

UCLA

UCLA Previously Published Works

Title

Associations of immunity-related single nucleotide polymorphisms with overall survival among prostate cancer patients.

Permalink

<https://escholarship.org/uc/item/6v26g889>

Journal

International Journal of Clinical and Experimental Medicine, 8(7)

ISSN

1940-5901

Authors

Miles, Fayth L
Rao, Jian-Yu
Eckhert, Curtis
[et al.](#)

Publication Date

2015

Peer reviewed

Original Article

Associations of immunity-related single nucleotide polymorphisms with overall survival among prostate cancer patients

Fayth L Miles¹, Jian-Yu Rao², Curtis Eckhert³, Shen-Chih Chang¹, Allan Pantuck⁴, Zuo-Feng Zhang^{1,5}

¹Department of Epidemiology, Fielding School of Public Health, University of California-Los Angeles, Los Angeles, CA, USA; ²Department of Pathology and Laboratory Medicine, David Geffen School of Medicine at UCLA, Los Angeles, CA, USA; ³Department of Environmental Health Sciences, Fielding School of Public Health, University of California-Los Angeles, Los Angeles, CA, USA; ⁴Department of Urology, David Geffen School of Medicine at UCLA, Los Angeles, CA, USA; ⁵Jonsson Comprehensive Cancer Center, University of California-Los Angeles, Los Angeles, CA, USA

Received May 10, 2015; Accepted June 24, 2015; Epub July 15, 2015; Published July 30, 2015

Abstract: The progression of prostate cancer is influenced by systemic inflammation, and may be attributed, in part, to genetic predisposition. Single nucleotide polymorphisms associated with the immune response may help mediate prostate cancer progression. We analyzed data from a hospital-based case-control study of 164 prostate cancer patients and 157 healthy male controls from the Memorial Sloan Kettering Cancer Center. We evaluated associations between six immunity-related polymorphisms (CRP rs1205 and rs1800947, FGFR2 rs1219648 and rs2981582, IFNGR1 rs11914, and IL10 rs1800871) and overall survival among prostate cancer patients, calculating adjusted hazard ratios (HR) and 95% confidence intervals (CI) using Cox proportional hazards regression. FGFR2 rs1219648 (GG vs. AA) and rs2981582 (TT vs. CC) polymorphisms were associated with more favorable overall survival (HR: 0.13, 95% CI: 0.03-0.62 and HR: 0.13, 95% CI: 0.03-0.53, respectively) in patients with primary prostate cancer. These observations highlight the need to validate and identify these and other immunity-related polymorphisms in larger studies examining survival of prostate cancer patients.

Keywords: Genetic predisposition, case-control, proportional hazards model, prostate cancer, immune response, polymorphisms

Introduction

Prostate cancer is the most commonly diagnosed cancer in American men, and is the second leading cause of cancer death in the U.S. Inflammation may play a role in prostate carcinogenesis and progression, although the association is complex and not fully understood [1]. Pro-inflammatory signals are initiated by various environmental stimuli in combination with genetic factors. This is characterized by accumulation of leukocytes and production of a number of immune-related cytokines and enzymatic mediators, which may induce oxidative stress, and promote signaling leading to tumor growth and metastasis [2, 3].

Alterations in genes or proteins related to innate immunity have been observed in pros-

tate cancer patients. Increases in gene expression or cytokine production of various members of the interleukin (IL) family such as IL-4, -6, -8 and -10, and C-reactive protein have been associated with malignant epithelium and metastasis [4, 5]. Alterations in expression of FGFR2, particularly through an isoform switch, may also correlate with prostate cancer progression [6].

Single nucleotide polymorphisms (SNPs) in immune response genes could potentially alter the susceptibility to or progression of cancer through modifications in the cancer-mediated inflammatory response, and related signaling events. Although SNPs in several inflammation-related genes such as cyclooxygenase-2 (COX-2), tumor necrosis factor-alpha (TNF- α) and various interleukins have been shown previously to be associated with increased prostate can-

Immune-response polymorphisms in prostate cancer

Table 1. Demographic and clinical characteristics of prostate cancer cases in the MSKCC study

	All, n ¹	Death, n (%)	Censored, n (%)	P-value ²
Survival	164	47 (29)	117 (71)	
Age at diagnosis				
mean, SD		63.0 ± 7.3	59.4 ± 6.0	
< 65	103	24 (33)	79 (77)	0.004
> 65	51	23 (45)	28 (55)	
Ethnicity				
Caucasian	136	41 (30)	95 (70)	0.45
Other	16	6 (27)	10 (63)	
Smoking				
< 100 cigarettes	56	18 (32)	38 (68)	0.88
> 100 cigarettes	97	29 (30)	68 (70)	
Pack-years				
mean, SD		26.8 ± 31.8	16.5 ± 19.2	
< 20	96	26 (27)	70 (73)	0.13
20-40	25	7 (28)	18 (72)	
> 40	32	14 (44)	18 (56)	
BMI (Kg/m ²)				
mean, SD		26.5 ± 4.3	27.4 ± 3.1	
< 25	33	11 (33)	22 (67)	0.42
> 25	131	36 (27)	95 (73)	
Family history, No. (%)				
Yes	101	32 (32)	69 (68)	0.83
No	51	15 (29)	36 (71)	
Education				
mean, SD		15.8 ± 3.8	14.5 ± 3.6	
< 12	56	16 (29)	40 (71)	0.54
12-16	82	27 (33)	55 (67)	
> 16	13	2 (15)	11 (85)	
Clinical stage				
I	19	15 (79)	4 (21)	0.0001
II	79	10 (13)	69 (87)	
III	41	12 (30)	29 (70)	
IV	13	4 (31)	9 (69)	
Gleason grade				
< 7	61	8 (13)	53 (87)	0.002
7-10	22	10 (45)	12 (55)	
Not graded ³	8	2 (25)	6 (75)	

¹All values may not sum to 164 due to missing data; ²Calculated by performing log-rank test for homogeneity across strata; ³Not graded because of prior hormone therapy.

cer risk [1-7], there have been very few reports of the association of immune-related SNPs with prostate cancer survival.

In light of the potential roles of genetic alterations in the immune-response pathway in chronic inflammation, we examined the association of these pro-inflammatory factors on prostate cancer survival in a hospital-based case-control study.

Materials and methods

Study population

The Memorial Sloan-Kettering Cancer Center (MSKCC) study was a hospital-based study conducted from May 1, 1994 to June 30, 1997. Details of the study design have been described previously [8]. The study was approved by and in accordance with the ethical standards of the Institutional Research Board on Human Subjects of both MSKCC and UCLA.

A total of 164 cases were identified as having newly diagnosed, pathologically confirmed prostate cancer. Eligibility of cases was confirmed by review of medical records and pathology reports. Controls consisted of 157 blood donors recruited from the MSKCC blood bank free of prostate cancer and in stable medical condition, who had resided in the US for at least one year. Data was collected on demographic characteristics, family history of prostate cancer, medical history, extensive dietary history, smoking and alcohol drinking, occupational and environmental exposures, and more. Eighty percent of prostate cancer cases provided blood for genotyping. DNA was extracted from tissues and blood samples using a modified phenol-chloroform method. Patients were followed for overall survival. The social security death index (SSDI) system was employed as a follow-up method for patient survival status and related dates.

The SSDI is generated from the public Death Master File of the U.S. Social Security Administration and provides death records of qualified social security recipients. These records were last retrieved on May 12, 2013. Follow-up time was calculated as the date of diagnosis to death or May 12, 2013. Patients who were not shown in the SSDI were considered alive (right-censored) on May 12, 2013. Among 164 prostate cancer patients with avail-

Immune-response polymorphisms in prostate cancer

Table 2. Cox proportional hazard model results for the association between immunity-related SNPs and survival among prostate cancer patients¹

<i>CRP</i> rs1205			<i>CRP</i> rs1800947			<i>FGFR2</i> rs1219648		
Genotype	Dead/All	HR (95% CI)	Genotype	Dead/All	HR (95% CI)	Genotype	Dead/All	HR (95% CI)
CC	12/48	1	GG	29/104	1	AA	15/44	1
CT	15/57	1.69 (0.70-4.07)	GC	1/13	0.32 (0.04-2.46)	GA	11/56	0.52 (0.23-1.15)
TT	1/12	0.49 (0.06-3.96)	CC	0/2	N/A	GG	3/18	0.13 (0.03-0.62)
Additive		0.98 (0.51-1.87)	Additive		0.30 (0.04-2.16)	Additive		0.43 (0.23-0.78)
Recessive		0.36 (0.05-2.70)	Recessive		N/A	Recessive		0.20 (0.05-0.86)
Dominant		1.45 (0.61-3.47)	Dominant		0.29 (0.04-2.22)	Dominant		0.40 (0.18-0.89)
<i>FGFR2</i> rs2981582			<i>IFNGR1</i> rs11914			<i>IL10</i> rs1800871		
Genotype	Dead/All	HR (95% CI)	Genotype	Dead/All	HR (95% CI)	Genotype	Dead/All	HR (95% CI)
CC	15/42	1	TT	19/83	1	CC	15/66	1
CT	10/55	0.50 (0.22-1.14)	TG	10/34	1.42 (0.59-3.40)	CT	11/42	1.35 (0.58-3.16)
TT	4/21	0.13 (0.03-0.53)	GG	1/2	10.30 (1.13-93.45)	TT	4/10	2.51 (0.64-9.85)
Additive		0.41 (0.23-0.73)	Additive		1.77 (0.81-3.85)	Additive		1.49 (0.81-2.76)
Recessive		0.18 (0.05-0.71)	Recessive		9.48 (1.06-84.72)	Recessive		2.19 (0.59-8.12)
Dominant		0.36 (0.16-0.80)	Dominant		1.59 (0.69-3.69)	Dominant		1.50 (0.67-3.35)

¹adjusted for race, age, pack-years of smoking, family history, body mass index, and stage.

Immune-response polymorphisms in prostate cancer

able follow-up data, 47 (29%) passed away before the date of May 12, 2013. The median follow-up time was 18.8 years.

Laboratory analysis

In this study we selected immunity-related SNPs that had a minor allele frequency > 5% in Caucasian populations in the National Center for Biotechnology Information SNP database, were functional or potentially functional SNPs located in the coding, 3', and 5'-untranslated regions, and near gene regions. SNPs were genotyped using the ABI (Applied Biosystems, Foster City, CA) TaqMan assay. Briefly, PCR was performed using fluorescently labeled sequence-specific probes. The denaturation process was performed at 92°C for 10 minutes followed by an annealing and extension phase of 60 cycles at 92°C for 15 seconds, and 62°C for 80 seconds. The genotyping call rate was \geq 95% and reproducibility (using a 5% random sample) was > 99%. SNPs that violated Hardy Weinberg equilibrium were excluded. The final group of SNPs used in this study included C-reactive protein (CRP) rs1205 and rs1800947, fibroblast growth factor receptor-2 (FGFR2) rs1219648 and rs2981582, interleukin-10 (IL-10) rs1800871, and interferon gamma receptor-1 (IFNGR1) rs11914.

Statistical analyses

All statistical analyses were performed using Statistical Analysis Software (SAS) version 9.3.

Descriptive statistics were performed for characteristics of interest, using the log-rank test for homogeneity over strata to calculate *P*-values for categorical variables. In survival analysis, survival time was defined as the difference in years between diagnosis and last follow-up (May 12, 2013) or death, whichever came first. Four models were used in SNP assessment, including genotype-specific, log-additive, dominant, and recessive genetic models. In the dominant model, the hazard associated with the variant allele, including homozygous recessive and heterozygous genotypes was compared to the homozygous dominant/ancestral genotype. In the recessive model, the hazard associated with presence of two variant alleles (homozygous recessive genotype) was compared to the homozygous dominant and heterozygous genotypes. The Cox proportional hazards model was used to generate adjusted ha-

zards ratios (HRs) and 95% confidence intervals (CIs) to determine the effect of genotype on overall survival. In the regression analysis, covariates included race (white versus non-white), pack-years of smoking (continuous), age (continuous), body mass index (BMI-continuous), family history (categorical) and stage (categorical). BMI was included because of a reported association with prostate cancer risk and mortality [9]. Missing values for BMI or smoking pack-years (< 11%) were imputed where possible using the median value among cases. Stage was defined using TNM staging criteria: stage 1-locally confined, clinically undetectable; stage 2-locally confined, palpable; stage 3-locally advanced; stage 4-advanced, metastatic prostate cancer.

Results

Demographic and clinical characteristics of prostate cancer cases from the MSKCC study population are presented in **Table 1**. Overall survival correlated with age, as significantly shorter survival was observed among cases over the age of 65 ($P = 0.004$). A notable difference in survival was noted according to pathological stage ($P = 0.0001$) and Gleason grade ($P = 0.002$). However, a considerably large number of patients over the age of 65 were diagnosed with stage I prostate cancer (not shown).

Cox proportional hazards estimates for the associations of immunity-related SNPs with overall survival were calculated for individuals diagnosed with prostate cancer (**Table 2**). The homozygous GG genotype of FGFR2 rs1219648 was associated with more favorable survival (HR: 0.13, 95% CI: 0.03-0.62), as compared to the AA genotype. A linear trend associated with the G allele in hazards ratios was observed (allelic HR: 0.43, 95% CI: 0.23-0.78). The homozygous FGFR2 rs2981582 TT genotype also correlated with better overall survival (HR: 0.13, 95% CI: 0.03-0.53), and a linear trend was again associated with the variant allele (HR: 0.41, 95% CI: 0.23-0.73). No other SNPs examined were associated with significant differences in survival among prostate cancer cases.

Discussion

Systemic inflammation promotes castration-resistance and metastasis [10]. Therefore, the role of inflammation-related SNPs in prostate

Immune-response polymorphisms in prostate cancer

cancer survival was analyzed. In the current study, FGFR2 rs2981582 and rs1219648, two SNPs in relatively high linkage disequilibrium, were associated with more favorable overall survival among prostate cancer cases.

FGFR2 is a receptor tyrosine kinase involved in regulation of cell growth, blood vessel formation, embryogenesis, and wound healing. Altered gene expression has potential consequences in tumor cell proliferation, migration, and angiogenesis. Alterations in FGFR2 mRNA have been reported in prostate cancer patients [11], although significant associations of rs2981582 and rs1219648 SNPs with prostate cancer remain to be identified. Interestingly, FGFR2 rs2981582 and rs1219648 variants are associated with increased breast cancer risk [12-14]. rs2981582, particularly, is associated with estrogen receptor positive, low-grade tumors [15, 16], consistent with what is observed in BRCA2 mutation carriers. Although not shown to alter alternative splicing of FGFR2 isoforms [17], it is possible that FGFR2 variants rs2981582 and rs1219648 alter hormone signaling to promote conditions that are less favorable for prostate cancer metastasis. To the best of our knowledge, this is the first report of the associations of rs2981582 and rs1219648 with prostate cancer survival. However, limitations due to sample size cannot be ignored, and additional studies examining these associations are warranted.

A better understanding of prostate cancer progression will require the discovery and analysis of pro-inflammatory biomarkers. Studies of sufficient sample size may allow for the utilization of SNPs and other potential environmental pro-inflammatory agents as prognostic indicators, and may prove useful in the prevention and control of aggressive prostate cancer.

Acknowledgements

This work was supported by National Institutes of Health (ES06718, ES01167, and CA09142), Seymour Family Gift for Innovative Investigator-Initiated Research in Bladder Cancer, Alper Research Center for Environmental Genomics of the University of California, and the University of California, Los Angeles Jonsson Comprehensive Cancer Center. We thank Drs. Victor Reuter and Howard Scher of Memorial Sloan-Kettering Cancer Center and Carlos Cordon-

Cardo of Mount Sinai School of Medicine for their contributions in the initial data collection.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Zuo-Feng Zhang, Department of Epidemiology, University of California-Los Angeles Fielding School of Public Health, 650 Charles E. Young Dr. South, Los Angeles, CA 90095-1772, U.S.A. Tel: 3108258418; Fax: 3102066039; E-mail: zfzhang@ucla.edu

References

- [1] Fernandez P, de Beer PM, van der Merwe L and Heyns CF. COX-2 promoter polymorphisms and the association with prostate cancer risk in South African men. *Carcinogenesis* 2008; 29: 2347-2350.
- [2] Kwon EM, Salinas CA, Kolb S, Fu R, Feng Z, Stanford JL and Ostrander EA. Genetic polymorphisms in inflammation pathway genes and prostate cancer risk. *Cancer Epidemiol Biomarkers Prev* 2011; 20: 923-933.
- [3] Panguluri RC, Long LO, Chen W, Wang S, Coulibaly A, Ukoli F, Jackson A, Weinrich S, Ahaghotu C, Isaacs W and Kittles RA. COX-2 gene promoter haplotypes and prostate cancer risk. *Carcinogenesis* 2004; 25: 961-966.
- [4] Saenz-Lopez P, Carretero R, Cozar JM, Romero JM, Canton J, Vilchez JR, Tallada M, Garrido F and Ruiz-Cabello F. Genetic polymorphisms of RANTES, IL1-A, MCP-1 and TNF-A genes in patients with prostate cancer. *BMC Cancer* 2008; 8: 382.
- [5] Tindall EA, Severi G, Hoang HN, Ma CS, Fernandez P, Southey MC, English DR, Hopper JL, Heyns CF, Tangye SG, Giles GG and Hayes VM. Comprehensive analysis of the cytokine-rich chromosome 5q31.1 region suggests a role for IL-4 gene variants in prostate cancer risk. *Carcinogenesis* 2010; 31: 1748-1754.
- [6] Tindall EA, Severi G, Hoang HN, Southey MC, English DR, Hopper JL, Giles GG and Hayes VM. Interleukin-6 promoter variants, prostate cancer risk, and survival. *Prostate* 2012; 72: 1701-1707.
- [7] Zabaleta J, Su LJ, Lin HY, Sierra RA, Hall MC, Sartor AO, Clark PE, Hu JJ and Ochoa AC. Cytokine genetic polymorphisms and prostate cancer aggressiveness. *Carcinogenesis* 2009; 30: 1358-1362.
- [8] Cao W, Cai L, Rao JY, Pantuck A, Lu ML, Dalbagni G, Reuter V, Scher H, Cordon-Cardo C, Figlin RA, Belldegrun A and Zhang ZF. Tobacco smoking, GSTP1 polymorphism, and bladder carcinoma. *Cancer* 2005; 104: 2400-2408.

Immune-response polymorphisms in prostate cancer

- [9] Cao Y and Ma J. Body mass index, prostate cancer-specific mortality, and biochemical recurrence: a systematic review and meta-analysis. *Cancer Prev Res (Phila)* 2011; 4: 486-501.
- [10] Ammirante M, Luo JL, Grivennikov S, Nedospasov S and Karin M. B-cell-derived lymphotoxin promotes castration-resistant prostate cancer. *Nature* 2010; 464: 302-305.
- [11] Naimi B, Latil A, Fournier G, Mangin P, Cussenot O and Berthon P. Down-regulation of (IIIb) and (IIIc) isoforms of fibroblast growth factor receptor 2 (FGFR2) is associated with malignant progression in human prostate. *Prostate* 2002; 52: 245-252.
- [12] Easton DF, Pooley KA, Dunning AM, Pharoah PD, Thompson D, Ballinger DG, Struewing JP, Morrison J, Field H, Luben R, Wareham N, Ahmed S, Healey CS, Bowman R, Meyer KB, Haiman CA, Kolonel LK, Henderson BE, Le Marchand L, Brennan P, Sangrajrang S, Gaborieau V, Odehrey F, Shen CY, Wu PE, Wang HC, Eccles D, Evans DG, Peto J, Fletcher O, Johnson N, Seal S, Stratton MR, Rahman N, Chenevix-Trench G, Bojesen SE, Nordestgaard BG, Axelsson CK, Garcia-Closas M, Brinton L, Chanock S, Lissowska J, Peplonska B, Nevanlinna H, Fagerholm R, Eerola H, Kang D, Yoo KY, Noh DY, Ahn SH, Hunter DJ, Hankinson SE, Cox DG, Hall P, Wedren S, Liu J, Low YL, Bogdanova N, Schurmann P, Dork T, Tollenaar RA, Jacobi CE, Devilee P, Klijn JG, Sigurdson AJ, Doody MM, Alexander BH, Zhang J, Cox A, Brock IW, MacPherson G, Reed MW, Couch FJ, Goode EL, Olson JE, Meijers-Heijboer H, van den Ouweland A, Uitterlinden A, Rivadeneira F, Milne RL, Ribas G, Gonzalez-Neira A, Benitez J, Hopper JL, McCredie M, Southey M, Giles GG, Schroen C, Justenhoven C, Brauch H, Hamann U, Ko YD, Spurdle AB, Beesley J, Chen X, Mannermaa A, Kosma VM, Kataja V, Hartikainen J, Day NE, Cox DR and Ponder BA. Genome-wide association study identifies novel breast cancer susceptibility loci. *Nature* 2007; 447: 1087-1093.
- [13] Hunter DJ, Kraft P, Jacobs KB, Cox DG, Yeager M, Hankinson SE, Wacholder S, Wang Z, Welch R, Hutchinson A, Wang J, Yu K, Chatterjee N, Orr N, Willett WC, Colditz GA, Ziegler RG, Berg CD, Buys SS, McCarty CA, Feigelson HS, Calle EE, Thun MJ, Hayes RB, Tucker M, Gerhard DS, Fraumeni JF Jr, Hoover RN, Thomas G and Chanock SJ. A genome-wide association study identifies alleles in FGFR2 associated with risk of sporadic postmenopausal breast cancer. *Nat Genet* 2007; 39: 870-874.
- [14] Ledwon JK, Hennig EE, Maryan N, Goryca K, Nowakowska D, Niwinska A and Ostrowski J. Common low-penetrance risk variants associated with breast cancer in Polish women. *BMC Cancer* 2013; 13: 510.
- [15] Antoniou AC, Beesley J, McGuffog L, Sinilnikova OM, Healey S, Neuhausen SL, Ding YC, Rebbeck TR, Weitzel JN, Lynch HT, Isaacs C, Ganz PA, Tomlinson G, Olopade OI, Couch FJ, Wang X, Lindor NM, Pankratz VS, Radice P, Manoukian S, Peissel B, Zaffaroni D, Barile M, Viel A, Allavena A, Dall'Olio V, Peterlongo P, Szabo CI, Zikan M, Claes K, Poppe B, Foretova L, Mai PL, Greene MH, Rennert G, Lejbkowitz F, Glendon G, Ozcelik H, Andrulis IL, Thomassen M, Gerdes AM, Sunde L, Cruger D, Birk Jensen U, Caligo M, Friedman E, Kaufman B, Laitman Y, Milgrom R, Dubrovsky M, Cohen S, Borg A, Jernstrom H, Lindblom A, Rantala J, Stenmark-Askmalin M, Melin B, Nathanson K, Domchek S, Jakubowska A, Lubinski J, Huzarski T, Osorio A, Lasa A, Duran M, Tejada MI, Godino J, Benitez J, Hamann U, Kriege M, Hoogerbrugge N, van der Luijt RB, van Asperen CJ, Devilee P, Meijers-Heijboer EJ, Blok MJ, Aalfs CM, Hogervorst F, Rookus M, Cook M, Oliver C, Frost D, Conroy D, Evans DG, Lalloo F, Pichert G, Davidson R, Cole T, Cook J, Paterson J, Hodgson S, Morrison PJ, Porteous ME, Walker L, Kennedy MJ, Dorkins H, Peock S, Godwin AK, Stoppa-Lyonnet D, de Pauw A, Mazoyer S, Bonadona V, Lasset C, Dreyfus H, Leroux D, Hardouin A, Berthet P, Faivre L, Loustalot C, Noguchi T, Sobol H, Rouleau E, Nogues C, Frenay M, Venat-Bouvet L, Hopper JL, Daly MB, Terry MB, John EM, Buys SS, Yassin Y, Miron A, Goldgar D, Singer CF, Dressler AC, Gschwantler-Kaulich D, Pfeiler G, Hansen TV, Jonson L, Agnarsson BA, Kirchoff T, Offit K, Devlin V, Dutra-Clarke A, Piedmonte M, Rodriguez GC, Wakeley K, Boggess JF, Basil J, Schwartz PE, Blank SV, Toland AE, Montagna M, Casella C, Imyanitov E, Tihomirova L, Blanco I, Lazaro C, Ramus SJ, Sucheston L, Karlan BY, Gross J, Schmutzler R, Wappenschmidt B, Engel C, Meindl A, Lochmann M, Arnold N, Heidemann S, Varon-Mateeva R, Niederacher D, Sutter C, Deissler H, Gadzicki D, Preisler-Adams S, Kast K, Schonbuchner I, Caldes T, de la Hoya M, Aittomaki K, Nevanlinna H, Simard J, Spurdle AB, Holland H, Chen X, Platte R, Chenevix-Trench G and Easton DF. Common breast cancer susceptibility alleles and the risk of breast cancer for BRCA1 and BRCA2 mutation carriers: implications for risk prediction. *Cancer Res* 2010; 70: 9742-9754.
- [16] Broeks A, Schmidt MK, Sherman ME, Couch FJ, Hopper JL, Dite GS, Apicella C, Smith LD, Hammet F, Southey MC, Van 't Veer LJ, de Groot R, Smit VT, Fasching PA, Beckmann MW, Jud S, Ekici AB, Hartmann A, Hein A, Schulz-Wendland R, Burwinkel B, Marme F, Schneeweiss A, Sinn HP, Sohn C, Tchatchou S, Bojesen SE, Nordestgaard BG, Flyger H, Orsted DD, Kaur-Knudsen D, Milne RL, Perez JI, Zamora P, Rodriguez PM, Benitez J, Brauch H, Justenhoven

Immune-response polymorphisms in prostate cancer

C, Ko YD, Hamann U, Fischer HP, Bruning T, Pesch B, Chang-Claude J, Wang-Gohrke S, Bremer M, Karstens JH, Hillemanns P, Dork T, Nevanlinna HA, Heikkinen T, Heikkila P, Blomqvist C, Aittomaki K, Aaltonen K, Lindblom A, Margolin S, Mannermaa A, Kosma VM, Kauppinen JM, Kataja V, Auvinen P, Eskelinen M, Soini Y, Chenevix-Trench G, Spurdle AB, Beesley J, Chen X, Holland H, Lambrechts D, Claes B, Vandorpe T, Neven P, Wildiers H, Flesch-Janys D, Hein R, Loning T, Kosel M, Fredericksen ZS, Wang X, Giles GG, Baglietto L, Severi G, McLean C, Haiman CA, Henderson BE, Le Marchand L, Kolonel LN, Alnaes GG, Kristensen V, Borresen-Dale AL, Hunter DJ, Hankinson SE, Andrulis IL, Mulligan AM, O'Malley FP, Devilee P, Huijts PE, Tollenaar RA, Van Asperen CJ, Seynaeve CS, Chanock SJ, Lissowska J, Brinton L, Peplonska B, Figueroa J, Yang XR, Hooning MJ, Hollestelle A, Oldenburg RA, Jager A, Kriege M, Ozturk B, van Leenders GJ, Hall P, Czene K, Humphreys K, Liu J, Cox A, Connley D, Cramp HE, Cross SS, Balasubramanian SP, Reed MW, Dunning AM, Easton DF, Humphreys MK, Caldas C, Blows F, Driver K, Provenzano E, Lubinski J, Jakubowska A, Huzarski T, Byrski T, Cybulski C, Gorski B, Gronwald J, Brennan P, Sangrajrang S, Gaborieau V, Shen CY, Hsiung CN, Yu JC, Chen ST, Hsu GC, Hou MF, Huang CS, Anton-Culver H, Ziogas A, Pharoah PD and Garcia-Closas M. Low penetrance breast cancer susceptibility loci are associated with specific breast tumor subtypes: findings from the Breast Cancer Association Consortium. *Hum Mol Genet* 2011; 20: 3289-3303.

[17] Huijts PE, van Dongen M, de Goeij MC, van Moolenbroek AJ, Blanken F, Vreeswijk MP, de Kruijf EM, Mesker WE, van Zwet EW, Tollenaar RA, Smit VT, van Asperen CJ and Devilee P. Allele-specific regulation of FGFR2 expression is cell type-dependent and may increase breast cancer risk through a paracrine stimulus involving FGF10. *Breast Cancer Res* 2011; 13: R72.