UCSF

UC San Francisco Previously Published Works

Title

The inextricable axis of targeted diagnostic imaging and therapy: An immunological natural history approach

Permalink

https://escholarship.org/uc/item/6nc4s61d

Journal

Nuclear Medicine and Biology, 43(3)

ISSN

0969-8051

Authors

Cope, Frederick O Abbruzzese, Bonnie Sanders, James et al.

Publication Date

2016-03-01

DOI

10.1016/j.nucmedbio.2015.11.007

Peer reviewed

HHS Public Access

Author manuscript

Nucl Med Biol. Author manuscript; available in PMC 2017 March 01.

Published in final edited form as:

Nucl Med Biol. 2016 March; 43(3): 215–225. doi:10.1016/j.nucmedbio.2015.11.007.

The Inextricable Axis Of Targeted Diagnostic Imaging And Therapy: An Immunological Natural History Approach

FO Cope¹, B Abbruzzese¹, J Sanders¹, W Metz¹, K Sturms¹, D Ralph¹, M Blue¹, J Zhang², P Bracci², W Bshara³, S Behr³, T Maurer³, A Beverly⁴, B Blay⁴, A Damughatla⁴, M Larsen⁴, C Mountain⁴, E Neylon⁴, K Parcel⁴, K Raghuraman⁴, K Ricks⁴, L Rose⁴, A Sivakumar⁴, N Streck⁴, B Wang³, C Wasco³, A Williams³, and M McGrath²

¹Navidea Biopharmaceuticals, Drug Development, 5600 Blazer Parkway, Dublin, OH 43017

²The University of California San Francisco and the San Francisco General Hospital, AIDS and Cancer Specimen Resource Center, The Department of Pathology, 1001 Potrero Ave, Bldg. 3, Rm 207 San Francisco, CA 94110

³Roswell Park Cancer Institute, Elm and Carlton Streets, Buffalo, NY 14263

⁴Navidea Biopharmaceuticals Drug Development Internship Program, 5600 Blazer Parkway, Dublin, OH 43017

Summary

In considering the challenges of approaches to clinical imaging, we are faced with choices that sometimes are impacted by rather dogmatic notions about what is a better or worse technology to achieve the most useful diagnostic image for the patient. For example, is PET or SPECT most useful in imaging any particular disease dissemination? The dictatorial approach would be to choose PET, all other matters being equal. But is such a totalitarian attitude toward imaging selection still valid? In the face of new receptor targeted SPECT agents one must consider the remarkable specificity and sensitivity of these agents. 99mTc-Tilmanocept is one of the newest of these agents, now approved for guiding sentinel node biopsy (SLNB) in several solid tumors. Tilmanocept has a K_d of 3×10^{-11} M, and it specificity for the CD206 receptor is unlike any other agent to date. This coupled with a number of facts, that specific disease-associated macrophages express this receptor (100 to 150 thousand receptors), the receptor has multiple binding sites for tilmanocept (>2 sites per receptor) and that these receptors are recycled every 15 minutes to bind more tilmanocept (acting as intracellular "drug compilers" of tilmanocept into non-degraded vesicles), give serious pause as to how we select our approaches to diagnostic imaging. Clinically, the size of SLNs varies greatly, some, anatomically, below the machine resolution of SPECT. Yet, with tilmanocept targeting, the SLNs are highly visible with macrophages stably accruing adequate ^{99m}Tc-tilmanocept counting statistics, as high target-to-background ratios can compensate for spatial resolution blurring. Importantly, it may be targeted imaging agents per se, again such as tilmanocept, which may significantly shrink any perceived chasm between the

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

imaging technologies and anchor the diagnostic considerations in the targeting and specificity of the agent rather than any lingering dogma about the hardware as the basis for imaging approaches. Beyond the elements of imaging applications of these agents is their evolution to therapeutic agents as well, and even in the neo-logical realm of *theranostics*. Characteristics of agents such as tilmanocept that exploit the natural history of diseases with remarkably high specificity are the expectations for the future of patient- and disease-centered diagnosis and therapy.

Diagnosis and Imaging: Setting Up the Therapeutic Continuum

In the preceding 25 years there has evolved a confluence of tumor biology ideology, nuclear medicine and surgical oncology that has led to the development of the theory of the "sentinel node" [1-6]. Data accrued over the intervening years has provided a confirmation of the sentinel node theory as it relates to the incorporation of sentinel lymph node detection/ biopsy in breast cancer and melanoma patient outcome in surgical practice [7–15]. Sentinel node theory holds that there is a predictable anatomical relationship between the immediate tumor environment and the proximate lymphatic system such that assessment of this nexus can provide a reliable appraisal of the nodal disease stage and reduce or eliminate the need for expanded surgery as this relates to lymphadenectomy, and be equally predictive of nodal status with similar outcomes with regard to any such expatiated surgery [16-25]. The initial sentinel lymph node biopsy (SLNB) forays relied on the application of dyes injected into or around the tumor area, with visual tracing of these dyes, or "chasing" the drainage of the dyes into the lymphatic ducts and nodes. The flow and adsorption of the dyes into proximal nodes (and in many cases distal nodes) was implicative of a node's anatomic or biological linkage to the tumor bed and increased potential for the residence of tumor cells whose derivation was from the primary tumor [26-32]. This procedure of SLNB was tested repeatedly in clinical studies and these studies provided validation of the concept and its positive impact on patient outcome [16-25].

However, other factors altered the clinical landscape of SLNB, in particular, the adoption of radiolabeled particulates used in other diagnostic procedures [33–40]. Although neither blue dyes nor particulate colloids provided any real specificity, the combined use of these two agents has improved SLNB detection reliability to singly employed dyes or colloids [41–43]. The results of this unlikely amalgam lead to their use in numerous clinical trials and the expanded adoption of the SLNB procedure, primarily for breast cancer followed by its use in melanoma surgery.

In the face of this seeming remedy for mitigating some unnecessarily extensive surgery, the adoption of SLNB in solid tumors other than melanoma and breast cancer seemed highly likely. However, out of the evolving SLNB clinical data sets for breast cancer and melanoma, and especially the experiential data of SLNB in other solid tumors failed to realize the efficacy found in breast and melanoma surgery, came the observation that there was a need for a SLN-discriminating agent in order to thoroughly potentiate the reliability and positive patient outcome of SLNB. These data strongly suggested that a SLN detection agent would have to provide true target specificity such that the tumor-node axis was more reliably mapped.

The agent that has risen to fill this void is ^{99m}Tc-tilmanocept (Lymphoseek[®], Navidea Biopharmaceuticals, Inc.) (Figures 1a and 1b). Tilmanocept functions through a mechanism of action that relies on its mannose moieties, which bind strongly (as multivalent ligands) and specifically to C-type lectin receptors (CD206), which are present on the surface of macrophages and dendritic cells residing in SLNs. It is this mechanism of action that gives the tilmanocept platform its tremendous capability as a diagnostic and SLN detection agent (Figures 2 and 3). It is now clear from the clinical studies of tilmanocept not only in breast cancer and melanoma, but in head/neck (oral) squamous cell carcinoma that such targeting significantly reduces the false negative rate for SLNB (Table 1).

Recognition of the Imaging-Therapeutic Axis

As noted, ^{99m}Tc-tilmanocept is currently used as a mapping agent for the identification of sentinel lymph nodes in multiple solid tumors. The approved indications for ^{99m}Tc-tilmanocept include guiding SLNB in clinically node negative breast cancer, melanoma, and oral cavity squamous cell carcinoma.

In the context of studying and developing ^{99m}Tc-tilmanocept, the cellular biology underlying the diagnostic targeting specificity is ineluctably linked to the potential for ^{99m}Tc-tilmanocept to be used as a targeting agent against those diseases studied. The basis for this eureka moment was the understanding of the pleotropic impact of the CD206-expressing macrophages in not only the oncology realm (the tumorigenesis process) but in diseases in general.

Tumor evolution and the necessity of tumor-associated macrophages (TAMs) even in the early carcinogenesis process are well established. It is no coincidence that such macrophages express CD206, the explicit receptor for Tilmanocept, and that this coincident convenience is an element of the efficacy of ^{99m}Tc-tilmanocept in SLNB. But the matter is more than this. Macrophages are an integral process element in a host of other diseases. We now realize that their integration into disease processes is remarkably complex, with many highly integrated macrophage and disease elements, testifying to the biologically synchronized transduction of macrophage roles in disease natural history.

In many maladies, including autoimmune diseases (such as multiple sclerosis, diabetes, rheumatoid arthritis (RA), systemic lupus erythematosus (SLE) and Crohn's disease), infectious diseases (HIV and tuberculosis), tumorigenesis, neurodegenerative disorders (dementias) and cardiovascular disease (vulnerable plaque and atherosclerosis), the presence of CD206-macrophages becomes an autogenous progression of the disease state. Table 2 lists a number of these diseases and macrophage involvement. This table effectively defines the not only the potential for diagnostic use of 99m Tc-tilmanocept, but also the opportunity for therapeutic targeting using tilmanocept as the targeting agent.

Selected Opportunities – Exploiting the Diagnostic/Therapeutic Axis: Tilmanocept to Manocepttm

An adaptation of tilmanocept from imaging to therapy is supported by both in vivo and ex vivo data, we have adapted the nomenclature to segregate these initiatives: "Manocept" agents that target the same CD206-macrophage mannose receptor and are derived from Tilmanocept now represent a therapeutic enterprise. In this vane, we outline below selected clinical sorties in which there is a validated transition of tilmanocept diagnostic imaging to Manocept therapeutic targeting.

Kaposi's sarcoma (KS)

AIDS-related KS is an aggressive, multifocal, angioproliferative neoplasm associated with Kaposi's sarcoma herpes virus (HHV8/KSHV) infection. It involves cutaneous and visceral tissues, with later forms of disease associated with widespread organ involvement. It is the most common cancer in patients infected with HIV. Effective antiviral therapy has produced a decline in the incidence of AIDS related KS, but HIV-infected individuals still have a 3,640-fold greater risk of developing KS than the uninfected population. In general, no imaging studies have been able to identify specific KS-involved tissues, apart from standard ultrasound and CT imaging, in which therapy-associated changes are implied to be associated with KS lesion shrinkage.

Inflammation appears to play a critical role in tumor development of HIV-associated KS. Specifically, emerging data show that KS tumor cells that co-express various macrophage antigens, especially CD206, the tilmanocept receptor, become resistant to current anti-viral therapies used to treat KS and AIDS. Macrophages also are a known source of KS tumor cell growth factors and substantial evidence suggests that TAMs represent a reservoir for HIV and its evolved retroviral variants. The macrophage pool driving these two pathological pathways share a common element rooted in the macrophages, the CD206 human macrophage mannose receptor. Manocept, as a molecular targeting agent, binds and enters macrophages via pinocytosis of holo-CD206, providing a cell portal for the evaluation of Manocept as a macrophage and KS targeting agent.

Kaposi's sarcoma lesions are comprised of KS spindle cells infected with HHV8/KSHV, as well as numerous macrophage antigen-expressing cells. In a study of over 100 KS lesions we demonstrated that both skin and visceral forms of KS express the CD206 molecule; CD206 was found on both KS tumor cells and TAMs, allowing the potential for use of tilmanocept as a tumor-specific imaging agent capable of identifying both tumor cells and TAMs in patients with KS (Figures 4 and 5). The results of imaging in vivo in both HIV+ and HIV- KS patients suggest that the therapeutic potential value of Manocept. The supraspecificity of tilmanocept for the KS lesions and the virtual lack of off-target (non-CD206+) background in vivo is strong evidence for employing Manocept congeners in KS and other solid tumors as tumor specific therapies.

Rheumatoid Arthritis (RA)

Rheumatoid arthritis is a common, chronic, systemic, and progressive autoimmune disease associated with inflammation and pathology throughout the body, but perhaps most noticeably in the peripheral joints of the skeleton (i.e., hands, feet, knees, hips, etc.). In the affected joints, RA is characterized by macrophage and lymphocyte infiltration, proliferation of synovial fibroblastic tissue (pannus), and joint destruction. RA causes joint pain, stiffness and reduced mobility. If not successfully treated, joint inflammation and destruction in RA patients can lead to crippling loss of function, severe chronic pain, and disfigurement of the joints. Also, RA patients have significantly higher risk of coronary heart disease including acute myocardial infarction (odds ratio = 3.17). Patients with uncontrolled RA may also suffer a reduction of 3–10 years in their life expectancies. RA can strike anyone at any age, but is diagnosed most frequently in women in their 40s and 50s. Worldwide, about one adult in every 200 has RA. In the United States, approximately 1.3 million adults have RA. RA is a chronic disease and the prevalence of RA increases with age. Because of aging demographics in the United States and elsewhere, the number of patients with RA and the burden of RA on society are expected to increase in the coming decades. Therefore, there is a significant current and growing need to manage RA patients more effectively to limit the morbidity and mortality caused by RA.

While many types of cells, including T-cells, B-cells, dendritic cells, and activated synovial fibroblasts, contribute significantly to the establishment and maintenance of RA, macrophages play a critical role in RA pathogenesis. They produce most of the TNFa that drives and perpetuates the inflammatory cycle in RA. In the synovial sub-lining of a joint affected by RA, macrophages are the dominant cell type. In the inflamed joint as a whole, macrophages in RA patients make up at least 30%-40% of all cells. Furthermore, macrophages participate directly in the destruction of bone and cartilage. Activated macrophage populations and synoviocytes are the predominant cell types at the interface between pannus and cartilage and secrete destructive proteases in abundance. As a result, it may not be surprising that synovial macrophage numbers—but not the numbers of other immune cell types—correlate with radiographically determined joint destruction in RA. While macrophages may play a role in other pathologies that cause joint pain and inflammation, the degree to which macrophages are involved in the pathological process of RA and the sheer mass or volume of macrophages that infiltrate the joints inflamed due to RA differentiates RA from other rheumatic diseases. Therefore, detection of the density or numbers of macrophages in inflamed joints may permit differentiation of patients with RA from those with other causes of arthritis. In addition, it is known that the RA pathology begins significantly before, perhaps years before, the onset of symptoms (i.e., joint pain and inflammation) and well before the beginning of bone destruction. Macrophage infiltration of synovial tissues precedes development of RA clinical signs in animal studies. In humans, macrophage infiltrates in synovial tissues are present when RA patients first develop clinical symptoms. Therefore, detection of the density or numbers of macrophages in inflamed joints may facilitate more sensitive and specific identification of RA patients as soon as they present with symptoms and early in the course of their illnesses when disease-modifying anti-rheumatic drugs (DMARDs) are likely to be most effective.

Below are select data from studies using tilmanocept as an imaging agent for synovial macrophages in the inflamed joints of RA patients and in animal models. We investigated CD206 expression and tilmanocept binding to synovial macrophages in an anti-type II collagen monoclonal antibody induced mouse model of RA. Male 8-week-old Dba1 mice (n=8) were injected intraperitoneally (IP) with 1.5 mg Athrogen-CIA arthrogenic five monoclonal antibody cocktail to Type II collagen followed in three days by an IP injection with E. coli 0111:B4 lipopolysaccharide. Control mice (n = 4) were injected with phosphate-buffered saline. Evidence of arthritis (joint swelling and redness) developed in 5–6 days, and the severity of arthritis was scored for each limb daily. On days 9 or 11, mice were imaged 1–2 hours after they had received an intravenous injection of fluorescent Cy3-tilmanocept. The mice were then euthanized followed by limb dissection and reimaging. The primary result of this experiment was that Cy3-tilmanocept administered intravenously localized to synovial macrophages in the affected joints of arthritic mice but not control mice (Figure 6).

Ex vivo experiments showed specific binding of Cy3-tilmanocept to human synovial macrophages obtained in samples from patients with active RA undergoing therapeutic surgical procedures (Figure 3). For these studies, RA patients were recruited through approved institutional review board protocols. Flash-frozen synovial tissue was sectioned to 4 µm onto glass slides for immunohistochemistry. Slides were incubated with DAPI nuclear stain (blue), an anti-CD206 antibody and/or Cy3-tilmanocept (red). Bound anti-CD206 antibody was visualized with a secondary antibody conjugated with Alexa Fluor 647 (green). Images were obtained using a Zeiss fluorescent microscope and merged to show colocalization. It is important to note that, first, human synovial macrophages from human RA patients abundantly express CD206, and (second) Cy3-tilmanocept binds to human synovial macrophages and co-localizes with CD206. Finally (third), CD206-expressing macrophages are a highly abundant cell type in RA synovia. The specificity of Cy3-tilmanocept binding to synovial macrophages was further demonstrated by pre-incubating with a tenfold excess of tilmanocept that had not been conjugated with Cy3. Pre-incubation with unconjugated tilmanocept completely abolished Cy3-tilmanocept binding (Figure 7).

In order to provide perspective around the specificity of tilmanocept binding/uptake into RA synovial macrophages, additional experiments performed with synovial tissues obtained from joints of patients with osteoarthritis and from healthy controls (obtained from a tissue bank), provided results showing that Cy3-tilmanocept binding was much greater in synovial tissues from RA patients than it was in similar tissues obtained from patients with osteoarthritis or healthy controls. It is well known that osteoarthritic joints contain macrophage infiltrates; however, in osteoarthritis, synovial macrophages are less numerous than in RA and as this experiment shows, synovial macrophages in osteoarthritis produce much less CD206 (Figure 8).

Similar to KS, the specificity of tilmanocept for the RA loci and the virtual lack of off-target (non-CD206+) background reactivity provides strong supportive evidence for employing Manocept congeners in RA therapies.

Tuberculosis (TB)

Mycobacterium tuberculous (Mtb) is an airborne pathogen that infects the lungs and then can disseminate throughout the body. Mtb is spread when an individual with symptomatic or active TB coughs, creating an Mtb-rich aerosol that can be inhaled into the lungs of previously uninfected persons. Alveolar macrophages recognize Mtb by various means but importantly through interactions between the mannose caps on the Mtb coat molecule, mannosyslated lipoarabinomannan (ManLAM), and CD206. The interaction between ManLAM and CD206 is important because it alters phagosome trafficking, inhibiting fusion of the Mtb-containing phagosome with lysosomes, and allowing Mtb to survive and replicate in these cells. The infected alveolar macrophage then secretes various cytokines that attract additional macrophages and various other immune cells. Mtb replication and dissemination combined with the accumulation of various immune cells leads to a robust systemic immune response and the early formation of a granuloma. The granuloma (granulomatous pulmonary Mtb) is comprised of T and B lymphocytes surrounding a fibrous cuff. Within the fibrous cuff there are macrophages (both infected and uninfected) and some neutrophils. Lipid metabolism in many of the macrophages becomes perturbed, perhaps in response to stimuli from the Mtb, causing them to further differentiate into foam cells containing lipid micro-droplets. The granuloma sequesters Mtb and prevents it from spreading to other parts of the body. However, Mtb can persist in a metabolically quiescent state for years or decades in granulomas. The large majority of individuals who become infected with Mtb do not exhibit symptoms, having their Mtb sequestered in granulomas. Such asymptomatic persons are said to have latent TB infections (LTBI). Unfortunately over time, there is risk that a granuloma can progress, with the patient developing active TB. In active TB, the granuloma enlarges with a necrotic center comprised of Mtb cells and a lipid rich material derived from the cellular debris of dead foam cells, which is called caseum due to an appearance resembling milky cheese. In time, the granuloma may rupture releasing Mtb and necrotic debris into the patient's airway, causing coughing that spreads the infection to other people. If a person with active TB is not treated, on average they will transmit Mtb to 10-15 people per year. A latently infected person without co-morbidities such as AIDS or diabetes has a 5%-10% chance of developing active TB over their lifetime. It is estimated that one third of the world's population is latently infected with Mtb.

Our recent data show (Figure 9) that the infection of macrophages with Mtb does not downregulate the synthesis and expression of CD206, nor does infection abrogate the uptake of tilmanocept via CD206 and its accrual intracellularly.

These data are, again, notable evidence for the use of a Manocept therapeutic congener for the treatment of Mtb and drug resistant Mtb to the extent that we believe these data suggest that the biology of the granuloma is now made vulnerable to attack using many congener strategies.

Cardiovascular Disease

Atherosclerosis is a chronic inflammatory syndrome that develops slowly in the walls of arteries over the course of many years or decades. The initiation and progression of atherosclerosis involves interactions between plasma lipoproteins, cytokines extracellular

matrix, inflammatory signaling molecules and several cell types including macrophages. The primary lesion of atherosclerosis is the atherosclerotic plaque. Atherosclerotic plaques can expand into the lumen of an artery, eventually impacting and diminishing the flow of blood. This impairment of blood flow can result in angina pectoris, which is painful and debilitating. Alternatively, atherosclerosis can progress to an advanced state without producing symptoms. Eventually, an atherosclerotic plaque can rupture causing the internal contents of the plaque to come into contact with the blood, thereby initiating thrombus (i.e., clot) formation. These clots can lead to ischemic events that manifest as myocardial infarctions, sudden cardiac deaths or strokes. For a tragically large proportion of patients, experiencing one of these potentially catastrophic and/or lethal events is the first observed symptom experienced by a patient with advanced atherosclerosis. To prevent these potentially catastrophic plaque rupture associated events, it is necessary to identify patients with atherosclerotic plaques that are at high risk of near term rupture so that appropriate intensive rupture preventing therapeutic interventions can be administered.

Autopsies performed on people who have died as a result of infarctions caused by ruptures of atherosclerotic plaques have identified the relevant ruptured plaques, which are termed the "culprit lesions." To identify individuals at high-risk of impending plaque rupture, atherosclerotic plaques that most closely resemble culprit plaques but which have not yet ruptured have been identified. These culprit-resembling plaques are termed "vulnerable plaques." Vulnerable plaques have morphological features and internal compositions that differentiate them from other atherosclerotic plaques (i.e., stable plaques). These differentiating features include the presence of large necrotic cores associated with a lipid pool and a thin (<65 micron) fibrous cap. This type of vulnerable plaque is also referred to as a thin cap fibroatheroma (TCFA). TCFA and culprit plaques are not typically highly calcified, but may display evidence of nascent calcification that is not yet extensive. Interestingly, despite their relatively large sizes, TCFA most commonly do not cause severe narrowing of the lumen of the arteries in which they occur. Instead TCFA are frequently associated with remodeling of their arterial walls that expands their arterial lumens to accommodate their relatively large volumes.

Macrophages contribute a key and evolving role at each stage of the pathological development of atherosclerosis. Following an injury to the arterial endothelium, low density lipoproteins (LDL) invade the endothelium and become oxidized, initiating an inflammatory response that attracts monocytes. These monocytes ingest the oxidized LDL and become macrophage "foam cells" that further propagate the inflammatory response by secreting proinflammatory cytokines. Eventually, the foam cells die or undergo apoptosis creating the lipid rich necrotic core of vulnerable plaque. The necrotic core further attracts macrophages. Narula et al examined atherosclerotic plaques from patients who died suddenly [133]. They observed that an increase in the number of infiltrating macrophages was a key discriminator between TCFA and stable plaques. Importantly, Tahara et al also examined atherosclerotic plaques from patients that had suffered sudden cardiac death [134]. They confirmed Narula's findings of increased macrophages in TCFA. In addition, immunohistochemical analyses on the plaques found that large proportions of macrophages in TCFA, but not in stable thick capped plaques, confirmed positive expression of CD206, the macrophage mannose receptor and the specific target for ^{99m}Tc-tilmanocept binding, and CD163, both

markers for alternative, M2, macrophage activation (30–33). These results suggest that Tc-tilmanocept will bind to abundant macrophage targets in TCFA and that CD206 imaging results can differentiate TCFA from stable atherosclerotic plaques.

Macrophages can be activated to differentiate into various gene expression phenotypes when stimulated by various combinations of cytokines and their local environment. Macrophages can be activated to a pro-inflammatory, M1, phenotype, or anti-inflammatory, M2a, M2b, and M2c, phenotypes. Early in the development of atherosclerosis, M1 macrophages predominate in atherosclerotic plaques, with foam cells expressing many pro-inflammatory cytokines characteristic of a M1 phenotype. M2a and M2c express high levels of CD206. M2c macrophages are particularly interesting because they accumulate in areas where apoptotic cells are present. The necrotic cores of vulnerable atherosclerotic plaques result largely from the apoptosis of foam cells and as such, would be expected to attract M2c macrophages. M2c macrophages abundantly express CD163 in addition to CD206.

These observations are consistent with our approach to exploiting this cardiovascular disease natural history with Manocept congeners for both imaging and therapy. Our ex vivo data showing both the expression of CD206 and the localization of fluorescent Manocept in the coronary arteries on this CD206 is strong evidence for the axis initiative in this disease (Figure 10).

Conclusions

In the wake of innovation, tilmanocept (and as Manocept) is self-transforming from a powerful diagnostic/imaging agent in cancer staging procedures into a potential multi-application molecule for diagnosis and treatment of an array of diseases. It has displayed its effectiveness in applications with regards to SLNB, much due to its unique structure and targeting. As a non-particulate, small molecular size, receptor targeted (CD206) molecule with the ability to interchange not only radionuclides but also lethal or biological modifier molecules, tilmanocept/Manocept has great potential for targeting macrophage mediated diseases and delivering an effective, concentrated dose for purposes of diagnosis or treatment. As Manocept continues to generate clinically relevant data for future applications in immunotherapies for KS (and other solid tumors), RA, TB, and cardiovascular disease, one thing remains clear, the selection of this approach, targeting CD206, appears to provide encouraging results with inter-disease consistency and reliability on the this biological strategy.

Terachem Manuscript References

- 1. Gould EA, Winship T, Philbin PH, Kerr HH. Observations on a "sentinel node" in cancer of the parotid. Cancer. 1960; 13:77–78. [PubMed: 13828575]
- 2. Cabanas RM. An approach for the treatment of penile carcinoma. Cancer. 1977; 39:456–466. PMID 837331. [PubMed: 837331]
- 3. Morton DL, Wen DR, Wong JH, et al. Technical details of intraoperative lymphatic mapping for early stage melanoma. Arch Surg. 1992; 127:392–399. [PubMed: 1558490]
- Morton DL, Wen DR, Wong JH, Economou JS, Cagle LA, Storm FK, Foshag LJ, Cochran AJ. Technical details of intraoperative lymphatic mapping for early stage melanoma. Arch Surg. 1992; 127:392–399. [PubMed: 1558490]

 Krag DN, Weaver DL, Alex JC, Fairbank JT. Surgical resection and radiolocalization of the sentinel lymph node in breast cancer using a gamma probe. Surg Oncol. 1993; 2:335–339. [PubMed: 8130940]

- Giuliano AE, Kirgan DM, Guenther JM, Morton DL. Lymphatic mapping and sentinel lymphadenectomy for breast cancer. Ann Surg. 1994; 220:391–398. [PubMed: 8092905]
- Giuliano AE. Mapping a pathway for axillary staging: a personal perspective on the current status of sentinel lymph node dissection for breast cancer. Arch Surg. 1999; 134:195–199. [PubMed: 10025463]
- 8. Turner RR, Ollila DW, Krasne DL, Giuliano AE. Histopathologic validation of the sentinel lymph node hypothesis for breast carcinoma. Ann Surg. 1997; 226:271–278. [PubMed: 9339933]
- 8. Hsueh EC, Turner RR, Glass EC, Brenner RJ, Brennan MB, Giuliano AE. Sentinel node biopsy in breast cancer. J Am Coll Surg. 1999; 189:207–213. [PubMed: 10437844]
- Giuliano AE, Haigh PI, Brennan MB, Hansen NM, Kelley MC, Ye W, Glass EC, Turner RR. Prospective observational study of sentinel lymphadenectomy without further axillary dissection in patients with sentinel node-negative breast cancer. J Clin Oncol. 2000; 18:2553–2559. [PubMed: 10893286]
- 11. Tanis PJ, Nieweg OE, Valdés Olmos RA, Th Rutgers EJ, Kroon BB. History of sentinel node and validation of the technique. Breast Cancer Research. 2001; 3(2):109–112. [PubMed: 11250756]
- 12. Krag DN. Is axillary dissection needed when the sentinel node is positive? Yes! J Surgical Oncology. 2008 Mar; 97(3):197–198. [PubMed: 18264980]
- Sadeghi R, Asadi M, Treglia G, Zakavi SR, Fattahi A, Krag DN. Determining axillary concordance rate for different injection locations in sentinel node mapping of breast cancer: how ambitious can we get? Breast Cancer Res Treat. 2014 Jul; 146(1):231–232. PubMed PMID: 24878987. [PubMed: 24878987]
- 14. Calhoun BC, Chambers K, Flippo-Morton T, Livasy CA, Armstrong EJ 3rd, Symanowski JT, Sarantou T, Greene FL, White RL Jr. Breast cancer detection in axillary sentinel lymph nodes: the impact of the method of pathologic examination. Hum Pathol. 2014 Dec; 45(12):2497–2501. Epub 2014 Oct 2. PubMed PMID: 25449631. [PubMed: 25449631]
- 15. Pesek S, Ashikaga T, Krag LE, Krag D. The false-negative rate of sentinel node biopsy in patients with breast cancer: a meta-analysis. World J Surg. 2012 Sep; 36(9):2239–2251. PubMed PMID: 22569745; PubMed Central PMCID: PMC3469260. [PubMed: 22569745]
- 16. Parrett BM, Kashani-Sabet M, Singer MI, Li R, Thummala S, Fadaki N, Leong SP. Long-term prognosis and significance of the sentinel lymph node in head and neck melanoma. Otolaryngol Head Neck Surg. 2012 Oct; 147(4):699–706. Epub 2012 Apr 24. PubMed PMID: 22535913. [PubMed: 22535913]
- Layfield DM, Agrawal A, Roche H, Cutress RI. Intraoperative assessment of sentinel lymph nodes in breast cancer. Br J Surg. 2011 Jan; 98(1):4–17. Epub 2010 Sep 1. Review. PubMed PMID: 20812233. [PubMed: 20812233]
- Holloway CM, Easson A, Escallon J, Leong WL, Quan ML, Reedjik M, Wright FC, McCready DR. Technology as a force for improved diagnosis and treatment of breast disease. Can J Surg. 2010 Aug; 53(4):268–277. Review. PubMed PMID: 20646402; PubMed Central PMCID: PMC2912014. [PubMed: 20646402]
- Clark RR, Shaw-Dunn J, Soutar DS. A cadaveric study of auricular lymphatics and implications for sentinel lymph node biopsy. Clin Anat. 2010 Oct; 23(7):792–797. PubMed PMID: 20641070. [PubMed: 20641070]
- Pajares M, Freire JM, Moreno P, Utor A, Tocino A, Alonso E. Evaluation of the quality of the sentinel lymph node biopsy procedure in patients with breast cancer. Rev Esp Med Nucl. 2010 Sep-Oct;29(5):236–240. Epub 2010 Jul 16. Spanish. PubMed PMID: 20637527. [PubMed: 20637527]
- 21. Wells BJ, Najjar H, Wright FC, Holloway CM, Fraser N, McCready D, Quan ML. Measuring the quality of sentinel lymph node biopsy in breast cancer using newly developed quality indicators: a feasibility study. Ann Surg Oncol. 2011 Jan; 18(1):78–85. Epub 2010 Jul 13. PubMed PMID: 20625839. [PubMed: 20625839]

22. Piñero A. Present and future of sentinel lymph node biopsy in breast cancer staging. Clin Transl Oncol. 2010 Jul; 12(7):457–458. PubMed PMID: 20615820. [PubMed: 20615820]

- 23. Shen J, Wallace AM, Bouvet M. The role of sentinel lymph node biopsy for melanoma. Semin Oncol. 2002 Aug; 29(4):341–352. Review. PubMed PMID: 12170437. [PubMed: 12170437]
- 24. Davis KG, Schriver JP, Waddell B. Implementation of sentinel lymph node biopsy for breast cancer by surgeons in the Department of Defense. Am J Surg. 2002 Aug; 184(2):94–96. PubMed PMID: 12169350. [PubMed: 12169350]
- 25. Tuttle TM, Zogakis TG, Dunst CM, Zera RT, Singletary SE. A review of technical aspects of sentinel lymph node identification for breast cancer. J Am Coll Surg. 2002 Aug; 195(2):261–268. Review. PubMed PMID: 12168974. [PubMed: 12168974]
- 26. Giménez J, Botella-Estrada R, Hernández D, Carbonell M, Martínez MA, Guillén C, Vázquez C. Anaphylaxis after peritumoral injection of sulphan blue 1% for identification of the sentinel node in lymphatic mapping of the breast. Eur J Surg. 2001 Dec; 167(12):921–923. PubMed PMID: 11841084. [PubMed: 11841084]
- 27. Li X, Wang J, Zhou Z. Experimental study of sentinel lymph node biopsy in thyroid by using three kinds of vital dyes at different concentration and dose. Lin Chung Er Bi Yan Hou Tou Jing Wai Ke Za Zhi. 2007 Nov; 21(21):988–990. Chinese. PubMed PMID: 18309656. [PubMed: 18309656]
- Zakaria S, Hoskin TL, Degnim AC. Safety and technical success of methylene blue dye for lymphatic mapping in breast cancer. Am J Surg. 2008 Aug; 196(2):228–233. PubMed PMID: 18367146. [PubMed: 18367146]
- Liang MI, Carson WE 3rd. Biphasic anaphylactic reaction to blue dye during sentinel lymph node biopsy. World J Surg Oncol. 2008 Jul 27.6:79. PubMed PMID: 18655732; PubMed Central PMCID: PMC2518154. [PubMed: 18655732]
- Bühler HS, Rojas PH, Cayazzo MD, Cunill CE, Vesperinas AG, Hamilton SJ. Exclusive use of blue dye to detect sentinel lymph nodes in breast cancer. Rev Med Chil. 2008 Aug; 136(8):1015– 1020. doi: /S0034-98872008000800008. Epub 2008 Oct 7. Spanish. PubMed PMID: 18949185. [PubMed: 18949185]
- 31. Teknos D, Ramcharan A, Oluwole SF. Pulmonary edema associated with methylene blue dye administration during sentinel lymph node biopsy. J Natl Med Assoc. 2008 Dec; 100(12):1483–1484. PubMed PMID: 19110921. [PubMed: 19110921]
- 32. East JM, Valentine CS, Kanchev E, Blake GO. Sentinel lymph node biopsy for breast cancer using methylene blue dye manifests a short learning curve among experienced surgeons: a prospective tabular cumulative sum (CUSUM) analysis. BMC Surg. 2009 Jan 27.9:2. PubMed PMID: 1917 3714; PubMed Central PMCID: PMC2640353. [PubMed: 19173714]
- 33. Gershenwald JE, Tseng CH, Thompson W, Mansfield PF, Lee JE, Bouvet M, Lee JJ, Ross MI. Improved sentinel lymph node localization in patients with primary melanoma with the use of radiolabeled colloid. Surgery. 1998 Aug; 124(2):203–210. PubMed PMID: 9706139. [PubMed: 9706139]
- 34. Wengenmair H, Kopp J, Vogt H, Wawroschek F, Gröber S, Dorn R, Heidenreich P. Sentinel lymph node diagnosis in prostatic carcinoma: II. Biokinetics and dosimetry of 99mTc-Nanocolloid after intraprostatic injection. Nuklearmedizin. 2002 Apr; 41(2):102–107. German. PubMed PMID: 11989296. [PubMed: 11989296]
- 35. Rasgon BM. Use of low-dose technetium Tc 99m sulfur colloid to locate sentinel lymph nodes in melanoma of the head and neck: preliminary study. Laryngoscope. 2001 Aug; 111(8):1366–1372. PubMed PMID: 11568570. [PubMed: 11568570]
- 36. Tafra L, Chua AN, Ng PC, Aycock D, Swanson M, Lannin D. Filtered versus unfiltered technetium sulfur colloid in lymphatic mapping: a significant variable in a pig model. Ann Surg Oncol. 1999 Jan-Feb;6(1):83–87. PubMed PMID: 10030419. [PubMed: 10030419]
- 37. Pijpers R, Borgstein PJ, Meijer S, Krag DN, Hoekstra OS, Greuter HN, Teule GJ. Transport and retention of colloidal tracers in regional lymphoscintigraphy in melanoma: influence on lymphatic mapping and sentinel node biopsy. Melanoma Res. 1998 Oct; 8(5):413–418. PubMed PMID: 9835454. [PubMed: 9835454]
- 38. Bartolomei M, Testori A, Chinol M, Gennari R, De Cicco C, Leonardi L, Zoboli S, Paganelli G. Sentinel node localization in cutaneous melanoma: lymphoscintigraphy with colloids and antibody

- fragments versus blue dye mapping. Eur J Nucl Med. 1998 Nov; 25(11):1489–1494. PubMed PMID: 9799344. [PubMed: 9799344]
- 39. Liptay MJ, D'amico TA, Nwogu C, Demmy TL, Wang XF, Gu L, Litle VR, Swanson SJ, Kohman LJ. Thoracic Surgery Subcommittee of the Cancer and Leukemia Group B. Intraoperative sentinel node mapping with technitium-99 in lung cancer: results of CALGB 140203 multicenter phase II trial. J Thorac Oncol. 2009 Feb; 4(2):198–202. PubMed PMID: 19179896. [PubMed: 19179896]
- Wallace AM, Vera DR, Stadalnik RC. Blue dye and 99mTc-labeled human serum albumin: sentinel node detection by magic bullets? J Nucl Med. 1999 Jul; 40(7):1149–1150. PubMed PMID: 10405135. [PubMed: 10405135]
- 41. Stolier A. Breast lymphatic mapping using blue dye and radiocolloid. J Am Coll Surg. 2002 May. 194(5):681. author reply 681-3. PubMed PMID: 12022613. [PubMed: 12022613]
- 42. Tafra L, Lannin DR, Swanson MS, Van Eyk JJ, Verbanac KM, Chua AN, Ng PC, Edwards MS, Halliday BE, Henry CA, Sommers LM, Carman CM, Molin MR, Yurko JE, Perry RR, Williams R. Multicenter trial of sentinel node biopsy for breast cancer using both technetium sulfur colloid and isosulfan blue dye. Ann Surg. 2001 Jan; 233(1):51–59. PubMed PMID: 11141225; PubMed Central PMCID: PMC1421166. [PubMed: 11141225]
- 43. van der Ent FW, Kengen RA, van der Pol HA, Hoofwijk AG. Sentinel node biopsy in 70 unselected patients with breast cancer: increased feasibility by using 10 mCi radiocolloid in combination with a blue dye tracer. Eur J Surg Oncol. 1999 Feb; 25(1):24–29. PubMed PMID: 10188850. [PubMed: 10188850]
- 44. Amr D, Broderick-Villa G, Haigh PI, Guenther JM, DiFronzo LA. Adverse drug reactions during lymphatic mapping and sentinel lymph node biopsy for solid neoplasms. Am Surg. 2005 Sep; 71(9):720–724. PubMed PMID: 16468505. [PubMed: 16468505]
- 45. Gipponi M. Clinical applications of sentinel lymph-node biopsy for the staging and treatment of solid neoplasms. Minerva Chir. 2005 Aug; 60(4):217–233. Review. English, Italian. PubMed PMID: 16166921. [PubMed: 16166921]
- 46. Alex JC. The application of sentinel node radiolocalization to solid tumors of the head and neck: a 10-year experience. Laryngoscope. 2004 Jan; 114(1):2–19. Review. PubMed PMID: 14709988. [PubMed: 14709988]
- 47. Krag DN, Weaver DL. Pathological and molecular assessment of sentinel lymph nodes in solid tumors. Semin Oncol. 2002 Jun; 29(3):274–279. Review. PubMed PMID: 12063680. [PubMed: 12063680]
- 48. Gershenwald JE, Coit DG, Sondak VK, Thompson JF. The challenge of defining guidelines for sentinel lymph node biopsy in patients with thin primary cutaneous melanomas. Ann Surg Oncol. 2012 Oct; 19(11):3301–3303. PubMed PMID: 22868918. [PubMed: 22868918]
- 49. Winter A, Vogt C, Weckermann D, Wawroschek F. Complications of pelvic lymphadenectomy in clinically localised prostate cancer: different techniques in comparison and dependency on the number of removed lymph nodes. Aktuelle Urol. 2011 May; 42(3):179–183. Epub 2011 Mar 15. German. PubMed PMID: 21409742. [PubMed: 21409742]
- Schurr PG, Behnke S, Kaifi JT, Bogoevski D, Link B, Mann O, Strate T, Pantel K, Izbicki JR, Yekebas E. Central mesenteric lymph node BER-Ep4+ cells in colorectal cancer: challenge to sentinel node concept? Dig Surg. 2007; 24(1):19–27. Epub 2007 Mar 16. PubMed PMID: 17369677. [PubMed: 17369677]
- Wallace AM, Hoh CK, Vera DR, Darrah DD, Schulteis G. Lymphoseek: a molecular radiopharmaceutical for sentinel node detection. Ann Surg Oncol. 2003 Jun; 10(5):531–538. PubMed PMID: 12794019. [PubMed: 12794019]
- 52. Baker JL, Pu M, Tokin CA, Hoh CK, Vera DR, Messer K, Wallace AM. Comparison of [(99m) Tc] tilmanocept and filtered [(99m) Tc] sulfur colloid for identification of SLNs in breast cancer patients. Ann Surg Oncol. 2015 Jan; 22(1):40–45. Epub 2014 Jul 29. PubMed PMID: 25069859; PubMed Central PMCID: PMC4273083. [PubMed: 25069859]
- 53. Wallace AM, Han LK, Povoski SP, Deck K, Schneebaum S, Hall NC, Hoh CK, Limmer KK, Krontiras H, Frazier TG, Cox C, Avisar E, Faries M, King DW, Christman L, Vera DR. Comparative evaluation of [(99m) Tc] tilmanocept for sentinel lymph node mapping in breast cancer patients: results of two phase 3 trials. Ann Surg Oncol. 2013 Aug; 20(8):2590–2599. Epub

- 2013 Mar 17. PubMed PMID: 23504141; PubMed Central PMCID: PMC3705144. [PubMed: 23504141]
- 54. Sondak VK, King DW, Zager JS, Schneebaum S, Kim J, Leong SP, Faries MB, Averbook BJ, Martinez SR, Puleo CA, Messina JL, Christman L, Wallace AM. Combined analysis of phase III trials evaluating [⁹⁹mTc]tilmanocept and vital blue dye for identification of sentinel lymph nodes in clinically node-negative cutaneous melanoma. Ann Surg Oncol. 2013 Feb; 20(2):680–688. Epub 2012 Oct 3. PubMed PMID: 23054107; PubMed Central PMCID: PMC3560941. [PubMed: 23054107]
- 55. Marcinow AM, Hall N, Byrum E, Teknos TN, Old MO, Agrawal A. Use of a novel receptor-targeted (CD206) radiotracer, 99mTc-tilmanocept, and SPECT/CT for sentinel lymph node detection in oral cavity squamous cell carcinoma: initial institutional report in an ongoing phase 3 study. JAMA Otolaryngol Head Neck Surg. 2013 Sep; 139(9):895–902. PubMed PMID: 24051744; PubMed Central PMCID: PMC4301415. [PubMed: 24051744]
- 56. Wallace AM, Hoh CK, Limmer KK, Darrah DD, Schulteis G, Vera DR. Sentinel lymph node accumulation of Lymphoseek and Tc-99m-sulfur colloid using a "2-day" protocol. Nucl Med Biol. 2009 Aug; 36(6):687–692. PubMed PMID: 19647175; PubMed Central PMCID: PMC4201841. [PubMed: 19647175]
- 57. Wallace AM, Hoh CK, Darrah DD, Schulteis G, Vera DR. Sentinel lymph node mapping of breast cancer via intradermal administration of Lymphoseek. Nucl Med Biol. 2007 Oct; 34(7):849–853. Epub 2007 Aug 9. PubMed PMID: 17921035; PubMed Central PMCID: PMC4189800. [PubMed: 17921035]
- de Paulis T. Drug evaluation: Lymphoseek Neoprobe's sentinel lymph node imaging agent for use in cancer patients. Curr Opin Investig Drugs. 2006 Dec; 7(12):1100–1107. Review. PubMed PMID: 17209528.
- Wallace AM, Hoh CK, Ellner SJ, Darrah DD, Schulteis G, Vera DR. Lymphoseek: a molecular imaging agent for melanoma sentinel lymph node mapping. Ann Surg Oncol. 2007 Feb; 14(2): 913–921. Epub 2006 Dec 5. PubMed PMID: 17146742. [PubMed: 17146742]
- Wallace AM, Ellner SJ, Méndez J, Hoh CK, Salem CE, Bosch CM, Orahood RC, Vera DR. Minimally invasive sentinel lymph node mapping of the pig colon with Lymphoseek. Surgery. 2006 Feb; 139(2):217–223. PubMed PMID: 16455331. [PubMed: 16455331]
- 61. Agrawal A, Civantos FJ, Brumund KT, Chepeha DB, Hall NC, Carroll WR, Smith RB, Zitsch RP, Lee WT, Shnayder Y, Cognetti DM, Pitman KT, King DW, Christman LA, Lai SY. [(99m)Tc]Tilmanocept Accurately Detects Sentinel Lymph Nodes and Predicts Node Pathology Status in Patients with Oral Squamous Cell Carcinoma of the Head and Neck: Results of a Phase III Multi-institutional Trial. Ann Surg Oncol. 2015 Feb 11. [Epub ahead of print] PubMed PMID: 25670018.
- 62. Laoui D, Movahedi K, Van Overmeire E, Van den Bossche J, Schouppe E, Mommer C, Nikolaou A, Morias Y, De Baetselier P, Van Ginderachter JA. Tumor-associated macrophages in breast cancer: distinct subsets, distinct functions. Int J Dev Biol. 2011; 55(7–9):861–867. [PubMed: 22161841]
- 63. Lanciotti M, Masieri L, Raspollini MR, Minervini A, Mari A, Comito G, Giannoni E, Carini M, Chiarugi P, Serni S. The Role of M1 and M2 Macrophages in Prostate Cancer in relation to Extracapsular Tumor Extension and biochemical Recurrence after Radical Prostatectomy. BioMedical Research International. 2014 Mar 11.
- 64. Zhang W, Zhu XD, Sun HC, Xiong YC, Zhuang PY, Xu HX, Kong LQ, Wang L, Wu WZ, Tang ZY. Depletion of tumor-associated macrophages enhances the effect of sorafenib in metastatic liver cancer models by antimetastatic and antiangiogenic effects. Clin Cancer Res. 2010 Jul 1; 16(13):3420–3430. Doi:10.1158/1078-0432.CCR-09-2904. [PubMed: 20570927]
- 65. Hussein Mahmoud R. Tumor-associated macrophages and melanoma tumourigenesis: integrating the complexity. Int J exp Pathol. 2006 Jun; 87(3):163–176. [PubMed: 16709225]
- 66. Ding H, Cai J, Mao M, Fang Y, Huang Z, Jia J, Li T, Xu L, Wang J, Zhou J, Yang Q, Wang Z. Tumor-associated macrophages induce lymphangiogenesis in cervical cancer via interactions with tumor cells. APMIS. 2014 Nov; 122(11):1059–1069. [PubMed: 24698523]
- 67. Steidl C, Lee T, Shah SP, Farinha P, Han G, Nayar T, Delaney A, Jones SJ, Iqbal J, Weisenburger DD, Bast MA, Rosenwald A, Muller-Hermelink HK, Rimsza L, Campo E, Delabie J, Braziel RM,

- Cook JR, Tubbs RR, Jaffe ES, Lenz G, Connors JM, Staudt LM, Chang WC, Gascoyne RD. N Engl J Med. 2010 Mar 11.362:875–885. [PubMed: 20220182]
- 68. Huysentruyt LC, McGrath MS. The role of macrophages in the development and progression of AIDS-related non-Hodgkin lymphoma. J Leuk Biol. 2010 Apr; 87(4):627–632.
- 69. Ribatti D, Vacca A. The role of monocytes-macrophages in vasculogenesis in multiple myeloma. Leukemia. 2009; 23:1535–1536. [PubMed: 19738626]
- 70. Fujiwara T, Fukushi J, Yamamoto S, Matsumoto Y, Setsu N, Oda Y, Yamada H, Okada S, Watari K, Ono M, Kuwano M, Kamura S, Iida K, Okada Y, Koga M, Iwamoto Y. Macrophage infiltration predicts a poor prognosis for human ewing sarcoma. Am J Pathol. 2011 Sep; 179(3):1157–1170. [PubMed: 21771572]
- 71. Ong SM, Tan YC, Beretta O, Jiang D, Weap WH, Tai JJ, Wong WC, Yang H, Schwarz H, Lim KH, Koh PK, Ling KL, Wong SC. Macrophages in human colorectal cancer are pro-inflammatory and prime T cells towards an anti-tumour type-1 inflammatory response. Eur J Immunol. 2012 Jan; 42(1):89–100. [PubMed: 22009685]
- 72. Kumar A, Herbein G. The macrophage: a therapeutic target in HIV-1 infection. Mol Cell Ther. 2014 Apr 2.2:10. [PubMed: 26056579]
- 73. Kinne RW, Bräuer R, Stuhlmüller B, Palombo-Kinne E, Burmester GR. Macrophages in rheumatoid arthritis. Arthritis Res. 2000 Apr 12; 2(3):189–202. [PubMed: 11094428]
- Podinovskaia M, Lee W, Caldwell S, Russel DG. Infection of macrophages with Mycobacterium tuberculosis induces global modifications to phagosomal function. Cell Microbiol. 2013 Jun; 15(6):843–859. [PubMed: 23253353]
- 75. Vogel DYS, Vereyken EJF, Glim JE, Heijnen PDAM, Moeton M, van der Valk P, Amor S, Teunissen CE, van Horssen J, Dijkstra CD. Macrophages in inflammatory multiple sclerosis lesions have an intermediate activation status. J Neurinflamm. 2013; 10:35.
- Brück W, Sommermeier N, Bergmann M, Zettl U, Geobel HH, Kertzschmar HA, Lassmann H. Macrophages in multiple sclerosis. Immunobiology. 1996 Oct; 195(4–5):588–600. [PubMed: 8933159]
- 77. Orme J, Mohan C. Macrophage subpopulations in systemic lupus erythematosus. Discov Med. 2012 Feb; 13(169):151–158. PMID: 22369974. [PubMed: 22369974]
- 78. Li Y, Lee PY, Reeves WH. Monocyte and macrophage abnormalities in systemic lupus erythematosus. Arch Immunol Ther Exp (Warsz). 2010 Oct; 58(5):355–364. [PubMed: 20676788]
- 79. Gate D, Rezai-Zadeh K, Jodry D, Tentsendorj A, Town T. Macrophages in Alzheimer's disease: the-blood-borne identity. J Neural Transm. 2010 Aug; 117(8):961–970. [PubMed: 20517700]
- 80. Olefsky JM, Glass CK. Macrophages, inflammation, and insulin resistance. Annu Rev Physiol. 2010; 72:219–246. [PubMed: 20148674]
- 81. Perry VH. Innate Inflammation in Parkinson's Disease. Cold Spring Harbor Persp in Med. 2012
- 82. Su X, Federoff HJ. Immune Responses in Parkinson's Disease: Interplay between Central and Peripheral Immune Systems. BioMed Res Int. 2014 Apr 13.
- 83. Liu G, Fiala M, Mizwicki MT, Sayre J, Magpantay L, Siani A, Mahanian M, Chattopadhyay M, Le Cava A, Weidau-Pazos M. Neuronal phagocytosis by inflammatory macrophages in ALS spinal cord: inhibition of inflammation by resolving D1. Am J Neurodeger Dis. 2012; 1(1):60–74.
- 84. Graves MC, Fiala M, Dinglasan LA, Liu NQ, Sayre J, Chiappelli F, van Kooten C, Vinters HV. Inflammation in amyotrophic lateral sclerosis spinal cord and brain is mediated by activated macrophages, mast cells and T cells. Amyotroph Lateral Scler Orther Motor Neuron Disord. 2004 Dec; 5(4):213–219. PMID: 15799549.
- 85. Li J, Hsu HC, Mountz JD. Managing macrophages in rheumatoid arthritis by reform or removal. Curr Rheumatol Rep. 2012 Oct; 14(5):445–454. [PubMed: 22855296]
- 86. Krown SE, Lee JY, Dittmer DP. AIDS Malignancy Consortium. More on HIV-associated Kaposi's sarcoma. N Engl J Med. 2008 Jan 31; 358(5):535–536. author reply 536. PMID: 18234764. [PubMed: 18234764]
- 87. Maskew M, Fox MP, van Cutsem G, Chu K, Macphail P, Boulle A, Egger M, Africa FI. Treatment response and mortality among patients starting antiretroviral therapy with and without Kaposi sarcoma: a cohort study. PLoS One. 2013 Jun 5.8(6):e64392. PMID: 23755122. [PubMed: 23755122]

88. Nasti G, Talamini R, Antinori A, Martellotta F, Jacchetti G, Chiodo F, Ballardini G, Stoppini L, Di Perri G, Mena M, Tavio M, Vaccher E, D'Arminio Monforte A, Tirelli U. AIDS-related Kaposi's Sarcoma: evaluation of potential new prognostic factors and assessment of the AIDS Clinical Trial Group Staging System in the Haart Era--the Italian Cooperative Group on AIDS and Tumors and the Italian Cohort of Patients Naive From Antiretrovirals. J Clin Oncol. 2003 Aug 1; 21(15):2876–2882. PMID: 12885804. [PubMed: 12885804]

- Uldrick TS, Whitby D. Update on KSHV epidemiology, Kaposi Sarcoma pathogenesis, and treatment of Kaposi Sarcoma. Cancer Lett. 2011 Jun 28; 305(2):150–162. PMID: 21377267.
 [PubMed: 21377267]
- Maurer T, Ponte M, Leslie K. HIV-associated Kaposi's sarcoma with a high CD4 count and a low viral load. N Engl J Med. 2007 Sep 27; 357(13):1352–1353. PMID: 17898112. [PubMed: 17898112]
- 91. Ma Y, Pope RM. The role of macrophages in rheumatoid arthritis. Curr Pharm Des. 2005; 11(5): 569–580. PMID: 15720276. [PubMed: 15720276]
- Maradit-Kremers H, Crowson CS, Nicola PJ, Ballman KV, Roger VL, Jacobsen SJ, Gabriel SE. Increased unrecognized coronary heart disease and sudden deaths in rheumatoid arthritis: a population-based cohort study. Arthritis Rheum. 2005 Feb; 52(2):402–411. PMID: 15693010. [PubMed: 15693010]
- 93. Carmona L, Cross M, Williams B, Lassere M, March L. Rheumatoid arthritis. Best Pract Res Clin Rheumatol. 2010 Dec; 24(6):733–745. PMID: 21665122. [PubMed: 21665122]
- 94. Oliver JE, Silman AJ. Why are women predisposed to autoimmune rheumatic diseases? Arthritis Res Ther. 2009; 11(5):252. PMID: 19863777. [PubMed: 19863777]
- 95. Helmick CG, Felson DT, Lawrence RC, Gabriel S, Hirsch R, Kwoh CK, Liang MH, Kremers HM, Mayes MD, Merkel PA, Pillemer SR, Reveille JD, Stone JH. National Arthritis Data Workgroup. Estimates of the prevalence of arthritis and other rheumatic conditions in the United States. Part I. Arthritis Rheum. 2008 Jan; 58(1):15–25. 18163481. [PubMed: 18163481]
- Scott DL, Wolfe F, Huizinga TW. Rheumatoid arthritis. Lancet. 2010 Sep 25; 376(9746):1094–1108. PMID: 20870100. [PubMed: 20870100]
- 97. Hootman JM, Helmick CG. Projections of US prevalence of arthritis and associated activity limitations. Arthritis Rheum. 2006; 54:226–229. PMID: 16385518. [PubMed: 16385518]
- 98. Raza K, Falciani F, Curnow SJ, Ross EJ, Lee CY, Akbar AN, Lord JM, Gordon C, Buckley CD, Salmon M. Early rheumatoid arthritis is characterized by a distinct and transient synovial fluid cytokine profile of T cell and stromal cell origin. Arthritis Res Ther. 2005; 7(4):R784–R795. PMID: 15987480. [PubMed: 15987480]
- Olszewski WL, Pazdur J, Kubasiewicz E, Zaleska M, Cooke CJ, Miller NE. Lymph draining from foot joints in rheumatoid arthritis provides insight into local cytokine and chemokine production and transport to lymph nodes. Arthritis Rheum. 2001 Mar; 44(3):541–549. PMID: 11263768.
 [PubMed: 11263768]
- 100. Meyer PW, Hodkinson B, Ally M, Musenge E, Wadee AA, Fickl H, Tikly M, Anderson R. Circulating cytokine profiles and their relationships with autoantibodies, acute phase reactants, and disease activity in patients with rheumatoid arthritis. Mediators Inflamm. 2010; 2010:158514. PMID: 21437211. [PubMed: 21437211]
- 101. Leizer T, Cebon J, Layton JE, Hamilton JA. Cytokine regulation of colony-stimulating factor production in cultured human synovial fibroblasts: I. Induction of GM-CSF and G-CSF production by interleukin-1 and tumor necrosis factor. Blood. 1990 Nov 15; 76(10):1989–1996. PMID: 1700731. [PubMed: 1700731]
- 102. Westra J, Doornbos-van der Meer B, de Boer P, van Leeuwen MA, van Rijswijk MH, Limburg PC. Strong inhibition of TNF-alpha production and inhibition of IL-8 and COX-2 mRNA expression in monocyte-derived macrophages by RWJ 67657, a p38 mitogen-activated protein kinase (MAPK) inhibitor. Arthritis Res Ther. 2004; 6(4):R384–R392. PMID: 15225374. [PubMed: 15225374]
- 103. Kraan MC, Reece RJ, Smeets TJ, Veale DJ, Emery P, Tak PP. Comparison of synovial tissues from the knee joints and the small joints of rheumatoid arthritis patients: Implications for pathogenesis and evaluation of treatment. Arthritis Rheum. 2002 Aug; 46(8):2034–2038. PMID: 12209505. [PubMed: 12209505]

104. Kennedy A, Fearon U, Veale DJ, Godson C. Macrophages in synovial inflammation. Front Immunol. 2011 Oct 10.2:52. PMID: 22566842. [PubMed: 22566842]

- 105. Bresnihan B. Pathogenesis of joint damage in rheumatoid arthritis. J Rheumatol. 1999 Mar; 26(3): 717–719. PMID: 10090189. [PubMed: 10090189]
- 106. Mulherin D1, Fitzgerald O, Bresnihan B. Synovial tissue macrophage populations and articular damage in rheumatoid arthritis. Arthritis Rheum. 1996 Jan; 39(1):115–124. PMID: 8546720. [PubMed: 8546720]
- 107. Deane KD, Norris JM, Holers VM. Preclinical rheumatoid arthritis: identification, evaluation, and future directions for investigation. Rheum Dis Clin North Am. 2010 May; 36(2):213–241. PMID: 20510231. [PubMed: 20510231]
- 108. El-Gabalawy H. The preclinical stages of RA: lessons from human studies and animal models.

 Best Pract Res Clin Rheumatol. 2009 Feb; 23(1):49–58. PMID: 19233045. [PubMed: 19233045]
- 109. Kraan MC, Versendaal H, Jonker M, Bresnihan B, Post WJ, t Hart BA, Breedveld FC, Tak PP. Asymptomatic synovitis precedes clinically manifest arthritis. Arthritis Rheum. 1998 Aug; 41(8): 1481–1488. PMID: 9704648. [PubMed: 9704648]
- 110. Demoruelle MK, Deane KD, Holers VM. When and where does inflammation begin in rheumatoid arthritis? Curr Opin Rheumatol. 2014 Jan; 26(1):64–71. PMID: 24247116. [PubMed: 24247116]
- 111. Azad AK, Rajaram MV, Schlesinger LS. Exploitation of the Macrophage Mannose Receptor (CD206) in Infectious Disease Diagnostics and Therapeutics. J Cytol Mol Biol. 2014 Jan 10.1(1) pii: 1000003. PMID: 24672807.
- 112. Schlesinger LS. Macrophage phagocytosis of virulent but not attenuated strains of Mycobacterium tuberculosis is mediated by mannose receptors in addition to complement receptors. J Immunol. 1993 Apr 1; 150(7):2920–2930. PMID: 8454864. [PubMed: 8454864]
- 113. D'Addio SM1, Baldassano S, Shi L, Cheung L, Adamson DH, Bruzek M, Anthony JE, Laskin DL, Sinko PJ, Prud'homme RK. Optimization of cell receptor-specific targeting through multivalent surface decoration of polymeric nanocarriers. J Control Release. 2013; 168:41–49. PMID: 23419950. [PubMed: 23419950]
- 114. Kang PB, Azad AK, Torrelles JB, Kaufman TM, Beharka A, Tibesar E, DesJardin LE, Schlesinger LS. The human macrophage mannose receptor directs Mycobacterium tuberculosis lipoarabinomannan-mediated phagosome biogenesis. J Exp Med. 2005 Oct 3; 202(7):987–999. PMID: 16203868. [PubMed: 16203868]
- 115. Clemens DL, Horwitz MA. Characterization of the Mycobacterium tuberculosis phagosome and evidence that phagosomal maturation is inhibited. J Exp Med. 1995 Jan 1; 181(1):257–270. PMID: 7807006. [PubMed: 7807006]
- 116. Sturgill-Koszycki S, Schaible UE, Russell DG. Mycobacterium-containing phagosomes are accessible to early endosomes and reflect a transitional state in normal phagosome biogenesis. EMBO J. 1996 Dec 16; 15(24):6960–6968. PMID: 9003772. [PubMed: 9003772]
- 117. Guirado E, Schlesinger LS, Kaplan G. Macrophages in tuberculosis: friend or foe. Semin Immunopathol. 2013 Sep; 35(5):563–583. PMID: 23864058. [PubMed: 23864058]
- 118. Russell DG, Cardona PJ, Kim MJ, Allain S, Altare F. Foamy macrophages and the progression of the human tuberculosis granuloma. Nat Immunol. 2009 Sep; 10(9):943–948. Epub 2009 Aug 19. PMID: 19692995. [PubMed: 19692995]
- 119. Peyron P, Vaubourgeix J, Poquet Y, Levillain F, Botanch C, Bardou F, Daffé M, Emile JF, Marchou B, Cardona PJ, de Chastellier C, Altare F. Foamy macrophages from tuberculous patients' granulomas constitute a nutrient-rich reservoir for M. tuberculosis persistence. PLoS Pathog. 2008 Nov.4(11):e1000204. PMID: 19002241. [PubMed: 19002241]
- 120. Kim MJ, Wainwright HC, Locketz M, Bekker LG, Walther GB, Dittrich C, Visser A, Wang W, Hsu FF, Wiehart U, Tsenova L, Kaplan G, Russell DG. Caseation of human tuberculosis granulomas correlates with elevated host lipid metabolism. EMBO Mol Med. 2010 Jul; 2(7):258–274. PMID: 20597103. [PubMed: 20597103]
- 121. Ehlers S, Schaible UE. The granuloma in tuberculosis: dynamics of a host-pathogen collusion. Front Immunol. 2013 Jan 7.3:411. PMID: 23308075. [PubMed: 23308075]

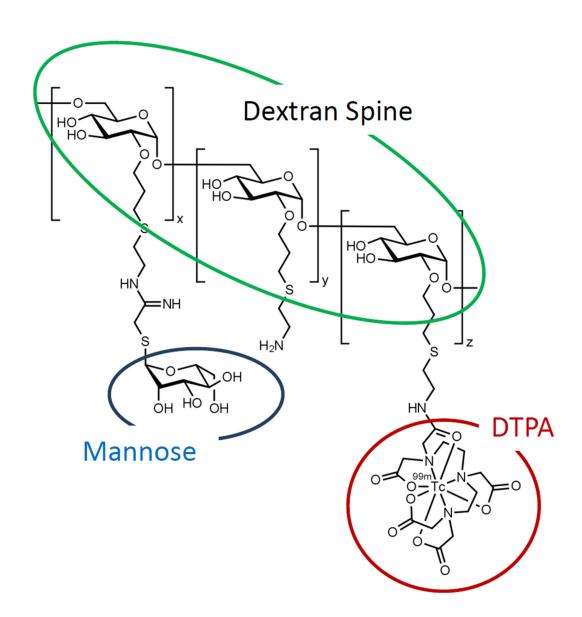
122. Fan J, Watanabe T. Inflammatory reactions in the pathogenesis of atherosclerosis. J Atheroscler Thromb. 2003; 10(2):63–71. PMID: 12740479. [PubMed: 12740479]

- Libby P. Inflammation in atherosclerosis. Arterioscler Thromb Vasc Biol. 2012 Sep; 32(9):2045– 2051. PMID: 22895665. [PubMed: 22895665]
- 124. Schwartz CJ, Valente AJ, Sprague EA, Kelley JL, Nerem RM. The pathogenesis of atherosclerosis: an overview. Clin Cardiol. 1991 Feb; 14(2 Suppl 1):I1–I16. PMID: 2044253. [PubMed: 2044253]
- 125. Liang M, Puri A, Devlin G. The vulnerable plaque: the real villain in acute coronary syndromes. Open Cardiovasc Med J. 2011; 5:123–129. PMID: 21673834. [PubMed: 21673834]
- 126. Kullo IJ, Edwards WD, Schwartz RS. Vulnerable plaque: pathobiology and clinical implications. Ann Intern Med. 1998 Dec 15; 129(12):1050–1060. PMID: 9867761. [PubMed: 9867761]
- 127. Benedek T, Jako B, Benedek I. Plaque quantification by coronary CT and intravascular ultrasound identifies a low CT density core as a marker of plaque instability in acute coronary syndromes. Int Heart J. 2014; 55(1):22–28. PMID: 24463925. [PubMed: 24463925]
- 128. Zhou J, Chew M, Ravn HB, Falk E. Plaque pathology and coronary thrombosis in the pathogenesis of acute coronary syndromes. Scand J Clin Lab Invest Suppl. 1999; 230:3–11. PMID: 10389196. [PubMed: 10389196]
- 129. Falk E, Nakano M, Bentzon JF, Finn AV, Virmani R. Update on acute coronary syndromes: the pathologists' view. Eur Heart J. 2013 Mar; 34(10):719–728. PMID: 23242196. [PubMed: 23242196]
- 130. Moore KJ, Sheedy FJ, Fisher EA. Macrophages in atherosclerosis: a dynamic balance. Nat Rev Immunol. 2013 Oct; 13(10):709–721. PMID: 23995626. [PubMed: 23995626]
- 131. Bekkering S, Quintin J, Joosten LA, van der Meer JW, Netea MG, Riksen NP. Oxidized Low-Density Lipoprotein Induces Long-Term Proinflammatory Cytokine Production and Foam Cell Formation via Epigenetic Reprogramming of Monocytes. Arterioscler Thromb Vasc Biol. 2014 Jun 5. pii: ATVBAHA.114.303887. [Epub ahead of print] PMID: 24903093.
- 132. Sakakura K, Nakano M, Otsuka F, Ladich E, Kolodgie FD, Virmani R. Pathophysiology of atherosclerosis plaque progression. Heart Lung Circ. 2013 Jun; 22(6):399–411. PMID: 23541627. [PubMed: 23541627]
- 133. Narula J, Nakano M, Virmani R, Kolodgie FD, Petersen R, Newcomb R, Malik S, Fuster V, Finn AV. Histopathologic characteristics of atherosclerotic coronary disease and implications of the findings for the invasive and noninvasive detection of vulnerable plaques. J Am Coll Cardiol. 2013 Mar 12; 61(10):1041–1051. PMID: 23473409. [PubMed: 23473409]
- 134. Tahara N, Mukherjee J, de Haas HJ, Petrov AD, Tawakol A, Haider N, Tahara A, Constantinescu CC, Zhou J, Boersma HH, Imaizumi T, Nakano M, Finn A, Fayad Z, Virmani R, Fuster V, Bosca L, Narula J. 2-deoxy-2-[18F]fluoro-D-mannose positron emission tomography imaging in atherosclerosis. Nat Med. 2014; 20:215–219. PMID: 24412923. [PubMed: 24412923]
- 135. Moore KJ, Sheedy FJ, Fisher EA. Macrophages in atherosclerosis: a dynamic balance. Nat Rev Immunol. 2013 Oct; 13(10):709–721. PMID: 23995626. [PubMed: 23995626]
- 136. Martinez FO, Gordon S. The M1 and M2 paradigm of macrophage activation: time for reassessment. F1000Prime Rep. 2014 Mar 3.6:13. eCollection 2014. PMID: 24669294. [PubMed: 24669294]
- 137. Zizzo G, Hilliard BA, Monestier M, Cohen PL. Efficient clearance of early apoptotic cells by human macrophages requires M2c polarization and MerTK induction. J Immunol. 2012 Oct 1; 189(7):3508–3520. PMID: 22942426. [PubMed: 22942426]
- 138. Thorp E, Cui D, Schrijvers DM, Kuriakose G, Tabas I. Mertk receptor mutation reduces efferocytosis efficiency and promotes apoptotic cell accumulation and plaque necrosis in atherosclerotic lesions of apoe-/- mice. Arterioscler Thromb Vasc Biol. 2008 Aug; 28(8):1421– 1428. PMID: 18451332. [PubMed: 18451332]
- 139. Weiss RA. How does HIV cause AIDS? Science. 1993 May 28; 260(5112):1273–1279. PMID: 8493571. [PubMed: 8493571]
- 140. Torres RA, Barr M. Impact of combination therapy for HIV infection on inpatient census. N Engl J Med. 1997 May 22; 336(21):1531–1532. PMID: 9157292. [PubMed: 9157292]

141. Martinez FO, Gordon S. The M1 and M2 paradigm of macrophage activation: time for reassessment. F1000Prime Rep. 2014 Mar 3.6:13. eCollection 2014. PMID: 24669294. [PubMed: 24669294]

- 142. Zizzo G, Hilliard BA, Monestier M, Cohen PL. Efficient clearance of early apoptotic cells by human macrophages requires M2c polarization and MerTK induction. J Immunol. 2012 Oct 1; 189(7):3508–3520. PMID: 22942426. [PubMed: 22942426]
- 143. Thorp E, Cui D, Schrijvers DM, Kuriakose G, Tabas I. Mertk receptor mutation reduces efferocytosis efficiency and promotes apoptotic cell accumulation and plaque necrosis in atherosclerotic lesions of apoe-/– mice. Arterioscler Thromb Vasc Biol. 2008 Aug; 28(8):1421–1428. PMID: 18451332. [PubMed: 18451332]
- 144. Zizzo G, Hilliard BA, Monestier M, Cohen PL. Efficient clearance of early apoptotic cells by human macrophages requires M2c polarization and MerTK induction. J Immunol. 2012 Oct 1; 189(7):3508–3520. PMID: 22942426. [PubMed: 22942426]
- 145. Zanni MV, Abbara S, Lo J, Wai B, Hark D, Marmarelis E, Grinspoon SK. Increased coronary atherosclerotic plaque vulnerability by coronary computed tomography angiography in HIV-infected men. AIDS. 2013 May 15; 27(8):1263–1272. PMID: 23324657. [PubMed: 23324657]

a -



b -

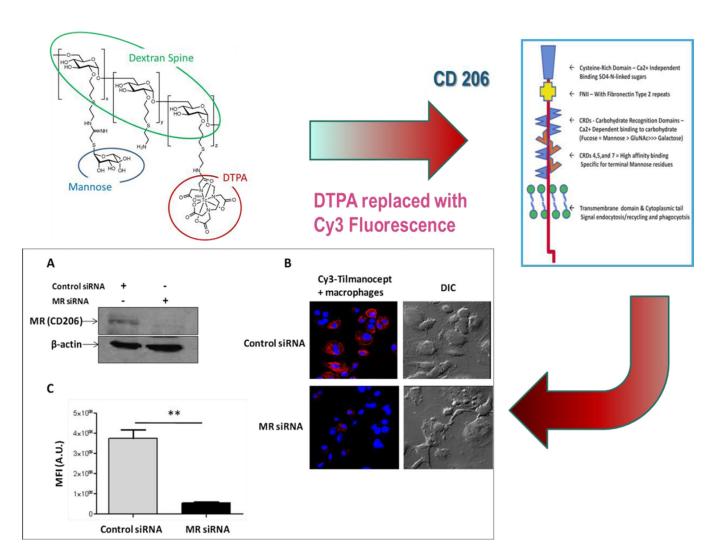


Figure 1. a - ^{99m}Tc-tilmanocept (Lymphoseek[®]; Navidea) b - Tilmanocept Specifically Binds to CD206

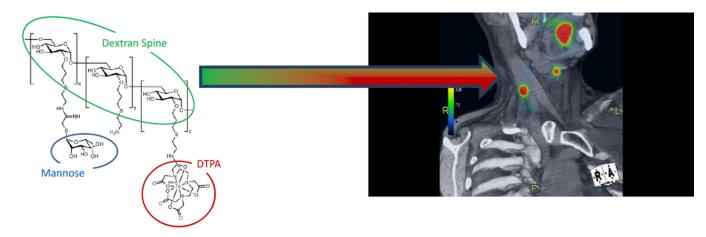


Figure 2.Sentinel node discrimination in patient with oral squamous cell carcinoma of the tongue; multiple sentinel nodes are visible 15 minutes after injection.

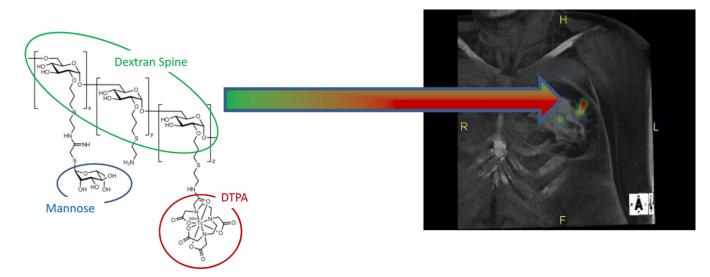


Figure 3.Sentinel node discrimination in patient with breast cancer; multiple sentinel nodes are visible 15 minutes after injection.

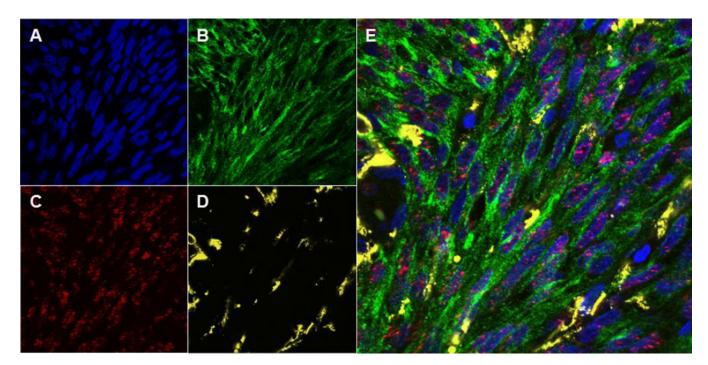


Figure 4.CD206 expresses on both Kaposi's sarcoma spindle cells and associated macrophages (A-Nuclear stain DAPI; B-Anti-CD206; C-Anti-LANA/HHV8; D-Anti-CD68 macrophage marker; E-merged images)".

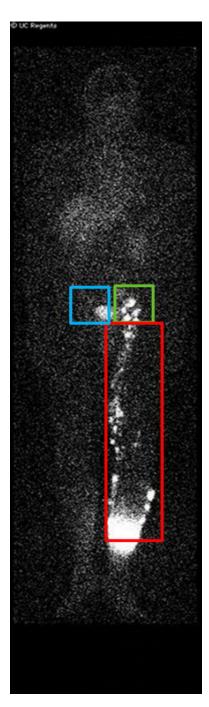


Figure 5. Patient injected subcutaneously with Tc99m-Tilmanocept (50 μ g; 2.0 mCi) and imaged 4 hr post-injection; SPECT WHOLE BODY SCAN: Injection site lower left leg; KS lesions match tracings of leg lesions; TC99m- tilmanocept localizes in KS chain linked by lymphatic ducts; localizes in (KS+?) inguinal nodes; Ift of nodes in image is bladder

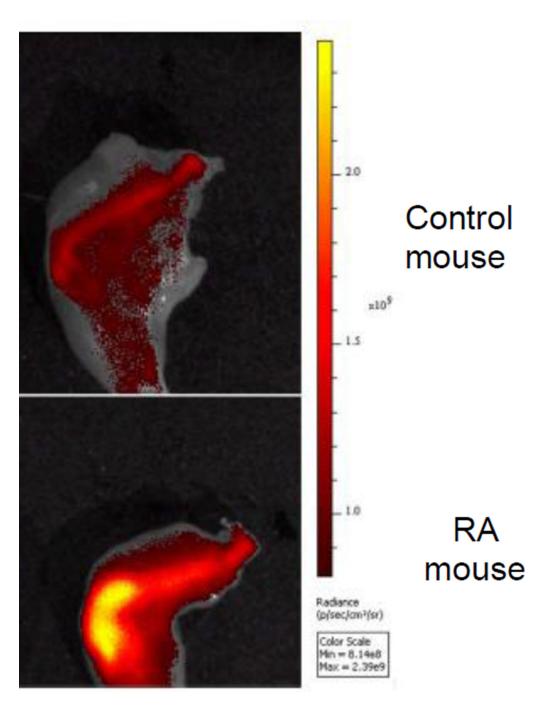


Figure 6. Mice were imaged in the IVIS (in vivo imaging system) to measure Cy-3 levels. In these ex vivo images, the limbs were detached and the skin was removed before imaging. On the bottom you can see the enhanced signal in the RA mice knees. The values are statistically significant when quantitated.

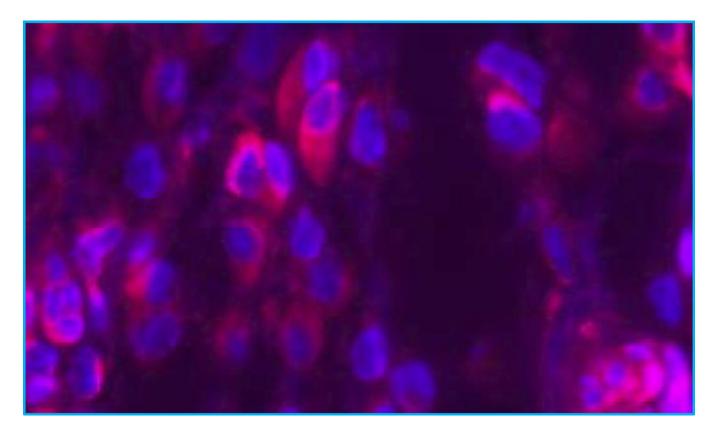


Figure 7. Staining of synovial tissue frozen sections from a patient with Rheumatoid arthritis. Sections were stained with 4',6-diamidino-2-phenylindole, BLUE (DAPI = DNA) and Cy3-tilmanocept – RED/PINK (CD206) and examined under fluorescence microscopy.

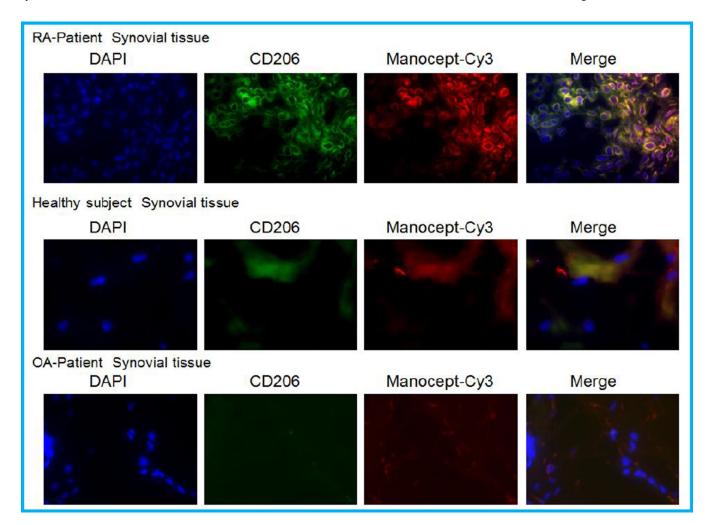
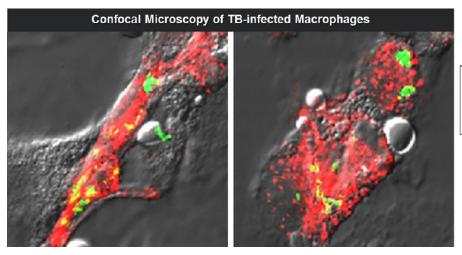


Figure 8. we then examined CD206 and Manocept in synovial tissue. In the merged image, you can clearly see the co-localization of CD206 and Manocept. Healthy and osteoarthritis synovial tissue samples were negative for both.



Red: Tilmanocept-Cy3
Green: GFP-M. tuberculosis
Yellow: Tilmanocept-Cy3 & M.
tuberculosis (TB) co-localization

Figure 9.Human monocyte-derived macrophages in monolayer culture that make up the components of the TB granulomas are infected with a GFP-expressing M. tuberculosis which is internalized by macrophages. The infected cells were exposed to tilmanocept-Cy3 and then analyzed by confocal microscopy.

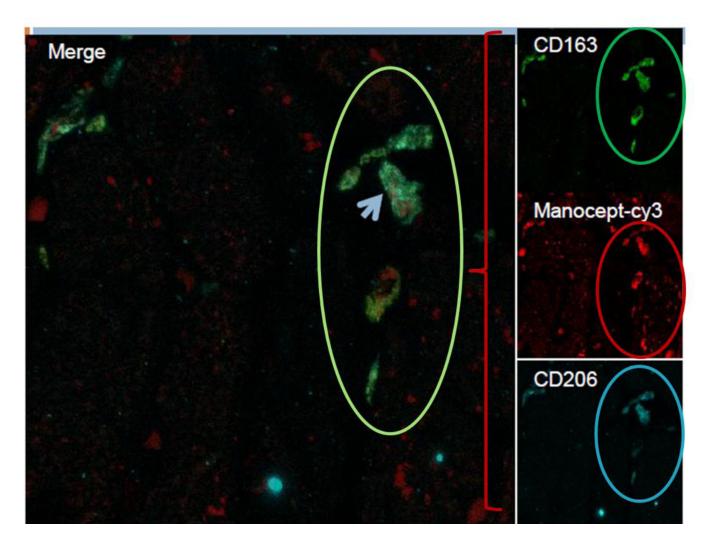


Figure 10.Cy3-Tilmanocept targets macrophages in arteries with atherosclerosis; figure shows colocalization CD206 antibodies (blue), Cy-3-tilmanocept (red) and CD163 (green) macrophage markers in arteries of SIV-infected macaque monkeys.

Table 1
Sentinel Lymph Node False Negative Rate (FNR) in Head and Neck Squamous Cell Carcinoma

		Pathology	
	Lymphoseek Detection	+ 1 Lymph Nodes (SLN or non-SLN) ^I Are Pathology Positive	All Lymph Nodes (SLN and non-SLN) Are Pathology Negative
+	1 Lymphoseek Detected Lymph Node(s) (SLNs) Are Pathology Positive	38 True Positive	N/A ² (0)
_	Lymphoseek Detected Lymph Nodes (SLNs) Are ALL Pathology Negative (or no SLNs exist)	1 False Negative ^{3,4}	44 True Negative

 $^{^{1}}$ SLN=sentinel lymph node; non-SLN=non-sentinel lymph node.

 $^{^{2}}$ N/A=Not applicable. False positives were no applicable to the analysis as lymph nodes did not fit into this category (Nodes cannot both be pathology positive and pathology negative).

³The FNR was 0.0256 and the exact binomial test of this result against the null hypothesis H0: FNR 0.14 was statistically significant at p = 0.0205. Of the 39 patients with at least 1 pathology-positive lymph node, Lymphoseek detected nodes with positive pathology in all but 1 patient.

⁴Due to a positive (low) FNR of an interim-analysis significance level for a 1-sided exact test of binomial proportion being 0.02486 after 38 patients with a pathology-positive lymph node, the DSMC recommended stopping the trial for efficacy (which ended in approximately 40 months).

Table 2

Macrophages in the Natural History of Disease

Innate Immunity		
Allergy		
Asthma		
Atherosclerosis		
Cancer		
COPD		
Diabetes		
Infection/HIV		
Rheumatoid Arthritis		
Sepsis		
Transplant Rejection		