beta; *HIV*: Human immunodeficiency virus; *IGF-1*: Insulin-like growth factor 1; *iPSC*: Induced pluripotent stem cell; *ITT*: Intent-to-treat; *KiTAP*: Test of attentional performance for children; *KO*: Knockout; *LTD*: Long-term depression; *LTP*: Long-term potentiation; *MDX*: Metadoxine ER; *mGluRs*: Group I metabotropic glutamate receptors; *MMP-9*: Matrix metalloproteinase 9; *MSEL*: Mullen Scales of Early Learning; *mTOR*: Mammalian target of rapamycin; *NMDA*: N-methyl-D-aspartate; *PET*: Positron emission tomography; *PPI*: Prepulse inhibition; *RBANS*: Repeatable Battery for the Assessment of Neuropsychological Status; *SRS*: Social responsiveness scale; *SSRIs*: Selective serotonin reuptake inhibitors; *TBI*: Traumatic brain injury; *UTR*: Untranslated region; *VAS*: Visual analog scale; *Vineland-II*: Vineland adaptive behavior scale

#### Acknowledgements

Not applicable.

#### Funding

NIH grant U54HD082008 (Drs. Erickson, Sweeney, Schaefer, Wink and Pedapati) supported this publication.

#### Availability of data and materials

Not Applicable.

#### Authors' contributions

All authors contributed their expertise to various sections, edited, and approved the final manuscript.

#### Competing interests

Dr. Erickson has received current/past support from Alcobra Pharmaceuticals, the American Academy of Child & Adolescent Psychiatry, Autism Speaks, Cincinnati Children's Hospital Medical Center, FRAXA Research Foundation, Hoffmann-La Roche Inc., Neuren Pharmaceuticals Limited, Simons Foundation Autism Research Initiative, Stemina Biomarker Discovery, Inc., SynapDx, The John Merck Fund, the National Fragile X Foundation, the National Institutes of Health, and the US Department of Defense. Dr. Erickson is a past consultant to Alcobra, Neurotrope, The Roche Group, Novartis, and a current consultant to Confluence Pharmaceuticals and Fulcrum Therapeutics. Additionally, Dr. Erickson is an equity holder in Confluence Pharmaceuticals, a company working to reformulate acamprostate for commercialization purposes in the field of developmental disorders. Drs. Wink and Dr. Pedapati receive research support from Children's Hospital Research Foundation and the American Academy of Child and Adolescent Psychiatry. Dr. Schaefer has received past support from Confluence Pharmaceuticals and FRAXA Research Foundation. Dr. Sweeney has consulted to Takeda Pharmaceuticals. Ms. Fitzpatrick and Mr. Davenport have no conflict of interests. Dr. Brown receives current support from the Center for Disease Control and has received past support from Hoffmann-La Roche Inc., Novartis, SynapDx, and the National Institutes of Health. Dr. Budimirovic has received support for clinical trials for FXS from Seaside Therapeutics and has done consulting work or received research funding from Asuragen Inc., the American Academy of Child & Adolescent Psychiatry, Ironshore, MEDACorp, and Sunovion. Dr. Randi Hagerman has received support from Neuren, Alcobra, Roche/Genentech, Novartis, Marinus, the Department of Defense, HRSA, and the National Fragile X Foundation to carry out trials in FXS. Dr. Hagerman has also consulted with Ovid, Zynerba, and Alcobra regarding trials in FXS. Dr. Hessler has received past support from Seaside Therapeutics, Novartis, Hoffmann-La Roche, Inc., and The John Merck Fund for consulting and clinical trials work in FXS. Dr. Kaufmann is a consultant for Neuren, Newron, Elox, EryDel, Marinus, Edison, and GW Pharmaceuticals. He has received past support from Seaside Therapeutics, Novartis, Hoffmann-La Roche, and Ipsen. Dr. Berry-Kravis has received current/past support for clinical trials or research from Alcobra Pharmaceuticals, Asuragen, FRAXA Research Foundation, Hoffmann-La Roche Inc., Neuren Pharmaceuticals Limited, Novartis, Simons Foundation Autism Research Initiative, SynapDx, The John Merck Fund, the National Fragile X Foundation, the National Institutes of Health, National Center for Advancement of Translational Science, and the Centers for Disease Control. Dr. Berry-Kravis is a past consultant to Neurotrope, Roche, Asuragen, Novartis, and a current consultant to Alcobra, Neuren, and Fulcrum Therapeutics.

#### Consent for publication

All authors consent to publish this manuscript.

#### Ethics approval and consent to participate

Not Applicable.

#### Author details

<sup>1</sup>Division of Child and Adolescent Psychiatry (MLC 4002), Cincinnati Children's Hospital Medical Center, 3333 Burnet Ave., Cincinnati, OH 45229-3039, USA. <sup>2</sup>Department of Psychiatry, College of Medicine, University of Cincinnati, Cincinnati, OH, USA. <sup>3</sup>Department of Biomedical Engineering, College of Engineering and Applied Science, University of Cincinnati, Cincinnati, OH, USA. <sup>4</sup>Medical Investigation of Neurodevelopmental Disorders (MIND) Institute, Davis Medical Center, University of California, Sacramento, CA, USA. <sup>5</sup>Department of Pediatrics, Davis Medical Center, University of California, Sacramento, California, USA. <sup>6</sup>Department of Psychiatry and Behavioral Sciences, Davis Medical Center, University of California, Sacramento, California, USA. <sup>7</sup>Greenwood Genetic Center, Greenwood, SC, USA. <sup>8</sup>Boston Children's Hospital, Boston, Massachusetts, USA. <sup>9</sup>Departments of Pediatrics, Neurological Sciences, Biochemistry, Rush University Medical Center, Chicago, Illinois, USA. <sup>10</sup>Institute for Basic Research in Developmental Disabilities, New York, NY, USA. <sup>11</sup>Clinical Research Center, Clinical Trials Unit, Fragile X Clinic, Kennedy Krieger Institute, The Johns Hopkins Medical Institutions, Baltimore, MD, USA. <sup>12</sup>Departments of Psychiatry & Behavioral Sciences, Child Psychiatry, The Johns Hopkins Medical Institutions, Baltimore, MD, USA.

Received: 13 July 2016 Accepted: 18 January 2017

Published online: 12 June 2017

#### References

- Turner G, Webb T, Wake S, Robinson H. Prevalence of fragile X syndrome. *Am J Med Genet*. 1996;64(1):196–7.
- Song FJ, Barton P, Sleightholme V, Yao GL, Fry-Smith A. Screening for fragile X syndrome: a literature review and modelling study. *Health Technol Assess*. 2003;7(16):1–106.
- Bailey Jr DB, Raspa M, Olmsted M, Holiday DB. Co-occurring conditions associated with FMR1 gene variations: findings from a national parent survey. *Am J Med Genet A*. 2008;146A(16):2060–9.
- Hagerman RJ, Berry-Kravis E, Kaufmann WE, Ono MY, Tartaglia N, Lachiewicz A, Kronk R, Delahunty C, Hessler D, Visootsak J, et al. Advances in the treatment of fragile X syndrome. *Pediatrics*. 2009;123(1):378–90.
- Budimirovic DB, Kaufmann WE. What can we learn about autism from studying fragile X syndrome? *Dev Neurosci*. 2011;33(5):379–94.
- Fu YH, Kuhl DP, Pizzuti A, Pieretti M, Sutcliffe JS, Richards S, Verkerk AJ, Holden JJ, Fenwick Jr RG, Warren ST, et al. Variation of the CGG repeat at the fragile X site results in genetic instability: resolution of the Sherman paradox. *Cell*. 1991;67(6):1047–58.
- Pieretti M, Zhang FP, Fu YH, Warren ST, Oostra BA, Caskey CT, Nelson DL. Absence of expression of the FMR-1 gene in fragile X syndrome. *Cell*. 1991;66(4):817–22.
- Bear MF, Huber KM, Warren ST. The mGluR theory of fragile X mental retardation. *Trends Neurosci*. 2004;27(7):370–7.
- Lozano R, Rosero CA, Hagerman RJ. Fragile X spectrum disorders. *Intractable Rare Dis Res*. 2014;3(4):134–46.
- Kidd SA, Lachiewicz A, Barbouth D, Blitz RK, Delahunty C, McBrien D, Visootsak J, Berry-Kravis E. Fragile X syndrome: a review of associated medical problems. *Pediatrics*. 2014;134(5):995–1005.
- Garber KB, Visootsak J, Warren ST. Fragile X syndrome. *Eur J Hum Genet*. 2008;16(6):666–72.
- Tsiouris JA, Brown WT. Neuropsychiatric symptoms of fragile X syndrome: pathophysiology and pharmacotherapy. *CNS Drugs*. 2004;18(11):687–703.
- Kaufmann WE, Cortell R, Kau AS, Bukelis I, Tierney E, Gray RM, Cox C, Capone GT, Stanard P. Autism spectrum disorder in fragile X syndrome: communication, social interaction, and specific behaviors. *Am J Med Genet A*. 2004;129A(3):225–34.
- Budimirovic DB, Bukelis I, Cox C, Gray RM, Tierney E, Kaufmann WE. Autism spectrum disorder in Fragile X syndrome: differential contribution of adaptive socialization and social withdrawal. *Am J Med Genet A*. 2006;140A(17):1814–26.



15. Gholizadeh S, Halder SK, Hampson DR. Expression of fragile X mental retardation protein in neurons and glia of the developing and adult mouse brain. *Brain Res.* 2015;1596:22–30.
16. Weiler IJ, Irwin SA, Klintsova AY, Spencer CM, Brazelton AD, Miyashiro K, Comery TA, Patel B, Eberwine J, Greenough WT. Fragile X mental retardation protein is translated near synapses in response to neurotransmitter activation. *Proc Natl Acad Sci U S A.* 1997;94(10):5395–400.
17. Napoli I, Mercaldo V, Boyl PP, Eleuteri B, Zalfa F, De Rubeis S, Di Marino D, Mohr E, Massimi M, Falconi M, et al. The fragile X syndrome protein represses activity-dependent translation through CYFIP1, a new 4E-BP. *Cell.* 2008;134(6):1042–54.
18. Corbin F, Bouillon M, Fortin A, Morin S, Rousseau F, Khandjian EW. The fragile X mental retardation protein is associated with poly(A)+ mRNA in actively translating polyribosomes. *Hum Mol Genet.* 1997;6(9):1465–72.
19. Feng Y, Absher D, Eberhart DE, Brown V, Malter HE, Warren ST. FMRP associates with polyribosomes as an mRNP, and the I304N mutation of severe fragile X syndrome abolishes this association. *Mol Cell.* 1997;1(1):109–18.
20. Santoro MR, Bray SM, Warren ST. Molecular mechanisms of fragile X syndrome: a twenty-year perspective. *Annu Rev Pathol.* 2012;7:219–45.
21. Sethna F, Moon C, Wang H. From FMRP function to potential therapies for fragile X syndrome. *Neurochem Res.* 2014;39(6):1016–31.
22. Hazlett HC, Poe MD, Lightbody AA, Styner M, MacFall JR, Reiss AL, Piven J. Trajectories of early brain volume development in fragile X syndrome and autism. *J Am Acad Child Adolesc Psychiatry.* 2012;51(9):921–33.
23. Rudelli RD, Brown WT, Wisniewski K, Jenkins EC, Laure-Kamionowska M, Connell F, Wisniewski HM. Adult fragile X syndrome. *Clinico-neuropathologic findings.* *Acta Neuropathol.* 1985;67(3-4):289–95.
24. Hinton VJ, Brown WT, Wisniewski K, Rudelli RD. Analysis of neocortex in three males with the fragile X syndrome. *Am J Med Genet.* 1991;41(3):289–94.
25. Comery TA, Harris JB, Willems PJ, Oostra BA, Irwin SA, Weiler IJ, Greenough WT. Abnormal dendritic spines in fragile X knockout mice: maturation and pruning deficits. *Proc Natl Acad Sci U S A.* 1997;94(10):5401–4.
26. Huber KM, Gallagher SM, Warren ST, Bear MF. Altered synaptic plasticity in a mouse model of fragile X mental retardation. *Proc Natl Acad Sci U S A.* 2002;99(11):7746–50.
27. Bhattacharya A, Kaphzan H, Alvarez-Dieppa AC, Murphy JP, Pierre P, Klann E. Genetic removal of p70 S6 kinase 1 corrects molecular, synaptic, and behavioral phenotypes in fragile X syndrome mice. *Neuron.* 2012;76(2):325–37.
28. Hou L, Antion MD, Hu D, Spencer CM, Paylor R, Klann E. Dynamic translational and proteasomal regulation of fragile X mental retardation protein controls mGluR-dependent long-term depression. *Neuron.* 2006; 51(4):441–54.
29. Michalon A, Sidorov M, Ballard TM, Ozmen L, Spooren W, Wettstein JG, Jaeschke G, Bear MF, Lindemann L. Chronic pharmacological mGlu5 inhibition corrects fragile X in adult mice. *Neuron.* 2012;74(1):49–56.
30. Auerbach BD, Bear MF. Loss of the fragile X mental retardation protein decouples metabotropic glutamate receptor dependent priming of long-term potentiation from protein synthesis. *J Neurophysiol.* 2010; 104(2):1047–51.
31. Zhao MG, Toyoda H, Ko SW, Ding HK, Wu LJ, Zhuo M. Deficits in trace fear memory and long-term potentiation in a mouse model for fragile X syndrome. *J Neurosci.* 2005;25(32):7385–92.
32. Yun SH, Trommer BL. Fragile X mice: reduced long-term potentiation and N-methyl-D-aspartate receptor-mediated neurotransmission in dentate gyrus. *J Neurosci Res.* 2011;89(2):176–82.
33. Martin HG, Lassalle O, Brown JT, Manzoni OJ. Age-Dependent Long-Term Potentiation Deficits in the Prefrontal Cortex of the Fmr1 Knockout Mouse Model of Fragile X Syndrome. *Cereb Cortex.* 2016;26(5):2084–92.
34. Gross C, Chang CW, Kelly SM, Bhattacharya A, McBride SM, Danielson SW, Jiang MQ, Chan CB, Ye K, Gibson JR, et al. Increased expression of the PI3K enhancer PIKE mediates deficits in synaptic plasticity and behavior in fragile X syndrome. *Cell Rep.* 2015;11(5):727–36.
35. Gross C, Raj N, Molinaro G, Allen AG, Whyte AJ, Gibson JR, Huber KM, Gourley SL, Bassell GJ. Selective role of the catalytic PI3K subunit p110beta in impaired higher order cognition in fragile X syndrome. *Cell Rep.* 2015;11(5):681–8.
36. Gross C, Nakamoto M, Yao X, Chan CB, Yim SY, Ye K, Warren ST, Bassell GJ. Excess phosphoinositide 3-kinase subunit synthesis and activity as a novel therapeutic target in fragile X syndrome. *J Neurosci.* 2010;30(32):10624–38.
37. Matic K, Eninger T, Bardoni B, Davidovic L, Macek B. Quantitative phosphoproteomics of murine Fmr1-KO cell lines provides new insights into FMRP-dependent signal transduction mechanisms. *J Proteome Res.* 2014;13(10):4388–97.
38. Curia G, Gualtieri F, Bartolomeo R, Vezzali R, Biagini G. Resilience to audiogenic seizures is associated with p-ERK1/2 dephosphorylation in the subiculum of Fmr1 knockout mice. *Front Cell Neurosci.* 2013;7:46.
39. Kim SH, Markham JA, Weiler IJ, Greenough WT. Aberrant early-phase ERK inactivation impedes neuronal function in fragile X syndrome. *Proc Natl Acad Sci U S A.* 2008;105(11):4429–34.
40. Bilousova TV, Dansie L, Ngo M, Aye J, Charles JR, Ethell DW, Ethell IM. Minocycline promotes dendritic spine maturation and improves behavioural performance in the fragile X mouse model. *J Med Genet.* 2009;46(2):94–102.
41. Dansie LE, Phommahaxay K, Okusanya AG, Uwadia J, Huang M, Rotschafer SE, Razak KA, Ethell DW, Ethell IM. Long-lasting effects of minocycline on behavior in young but not adult Fragile X mice. *Neuroscience.* 2013;246:186–98.
42. Qin M, Zeidler Z, Moulton K, Krych L, Xia Z, Smith CB. Endocannabinoid-mediated improvement on a test of aversive memory in a mouse model of fragile X syndrome. *Behav Brain Res.* 2015;291:164–71.
43. Tang AH, Alger BE. Homer protein-metabotropic glutamate receptor binding regulates endocannabinoid signaling and affects hyperexcitability in a mouse model of fragile X syndrome. *J Neurosci.* 2015;35(9):3938–45.
44. Jung KM, Sepers M, Henstridge CM, Lassalle O, Neuhofer D, Martin H, Ginger M, Frick A, DiPatrizio NV, Mackie K, et al. Uncoupling of the endocannabinoid signalling complex in a mouse model of fragile X syndrome. *Nat Commun.* 2012;3:1080.
45. Zhang L, Alger BE. Enhanced endocannabinoid signaling elevates neuronal excitability in fragile X syndrome. *J Neurosci.* 2010;30(16):5724–9.
46. Kim SW, Cho KJ. Activity-dependent alterations in the sensitivity to BDNF-TrkB signaling may promote excessive dendritic arborization and spinogenesis in fragile X syndrome in order to compensate for compromised postsynaptic activity. *Med Hypotheses.* 2014;83(4):429–35.
47. Castren M, Lampinen KE, Miettinen R, Koponen E, Sipola I, Bakker CE, Oostra BA, Castren E. BDNF regulates the expression of fragile X mental retardation protein mRNA in the hippocampus. *Neurobiol Dis.* 2002;11(1):221–9.
48. Sharma A, Hoefler CA, Takayasu Y, Miyawaki T, McBride SM, Klann E, Zukin RS. Dysregulation of mTOR signaling in fragile X syndrome. *J Neurosci.* 2010;30(2):694–702.
49. Price TJ, Rashid MH, Millicamps M, Sanoja R, Entrena JM, Cervero F. Decreased nociceptive sensitization in mice lacking the fragile X mental retardation protein: role of mGluR1/5 and mTOR. *J Neurosci.* 2007;27(51):13958–67.
50. Amiri A, Sanchez-Ortiz E, Cho W, Birnbaum SG, Xu J, McKay RM, Parada LF. Analysis of FMR1 deletion in a subpopulation of post-mitotic neurons in mouse cortex and hippocampus. *Autism Res.* 2014;7(1):60–71.
51. Davenport MH, Schaefer TL, Friedmann KJ, Fitzpatrick SE, Erickson CA. Pharmacotherapy for Fragile X syndrome: progress to date. *Drugs.* 2016; 76(4):431–45.
52. Schaefer TL, Davenport MH, Erickson CA. Emerging pharmacologic treatment options for fragile X syndrome. *Appl Clin Genet.* 2015;8:75–93.
53. Brown V, Jin P, Ceman S, Darnell JC, O'Donnell WT, Tenenbaum SA, Jin X, Feng Y, Wilkinson KD, Keene JD, et al. Microarray identification of FMRP-associated brain mRNAs and altered mRNA translational profiles in fragile X syndrome. *Cell.* 2001;107(4):477–87.
54. Chuang SC, Zhao W, Bauchwitz R, Yan Q, Bianchi R, Wong RK. Prolonged epileptiform discharges induced by altered group I metabotropic glutamate receptor-mediated synaptic responses in hippocampal slices of a fragile X mouse model. *J Neurosci.* 2005;25(35):8048–55.
55. Karachot L, Shirai Y, Vigot R, Yamamori T, Ito M. Induction of long-term depression in cerebellar Purkinje cells requires a rapidly turned over protein. *J Neurophysiol.* 2001;86(1):280–9.
56. Zho WM, You JL, Huang CC, Hsu KS. The group I metabotropic glutamate receptor agonist (S)-3,5-dihydroxyphenylglycine induces a novel form of depotentiation in the CA1 region of the hippocampus. *J Neurosci.* 2002; 22(20):8838–49.
57. Huber KM, Kayser MS, Bear MF. Role for rapid dendritic protein synthesis in hippocampal mGluR-dependent long-term depression. *Science.* 2000; 288(5469):1254–7.
58. Raymond CR, Thompson VL, Tate WP, Abraham WC. Metabotropic glutamate receptors trigger homosynaptic protein synthesis to prolong long-term potentiation. *J Neurosci.* 2000;20(3):969–76.
59. Yan QJ, Rammal M, Tranfaglia M, Bauchwitz RP. Suppression of two major Fragile X syndrome mouse model phenotypes by the mGluR5 antagonist MPEP. *Neuropharmacology.* 2005;49(7):1053–66.

60. de Vrij FM, Levens J, van der Linde HC, Koekoek SK, De Zeeuw CI, Nelson DL, Oostra BA, Willemsen R. Rescue of behavioral phenotype and neuronal protrusion morphology in *Fmr1* KO mice. *Neurobiol Dis*. 2008;31(1):127–32.
61. Lindemann L, Jaeschke G, Michalon A, Vieira E, Honer M, Spooren W, Porter R, Hartung T, Kolczewski S, Buttelmann B, et al. CTEP: a novel, potent, long-acting, and orally bioavailable metabotropic glutamate receptor 5 inhibitor. *J Pharmacol Exp Ther*. 2011;339(2):474–86.
62. Dolen G, Osterweil E, Rao BS, Smith GB, Auerbach BD, Chattarji S, Bear MF. Correction of fragile X syndrome in mice. *Neuron*. 2007;56(6):955–62.
63. Thomas AM, Bui N, Graham D, Perkins JR, Yuva-Paylor LA, Paylor R. Genetic reduction of group 1 metabotropic glutamate receptors alters select behaviors in a mouse model for fragile X syndrome. *Behav Brain Res*. 2011;223(2):310–21.
64. Olmos-Serrano JL, Paluszkiwicz SM, Martin BS, Kaufmann WE, Corbin JG, Huntsman MM. Defective GABAergic neurotransmission and pharmacological rescue of neuronal hyperexcitability in the amygdala in a mouse model of fragile X syndrome. *J Neurosci*. 2010;30(29):9929–38.
65. El Idrissi A, Yan X, L'Amoreaux W, Brown WT, Dobkin C. Neuroendocrine alterations in the fragile X mouse. *Results Probl Cell Differ*. 2012;54:201–21.
66. D'Hulst C, Heulens I, Brouwer JR, Willemsen R, De Geest N, Reeve SP, De Deyn PP, Hassan BA, Kooy RF. Expression of the GABAergic system in animal models for fragile X syndrome and fragile X associated tremor/ataxia syndrome (FXTAS). *Brain Res*. 2009;1253:176–83.
67. D'Hulst C, De Geest N, Reeve SP, Van Dam D, De Deyn PP, Hassan BA, Kooy RF. Decreased expression of the GABA<sub>A</sub> receptor in fragile X syndrome. *Brain Res*. 2006;1121(1):238–45.
68. Heulens I, D'Hulst C, Van Dam D, De Deyn PP, Kooy RF. Pharmacological treatment of fragile X syndrome with GABAergic drugs in a knockout mouse model. *Behav Brain Res*. 2012;229(1):244–9.
69. Chang S, Bray SM, Li Z, Zarnescu DC, He C, Jin P, Warren ST. Identification of small molecules rescuing fragile X syndrome phenotypes in *Drosophila*. *Nat Chem Biol*. 2008;4(4):256–63.
70. Henderson C, Wijetunge L, Kinoshita MN, Shumway M, Hammond RS, Postma FR, Brynczka C, Rush R, Thomas A, Paylor R, et al. Reversal of disease-related pathologies in the Fragile X mouse model by selective activation of GABA<sub>B</sub> receptors with arbaclofen. *Sci Transl Med*. 2012;4(152):152ra128.
71. Pacey LK, Heximer SP, Hampson DR. Increased GABA(B) receptor-mediated signaling reduces the susceptibility of fragile X knockout mice to audiogenic seizures. *Mol Pharmacol*. 2009;76(1):18–24.
72. Porter RH, Jaeschke G, Spooren W, Ballard TM, Buttelmann B, Kolczewski S, Peters JU, Prinsens E, Wichmann J, Vieira E, et al. Fenobam: a clinically validated nonbenzodiazepine anxiolytic is a potent, selective, and noncompetitive mGlu5 receptor antagonist with inverse agonist activity. *J Pharmacol Exp Ther*. 2005;315(2):711–21.
73. Berry-Kravis E, Hessel D, Coffey S, Hervey C, Schneider A, Yuhas J, Hutchison J, Snape M, Tranfaglia M, Nguyen DV, et al. A pilot open label, single dose trial of fenobam in adults with fragile X syndrome. *J Med Genet*. 2009;46(4):266–71.
74. Friedmann CTH, Davis LJ, Ciccone PE, Rubin RT. Phase-II double-blind controlled-study of a new anxiolytic, fenobam (Mcn-3377) vs placebo. *Curr Ther Res Clin E*. 1980;27(2):144–51.
75. Jacquemont S, Curie A, des Portes V, Torrioli MG, Berry-Kravis E, Hagerman RJ, Ramos FJ, Cornish K, He Y, Paulding C, et al. Epigenetic modification of the *FMR1* gene in fragile X syndrome is associated with differential response to the mGluR5 antagonist AFQ056. *Sci Transl Med*. 2011;3(64):64ra61.
76. Berry-Kravis E, Des Portes V, Hagerman R, Jacquemont S, Charles P, Visootsak J, Brinkman M, Rerat K, Koumaras B, Zhu L, et al. Mavoglurant in fragile X syndrome: results of two randomized, double-blind, placebo-controlled trials. *Sci Transl Med*. 2016;8(321):321ra325.
77. Sansone SM, Widaman KF, Hall SS, Reiss AL, Lightbody A, Kaufmann WE, Berry-Kravis E, Lachiewicz A, Brown EC, Hessel D. Psychometric study of the aberrant behavior checklist in fragile x syndrome and implications for targeted treatment. *J Autism Dev Disord*. 2012;42(7):1377–92.
78. Aman MG, Singh NN, Stewart AW, Field CJ. The Aberrant Behavior Checklist: a behavior rating scale for the assessment of treatment effects. *Am J Ment Defic*. 1985;5:485–91.
79. Novartis discontinues development of mavoglurant (AFQ056) for Fragile X syndrome. <http://www.fraxa.org/novartis-discontinues-development-mavoglurant-afq056-fragile-x-syndrome/>.
80. Roche Letter. <http://www.fiercebitech.com/r-d/roche-abandons-another-fragile-x-r-d-program-after-phii-trials-flunk-out>.
81. Hu H, Qin Y, Bochorishvili G, Zhu Y, van Aelst L, Zhu JJ. Ras signaling mechanisms underlying impaired GluR1-dependent plasticity associated with fragile X syndrome. *J Neurosci*. 2008;28(31):7847–62.
82. Pilpel Y, Kollekter A, Berberich S, Ginger M, Frick A, Mientjes E, Oostra BA, Seeburg PH. Synaptic ionotropic glutamate receptors and plasticity are developmentally altered in the CA1 field of *Fmr1* knockout mice. *J Physiol*. 2009;587(Pt 4):787–804.
83. Eadie BD, Cushman J, Kannagara TS, Fanselow MS, Christie BR. NMDA receptor hypofunction in the dentate gyrus and impaired context discrimination in adult *Fmr1* knockout mice. *Hippocampus*. 2012;22(2):241–54.
84. Erickson CA, Mullett JE, McDougle CJ. Open-label memantine in fragile X syndrome. *J Autism Dev Disord*. 2009;39(12):1629–35.
85. Berry-Kravis E, Krause SE, Block SS, Guter S, Wu J, Leurgans S, Declé P, Potanos K, Cook E, Salt J, et al. Effect of CX516, an AMPA-modulating compound, on cognition and behavior in fragile X syndrome: a controlled trial. *J Child Adolesc Psychopharmacol*. 2006;16(5):525–40.
86. Gallagher SM, Daly CA, Bear MF, Huber KM. Extracellular signal-regulated protein kinase activation is required for metabotropic glutamate receptor-dependent long-term depression in hippocampal area CA1. *J Neurosci*. 2004;24(20):4859–64.
87. Wang X, Snape M, Klann E, Stone JG, Singh A, Petersen RB, Castellani RJ, Casadesus G, Smith MA, Zhu X. Activation of the extracellular signal-regulated kinase pathway contributes to the behavioral deficit of fragile x-syndrome. *J Neurochem*. 2012;121(4):672–9.
88. Osterweil EK, Krueger DD, Reinhold K, Bear MF. Hypersensitivity to mGluR5 and ERK1/2 leads to excessive protein synthesis in the hippocampus of a mouse model of fragile X syndrome. *J Neurosci*. 2010;30(46):15616–27.
89. Zarate Jr CA, Payne JL, Quiroz J, Sporn J, Denicoff KK, Luckenbaugh D, Charney DS, Manji HK. An open-label trial of riluzole in patients with treatment-resistant major depression. *Am J Psychiatry*. 2004;161(1):171–4.
90. Grant P, Lougee L, Hirschtritt M, Swedo SE. An open-label trial of riluzole, a glutamate antagonist, in children with treatment-resistant obsessive-compulsive disorder. *J Child Adolesc Psychopharmacol*. 2007;17(6):761–7.
91. Martin D, Thompson MA, Nadler JV. The neuroprotective agent riluzole inhibits release of glutamate and aspartate from slices of hippocampal area CA1. *Eur J Pharmacol*. 1993;250(3):473–6.
92. Jahn K, Schlesinger F, Jin LJ, Dengler R, Bufler J, Krampf K. Molecular mechanisms of interaction between the neuroprotective substance riluzole and GABA(A)-receptors. *Naunyn Schmiedeberg Arch Pharmacol*. 2008;378(1):53–63.
93. Erickson CA, Weng N, Weiler IJ, Greenough WT, Stigler KA, Wink LK, McDougle CJ. Open-label riluzole in fragile X syndrome. *Brain Res*. 2011;1380:264–70.
94. Weiler IJ, Spangler CC, Klintsova AY, Grossman AW, Kim SH, Bertaina-Anglade V, Khalil H, de Vries FE, Lambers FA, Hatia F, et al. Fragile X mental retardation protein is necessary for neurotransmitter-activated protein translation at synapses. *Proc Natl Acad Sci U S A*. 2004;101(50):17504–9.
95. Boismare F, Daoust M, Moore N, Saligaut C, Lhuintre JP, Chretien P, Durlach J. A homotaurine derivative reduces the voluntary intake of ethanol by rats: are cerebral GABA receptors involved? *Pharmacol Biochem Behav*. 1984;21(5):787–9.
96. Harris BR, Prendergast MA, Gibson DA, Rogers DT, Blanchard JA, Holley RC, Fu MC, Hart SR, Pedigo NW, Littleton JM. Acamprosate inhibits the binding and neurotoxic effects of trans-ACPD, suggesting a novel site of action at metabotropic glutamate receptors. *Alcohol Clin Exp Res*. 2002;26(12):1779–93.
97. Mann K, Kiefer F, Spanagel R, Littleton J. Acamprosate: recent findings and future research directions. *Alcohol Clin Exp Res*. 2008;32(7):1105–10.
98. Erickson CA, Mullett JE, McDougle CJ. Brief report: acamprosate in fragile X syndrome. *J Autism Dev Disord*. 2010;40(11):1412–6.
99. Erickson CA, Wink LK, Ray B, Early MC, Stiegelmeier E, Mathieu-Frasier L, Patrick V, Lahiri DK, McDougle CJ. Impact of acamprosate on behavior and brain-derived neurotrophic factor: an open-label study in youth with fragile X syndrome. *Psychopharmacology (Berl)*. 2013;228(1):75–84.
100. Erickson C, Ray B, Maloney B, Wink LK, Bowers K, Schaefer JL, McDougle C, Sokol DK, Lahiri D. Impact of acamprosate on plasma amyloid- $\beta$  precursor protein in youth: a pilot analysis in fragile X syndrome-associated and idiopathic autism spectrum disorder suggests a pharmacodynamic protein marker. *J Psychiatr Res*. 2014, in press.
101. Braat S, D'Hulst C, Heulens I, De Rubeis S, Mientjes E, Nelson DL, Willemsen R, Bagni C, Van Dam D, De Deyn PP, et al. The GABA<sub>A</sub> receptor is an FMRP target with therapeutic potential in fragile X syndrome. *Cell Cycle*. 2015;14(18):2985–95.

102. Berry-Kravis EM, Hessel D, Rathmell B, Zarevics P, Cherubini M, Walton-Bowen K, Mu Y, Nguyen DV, Gonzalez-Heydrich J, Wang PP, et al. Effects of STX209 (Arbaclofen) on Neurobehavioral Function in Children and Adults with Fragile X Syndrome: A Randomized, Controlled, Phase 2 Trial. *Sci Transl Med*. 2012;4(152):152ra127.
103. Fornai F, Grazia Alessandri M, Bonuccelli U, Scali V, Corsini GU. Effect of metadoxine on striatal dopamine levels in C57 black mice. *J Pharm Pharmacol*. 1993;45(5):476–8.
104. Alcobra announces results from phase 2 clinical trial of MDX for fragile X syndrome. <http://www.alcobra-pharma.com/releasedetail.cfm?ReleaseID=919218>.
105. Bailey Jr DB, Berry-Kravis E, Wheeler A, Raspa M, Merrier F, Ricart J, Koumaras B, Rosenkranz G, Tomlinson M, von Raison F, et al. Mavoglurant in adolescents with fragile X syndrome: analysis of Clinical Global Impression-Improvement source data from a double-blind therapeutic study followed by an open-label, long-term extension study. *J Neurodev Disord*. 2016;8:1.
106. Jope RS, Roh MS. Glycogen synthase kinase-3 (GSK3) in psychiatric diseases and therapeutic interventions. *Curr Drug Targets*. 2006;7(11):1421–34.
107. Portis S, Giunta B, Obregon D, Tan J. The role of glycogen synthase kinase-3 signaling in neurodevelopment and fragile X syndrome. *Int J Physiol Pathophysiol Pharmacol*. 2012;4(3):140–8.
108. Liu ZH, Chuang DM, Smith CB. Lithium ameliorates phenotypic deficits in a mouse model of fragile X syndrome. *Int J Neuropsychopharmacol*. 2011;14(5):618–30.
109. Liu ZH, Huang T, Smith CB. Lithium reverses increased rates of cerebral protein synthesis in a mouse model of fragile X syndrome. *Neurobiol Dis*. 2012;45(3):1145–52.
110. Choi CH, Schoenfeld BP, Bell AJ, Hinchey P, Kollaras M, Gertner MJ, Woo NH, Tranfaglia MR, Bear MF, Zukin RS, et al. Pharmacological reversal of synaptic plasticity deficits in the mouse model of Fragile X syndrome by group II mGluR antagonist or lithium treatment. *Brain Res*. 2011;1380:106–19.
111. Yuskaitis CJ, Mines MA, King MK, Sweatt JD, Miller CA, Jope RS. Lithium ameliorates altered glycogen synthase kinase-3 and behavior in a mouse model of fragile X syndrome. *Biochem Pharmacol*. 2010;79(4):632–46.
112. Franklin AV, King MK, Palomo V, Martinez A, McMahon LL, Jope RS. Glycogen synthase kinase-3 inhibitors reverse deficits in long-term potentiation and cognition in fragile X mice. *Biol Psychiatry*. 2014;75(3):198–206.
113. Berry-Kravis E, Sumis A, Nelson M, Porges SW, Weng N, Weiler IJ, Greenough WT. Open-label treatment trial of lithium to target the underlying defect in fragile X syndrome. *J Dev Behav Pediatr*. 2008;29(4):293–302.
114. Bilousova TV, Rusakov DA, Ethell DW, Ethell IM. Matrix metalloproteinase-7 disrupts dendritic spines in hippocampal neurons through NMDA receptor activation. *J Neurochem*. 2006;97(1):44–56.
115. Bilousova T, Dansie L, Ngo M, Aye J, Charles JR, Ethell DW, Ethell IM: Minocycline promotes dendritic spine maturation and improves behavioral performance in the fragile X mouse model. *J Med Genet*. 2009;46(2):94–102.
116. Paribello C, Tao L, Folino A, Berry-Kravis E, Tranfaglia M, Ethell IM, Ethell DW. Open-label add-on treatment trial of minocycline in fragile X syndrome. *BMC Neurol*. 2010;10:91.
117. Leigh MJ, Nguyen DV, Mu Y, Winarni TI, Schneider A, Chechi T, Polussa J, Doucet P, Tassone F, Rivera SM, et al. A randomized double-blind, placebo-controlled trial of minocycline in children and adolescents with fragile X syndrome. *J Dev Behav Pediatr*. 2013;34(3):147–55.
118. Schneider A, Leigh MJ, Adams P, Nanakul R, Chechi T, Olichney J, Hagerman R, Hessel D. Electrocardiographic changes associated with minocycline treatment in fragile X syndrome. *J Psychopharmacol*. 2013;27(10):956–63.
119. Dziembowska M, Pretto DI, Janusz A, Kaczmarek L, Leigh MJ, Gabriel N, Durbin-Johnson B, Hagerman RJ, Tassone F. High MMP-9 activity levels in fragile X syndrome are lowered by minocycline. *Am J Med Genet A*. 2013;161(8):1897–903.
120. Descamps OS, Tenoutasse S, Stephenne X, Gies I, Beauloye V, Lebrethon MC, De Beaufort C, De Waele K, Scheen A, Rietzschel E, et al. Management of familial hypercholesterolemia in children and young adults: consensus paper developed by a panel of lipidologists, cardiologists, paediatricians, nutritionists, gastroenterologists, general practitioners and a patient organization. *Atherosclerosis*. 2011;218(2):272–80.
121. Cerezo-Guisado MI, Garcia-Roman N, Garcia-Marin LJ, Alvarez-Barrientos A, Bragado MJ, Lorenzo MJ. Lovastatin inhibits the extracellular-signal-regulated kinase pathway in immortalized rat brain neuroblasts. *Biochem J*. 2007;401(1):175–83.
122. Xu XQ, McGuire TF, Blaskovich MA, Sebt SM, Romero G. Lovastatin inhibits the stimulation of mitogen-activated protein kinase by insulin in HIRcB fibroblasts. *Arch Biochem Biophys*. 1996;326(2):233–7.
123. Osterweil EK, Chuang SC, Chubykin AA, Sidorov M, Bianchi R, Wong RK, Bear MF. Lovastatin corrects excess protein synthesis and prevents epileptogenesis in a mouse model of fragile X syndrome. *Neuron*. 2013;77(2):243–50.
124. Caku A, Pellerin D, Bouvier P, Riou E, Corbin F. Effect of lovastatin on behavior in children and adults with fragile X syndrome: an open-label study. *Am J Med Genet A*. 2014;164A(11):2834–42.
125. Berry-Kravis E, Levin R, Shah H, Mathur S, Darnell JC, Ouyang B. Cholesterol levels in fragile X syndrome. *Am J Med Genet A*. 2015;167A(2):379–84.
126. Cartagena CM, Phillips KL, Williams GL, Konopko M, Tortella FC, Dave JR, Schmid KE. Mechanism of action for NNZ-2566 anti-inflammatory effects following PBBI involves upregulation of immunomodulator ATF3. *Neuromolecular Med*. 2013;15(3):504–14.
127. Wei HH, Lu XC, Shear DA, Waghray A, Yao C, Tortella FC, Dave JR. NNZ-2566 treatment inhibits neuroinflammation and pro-inflammatory cytokine expression induced by experimental penetrating ballistic-like brain injury in rats. *J Neuroinflammation*. 2009;6:19.
128. Lu XC, Chen RW, Yao C, Wei H, Yang X, Liao Z, Dave JR, Tortella FC. NNZ-2566, a glypromate analog, improves functional recovery and attenuates apoptosis and inflammation in a rat model of penetrating ballistic-type brain injury. *J Neurotrauma*. 2009;26(1):141–54.
129. Deacon RM, Glass L, Snape M, Hurley MJ, Altimiras FJ, Biekofsky RR, Cogran P. NNZ-2566, a novel analog of (1–3) IGF-1, as a potential therapeutic agent for fragile X syndrome. *Neuromolecular Med*. 2015;17(1):71–82.
130. Abitbol M, Menini C, Delezoide AL, Rhyner T, Vekemans M, Mallet J. Nucleus basalis magnocellularis and hippocampus are the major sites of FMR-1 expression in the human fetal brain. *Nat Genet*. 1993;4(2):147–53.
131. Kesler SR, Lightbody AA, Reiss AL. Cholinergic dysfunction in fragile X syndrome and potential intervention: a preliminary 1H MRS study. *Am J Med Genet A*. 2009;149A(3):403–7.
132. D'Antuono M, Merlo D, Avoli M. Involvement of cholinergic and gabaergic systems in the fragile X knockout mice. *Neuroscience*. 2003;119(1):9–13.
133. Sahu JK, Gulati S, Sapra S, Arya R, Chauhan S, Chowdhury MR, Gupta N, Kabra M, Gupta YK, Dwivedi SN, et al. Effectiveness and safety of donepezil in boys with fragile X syndrome: a double-blind, randomized, controlled pilot study. *J Child Neurol*. 2013;28(5):570–5.
134. Chugani DC, Muzik O, Chakraborty P, Mangner T, Chugani HT. Human brain serotonin synthesis capacity measured in vivo with a-(C-11) methyl-L-tryptophan. *Synapse-New York*. 1998;28:33–43.
135. Hanson AC, Hagerman RJ. Serotonin dysregulation in fragile X syndrome: Implications for treatment. *Intractable Rare Dis Res*. 2014;3(4):110–7.
136. Jeste SS, Geschwind DH. Clinical trials for neurodevelopmental disorders: at a therapeutic frontier. *Sci Transl Med*. 2016;8(321):321fs321.
137. Budimirovic D, Duy P. Challenges in translating therapeutic frontiers in clinical trials: where are we now and what's next. *Madridge J Neuro Sci*. 2016;1(1):e1–3.
138. McCracken JT, McGough J, Shah B, Cronin P, Hong D, Aman MG, Arnold LE, Lindsay R, Nash P, Hollway J, et al. Risperidone in children with autism and serious behavioral problems. *N Engl J Med*. 2002;347(5):314–21.
139. Marcus RN, Owen R, Kamen L, Manos G, McQuade RD, Carson WH, Aman MG. A placebo-controlled, fixed-dose study of aripiprazole in children and adolescents with irritability associated with autistic disorder. *J Am Acad Child Adolesc Psychiatry*. 2009;48(11):1110–9.
140. Kaufmann WE, Capone GT, Clarke M, Budimirovic DB. Autism in genetic intellectual disability. In: *Autism*. Springer; 2008:81–108. <http://www.springer.com/us/book/9781603274883>.
141. Wolff JJ, Bodfish JW, Hazlett HC, Lightbody AA, Reiss AL, Piven J. Evidence of a distinct behavioral phenotype in young boys with fragile X syndrome and autism. *J Am Acad Child Adolesc Psychiatry*. 2012;51(12):1324–32.
142. Ethridge L, White S, Mosconi M, Wang J, Byerly M, Sweeney J. Reduced habituation of auditory evoked potentials indicate cortical hyper-excitability in Fragile X Syndrome. *Transl Psychiatry*. 2016;6(4), e787.
143. Hall SS, Lightbody AA, McCarthy BE, Parker KJ, Reiss AL. Effects of intranasal oxytocin on social anxiety in males with fragile X syndrome. *Psychoneuroendocrinology*. 2012;37(4):509–18.