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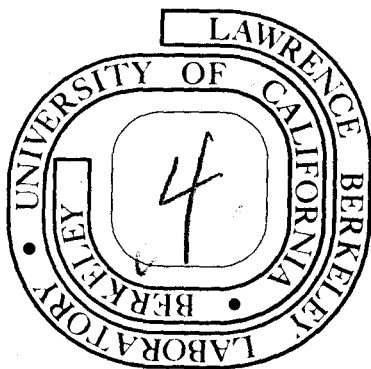
Thomas Vincent Rowland
(Ph. D. thesis)

October 1975

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OXYGEN-17 NMR STUDIES OF THE RATE OF WATER EXCHANGE
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OXYGEN-17 NMR STUDIES OF THE RATE OF WATER EXCHANGE
FROM PARTIALLY COMPLEXED NICKEL ION

Thomas Vincent Rowland

Inorganic Materials Research Division, Lawrence Berkeley Laboratory
and Department of Chemistry; University of California,
Berkeley, California

ABSTRACT

The oxygen-17 bulk water NMR linewidth and chemical shifts for an aqueous solution of the nickel iminodiacetate complex, NiIDA, have been measured from 0°C to 140°C at 8.134 MHz in order to determine both the rate of chemical exchange between the two chemically non-equivalent first coordination sphere sites and the bulk solvent and also the scalar coupling constants, A/h. Linewidth measurements from 0°C to 100°C for a Ni(IDA)₂²⁻ sample yielded estimates of the per cent time of dissociation for the carboxylate arms and the resulting rate of water exchange.

Two distinct rates of water exchange were measured for the three waters of the NiIDA complex: one slow exchanging site with $k_{298^\circ} = 5.3 \times 10^4 \text{ sec}^{-1}$, $\Delta H^\ddagger = 11.8 \text{ kcal.}$, $\Delta S^\ddagger = 2.7 \text{ eu}$, and $A/h = 1.8 \times 10^7 \text{ Hz}$ and two equivalent fast exchanging sites with $k_{298^\circ} = 2.4 \times 10^5 \text{ sec}^{-1}$, $\Delta H^\ddagger = 11.4 \text{ kcal.}$, $\Delta S^\ddagger = 4.6 \text{ eu}$, and $A/h = 1.9 \times 10^7 \text{ Hz}$. The difference in exchange rates is thought to be due to the bond strain about the coordinated nitrogen group which is partially relieved by a tendency to eliminate one of the two water positions cis to the nitrogen group, while the third water position remains unaffected by the strain.

For the $\text{Ni}(\text{IDA})_2^{2-}$ complex, the per cent time of dissociation of some one of the four carboxylate arms was measured to be ca. 1.5%, while the rate of water exchange was estimated to be $4 \times 10^5 \text{ sec}^{-1}$, assuming that the scalar coupling constants were similar to those measured for NiIDA.

Ligand labilization trends are discussed relative to these and other complexes containing coordinated nitrogen and carboxylate groups for which water exchange rates appear in the literature. Electron donor effects, configurational strain and conformational effects are considered in this treatment, and it is concluded that the effect of bond strain is important for the IDA complex in the labilization of the remaining water molecules.

I. INTRODUCTION

Nuclear magnetic resonance spectroscopy has become a widely applied technique used in measuring the rate of water exchange between the first hydration sphere of metal ions and the bulk solvent in cases where rapid exchange occurs. Temperature studies of the broadening of the bulk water oxygen-17 NMR signal by paramagnetic transition metal ions have in the past provided much information about water exchange rates for these ions.^{1a,b,c} More recently similar temperature studies on partially substituted metal ion complexes have been reported.² Much of the work has been done on systems utilizing ligands coordinated at nitrogen and/or oxygen positions such as NH_3 , en, dien, EDTA, etc, and paramagnetic transition metal ions such as cobalt and nickel. It has been generally shown that the rate of replacement of water in the first sphere of hydration is definitely affected by the presence of the other coordinated ligands bound to the metal ion. In general, coordinated nitrogens appear to increase the lability of the remaining water molecules while for coordinated oxygens the case is not as clear.^{2,3a,b}

The purpose of this thesis is to measure the rates of water exchange from the partially substituted nickel iminodiacetate complex, $\text{NiNH}(\text{CH}_2\text{COO})_2$ or NiIDA, and to attempt to determine the reasons for the labilization of the remaining waters. The NiIDA complex has three remaining water molecules in the first coordination sphere, two of which are equivalent. This ligand is believed to be complexed with nickel in a cis (facial) manner, that is with the carboxylate groups on axes 90° from each other rather than 180° .^{4a,b} Nickel has been chosen as

the paramagnetic metal ion in this experiment since the oxygen-17 nuclear magnetic relaxation has been well characterized for the hexaquo ion, with both the chemical exchange control and relaxation control regions reached over the easily attainable temperature range of 0°C to 140°C. At lower temperatures (below about 60°C) relaxation is controlled by the rate of chemical exchange of the water molecules, while in the mid temperature region relaxation is controlled by the $\Delta\omega$ mechanism. At higher temperatures scalar coupling relaxation also contributes appreciably.^{1c} These same relaxation regions are expected to be found over this temperature range for the partially complexed NiIDA.

Previous work^{5a,b} on the nickel nitrilotriacetate complex, $\text{NiN}(\text{CH}_2\text{COO})_3^-$ or NiNTA, which has two nonequivalent waters remaining in the first coordination sphere, one trans and the other cis to the coordinated nitrogen, has indicated that these two water molecules exchange at considerably different rates ($k_1 = \sim 2 \times 10^4 \text{ sec}^{-1}$, $k_2 = \sim 1 \times 10^6 \text{ sec}^{-1}$). From the results of the oxygen-17 line broadening temperature study it is not possible to determine which site is the faster exchanging site. However, the case for the IDA complex, similar to the NTA complex but with one less coordinated carboxylate arm, is theoretically different. Since the broadening of the NMR signal is a function of the concentration of both the paramagnetic ion and the number of identical water coordination positions, and because for the NiIDA complex there are two water molecules of one type (cis to nitrogen)

and one of another type (trans to nitrogen), the fast exchanging position (s) may be distinguished from the slow position(s) as long as the difference in exchange rates is large enough to be detectable.

In order to measure accurately the paramagnetic line broadening of the oxygen-17 signal, three corrections must be known. First, the natural oxygen-17 pure water linewidth must be subtracted from the experimentally observed bulk water linewidth. Although normally a trivial process, in this case where for stoichiometric reasons the NiIDA sample solution was held at a pH of 8.5, this correction was more difficult to make due to the dependency of the oxygen-17 linewidth on pH in the pH range of about 4.0 to 9.0 at room temperature and lower. In the neutral region where the proton exchange rate from a water molecule is considerably slower than in an acidic or basic solution, the oxygen-17 linewidth becomes broadened due to the spin-spin splitting of the signal by the protons.⁶ A study was undertaken to determine the pH of the sample solution as a function of temperature and NMR data were collected so as to know the proper linewidth blank correction both as a function of pH and temperature.

A second correction which is of interest in itself was determined by measuring the paramagnetic broadening which occurs in a fully complexed $\text{Ni}(\text{IDA})_2^{2-}$ sample. Theoretically if the IDA were bound strongly enough to the nickel ion, there would be no paramagnetic broadening since the totally complexed nickel would not interact with the bulk water in a manner which would cause nuclear magnetic relaxation,

unless of course second sphere interactions were strong enough to contribute to the bulk water oxygen-17 relaxation. Although it was determined that the correction required due to the presence of $\text{Ni}(\text{IDA})_2^{2-}$ in the NiIDA sample was negligible, it still provides a direct measurement of the complexing strength of the carboxylate groups in the IDA molecule. Whenever a carboxylate arm comes free from its coordinated position and allows a water molecule to exchange in and out of this position, paramagnetic broadening will occur. This effect has been measured and will be discussed. Additionally, the correction for broadening due to free nickel ion in the NiIDA sample solution was easily made using Neely's data for the nickel hexaquo ion.^{1c}

Measurement of the chemical shift of the bulk water resonance position of the NiIDA sample solution relative to the resonance position of a pure H_2O^{17} sample has also been undertaken. This experiment also provides information on the water exchange rates of the NiIDA complex as well as assisting in the determination of the relevant scalar coupling constants.

II. THEORY

In general, the theory involved in interpreting the experimental results obtained consists of arriving at equations to handle both the NMR linewidth and chemical shift data. The well known Bloch equations⁷ provide a phenomenological description of the macroscopic properties of nuclei in an external magnetic field H_0 in terms of a longitudinal relaxation time T_1 and a transverse relaxation time T_2 . When solved for the steady state, slow passage condition with no RF saturation, using audio frequency modulation side band detection methods,⁸ the Bloch equations yield for the absorption mode first side band signal strength

$$S = (2/\beta) \frac{[J_1(\beta)]^2 \gamma H_1 T_2 M_0}{1 + T_2^2 (\Delta\omega_0 \pm \omega_m)^2} \quad (1)$$

where $\beta = \gamma H_m / \omega_m$, γ is the magnetogyric ratio of the observed nucleus, H_m is the amplitude of the audio modulation in gauss, J_1 is the first Bessel function of the first kind, H_1 is one half the amplitude of the RF field in gauss, M_0 is the equilibrium magnetization, $\Delta\omega_0 = \omega_0 - \omega$, ω_0 is the Larmor precessional frequency of the observed nucleus in the field H_0 , and ω_m is the frequency of the audio modulation. The second term in the denominator of this equation may be more meaningful if rearranged and written in terms of the first side band resonance frequency, $\omega_0 \pm \omega_m$, giving $T_2^2 [(\omega_0 \pm \omega_m) - \omega]^2$. The signal will have a maximum height when $(\omega_0 \pm \omega_m) - \omega = 0$ or $\omega_0 = \gamma H_0 = \pm \omega_m + \omega$, and S will be at half height when $\{T_2 [(\omega_0 \pm \omega_m) - \omega]\}^2 = 1$ or

$\frac{1}{T_2} = (\omega_0 \pm \omega_m) - \omega_{1/2} = \Delta\omega_{1/2}$, where $\Delta\omega_{1/2}$ is seen to be the half width of the signal at half height.

In general for dilute aqueous solutions of paramagnetic ions, the oxygen-17 transverse relaxation rate can be divided into two contributions,

$$\frac{1}{T_2} = \frac{1}{T_{2H_2O}} + \frac{1}{T_{2p}} \quad (2)$$

where $\frac{1}{T_{2H_2O}}$ is the natural oxygen-17 linewidth of pure water and $\frac{1}{T_{2p}}$ represents the line broadening due to the presence of paramagnetic ions. For dilute solutions, where exchange is occurring between bulk and bound waters, $\frac{1}{T_{2p}}$ can be described by the following equation, ^{1a}

$$\frac{1}{T_{2p}} = \frac{P_m}{\tau_m} \frac{\frac{1}{T_{2m}^2} + \frac{1}{T_{2m}\tau_m} + \Delta\omega_m^2}{\left[\left(\frac{1}{T_{2m}} + \frac{1}{\tau_m} \right)^2 + \Delta\omega_m^2 \right]} \quad (3)$$

where τ_m is the lifetime of a water molecule in the first coordination sphere of the metal ion, $\frac{1}{T_{2m}}$ is the transverse relaxation rate of the coordinated or bound water molecules, $\Delta\omega_m$ is the frequency difference between the bound water oxygen-17 and the bulk water oxygen-17 signals in the paramagnetic solution, and P_m is the mole ratio of the bound waters to the bulk waters, defined by

$$P_m = \frac{x_m}{x_{H_2O}} = \frac{n[Ni^{2+}]}{55.5 - n[Ni^{2+}]} \quad (4)$$

where n is the number of water coordination sites on the paramagnetic ion and $[\text{Ni}^{2+}]$ is the molality of the paramagnetic ion.

To handle the NiIDA problem, where there are two chemically nonequivalent bound water environments, the above equation must be written as derived by Swift and Connick^{1a} for three sites, where exchange is allowed from bound positions to bulk positions only and not from one bound position to another bound position. The equation then becomes

$$\frac{1}{T_{2p}} = \frac{P_{m_1}}{\tau_{m_1}} \left[\frac{\frac{1}{T_{2m_1}^2} + \frac{1}{T_{2m_1} \tau_{m_1}} + \Delta\omega_{m_1}^2}{\left(\frac{1}{T_{2m_1}} + \frac{1}{\tau_{m_1}}\right)^2 + \Delta\omega_{m_1}^2} \right] + \frac{P_{m_2}}{\tau_{m_2}} \left[\frac{\frac{1}{T_{2m_2}^2} + \frac{1}{T_{2m_2} \tau_{m_2}} + \Delta\omega_{m_2}^2}{\left(\frac{1}{T_{2m_2}} + \frac{1}{\tau_{m_2}}\right)^2 + \Delta\omega_{m_2}^2} \right] \quad (5)$$

where the subscript "1" signifies the single water coordination position and the subscript "2" signifies the remaining two equivalent water positions on the NiIDA complex, and thus $P_{m_1} = [\text{NiIDA}]/(55.5 - 3[\text{NiIDA}])$ and $P_{m_2} = 2[\text{NiIDA}]/(55.5 - 3[\text{NiIDA}])$. This equation cannot be simplified and still describe the full temperature range over which data were collected, since a T_{2m} relaxation region, a $\Delta\omega_m$ relaxation region, and an exchange controlled region are all present.

The chemical shift measurements made on the NiIDA complex system may be handled in the following manner. The bulk water chemical shift, $\Delta\omega_{H_2O}$, which is the difference between the resonance frequency of pure water and that of the bulk water in the paramagnetic solution, for dilute solutions obeys the equation^{1a}

$$\Delta\omega_{H_2O} = \frac{-P_m \Delta\omega_m}{\tau_m^2 \left[\left(\frac{1}{T_{2m}} + \frac{1}{\tau_m} \right)^2 + \Delta\omega_m^2 \right]} \quad (6)$$

or for the NiIDA case with two chemically different bound water sites

$$\Delta\omega_{H_2O} = \frac{-P_{m_1} \Delta\omega_{m_1}}{\tau_{m_1}^2 \left[\left(\frac{1}{T_{2m_1}} + \frac{1}{\tau_{m_1}} \right)^2 + \Delta\omega_{m_1}^2 \right]} + \frac{-P_{m_2} \Delta\omega_{m_2}}{\tau_{m_2}^2 \left[\left(\frac{1}{T_{2m_2}} + \frac{1}{\tau_{m_2}} \right)^2 + \Delta\omega_{m_2}^2 \right]} \quad (7)$$

where all terms have been previously defined. At high temperature, or more specifically when $\frac{1}{\tau_m} \gg \frac{1}{T_{2m}}$, $\Delta\omega_m^2 \tau_m$, then

$$\Delta\omega_{H_2O} = -P_m \Delta\omega_m = -P_{m_1} \Delta\omega_{m_1} - P_{m_2} \Delta\omega_{m_2} \quad (8)$$

and the bulk water chemical shift experiment gives a direct measurement of the value of the relaxation parameter $\Delta\omega_m$. Bloembergen's equation⁹ gives the chemical shift of the bound waters relative to pure water, $\Delta\omega_{H_2O-m}$, and can be set approximately equal to $\Delta\omega_m$ for the dilute solution case where the bound water shift will be considerably greater than the bulk water shift,

$$\Delta\omega_{\text{H}_2\text{O}-m} = \Delta\omega_m - \Delta\omega_{\text{H}_2\text{O}} = \omega S(S+1) \frac{\gamma_S}{\gamma_I} \frac{A}{3kT} \cong \Delta\omega_m \quad (9)$$

where ω is the precessional frequency of the observed nuclei, S is the electron spin quantum number, γ_S and γ_I are the gyromagnetic ratios of the unpaired electron and the observed nucleus, A is the scalar coupling constant in ergs, which will be discussed later, k is Boltzmann's constant, and T is the temperature. Again, for the NiIDA case there will be two different scalar coupling constants, A_1 and A_2 , representing the two different water sites. The value of γ_S was calculated from the relationship

$$\gamma_S = g_{\text{eff}} \beta = \frac{\mu_{\text{eff}}}{\sqrt{S(S+1)}} \quad (10)$$

where g_{eff} is the effective spectroscopic splitting factor or g-factor, β is the Bohr magneton, and μ_{eff} is the effective magnetic moment for nickel ion in aqueous solution.¹⁰ Using μ_{eff} equal to 3.22 as the observed value reported in Tables of Constants and Numerical Data (U.I.C.P.A.), No. 7, Constantes Selectionnees Diamagnetisme et Paramagnetisme by G. Foex, Masson, and Cie, Paris (1957), a value of 2.28 was obtained for g_{eff} . This is in good agreement with the value of 2.25 obtained from ESR work on hydrated nickel salts.¹¹

Both the equation for the paramagnetic line broadening, $\frac{1}{T_{2p}}$, and the equation for the bulk water chemical shift, $\Delta\omega_{\text{H}_2\text{O}}$, contain the parameters τ_m and T_{2m} in addition to the above defined $\Delta\omega_m$. The exchange lifetime, τ_m , should have the normal temperature dependence expressed by the Eyring equation,

$$\tau_m = \frac{h}{kT} \exp \left[\frac{\Delta H^\ddagger}{RT} - \frac{\Delta S^\ddagger}{R} \right] \quad (11)$$

where ΔH^\ddagger and ΔS^\ddagger are the enthalpy and entropy of activation for water exchange.

It has been shown that the oxygen-17 relaxation caused by the hydrated paramagnetic nickel ion, $\frac{1}{T_{2p}}$, is due to the scalar coupling mechanism,^{1c} and the same is expected for the partially complexed nickel case. The scalar coupling interaction has the form $A\vec{I} \cdot \vec{S}$ where A is called the scalar coupling constant and is in effect a measure of the amount of unpaired electron overlap at the position of the nucleus I. The relaxation rate due to this scalar interaction is described by the equation¹²

$$\frac{1}{T_{2m}} = \frac{S(S+1)}{3} \left(\frac{A}{h}\right)^2 \left[\tau_{c1} + \frac{\tau_{c2}}{1 + \omega_S^2 \tau_{c2}^2} \right] \quad (12)$$

where it has been assumed that $\omega_I \ll \omega_S$ and where

$$\frac{1}{\tau_{c1}} = \frac{1}{\tau_m} + \frac{1}{T_{1e}}, \quad \frac{1}{\tau_{c2}} = \frac{1}{\tau_m} + \frac{1}{T_{2e}} \quad (13)$$

where T_{1e} and T_{2e} are the longitudinal and transverse electronic relaxation times. In the case of nickel, since $\tau_m \gg T_{1e}, T_{2e}$, the scalar coupling correlation time which will be designated τ_e may be written

$$\tau_e = T_{1e} + \frac{T_{2e}}{1 + \omega_S^2 T_{2e}^2} \quad (14)$$

Again, similar equations for $\frac{1}{T_{2m_1}}$ and $\frac{1}{T_{2m_2}}$ must be written utilizing the scalar coupling constants A_1 and A_2 respectively for the NiIDA case. In general, since the T_{1e} and T_{2e} contributions to the correlation time cannot be distinguished in the following experiments, for computer fitting purposes the equation for the correlation time can be simplified to the form

$$\tau_e = \tau_e^o \exp \frac{V}{RT} \quad (15)$$

where V represents the activation energy for the electronic relaxation.

III. EXPERIMENTAL

A. Sample Preparation

The pH vs. temperature line broadening study of pure water to determine blank corrections for the NiIDA samples was conducted using 10% H_2O^{17} buffered with 0.1 M NH_4ClO_4 and adjusted to the desired pH with NaOH. Multiple samples were run with pH's varying from 8.00 to 9.00 over the temperature range of 0°C to 45°C. Above 45°C at pH 8.2 or higher no additional correction above the normal (acidic or basic) H_2O^{17} linewidth is required.

NiIDA sample solutions were prepared from recrystallized $\text{Ni}(\text{ClO}_4)_2 \cdot 6\text{H}_2\text{O}$ (GFS) and iminodiacetic acid (Eastman Kodak) recrystallized successively from acidic water and ethanol. The recrystallized $\text{Ni}(\text{ClO}_4)_2$ was analyzed spectroscopically to be free of other paramagnetic impurities. Solutions of about 2 ml. volume were prepared by weight to be approximately 0.025 M in both the mono and bis complexes of nickel IDA using ~10% enriched H_2O^{17} . The pH of the sample solutions was adjusted to 8.5 using high purity NaOH, and all samples contained $\sim 5 \times 10^{-4}$ M NH_4ClO_4 as a buffer. Concentration calculations were made using the complexing constants shown in Table I taken from Sillen and Martell's compilation, Stability Constants of Metal-Ion Complexes,¹³ and sample conditions (pH, concentration of constituents) were such as to maximize NiIDA concentration while minimizing dependency on the nickel iminodiacetate complexing constants without suffering nickel hydroxide solubility problems.

Table I. Equilibrium Constants for the NiIDA and Ni(IDA)₂²⁻ Complexes¹³

| Reaction | log K _{30°K} (mole ⁻¹ litre) | ΔH ^o (kcal) | ΔS ^o (eu) |
|--|---|---------------------------|-------------------------|
| Acid Dissociation Constants: | | | |
| H ⁺ + IDA ²⁻ = H(IDA) ⁻ | 9.12 | -8.15 | 15.4 |
| H ⁺ + H(IDA) ⁻ = H ₂ (IDA) | 2.98 | | |
| Nickel Complexing Constants: | | | |
| Ni ²⁺ + IDA ²⁻ = Ni(IDA) | 8.26 | -5.05 | 20.0 |
| Ni(IDA) + IDA ²⁻ = Ni(IDA) ₂ ²⁻ | 6.35 | -4.5 | 13.1 |
| Nickel Hydroxide Solubility Product: | | | |
| Ni ²⁺ + 2(OH ⁻) = Ni(OH) ₂ ↓ | -15.0 | -6.7 | |

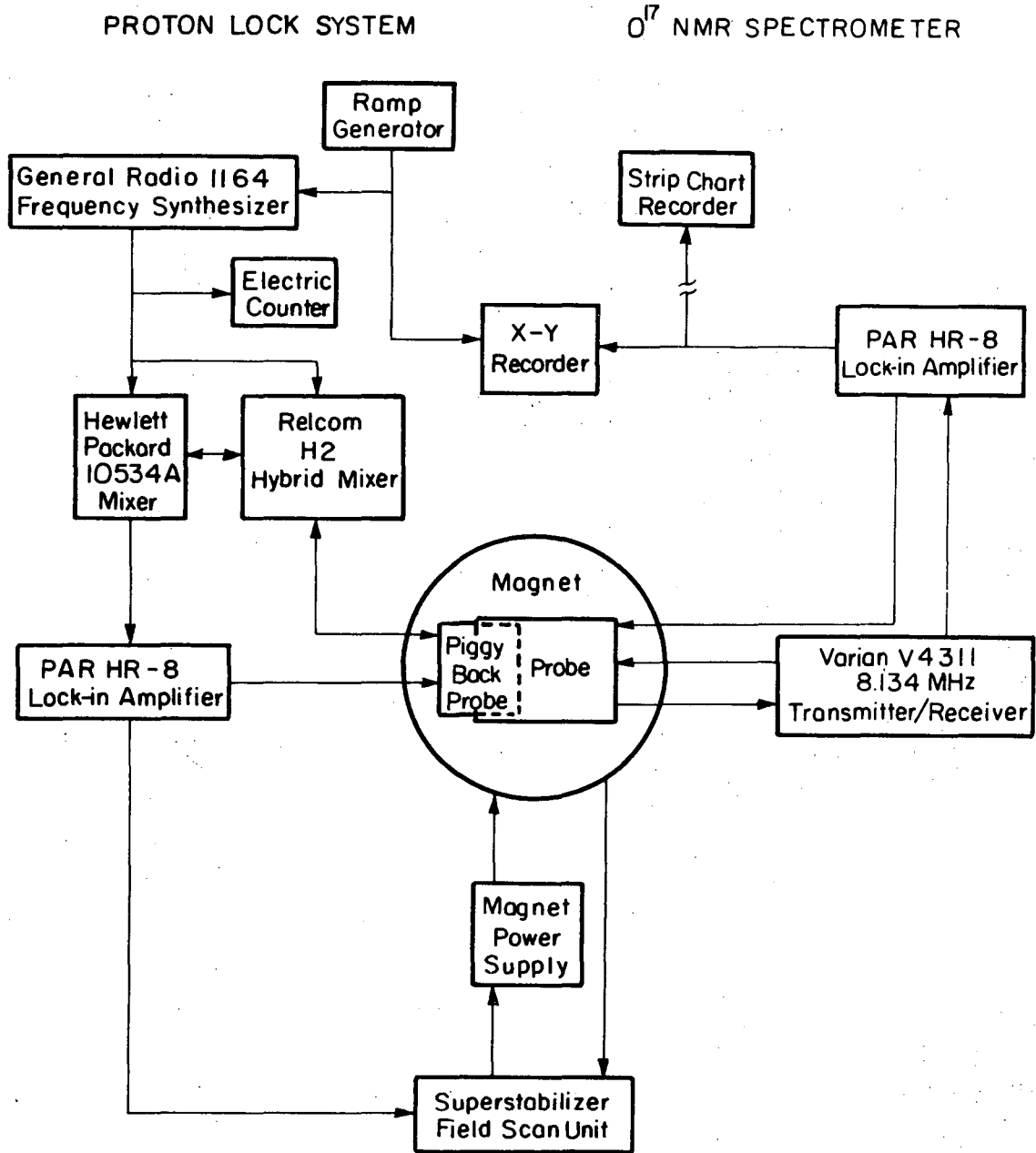
The bis nickel IDA complex solutions were prepared in a similar manner, but with a $\text{Ni}(\text{IDA})_2^{2-}$ concentration of approximately 0.50 M and negligible NiIDA concentration, and with pH adjusted to about 9.2. A similar $\text{Zn}(\text{IDA})_2^{2-}$ sample was prepared as a diamagnetic water blank having similar properties.

Between the temperatures of 0°C and 60°C all samples were run in 11 mm. O.D. Pyrex tubes (~2ml. volume) constructed with thermocouple wells immersed in the solution, and were sealed under vacuum. Above 60°C, machined teflon sample tubes of similar dimensions were used for reasons explained below. These tubes, which utilized threaded pressure caps, were tested to 140°C (3.5 atm.) for leaks and were sealed at atmospheric pressure. Difficulty was experienced in holding the pH of the 0.025 NiIDA samples constant for the duration of an NMR temperature study. At high temperatures a reaction occurred within the sample cell manifesting itself by the appearance of a small amount of white precipitate and a decline in pH. Depending on the temperature and length of heating, the initial pH of 8.4 declined as much as one pH unit, causing a considerable amount of neutral region linebroadening. The precipitate was analyzed to be high in both boron and silicon while all sample constituents were analyzed free of these contaminants, leading one to suspect the glass. Using a teflon sample cell and 0.025 M NiIDA at a pH of 8.44 resulted in considerably less pH change (-0.24 pH units) and the appearance of no ppt. after heating for six hours at 105°C, while the same treatment resulted in a much larger change (-0.84 pH units) for an identical sample in a pyrex tube.

Care was taken in sample preparation to use no soft glassware and all glass sample cells were sealed in such a manner as to preclude direct heating of the sample solution. The pH of the sample was checked upon transfer from pyrex to teflon tube for the high temperature work and after each run above 100°C to ensure that a negligible pH change had taken place.

B. NMR Linewidth Measurements

For all linewidth measurements a Varian Associates model V4200 wide line spectrometer was used in conjunction with a model V4311 fixed frequency transmitter/receiver operating at 8.134 MHz. The diode or leakage detection mode was employed and care was taken to avoid RF power saturation, with power settings between 0.04 and 0.23 volts rms depending on the linewidth. A Princeton Applied Research model HR-8 lock-in amplifier was used to provide audio modulation and phase sensitive detection. The modulation amplitude setting was determined to maximize the peak height of the first sidebands and the audio frequency was set such that the half width of the resonance signal was always at least an order of magnitude smaller than the modulation frequency. A diagram of the spectrometer as used for the linewidth measurements is shown in the right half of Fig. 1. Data were collected by scanning continuously through the upfield and downfield first sidebands four times, allowing eight linewidth measurements. Since the separation of the first sidebands is exactly twice the modulation frequency, the linewidths may be easily calculated.



XBL 759-7194

Fig. 1. Schematic of oxygen-17 NMR spectrometer and proton lock system as used in linewidth and chemical shift measurements.

C. Chemical Shift Measurements

Chemical shift measurements of the bulk water resonance relative to the pure H_2O^{17} resonance were made using the same spectrometer, but additionally utilizing an external NMR proton lock system which was used for both locking and scanning purposes. This system consisted of a scannable 60 MHz radio frequency source, a Varian piggy back probe containing a doped water sample which was fitted inside the main probe body, a passive RF detection system, and an additional phase sensitive lock-in amplifier for audio modulation and detection in the lock system as shown in the left half of Fig. 1. During the chemical shift experiment the magnetic field was scanned by driving the frequency synthesizer with a ramp generator, causing the proton RF to change which in turn caused the superstabilizer/scan unit to scan the magnetic field. This system allowed accurate calibration of the magnetic field strength by counting the proton lock frequency at the beginning and end of each scan, and thus the absolute position of the resonance lines could be measured accurately to determine the chemical shifts. The results of four scans on both the water blank and the NiIDA samples were averaged to give a single data point.

The temperature of the sample in the probe was controlled by flowing heated or cooled dry nitrogen gas past the sample tube which was surrounded by a Dewared probe insert. The temperature for each run was taken as the average reading of two copper-constantan thermocouples read to 0.1°C , one placed in the nitrogen stream directly below the sample tube, the other in the thermocouple well in the top portion of

the sample tube. Both thermocouples were read immediately before and after each run, with temperature differences between top and bottom ranging from zero at room temperature to 5°C at 140°C. In all runs utilizing teflon sample tubes, in order to facilitate rapid temperature equilibration within the probe, the sample tubes were preheated in an external oil heat bath before being transferred to the probe. This reduced the total time the sample remained at elevated temperatures in order to minimize the possibility of decomposition due to heat.

D. Computer Fitting Programs

The linewidth data were fitted to Eq. (5) using the Oak Ridge General Least Squares (ORGLS) computer program.¹⁴ The standard deviation (68% confidence interval) in the eight linewidths measured for each data point was used as the weighting factor. A correlation matrix consisting of an array of correlation coefficients which describe the linear dependence of each parameter on each other parameter (where one represents complete correlation)¹⁵ was printed at the end of the output section.

The results of the linewidth and chemical shift measurements were simultaneously fitted to Eq. (5) and Eq. (7) using the Dye and Nicely General Purpose Curvefitting Program.¹⁶ Again standard deviations were used as weighting factors for both sets of data and a list of multiple correlation coefficients which describe the total linear dependence of each parameter on all other parameters¹⁵ was printed in the output of the program. The curvefitting subroutine contained IF statements directing the program to fit the proper data set to the proper equation,

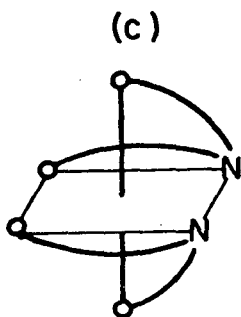
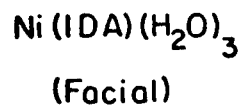
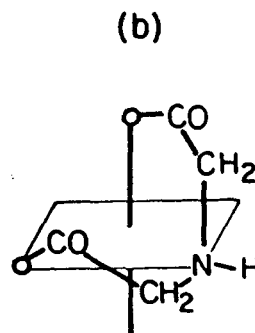
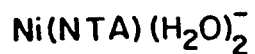
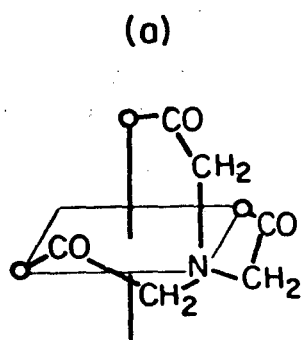
as shown in Appendix II. Fortuitously, the numerical values of the linewidths and chemical shifts were of the same order of magnitude in the NiIDA experiment as were the weighting factors for both experimental data sets. Were this not true, one data set would have to be normalized to the other to provide a reasonably weighted fit.

IV. RESULTS AND DISCUSSION

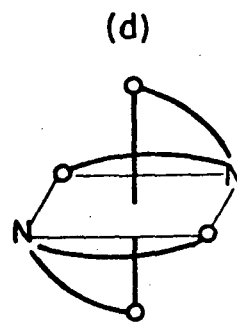
A. Introduction: NiNTA Complex

In 1968 Kustin and Vriesenga, in work done in this laboratory,⁵ observed two distinct water exchange rates from the first coordination sphere of the nickel nitrilotriacetate complex, $\text{NiNTA}(\text{H}_2\text{O})_2^-$, which is shown in Fig. 2(a). It is evident from the figure that the two remaining water molecules are not structurally equivalent, one being trans to a tertiary nitrogen, the other being cis to the nitrogen and in the plane of coordination of the carboxylate groups. Preliminary results from this experiment, to be published later, have shown that the two exchange rates differ by about a factor of seventy at room temperature. The linebroadening measurements, as a function of temperature, produce two easily distinguishable minima when plotted as T_{2p} vs. $10^3/T$, showing two exchange controlled and two $\Delta\omega_m$ relaxation regions. The chemical shift measurements also show two flat "rapid exchange" areas as expected when plotted as $\Delta\omega_{\text{H}_2\text{O}}$ vs. $10^3/T$. Unfortunately, while the large difference in rates for the two exchange positions appears to confirm the structural non-equivalence of these sites, the oxygen-17 NMR experiment does not identify the fast or slow site.

Vriesenga has treated the linewidth and chemical shift data in two manners.^{5b} The first method was to determine the scalar coupling constants, A_1 and A_2 , from the best visual fit of the chemical shift plot, then choosing the activation parameters ΔH_1^\ddagger , ΔS_1^\ddagger , ΔH_2^\ddagger and ΔS_2^\ddagger



Cis-Facial



Trans - Facial

XBL 759-7195

Fig. 2. (a) Configuration of the nickel nitrilotriacetate (NiNTA^-) complex.
(b) Configuration of the facial isomer of the mono nickel iminodiacetate (NiIDA) complex.
(c) and (d) Configuration of the two possible facial isomers of the bis nickel iminodiacetate [$\text{Ni}(\text{IDA})_2^-$] complex.

to give the best visual fit of the linewidth plot. A constant value for the contribution due to the scalar coupling relaxation mechanism, $1/T_{2m}$, which becomes appreciable only at high temperatures was assumed. This procedure yielded the following parameters:

| | |
|--|--|
| $A_1/h = 3.2 \times 10^7 \text{ Hz}$ | $A_2/h = 2.9 \times 10^7 \text{ Hz}$ |
| $\Delta H_1^\ddagger = 11.0 \text{ kcal}$ | $\Delta H_2^\ddagger = 11.3 \text{ kcal}$ |
| $\Delta S_1^\ddagger = 5.6 \text{ eu}$ | $\Delta S_2^\ddagger = -0.7 \text{ eu}$ |
| $k_{1,298^\circ} = 1.1 \times 10^6 \text{ sec}^{-1}$ | $k_{2,298^\circ} = 1.7 \times 10^4 \text{ sec}^{-1}$ |

The rate constant k_{298° represents the rate of exchange of a water molecule at a particular site on the coordinated ion. Rigorously, k_{298° is defined as $1/\tau_m$ (the reciprocal of Eq. 11), where τ_m is the mean lifetime of a water in a bound site.

However, a second treatment of the NiNTA linewidth data utilizing a six parameter non-linear least square computer fitting program yielded a slightly different set of parameters:

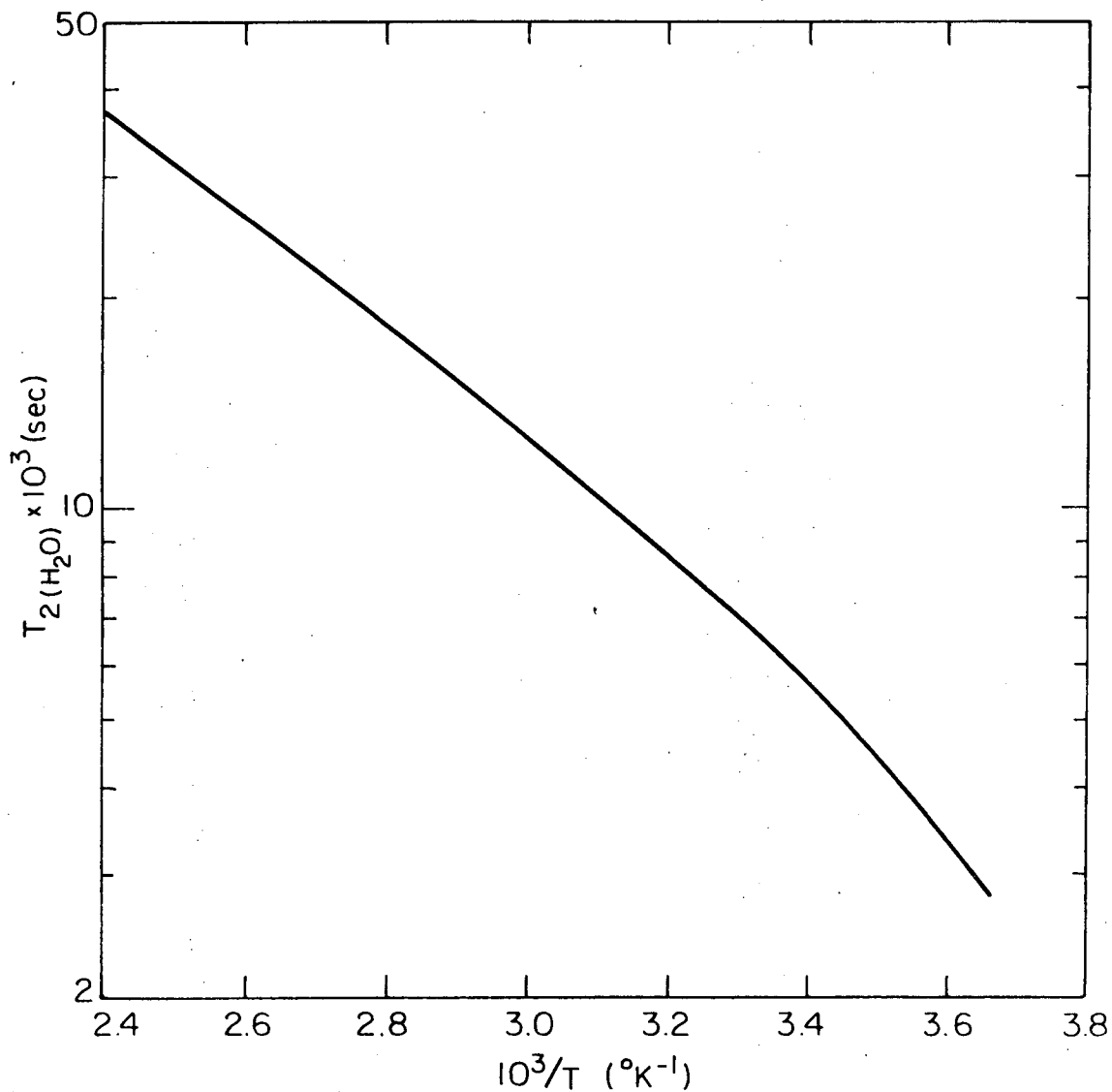
| | |
|--|--|
| $A_1/h = 2.9 \times 10^7 \text{ Hz}$ | $A_2/h = 2.6 \times 10^7 \text{ Hz}$ |
| $\Delta H_1^\ddagger = 13.6 \text{ kcal}$ | $\Delta H_2^\ddagger = 13.6 \text{ kcal}$ |
| $\Delta S_1^\ddagger = 15.1 \text{ eu}$ | $\Delta S_2^\ddagger = 6.6 \text{ eu}$ |
| $k_{1,298^\circ} = 1.3 \times 10^6 \text{ sec}^{-1}$ | $k_{2,298^\circ} = 1.8 \times 10^4 \text{ sec}^{-1}$ |

Both the ΔH^\ddagger and ΔS^\ddagger values are larger in this treatment than from the original hand fitting procedure. However, the computer program does show a strong correlation between the ΔH^\ddagger , ΔS^\ddagger pairs, suggesting that the curve can be fit fairly well with a number of different enthalpy/entropy values. Additionally, the rates calculated from the two different data treatments do not differ greatly and the ratio k_1/k_2 is quite similar in both cases, 65 for the hand fit and 72 for the computer fit.

Data have also been collected by Vriesenga at 2 MHz as well as at 8 MHz in the work described above in the hope of fixing the parameters more accurately. Proton NMR work where the proton signals of the NTA ligand have been studied as a function of temperature suggests that the faster water exchanging site is cis to the nitrogen coordinating position. This work has served as background and stimulation for the NiIDA investigation, whose main purpose has been in measuring the water exchange rates to determine which site exchanges more rapidly and the reasons for this labilization.

B. NiIDA Linewidth Measurements and Results

A plot of the oxygen-17 water blank relaxation time, $T_2(\text{H}_2\text{O})$, used in determining T_{2p} for the NiIDA system is shown in Fig. 3 plotted against $10^3/T$. This curve has been drawn to represent the proper water blank correction for a buffered 0.025 M NiIDA sample. Since the pH of the sample varies slightly with temperature (from pH 8.3 at 45°C to 8.7 at 0°C) and the linewidth varies with pH, a number of water samples adjusted to slightly different pH values had to be run within the

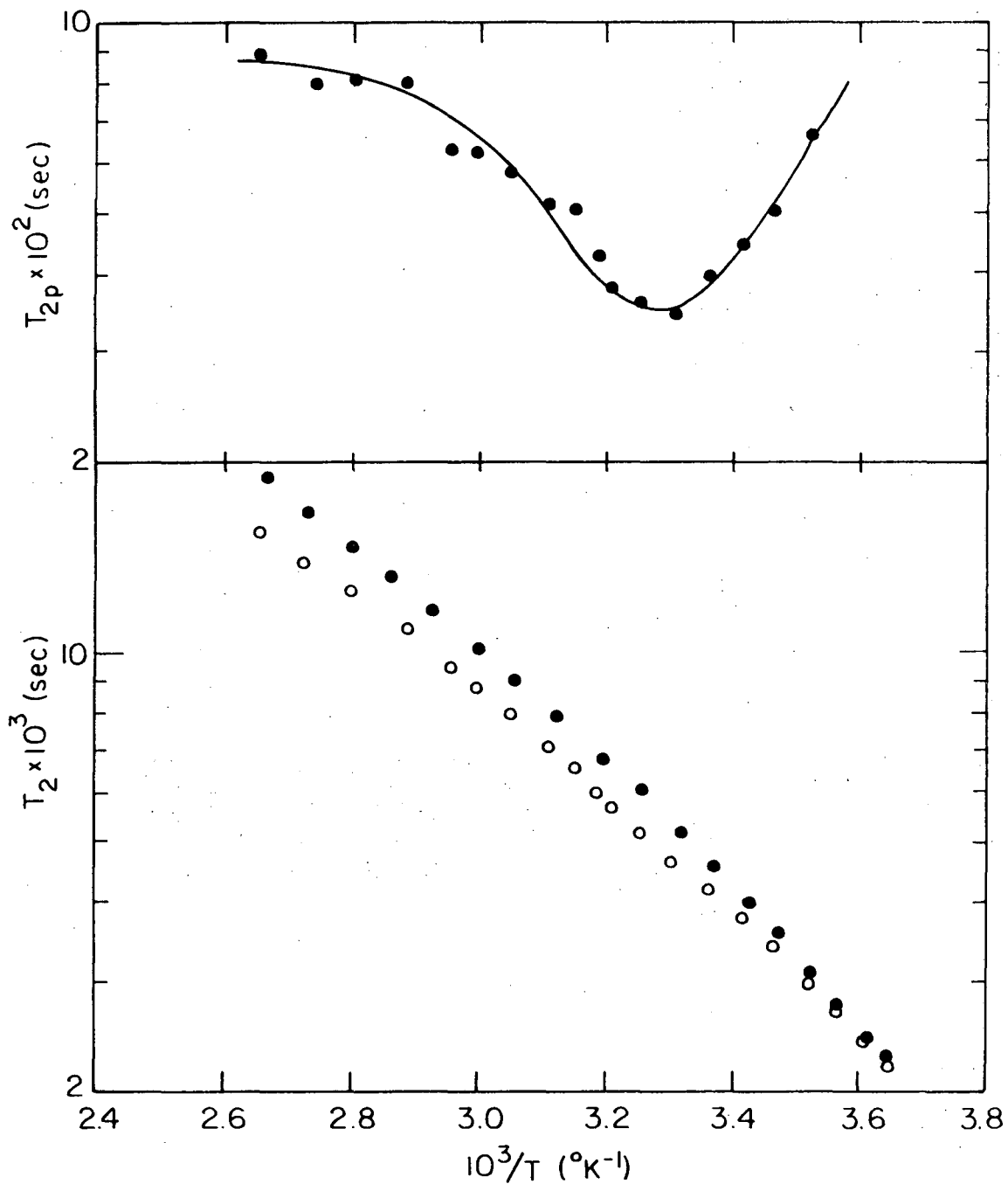


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Fig. 3. $T_{2(H_2O)} \times 10^3$ versus the reciprocal of absolute temperature for the oxygen-17 NMR of a water solution buffered with 0.10 M NH_4ClO_4 and adjusted to pH ~ 8.4 at 25° with NaOH. For exact pH adjustment as a function of temperature see data table in the appendix.

temperature range of 0°C to 45°C. Data for these samples and their pH values are listed in Appendix I. Above 45°C the curve in Fig. 3 represents the relaxation time of pure basic or acidic water, the rate of proton exchange from the water molecule being fast enough to eliminate the neutral region linebroadening effect.⁶ Water blank correction values for the NiIDA data points are also listed in Appendix I.

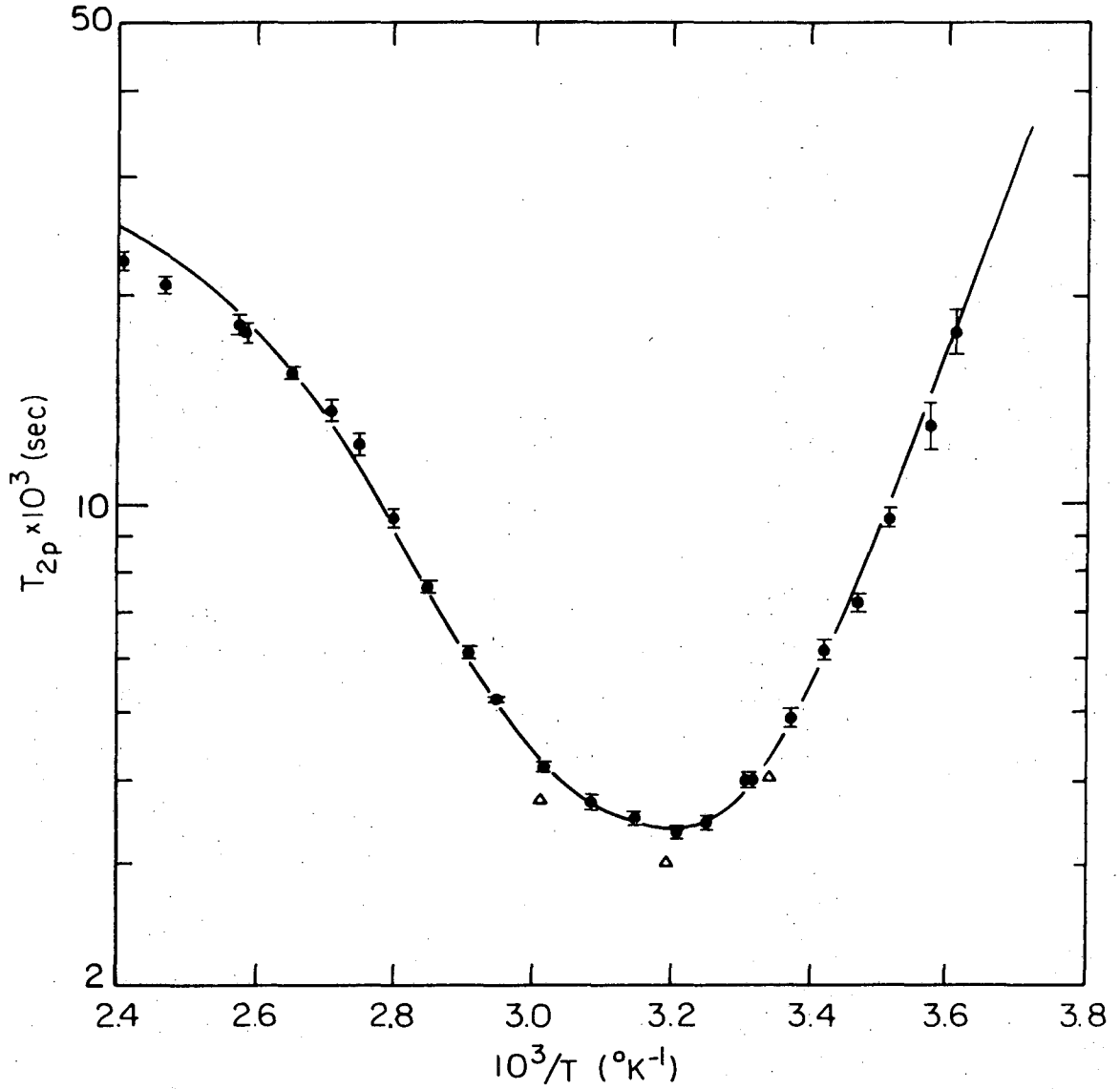
The correction for the free hexaquo nickel ion in the NiIDA solution was calculated from Neely's data^{1c} for a room temperature concentration of 2.99×10^{-4} M Ni^{2+} and is listed in Appendix I as $T_{2(\text{Ni}^{2+})}$. In addition, a correction for the cis complex, $\text{Ni}(\text{IDA})_2^{2-}$, whose concentration is approximately equivalent to that of the NiIDA species, must be considered. A plot of T_{2p} vs. $10^3/T$ for this complex is shown in the upper portion of Fig. 4. Although detectable linebroadening has been observed with a sample of 0.5 M $\text{Ni}(\text{IDA})_2^{2-}$, the broadening due to the presence of this species in the NiIDA sample is negligible relative to the NiIDA linebroadening itself. For example, at room temperature for the 0.0249 M NiIDA sample, the linewidth contribution due to the 0.0249 M $\text{Ni}(\text{IDA})_2^{2-}$ species will be approximately 0.3 Hz where the total paramagnetic linewidth is about 40 Hz; thus the $\text{Ni}(\text{IDA})_2^{2-}$ can be seen to contribute less than 1% of the total linebroadening. At higher and lower temperatures the per cent contribution is similar. The linebroadening data for $\text{Ni}(\text{IDA})_2^{2-}$ will be discussed in a later section for additional meaning.



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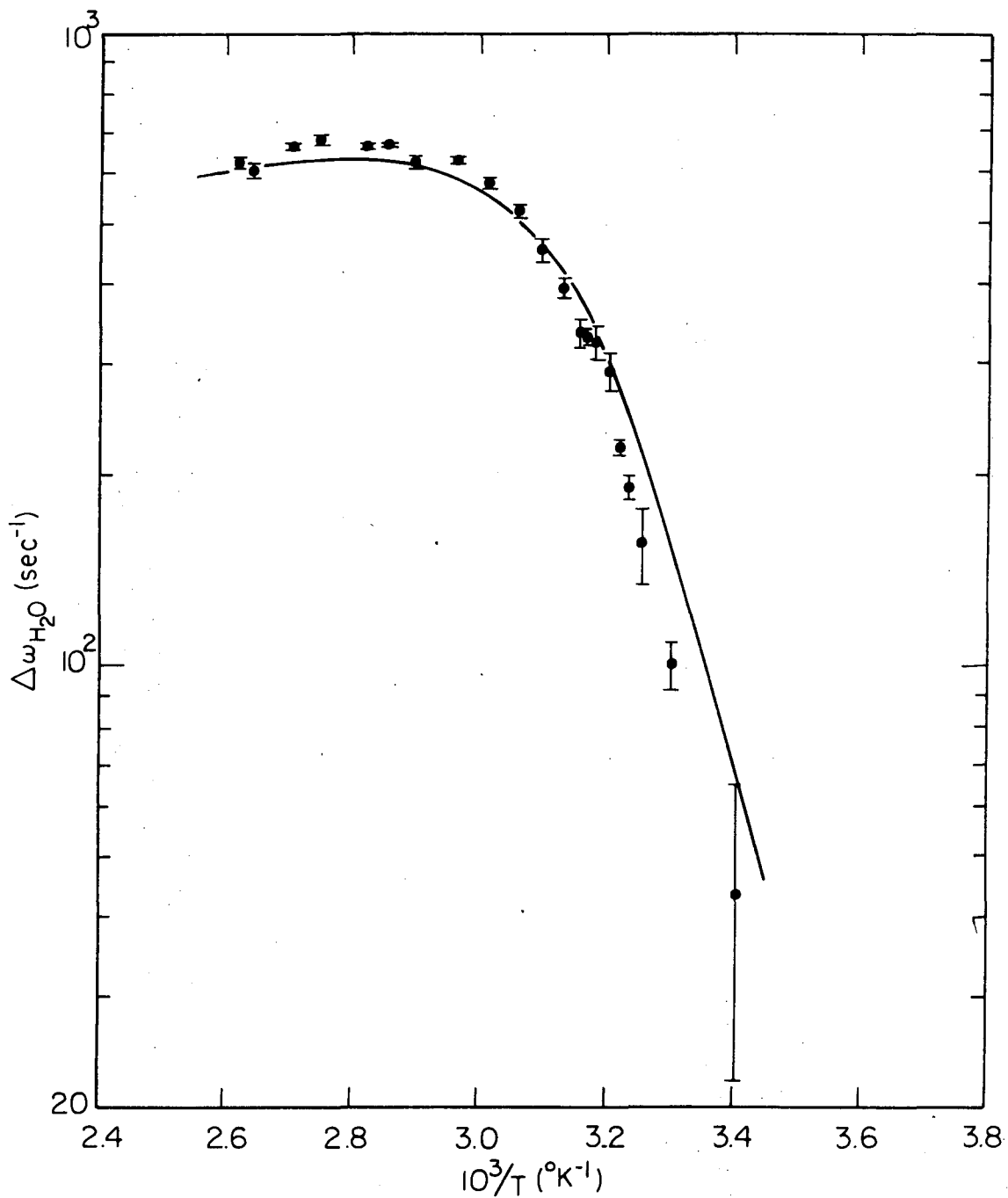
Fig. 4. Upper: $T_{2p} \times 10^2$ versus the reciprocal of absolute temperature for the oxygen-17 NMR of a 0.4970 M $\text{Ni}(\text{IDA})_2^{2-}$ sample at pH 9.2 at 25° . Lower: $T_2 \times 10^3$ versus the reciprocal of absolute temperature for \bullet - 0.515 M $\text{Zn}(\text{IDA})_2^{2-}$ at pH 9.2. \circ - 0.497 M $\text{Ni}(\text{IDA})_2^{2-}$ at pH 9.2.

A plot of T_{2p} vs. $10^3/T$ for the NiIDA complex is shown in Fig. 5, where the curve drawn through the data points represents a six parameter computer fit (ORGLS) for these linewidth data. Figure 6 shows the chemical shift of the bulk water resonance position relative to pure H_2O^{17} , $\Delta\omega_{H_2O}$, plotted against $10^3/T$ for a similar NiIDA sample where the solid line has been calculated using the parameters from the linewidth fit. Figure 7 shows the linewidth curve with the individual fitting for chemical exchange, $\Delta\omega_m$ relaxation, and scalar coupling relaxation for both the fast and slow exchanging sites. These lines were also calculated from the best fit parameters of the computer program. The intersection of the chemical exchange line, τ_{m_1}/P_{m_1} , and the $\Delta\omega_m$ line, $1/(P_{m_1} \tau_{m_1} \Delta\omega_{m_1}^2)$, can be seen to be to the left of and about a factor of two higher than the intersection of the other exchange line and $\Delta\omega_m$ line. Thus the fit indicates that there are two fast exchange sites and one slow site. Note that neither T_{2p} nor $\Delta\omega_{H_2O}$ has been multiplied by the concentration factor P_m as this may prove to be misleading when dealing with a curve fitted by an equation involving more than one P_m value (in this case P_{m_1} and P_{m_2} where $P_{m_2} = 2P_{m_1}$). The data points may of course be multiplied by $P_m = P_{m_1} + P_{m_2} = 3P_{m_1}$ for the purpose of comparison with samples of different concentration or with different chemical systems, but the fitting lines representing chemical exchange in Fig. 7 then become $\frac{3}{2} \tau_{m_2}$ and $3 \tau_{m_1}$ instead of the simpler τ_{m_2}/P_{m_2} and τ_{m_1}/P_{m_1} , and similarly for the chemical shift curve.



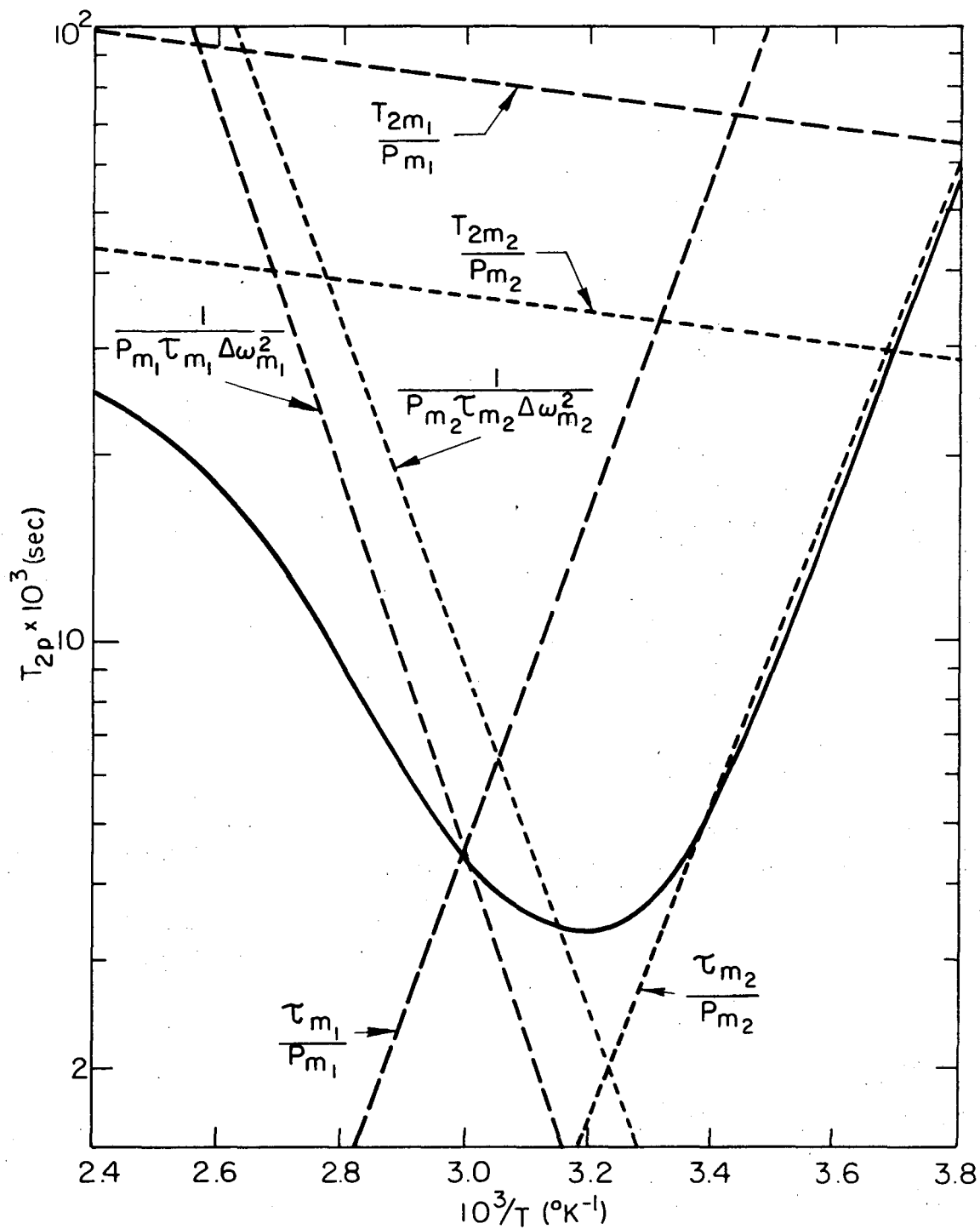
XBL 759-7198

Fig. 5. $T_{2p} \times 10^3$ versus the reciprocal of absolute temperature for the oxygen-17 NMR of a 0.0249 M NiIDA sample at pH 8.4 at 25°. The curve was calculated using Eq. (5) and the "linewidth only" computer fitting parameters in Table II. Δ - 0.0147 M NiIDA at pH 3.1.



XBL 759-7199

Fig. 6. $\Delta\omega_{H_2O}$ versus the reciprocal of absolute temperature for the oxygen-17 chemical shift of the bulk water in an aqueous solution of 0.0259 M NiIDA. The curve was calculated using Eq. (7) and the "linewidth only" computer fitting parameters in Table II.



XBL 759-7200

Fig. 7. $T_{2p} \times 10^3$ versus the reciprocal of absolute temperature for the oxygen-17 NMR of 0.0249 M NiIDA where the curve represents the "linewidth only" computer fit. The long dashed lines represent the individual fit lines as labeled for site 1; the short dashed lines represent the same for site 2.

A preliminary oxygen-17 NMR study of an acidic (pH 3.09) NiIDA sample was undertaken by Vriesenga^{5b} and is in general agreement with the high pH results reported here. In the present study a number of linewidth points were also recorded for an acidic (pH 3.13) 0.0147 M NiIDA sample and are shown with the proper concentration normalization as triangles in Fig. 5 and in the data table in Appendix I. The appropriate corrections have been made for other nickel species in the solution. The agreement is again reasonable and suggests that there are no large pH effects on the rate of water exchange from the NiIDA complex.

C. NiIDA Chemical Shift Measurements

The correction for the shift caused by hexaquo nickel ion in the NiIDA solution was also calculated from Neely's data.^{1c} Since no appreciable chemical exchange was observed from the $\text{Ni}(\text{IDA})_2^{2-}$ species, no correction to the shift data was required. However, because the chemical shift measurements were done in cylindrical tubes rather than spherical tubes in order to improve the signal to noise ratio, a magnetic susceptibility correction was necessary. This correction was determined using the normal equation for the field inside a long cylindrical tube,¹⁷

$$H = H_o \left(1 + \frac{2\pi}{3} \kappa\right) \quad (16)$$

where κ , the volume susceptibility of the sample, was determined from the relationships¹⁸

$$\kappa = [\text{Ni}] \chi_m \quad (17)$$

and

$$\chi_m = \frac{Ng_{\text{eff}}^2 \beta^2}{3kT} S(S+1) + 2N\beta^2 \Lambda \quad (18)$$

where N is Avogadro's number, and $\Lambda = \frac{4}{\Delta}$ for nickel where Δ is the crystal field splitting in cm^{-1} . The first term is the spin only term (quenched orbital angular momentum is assumed) written with g_{eff} to take into account the spin orbit coupling. The second term is a second order term arising from orbital contributions only. These corrections are tabulated in the NiIDA chemical shift data table in the appendix.

An additional correction was made to the chemical shift data in order to compensate for the fact that a non-logarithmic plot of $\Delta\omega_{\text{H}_2\text{O}}$ vs. $10^3/T$ as shown by the solid circles in Fig. 8 does not appear to approach zero asymptotically at low temperature as is expected from the equation for the shift, but rather approaches some negative value. At low temperature where for a nickel sample

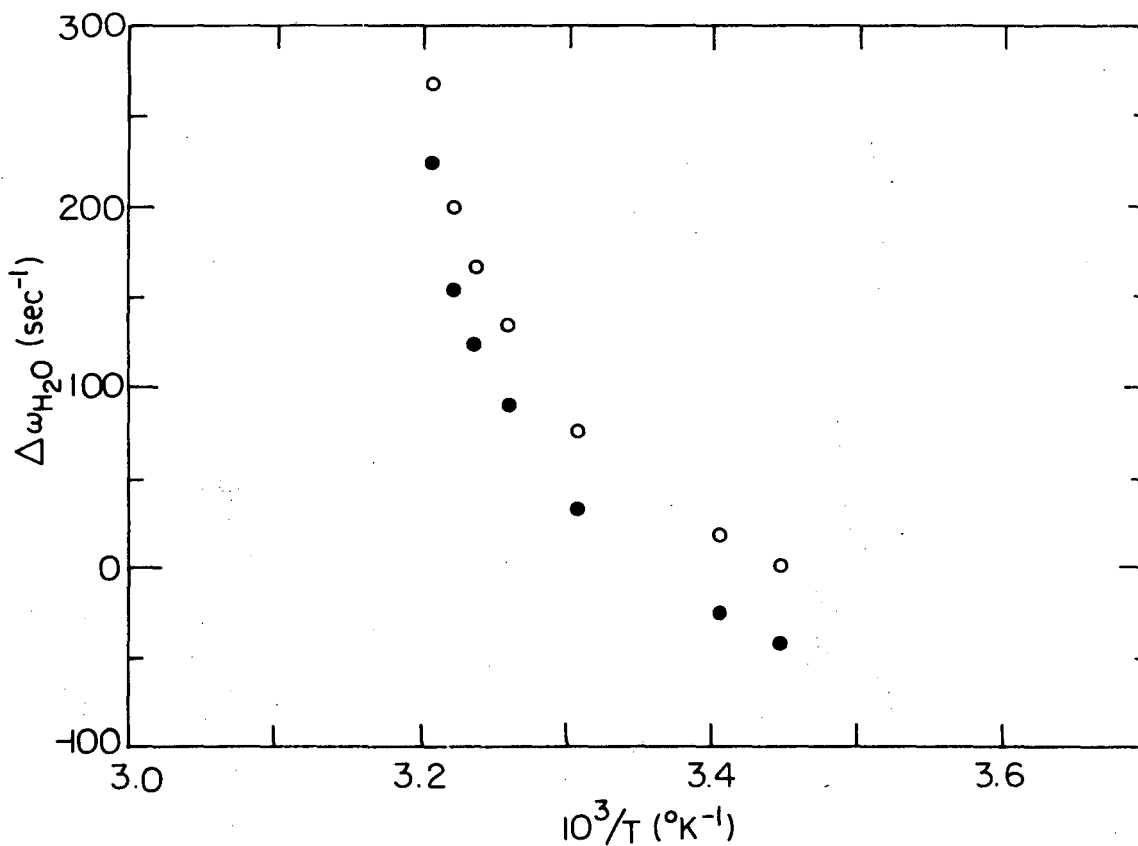
$$\Delta\omega_m^2 \gg \frac{1}{\tau_m^2}, \frac{1}{T_{2m}^2},$$

the chemical shift equation can be reduced to

$$\Delta\omega_{\text{H}_2\text{O}} = \frac{-P_m}{\tau_m^2 \Delta\omega_m} \quad (19)$$

and thus the correctional factor can be determined by adding a constant to the data points in the non-logarithmic plot until the curve approaches zero at low temperature or by adding a constant to the

low temperature points in a semi-log plot and checking them against a straight line fit. The second method was employed, using the seven lowest temperature data points, adding a small constant and checking the straight line fit by programmable calculator until a constant was found which best allowed the log of the data points to approximate a straight line. Although the higher temperature points used in this fitting procedure do not meet the condition $\Delta\omega_m^2 \gg 1/\tau_m^2$, the resulting constant, when added to the data points can be seen (open circles in Fig. 8) to give values which closely approach zero at low temperature. The value of this constant as shown in Fig. 8 and the chemical shift data table was determined to be 44 sec^{-1} which is approximately a 7% correction to the high temperature region of the curve. Although this baseline correction is not based on direct experimental observation, since there is no reason to expect that Eq. (7) is not valid for this experimental system, it was assumed that the normal low temperature chemical shift behavior would occur. Such a residual shift might be attributable to some unknown systematic error or to improper susceptibility corrections due either to irregular sample cell shape or to the paramagnetism of the glass sample cell itself. The susceptibility correction calculations made using Eq. (16) for an infinite cylinder yielded values on the order of 21 sec^{-1} . This correction is of the same size as the probable error for each data point, which is on the average about 15 sec^{-1} . A quick check at room temperature using a spherical sample cell for the paramagnetic nickel sample solution showed a chemical shift virtually identical to



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Fig. 8. Cartesian plot of $\Delta\omega_{\text{H}_2\text{O}}$ versus the reciprocal of absolute temperature for the H_2O oxygen-17 chemical shift of the bulk water in a 0.0259 M NiIDA solution showing:

- - original low temperature data points approaching a negative value at low temperature.
- - corrected data points, raised by a constant 44 sec^{-1} , approaching zero at low temperature.

the shift measured using the cylindrical sample tube where the infinite cylinder correction had been made. However the probable error in this set of measurements was larger (25 sec^{-1}) than for those made in cylinders due to the loss in the signal to noise ratio because of the diminished sample volume. According to Dickenson,^{17b} for a sample tube with this length to diameter ratio, approximately 70% of the full infinite cylinder correction should be used. Because of the large probable errors involved, this comparison does not really allow a more accurate calculation of the susceptibility correction, but serves only to show that within experimental error compatible chemical shift values could be measured in both spherical and cylindrical tubes. Thus, the full infinite cylinder correction was used, the difference between this and Dickenson's suggested 70% value being small relative to the probable error of the measurements. Although the spherical sample comparison does not rule out the possibility of an improper susceptibility correction (indeed, the combination of the probable errors from the cylinder and sphere measurements is not much less than the 44 sec^{-1} correction), the baseline correction made by fitting the low temperature data appears to be a reasonable manner in which to handle the data.

D. Computer Fitting Results

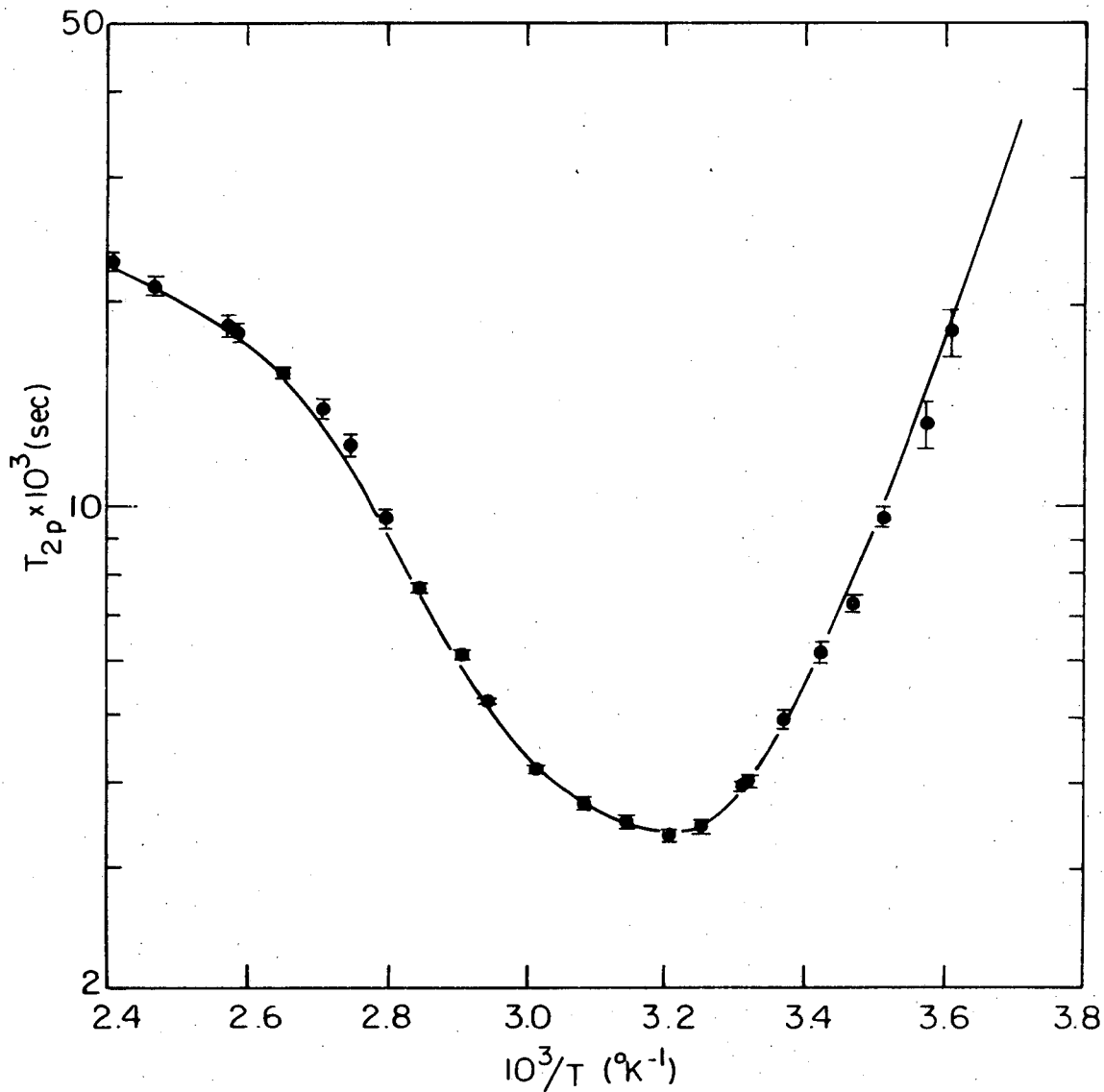
The curves through the data points of Figs. 5 and 6 represent the theoretically calculated curves using Eqs. (5) and (7) respectively and the computer fitted parameters listed in Table II, where the computer fit is to the linewidth data only.

Table II. Curve Fitting Parameters for 0.0249 M NiIDA Linewidth Data
and for 0.0259 M NiIDA Chemical Shift Data

| Linewidth Fit Only | ΔH^\ddagger (kcal/mole) | ΔS^\ddagger (eu) | $\frac{A}{h}$ (Hz) | τ_e^o (sec ⁻¹) | k_{298° (sec ⁻¹) |
|-------------------------------|------------------------------------|-----------------------------|--------------------|---------------------------------|---|
| Site 1 | 11.8 | 2.7 | 1.8×10^7 | 1.3×10^{-12} | 5.3×10^4 |
| Site 2 | 11.4 | 4.6 | 1.9×10^7 | 1.3×10^{-12} | 2.4×10^5 |
| Linewidth & Chemical Shift | | | | | |
| Site 1 | 13.8 | 8.8 | 1.9×10^7 | 1.5×10^{-12} | 4.3×10^4 |
| Site 2 | 12.1 | 7.2 | 2.0×10^7 | 1.5×10^{-12} | 2.9×10^5 |

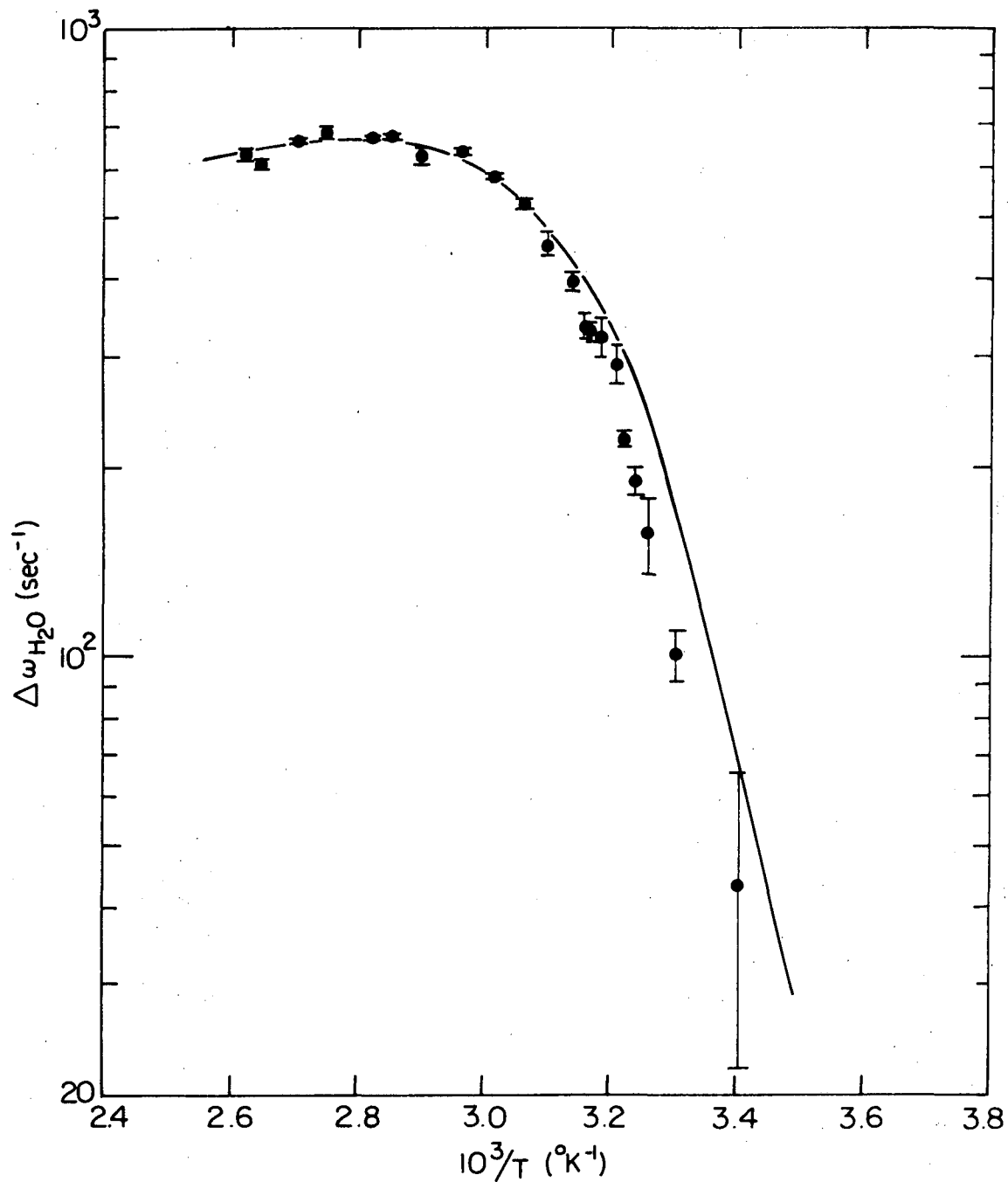
The second set of values comes from a similar computer fit, but where both the linewidth and chemical shift data were fit simultaneously as shown by the solid lines in Figs. 9 and 10. The quantities ΔH^\ddagger , ΔS^\ddagger and A/h were fitted parameters in all programs; however, the value of V in the electronic correlation time, τ_e , as defined by Eq. (15), was assumed to be 0.6 kcal for all sites. The fit is quite insensitive to the value of V , and 0.6 kcal was chosen as this value was suggested in the case of a similar treatment of Ni^{2+} by Neely.^{1c} Values between 0.3 and 1.2 kcal change the resulting exchange parameters negligibly. The value of τ_e^0 was varied manually in the computer program about a value determined by a hand fit of the high temperature linewidth data where the scalar coupling relaxation mechanism contributes appreciably, until the best fit of the data was obtained. No attempt was made to fit τ_e^0 and V simultaneously with the other parameters since there are few data points in the region where scalar coupling relaxation becomes the dominant mechanism.

It is, in fact, difficult to fit the linewidth data well with a six parameter (ΔH_1^\ddagger , ΔH_2^\ddagger , ΔS_1^\ddagger , ΔS_2^\ddagger , A_1 , A_2) computer program, as seen in Fig. 5. The computer shows a best fit for values of τ_e^0 and the scalar coupling constants A_1 and A_2 which visually do not fit the high temperature region well, while fitting the remainder of the curve quite well. As seen in Fig. 6 the solid curve calculated from these linewidth parameters is in reasonably good quantitative agreement with the chemical shift data collected; however the fit is not excellent in either the high or low temperature regions. In order to determine better the scalar



XBL 759-7201

Fig. 9. $T_{2p} \times 10^3$ versus the reciprocal of absolute temperature for the oxygen-17 NMR of 0.0249 M NiIDA at pH 8.4 at 25° where the curve represents the best fit to the linewidth and chemical shift data simultaneously. Fitting parameters are listed in Table II.



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Fig. 10. $\Delta\omega_{\text{H}_2\text{O}}$ versus the reciprocal of absolute temperature for the oxygen-17 chemical shift of the bulk water in an aqueous solution of 0.0259 M NiIDA. The curve represents the best fit to the linewidth and chemical shift data simultaneously. Fitting parameters are listed in Table II.

coupling constants a computer fitting routine was developed which could simultaneously fit the linewidth and the chemical shift data. It was hoped that this more complete utilization of the experimental data would provide a more accurate fix on the exchange parameters as well. The simultaneous fitting procedure can be seen in Fig. 9 to give a visually satisfying fit to the linewidth data over the full temperature range studied, including the scalar coupling region. However, it should be noted that although the values for ΔH_1^\ddagger , ΔS_1^\ddagger and ΔH_2^\ddagger , ΔS_2^\ddagger differ somewhat in the two different fitting procedures, the values for the rate of water exchange as shown in Table II are reasonably similar. The correlation matrices and the multiple correlation coefficients in the output of the computer programs indicated a strong correlation between the ΔH^\ddagger and ΔS^\ddagger values for a particular site in both the linewidth only and the simultaneous fitting procedures, as one would suspect in fitting an equation of this nature. There is a lesser degree of correlation between the two scalar coupling constants A_1 and A_2 as shown in Table III below, where plus or minus one represents complete correlation and zero no correlation at all.

E. Discussion of the Electronic Correlation Time, τ_e

The value of τ_e at 120°C, which is in the region of interest, as defined by Eq. (15) can be calculated from the parameters in Table I to be 2.8×10^{-12} sec for the linewidth fitting case and 3.2×10^{-12} sec for the simultaneous fitting routine. Neely^{1c} found the value of τ_e for the hexaquo nickel ion to be 8.2×10^{-12} sec at 120°C while Hynes¹⁹

Table III. Correlation Matrix for the "Linewidth-Only" Fit of the NiIDA Data.

| | A_1 | A_2 | ΔH_1^\ddagger | ΔH_2^\ddagger | ΔS_1^\ddagger | ΔS_2^\ddagger |
|-----------------------|--------|---------|-----------------------|-----------------------|-----------------------|-----------------------|
| A_1 | 1.0000 | -0.9748 | -0.8812 | 0.8331 | -0.8499 | 0.8288 |
| A_2 | | 1.0000 | 0.9369 | -0.7264 | 0.9136 | -0.7188 |
| ΔH_1^\ddagger | | | 1.0000 | -0.6626 | 0.9979 | -0.6511 |
| ΔH_2^\ddagger | | | | 1.0000 | -0.6336 | 0.9996 |
| ΔS_1^\ddagger | | | | | 1.0000 | -0.6216 |
| ΔS_2^\ddagger | | | | | | 1.0000 |

has calculated from proton NMR work a value of approximately 5×10^{-12} sec at 40°C for the same ion. Other values in this range have been reported elsewhere.^{20,21} Although the value of τ_e is not well defined by the present experimental work since such a small portion of the curve is determined by the scalar coupling relaxation mechanism, the values reported here for the NiIDA system are reasonable when compared to those reported for $\text{Ni}(\text{H}_2\text{O})_6^{2+}$. One would in general expect a shorter T_{1e} and T_{2e} and thus τ_e (as related in Eq. 14) for the NiIDA complex where there is a lower degree of symmetry than the hexaquo ion,²² as is observed here.

F. Discussion of Fitting Parameters and Curves

As previously noted, the visual fit of the linewidth data in the chemical exchange controlled region of the curve in Fig. 5 is excellent, with only three points further than one standard deviation from the calculated line, while these same parameters give substantial agreement with the chemical shift data in Fig. 6. It should be noted that although the high temperature linewidth data in the scalar coupling relaxation region could possibly have been fit better using a higher value for V in the expression for the electronic correlation time (Eq. 15), this would have little effect on the fitting of the same temperature region of the chemical shift curve where in the limit of rapid exchange Eq. (8), which is dependent only on $\Delta\omega_m$ or the fitting parameters A_1 and A_2 , determines the shape of the curve. Obviously a slightly increased value for $A_1 + A_2$ would produce a better fit in this region, but at the cost

of the fit of the linewidth data, as seen below. This would also raise the calculated chemical shift curve in the low temperature region, moving it away from the experimental points even further.

The linewidth data fit obtained from the simultaneous fitting procedure, while giving an excellent overall fit, greatly improves the fit in the high temperature scalar coupling relaxation region. Only seven of the twenty five points are more than one standard deviation from the calculated T_{2p} line over the entire temperature range studied. In the high temperature portion of the chemical shift curve the fit is excellent suggesting a good fix on the two scalar coupling constants. However, below 50°C there is an obvious, consistent positive deviation of the calculated line from the experimental points. This inconsistency does not appear to be due to any instrumental or experimental error of a constant nature as this was tested by the addition of a constant value equal to the difference between the experimental and calculated points at 29°C ($10^3/T = 3.305$) to each experimental point and this new data set was then used in the simultaneous fitting routine. While this procedure results in a calculated chemical shift curve much better fitted to this contrived data set than the former curve to the original data set, the fit of the linewidth data deteriorates greatly. This is in large part due to the increased value of the scalar coupling constants produced by the addition of the constant to the high temperature chemical shift data. This artificially high value for the scalar coupling constants gives a bad fit of the minima of the linewidth curve. The result is in a sense gratifying as it is reassuring to see

that with this six parameter fitting program the other variables do not tend to easily hide the fact that a particular variable or pair of variables has taken on an unreasonable value. Thus this procedure actually allows one to gain a measure of confidence in the sensitivity of the fitting procedure.

In order to remedy the discrepancy in the chemical shift fit it appears that a correction which is large at low temperature (on the order of 70 sec^{-1}) and decreases to little or no correction above 50°C is necessary. Actually it may be necessary to add to this the baseline correction which has already been made earlier in order to determine the real size of the discrepancy which must be dealt with. Thus, at low temperature the calculated curve would be about 112 sec^{-1} from the data points while at high temperature the discrepancy would be reduced to the baseline correction value of 44 sec^{-1} . A correction with such a temperature dependence would not arise from a paramagnetic impurity, nor is there any reason from the linebroadening data to suspect that an additional relaxation mechanism might be operative. There is of course a possibility that some unknown complexed nickel species is present in varying concentration over the affected temperature range, but this also is reasonably well precluded by both the linebroadening data and a visible spectroscopy study which showed no evidence of such an additional species. Although the reason(s) for this discrepancy cannot be pin-pointed, it should not in any case greatly alter the results of the computer fit of the linebroadening data or conclusions drawn from the fitting parameters.

In general, the linebroadening data appear to be more reliable than the chemical shift data as the blank corrections to the linewidth data were known more accurately. Also, the standard deviations for the linewidth points were generally smaller for a series of measurements than for the chemical shift points. For these reasons, the fitting parameters from the linewidth only computer fit will be used in the section on water exchange rate comparisons.

G. Ni(IDA)₂²⁻ Results and Discussion

As shown in Fig. 4, a solution of approximately 0.5 M Ni(IDA)₂²⁻ produces an observable linebroadening compared to a Zn(IDA)₂²⁻ blank. The effect is thought to be caused by water exchange at the carboxylate coordinating sites. These samples, unlike the NiIDA samples, were run at a pH of 9.2, eliminating the problem of neutral region broadening of the water blank. Since the effect was small relative to the blank, with the maximum broadening being only about 15% of the blank linewidth, and since the total metal complex concentration was fairly high, it was necessary to use a blank which would be, if possible, a diamagnetic duplicate of the Ni(IDA)₂²⁻, giving a sample with virtually identical non-magnetic properties such as viscosity which would affect the unbound water oxygen-17 linewidth. It should be noted that the curve drawn in the upper section of Fig. 4 is quite approximate due to the size of the paramagnetic broadening relative to the zinc blank (about 4 Hz broadening to a 30 Hz zinc blank at room temperature). However, the shape of this curve itself precludes the broadening from being a second

sphere effect, as the rate of chemical exchange from a second sphere position for nickel ion is fast enough that an exchange controlled region such as appears at the low temperature end of this plot could not be obtained for either a dipolar or scalar relaxation mechanism. Although the high temperature end of the curve bends over more rapidly than the curve for the mono nickel IDA complex, rough calculations show that neither a scalar or dipolar interaction would cause a broadening of this order of magnitude, and thus this effect is probably attributable either to the uncertainty in the data, or to a longer electronic correlation time (closer to that of nickel hexaquo ion) for this complex.

The minimum seen in Fig. 4 resembles that of Fig. 5, although not as wide and shifted to a slightly lower temperature. This broadening effect can be used in a qualitative numerical calculation to determine the per cent of the time that some one of the carboxylate groups will be dissociated from its coordinating position, along with the rate of water replacement at the carboxylate position by either another water or the carboxylate group. It would be advantageous to measure the chemical shift for the $\text{Ni}(\text{IDA})_2^{2-}$ system as a function of temperature in the limit of rapid exchange and, by assuming scalar coupling constants similar to those for NiIDA, determine P_m , thereby allowing the direct calculation of the per cent of the time that some one of the carboxylate arms is dissociated; however the shift is too small to be measured accurately with present instrumentation.

Although the structure of the $\text{Ni}(\text{IDA})_2^{2-}$ complex has not been resolved satisfactorily, with one group reporting approximately a 60/40

cis-facial/trans facial ratio^{4a} while another reported an 80/20 ratio,^{4b} it is not unreasonable to assume a mixture having a two to one ratio of the cis-facial species. As seen in Fig. 2, the cis-facial complex has two carboxylate sites which when dissociated provide water exchange positions which are cis to the coordinated nitrogen of the other ligand. These sites approximate the fast exchange positions of the mono complex where the strain on the carboxylate arms causes a labilization of the cis water positions, as will be discussed later. The remaining two sites on the cis-facial complex provide water sites trans to the coordinated nitrogen of the other ligand corresponding to the slow exchange position of the mono complex. For the trans-facial complex, all possible sites resemble fast exchanging positions. Thus with a two to one concentration ratio of these isomers, one also obtains a two to one fast site to slow site ratio, which is identical to the fast site to slow site ratio in the NiIDA complex. Assuming, then, that the ratio of fast to slow exchange lifetimes are the same for both the NiIDA complex and the $\text{Ni}(\text{IDA})_2^{2-}$ complex and also that the scalar coupling constants are identical for both systems, the two linebroadening curves (Figs. 4 and 5) can be compared and both a value of the per cent time of dissociation and the rate of water replacement can be estimated. Thus for single exchange site systems at the cross point of the exchange line, τ_m/P_m , and the $\Delta\omega_m$ relaxation line, $1/(P_m \tau_m \Delta\omega_m^2)$, a comparison of the paramagnetic linewidths gives the ratio of the P_m 's for the two systems if it is assumed that the $\Delta\omega_m$ values are the same. Using the

values of $1/T_{2p}$ at the minima of the two curves and the known value of P_m for the NiIDA system, the value of P_m for the $Ni(IDA)_2^{2-}$ system was calculated to be 1.3×10^{-4} and the per cent of the time that some one of the four carboxylate groups is dissociated was calculated to be 1.5%. Assuming the same percentage dissociation will hold at 25°C, and that the activation energy is about the same as for the NiIDA fast site, the rate of water replacement from a fast site on the bis complex was calculated to be $4 \times 10^5 \text{ sec}^{-1}$. It should be noted that it is not known whether the water is being replaced by another water molecule or by the acetate arm.

Since the T_{2p} minimum in the $Ni(IDA)_2^{2-}$ curve is somewhat narrower than that for the NiIDA, a value for the rate of water replacement and the per cent time of dissociation was also estimated assuming that only one exchange rate exists for the $Ni(IDA)_2^{2-}$ complex. In this case the site 2 theoretical lines of Fig. 7 were used for comparison and the per cent of the time that some one of the carboxylate groups is dissociated was calculated to be 1.2%. The rate of water replacement was calculated to be $3 \times 10^5 \text{ sec}^{-1}$ as compared with $4 \times 10^5 \text{ sec}^{-1}$ from above. These values are similar to the rate measured for the fast site of the NiIDA complex ($2.4 \times 10^5 \text{ sec}^{-1}$) which is not unreasonable. Although the uncertainty in these calculations is large, they serve to give a good idea of the strength of the carboxylate coordination in these complexes and insure that the linewidth correction to the NiIDA data from the $Ni(IDA)_2^{2-}$ complex is negligible.

V. DISCUSSION

Ligand Labilization Effects

In the past the rates of water exchange from partially complexed nickel ion have been measured for a number of different complexes and in particular for those with coordinated nitrogen sites. A number of complexes with both coordinated nitrogen and carboxylate oxygen have also been investigated. Much of this experimental work has been done using oxygen-17 NMR and the resulting rates of water exchange and activation parameters are listed in Table IV for a number of these complexes. Also included for comparative purposes are rates of water replacement from the first sphere of coordination by ammonia for some of the same nickel complexes, where the temperature jump technique was employed.^{23,24} Often both the exchange rates and activation parameters, ΔH^\ddagger and ΔS^\ddagger , have been reported and thus the effect of ligand substitutions may be examined in terms of the effect on these parameters as well as the exchange rate in general. It should be remembered in comparing data for the different chemical systems that the rate constants reported often have standard deviations of about 10%, with ΔH^\ddagger values reliable at best to about ± 0.5 kcal and ΔS^\ddagger values to an entropy unit or two. Differences in sample variables such as ionic strength, pH, and type of anion present may also result in differing values being reported. Scalar coupling constants are usually reported for the NMR experiments and in general are similar for most nickel complexes. As previously defined, k_{298}° is the reciprocal of the lifetime of a water molecule

Table IV. Water exchange rates and activation parameters.

| Complex | H ₂ O Exchange | | | | NH ₃ Replacement | | |
|---|--|------------------------|----------------------|---------------------|-----------------------------|---|------|
| | k _{298°} (sec ⁻¹) | ΔH [‡] (kcal) | ΔS [‡] (eu) | A/h(Hz) | Ref. | k _{298°} (sec ⁻¹) [†] | Ref. |
| Ni(H ₂ O) ₆ ²⁺ | 3.2×10 ⁴ | 13.9 | 10 | 2.7×10 ⁷ | 1c | 2.8×10 ⁴ | 24 |
| | 3.0×10 ⁴ | 10.8 | | 3.0×10 ⁷ | 1b | | |
| | 2.7×10 ⁴ | 11.6 | 0.6 | 4.3×10 ⁷ | 1a | | |
| | 4.4×10 ⁴ | 10.3 | -5.2 | 2.2×10 ⁷ | 25 | | |
| | 3.4×10 ⁴ | 12.1 | 2.9 | | 26 | | |
| | 3.6×10 ⁴ | 12.3 | 3.6 | 2.3×10 ⁷ | 27 | | |
| Ni(H ₂ O) ₅ (NH ₃) ²⁺ | 2.5×10 ⁵ | 10.6 | 1.8 | 2.5×10 ⁷ | 27 | | |
| Ni(H ₂ O) ₄ (NH ₃) ₂ ²⁺ | 6.1×10 ⁵ | 7.8 | -6.0 | 1.9×10 ⁷ | 27 | | |
| Ni(H ₂ O) ₄ en ²⁺ | 4.4×10 ⁵ | 10.0 | 10 | 2.2×10 ⁷ | 25 | 1.8×10 ⁵ | 24 |
| Ni(H ₂ O) ₃ (NH ₃) ₃ ²⁺ | 2.5×10 ⁶ | 10.2 | 5.0 | 2.6×10 ⁷ | 27 | | |
| Ni(H ₂ O) ₃ dien ²⁺ | 1.2×10 ⁶ | 5.5 | | 2.3×10 ⁷ | 2 | 8.6×10 ⁵ | 24 |
| Ni(H ₂ O) ₂ en ₂ ²⁺ | 5.4×10 ⁶ | 9.1 | | 2.3×10 ⁷ | 2 | 2.1×10 ⁸ | 24 |
| Ni(H ₂ O) ₂ trien ²⁺ | 2.9×10 ⁶ | 7 | | 2.1×10 ⁷ | 2 | 3.6×10 ⁶ | 24 |

Table IV. Continued.

| Complex | H ₂ O Exchange | | | | Ref. | NH ₃ Replacement | |
|---|--|------------------------|----------------------|---------------------|------|---|-------|
| | k _{298°} (sec ⁻¹) | ΔH [‡] (kcal) | ΔS [‡] (eu) | A/h(Hz) | | ^{-H₂O} k _{298°} (sec ⁻¹) [†] | Ref. |
| Ni(H ₂ O) ₂ tet ²⁺ | | | | | | 3.9×10 ⁴ | 3a |
| Ni(H ₂ O)(NH ₃) ₅ ²⁺ | | | | | | 4.3×10 ⁶ | 24 |
| Ni(H ₂ O) ₃ IDA | 5.3×10 ⁴ | 11.8 | 2.7 | 1.8×10 ⁷ | | 6.4×10 ^{4*} | 23,24 |
| | 2.4×10 ⁵ | 11.4 | 4.6 | 1.9×10 ⁷ | | | |
| Ni(H ₂ O) ₂ EDDA | | | | | | 1.8×10 ^{5*} | 23,24 |
| Ni(H ₂ O) ₂ NTA ⁻ | 1.7×10 ⁴ | 11.3 | -0.7 | 2.9×10 ⁷ | 32 | 1.6×10 ^{5*} | 23,24 |
| | 1.1×10 ⁶ | 11.0 | 5.6 | 3.2×10 ⁷ | | | |
| Ni(H ₂ O) ₂ tren ²⁺ | 8.2×10 ⁵ | 8.0 | | | 28 | 7.8×10 ^{6*} | 23,24 |
| | 9 ×10 ⁶ | 8.0 | | | | | |
| Ni(H ₂ O)EDTA ²⁻ | 7.0×10 ⁵ | 8.0 | -7.0 | 2.6×10 ⁷ | 29 | 2.6×10 ^{6*} | 23,24 |

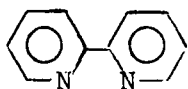
[†] Margerum's experiments gave no evidence of two different exchange rates for any of the complexes studied. In cases where two rates do actually exist, this technique probably reflects the faster rate primarily.

* Calculated from the value of k₁₂ in reference 24 using Eq. (20) and K_{os} = 0.1 M⁻¹. All other values for k_{298°}^{-H₂O} were calculated by Margerum et al. using the same equation and value for K_{os}.

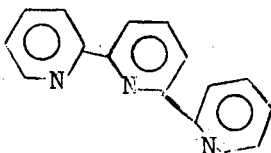
Abbreviations:

- IDA iminodiacetate $\text{NH}(\text{CH}_2\text{COO}^-)_2$
 NTA nitrilotriacetate $\text{N}(\text{CH}_2\text{COO}^-)_3$
 EDTA ethylenediaminetetraacetate $(\text{CH}_2\text{COO}^-)_2\text{N}(\text{CH}_2)_2\text{N}(\text{CH}_2\text{COO}^-)_2$
 en ethylenediamine $\text{NH}_2(\text{CH}_2)_2\text{NH}_2$
 dien diethylenetriamine $(\text{NH}_2\text{CH}_2\text{CH}_2)_2\text{NH}$
 trien triethylenetetramine $\text{NH}_2\text{CH}_2(\text{CH}_2\text{NHCH}_2)_2\text{CH}_2\text{NH}_2$
 tren β, β', β'' -triaminotriethylamine $(\text{NH}_2\text{CH}_2\text{CH}_2)_3\text{N}$
 EDDA ethylenediamine-N,N'-diacetate $(\text{CH}_2\text{COO}^-)\text{NHCH}_2\text{CH}_2\text{NH}(\text{CH}_2\text{COO}^-)$
 tet 1,4,8,11-tetraazaundecane $\text{NH}_2(\text{CH}_2)_2\text{NH}(\text{CH}_2)_3\text{NH}(\text{CH}_2)_2\text{NH}_2$

bipy 2,2'-bipyridyl



terpy 2,2',2''-terpyridyl



coordinated to the nickel at 25°C. Where more than one type of site exists for the water (e.g. NiIDA with two types of sites), each type of site should have its characteristic value of k_{298° . Where more than one type of water coordination site exists but the separate rate constants could not be resolved from the data (e.g. $\text{Ni}(\text{H}_2\text{O})_5(\text{NH}_3)^{2+}$), the results have been treated as if a single type of site were present. The ammonia substitution rates listed in Table IV as $k_{298^\circ}^{-\text{H}_2\text{O}}$ are the rate constants for the exchange of water in the inner coordination sphere as defined by the relationship

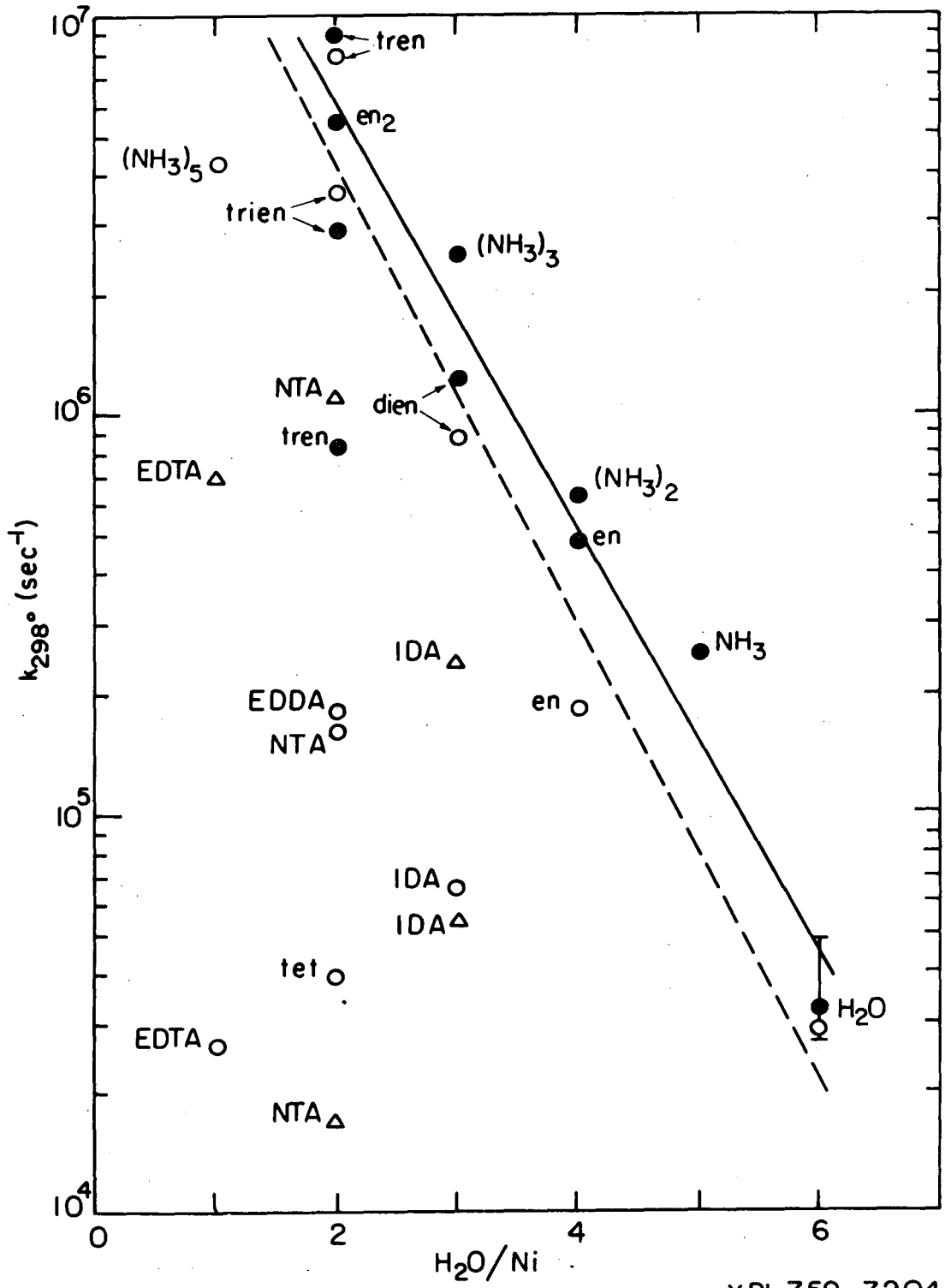
$$k_{298^\circ}^{-\text{H}_2\text{O}} = \frac{k_{12}}{K_{\text{os}}} \frac{6}{\text{no. of H}_2\text{O}} \quad (20)$$

where k_{12} is the forward rate constant for the ammonia substitution reaction and K_{os} is the outer sphere association constant of the amine complex.

Experimental results have tended to support the theory that coordinated aliphatic nitrogens tend to increase the rate of water exchange from the first sphere of hydration. In fact, Hunt² has shown in general that for the mono-, di, and tri-amine, en, en₂, dien and trien complexes of nickel, the log of the exchange rate, k_{298° , increases roughly linearly as the number of waters per nickel ion decreases, and this is shown in Fig. 11. The log of the rate of water replacement by ammonia, $k_{298^\circ}^{-\text{H}_2\text{O}}$, can also be seen in Fig. 11 to increase linearly as the

Fig. 11. The first order rate constant for water exchange from a particular site, k_{298° , versus the number of coordinated waters per nickel complex, H_2O/Ni , for:

- - nickel complexes with only aliphatic nitrogen coordinating ligands as labeled. The solid line is drawn through these points.
- Δ - nickel complexes containing both aliphatic nitrogen and carboxylate coordinating ligands.
- - The first order rate constant for water replacement by ammonia for a particular site, $k_{298^\circ}^{-H_2O}$, for nickel complexes as labeled. The dashed line is drawn through the points containing only aliphatic nitrogen coordinating positions.



XBL 759-7204

Fig. 11

number of waters per nickel ion decreases for the en, dien, trien and pentamine complexes. Only the en₂ and tet complexes are in gross disagreement with both the linear trend and the value expected from the water exchange rate determined by the NMR method. For the rates determined by NMR there does not appear to be any well defined trend in ΔH^\ddagger and ΔS^\ddagger values as the number of coordinated nitrogens increases (no activation parameters given for the ammonia substitution reactions). Whereas there is a ΔH^\ddagger increase in going from the en to the en₂ complex, the mono-, di-, and tri-amine complexes give irregular values. The same holds true for ΔS^\ddagger values although they are most commonly small and positive. The same rate enhancement factors are not found for ligands coordinated at unsaturated nitrogen sites such as bipy, terpy, bipy₂, or NCS⁻, although again for the first three complexes the logarithm of the exchange rates increase roughly linearly as the number of water coordinating positions decreases, but with a much smaller slope than for the aliphatic nitrogen complex.

These trends may be satisfactorily explained based on the electron donating/withdrawing ability of the complexing ligand. Aliphatic nitrogens, which are good electron donors, increase the electron density at the metal center, thereby weakening the remaining coordinated water bonds as is manifested by increased exchange rates. However for unsaturated or aromatic nitrogens, the electron donor properties are to some degree neutralized by π backbonding and thus labilization is greatly diminished from the value expected from aliphatic nitrogen coordination. Certainly this is an overly simplified view of the

problem as ligand geometry and configuration have not been taken into account. In view of this fact, the degree of correlation between $\log k_{298^\circ}$ and H_2O/Ni is somewhat surprising.

The effect of configuration is immediately evident in a direct comparison of the nickel tren and trien exchange rates and this will be discussed along with the EDTA, IDA, and NTA complexes later. Margerum²⁴ has reported the effects of N-alkyl substitution in diamine ligands on the rate of water replacement by ammonia. For example, the addition of an ethyl group to one nitrogen of en produced a 2 1/2 fold increase in the rate of ammonia substitution. Also based on a study of the rate of water replacement by ammonia, Cobb and Hague have concluded that the labilizing influence on the remaining water molecules is larger for trans- than for cis-nitrogen molecules.^{3a} They have reached this conclusion by comparing the data for the nickel en_2 , trien, and tren complexes which are all thought to have two remaining waters trans to coordinated nitrogens^{30a,b,c,d} with that for the nickel tet complex, where the two waters are opposite each other. Whereas the first three complexes have drastically labilized waters, water replacement by ammonia on the tet complex occurs barely faster than for the hexaquo nickel ion. However there appears to be no other evidence at this time to establish this trend; indeed, with such a drastic cis-trans labilization effect as claimed, one might have expected to measure two water exchange rates for complexes such as $Ni(H_2O)_4 en^{2+}$, where two remaining waters are cis and two are trans to coordinated nitrogens. Thus, although the number of coordinated nitrogen sites is indeed extremely

important, other factors may also greatly affect the degree of labilization produced by the ligand.

Carboxylate coordination labilization trends have not been as well determined, with cases of both slight rate enhancement and more often small decreases being reported.²⁴ It is difficult to determine directly the effect of the carboxylate coordination since the ligands most often also contain one or two coordinated nitrogens. EDTA (five coordinate) gives a slightly faster exchange rate for its remaining water than does its precursor en. Again, this may be more a configurational effect rather than an enhancement due to the three carboxylate groups and will be treated as such later. Unfortunately, there are no NMR water exchange data on either the N, N'-EDDA or the N,N-EDDA complexes to compare with those of the nickel en system. The N,N'-EDDA system in particular should yield straightforward information on the effects of carboxylate coordination with minimal configurational considerations, while a comparison of the two EDDA complexes would yield configurational information. Margerum²⁴ has measured a rate of $1.8 \times 10^5 \text{ sec}^{-1}$ for water replacement by ammonia in the nickel en complex and from data given in this same reference an identical rate of $1.8 \times 10^5 \text{ sec}^{-1}$ can be calculated for the N,N'-EDDA complex, suggesting that the carboxylate groups have little labilizing effect. It is not surprising that the carboxylate group does not provide a large labilization of the remaining coordinated waters, as it is not nearly the electron donor that nitrogen is. In fact, the carboxylate oxygen may not differ drastically from the water oxygen

in its static electronic (point dipole) properties, and thus would not be expected to have a much different effect from that of water itself on the remaining coordinated water molecules.

For the complexes of nickel with IDA, NTA and tren a different approach is necessary in order to explain the observance of two different water exchange rates. The results for the EDTA complex will also be considered here since this complex is to some degree structurally similar to these three ligands. In both the NiNTA^- and $\text{Ni}(\text{tren})^{2+}$ complexes the two remaining waters are located at structurally nonequivalent positions and similarly for NiIDA the cis and trans to nitrogen water sites are structurally quite different. Since in the NiIDA study it has been determined that the slower exchanging site is trans to the nitrogen group, it is assumed here that the same should hold true for the NiNTA^- complex (as postulated by Vriesenga on the basis of proton NMR^{5b}), and for the $\text{Ni}(\text{tren})$ complex the equivalent position will be that site trans to the tertiary nitrogen. The NiNTA^- and NiIDA trans site exchange rates generally fit the expected trend with a labilizing effect due to nitrogen being partially balanced by a small delabilizing effect from each carboxylate coordinating position. Although slightly slower than would be predicted from the $\text{Ni}(\text{trien})^{2+}$ and $\text{Ni}(\text{en})_2^{2+}$ results, the trans site rate for $\text{Ni}(\text{tren})^{2+}$ is considerably faster than for the NTA or IDA complexes as is expected due to the four nitrogen coordination sites.

The water exchange site on the five coordinate EDTA complex may either be cis to one nitrogen and trans to the other, or cis to both nitrogens. A structural study of the solid nickel EDTA complex by Smith and Hoard has shown that one of the cis/trans positions remains uncoordinated by the EDTA ligand.³² This isomer appears to provide a water exchange site similar to the fast IDA site in terms of the effects of bond strain about the coordinated nitrogens, while the latter provides a site similar to the fast NTA site. Since the EDTA complex has one more coordinated nitrogen than the NTA complex, should the cis/trans isomer prevail, an exchange rate enhanced by about a factor of three might be predicted along with an additional enhancement of about a factor of four and a half due to the IDA type strain thought to be present. This procedure yields an exchange rate of about $2 \times 10^5 \text{ sec}^{-1}$ compared with the reported value of 7×10^5 . For the cis/cis isomer the strain enhancement factor would be expected to be considerably greater as with the NTA complex (enhancement factor of about 70), and thus including the enhancement of the additional coordinated nitrogen a rate of about $3 \times 10^6 \text{ sec}^{-1}$ would be predicted. Although the strain produced in each of these isomers may not be exactly like that of either the IDA or NTA complexes due to the steric interactions of the ethylene link in the ligand, it seems reasonable that the observed value falls somewhere between the two predicted values.

The fast exchanging sites on the NTA and tren complexes are both cis to a tertiary nitrogen and the two fast IDA sites are cis to a secondary nitrogen. Fisher-Hirshfelder models of the complexes show a large degree of bond strain about the tertiary or secondary nitrogens,

making it hard to fasten all of the remaining coordinating positions simultaneously. This strain about the nitrogen would of course operate to push the complex into such a configuration as to allow tetrahedral bonding to the central nitrogen. This can be achieved if the complex becomes trigonal bipyramidal in shape, eliminating one cis coordinating position. A number of similar Ni^{2+} complexes such as nickel tris (2-methylaminoethyl)amine, where the aliphatic substitution to the tren ligand has sterically increased the strain in the octahedral configuration, are known to have trigonal bipyramidal structure in the solid state and are thought to retain this structure in solution.^{31a,b} For the nickel IDA, NTA, tren, and EDTA complexes a tendency toward such a configurational intermediate would explain the unusually high rates of water exchange from the cis water sites. One would expect the greater strain in the tren and NTA tertiary nitrogens to produce a faster water exchange rate than the strain in the secondary nitrogen of the IDA complex, as is observed, NTA's fast rate being approximately five times that of IDA. Additionally, the faster exchange rate for the tren complex relative to the NTA is in line with the fourfold increase in coordinated nitrogens. The $\text{Ni}(\text{IDA})_2^{2-}$ where exchange is occurring at a dissociated carboxylate group also appears to fit the general pattern described above, giving a rate somewhat faster than for the fast site of the mono complex, as would be expected due to the addition of both a nitrogen and a carboxylate coordinating group.

The above observations lead to a set of conclusions which establish or reinforce a number of ligand labilization trends. First, coordinated aliphatic nitrogen labilizes the remaining water molecules present in the complex, although the degree of labilization may be affected by substitution on the coordinating nitrogen group. Secondly, coordinated carboxylate groups have a small delabilizing effect. Third, the strain produced by certain ligand configurations may cause a large degree of labilization. In particular, tertiary nitrogen bond strain can produce a water exchange rate enhancement as great as that expected from the coordination of three aliphatic nitrogens. Secondary nitrogens appear to have a similar but diminished effect. Fourth, conformation of the ligand causes measureable ligand labilization effects which are yet of an unclear nature. However, based on the currently available data, one or more exceptions can be found to any one of these trends, which illustrate, in general, the complexity of predicting ligand labilization trends where the effects of ligand electron donor ability of the particular ligand involved, configurational strain, and conformation may all have roles of nearly equal importance in determining the degree of labilization of the remaining coordinated water molecules. Due to the limited number of systems which have been studied to this date and due to the complexity of these systems, no numerical treatment of either the carboxylate labilization or the nitrogen bond strain effects can be given.

ACKNOWLEDGMENTS

This thesis was prepared under the direction of Professor Robert E. Connick and I wish to express my sincere gratitude to him for his patient counsel and enthusiastic support of my research. The proficient and good natured electronics assistance of Phil Eggars is gratefully acknowledged as is the cooperation of Phil Frazier in providing a loan instrument vital to these experiments. Dr. Mark Zetter is thanked for sharing his expertise in computer programming.

I wish to thank all the members of my research group, past and present, and in particular Bill Earl for his continual assistance both inside and outside the chemistry laboratory, and Kevin Klotter for his help while filing this thesis. My sincerest thanks to: Jere Rowland for continual support of my undertakings and her ability to put chemical problems into a cosmic perspective, Kevin Leary for his ability to put cosmic problems into a scientific perspective, and lastly to my parents for their continual interest in my well being from whence, no doubt, springs my interest in science.

Financial assistance from the Lawrence Berkeley Laboratory is gratefully acknowledged. This work was performed under the auspices of the U. S. Energy, Research and Development Administration.

APPENDIX I. EXPERIMENTAL DATA

Table Ia. Oxygen-17 bulk water linewidth data as a function of temperature (below 40°C) and pH using 0.10 M NH_4ClO_4 adjusted to the desired pH with NaOH.*

| Temp(°C) | $10^3/T(^{\circ}\text{K}^{-1})$ | $T_{2\text{H}_2\text{O}} \times 10^3(\text{sec})$ |
|---------------------------|---------------------------------|---|
| Sample 1: pH 8.42 at 26°C | | |
| 40.36 | 3.190 | 8.73 |
| 36.07 | 3.234 | 7.97 |
| 28.85 | 3.311 | 6.96 |
| Sample 2: pH 8.53 at 16°C | | |
| 20.07 | 3.410 | 5.60 |
| 15.75 | 3.461 | 4.92 |
| 11.26 | 3.516 | 4.20 |
| Sample 3: pH 8.61 at 5°C | | |
| 8.56 | 3.550 | 3.88 |
| 4.32 | 3.604 | 3.17 |
| 1.41 | 3.642 | 2.81 |

*The change in pH of the 0.10 M NH_4ClO_4 solution as a function of temperature was similar to, but slightly larger than the 0.025 M NiIDA samples.

Table Ib. The pH of a 0.025 M NiIDA sample as a function of temperature.

| Temp(°C) | pH |
|----------|------|
| 31.2 | 8.37 |
| 21.4 | 8.45 |
| 17.6 | 8.49 |
| 9.2 | 8.60 |
| 1.4 | 8.65 |

Table IIa. Linewidth data for a 0.0249 M NiIDA sample adjusted to pH 8.4 at 25°C. Sample also contains 0.0249 M Ni(IDA)₂²⁻ and 2.99×10⁻⁴ M Ni²⁺ and was made by adding 32.6 mg Ni(ClO₄)₂·6H₂O, 17.8 mg NH(CH₂COOH), and 1.1 mg NH₄ClO₄ to 1.8474 g of 12% H₂O¹⁷ (~24% H₂O¹⁸). P_m(Ni²⁺)=3.232×10⁻⁵; P_m(NiIDA)**=1.346×10⁻³

| Temp. (°C) | 10 ³ /T(°K ⁻¹) | T _{2H₂O} ×10 ³ (sec)* | T _{2p} (Ni ²⁺)(sec) | T _{2p} (NiIDA)×10 ³ (sec) |
|------------|---------------------------------------|--|--|---|
| 3.93 | 3.609 | 3.10 | 7.18 | 17.80 |
| 6.79 | 3.572 | 3.62 | 4.55 | 13.05 |
| 11.49 | 3.513 | 4.27 | 2.99 | 9.59 |
| 15.18 | 3.468 | 4.80 | 2.16 | 7.72 |
| 19.10 | 3.422 | 5.38 | 1.57 | 6.18 |
| 23.54 | 3.370 | 6.03 | 1.08 | 4.92 |
| 28.24 | 3.318 | 6.81 | 0.743 | 4.01 |
| 28.39 | 3.316 | 6.86 | 0.733 | 3.97 |
| 28.80 | 3.312 | 6.91 | 0.712 | 3.98 |
| 34.80 | 3.247 | 7.91 | 0.446 | 3.44 |
| 39.05 | 3.203 | 8.60 | 0.325 | 3.34 |
| 45.02 | 3.143 | 9.67 | 0.217 | 3.51 |
| 51.40 | 3.081 | 10.8 | 0.154 | 3.70 |
| 58.72 | 3.013 | 12.3 | 0.112 | 4.19 |
| 66.39 | 2.945 | 14.0 | 0.0938 | 5.28 |
| 70.84 | 2.907 | 15.0 | 0.0913 | 6.14 |
| 78.24 | 2.846 | 16.7 | 0.0962 | 7.64 |
| 84.27 | 2.798 | 18.4 | 0.106 | 9.57 |
| 91.02 | 2.746 | 20.2 | 0.119 | 12.3 |
| 96.28 | 2.707 | 21.8 | 0.130 | 13.8 |
| 104.38 | 2.649 | 24.4 | 0.148 | 15.6 |
| 113.98 | 2.583 | 27.1 | 0.166 | 17.5 |
| 115.15 | 2.575 | 28.0 | 0.168 | 18.02 |
| 132.24 | 2.467 | 34.3 | 0.191 | 20.5 |
| 142.88 | 2.404 | 38.7 | 0.199 | 21.3 |

* Values for T_{2H₂O} taken from Fig. 3. ** where P_m(NiIDA) = P_{m1} + P_{m2}.

Table IIb. Linewidth data for a 0.0147 M NiIDA sample adjusted to pH 3.1 at 25°C. Sample also contains 3.04×10^{-3} M $\text{Ni}(\text{IDA})_2^{2-}$ and 8.57×10^{-4} M Ni^{2+} and was made by adding 9.5 mg $\text{Ni}(\text{ClO}_4)_2 \cdot 6\text{H}_2\text{O}$, and 20.7 mg $\text{NH}(\text{CH}_2\text{COOH})$ to 1.4220 g of 6% H_2O^{17} (12% H_2O^{18}).
 $P_m(\text{Ni}^{2+}) = 9.265 \times 10^{-5}$; $P_m(\text{NiIDA})^* = 7.946 \times 10^{-4}$.

| Temp. (°C) | $10^3/T(^{\circ}\text{K}^{-1})$ | $T_{2\text{H}_2\text{O}} \times 10^3 (\text{sec})^{**}$ | $T_{2p}(\text{Ni}^{2+})$ (sec) | $T_{2p}(\text{NiIDA}) \times 10^3$ (sec) |
|------------|---------------------------------|---|-----------------------------------|---|
| 27.02 | 3.339 | 6.81 | 0.287 | 6.74 |
| 40.21 | 3.191 | 8.87 | 0.105 | 5.11 |
| 59.57 | 3.005 | 12.55 | 3.78×10^{-2} | 6.34 |

* where $P_m(\text{NiIDA}) = P_{m_1} + P_{m_2}$.

** normal acidic water blank data used.

Table III. Chemical shift data for a 0.0259 M NiIDA sample adjusted to pH 8.4 at 25°C. Sample also contains 0.0222 M Ni(IDA)₂²⁻ and 3.63×10⁻⁴ M Ni²⁺ and was made by adding 31.4 mg Ni(ClO₄)₂·6H₂O, 16.9 mg NH(CH₂COOH), and 1.2 mg NH₄ClO₄ to 1.8312 g of 10% H₂O¹⁷ (~20% H₂O¹⁸). γ_{H_K} is the magnetic susceptibility correction. All measured shifts were downfield shifts. P_m(Ni²⁺) = 3.922×10⁻⁵; P_m(NiIDA)^{*} = 1.400×10⁻³.

| 10 ³ /T(°K ⁻¹) | -Δω _{H₂O} ^{Ni²⁺} (sec ⁻¹) | γ _{H_K} (sec ⁻¹) | -Δω _{H₂O} (sec ⁻¹) | -Δω _{H₂O} +C ^{**} (sec ⁻¹) |
|---------------------------------------|---|---|--|---|
| 3.446 | 0.01 | 25.1 | -42.8 | 0.7 |
| 3.404 | 0.05 | 24.8 | -0.17 | 43.3 |
| 3.305 | 0.11 | 24.1 | 56.8 | 100 |
| 3.258 | 0.22 | 23.7 | 113 | 157 |
| 3.236 | 0.28 | 23.6 | 148 | 191 |
| 3.220 | 0.39 | 23.4 | 178 | 222 |
| 3.207 | 0.45 | 23.3 | 248 | 291 |
| 3.183 | 0.67 | 23.1 | 281 | 325 |
| 3.167 | 0.82 | 23.0 | 286 | 329 |
| 3.159 | 0.86 | 22.9 | 293 | 336 |
| 3.131 | 1.33 | 22.7 | 352 | 396 |
| 3.095 | 2.00 | 22.4 | 410 | 454 |
| 3.060 | 2.86 | 22.2 | 480 | 524 |
| 3.010 | 4.71 | 21.7 | 537 | 581 |
| 2.962 | 7.26 | 21.4 | 590 | 634 |
| 2.894 | 11.9 | 20.8 | 582 | 626 |

* where P_m(NiIDA) = P_{m₁} + P_{m₂}.

** C = 44 (sec⁻¹).

Table III. Continued.

| $10^3/T(^{\circ}\text{K}^{-1})$ | $-\Delta\omega_{\text{H}_2\text{O}}^{\text{Ni}^{2+}} (\text{sec}^{-1})$ | $\gamma_{\text{H}_\kappa} (\text{sec}^{-1})$ | $-\Delta\omega_{\text{H}_2\text{O}} (\text{sec}^{-1})$ | $-\Delta\omega_{\text{H}_2\text{O}} + \text{C}^{**} (\text{sec}^{-1})$ |
|---------------------------------|---|--|--|--|
| 2.850 | 14.9 | 20.4 | 628 | 672 |
| 2.819 | 16.5 | 20.2 | 620 | 664 |
| 2.747 | 19.4 | 19.6 | 636 | 680 |
| 2.706 | 20.0 | 19.2 | 616 | 660 |
| 2.642 | 20.6 | 18.8 | 562 | 606 |
| 2.617 | 20.8 | 18.5 | 585 | 629 |

Table IVa. Linewidth data for a 0.497 M Ni(IDA)₂²⁻ sample adjusted to pH 9.2 at 25°C. The sample was made by adding 328.8 mg Ni(ClO₄)₂·6H₂O, 264.0 mg NH(CH₂COOH), and 10.7 mg NH₄ClO₄ to 1.8080 g of 7% H₂O¹⁷ (~14% H₂O¹⁸).

| Temp. (°C) | 10 ³ /T(°K ⁻¹) | T _{2H₂O} *×10 ³ (sec) | T _{2p} ×10 ² (sec) |
|------------|---------------------------------------|--|--|
| 10.80 | 3.522 | 3.12 | 6.62 |
| 15.60 | 3.463 | 3.66 | 4.98 |
| 19.78 | 3.414 | 4.13 | 4.48 |
| 24.54 | 3.359 | 4.70 | 3.98 |
| 29.39 | 3.305 | 5.38 | 3.43 |
| 34.29 | 3.253 | 6.06 | 3.60 |
| 38.64 | 3.207 | 6.66 | 3.80 |
| 40.93 | 3.184 | 7.00 | 4.29 |
| 44.26 | 3.150 | 7.51 | 5.10 |
| 48.74 | 3.107 | 8.19 | 5.18 |
| 55.02 | 3.047 | 9.26 | 5.81 |
| 61.20 | 2.991 | 10.3 | 6.21 |
| 65.61 | 2.952 | 11.2 | 6.21 |
| 73.42 | 2.885 | 12.6 | 8.02 |
| 84.33 | 2.797 | 14.8 | 8.05 |
| 94.13 | 2.723 | 16.7 | 7.95 |
| 103.94 | 2.652 | 18.9 | 8.83 |

*T_{2H₂O} determined by measuring T₂ of a 0.515 M Zn(IDA)₂²⁻ sample.

Table IVb. Linewidth data for a 0.515 M $\text{Zn}(\text{IDA})_2^{2-}$ sample adjusted to pH 9.2 at 25°C. The sample was made by adding 334.3 mg $\text{Zn}(\text{ClO}_4)_2 \cdot 6\text{H}_2\text{O}$, 263.7 mg $\text{NH}(\text{CH}_2\text{COOH})$, and 11.3 mg NH_4ClO_4 to 1.7740 g of 7% H_2O^{17} (14% H_2O^{18}).

| Temp. (°C) | $10^3/\text{T}(\text{°K}^{-1})$ | $\text{T}_{2\text{H}_2\text{O}} \times 10^3 = \text{T}_2(\text{Zn}(\text{IDA})_2^{2-})$ (sec) |
|------------|---------------------------------|---|
| 1.46 | 3.641 | 2.27 |
| 3.74 | 3.611 | 2.46 |
| 7.35 | 3.565 | 2.75 |
| 10.58 | 3.524 | 3.11 |
| 14.88 | 3.472 | 3.57 |
| 18.88 | 3.424 | 4.00 |
| 23.59 | 3.370 | 4.57 |
| 28.24 | 3.318 | 5.20 |
| 34.10 | 3.255 | 6.07 |
| 39.69 | 3.196 | 6.82 |
| 47.07 | 3.123 | 7.94 |
| 53.88 | 3.058 | 9.05 |
| 60.39 | 2.998 | 10.2 |
| 68.73 | 2.925 | 11.7 |
| 76.20 | 2.862 | 13.2 |
| 84.40 | 2.797 | 14.7 |
| 93.39 | 2.728 | 16.7 |
| 101.60 | 2.668 | 18.9 |

APPENDIX II.

Computer Programs

An example of a computer subroutine for the Dye and Nicely General Purpose Curve Fitting Program¹⁶ for fitting two equations (linewidth and chemical shift) simultaneously:

C*****SIMULTANEOUS CHEMICAL SHIFT AND LINEWIDTH SUBROUTINE*****

```
      SUBROUTINE EGN
      COMMON KOUNI,ITAPE,JTAPE,IWT,LAP,AINCR,NOPT,NOVAR,NOUNK,A,U,ITMAX,
      1*IX,IECT,IAV,RESID,IAK,EPS,ITYP,AX,RXTYP,DALI,FOP,FU,FU,P,ZL,TO,
      2EIGVAL,AST,T,D,L,M,JUJ,Y,DY,VECT,NCST,CONST,LOG,JJ
      DIMENSION X(4,100), U(20), WIX(4,100), XX(4), FOP(100), FU(100),
      1FU(100), P(20,21), VECT(20,21), ZL(100), TO(20), EIGVAL(20),
      2AST(100), Y(10), DY(10), CONST(16), CMUKH7(4), CMUK1(4), PH25(4),
      3TL(4), IL(4), TA(4), ZH(100), ZK7(100), ZKH(100), EM(100),ZK1(100)
      GO TO (3,4,5,6,1,7,8), ITP
1 CONTINUE
      ITAPE = 5
      JTAPE = 6
      LOG = 0
      WRITE (JTAPE,2)
2 FORMAT (1H,'CHEMICAL SHIFT AND LINEWIDTH DATA')
      NUNK = 6
      NVAR = 2
      CC = 0.0247
      CA = 0.0259
      TE = 1.24E-12
      PROGRAM WRITTEN SO THAT
      C      CC IS CONCENTRATION OF NI(L) FOR LINEWIDTH DATA
      C      CA IS CONCENTRATION OF NI(L) FOR CHEMICAL SHIFT DATA
      C      TE IS ELECTRONIC CORR. TIME TAU E U
      C      U(1) IS SCALAR COUPLING CONSTANT OF THE FAST SITE
      C      U(2) IS SCALAR COUPLING CONSTANT OF THE SLOW SITE
      C      U(3) IS DELTA H FOR FAST SITE
      C      U(4) IS DELTA H FOR SLOW SITE
      C      U(5) IS DELTA J FOR FAST SITE
      C      U(6) IS DELTA J FOR SLOW SITE
      C      AX(1) IS 1/T TIMES 1.0E3
      C      AX(2) IS DELTA OMEGA OR 1/T2P
```

```
      RETURN
3 CONTINUE
      IF (JJ.LE.20) GO TO 100
      * = (CC*((2.699E119)*(U(1)**4)*TE*TE*EXP((-U(3)*AX(1)*0.5032E-3)
      1+0.5032*U(5)+AX(1)*6.038E-1)/AX(1))+((9.380E78)*(U(1)**2)*TE*
      1EXP((-U(3)*AX(1)*1.0064E-3)+1.0064*U(5) + (AX(1)*3.019E-1)/
      2AX(1)**2)+(1.362E00)*(U(1)**2)*AX(1)*EXP((-U(3)*AX(1)*0.5032E-3)
      3+0.5032*U(5)))/((3.394E107)*(U(1)**4)*TE*TE*EXP(AX(1)*6.038E-1)
      4+((2.498E67)*(U(1)**2)*TE*EXP((-U(3)*AX(1)*0.5032E-3)+0.5032*U(5)+
```

```
5((XX(1)*3.019E-1)/XX(1))+((4.343E26)*EXP((-U(3)*XX(1)*1.0064E-3)
6+1.0064*U(5))/XX(1)**2)+((1.8133E48)*(U(1)**2)*(XX(1)**2)))
7+(CC*((1.350E119)*(U(2)**4)*TE*TE*EXP((-U(4)*XX(1)*U.5032E-3)
8+U.5032*U(6)+(XX(1)*6.038E-1))/XX(1))+((4.690E76)*(U(2)**2)*TE*
9EXP((-U(4)*(XX(1)*1.0064E-3)+1.0064*U(6) +(XX(1)*3.019E-1))/
10XX(1)**2)+(6.809E59)*(U(2)**2)*XX(1)*EXP((-U(4)*XX(1)*U.5032E-3)
11+U.5032*U(6)))/((3.594E107)*(U(2)**4)*TE*TE*EXP(XX(1)*6.038E-1)
12C+((2.498E67)*(U(2)**2)*TE*EXP((-U(4)*XX(1)*U.5032E-3)+U.5032*U(6)+
13U(XX(1)*3.019E-1)/XX(1))+((4.343E26)*EXP((-U(4)*XX(1)*1.0064E-3)
14+1.0064*U(6))/XX(1)**2)+((1.8133E48)*(U(2)**2)*(XX(1)**2)))
15 GO TO 101
100 * = (((2.1073E49)*CA*U(1)*EXP((-U(3)*XX(1)*1.0064E-3)+1.0064*U(5))
16/XX(1))/((3.594E107)*(U(1)**4)*(TE**2)*EXP(XX(1)*6.038E-1)
17+((2.498E67)*(U(1)**2)*TE*EXP((-U(3)*XX(1)*U.5032E-3)+U.5032*U(5)
18+((XX(1)*3.019E-1)/XX(1))+((4.343E26)*EXP((-U(3)*XX(1)*1.0064E-3)+
191.0064*U(5))/XX(1)**2))+((1.8133E48)*(U(1)**2)*(XX(1)**2)))
20+(((1.0538E49)*CA*U(2)*EXP((-U(4)*XX(1)*1.0064E-3)+1.0064*U(6))/
216XX(1))/((3.594E107)*(U(2)**4)*(TE**2)*EXP(XX(1)*6.038E-1)
22+((2.498E67)*(U(2)**2)*TE*EXP((-U(4)*XX(1)*U.5032E-3)+U.5032*U(6)
23+((XX(1)*3.019E-1)/XX(1))+((4.343E26)*EXP((-U(4)*XX(1)*1.0064E-3)+
241.0064*U(6))/XX(1)**2))+((1.8133E48)*(U(2)**2)*(XX(1)**2)))
101 RESID = * - XX(2)
    RETURN
    4 CONTINUE
    RETURN
    5 CONTINUE
    RETURN
    6 CONTINUE
    RETURN
    7 CONTINUE
    DO 300 J = 1,NOPT
    READ (5,61) X(1,J), X(2,J), WTX(2,J)
    61 FORMAT (3E12,4)
    300 WTX(1,J) = 0.005
    RETURN
    8 CONTINUE
    RETURN
    END
```

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