

# UC Davis

## UC Davis Previously Published Works

### Title

The Influence of Spin-Labeled Fluorene Compounds on the Assembly and Toxicity of the A $\beta$  Peptide

### Permalink

<https://escholarship.org/uc/item/6w79g1sh>

### Journal

PLOS ONE, 7(4)

### ISSN

1932-6203

### Authors

Petrlova, Jitka

Kálai, Tamás

Maezawa, Izumi

et al.

### Publication Date

2012

### DOI

10.1371/journal.pone.0035443

Peer reviewed

# The Influence of Spin-Labeled Fluorene Compounds on the Assembly and Toxicity of the A $\beta$ Peptide

Jitka Petrlova<sup>1</sup>, Tamás Kálai<sup>2</sup>, Izumi Maezawa<sup>3</sup>, Robin Altman<sup>1</sup>, Ghimire Harishchandra<sup>4</sup>, Hyun-Seok Hong<sup>3</sup>, Daniel A. Bricarello<sup>5</sup>, Atul N. Parikh<sup>5</sup>, Gary A. Lorigan<sup>4</sup>, Lee-Way Jin<sup>3</sup>, Kálmán Hideg<sup>2</sup>, John C. Voss<sup>1\*</sup>

**1** Department of Biochemistry and Molecular Medicine, University of California Davis, Davis, California, United States of America, **2** Institute of Organic and Medicinal Chemistry, University of Pécs, Pécs, Hungary, **3** Laboratory Medicine, Department of Pathology, M.I.N.D. Institute, Miami University, Oxford, Ohio, United States of America, **4** Department of Chemistry and Biochemistry, Miami University, Oxford, Ohio, United States of America, **5** Department of Applied Science, University of California Davis, Davis, California, United States of America

## Abstract

**Background:** The deposition and oligomerization of amyloid  $\beta$  (A $\beta$ ) peptide plays a key role in the pathogenesis of Alzheimer's disease (AD). A $\beta$  peptide arises from cleavage of the membrane-associated domain of the amyloid precursor protein (APP) by  $\beta$  and  $\gamma$  secretases. Several lines of evidence point to the soluble A $\beta$  oligomer (A $\beta$ O) as the primary neurotoxic species in the etiology of AD. Recently, we have demonstrated that a class of fluorene molecules specifically disrupts the A $\beta$ O species.

**Methodology/Principal Findings:** To achieve a better understanding of the mechanism of action of this disruptive ability, we extend the application of electron paramagnetic resonance (EPR) spectroscopy of site-directed spin labels in the A $\beta$  peptide to investigate the binding and influence of fluorene compounds on A $\beta$ O structure and dynamics. In addition, we have synthesized a spin-labeled fluorene (SLF) containing a pyrroline nitroxide group that provides both increased cell protection against A $\beta$ O toxicity and a route to directly observe the binding of the fluorene to the A $\beta$ O assembly. We also evaluate the ability of fluorenes to target multiple pathological processes involved in the neurodegenerative cascade, such as their ability to block A $\beta$ O toxicity, scavenge free radicals and diminish the formation of intracellular A $\beta$ O species.

**Conclusions:** Fluorene modified with pyrroline nitroxide may be especially useful in counteracting A $\beta$  peptide toxicity, because they possess both antioxidant properties and the ability to disrupt A $\beta$ O species.

**Citation:** Petrlova J, Kálai T, Maezawa I, Altman R, Harishchandra G, et al. (2012) The Influence of Spin-Labeled Fluorene Compounds on the Assembly and Toxicity of the A $\beta$  Peptide. PLoS ONE 7(4): e35443. doi:10.1371/journal.pone.0035443

**Editor:** Koichi M. Iijima, Thomas Jefferson University, United States of America

**Received:** November 11, 2011; **Accepted:** March 16, 2012; **Published:** April 30, 2012

**Copyright:** © 2012 Petrlova et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This work was funded by grants from the U.S. National Institutes of Health (R01 AG029246) and the Hungarian National Research Fund OTKA T81123. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: jcvoss@ucdavis.edu

## Introduction

Alzheimer's disease (AD) is characterized by the deposition of various amyloid  $\beta$  (A $\beta$ ) aggregates forming amyloid in the brain. Evidence from a variety of studies has established that the oligomeric species of A $\beta$  (A $\beta$ O) carries the greatest toxicity, triggering a variety of downstream effects resulting in neurotoxicity and cognitive deficits [1,2,3,4]. A major impediment to the development of effective anti-A $\beta$  compounds for AD therapy is that essentially 100% of large-molecule drugs and >98% of small-molecule drugs fail to cross the blood-brain barrier (BBB) [5]. Recently [6], we explored a series of compounds based on a highly rigid tricyclic fluorene ring that were developed as amyloid imaging agents [7]. These compounds contain a tertiary amine electron-donating group attached to one aromatic ring and display excellent pharmacokinetics properties and brain bioavailability. In that work, we reported on the ability of two fluorene compounds to disrupt A $\beta$ O assemblies and reduce A $\beta$  toxicity [6]. These compounds (K01-162 and K01-186) were identified based on their

ability to block cell death secondary to intracellular A $\beta$ O production. Both fluorene compounds bind and destabilize A $\beta$ O, and are capable of penetrating the brain and reducing the cerebral amyloid burden in APP transgenic mice. Fluorenes therefore have a potential use in AD therapy by targeting A $\beta$ O toxicity at both intraneuronal and extracellular sites [6,8].

In AD, accumulating evidence points to oxidative stress as the ultimate downstream component of A $\beta$ -induced toxicity [9,10]. For example, A $\beta$  increases NMDA receptor activation, and one of the newer drugs for the treatment of AD (Memantine) targets NMDA receptors in order to block glutamate excitotoxicity. Among other pathways, over-stimulation of NMDA receptors activates phospholipase A, leading to elevated arachidonic acid levels, which in turn generates oxygen free radicals and further activation of phospholipases [11]. Thus the excitotoxicity involves a feedback loop that ultimately leads to neuronal self-digestion via increased Ca<sup>2+</sup> levels, protein breakdown, free radical formation and lipid peroxidation [10]. As shown previously [6], the anti-amyloid fluorenes have antioxidant properties. Furthermore,

because nitroxides such as the pyrroline species can cycle within a redox cascade via a relatively stable non-damaging N-oxyl (nitroxyl) radical intermediate [12,13], compounds carrying this moiety are likely to have the added potential for decreasing oxidative stress and attenuating the damage caused by reactive oxygen species.

In this study, we apply electron paramagnetic resonance (EPR) spectroscopy to a novel fluorene compound containing a pyrroline nitroxide. This spin-labeled fluorene (SLF) exerts similar potency in A $\beta$ O disruption and protection against A $\beta$ O-induced toxicity, while also having superior free radical scavenging compared to the model fluorene compounds. Furthermore, the nitroxide moiety provides an intrinsic reporter group that can be probed by EPR spectroscopy, which may provide a sensitive diagnostic tool for *in vivo* detection of A $\beta$  plaques in patients with AD [14]. Thus, in addition to its potential as a novel bifunctional candidate to address A $\beta$ O toxicity, the SLF compound may also help as a diagnostic and research tool in elucidating fluorene mechanism of action.

## Results and Discussion

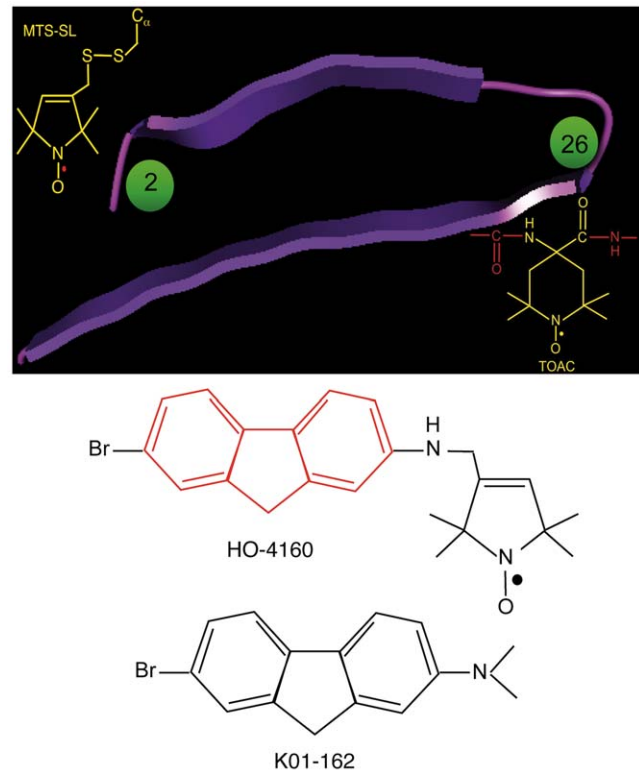
### Bifunctional structure of spin-labeled fluorenes

Previously [6], we found the substituted fluorene 7-bromo-2-N,N-dimethylaminofluorene (K01-162) is able to reduce the amyloid burden in mice and block A $\beta$  toxicity in cultured neurons. In addition, K01-162 binds and disaggregates A $\beta$  in its toxic, soluble oligomeric state (A $\beta$ O). In order to extend the experimental, diagnostic and therapeutic potential of the fluorene agents, the methyl group of dimethylamino was replaced with a pyrroline or piperidine ring to incorporate a spin probe on the fluorene ligand [15]. The structure of spin-labeled fluorene (SLF) HO-4160 (7-bromo-N-methyl-N-[(2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-yl)methyl]-9H-fluoren-2-amine radical) is shown in Figure 1.

### SLFs protect against A $\beta$ O toxicity in cultured neurons

As demonstrated earlier [6,8] K01-162 attenuates intracellular A $\beta$ O accumulation and protects against A $\beta$ O-induced toxicity in cultured neurons. To test the efficacy of SLFs to block A $\beta$  toxicity in cultured neurons, we examined the potential of a recently described class of SLF molecules to influence the survivability of MC65 neuroblastoma cells [16] containing conditional expression of the C-terminal region (C99) of the amyloid precursor protein (APP). In the MC65 model system, expression of the APP fragment is turned on in the absence of the transgene suppressor, tetracycline (TC). Upon APP-C99 induction, A $\beta$  is generated after proteolysis by cellular  $\gamma$ -secretase [16]. As shown in Figure 2, the viability of cells is severely diminished when the expression of APP-C99 is turned on (-TC), compared to the control cells (+TC) lacking A $\beta$  generation. However, increasing levels of SLF HO-4160 restore cell viability to near control levels, with an EC<sub>50</sub> of 30 nM. This response is superior to the K01-162 model fluorene (Figure 2, inset). At concentrations above 1  $\mu$ M, the viability of both APP-induced and un-induced cells declines, reflecting a tolerance limit of the cells to the SLF.

As mentioned above, HO-4160 is one of a group of related SLF compounds that we have recently described [15]. We also measured the ability of the other related SLF analogs (see Fig. S1 in Supporting Information) to block A $\beta$  toxicity. As shown in Table 1, except for compound 5 in this series, each of the compounds offers protection with potency comparable or superior to K01-162. Because HO-4160 provides the greatest amount of

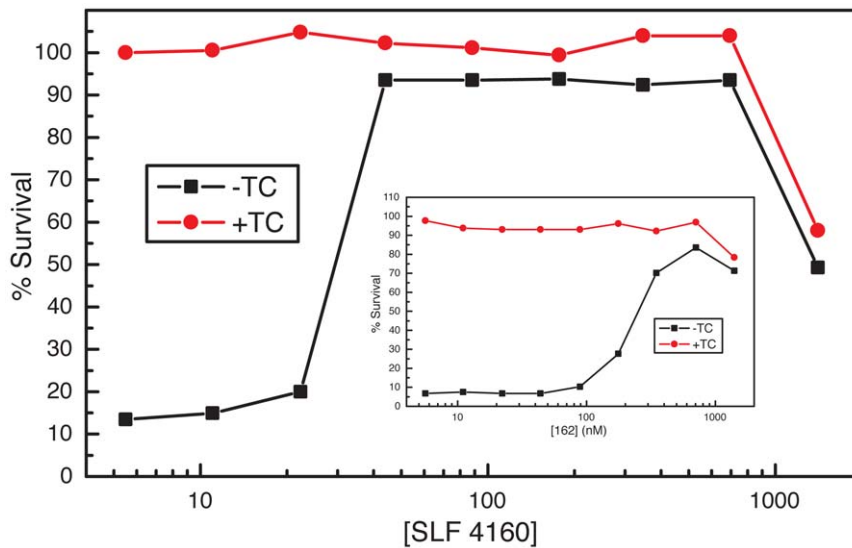


**Figure 1. Diagram showing the locations of spin labels in A $\beta$ <sub>(1-40)</sub> and the structures of the HO-4160 and K01-162 fluorene compounds.** Top panel: the spin labels were targeted to either position 2 (MTS nitroxide spin label) or position 26 (TOAC nitroxide spin label) of A $\beta$ . Position 26 lies within a putative hairpin loop connecting the terminal domains of the peptide, while position 2 is found within the N-terminal domain. Bottom panel: the structure of the SLF compound HO-4160, which is a derivative of the active fluorene K01-162 described in [6]. The core fluorene ring structure is shown in red. doi:10.1371/journal.pone.0035443.g001

protection against A $\beta$  toxicity, we therefore selected this SLF for more detailed analysis of its molecular effects on A $\beta$ O.

### SLF HO-4160 reduces A $\beta$ O accumulation in cultured neurons

Our results clearly demonstrate the protective effect of SLF HO-4160 on MC65 cells expressing APP-C99. To determine whether this protective effect corresponds to a reduction in the A $\beta$ O burden of the cells, we used both Western blot and immunofluorescent staining to analyze the A $\beta$ O levels in SLF-treated cells. Western blot analysis of MC65 cellular extracts reveals decreased levels of A $\beta$ O assemblies of varying sizes in SLF-treated cells expressing APP-C99 (-TC) compared to control cells (+TC). As shown in Figure 3, the A $\beta$  fragments produced by  $\gamma$ -secretase action fail to form any readily apparent oligomers when the cells are treated with 300 nM HO-4160. Complementary results were achieved by using immunofluorescence to probe for A $\beta$ O using the oligomer-specific antibody A11 [17]. Figure 4 shows greatly decreased levels of A $\beta$ O assemblies in MC65 cells expressing APP-C99 and treated with SLF (Fig. 4D), compared to untreated cells (Fig. 4B). These results indicate that not only does SLF HO-4160 block the toxic effects associated with A $\beta$  production in cultured neuronal cells, but also directly attenuates the formation of oligomeric A $\beta$  species in the same model.



**Figure 2. SLF HO-4160 protects against A $\beta$ O toxicity in cultured neurons.** Plotted are the viability values for neuroblastoma MC65 cells with conditional APP-C99 expression as a function of compound concentration. The protective effect against A $\beta$ O toxicity exceeds that of the previously described base compound, K01-162 (insert). doi:10.1371/journal.pone.0035443.g002

### Disruption of A $\beta$ O aggregates by SLF

To test whether SLF HO-4160 blocks the formation of A $\beta$ O assemblies that can be imaged by atomic force microscopy (AFM) [6], we acquired AFM images of A $\beta$  preparations (50  $\mu$ M peptide) incubated in PBS buffer for 1 hour, with and without 50  $\mu$ M SLF treatment. As shown in Figure 5 (A&B), there is a lack of particles >5 nm when A $\beta$  is incubated with the SLF. We also tested whether SLF decreases the formation of particles rich in beta-strand secondary structure using the amyloidophilic dye thioflavin T (ThT). When SLF is included in a 24-hour incubation of A $\beta$ , the ThT fluorescence is decreased by nearly one-half (Fig. 5C). Both the AFM and ThT observations are consistent with a mechanism where the protective activity of HO-4160, like fluorene K01-162, is related to its ability to bind and disaggregate A $\beta$ .

### EPR spectroscopy detects the binding of SLF to A $\beta$

Due to its small size, the rotational diffusion of the SLF molecule when free in solution will average the hyperfine anisotropy of its EPR spectrum, producing the narrow, time-

averaged line shape seen in Figure 6 (black line) indicative of sub-nanosecond diffusion. However, in the presence of A $\beta$ O, substantial broadening of the SLF (10  $\mu$ M) signal is apparent (Fig. 6, red line). The most likely cause of the spectral broadening is a reduction in the rate of rotational diffusion by the SLF molecule in solution upon binding to the A $\beta$  peptide. While a small broad peak appears to arise uniquely in the sample containing A $\beta$ O (see arrow, Fig. 6), we are unable to resolve this feature sufficiently above background. Therefore it is difficult to conclude with certainty whether the resulting spectrum of the SLF in the presence of A $\beta$  represents the compound docked to a mixture of monomers and higher oligomers, or a mixture of bound and unbound SLF. Given the high affinity of fluorenes for A $\beta$ O [6], we expect that the SLF compound is entirely occupied by the excess A $\beta$  peptide.

### Oligomerization of A $\beta$ as reported by peptide containing the TOAC spin label at position 26

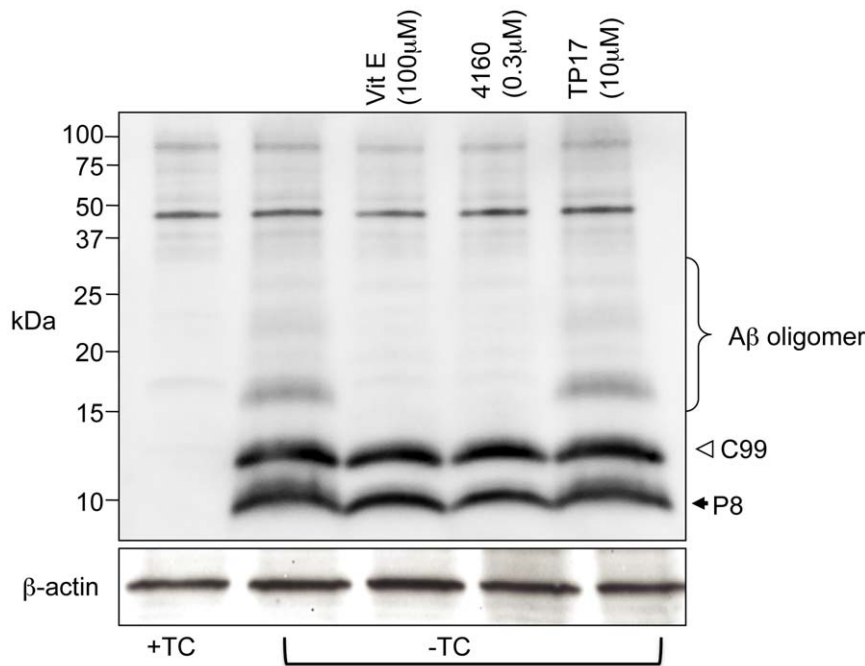
We recently [18] reported on A $\beta$  synthesized with a TOAC spin label positioned in the central region of the peptide, at position 26 (A $\beta$ <sup>(26TOAC)</sup>). TOAC is a backbone-restricted nitroxide that offers improved detection of the dynamics arising from movement of a peptide's backbone and/or global rotational diffusion. We have previously shown that the local backbone at position 26 is sufficiently ordered such that the EPR spectrum of A $\beta$ <sup>(26TOAC)</sup> is sensitive to changes in the peptide's rate of global rotational diffusion [18]. Since oligomerization of A $\beta$ <sup>(26TOAC)</sup> will have profound effects on the rate of global rotational diffusion, this modified peptide provides direct insights into the oligomeric state of A $\beta$  in solution. As shown in Figure 7A, there is a time-dependent broadening of the A $\beta$ <sup>(26TOAC)</sup> EPR line shape, consistent with an increasing molecular volume resulting from oligomerization. Because of the close proximity of peptides in A $\beta$ O, the samples in Figure 7A contained a 25% molar fraction of A $\beta$ <sup>(26TOAC)</sup> that was spin-diluted with wild-type A $\beta$ . Thus the increased line-broadening in these samples can be attributed to changes in spin-label correlation time, and not spin coupling. However, the strong influence of dipolar coupling in the oligomer

**Table 1. Potency of SLF compounds [15] against A $\beta$  toxicity as determined by the MTT assay.**

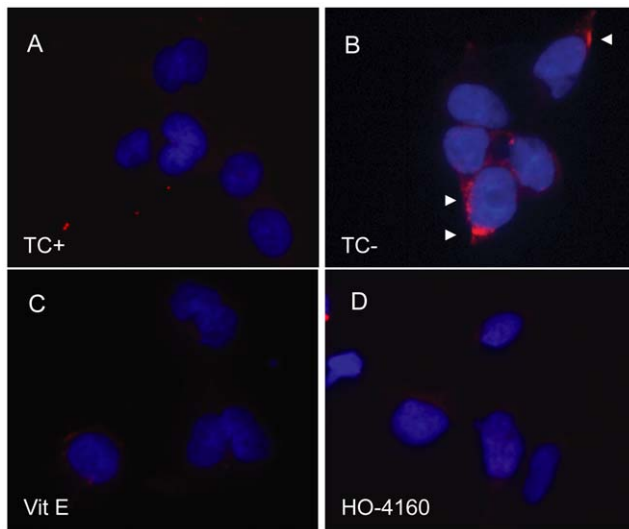
Compound	EC <sub>50</sub> ( $\mu$ M)	Tx <sub>50</sub> ( $\mu$ M)
3a (HO-4160)	0.03	3.1
3b	0.05	1.4
3b/OH/2HCl	0.1	2.5
3c	0.05	1.3
3d	0.1	2.3
4	0.07	1.6
5	1.2	3.1
K01-162	0.12	10

The structures of 3a and K01-162 are shown in Fig. 1, see Fig. S1 in Supporting Information for other structures.

doi:10.1371/journal.pone.0035443.t001



**Figure 3. SLF HO-4160 greatly diminishes the intracellular population of oligomeric A $\beta$**  Shown are Western blots of extracts from the MC65 human neuroblastoma cell line that conditionally expresses C99, a 99-residue carboxyl terminal fragment of APP in the absence of tetracycline (–TC). C99 is subsequently cleaved by cellular  $\gamma$ -secretase to generate A $\beta$ . TP17, an inactive tricyclic pyrone, serves as a negative control [31], while vitamin E (Vit E), a potent antioxidant that was shown to also block A $\beta$ O formation in MC65 cells [30], serves as a positive control. p8 is an unresolved band that could be an A $\beta$  homodimer, or a heterodimer of A $\beta$  and APP $\Delta$ 31 [31], a caspase cleavage product involving residues just downstream from the A $\beta$  origin on APP. The presence of this band does not correlate with MC65 cell death [31]. Blotting was carried out using the A $\beta$  antibody 6E10 (upper panel) and the loading control (lower panel) was probed using an antibody directed against  $\beta$ -actin. The blot shown is representative of 3 replicates.  
doi:10.1371/journal.pone.0035443.g003



**Figure 4. SLF blocks the appearance of oligomeric A $\beta$  species within cultured cells.** MC65 cells were treated with the indicated compounds (1  $\mu$ M) immediately after the removal of the transgene suppressor tetracycline (–TC; panels B–D) to induce A $\beta$ O accumulation. At 24 hours, cells were fixed, immunofluorescently stained with the oligomer-specific antibody A11 (red) and counterstained with the nuclear dye DAPI (blue). As shown in panel (D), the cytoprotective SLF HO-4160, as well as the antioxidant vitamin E (Vit E) attenuate the accumulation of intracellular A $\beta$ O (red fluorescent puncta).  
doi:10.1371/journal.pone.0035443.g004

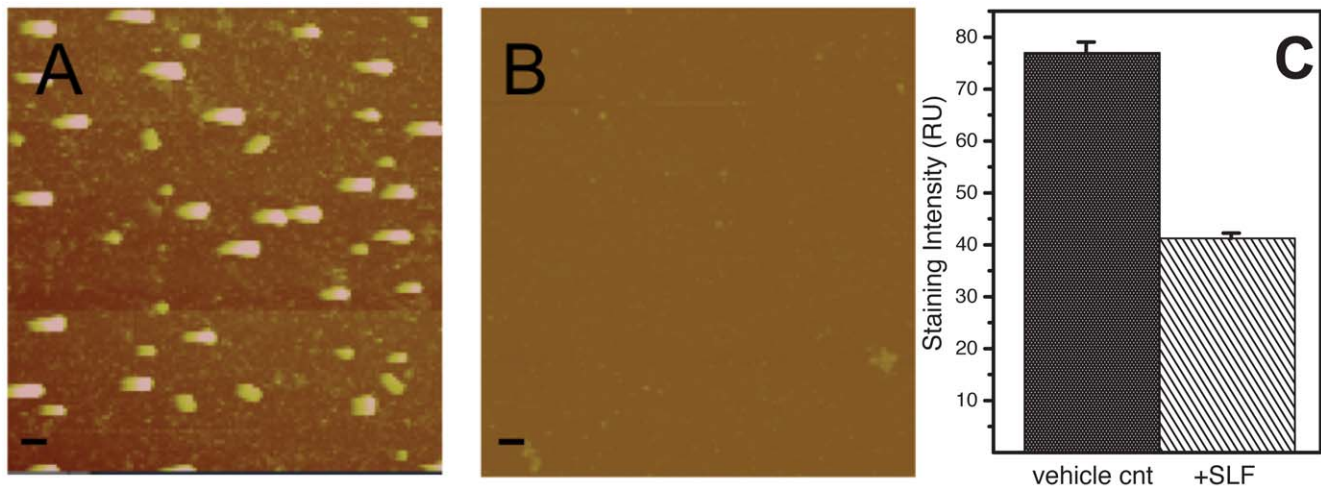
is evident if spin dilution of A $\beta$ <sup>(26TOAC)</sup> is not carried out (Figure 7B).

#### Effects of SLF observed by spin labels located within A $\beta$

As shown previously [6], the dynamics of A $\beta$  containing a spin label near its N-terminus serve to indicate the disruption of A $\beta$ O by active fluorene compounds. Because A $\beta$ O disruption should be accompanied by increased rates of global rotational diffusion in solution, the spin labels attached to A $\beta$  provide an additional means of observing A $\beta$ O disruption by the SLF. Furthermore, if the proximity of the nitroxide moieties located on the fluorene ligand and the A $\beta$  peptide are close enough to interact magnetically, the dipolar broadening may be evident in the composite spectrum.

As shown in Figure 8, 2 hours after addition of the compound to A $\beta$ O harboring a spin label at position 2 or 26, the spectrum greatly increases in amplitude (red trace). This demonstrates that the SLF is able to greatly disrupt the A $\beta$ O after 2 hours. Since spin labels are only attached to 25% of the A $\beta$  peptides, the level of broadening suggests the nitroxides on the oligomeric peptides can interact with more than one docked fluorene. This is supported by the observation that after complete disruption of A $\beta$ O, there is a large increase in the spectral amplitude. Some increase in the amplitude is expected due to faster diffusion of monomeric A $\beta$  [18]. Our investigation of the ability of 10  $\mu$ M SLF to disrupt oligomers formed after 24 hours produced varied results that most likely reflect the heterogeneity of assembly size and structure, some of which appear to be resistant to SLF disruption.





**Figure 5. Clearance of A $\beta$ O aggregates observed by AFM imaging and ThT binding assay.** Small oligomers used in this study are formed within 60 minutes (scale bar 10 nm). The direct visualization of oligomers by using AFM revealed the formation of A $\beta$ O aggregates. (A) 50  $\mu$ M of A $\beta$  after 24 hours of incubation at room temperature (diameter 5–10 nm). (B) 50  $\mu$ M of A $\beta$  and 50  $\mu$ M of SLF after 24 hours of incubation at room temperature. (C) Staining for beta-rich assemblies by the amyloid dye thioflavin T for incubations of A $\beta$  with and without SLF HO-4160 as described in Methods. Data are the averages from 3 separate experiments with the error bars representing the SEM. doi:10.1371/journal.pone.0035443.g005

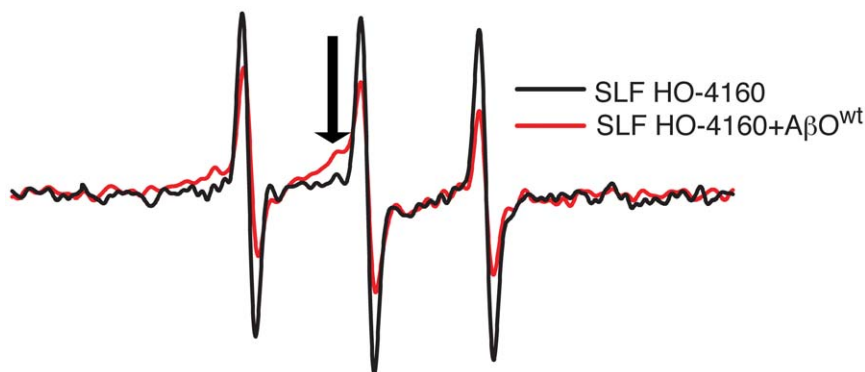
### Interaction between the nitroxides on the SLF ligand and A $\beta$

Figure 9 compares EPR spectra from the following conditions using A $\beta$  spin-labeled at either position 2 or 26: a sample containing spin-labeled A $\beta$ O alone, a sample containing both spin-labeled A $\beta$ O and SLF, and a composite spectrum generated from the individual SLF and spin-labeled A $\beta$ O spectra. From these comparisons, it is evident that both of the spin-labeled positions in A $\beta$ O show at least some interaction with the nitroxide moiety on the SLF. This is based on the observation that the spectrum of the sample with both species labeled is lower in amplitude than the sample containing labeled A $\beta$ O alone. Position 26 in A $\beta$  (Fig. 9B) experiences a stronger interaction with the SLF than position 2 of A $\beta$  (Fig. 9A). The blue trace in Figure 9A or 8B is a sum of the individual spectra of the two spin-labeled species (A $\beta$  and SLF) and indicates what the actual mixed sample spectrum would look like if no interaction were present. However, the observed spectrum of the mixed sample (black trace) is lower than the composite spectrum. The broadening of the mixed spectrum is slight when A $\beta$  contains the spin label at position 2,

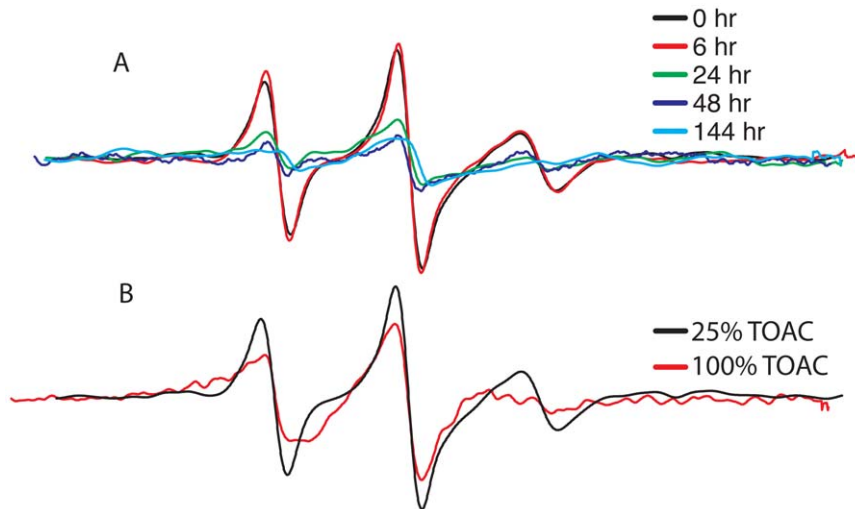
indicating the nitroxide of SLF approaches within 2 nm of the N-terminal region. However, the broadening of the sample containing a spin label at position 26 is more substantial, demonstrating the nitroxide of SLF is in closer proximity (1.5 nm or less).

### Circular dichroism analysis of SLF influences on A $\beta$

Circular dichroism (CD) spectroscopy provides a global indicator for the increase in beta-strand content that accompanies the oligomerization of the A $\beta$  peptide [19]. To determine whether SLF affects the transition of A $\beta$  monomers into secondary structures, CD spectra of A $\beta$  were collected immediately after its initial introduction into aqueous buffer and then throughout a time course of 24 hours. In the absence of SLF, the CD spectra of A $\beta$  reflect a transition from an unstructured random coil to a spectrum characteristic of beta-strand secondary structure (Fig. 10) [20]. The most substantial effect of SLF on the CD spectrum of A $\beta$  is seen at 1 hour, where in the absence of SLF the spectrum shows a distinct beta absorption pattern. However in the presence of SLF, the peptide maintains a largely disordered structure after



**Figure 6. EPR examination indicates the motional freedom of SLF is reduced by interaction with A $\beta$ O.** A $\beta$ O imparts a broad component (arrow) into the EPR spectrum of HO-4160, reflecting a population of SLF with a slower correlation time. doi:10.1371/journal.pone.0035443.g006



**Figure 7. Spectral broadening due to immobilization and dipolar interaction in assemblies of A $\beta$ .** (A) Changes in the correlation time of A $\beta^{(26\text{TOAC})}$  with increased incubation time. To mainly observe the effects on dynamics, the spins in the A $\beta$  preparation were diluted with wild-type peptide (A $\beta_{(1-40)}$ ), so that the sample contained only 25% A $\beta^{(26\text{TOAC})}$ . (B) Comparisons of EPR spectra of 80  $\mu\text{M}$  A $\beta\text{O}$  preparations containing either 25% (black line) or 100% (red line) A $\beta^{(26\text{TOAC})}$ . Both spectra were acquired after 1 hour at room temperature. doi:10.1371/journal.pone.0035443.g007

1 hour. Although SLF does slow the development of beta secondary structure, by 24 hours the CD spectrum of A $\beta$  in the presence of SLF shows strong absorption in the wavelength range that interacts most efficiently with a beta-sheet fold. As shown in Figure 10, however, the 24-hour spectra for A $\beta$  in the presence and absence of SLF are significantly different, suggesting a unique conformation for the peptide in the presence of SLF. This is not unexpected, as the peptide's structure likely adopts a distinct fold as it incorporates into larger assemblies. As the neurotoxicity of A $\beta$  correlates more strongly with aggregative ability than secondary structure [21], compounds that influence the former provide better candidates for intervening in the molecular pathology of AD.

### SLF scavenges free radicals

Sterically hindered alpha, alpha'-tetrasubstituted 5-membered cyclic secondary amines are sensitive to oxidation by ROS to N-oxyls. In turn, the N-oxyls can be oxidized to the N-oxoimmonium, or reduced to diamagnetic N-hydroxylamines [22,23,24]. A major advantage of the N-oxyl radical is that it represents a stable free radical that does *not* induce damage to DNA, proteins, lipids or sugars. Thus, this added feature of the fluorene should improve its ability to lower oxidative stress by either donating or accepting electrons with radicals ( $\bullet\text{R}$ ), such as reactive oxygen species ( $\bullet\text{ROS}$ ).

To measure the free-radical scavenging potential of candidate compounds, spin-trapping in the presence and absence of SLF was measured by EPR spectroscopy to look for the depletion of superoxide and hydroxyl radicals. The EPR spectra of BMPO adducts with superoxide and hydroxyl radicals (generated via horseradish peroxidase/ $\text{H}_2\text{O}_2$  and iron sulfate/ $\text{H}_2\text{O}_2$ , respectively) are shown in green (Fig. 11). The spectrum in the presence of SLF is shown in red, reflecting a decrease in the amount of BMPO adduct formed. The antioxidant activity of the SLF further extends to cellular systems. In the same MC65 neuroblastoma cell model described previously, addition of SLF attenuates the production of hydrogen peroxide in response to APP-C99 expression ( $-\text{TC}$ ) compared to control cells ( $+\text{TC}$ ) lacking APP-C99 (Fig. 12). The reduced hydrogen peroxide production approaches control levels and is similar to the reduced levels

achieved by treatment of A $\beta$ -generating cells with the antioxidant vitamin E.

### Summary

We have previously shown that selected fluorene compounds can rapidly disrupt A $\beta\text{O}$  in solution, and dramatically attenuate A $\beta\text{O}$  toxicity *in vivo* [6]. The results reported here demonstrate how SLFs help elucidate the mechanism of fluorene action and, more importantly, that SLFs have superior potency in alleviating A $\beta\text{O}$ -induced toxicity. Simulations of the SLF parent K01-162 and A $\beta_{(1-42)}$  show preferential interaction of the compound with a hydrophobic core region of the peptide constituted by residues 17–21 [25]. Figure 13 illustrates one potential mode of interaction between A $\beta\text{O}$  and SLF consistent with our findings regarding substituted fluorenes [6], where the binding of SLF to A $\beta$  occludes a hydrophobic interface that facilitates peptide oligomerization.

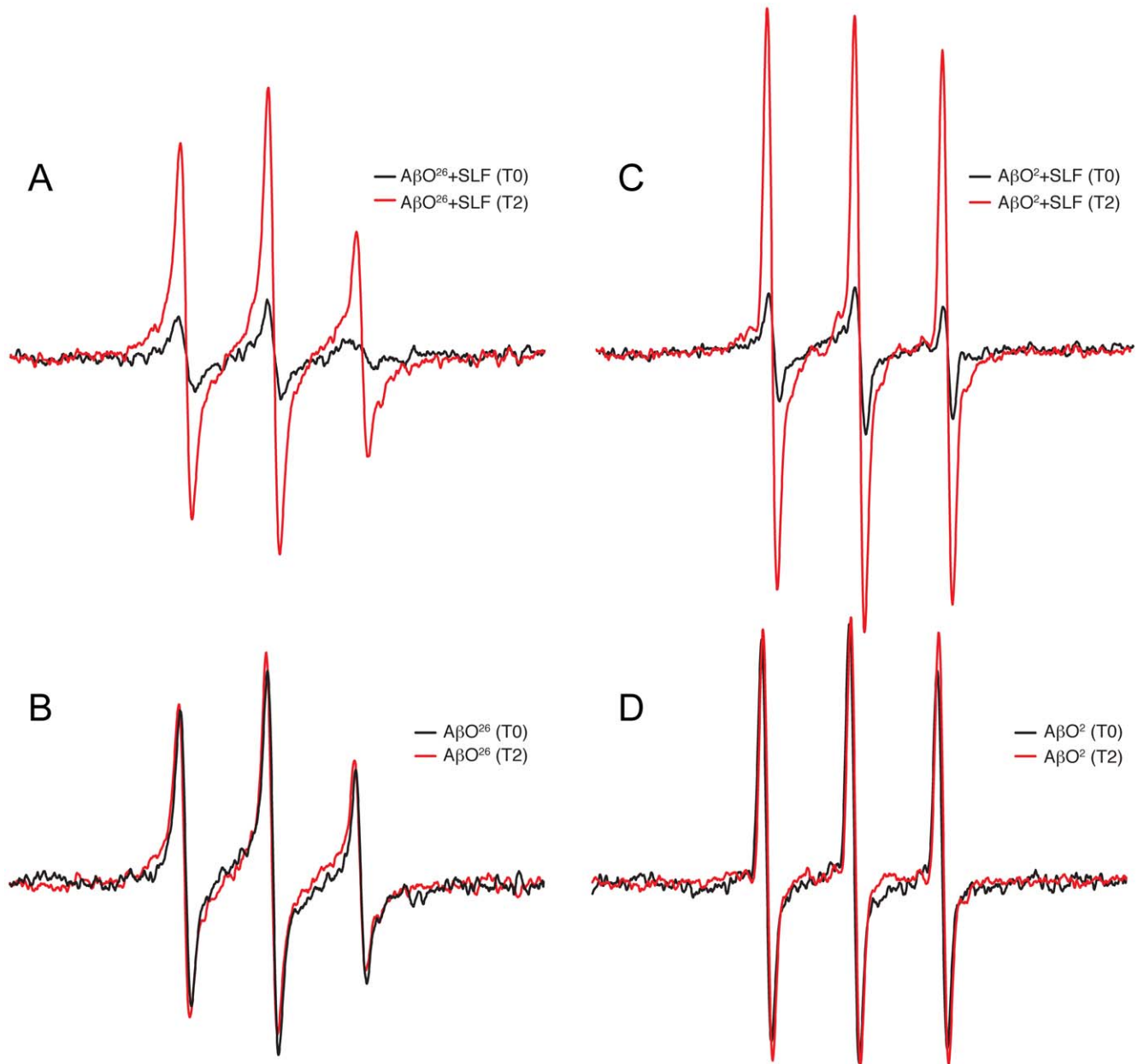
### Materials and Methods

#### Materials

Spin-labeled fluorene HO-4160 (7-Bromo-N —[(1-Oxyl-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-yl)methyl]-9H-fluorene-2-amine radical) was synthesized as previously described [15]. Peptide labeled at position 26 (A $\beta^{(26\text{TOAC})}$ ) was synthesized with the TOAC nitroxide incorporated at position 26 as previously described [18]. Peptide labeled at position 2 (A $\beta^{(2\text{R}1)}$ ), was first synthesized with a Cys residue substituted at position 2, and then reacted with the methanethiosulfonate spin label (MTS-SL) [26], as previously described [6]. The structures and approximate locations of each spin label are indicated in Figure 1. MC65 cells were generated by Sopher et. al. [27], and gifted to L-W.J.

#### Preparation of A $\beta$ oligomers

Solid A $\beta_{(1-40)}$  peptide (Bachem Cat # 1194.0001, Torrance, CA) was dissolved in Hexa-Fluoro-Iso-Propanol (HFIP, Sigma, St. Louis, MO). The solution was incubated at room temperature for 1 day until the solution became clear and colorless. HFIP is a strong reducing reagent that can break hydrogen bonds and keep A $\beta_{(1-40)}$  in the monomeric form. All the HFIP was removed by



**Figure 8. Fluorenes bind A $\beta$ O and destabilize highly organized secondary structure.** EPR spectra of SLF (10  $\mu$ M) with single-cysteine mutant A $\beta$ <sub>(1-40)</sub> oligomers (40  $\mu$ M) show that motional freedom of the mixture increased following a 2-hour incubation at room temperature (red) compared with 0 hours (black). A) and B) 26 A $\beta$ O: spin labeled at 26<sup>th</sup> residue C) and D) 2 A $\beta$ O: spin labeled at 2<sup>nd</sup> residue. doi:10.1371/journal.pone.0035443.g008

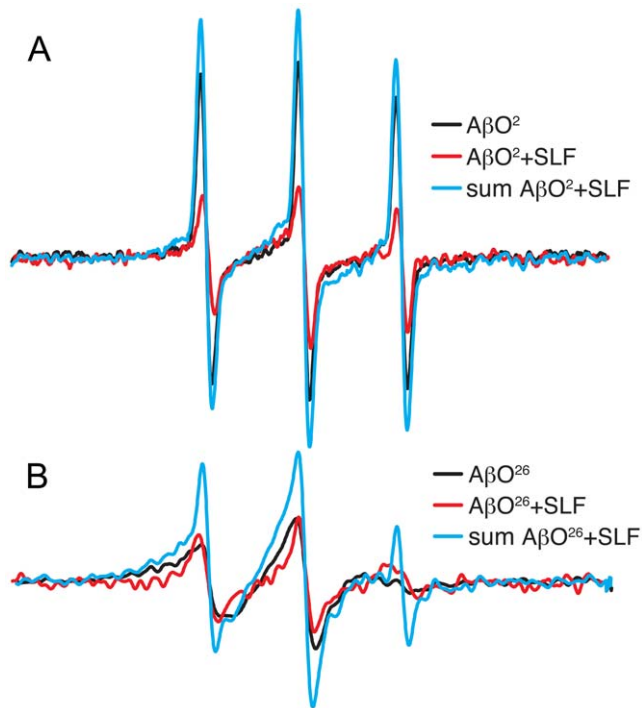
SpeedVac Concentrator (Savant, SV100H, Thermo Scientific, Waltham, MA). To generate A $\beta$ <sub>(1-40)</sub> oligomers, a 100% DMSO stock solution of 1 mM A $\beta$ <sub>(1-40)</sub> was diluted in cold PBS buffer pH 7.4 to a total concentration of 100  $\mu$ M. For EPR experiments using spin-labeled A $\beta$ , the A $\beta$ O preparations contained a mixture of 25% spin-labeled peptide to 75% native A $\beta$ <sub>(1-40)</sub>. This dilution of labeled peptide in the A $\beta$ O sample minimizes the broadening of the EPR spectrum that arises from the dipolar interaction of spins in close proximity (<2 nm) [28].

#### Cell culture models exhibiting intraneuronal A $\beta$ O

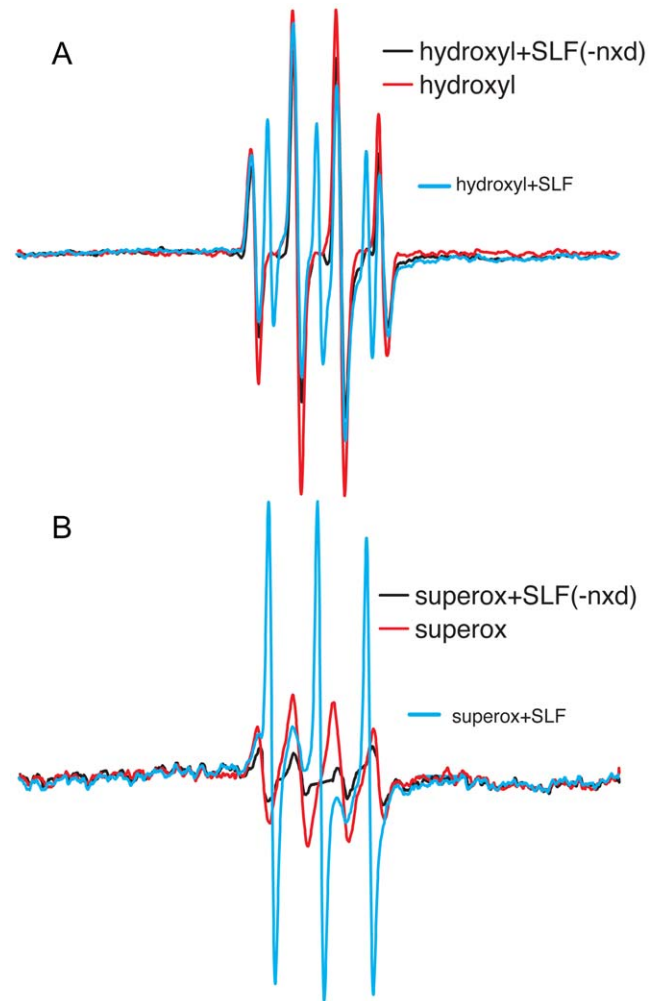
The cell culture model used for these studies was the human neuroblastoma cell line (MC65) equipped with conditional

expression of the carboxyl-terminal 99 residues of the amyloid- $\beta$  precursor protein (APP-C99). A $\beta$  is generated from APP-C99 after proteolysis by cellular  $\gamma$ -secretase. To induce cellular A $\beta$  production, the transgene suppressor, tetracycline (TC), was removed from the media, as described previously [3]. Intracellular A $\beta$ O started to accumulate as early as 4 hours after TC removal. The cytotoxicity was determined on day 3 using a colorimetric MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] assay, the results of which were comparable to data obtained using counts of viable cells based on trypan blue exclusion and the LIVE/DEAD assay (Invitrogen, Grand Island, NY). To test the fluorene compounds, the compounds were added

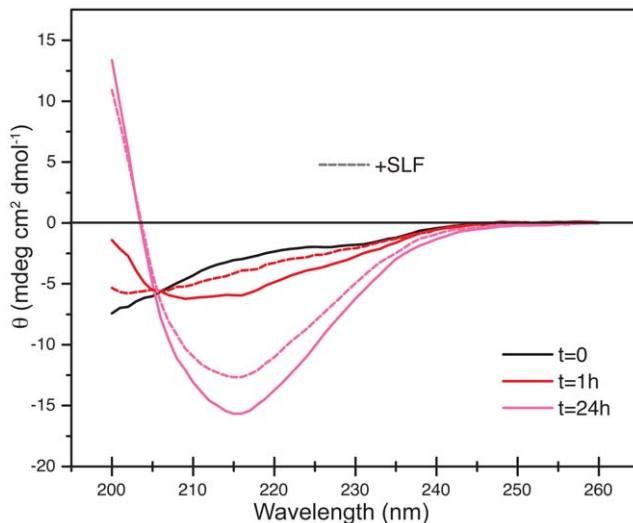




**Figure 9. The comparison of EPR spectra from A $\beta$  alone, an actual mixture of SLF and A $\beta$ , and the mathematical sum of the individual EPR signals from SLF and A $\beta$ .** Data were taken for samples containing A $\beta$ O spin labeled at the 2<sup>nd</sup> residue (A) and for samples containing A $\beta$ O spin labeled at the 26<sup>th</sup> residue (B).  
doi:10.1371/journal.pone.0035443.g009



**Figure 11. The EPR spectra of BMPO adducts with hydroxyl (A) and superoxide (B) radicals (generated via iron sulfate/H<sub>2</sub>O<sub>2</sub> and horseradish peroxidase/H<sub>2</sub>O<sub>2</sub>, respectively) are shown in green.** The spectra in the presence of SLF are shown in red, reflecting a decrease in the amount of BMPO adduct formed. Based on the difference in the spectral intensities of the generated BMPO-adduct, SLF HO-4160 is able to scavenge approximately 80% and 25% of the superoxide and hydroxyl radicals, respectively.  
doi:10.1371/journal.pone.0035443.g011



**Figure 10. Circular dichroism analysis of secondary structural changes in A $\beta$ .** The time zero ( $t=0$ ) trace was obtained by scanning A $\beta$  immediately after introduction into buffer. Inclusion of SLF had no significant effect on the  $t=0$  spectrum (not shown). Additional traces were obtained at 1, 2, 4, 6 and 24 hours. For both the control and +SLF samples, the  $t=2$ , 4, and 6 hour traces are not shown, as these curves changed in a stepwise manner between the  $t=1$  and  $t=24$  hour time points. See Methods for additional details.  
doi:10.1371/journal.pone.0035443.g010

immediately after TC removal, and the cells were maintained for 3 days without media change before the MTT assay.

#### Western blotting

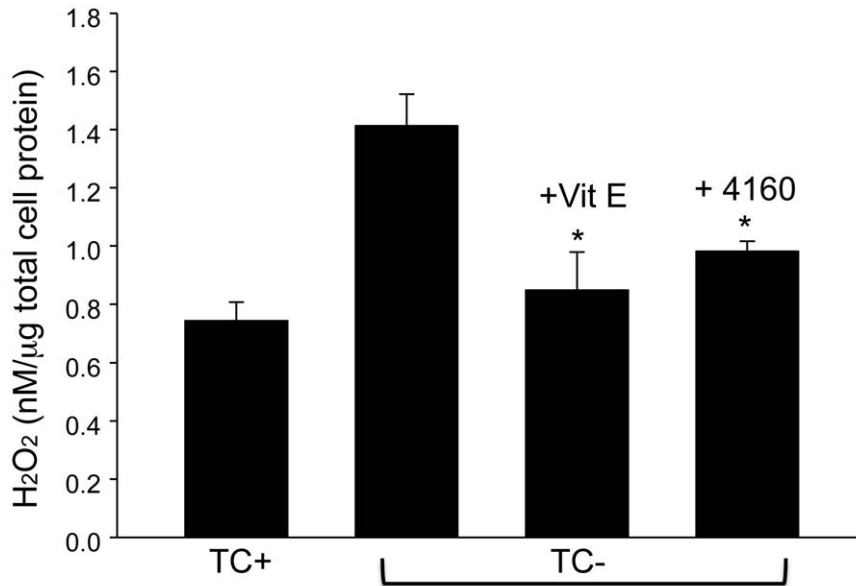
The preparation of cell homogenates and Western blotting were performed as previously described [29,30].

#### Immunofluorescence staining

Immunofluorescent labeling of A $\beta$ O in cultured MC65 cells by the A11 anti-oligomer antibody (Chemicon-Millipore, Billerica, MA) was performed according to our previously published protocols [16].

#### Atomic force microscopy (AFM)

AFM was employed to analyze the oligomer formation of wild-type A $\beta$ O. All surface scans employed a Dimension 3100 Scanning Probe Microscope with a Hybrid closed-loop XYZ



**Figure 12. Cultured neuronal cells treated with SLF HO-4160 decrease production of hydrogen peroxide.** MC65 cells were cultured in the presence (TC+) or absence (TC-) of tetracycline, a transgene suppressor of the APP fragment C99. In the absence of tetracycline, cells express the C99 fragment which is then further cleaved to form A $\beta$ . Treatment with either the HO-4160 compound (+4160, 0.3  $\mu$ M) or the antioxidant vitamin E (+Vit E, 100  $\mu$ M) reduces the extracellular accumulation of hydrogen peroxide in cells expressing APP-C99. Error bars represent the SEM for N=3. \* $p$ <0.05 TC- compared to both + Vit E and +4160. doi:10.1371/journal.pone.0035443.g012

head and Nanoscope IVa controller (Veeco, Santa Barbara, CA). All samples were prepared on freshly-cleaved mica (Ted Pella, Redding, CA) and imaged in tapping mode in air by a phosphorous-doped silicon cantilever with a nominal spring constant of 40 N/m. Particle dimension measurements and image enhancement were performed with the Nanoscope software supplied by Veeco, version 6.14. For each measurement, an aliquot of A $\beta$ O was removed from the 1 mM DMSO stock solution and diluted to 50  $\mu$ M of A $\beta$ O in PBS pH 7.4, and immediately spotted on freshly cleaned mica. After 2 minutes the samples were washed with 200  $\mu$ L distilled water and then

partially dried by compressed air and completely dried at room temperature [6].

#### Thioflavin T (ThT) binding assay

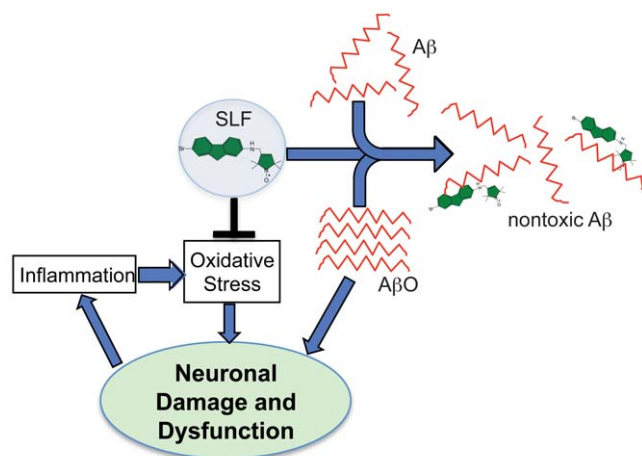
A $\beta$ <sub>(1-40)</sub> (40  $\mu$ M) in the presence or absence of SLF (10  $\mu$ M) was incubated at room temperature for 24 hours. A $\beta$ <sub>(1-40)</sub> (20  $\mu$ L) with or without SLF was incubated in 170  $\mu$ L PBS buffer and 10  $\mu$ L ThT (ThT stock: 0.1 mM stored in the dark at 4°C) for 15 minutes in the dark. Then ThT fluorescence was measured by a VICTOR3 Multilabel Plate Counter (PerkinElmer, Waltham, MA) spectrofluorometer at an excitation of 450 nm, with excitation and emission slit widths of 10 nm.

#### Circular dichroism spectroscopy

CD measurements were performed on a Jasco J-810 spectropolarimeter equipped with a Jasco CDF-426S Peltier set to 25°C (Jasco, Easton, MD). A $\beta$ <sub>(1-40)</sub> was diluted to 40–80  $\mu$ M in phosphate buffer (25 mM, pH 7.4). SLF in DMSO (final concentration 10–20  $\mu$ M) or DMSO alone (final concentration 0.01–0.02%) was added to the A $\beta$ , and the samples incubated for 0, 1, 2, 4, 6 and 24 hours at room temperature. After each incubation time point the samples were placed in a 0.1 mm quartz cuvette and, after extensive purging with nitrogen, scanned in the 200 to 260 nm region (scan speed was 20 nm/min). Averages of five scans were baseline-subtracted (25 mM phosphate with 0.01–0.02% DMSO, or 25 mM phosphate with 10–20  $\mu$ M of SLF in DMSO).

#### EPR spectroscopy

EPR measurements were carried out in a JEOL TE-100 X-band spectrometer fitted with a loop-gap resonator as described previously [23] (JEOL USA, Peabody, MA). SLF (10  $\mu$ M) was added to the spin-labeled A $\beta$ O (40  $\mu$ M) at a final concentration of 32  $\mu$ M for 0 and 2 hours prior to EPR measurements. Appropriate vehicle controls were used for all samples. Approx-



**Figure 13. Model of action of SLFs on A $\beta$ O assemblies.** The illustration highlights the bifunctional properties of the SLF, including its ability to block the formation of nm particles and disrupt small oligomers, as well as its antioxidant activity. The ability of SLF to disrupt fibrils or more mature fibrillar oligomers [32] is undetermined. doi:10.1371/journal.pone.0035443.g013

imately 5  $\mu$ l of the protein, at a final concentration of 32  $\mu$ M was loaded into a sealed quartz capillary tube. The spectra were obtained by averaging two 2-minute scans with a sweep width of 100 G at a microwave power of 4 mW and modulation amplitude optimized to the natural line width of the attached spin probe. All the spectra were recorded at room temperature.

### Antioxidant activity

**Measurement of superoxide and hydroxyl free radicals by EPR.** The free radical scavenging activity of SLF compounds was determined by measuring the adduct levels accumulated by the spin trap BMPO. Briefly, a mixture of horseradish peroxidase (100 ng) and 0.03% hydrogen peroxide in PBS pH 7.4 was used to generate superoxide radicals. Hydroxyl radicals were generated by mixing ferrous ammonium sulfonate (0.1 mM) and hydrogen peroxidase (0.1 mM) in PBS. All EPR measurements were performed in PBS buffer pH 7.4 which contained BMPO (1 mM) in the presence or absence of SLFs at varied concentrations. The superoxide and hydroxyl radicals were measured as BMPO-OOH and BMPO-OH adducts, respectively.

**Scavenging of peroxyl and hydroxyl radicals measured by fluorescence.** Fluorescence detection was determined using the Radical Absorbance Capacity Assay (Cell Biolabs, Inc., San Diego, CA) according to the manufacturer's instructions. Briefly, the indicator is oxidized by ROS species resulting in a loss of

fluorescence. Thus ROS scavenging activity is determined by the intensity of fluorescence following addition of the hydroxyl or peroxyl challenge.

**Measurement of cellular hydrogen peroxide levels.** MC65 cells were plated onto 12-well plates at  $2 \times 10^5$  cells per well in Opti-MEM with and without tetracycline (1  $\mu$ g/ml). Compounds, such as HO-4160 (0.3  $\mu$ M) or  $\alpha$ -Tocopherol (referred to as vitamin E, 100  $\mu$ M, Sigma, St. Louis, MO) were added to the cultures immediately after plating. Culture medium was collected after a 24-hour incubation at 37°C. Hydrogen peroxide in the conditioned medium was analyzed by the Amplex Red Hydrogen Peroxide/Peroxidase Assay kit following the instructions of the manufacturer (Invitrogen, Grand Island, NY).

### Supporting Information

**Figure S1** Structures of SLF compounds [15] evaluated in Table 1. (TIF)

### Author Contributions

Conceived and designed the experiments: JCV KH LWJ. Performed the experiments: JP RA IM HSH DAB. Analyzed the data: JCV JP LWJ. Contributed reagents/materials/analysis tools: TK KH LWJ GAL GH ANP. Wrote the paper: JP RA JCV.

### References

- LaFerla FM, Green KN, Oddo S (2007) Intracellular amyloid-beta in Alzheimer's disease. *Nat Rev Neurosci* 8: 499–509.
- Klein WL, Stine WB, Jr., Teplow DB (2004) Small assemblies of unmodified amyloid beta-protein are the proximate neurotoxin in Alzheimer's disease. *Neurobiol Aging* 25: 569–580.
- Maezawa I, Hong HS, Liu R, Wu CY, Cheng RH, et al. (2008) Congo red and thioflavin-T analogs detect Abeta oligomers. *J Neurochem* 104: 457–468.
- Lesne S, Koh MT, Kotilinek L, Kaye R, Glabe CG, et al. (2006) A specific amyloid-beta protein assembly in the brain impairs memory. *Nature* 440: 352–357.
- Pardridge WM (2007) Blood-brain barrier delivery. *Drug Discov Today* 12: 54–61.
- Hong HS, Maezawa I, Budamagunta M, Rana S, Shi A, et al. (2010) Candidate anti-Abeta fluorene compounds selected from analogs of amyloid imaging agents. *Neurobiol Aging* 31: 1690–1699.
- Lee CW, Kung MP, Hou C, Kung HF (2003) Dimethylamino-fluorenes: ligands for detecting beta-amyloid plaques in the brain. *Nucl Med Biol* 30: 573–580.
- Hong HS, Rana S, Barrigan L, Shi A, Zhang Y, et al. (2009) Inhibition of Alzheimer's amyloid toxicity with a tricyclic pyrone molecule in vitro and in vivo. *J Neurochem* 108: 1097–1108.
- Lin MT, Beal MF (2006) Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* 443: 787–795.
- Lipton SA (2006) Paradigm shift in neuroprotection by NMDA receptor blockade: memantine and beyond. *Nat Rev Drug Discov* 5: 160–170.
- Farooqui AA, Yang HC, Horrocks L (1997) Involvement of phospholipase A2 in neurodegeneration. *Neurochem Int* 30: 517–522.
- Hideg K, Kalai T (2007) Novel antioxidants in anthracycline cardiotoxicity. *Cardiovasc Toxicol* 7: 160–164.
- Mandal R, Kutala VK, Khan M, Mohan IK, Varadharaj S, et al. (2007) N-hydroxy-pyrroline modification of verapamil exhibits antioxidant protection of the heart against ischemia/reperfusion-induced cardiac dysfunction without compromising its calcium antagonistic activity. *J Pharmacol Exp Ther* 323: 119–127.
- Cai L, Innis RB, Pike VW (2007) Radioligand development for PET imaging of beta-amyloid (A $\beta$ )—current status. *Curr Med Chem* 14: 19–52.
- Kalai T, Petrlova J, Balog M, Aung HH, Voss JC, et al. (2011) Synthesis and study of 2-amino-7-bromofluorenes modified with nitroxides and their precursors as dual anti-amyloid and antioxidant active compounds. *Eur J Med Chem* 46: 1348–1355.
- Hong HS, Maezawa I, Yao N, Xu B, Diaz-Avalos R, et al. (2007) Combining the rapid MTT formazan exocytosis assay and the MC65 protection assay led to the discovery of carbazole analogs as small molecule inhibitors of A $\beta$  oligomer-induced cytotoxicity. *Brain Res* 1130: 223–234.
- Kayed R, Head E, Thompson JL, McIntire TM, Milton SC, et al. (2003) Common structure of soluble amyloid oligomers implies common mechanism of pathogenesis. *Science* 300: 486–489.
- Petrlova J, Hong HS, Bricarello DA, Harishchandra G, Lorigan GA, et al. (2011) A differential association of Apolipoprotein E isoforms with the amyloid-beta oligomer in solution. *Proteins* 79: 402–416.
- Barrow CJ, Yasuda A, Kenny PT, Zagorski MG (1992) Solution conformations and aggregational properties of synthetic amyloid beta-peptides of Alzheimer's disease. Analysis of circular dichroism spectra. *J Mol Biol* 225: 1075–1093.
- Kelly SM, Jess TJ, Price NC (2005) How to study proteins by circular dichroism. *Biochim Biophys Acta* 1751: 119–139.
- Liao MQ, Tzeng YJ, Chang LY, Huang HB, Lin TH, et al. (2007) The correlation between neurotoxicity, aggregative ability and secondary structure studied by sequence truncated A $\beta$  peptides. *FEBS Lett* 581: 1161–1165.
- Lam MA, Pattison DL, Bottle SE, Keddie DJ, Davies MJ (2008) Nitric oxide and nitroxides can act as efficient scavengers of protein-derived free radicals. *Chem Res Toxicol* 21: 2111–2119.
- Twomey P, Taira J, DeGraff W, Mitchell JB, Russo A, et al. (1997) Direct evidence for in vivo nitroxide free radical production from a new antiarrhythmic drug by EPR spectroscopy. *Free Radic Biol Med* 22: 909–916.
- Glebska J, Pulaski L, Gwozdziński K, Skolimowski J (2001) Structure-activity relationship studies of protective function of nitroxides in Fenton system. *Biomaterials* 14: 159–170.
- Li J, Liu R, Lam KS, Jin LW, Duan Y (2011) Alzheimer's disease drug candidates stabilize A $\beta$ -protein native structure by interacting with the hydrophobic core. *Biophys J* 100: 1076–1082.
- Berliner LJ, Grunwald J, Hankovszky HO, Hideg K (1982) A novel reversible thiol-specific spin label: papain active site labeling and inhibition. *Anal Biochem* 119: 450–455.
- Sopher BL, Fukuchi K, Smith AC, Leppig KA, Furlong CE, et al. (1994) Cytotoxicity mediated by conditional expression of a carboxyl-terminal derivative of the beta-amyloid precursor protein. *Brain Res Mol Brain Res* 26: 207–217.
- Likhtenshtein GI (1993) Biophysical labeling methods in molecular biology. New York: Cambridge University Press. pp 46–79.
- Jin LW, Shie FS, Maezawa I, Vincent I, Bird T (2004) Intracellular accumulation of amyloidogenic fragments of amyloid-beta precursor protein in neurons with Niemann-Pick type C defects is associated with endosomal abnormalities. *Am J Pathol* 164: 975–985.
- Maezawa I, Jin LW, Woltjer RL, Maeda N, Martin GM, et al. (2004) Apolipoprotein E isoforms and apolipoprotein AI protect from amyloid precursor protein carboxy terminal fragment-associated cytotoxicity. *J Neurochem* 91: 1312–1321.
- Maezawa I, Hong HS, Wu HC, Battina SK, Rana S, et al. (2006) A novel tricyclic pyrone compound ameliorates cell death associated with intracellular amyloid-beta oligomeric complexes. *J Neurochem* 98: 57–67.
- Wu J, Breydo L, Isas JM, Lee J, Kuznetsov YG, et al. (2009) Fibrillar oligomers nucleate the oligomerization of monomeric A $\beta$ , but do not seed fibril formation. *J Biol Chem*.