Check for updates

Effect of atorvastatin, cholesterol ester transfer protein inhibition, and diabetes mellitus on circulating proprotein subtilisin kexin type 9 and lipoprotein(a) levels in patients at high cardiovascular risk

Benoit J. Arsenault, PhD¹, Francine Petrides, PhD¹, Fatiha Tabet, PhD¹, Weihang Bao, PhD, G. Kees Hovingh, MD, PhD, S. Matthijs Boekholdt, MD, PhD, Stéphane Ramin-Mangata, BSc, Olivier Meilhac, PhD, David DeMicco, PharmD, Kerry-Anne Rye, PhD, David D. Waters, MD, John J. P. Kastelein, MD, PhD, Philip Barter, MD, PhD, Gilles Lambert, PhD*

Centre de Recherche de l'Institut Universitaire de Cardiologie et de Pneumologie de Québec, Québec, Québec, Canada (Dr Arsenault); Department of Medicine, Faculty of Medicine, Université Laval, Québec, Québec, Canada (Dr Arsenault); School of Medical Sciences, The University of New South Wales, Sydney, New South Wales, Australia (Drs Petrides, Tabet, Rye, and Barter); Pfizer Inc, New York, NY, USA (Drs Bao and DeMicco); Department of Vascular Medicine, Academic Medical Center, Amsterdam, The Netherlands (Drs Hovingh and Kastelein); Department of Cardiology, Academic Medical Center, Amsterdam, The Netherlands (Dr Boekholdt); Inserm, UMR 1188 DéTROI, Université de La Réunion, Sainte–Clotilde, France (Drs Ramin-Mangata, Meilhac, and Lambert); and Division of Cardiology, University of California, San Francisco, CA, USA (Dr Waters)

KEYWORDS:

Atorvastatin; Diabetes mellitus; Lipoprotein(a); PCSK9; CETP **BACKGROUND:** Proprotein subtilisin kexin type 9 (PCSK9) and lipoprotein (a) [Lp(a)] levels are causative risk factors for coronary heart disease.

OBJECTIVES: The objective of the study was to determine the impact of lipid-lowering treatments on circulating PCSK9 and Lp(a).

METHODS: We measured PCSK9 and Lp(a) levels in plasma samples from Investigation of Lipid Level Management to Understand its Impact in Atherosclerotic Events trial patients with coronary heart disease and/or type II diabetes (T2D) mellitus. Patients received atorvastatin, which was titrated (10, 20, 40, or 80 mg/d) to achieve low-density lipoprotein cholesterol levels <100 mg/dL (baseline) and were subsequently randomized either to atorvastatin + torcetrapib, a cholesterol ester transfer protein inhibitor, or to atorvastatin + placebo.

Clinical Trial Registration: NCT00134264.

* Corresponding author. Inserm UMR 1188, Plateforme CYROI, 2 Rue Maxime Rivière, 97490 Sainte Clotilde, France. E-mail address: gilles.lambert@univ-reunion.fr Submitted April 11, 2017. Accepted for publication October 3, 2017.

1933-2874/© 2017 National Lipid Association. All rights reserved. https://doi.org/10.1016/j.jacl.2017.10.001

¹ These authors are considered as first equal authors.

RESULTS: At baseline, both plasma PCSK9 and Lp(a) were dose-dependently increased with increasing atorvastatin doses. Compared with patients without T2D, those with T2D had higher PCSK9 (357 ± 123 vs 338 ± 115 ng/mL, P = .0012) and lower Lp(a) levels (28 ± 32 vs 32 ± 33 mg/dL, P = .0005). Plasma PCSK9 levels significantly increased in patients treated with torcetrapib (+13.1 \pm 125.3 ng/mL [+3.7%], P = .005), but not in patients treated with placebo (+2.6 \pm 127.9 ng/mL [+0.7%], P = .39). Plasma Lp(a) levels significantly decreased in patients treated with placebo (+0.3 \pm 9.4 mg/dL [+0.1%], P = .92).

CONCLUSION: In patients at high cardiovascular disease risk, PCSK9 and Lp(a) are positively and dose-dependently correlated with atorvastatin dosage, whereas the presence of T2D is associated with higher PCSK9 but lower Lp(a) levels. Cholesterol ester transfer protein inhibition with torcetrapib slightly increases PCSK9 levels and decreases Lp(a) levels. © 2017 National Lipid Association. All rights reserved.

Introduction

Statin therapy decreases low-density lipoprotein (LDL) cholesterol levels and thereby reduces cardiovascular disease (CVD) risk.¹ By inhibiting intracellular cholesterol synthesis, statins increase the expression of the LDL receptor (LDLR), thus promoting an enhanced clearance of LDL particles. However, stating also increase the expression of proprotein convertase subtilisin kexin type 9 (PCSK9), a natural circulating inhibitor of the LDLR.² PCSK9 binds to the LDLR and after endocytosis targets the LDLR that normally recycles back to the cell surface, for lysosomal degradation. The efficacy of statins in reducing LDL cholesterol levels appears to be partially offset by a concomitant rise in PCSK9.^{3,4} Pharmacologic inhibition of PCSK9 with monoclonal antibodies lowers circulating LDL cholesterol further in patients at high CVD risk and not at LDL cholesterol therapeutic goals despite aggressive statin treatment.^{5,6} It is therefore important to determine whether and to what extent statins dose-dependently increase circulating PCSK9 levels in such patients.

In contrast to statins, anti-PCSK9 monoclonal antibodies promote an unexplained 25% to 30% reduction in circulating lipoprotein (a) [Lp(a)] levels.⁷ Lp(a) is a lipoprotein subfraction analogous to LDL, where a unique protein homologous to plasminogen, apolipoprotein (a) [apo(a)], is covalently tethered to apolipoprotein B100 by a unique disulfide bond.⁸ Approximately 20% of the Caucasian population have high Lp(a) levels (above 50 mg/dL)⁹ and a consequent increased risk of coronary heart disease, stroke, calcific aortic valve stenosis, and heart failure.¹⁰⁻¹³ The molecular mechanisms of Lp(a) assembly that likely occurs at the surface of hepatocytes between a newly synthesized apo(a) and apoB100 containing lipoproteins (LpB) remain elusive.¹⁴ Apo(a) is never found associated with triglyceride-rich lipoproteins but rather on cholesterol-rich LDL particles.^{15,16} The impact of statin therapy on plasma Lp(a) levels is somewhat controversial with studies documenting slight decreases in Lp(a) while others suggest that statins actually increase Lp(a) levels as well as the amount of oxidized phospholipids carried by Lp(a).¹⁷ Similar to PCSK9, whether statins dose-dependently influence Lp(a) plasma levels and whether this could be dependent on metabolic disturbances is unknown.

To shed light on the metabolic states favoring Lp(a) assembly, and thus elevated Lp(a) levels, in conjunction with the ongoing development of PCSK9 inhibitors, we aimed to determine the impact of different doses of statins, with and without metabolic disturbances of triglyceriderich lipoproteins (eg, in type II diabetes [T2D]) and with and without modulation of their cholesterol content (eg, by inhibition of the cholesterol ester transfer protein [CETP]) on circulating PCSK9 and Lp(a) levels in patients at high cardiovascular risk.

Methods

Study design

The Investigation of Lipid Level Management to Understand its Impact in Atherosclerotic Events (ILLUMI-NATE) trial included 15,067 men and women at high cardiovascular risk (Trial Registration Number: NCT00134264). The details of the study population have been previously described.¹⁸ Briefly, men and women aged 45 to 75 years were eligible if they had a prior history of myocardial infarction, stroke, acute coronary syndrome, unstable angina, peripheral vascular disease, or cardiac revascularization within the period of 30 days to 5 years before screening. Patients with T2D who met American Diabetes Association criteria or who were currently on hypoglycemic agents were also eligible. During a run-in period of 4 to 10 weeks, patients received atorvastatin, which was titrated (if needed) at 2-week intervals to achieve LDL cholesterol levels <100 mg/dL with atorvastatin 10, 20, 40, or 80 mg/d. Patients whose LDL cholesterol level met the target were randomly assigned to receive either atorvastatin (at a dose established during the run-in period) plus 60 mg of torcetrapib or atorvastatin plus placebo. Noteworthy, the use of torcetrapib in ILLU-MINATE was found to have serious adverse off-target effects with an increase in cardiovascular events and related deaths.¹⁸ We randomly selected 594 patients receiving torcetrapib and atorvastatin matched in a 1:2 ratio to 1151 patients receiving placebo and atorvastatin. This subset of patients was selected to ensure patients with T2D were overrepresented (48%) and that all atorvastatin doses (10, 20, 40, and 80 mg/d) were also equally represented.

Laboratory analyses

The level of high-density lipoprotein (HDL) cholesterol was determined through enzymatic analyses with dextran sulfate, polyethylene glycol-modified cholesterol esterase, and cholesterol oxidase, to generate a peroxide that was measured colorimetrically. Total cholesterol and triglyceride levels were determined by standard enzymatic techniques. LDL cholesterol was quantified by the Friedewald formula, except when the triglyceride level was above 400 mg/dL, in which case LDL cholesterol levels were measured by direct beta quantification. Lp(a) was measured by immunoturbidimetry (Randox, Parramata, Australia) and PCSK9 measured by enzyme-linked immunosorbent assay (Cyclex, Nagano, Japan). Homeostasis model assessment of insulin resistance (HOMA-IR) was calculated as follows: glucose (mg/dL) \times insulin (pmol/L) \div $7.175 \times 405.$

Statistical analyses

Comparisons of continuous variables were conducted using the 2-sided Wilcoxon rank-sum test. A chi-square test was used to compare categorical variables. Signed-rank tests were used to compare baseline and 3-month PCSK9 and Lp(a) levels.

Results

Plasma PCSK9 and Lp(a) levels were measured at baseline (at the end of the run-in period) and 3 months after randomization in a subset of 594 patients receiving torcetrapib and atorvastatin and in 1151 patients receiving placebo and atorvastatin. The characteristics of the study participants at baseline are presented in Table 1.

At baseline, that is, after the run-in period, both plasma PCSK9 (Fig. 1) and Lp(a) levels (Fig. 2) were dosedependently elevated with increasing atorvastatin doses. Compared with patients without T2D, those with T2D at baseline had higher PCSK9 (357 ± 123 vs 338 ± 115 ng/mL, P = .0012) and lower Lp(a) levels $(28 \pm 32 \text{ vs } 32 \pm 33 \text{ mg/dL}, P = .0005)$. Regardless of T2D status, higher atorvastatin doses were associated with higher PCSK9 levels (Fig. 1). Similar to PCSK9 levels, higher atorvastatin doses were associated with higher Lp(a) levels (Fig. 2). Spearman correlation coefficients between PCSK9 and Lp(a) levels with parameters of lipid metabolism and glucose homeostasis are presented in Table 2. Despite statin therapy, which increases PCSK9^{3,4} and reduces total and LDL cholesterol levels, circulating PCSK9 levels were positively associated with total cholesterol, LDL cholesterol, triglycerides, and apoB as well as with HDL cholesterol and apoA-I levels, albeit to a lower extent. Plasma PCSK9 levels were also positively associated with fasting blood glucose, insulin, and HOMA-IR. Circulating Lp(a) levels were positively

Table 1	Baseline characteristics	of study	participants of	classified on t	the basis of	^t treatment group

	Atorvastatin and placebo, N $=$ 1151	Atorvastatin and torcetrapib, N = 594 $$	P value
Male gender, %	83	85	.24
Diabetes mellitus, %	48	50	.42
Age, y	62 ± 7.5	62 ± 7.6	.75
Total cholesterol, mg/dL	153 ± 26	153 ± 27	.99
LDL cholesterol, mg/dL	77 ± 20	77 ± 21	.79
HDL cholesterol, mg/dL	46 ± 11	47 ± 12	.49
Triglycerides, mg/dL	151 ± 75	146 \pm 78	.02
Apolipoprotein B, mg/dL	72 ± 15	71 ± 16	.06
Apolipoprotein A-I, mg/dL	123 ± 22	122 ± 22	.06
Glucose, mg/dL	116 \pm 36	118 ± 41	.95
Insulin, pmol/L	118 ± 135	117 \pm 109	.88
HOMA-IR	5.02 ± 8.09	5.04 ± 5.80	.78
Lipoprotein(a), mg/dL	30 ± 33	31 ± 33	.99
PCSK9, ng/mL	350 ± 120	340 ± 117	.11

HDL, high-density lipoprotein; HOMA-IR, homeostasis model assessment of insulin resistance; LDL, low-density lipoprotein; PCSK9, proprotein convertase subtilisin/kexin type 9.

Data are presented as percentage or mean \pm standard deviation.

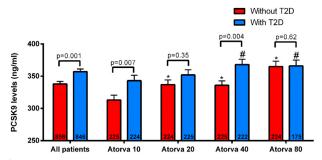


Figure 1 Plasma PCSK9 levels at baseline in patients with or without type II diabetes in the entire study sample and in patients separated on the basis of atorvastatin dose. *Significantly different from atorvastatin 10 mg group in patients without type II diabetes. *Significantly different from atorvastatin 10 mg group in patients with type II diabetes.

associated with LDL cholesterol, and to a much lower extent with total cholesterol, apoB, and HDL cholesterol. There was a weak but significant negative association between Lp(a) and triglycerides, fasting blood glucose, insulin, and HOMA-IR (Table 2). There was no significant correlation between PCSK9 and Lp(a) plasma levels.

We also measured PCSK9 and Lp(a) levels after 3 months of treatment either with placebo or with torcetrapib on top of atorvastatin doses established during the run-in period. Plasma PCSK9 levels significantly increased in patients treated with torcetrapib $(+13.1 \pm 125.3 \text{ ng/mL} [ie, +3.7\%], P = .005)$, but not in patients treated with placebo (+2.6 \pm 127.9 ng/mL [ie, +0.7%], P = .39). Increases in PCSK9 levels with torcetrapib were statistically significant in patients with T2D only (Fig. 3). In contrast, plasma Lp(a) levels significantly with decreased in patients treated torcetrapib $(-3.4 \pm 10.7 \text{ mg/dL} [-11.1\%], P < .0001)$, but not in patients treated with placebo (+0.3 \pm 9.4 mg/dL [+1%], P = .92). Decreases in Lp(a) were observed in patients with and without T2D (Fig. 4). Changes in PCSK9 were not associated with changes in Lp(a) levels in patients treated with torcetrapib (Spearman correlation coefficient r = -0.03, P = .21).

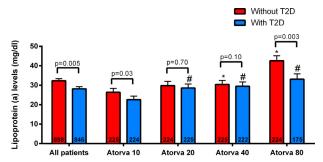


Figure 2 Plasma Lp(a) levels at baseline in patients with or without type II diabetes in the entire study sample and in patients separated on the basis of atorvastatin dose. *Significantly different from atorvastatin 10 mg group in patients without type II diabetes. #Significantly different from atorvastatin 10 mg group in patients with type II diabetes.

Table 2Spearman correlation coefficients between PCSK9and lipoprotein(a) levels and parameters of lipid metabolism orof glucose homeostasis

	PCSK9	Lipoprotein(a)
Total cholesterol	0.25 (<i>P</i> < .0001)	0.11 (<i>P</i> < .0001)
LDL cholesterol	0.15 (<i>P</i> < .0001)	0.21 (<i>P</i> < .0001)
HDL cholesterol	$0.08 \ (P = .0007)$	0.05 (P = .03)
Triglycerides	0.16 (<i>P</i> < .0001)	$-0.11 \ (P < .0001)$
Apolipoprotein B	0.22 (<i>P</i> < .0001)	$0.06 \ (P = .01)$
Apolipoprotein A-I	0.15 (<i>P</i> < .0001)	$0.01 \ (P = .63)$
Lipoprotein(a)	$0.03 \ (P = .38)$	-
PCSK9	-	$0.03 \ (P = .38)$
Glucose	0.14 (<i>P</i> < .0001)	-0.05 (P = .04)
Insulin	0.17 (<i>P</i> < .0001)	-0.05 (P = .05)
HOMA-IR	0.18 (<i>P</i> < .0001)	$-0.06 \ (P = .01)$

HDL, high-density lipoprotein; HOMA-IR, homeostasis model assessment of insulin resistance; LDL, low-density lipoprotein; PCSK9, proprotein convertase subtilisin/kexin type 9.

Discussion

In this sub-study of ILLUMINATE, a randomized clinical trial that documented the impact of CETP inhibition with torcetrapib on cardiovascular outcomes, we found that atorvastatin therapy was dose-dependently associated with higher PCSK9 and Lp(a) levels. The presence of T2D was also associated with higher PCSK9, but lower Lp(a) levels. We also showed that CETP inhibition with torce-trapib for 3 months slightly increased PCSK9 levels and decreased Lp(a) levels.

We found that atorvastatin therapy was dosedependently associated with higher PCSK9 levels. It is well established that all statins significantly increase circulating PCSK9 levels compared either with placebo or pretreatment values in humans (as reviewed by Sahebkar

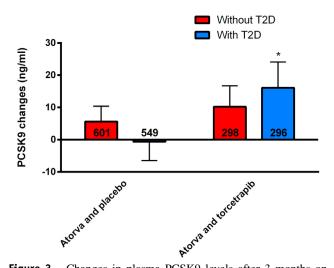


Figure 3 Changes in plasma PCSK9 levels after 3 months on placebo or torcetrapib in patients with and without type II diabetes. *Significantly different from baseline.

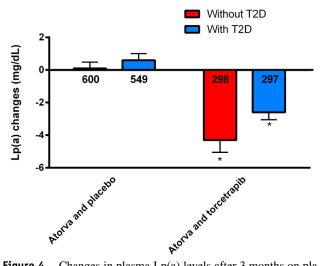


Figure 4 Changes in plasma Lp(a) levels after 3 months on placebo or torcetrapib in patients with and without type II diabetes. *Significantly different from baseline.

et al.¹⁹). Our study is the largest ever conducted on this topic and extends the results of a previous study on rosuvastatin⁴ by comprehensively documenting for the first time a dose response association between statin and PCSK9 levels. Compared with patients on 10 mg atorvastatin, those on the maximal 80 mg atorvastatin treatment dose had 37 ng/mL higher plasma PCSK9 levels. This increase in PCSK9 levels is of the same magnitude to that reported in patients on 10 mg atorvastatin vs placebo or pretreatment values (ie, +40 ng/mL).¹⁹ This observation underlines that the effects of atorvastatin on circulating PCSK9 levels are not linearly proportional to the treatment dose effects.²⁰

We also observed in ILLUMINATE that higher atorvastatin doses were associated with elevated Lp(a) levels. The impact of statin therapy on Lp(a) levels is somewhat controversial. For instance, in the Aortic Stenosis Progression Observation: Measuring Effects of Rosuvastatin trial, 40 mg rosuvastatin promoted a 20% increase in circulating Lp(a) levels.¹⁷ In the Treating to New Targets trial, we have shown that uptitration of atorvastatin doses from 10 to 80 mg/d was associated with a small but significant increase in Lp(a).²¹ In the Justification for the Use of Statins in Prevention: an Intervention Trial Evaluating Rosuvastatin trial however, 20 mg rosuvastatin did not significantly increase Lp(a) levels compared with pretreatment values.² In contrast, both atorvastatin and simvastatin reduced Lp(a) in a smaller cohort of heterozygous familial hypercholesterolemic patients.²³ We therefore cannot rule out that ILLU-MINATE patients requiring higher doses of atorvastatin to achieve LDL cholesterol <100 mg/dL were those with higher untreated baseline Lp(a) levels because assays that measure LDL cholesterol also include the contribution of Lp(a) cholesterol.²⁴ In that respect, the single nucleotide polymorphism rs10455872, that is a strong predictor of plasma Lp(a) levels, is also the second most important genetic determinant of LDL cholesterol lowering with statins.²⁵

Next, we showed that patients with T2D had higher PCSK9 levels than patients without T2D, as recently reported in a smaller cohort.²⁶ However, PCSK9 levels were not different between patients with vs without T2D in another smaller study.²⁷ Important questions still remain regarding the directionality and causality of this association. In that respect, insulin has been shown to enhance PCSK9 expression, which may argue in favor of T2D causing elevations in PCSK9 levels.^{28,29} This observation is also supported by the recent publication of 2 large-scale genetic associations studies that have shown that loss-of-function variants at the PCSK9 loci associated with impaired PCSK9 function may be associated with an increased risk of T2D.^{30,31}

In contrast, we found that Lp(a) levels were reduced in patients with T2D compared with patients without T2D. This result confirms previous studies showing lower Lp(a) levels in patients with T2D vs patients without T2D and extends the observations of 3 large prospective studies demonstrating a strong inverse association of Lp(a) levels with the risk of incident T2D (as reviewed by Tsimikas⁸). However, a genetic study challenged this association by showing that 4 single nucleotide polymorphism strongly correlated with the number of apo(a) kringle IV₂ repeats and Lp(a) levels did not predict T2D risk.³² A potential explanation for reduced Lp(a) levels in patients with T2D is that insulin reduces apo(a) synthesis in primary hepatocytes.³³ More likely, because T2D is characterized by increased levels of triglyceride-rich apoB-containing lipoprotein, which is (unlike LDL) a poor acceptor for apo(a), there is probably a reverse causality between Lp(a) levels and T2D. In line with this hypothesis, Lp(a)was significantly and negatively correlated with plasma triglycerides in the present study.

We also found that torcetrapib reduced Lp(a) by 11%. Noteworthy, more potent CETP inhibitors such as anacetrapib, evacetrapib, and TA-8995 provide 25% to 40% reductions in Lp(a) levels.^{34–36} Because CETP inhibition limits the net flux of triglycerides from triglyceride-rich lipoproteins to HDL and the net flux of cholesterol esters from HDL to triglyceride-rich lipoproteins particles, we speculate that fewer LDL-like particles might serve as substrate for Lp(a) assembly. In that respect, a recent genetic association study showed that genetic variants at the *LPA* locus linked with high Lp(a) levels were causally related with lower levels of circulating and triglyceride-rich lipoproteins particles. The directionality of this association clearly needs to be clarified.³⁷

Finally, we found that CETP inhibition with torcetrapib provides a small but significant increase in PCSK9 levels, in patients with T2D, but not in patients without T2D. The fact that torcetrapib was found to have serious off-target effects¹⁸ raises the possibility that the effects of torcetrapib observed in our study may have been unrelated to CETP inhibition. In contrast to our findings, other CETP inhibitors were shown to reduce PCSK9 expression in vitro, in mice and monkeys.^{38–41} Additional research will be needed to determine whether changes in PCSK9 levels result from a modulation of the canonical sterol regulatory elementbinding protein 2 transcription factor pathway or from off-target effects of torcetrapib.

In humans, the impact of CETP inhibition on PCSK9 appears to rely on background lipid-lowering therapy.⁴² In a study that included 39 mildly hypercholesterolemic patients, anacetrapib (100 mg/d) decreased PCSK9 levels by 18% in monotherapy but had no significant influence on PCSK9 levels (+3.4%) on top of atorvastatin 20 mg/d. Interestingly, plasma PCSK9 levels were weakly but positively associated with CETP activity in a cross-sectional study of 450 participants in a high-risk primary prevention setting.⁴³ Carriers of the PCSK9-R46L loss-of-function variant (linked with lower LDL cholesterol levels) also presented with lower CETP activity. This association has not been replicated in another study.⁴⁴ Thus, targeting CETP appears to influence PCSK9. Whether targeting PCSK9 also influences CETP activity will require confirmation.

We conclude that (1) both PCSK9 and Lp(a) levels are positively and dose-dependently increased by atorvastatin. The mechanisms by which statins increase PCSK9 levels are well known,^{3,4} but those by which higher atorvastatin doses may increase Lp(a) levels remain elusive. (2) The presence of T2D is associated with higher PCSK9, but lower Lp(a) levels. Whereas impaired apoB-containing lipoprotein remodeling in T2D and on CETP inhibition appears to be a plausible causative mechanisms for reduced Lp(a) in those patients, it is not clear as to why patients with T2D have higher PCSK9 levels and why CETP inhibition with torcetrapib increased PCSK9 levels in these patients. The physiological link(s) between glycemic parameters, PCSK9, LDL receptor function, and Lp(a) metabolism clearly needs to be addressed in future studies.

Acknowledgments

Authors' contributions: B.J.A. co-designed the statistical analysis plan and wrote the article. F.P. and F.T. conducted experiments. W.B. performed statistical analyses. G.K.H., S.M.B., S.R.-M., O.M., and K.-A.R. reviewed the article. D.D., D.D.W., J.J.P.K., and P.B. participated to the study design and reviewed the article. G.L. participated in study design, co-designed the statistical analysis plan, conducted experiments, and reviewed the article. All authors have approved the final article.

Financial disclosure

This study was sponsored by Pfizer. B.J.A. holds a junior scholar award from the Fonds de recherche du Québec: Santé (FRQS), has received consulting fees from Pfizer and research support from Pfizer and Ionis Pharmaceuticals. F.P. is an Amgen employee. W.B. and D.D. are Pfizer employees. G.K.H. is holder of a Vidi grant (016.156.445) from the Netherlands Organisation for Scientific Research (NWO), is supported by a grant from the CardioVascular Research Initiative (CVON2011-19; Genius) and the European Union (TransCard: FP7-603091-2). G.K.H. or his institution received honoraria for consultancy, advisory boards, and/or conduct of clinical trials from AMGEN, Aegerion, Pfizer, Astra Zeneca, Sanofi, Regeneron, KOWA, Ionis pharmaceuticals, and Cerenis. G.K.H. has received research support from Aegerion, AMGEN, Sanofi, Astra Zeneca, and Synageva. S.M.B. has received consulting fees from Pfizer. D.D.W. has received consulting fees and honoraria for lectures from Pfizer Inc and remuneration for serving on clinical trial committees from Sanofi Aventis, Regeneron, Resverlogix, and the Medicines Company. J.J.P.K. reports personal fees from Dezima, Cerenis, The Medicines Company, CSL, Behring, Amgen, Sanofi, Regeneron, Eli Lilly, Genzyme, Aegerion, Esperion, AstraZeneca, Omthera, Pronova, Vascular Biogenics, Boehringer Ingelheim, Catabasis, AtheroNova, UniQure, Novartis, Merck, Ionis Pharmaceuticals, and Kowa. P.B. is the chairman of the ILLUMINATE trial. He has received honoraria from Amgen, AstraZeneca, CSL-Behring, Lilly, Merck, Novartis, Pfizer, and Sanofi-Regerenon, has served on advisor boards for Amgen, AstraZeneca, CSL-Behring, Dezima, Lilly, Merck, Novartis, Pfizer, Sanofi-Regeneron, and has received research support from Merck and Pfizer. G.L. is the recipient of an Allocation de Recherche Chaire-Mixte (Inserm-Université de La Réunion) and a Programme de Recherche Hospitalière en Santé ANR-16-RHUS-0007. He has received research funding and honoraria from Affiris AG, Pfizer Inc, AMGEN, and Sanofi-Regeneron. F.T., S.R.-M., O.M., and K.-A.R. report no disclosure.

References

- 1. Baigent C, Keech A, Kearney PM, et al. Efficacy and safety of cholesterol-lowering treatment: prospective meta-analysis of data from 90,056 participants in 14 randomised trials of statins. *Lancet*. 2005;366:1267–1278.
- 2. Marais AD, Kim JB, Wasserman SM, Lambert G. PCSK9 inhibition in LDL cholesterol reduction: genetics and therapeutic implications of very low plasma lipoprotein levels. *Pharmacol Ther.* 2015;145: 58–66.
- **3.** Dubuc G, Chamberland A, Wassef H, et al. Statins upregulate PCSK9, the gene encoding the proprotein convertase neural apoptosis-regulated convertase-1 implicated in familial hypercholesterolemia. *Arterioscler Thromb Vasc Biol.* 2004;24:1454–1459.
- Awan Z, Seidah NG, Macfadyen JG, et al. Proprotein convertase subtilisin/kexin type 9 concentrations, and LDL cholesterol response: the JUPITER trial. *Clin Chem.* 2012;58:183–189.
- Robinson JG, Farnier M, Krempf M, et al, ODYSSEY LONG TERM Investigators. Efficacy and safety of alirocumab in reducing lipids and cardiovascular events. *N Engl J Med.* 2015;372:1489–1499.
- Sabatine MS, Giugliano RP, Wiviott SD, et al, Open-Label Study of Long-Term Evaluation against LDLCholesterol (OSLER) Investigators. Efficacy and safety of evolocumab in reducing lipids and cardiovascular events. *N Engl J Med.* 2015;372:1500–1509.

Journal of Clinical Lipidology, Vol 12, No 1, February 2018

- 7. Desai NR, Kohli P, Giugliano RP, et al. AMG145, a monoclonal antibody against proprotein convertase subtilisin kexin type 9, significantly reduces lipoprotein(a) in hypercholesterolemic patients receiving statin therapy: an analysis from the LDL-C assessment with proprotein convertase subtilisin kexin type 9 monoclonal antibody inhibition combined with statin therapy (LAPLACE)-thrombolysis in myocardial infarction (TIMI) 57 trial. *Circulation*. 2013;128: 962–969.
- Tsimikas S. Lipoprotein(a): novel target and emergence of novel therapies to lower cardiovascular disease risk. *Curr Opin Endocrinol Diabetes Obes*. 2016;23:157–164.
- Nordestgaard BG, Chapman MJ, Ray K, et al. Lipoprotein(a) as a cardiovascular risk factor: current status. *Eur Heart J.* 2010;31: 2844–2853.
- Clarke R, Peden JF, Hopewell JC, et al. Genetic variants associated with Lp(a) lipoprotein level and coronary disease. *N Engl J Med.* 2009;361:2518–2528.
- Thanassoulis G, Campbell CY, Owens DS, et al. Genetic associations with valvular calcification and aortic stenosis. *N Engl J Med.* 2013; 368:503–512.
- Arsenault BJ, Boekholdt SM, Dube MP, et al. Lipoprotein(a) levels, genotype, and incident aortic valve stenosis: a prospective Mendelian randomization study and replication in a case-control cohort. *Circ Cardiovasc Genet*. 2014;7:304–310.
- Kamstrup PR, Nordestgaard BG. Elevated lipoprotein(a) levels, LPA risk genotypes, and increased risk of heart failure in the general population. JACC Heart Fail. 2016;4:78–87.
- Kronenberg F, Utermann G. Lipoprotein(a): resurrected by genetics. J Intern Med. 2013;273:6–30.
- van Capelleveen JC, van der Valk FM, Stroes ES. Current therapies for lowering lipoprotein(a). J Lipid Res. 2016;57:1612–1618.
- Boerwinkle E, Leffert CC, Lin J, Lackner C, Chiesa G, Hobbs HH. Apolipoprotein(a) gene accounts for greater than 90% of the variation in plasma lipoprotein(a) concentrations. *J Clin Invest*. 1992;90:52–60.
- Capoulade R, Chan KL, Yeang C, et al. Oxidized phospholipids, lipoprotein(a), and progression of calcific aortic valve stenosis. *J Am Coll Cardiol.* 2015;66:1236–1246.
- Barter PJ, Caulfield M, Eriksson M, et al. Effects of torcetrapib in patients at high risk for coronary events. *N Engl J Med.* 2007;357: 2109–2122.
- Sahebkar A, Simental-Mendia LE, Guerrero-Romero F, Golledge J, Watts GF. Effect of statin therapy on plasma proprotein convertase subtilisin kexin 9 (PCSK9) concentrations: a systematic review and meta-analysis of clinical trials. *Diabetes Obes Metab.* 2015;17: 1042–1055.
- 20. Jones P, Kafonek S, Laurora I, Hunninghake D. Comparative dose efficacy study of atorvastatin versus simvastatin, pravastatin, lovastatin, and fluvastatin in patients with hypercholesterolemia (the CURVES study). *Am J Cardiol.* 1998;81:582–587.
- Arsenault BJ, Barter P, DeMicco DA, et al, Treating to New Targets (TNT) Investigators. Prediction of cardiovascular events in statintreated stable coronary patients of the treating to new targets randomized controlled trial by lipid and non-lipid biomarkers. *PLoS One*. 2014;9:e114519.
- 22. Khera AV, Everett BM, Caulfield MP, et al. Lipoprotein(a) concentrations, rosuvastatin therapy, and residual vascular risk: an analysis from the JUPITER trial (Justification for the Use of Statins in Prevention: an Intervention Trial Evaluating Rosuvastatin). *Circulation*. 2014;129: 635–642.
- van Wissen S, Smilde TJ, Trip MD, de Boo T, Kastelein JJ, Stalenhoef AF. Long term statin treatment reduces lipoprotein(a) concentrations in heterozygous familial hypercholesterolaemia. *Heart*. 2003;89:893–896.
- 24. Yeang C, Witztum JL, Tsimikas S. 'LDL-C' = LDL-C + Lp(a)-C: implications of achieved ultra-low LDL-C levels in the proprotein convertase subtilisin/kexin type 9 era of potent LDL-C lowering. *Curr Opin Lipidol*. 2015;26:169–178.

- Postmus I, Trompet S, Deshmukh HA, et al. Pharmacogenetic metaanalysis of genome-wide association studies of LDL cholesterol response to statins. *Nat Commun.* 2014;5:5068.
- 26. Ibarretxe D, Girona J, Plana N, et al. Circulating PCSK9 in patients with type 2 diabetes and related metabolic disorders. *Clin Investig Arterioscler*. 2016;28:71–78.
- 27. Brouwers MC, Troutt JS, van Greevenbroek MM, et al. Plasma proprotein convertase subtilisin kexin type 9 is not altered in subjects with impaired glucose metabolism and type 2 diabetes mellitus, but its relationship with non-HDL cholesterol and apolipoprotein B may be modified by type 2 diabetes mellitus: the CODAM study. *Atherosclerosis*. 2011;217:263–267.
- Costet P, Cariou B, Lambert G, et al. Hepatic PCSK9 expression is regulated by nutritional status via insulin and sterol regulatory element-binding protein 1c. *J Biol Chem.* 2006;281:6211–6218.
- Miao J, Manthena PV, Haas ME, et al. Role of insulin in the regulation of proprotein convertase subtilisin/kexin type 9. *Arterioscler Thromb Vasc Biol.* 2015;35:1589–1596.
- Ference BA, Robinson JG, Brook RD, et al. Variation in PCSK9 and HMGCR and risk of cardiovascular disease and diabetes. N Engl J Med. 2016;375:2144–2153.
- Schmidt AF, Swerdlow DI, Holmes MV, et al. PCSK9 genetic variants and risk of type 2 diabetes: a Mendelian randomisation study. *Lancet Diabetes Endocrinol*. 2017;5:97–105.
- Emdin CA, Khera AV, Natarajan P, et al. Phenotypic characterization of genetically lowered human lipoprotein(a) levels. *J Am Coll Cardiol*. 2016;68:2761–2772.
- Neele DM, de Wit EC, Princen HM. Insulin suppresses apolipoprotein(a) synthesis by primary cultures of cynomolgus monkey hepatocytes. *Diabetologia*. 1999;42:41–44.
- 34. Cannon CP, Shah S, Dansky HM, et al. Safety of anacetrapib in patients with or at high risk for coronary heart disease. *N Engl J Med.* 2010;363:2406–2415.
- 35. Nicholls SJ, Ruotolo G, Brewer HB, et al. Evacetrapib alone or in combination with statins lowers lipoprotein(a) and total and small LDL particle concentrations in mildly hypercholesterolemic patients. *J Clin Lipidol.* 2016;10:519–527.e4.
- 36. Hovingh GK, Kastelein JJ, van Deventer SJ, et al. Cholesterol ester transfer protein inhibition by TA-8995 in patients with mild dyslipidaemia (TULIP): a randomised, double-blind, placebo-controlled phase 2 trial. *Lancet*. 2015;386:452–460.
- Kettunen J, Demirkan A, Wurtz P, et al. Genome-wide study for circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA. *Nat Commun.* 2016;7:11122.
- Dong B, Singh AB, Fung C, Kan K, Liu J. CETP inhibitors downregulate hepatic LDL receptor and PCSK9 expression in vitro and in vivo through a SREBP2 dependent mechanism. *Atherosclerosis*. 2014;235:449–462.
- Miyosawa K, Watanabe Y, Murakami K, et al. New CETP inhibitor K-312 reduces PCSK9 expression: a potential effect on LDL cholesterol metabolism. *Am J Physiol Endocrinol Metab.* 2015;309:E177–E190.
- 40. van der Tuin SJ, Kuhnast S, Berbee JF, et al. Anacetrapib reduces (V) LDL cholesterol by inhibition of CETP activity and reduction of plasma PCSK9. J Lipid Res. 2015;56:2085–2093.
- **41.** Roddy TP, McLaren DG, Chen Y, et al. Effects of anacetrapib on plasma lipids, apolipoproteins and PCSK9 in healthy, lean rhesus macaques. *Eur J Pharmacol.* 2014;740:410–416.
- 42. Millar JS, Reyes-Soffer G, Jumes P, et al. Anacetrapib lowers LDL by increasing ApoB clearance in mildly hypercholesterolemic subjects. *J Clin Invest*. 2015;125:2510–2522.
- **43.** Girona J, Ibarretxe D, Plana N, et al. Circulating PCSK9 levels and CETP plasma activity are independently associated in patients with metabolic diseases. *Cardiovasc Diabetol.* 2016;15:107.
- 44. Verbeek R, Boyer M, Boekholdt SM, et al. Carriers of the PCSK9 R46L variant are characterized by an antiatherogenic lipoprotein profile assessed by nuclear magnetic resonance spectroscopy—Brief report. Arterioscler Thromb Vasc Biol. 2017;37:43–48.