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**Original Research** 



# Recombinant Alpha-1 Antitrypsin–Fc Fusion Protein INBRX-101 in Adults With Alpha-1 Antitrypsin Deficiency: A Phase 1 Study

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# Abstract

**Background:** Alpha-1 antitrypsin deficiency (AATD) is characterized by low alpha-1 antitrypsin (AAT) levels, predisposing individuals to lung disease. The standard of care, plasma-derived AAT (pdAAT), is delivered as weekly infusions to maintain serum AAT concentrations  $\geq 11 \mu M$  ( $\approx 50\%$  of those in healthy individuals). INBRX-101, a recombinant human AAT-Fc fusion protein, was designed to have a longer half-life and achieve higher AAT levels than pdAAT.

**Methods:** In this phase 1 dose-escalation study (N=31), adults with AATD received 1 dose (part 1) or 3 doses (part 2) of 10 (part 1), 40, 80, or 120mg/kg INBRX-101 every 3 weeks (Q3W) via intravenous infusion. The primary endpoint was safety and tolerability. Secondary endpoints were pharmacokinetics (PK), pharmacodynamics (PD), and immunogenicity of INBRX-101.

**Results:** INBRX-101 was well tolerated. Most treatment-emergent adverse events were grade  $\leq 2$ . In part 2 (n=18; each dose, n=6), dose-related increases in serum functional AAT (fAAT) were observed; mean fAAT levels remained above the 21µM target for up to 4 weeks after the final dose in the 120-mg/kg cohort. Antidrug antibodies had no meaningful impact on PK or PD. INBRX-101 was detected in pulmonary epithelial lining fluid (PELF) from all patients assessed (n=11), and PELF fAAT increased after dosing. PK/PD modeling projected steady-state serum fAAT $\geq$ 21µM at 120mg/kg Q3W (average concentration  $\approx$ 43µM; trough concentration  $\approx$ 28µM) and Q4W ( $\approx$ 34µM;  $\approx$ 21µM).

**Conclusion:** The favorable safety profile and ability to maintain serum fAAT levels  $>21\mu$ M with extended-interval dosing, support a phase 2 trial evaluating Q3W and Q4W dosing of INBRX-101.

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### Abbreviations:

**AAT**=alpha-1 antitrypsin; **AATD**=alpha-1 antitrypsin deficiency; **ADA**=antidrug antibody; **AE**=adverse event; **AUC**<sub>0-7</sub> days,ss</sub>=steadystate area under the plasma drug concentration-time curve, days 0 to 7; **BALF**=bronchoalveolar lavage fluid; **C**<sub>avg</sub>=average concentration; **C**<sub>avg,ss</sub>=average concentration at steady state; **C**<sub>max</sub>=maximum concentration; **C**<sub>max,ss</sub>=maximum concentration at steady state; **C**<sub>trough,ss</sub>=trough concentration at steady state; **CT**=computed tomography; **DLCO**=diffusing capacity of lung for carbon monoxide; **fAAT**=functional alpha-1 antitrypsin; **FEV**<sub>1</sub>=forced expiratory volume in 1 second; **NE**=neutrophil elastase; **PD**=pharmacodynamics; **pdAAT**=human plasma-derived alpha-1 antitrypsin; **PELF**=pulmonary epithelial lining fluid; **PK**=pharmacokinetics; **Q1W**=once weekly; **Q3W**=every 3 weeks; **Q4W**=every 4 weeks; **SD**=standard deviation

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#### This article has an online supplement.

## Introduction

Alpha-1 antitrypsin deficiency (AATD) is a rare, chronic genetic disorder characterized by low serum levels of the serine protease inhibitor alpha-1 antitrypsin (AAT). In AATD, neutrophil elastase (NE) activity in the lungs is insufficiently controlled by AAT, which can result in progressive lung damage and pulmonary complications. Patients with severe AATD (e.g., *SERPINA1* genotypes PiZZ, PiZ/Null, and PiNull/Null) have serum AAT concentrations <15%–20% of those in individuals with the wild-type genotype (PiMM); individuals with moderate deficiency (e.g., PiMZ and PiSZ) may have levels  $\geq$ 49% of those in individuals with the PiMM genotype.<sup>1-5</sup> Generally, lower serum AAT levels are correlated with a greater risk for pulmonary deterioration, poorer quality of life, and an increased mortality rate.<sup>5-10</sup>

The standard of care and only specific treatment available for AATD is augmentation therapy using human plasma– derived AAT (pdAAT) at  $60 \text{mg/kg.}^{2,10,11}$  With a mean half-life of 5–7 days,<sup>12-16</sup> once-weekly infusion is required, creating a considerable burden for patients.<sup>8,17</sup> Informed by studies from the 1980s, augmentation therapy with pdAAT aims to increase serum AAT levels above a theoretically "protective" threshold of 11µM.<sup>12,13,18-21</sup> The relevance and validity of this threshold, which is approximately 50% below the lower limit of serum AAT levels in individuals with the PiMM genotype (range, 20–53µM),<sup>22</sup> have been increasingly questioned.<sup>23-28</sup> However, studies of higher dosing and the impact of elevating serum AAT levels further, including in patients with more moderate deficiency of serum AAT (e.g., PiMZ genotype), have historically been limited by the high cost of pdAAT and concerns about diverting limited supply from patients with severe deficiency.<sup>2,26,27,29</sup>

Evidence suggests that achieving higher AAT levels closer to those observed in PiMM individuals may lead to improved clinical outcomes. In the randomized, placebocontrolled trial RAPID, a post hoc analysis found an association between higher median trough levels of total serum AAT (90th percentile, >20µM) and a slower annual rate of lung density decline.<sup>30</sup> Notably, this relationship between response and trough levels did not plateau, suggesting that higher serum AAT levels could further improve response. In a small study,<sup>31</sup> a weekly double dose of pdAAT (120mg/kg), for which mean total serum AAT trough levels were within the range observed in PiMM individuals (20-53µM),<sup>22,32</sup> led to greater reductions in bronchoalveolar lavage fluid (BALF) and plasma protease activity, BALF elastin degradation, and markers of airway inflammation compared with the standard dosing (60mg/kg weekly).

As total serum AAT levels with pdAAT 60mg/kg once weekly typically fall below 20µM (the lower limit of the range observed in PiMM individuals<sup>22</sup>) within 3-5 days of dosing,<sup>19,21,32</sup> therapies are needed that lead to higher AAT levels with less frequent dosing. INBRX-101 is a recombinant human AAT-Fc fusion protein designed to overcome the limitations of pdAAT.<sup>33,34</sup> INBRX-101 is composed of 2 recombinant human AAT molecules covalently linked to a human IgG4 Fc region and was optimized to have a longer half-life and higher therapeutic exposure (i.e., achieve higher functional AAT [fAAT] levels as measured by NE inhibition) than pdAAT (Figure 1A). The hinge and Fc regions have been modified for hinge stabilization, effector function silencing, and extended plasma half-life. Amino acid changes in the AAT reactive site loop mitigate loss of activity due to oxidation without impacting specificity for NE. In preclinical studies, INBRX-101 demonstrated NE inhibitory capacity comparable to that of pdAAT in vitro and in vivo.33 Importantly, INBRX-101 could penetrate the lungs in a murine model of pulmonary inflammation. Building upon these findings, a phase 1 study of INBRX-101 in patients with AATD was conducted. We present findings from the final analysis of this trial.

## Methods

## **Study Design and Patients**

This open-label, multicenter, 2-part, phase 1, doseescalation study evaluated INBRX-101 in adults with AATD (Supplementary Figure 1 in the online supplement) (ClinicalTrials.gov identifier, NCT03815396). In part 1 (single ascending dose escalation), patients received 1 dose of 10, 40, 80, or 120mg/kg INBRX-101. In part 2 (multiple ascending dose escalation), patients received 3 doses of 40, 80, or 120mg/kg every 3 weeks (Q3W). Patients from

Figure 1. INBRX-101 (A), Serum Pharmacokinetics Profiles of INBRX-101 (B), and Functional Alpha-1 Antitrypsin (C) in Part 2



(A) Structure and key features of INBRX-101 are shown. (B and C) Arrows indicate dosing. Patients were assessed weekly from day 7 to day 42 to reduce patient burden. No PK samples were collected immediately following the second dose (day 21). The range for fAAT<sup>34</sup> shown in panel C was determined in 65 healthy volunteers with the PiMM genotype (see Methods).

AAT=alpha-1 antitrypsin; Q3W=every 3 weeks; PK=pharmacokinetic; fAAT=functional alpha-1 antitrypsin

part 1 could continue to part 2; participation in part 1 was not required for part 2. INBRX-101 was administered by intravenous infusion over 30 or 45 minutes. The primary endpoint was safety and tolerability of INBRX-101 in patients who received  $\geq 1$  dose of INBRX-101 (safety population). Adverse events (AEs) were graded using the National Cancer Institute Common Terminology

Criteria for Adverse Events, version 5.0, and coded using the Medical Dictionary for Regulatory Activities, version 20.1. Secondary endpoints included: (1) pharmacodynamics (PD) of fAAT in serum, (2) pharmacokinetics (PK) of INBRX-101 in serum and pulmonary epithelial lining fluid (PELF), (3) immunogenicity of INBRX-101, and (4) PD of fAAT in PELF. The PK population comprised all dosed patients with sufficient serum concentrations for PK evaluations; the PD population included dosed patients with  $\geq 1$  measurable PD value.

Eligible patients were aged 18-80 years (18-75 years in the 80- and 120-mg/kg cohorts in part 2) with a diagnosis of AATD (any allelic combination except those resulting in a null/null genotype) and previously documented antigenic serum AAT level of <11µM prior to starting pdAAT augmentation therapy. A washout period of ≥4 weeks before study dosing was required for patients receiving augmentation therapy; dosing could not be resumed for the duration of the study, including follow-up. Smoking was not allowed for the study duration, and patients had to be nonsmokers for  $\geq 6$  months. All patients provided written informed consent before participation. This study was conducted in compliance with the protocol, Declaration of Helsinki, the International Council for Harmonisation Guidelines for Good Clinical Practice, and applicable national and local laws and regulatory requirements. The protocol was approved by the independent ethics committee or institutional review board at each site (Supplementary Table 1 in the online supplement).

## Procedures

Concentrations of INBRX-101 in serum samples were determined using a validated antigenic plate-based assay. A liquid chromatography multiple reaction monitoring mass spectrometry-based method was used to quantify INBRX-101 in human PELF. PK parameters were estimated via noncompartmental analysis using Phoenix WinNonlin version 8.3 (Certara, Radnor, Pennsylvania).

fAAT levels in serum and PELF were determined using validated anti-NE capacity methods. AAT calibrators (Prolastin-C), quality controls, and samples were incubated with a fixed amount of human NE; a fluorogenic NE substrate (N-methoxysuccinyl-Ala-Ala-Pro-Val-7-amino-4-methylcoumarin) was added and the fluorescent signal measured over 10 minutes. To estimate a preliminary range of fAAT in individuals with wild-type *SERPINA1* genotype PiMM, serum samples from 65 healthy volunteers (verified as PiMM by the A1ALC test at Mayo Clinic Laboratories) were assayed using the same validated assay.

BALF was collected from patients in the 80- and 120mg/kg cohorts in part 2 at screening and day 56 (i.e., 2 weeks after the third dose of INBRX-101); 3 lobes were sampled at each time point. Bronchoscopies were performed at 5 of the study sites by qualified medical staff per the Inhibrx-specified Bronchoscopy Procedure Manual; BALF was assessed by 3 specialized laboratories.

Immunogenicity to INBRX-101 and AAT were separately determined using traditional 3-tiered bridging electrochemiluminescence assays with sample pretreatment steps to improve tolerance to native AAT and INBRX-101. Characterization of antidrug antibody (ADA) domain specificity to the Fc/hinge portion of INBRX-101 was performed in a modified confirmatory tier using a domainspecific competitive reagent. Neutralizing antibodies to INBRX-101 and AAT were characterized using validated enzymatic activity assays.

Monolix 2021R1 and Simulx software (Lixoft, Antony, France) were used for population PK modeling and simulations. A 2-compartment model with linear elimination was used to describe INBRX-101 PK. Prediction-corrected Visual Predictive Checks and summaries of simulation results were created using RStudio (2022.02.3) and R (version 4.1.0). To simulate fAAT levels for phase 2 dose selection, a distribution of baseline fAAT values was generated based on the pooled mean (6.24µM) and standard deviation values (3.79µM) of baseline fAAT levels in patients with AATD in this phase 1 study and previously published analyses.<sup>30</sup> PK-fAAT time course data from 500 individuals were simulated; 250 individuals with total baseline serum AAT levels of  $\leq 11\mu$ M were randomly sampled for further summary.

## Results

## Patients

Overall, 31 patients were administered INBRX-101. In part 1, 24 patients were enrolled and received 1 intravenous infusion of 10, 40, 80, or 120mg/kg INBRX-101 (each n=6). In part 2, 18 patients (rollover patients from part 1, n=11) received 3 doses of 40, 80, or 120mg/kg INBRX-101 Q3W (each n=6). Per study criteria, patients had documented antigenic serum AAT of <11 $\mu$ M in their medical history. All except 5 patients had a PiZZ genotype, and baseline serum fAAT concentrations were 2.0–17.7 $\mu$ M (Table 1). Approximately half of patients had evidence of emphysema (51.6%), and the median forced expiratory volume in 1 second was 81.0% of predicted. Six patients (19.4%) had previously received pdAAT.

All patients completed dosing, and most completed the study (n=26 [83.9%]; data cutoff, September 26, 2022). Five patients discontinued (consent withdrawal, n=1; inability to complete study visits, n=4 [attributed to the COVID-19 pandemic, n=2]).

## Safety

AEs were reported in 28 of 31 patients (90.3%) (Table 2). Most were transient and self-limited, and only 1 grade  $\geq$ 3 AE was

## **Table 1. Patient Demographics and Baseline Characteristics**

Characteristics	INBRX-101 (N=31)	
Age, Median (Range), Years	55 (37–76)	
Sex, n (%)		
Female	19 (61.3)	
Male	12 (38.7)	
Race, n (%)		
White	30 (96.8)	
Other	1 (3.2)	
Genotype, n (%)ª		
PiZZ	26 (83.9)	
PiSZ	3 (9.7)	
PiMZ	2 (6.5)	
History of Smoking, n (%)	16 (51.6)	
Emphysema, n (%) <sup>b</sup>	16 (51.6)	
Prior Treatment With pdAAT, n (%)	6 (19.4)	
Functional AAT Serum Concentration, Median (Range), µM	4.91 (2.0–17.7) <sup>c</sup>	
FEV <sub>1</sub> , Median (Range), % of Predicted	81.0 (20.1–104.0)	

<sup>a</sup>As determined by phenotyping of AAT protein in plasma/serum by Mayo Clinic Laboratories.

<sup>b</sup>Evidence of emphysema per DLCO and CT.

<sup>c</sup>Patients were allowed to enroll in the trial if they had documented an antigenic AAT level of <11 µM in their past medical history. In the 26 patients with a PiZZ genotype, the median functional AAT serum concentration at baseline was 4.65 µM (range, 2.0–16.8 µM).

pdAAT=human plasma-derived alpha-1 antitrypsin; AAT=alpha-1 antitrypsin; FEV1=forced expiratory volume in 1 second; DLCO=diffusing capacity of lung for carbon monoxide; CT=computed tomography

reported (progression of preexisting follicular lymphoma). Fatigue was most common, occurring in 29.0% of patients (treatment related in 7 of 9 patients), followed by upper respiratory tract infections (16.1%). Treatment-related AEs, as determined by the investigator, occurred in 14 patients and were all grade 1 or 2. In 2 patients, serious AEs, both unrelated to the study drug, were reported: the lymphoma described above and sinus tachycardia grade 2, which resolved without treatment. No AEs led to treatment discontinuation or death.

AEs considered to be infusion related occurred in 8 patients. All were grade 1 or 2 and included pruritus, flushing, and urticaria (Supplementary Table 2 in the online supplement). Grade 2 infusion-related AEs were reported in 3 patients but were manageable and resolved on the same day; 2 patients received diphenhydramine and/or famotidine as supportive care. No eosinophil elevations were observed in these patients, and reported event terms suggested that infusion-related events were not due to INBRX-101 hypersensitivity.

## Pharmacokinetics and Immunogenicity

In part 1, INBRX-101 maximal concentration was achieved at or near the end of infusion; dose-related increases in maximal and total INBRX-101 exposure were observed across evaluated doses. The mean terminal elimination half-life was between 12.2 (10mg/kg) and 20.0 days (120mg/kg).

In part 2, similar trends were observed, with doseproportional increases in plasma area under the INBRX-101 concentration-time curve and maximal concentration (Figure 1B; Supplementary Table 3 in the online supplement). The mean terminal elimination half-life was 15.7 (40mg/kg) to 18.2 days (120mg/kg). Measurable INBRX-101 exposure was observed in all 3 cohorts for  $\geq 28$  days after the third dose.

Of the 29 patients evaluable for immunogenicity, ADAs against INBRX-101 were confirmed in 9 patients (treatment emergent, n=6; preexisting, n=3), of whom 1 developed low-titer neutralizing antibodies. Five patients had ADAs (treatment emergent, n=4; preexisting, n=1) reactive to INBRX-101 and AAT; none were neutralizing to AAT. Four patients still had ADAs (treatment emergent, n=2; preexisting, n=2) against INBRX-101 at their final testing.

ADA titers were low and had no meaningful impact on INBRX-101 PK or PD, including in the patient with neutralizing antibodies. PK parameters were comparable between patients regardless of ADA status. ADAs did not result in accelerated mean clearance of INBRX-101 (ADApositive patients,  $0.46mg/(day \times \mu M)/kg$ ; ADA-negative patients,  $0.49mg/(day \times \mu M)/kg$ ).

## Pharmacodynamics of Serum Functional Alpha-1 Antitrypsin

As measured by changes in serum NE inhibitory activity, dose-related increases in serum fAAT were observed in parts 1 and 2 (Figure 1C). Accumulation of serum fAAT corresponded with the long terminal elimination half-life of INBRX-101. For the duration of the dosing interval,

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## **Table 2. Adverse Event Summary**

	INBRX-101 (N=31)		
	Any Grade	Grade ≥3	
Treatment-Related AEs in ≥5% of Patients <sup>a</sup>			
Any AE	14 (45.2)	0	
Fatigue	7 (22.6)	0	
Blood Pressure Increased	4 (12.9)	0	
Pruritus	3 (9.7)	0	
Urticaria	2 (6.5)	0	
All-Cause AEs in ≥10% of Patients			
Any AE	28 (90.3)	1 (3.2)	
Fatigue	9 (29.0)	0	
Upper Respiratory Tract Infection	5 (16.1)	0	
Blood Pressure Increased	4 (12.9)	0	
Headache	4 (12.9)	0	
Coronavirus Infection	4 (12.9)	0	

All values are n (%).

<sup>a</sup>Reported by the investigator as possible, probable, or very likely/certainly related to INBRX-101.

#### AE=adverse event

80 and 120mg/kg Q3W INBRX-101 resulted in serum fAAT levels well above the historical target threshold of 11µM antigenic AAT and above 21µM, which was the lower limit of fAAT levels that we estimated previously using our validated fAAT assay in a pilot study of PiMM individuals (range,<sup>34</sup> 21-54µM). The mean serum fAAT level 21 days after the third dose of INBRX-101 (study day 63) was 35.64µM in the 80-mg/kg cohort and 29.82µM in the 120-mg/kg cohort; mean baseline concentrations were 10.76µM and 8.73µM, respectively (Table 3). The mean serum fAAT level remained >21 $\mu$ M in the 120-mg/kg dose cohort (27.28µM) even 28 days after the third dose (study day 70). The PD profile of serum fAAT in the 26 patients with a PiZZ genotype (Supplementary Figure 2 in the online supplement) was comparable to that observed in the total patient population. ADAs did not impact serum fAAT levels over time (Supplementary Figure 3 in the online supplement).

## INBRX-101 and Functional Alpha-1 Antitrypsin in Pulmonary Epithelial Lining Fluid

BALF samples were collected before and after dosing in part 2 from 11 patients (80mg/kg, n=5; 120mg/kg, n=6) to determine INBRX-101 and fAAT concentrations in PELF. Using a specific validated mass spectrometry assay, INBRX-101 was detected at day 56 (i.e., 14 days after the third dose) in PELF from all assessed lung lobes of every patient evaluated (Figure 2A). INBRX-101 was also detected in predose samples from part 1 rollover patients, even though  $\geq 84$  days had elapsed since their single dose in part 1. Dose-dependent increases of INBRX-101 were seen in PELF, with higher levels on average in patients administered 120 versus 80mg/kg. INBRX-101 remained active in PELF, with fAAT levels in PELF generally higher post baseline, as measured using a validated anti-NE capacity assay (Figure 2B).

## Phase 2 Dose Determination

We used PK/PD modeling and simulation to predict INBRX-101 and fAAT levels following steady-state dosing of 120mg/kg INBRX-101 Q3W or every 4 weeks (Q4W). Our modeling projected 120mg/kg Q4W would achieve a mean steady-state average concentration of  $\approx 34 \mu$ M and a mean steady-state trough concentration of  $\approx 21 \mu$ M (Figure 3)<sup>35</sup> versus  $\approx 43 \mu$ M and  $\approx 28 \mu$ M, respectively, with Q3W dosing. Based on these projections and the results from part 2, both dosing schedules will be evaluated in a pivotal phase 2 trial.

## Discussion

This study evaluated the safety of a novel recombinant AAT, INBRX-101, in patients with AATD. INBRX-101 was designed to overcome the limitations of pdAAT, with an improved PK profile, including a longer half-life. In part 2 of this phase 1 study, INBRX-101 exhibited a mean half-life  $\approx 2.5$  to >3 times of that documented for pdAAT (16–18 days versus 5–7 days<sup>12-16</sup>). The greater exposure to INBRX-101 corresponded with dose-dependent increases in serum fAAT levels. As INBRX-101 contains 2 AAT molecules, the serum concentration of fAAT was approximately double that of INBRX-101, with some deviation from a 2:1 ratio expected due to the contributions of endogenous fAAT. Consistent with the prolonged half-life of INBRX-101, accumulation of INBRX-101, and consequently serum functional AAT

# Table 3. Functional Serum Alpha-1 Antitrypsin Levels

Serum Functional AAT Level, Mean (SD), µM	Part 2 Cohort Dose, mg/kg Q3W		
	40 (n=6)	80 (n=6)	120 (n=6)
Baseline	7.24 (3.14)	10.76 (6.95)	8.73 (6.24)
Day 63	14.16 (3.90)	35.64 (11.39) <sup>a</sup>	29.82 (7.02)
Day 70	15.06 (5.10)	19.74 (3.76) <sup>b</sup>	27.28 (5.25)
C <sub>max</sub> Following Third Dose of INBRX-101	31.7 (6.61)	59.2 (16.9)	74.3 (20.9)
C <sub>avg</sub> , Days 42–63	18.4 (2.6)	39.9 (8.9)	43.3 (8.4)

<sup>a</sup>Data shown are from 4 patients. Two patients could not complete the day 63 study visit due to COVID-19-related restrictions.

<sup>b</sup>Data shown are from 4 patients. Two patients were not able to attend the day 70 study visit.

AAT=alpha-1 antitrypsin; SD=standard deviation; Q3W=every 3 weeks; Cmax=maximum concentration; Cavg=average concentration

# Figure 2. INBRX-101 (A) and Functional Alpha-1 Antitrypsin Levels (B) in Pulmonary Epithelial Lining Fluid



Baseline samples were acquired prior to the first dose in part 2 of the study regardless of whether patients had participated in part 1; the concentration in PELF was determined by urea normalization of results from bronchoalveolar lavage fluid. Horizontal lines indicate mean.

(A) Each point represents the mean INBRX-101 concentration measured across 3 lobes for 1 patient. Stability testing of frozen INBRX-101 in PELF is ongoing to cover the longest storage duration of tested samples. The overall mean concentrations (SD) of INBRX-101 in PELF in units of µg/mL for the 80- and 120-mg/kg cohorts were 42 (29) and 85 (61), respectively.

(B) For each of the 10 evaluable patients, change in functional PELF AAT level was determined for each lobe that had matched PELF samples from baseline and day 56. Shown are data from the lobe with the greatest change in functional PELF AAT levels from baseline (set as 0µM) to day 56. For the 1 patient (genotype PiMZ) in whom functional AAT levels decreased over time in all 3 lobes evaluated, data are shown for the lobe with the smallest decrease. The mean (SD) change from baseline in PELF functional AAT concentration in units of µM at day 56 was 1.34 (1.15).

PELF=pulmonary epithelial lining fluid; AAT=alpha-1 antitrypsin; SD=standard deviation

## Figure 3. Pharmacokinetics and Pharmacodynamics Modeling to Determine Phase 2 Dosing

	Regimen		
PD parameter, mean (SD), μΜ	pdAAT 60 mg/kg Q1W	INBRX-101 120 mg/kg Q3W⁰	INBRX-101 120 mg/kg Q4W <sup>c</sup>
C <sub>trough,ss</sub>	13.6 (3.0) <sup>a</sup>	28.0 (7.8)	20.9 (7.8)
$C_{avg,ss}$	17.8 <sup>b</sup>	43.0 (7.9)	33.7 (6.6)
C <sub>max,ss</sub>	34.5 <sup>b</sup>	65.9 (14.4)	57.2 (14.8)



Values shown in the table represent serum fAAT concentrations. The range shown in the plot was determined in 65 healthy volunteers with the PiMM genotype.<sup>34</sup> All PK and PD parameters for the INBRX-101 dosing regimens were estimated with relatively good precision, and goodness-of-fit and visual predictive checks demonstrated that the model adequately fit the data. The predicted fAAT concentrations shown represent the contribution of endogenous AAT plus INBRX-101.

#### <sup>a</sup>Value for Prolastin-C.<sup>35</sup>

<sup>b</sup>Mean C<sub>avg,ss</sub> was calculated by dividing the functional AAT AUC<sub>0-7 days,ss</sub> of lyophilized Prolastin-C expressed in mg×h/mL by 168 hours (7 days). Values originally presented in mg/mL were converted to molarity (1mg/mL equivalent to 19.2µM).<sup>12</sup>

PD=pharmacodynamic; SD=standard deviation; pdAAT=human plasma–derived alpha-1 antitrypsin; Q1W=once weekly; Q3W=every 3 weeks; Q4W=every 4 weeks; Ctrough, ss=trough concentration at steady state; Cavg, ss=average concentration at steady state; Cmax, ss=maximum concentration at steady state; AAT=alpha-1 antitrypsin; fAAT=functional alpha-1 antitrypsin; PK=pharmacokinetic; AUC<sub>0-7 days,ss</sub>=steady-state area under the plasma drug concentration-time curve, days 0 to 7

activity, was observed following each dose. The mean average concentration of serum fAAT over the 21-day interval following the third dose of INBRX-101 in the 80- and 120-mg/kg cohorts (39.9 $\mu$ M and 43.3 $\mu$ M, respectively) was well above the 21 $\mu$ M lower limit of serum fAAT levels that was observed in PiMM individuals. In patients who received 120mg/kg INBRX-101, mean serum fAAT levels continued to be >21 $\mu$ M at 21 and 28 days after the third dose (29.8 $\mu$ M and 27.3 $\mu$ M, respectively). In contrast, for pdAAT, a mean steady-state trough concentration of 16.7 $\mu$ M to 16.9 $\mu$ M has been reported for total antigenic serum AAT, of which only a portion is functional.<sup>12</sup>

Importantly, INBRX-101 penetrated the lungs and was detected in all 3 lobes assessed in each of the 11 evaluable patients. INBRX-101 was detected prior to dosing in part 2 only in patients dosed during part 1, confirming our assay's specificity. Mean INBRX-101 PELF concentrations were greater in patients who received 120mg/kg INBRX-101, and in general, fAAT levels increased following INBRX-101 administration. The latter suggests that INBRX-101 contributes to the greater anti-NE activity in PELF, but extending these analyses to a larger patient population is the subject of future research.

INBRX-101 was generally well tolerated, with only 1 grade  $\geq$ 3 AE reported (progression of preexisting follicular lymphoma). This was expected given that pdAAT has not been associated with overt toxicity, even at doses of >60mg/kg,<sup>32,36-40</sup> and serum AAT levels can become considerably elevated as part of the body's natural immune response.<sup>41,42</sup> All patients completed dosing; 5 discontinued before completing all study visits for reasons unrelated to INBRX-101. As INBRX-101 is a recombinant protein, infusion-related AEs were of special interest. Such AEs were reported in 8 patients, were all grade 1 or 2, resolved on the same or next day with no to minimal supportive care, and appeared to be a function of infusion volume versus hypersensitivity to INBRX-101. The incidence of ADAs against INBRX-101 was low overall, occurring in 9 of 29 evaluable patients. ADA titers were low and had no observed impact on serum INBRX-101 PK or fAAT PD.

pdAAT has been the standard of care for patients with severe AATD since the 1980s; however, it is associated with several limitations. As a consequence of its relatively short half-life,<sup>12,13</sup> patients need weekly infusions, which can negatively impact quality of life and affect treatment adherence.<sup>43</sup> Furthermore, the approved regimen of pdAAT 60mg/kg once weekly strives for serum AAT levels above a target threshold of 11µM AAT,<sup>2,12,13,21</sup> which is approximately half of the lower limit (20µM) observed in PiMM individuals.<sup>22</sup> pdAAT 60mg/kg fails to maintain serum antigenic AAT levels of >20µM beyond the first 3–5 days of the weekly dosing period.<sup>19,21,32</sup> Although historically considered the minimal protective concentration of serum AAT,  $11\mu$ M as the target threshold of treatment has been increasingly questioned,<sup>23-28</sup> with recent evidence of greater benefit at higher levels.<sup>30-32</sup> Additionally, as a human blood product, pdAAT sourcing is vulnerable to disruptions of the human blood supply, as experienced during the COVID-19 pandemic.<sup>44</sup>

Fc fusion protein technology, which has been successfully used in therapies for other disease areas,45 enables the extended half-life observed for INBRX-101. Unlike pdAAT, INBRX-101 allows patients to sustain serum antigenic AAT levels of >20µM for 3-4 weeks between doses as opposed to the 3-5 days observed for pdAAT, 19,21,32 potentially leading to better outcomes. The longer halflife of INBRX-101 also results in less frequent dosing and thus, less patient burden, which may improve treatment compliance. Additionally, as INBRX-101 is a recombinant protein, manufacturing is scalable, resulting in a predictable and reliable supply independent of human blood donations. This could more readily allow for dose-optimization studies or evaluations in patients with more moderate AATD without the accompanying ethical concerns of diverting limited supply.<sup>2,26,27,29</sup> Furthermore, INBRX-101 is free of the inherent risk for transmission of bloodborne pathogens.

Overall, 80 and 120mg/kg INBRX-101 Q3W led to serum fAAT levels of >21 $\mu$ M for the entire dosing interval, with a safety profile similar to that of pdAAT. These levels of fAAT were achieved by administering INBRX-101 Q3W instead of weekly, as done with pdAAT.<sup>2,10,11</sup> Notably, 120mg/kg Q3W led to fAAT serum levels of >21 $\mu$ M even 4 weeks after the last dose of INBRX-101. Furthermore, INBRX-101 was shown to penetrate the lungs, which suggested activity in the target organ. PK/PD modeling using data from this trial predicts that 120mg/kg INBRX-101 dosed on a less frequent Q4W schedule would still maintain a mean steady-state trough concentration of serum fAAT at 21 $\mu$ M.

The findings of this trial support further evaluation of 120mg/kg INBRX-101 Q3W and Q4W as possible treatments for patients with AATD. A double-blind phase 2 trial assessing INBRX-101 in patients with AATD at a dose of 120mg/kg Q3W or Q4W is underway (ElevAATe, NCT05856331). This randomized trial will evaluate INBRX-101 versus pdAAT, with a primary endpoint of change in serum fAAT concentration from baseline to trough at steady state. Patients eligible for ElevAATe will have centrally confirmed serum antigenic AAT concentrations of  $<11\mu$ M at screening and genotypes associated with more severe deficiency (e.g., PiZZ and PiZ/Null). Patients with PiSZ, PiMS, or PiMZ will be excluded. In addition to a more homogenous population, ElevAATe will have a greater number of patients from whom BALF samples are collected, allowing further evaluation of the connection between INBRX-101 and functional AAT concentrations in PELF.

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**Data sharing statement:** The protocol for and the data generated and/or analyzed in the current study are not publicly available but are available upon reasonable request after the entire study is completed. Sharing of data is subject to protection of patient privacy and respect for the patient's informed consent. For more information on the process or to submit a request, contact clinicaltrials@inhibrx.com.

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#### **Declaration of Interest**

MLB reports a relationship with the University of Florida College of Medicine that includes nonfinancial support. BTK reports a relationship with Takeda that includes consulting or advisory and speaking and lecture fees; a relationship with Boehringer Ingelheim that includes consulting or advisory fees; a relationship with Grifols that includes speaking and lecture fees; a relationship with Inhibrx, Inc., that includes consulting or advisory fees and travel reimbursement; and a relationship with the California Thoracic Society that includes nonfinancial support. HWF reports financial support provided by Hannibal Clinic Operations, LLC, and a relationship with Hannibal Clinic Operations, LLC, that includes employment. RM reports a relationship with Intellia Therapeutics, Inc., that includes consulting or advisory fees. CDB is an investigator for the ElevAATe trial (NCT05856331) that is sponsored by Inhibrx, Inc. MAC reports a relationship with Inhibrx, Inc., that includes consulting or advisory fees. EB reports a relationship with Inhibrx, Inc., that includes employment and equity or stocks. SB reports a relationship with Inhibrx, Inc., that includes employment and equity or stocks and has a patent titled "Effective Dosage of Recombinant Serpin-Fc Fusion Protein for Use in a Method of Treating AAT Deficiency in a Subject" pending to Inhibrx, Inc. TB reports a relationship with Inhibrx, Inc., that includes employment and equity or stocks. CV reports a relationship with Inhibrx, Inc., that includes employment and equity or stocks. VA reports a relationship with Inhibrx, Inc., that includes employment and equity or stocks. JK reports a relationship with Inhibrx, Inc., that includes employment and equity or stocks and reports a patent titled "Effective Dosage of Recombinant Serpin-Fc Fusion Protein for Use in a Method of Treating AAT Deficiency in a Subject" pending to Inhibrx, Inc. BE reports a relationship with Inhibrx, Inc., that includes employment and equity or stocks and reports patent #10,400,029 "Serpin Fusion Polypeptides and Methods of Use Thereof" issued to Inhibrx, Inc. AC, CLC, JEL, and AGV have nothing to disclose.

## References

- Blanco I, Bueno P, Diego I, et al. Alpha-1 antitrypsin Pi\*Z gene frequency and Pi\*ZZ genotype numbers worldwide: an update. *Int J Chron Obstruct Pulmon Dis.* 2017;12:561-569. https://doi.org/10.2147/COPD.S125389
- Sandhaus RA, Turino G, Brantly ML, et al. The diagnosis and management of alpha-1 antitrypsin deficiency in the adult. *Chronic Obstr Pulm Dis.* 2016;3(3):668-682. https://doi.org/10.15326/jcopdf.3.3.2015.0182
- 3. de Serres FJ, Blanco I. Prevalence of alpha-1 antitrypsin deficiency alleles PI\*S and PI\*Z worldwide and effective screening for each of the five phenotypic classes PI\*MS, PI\*MZ, PI\*SS, PI\*SZ, and PI\*ZZ: a comprehensive review. *Ther Adv Respir Dis.* 2012;6(5):277-295. https://doi.org/10.1177/1753465812457113
- Craig TJ, Henao MP. Advances in managing COPD related to alpha-1 antitrypsin deficiency: an under-recognized genetic disorder. *Allergy*. 2018;73(11):2110-2121. https://doi.org/10.1111/all.13558
- Strnad P, McElvaney NG, Lomas DA. Alpha-1 antitrypsin deficiency. *N Engl J Med.* 2020;382(15):1443-1455. https://doi.org/10.1056/NEJMra1910234
- 6. Stoller JK, Smith P, Yang P, Spray J. Physical and social impact of alpha-1 antitrypsin deficiency: results of a survey. *Cleve Clin J Med.* 1994;61(6):461-467. https://www.ccjm.org/content/61/6/461
- Miravitlles M, Herepath M, Priyendu A, et al. Disease burden associated with alpha-1 antitrypsin deficiency: systematic and structured literature reviews. *Eur Respir Rev.* 2022;31(163):210262. https://doi.org/10.1183/16000617.0262-2021
- Beiko T, Strange C. Anxiety and depression in patients with alpha-1 antitrypsin deficiency: current insights and impact on quality of life. *Ther Clin Risk Manag.* 2019;15:959-964. https://doi.org/10.2147/TCRM.S175369
- 9. Tanash HA, Ekström M, Rönmark E, Lindberg A, Piitulainen E. Survival in individuals with severe alpha-1 antitrypsin deficiency (PiZZ) in comparison to a general population with known smoking habits. *Eur Respir J.* 2017;50(3):1700198. https://doi.org/10.1183/13993003.00198-2017
- 10. American Thoracic Society/European Respiratory Society statement: standards for the diagnosis and management of individuals with alpha-1 antitrypsin deficiency. *Am J Respir Crit Care Med.* 2003;168(7):818-900. https://doi.org/10.1164/rccm.168.7.818
- 11. Miravitlles M, Dirksen A, Ferrarotti I, et al. European Respiratory Society statement: diagnosis and treatment of pulmonary disease in alpha-1 antitrypsin deficiency. *Eur Respir J.* 2017;50(5):1700610. https://doi.org/10.1183/13993003.00610-2017
- 12. Stocks JM, Brantly ML, Wang-Smith L, et al. Pharmacokinetic comparability of Prolastin®-C to Prolastin® in alpha-1 antitrypsin deficiency: a randomized study. *BMC Clin Pharmacol.* 2010;10:13. https://doi.org/10.1186/1472-6904-10-13

- 13. Barker AF, Campos MA, Brantly ML, et al. Bioequivalence of a liquid formulation of alpha1-proteinase inhibitor compared with Prolastin®-C (lyophilized alpha1-Pi) in alpha-1 antitrypsin deficiency. *COPD*. 2017;14(6):590-596. https://doi.org/10.1080/15412555.2017.1376044
- 14. Zemaira. Prescribing information. CSL Behring LLC; 2022.
- Glassia. Prescribing information. Takeda Pharmaceuticals USA, Inc; 2023.
- 16. Aralast NP. Prescribing information. Takeda Pharmaceuticals USA, Inc; 2023.
- Colello J, Ptasinski A, Zhan X, Kaur S, Craig T. Assessment of patient perspectives and barriers to self-infusion of augmentation therapy for alpha-1 antitrypsin deficiency during the COVID-19 pandemic. *Pulm Ther.* 2022;8:95-103. https://doi.org/10.1007/s41030-022-00182-z
- Gadek JE, Klein HG, Holland PV, Crystal RG. Replacement therapy of alpha-1 antitrypsin deficiency. Reversal of protease-antiprotease imbalance within the alveolar structures of PiZ subjects. *J Clin Invest.* 1981;68(5):1158-1165. https://doi.org/10.1172/JCI110360
- Wewers MD, Casolaro MA, Sellers SE, et al. Replacement therapy for alpha-1 antitrypsin deficiency associated with emphysema. N Engl J Med. 1987;316(17):1055-1062. https://doi.org/10.1056/NEJM198704233161704
- 20. Gadek JE, Fells GA, Zimmerman RL, Rennard SI, Crystal RG. Antielastases of the human alveolar structures. Implications for the protease-antiprotease theory of emphysema. *J Clin Invest.* 1981;68(4):889-898. https://doi.org/10.1172/JCI110344
- 21. Stocks JM, Brantly M, Pollock D, et al. Multi-center study: the biochemical efficacy, safety and tolerability of a new alpha-1 proteinase inhibitor, Zemaira. *COPD*. 2006;3(1):17-23. https://doi.org/10.1080/15412550500493220
- 22. Brantly ML, Wittes JT, Vogelmeier CF, Hubbard RC, Fells GA, Crystal RG. Use of a highly purified alpha-1 antitrypsin standard to establish ranges for the common normal and deficient alpha-1 antitrypsin phenotypes. *Chest.* 1991;100(3):703-708. https://doi.org/10.1378/chest.100.3.703
- Ferrarotti I, Thun GA, Zorzetto M, et al. Serum levels and genotype distribution of alpha-1 antitrypsin in the general population. *Thorax.* 2012;67(8):669-674. https://doi.org/10.1136/thoraxjnl-2011-201321
- 24. Franciosi AN, Fraughen D, Carroll TP, McElvaney NG. Alpha-1 antitrypsin deficiency: clarifying the role of the putative protective threshold. *Eur Respir J.* 2022;59(2):210410. https://doi.org/10.1183/13993003.01410-2021
- 25. Bornhorst JA, Greene DN, Ashwood ER, Grenache DG. Alpha-1 antitrypsin phenotypes and associated serum protein concentrations in a large clinical population. *Chest.* 2013;143(4):1000-1008. https://doi.org/10.1378/chest.12-0564

- 26. Brantly ML, Lascano JE, Shahmohammadi A. Intravenous alpha-1 antitrypsin therapy for alpha-1 antitrypsin deficiency: the current state of the evidence. *Chronic Obstr Pulm Dis.* 2019;6(1):100-114. https://doi.org/10.15326/jcopdf.6.1.2017.0185
- 27. Barjaktarevic I, Miravitlles M. Alpha-1 antitrypsin (AAT) augmentation therapy in individuals with the PI\*MZ genotype: a pro/con debate on a working hypothesis. *BMC Pulm Med.* 2021;21:99. https://doi.org/10.1186/s12890-021-01466-x
- 28. McElvaney GN, Sandhaus RA, Miravitlles M, et al. Clinical considerations in individuals with alpha-1 antitrypsin PI\*SZ genotype. *Eur Respir J.* 2020;55(6):1902410. https://doi.org/10.1183/13993003.02410-2019
- 29. Sandhaus RA, Turino G, Stocks J, et al. Alpha-1 antitrypsin augmentation therapy for PI\*MZ heterozygotes: a cautionary note. *Chest.* 2008;134(4):831-834. https://doi.org/10.1378/chest.08-0868
- 30. Chapman KR, Burdon JG, Piitulainen E, et al. Intravenous augmentation treatment and lung density in severe alpha-1 antitrypsin deficiency (RAPID): a randomised, double-blind, placebo-controlled trial. *Lancet.* 2015;386(9991):360-368. https://doi.org/10.1016/S0140-6736(15)60860-1
- Campos MA, Geraghty P, Holt G, et al. The biological effects of double-dose alpha-1 antitrypsin augmentation therapy. A pilot clinical trial. *Am J Respir Crit Care Med.* 2019;200(3):318-326. https://doi.org/10.1164/rccm.201901-00100C
- 32. Campos MA, Kueppers F, Stocks JM, et al. Safety and pharmacokinetics of 120mg/kg versus 60mg/kg weekly intravenous infusions of alpha-1 proteinase inhibitor in alpha-1 antitrypsin deficiency: a multicenter, randomized, double-blind, crossover study (SPARK). *COPD*. 2013;10(6):687-695. https://doi.org/10.3109/15412555.2013.800852
- 33. Veale A, Kalabus J, Farah H, et al. Evaluation of safety and pharmacokinetics of the recombinant human AAT-Fc fusion protein INBRX-101 in patients with alpha-1 antitrypsin deficiency. *Am J Respir Crit Care Med.* 2022;205:A4114. https://doi.org/10.1164/ ajrccm-conference.2022.205.1\_MeetingAbstracts.A4114
- 34. Veale A, Farah H, Mahadeva R, et al. Recombinant AAT-Fc fusion protein INBRX-101 achieves normal serum AAT levels in patients with alpha-1 antitrypsin deficiency (AATD). *Eur Respir J.* 2022;60(suppl 66):3599. https://doi.org/10.1183/13993003.congress-2022.3599
- 35. Pierce RL. Summary basis for regulatory action. Prolastin-C. US Food and Drug Administration website. Published September 2017. Accessed May 2023. https://www.fda.gov/media/107485/download.
- 36. Dirksen A, Dijkman JH, Madsen F, et al. A randomized clinical trial of alpha-1 antitrypsin augmentation therapy. *Am J Respir Crit Care Med.* 1999;160(5):1468-1472. https://doi.org/10.1164/ajrccm.160.5.9901055
- Hubbard RC, Crystal RG. Alpha-1 antitrypsin augmentation therapy for alpha-1 antitrypsin deficiency. *Am J Med.* 1988;84(6, suppl 1):52-62. https://doi.org/10.1016/0002-9343(88)90159-3

- 38. Barker AF, Iwata-Morgan I, Oveson L, Roussel R. Pharmacokinetic study of alpha1-antitrypsin infusion in alpha-1 antitrypsin deficiency. *Chest.* 1997;112(3):607-613. https://doi.org/10.1378/chest.112.3.607
- 39. Miravitlles M, Vidal R, Torrella M, Bofill JM, de Gracia J, Cotrina M. Evaluation of replacement therapy in emphysema caused by alpha-1 antitrypsin deficiency [in Spanish]. Arch Bronconeumol. 1994;30(10):479-484. https://doi.org/10.1016/S0300-2896(15)30997-2
- 40. Vidal Pla R, Padullés Zamora N, Sala Piñol F, Jardí Margaleff R, Rodríguez Frías F, Montoro Ronsano JB. Pharmacokinetics of alpha-1 antitrypsin replacement therapy in severe congenital emphysema [in Spanish]. *Arch Bronconeumol.* 2006;42(10):553-556. https://doi.org/10.1157/13093399
- 41. Sanders CL, Ponte A, Kueppers F. The effects of inflammation on alpha-1 antitrypsin levels in a national screening cohort. *COPD*. 2018;15(1):10-16. https://doi.org/10.1080/15412555.2017.1401600
- 42. Kueppers F. Genetically determined differences in the response of alpha-1 antitrypsin levels in human serum to typhoid vaccine. *Humangenetik.* 1968;6:207-214. https://doi.org/10.1007/BF00291864
- 43. Strange C, Stoller JK, Sandhaus RA, Dickson R, Turino G. Results of a survey of patients with alpha-1 antitrypsin deficiency. *Respiration*. 2006;73(2):185-190. https://doi.org/10.1159/000088061
- 44. Hartmann J, Klein HG. Supply and demand for plasma-derived medicinal products a critical reassessment amid the COVID-19 pandemic. *Transfusion*. 2020;60(11):2748-2752. https://doi.org/10.1111/trf.16078
- 45. Strohl WR. Fusion proteins for half-life extension of biologics as a strategy to make biobetters. *BioDrugs*. 2015;29:215-239. https://doi.org/10.1007/s40259-015-0133-6