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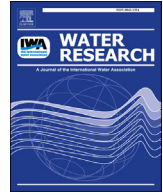
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Coupling stable isotopes and water chemistry to assess the role of hydrological and biogeochemical processes on riverine nitrogen sources



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ABSTRACT

Accurate source identification is critical for optimizing water pollution control strategies. Although the dual stable isotope ($^{15}\text{N}\text{-NO}_3/^{18}\text{O}\text{-NO}_3$) approach has been widely applied for differentiating riverine nitrogen (N) sources, the relatively short-term (<1 yr) $^{15}\text{N}\text{-NO}_3/^{18}\text{O}\text{-NO}_3$ records typically used in previous studies often hinders rigorous assessment due to high temporal variability associated with watershed N dynamics. Estimated contributions of legacy N sources in soils and groundwater to riverine N export by modeling approaches in many previous studies also lack validation from complementary information, such as multiple stable isotopes. This study integrated three years of multiple stable isotope ($^{15}\text{N}\text{-NO}_3/^{18}\text{O}\text{-NO}_3$ and $^2\text{H}\text{-H}_2\text{O}/^{18}\text{O}\text{-H}_2\text{O}$) and hydrochemistry measurements for river water, groundwater and rainfall to elucidate N dynamics and sources in the Yongan watershed (2474 km²) of eastern China. Nonpoint source N pollution dominated and displayed considerable seasonal and spatial variability in N forms and concentrations. Information from $\delta^{15}\text{N}\text{-NO}_3$ and $\delta^{18}\text{O}\text{-NO}_3$ indicated that riverine N dynamics were regulated by contributing sources, nitrification and denitrification, as well as hydrological processes. For the three examined catchments and entire watershed, slow subsurface and groundwater flows accounted for >75% of river discharge and were likely the major hydrological pathways for N delivery to the river. Riverine NO_3^- sources varied with dominant land use ($p < 0.001$), with the highest contributions of groundwater (60%), wastewater (35%), and soil (50%) occurring in agricultural, residential and forest catchments, respectively. For the entire watershed, groundwater (~50%) and soil N (>30%) were the dominant riverine NO_3^- sources, implying considerable potential for N pollution legacy effects. Results were consistent with observed nitrous oxide dynamics and N sources identified in previous modeling studies. As the first attempt to apply multiple isotope tracers for exploring and quantifying N transformation and transport pathways, this study provides an integrated approach for verifying and understanding the N pollution legacy effects observed in many watersheds worldwide. This study highlights that river N pollution control in many watersheds requires particular attention to groundwater restoration and soil N management in addition to N input control strategies.

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1. Introduction

Increasing anthropogenic nitrogen (N) inputs have resulted in water quality impairments, degradation of aquatic ecosystem

health, eutrophication and hypoxia in aquatic ecosystems worldwide (Galloway et al., 2004). Accurate identification and quantification of riverine N sources are critical for optimizing pollution control strategies at the watershed scale. Various lumped watershed models (e.g., export coefficient models, SPARROW, and Pol-Flow) and mechanistic models (e.g., AGNPS, HSPF, and SWAT) are available for estimating riverine N sources (Borah and Bera, 2004; Wellen et al., 2015). However, riverine N source apportionment remains a challenge, especially because of insufficient data. For

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example, mechanistic models typically require large amounts of data for calibration and validation, resulting in difficulties for application in many watersheds (Borah and Bera, 2004; Chen et al., 2014b). Lumped models, which usually provide a simplified understanding of N sources and transfer dynamics from the watershed to rivers, also require information on many watershed attributes, as well as N discharge from sewage treatment facilities and industries. In practice, current models are usually calibrated to measured river discharge and N loads at the watershed outlet, hindering assessment of how realistic these models simulate hydrological and biogeochemical processes throughout various subcatchments within a watershed (Wellen et al., 2015). Thus, watershed-scale simulation modeling has many challenges for practical application, especially for elucidating and validating specific sources and processes.

The dual nitrate stable isotope ($^{15}\text{N-NO}_3/^{18}\text{O-NO}_3$) approach provides an efficient alternative/complementary tool for differentiating riverine N sources, since each N source often has an unique isotopic signature (Kendall et al., 2007; Xue et al., 2009). Transformation processes (e.g., nitrification and denitrification) associated with N delivery from sources to receiving waters are also recorded by nitrate (NO_3^-) isotopic composition (Kendall et al., 2007; Xue et al., 2009; Xia et al., 2017). The dual NO_3^- stable isotope approach (usually coupled with Bayesian statistics, Parnell et al., 2010) has been successfully applied to assess N transformation processes and sources (e.g., atmospheric deposition, soil pools, chemical fertilizers, manure/sewage) in surface waters and groundwater. For example, the denitrification process in groundwater (Xue et al., 2009; Ji et al., 2017; Minet et al., 2017) and river water (Kaushal et al., 2011; Xia et al., 2017; Yi et al., 2017) was quantified using dual NO_3^- stable isotope data. Furthermore, the dual stable isotope approach was applied to quantify the major contributors of riverine NO_3^- in forest (Ohle, 2013), agricultural, sub-urban (Yang et al., 2013; Ji et al., 2017; Xia et al., 2017) and urban (Yang and Toor, 2016) catchments. In the majority of these studies, dual isotope records were collected for a short-term period (<1 yr), which hinders rigorous assessment of watershed N dynamics and sources given the large temporal variability in watershed processes (Yi et al., 2017; Yang et al., 2013). Additionally, few studies have attempted to verify dual NO_3^- stable isotope results by comparing to hydrological and biogeochemical information from other isotopes and hydrochemistry (Xue et al., 2009; Abbott et al., 2016; Minet et al., 2017; Yi et al., 2017).

Increasing evidence indicates that N leaching from legacy sources in soils and groundwater is a major cause for increasing riverine N exports despite significant declines in anthropogenic N inputs (Meals et al., 2010; Hamilton, 2012; Van Meter et al., 2017; Chen et al., 2018). Such legacy effects are related to the lag time (ranging from months to decades) elapsed between watershed N inputs and riverine export as regulated by hydrological and biogeochemical processes (Howden et al., 2011; Chen et al., 2018; Van Meter et al., 2018). However, the legacy effect is not rigorously addressed and formulated in most current watershed models (e.g., SWAT, HSPF, and AGNPS) due to limited knowledge concerning hydrological and biogeochemical mechanisms affecting the fate and transport of N while passing through soil, vadose zone, and groundwater to the river network (Meals et al., 2010; Covino, 2017; Moatar et al., 2017; Chen et al., 2018; Thomas and Abbott, 2018). Groundwater/baseflow N contributions to riverine N export have been estimated by combining hydrological models and water chemistry monitoring data or regression models for estimating riverine N loads (Table 1). However, these estimates are subject to uncertainties associated with use of empirical relationships in hydrological models (He and Lu, 2016) and lack of validation from complementary information, such as multiple stable isotopes.

Quantitative information concerning legacy N contributions from soils and groundwater to riverine N loads is critical for optimizing pollution control strategies (Sanford and Pope, 2013; Bouraoui and Grizzetti, 2014). Furthermore, isotope signals for legacy N in soils and groundwater are mixed and overlapping due to different N sources and cycling processes over time. This could introduce considerable uncertainty in apportioning riverine N to conventional sources (e.g., atmospheric deposition, soils, chemical fertilizers, and manure/sewage) by the dual stable isotope approach alone.

To overcome limitations associated with previous applications of the dual stable isotope approach, as well as previous estimates of legacy N contributions, this study addressed watershed N sources and cycling over a 3-year period using monthly records of multiple isotopes ($^{15}\text{N-NO}_3/^{18}\text{O-NO}_3$ and $^2\text{H-H}_2\text{O}/^{18}\text{O-H}_2\text{O}$) and hydrochemistry for river water, groundwater and rainfall at multiple sites in the Yongan River watershed of eastern China. Major objectives of this integrated analysis were to (1) reveal spatial and seasonal variations of riverine N dynamics; (2) assess roles of hydrological and biogeochemical processes on riverine N dynamics; and (3) apportion riverine N to soil, groundwater and wastewater sources. Results of this study provide a level of verification for previous studies examining legacy N contributions in the Yongan watershed, as well as improve our quantitative understanding of N dynamics and the N-leaching lag effect at the watershed scale. As the first attempt to apply multiple isotope tracers for exploring and quantifying N transformation and transport pathways, this study provides an integrated approach for verifying and understanding N pollution legacy effects observed in many watersheds worldwide.

2. Material and methods

2.1. Study area

The Yongan River watershed is a typical rainfall-dominated watershed with predominantly forest and agricultural land use in eastern Zhejiang Province, China (Fig. 1). The river drains 2474 km² and has an average discharge of 72.9 m³ s⁻¹ at the downstream outlet sampling site (Fig. 1). Climate is subtropical monsoon with an average annual temperature and precipitation of 17.4 °C and 1400 mm, respectively. Rainfall mainly occurs in May–October (~67%) with a typhoon season in July–September. Agricultural land (including paddy field, garden plot and dryland) averaged ~12% of total watershed area in recent decades, with developed lands, forest, and barren lands contributing ~3, ~67, and ~18%, respectively (Table 2). Red (Oxisols), yellow (Ultisols), and lithological (Entisols) soil groups are the dominant soil types and accounted for 64.6, 15.4, and 1.5% of total soil area, respectively (Chen et al., 2014a). Daily water discharge for the downstream outlet site and daily precipitation and temperature (2014–2017) for the watershed were obtained from the local Hydrology Bureau and Weather Bureau, respectively. Land-use distribution and elevation were obtained from the local Land Resource Bureau by extracting from remote sensing images and digital elevation models and subsequent verification by field investigation. Additional information on Yongan watershed climate, hydrology and river networks can be found in Section 1 of Supporting Information (SI).

2.2. Field sampling

Water samples were collected on a monthly basis during 2014–2017 (Fig. 2) from upstream to downstream, including 12 sampling sites (4 mainstem sites, 5 major tributaries, and 3 groundwater sites) (Fig. 1). Surface water samples were collected at 3–5 points across the river channel and composited to obtain a

Table 1
Baseflow contributions of water and nitrogen to riverine export in reported references.

| Study region | Water contribution (%) | N contribution (%) | Methods | Reference |
|--|------------------------|--------------------|--|----------------------------|
| Lake Michigan, USA | 63 | 38 | Hydrograph separation approaches (e.g., HYSEP1, HYSEP2, HYSEP3, UKIH, PART, BFLOW) for baseflow separation; Two-dimensional, finite-element, transport model SUTRA for N load estimation | Cherkauer et al. (1992) |
| MCR watershed, north of Harrisburg, USA | 56 | 56 | Water chemistry monitoring during baseflow conditions and storm events | Pionke et al. (1996) |
| A monitoring station site in Daejeon, Korea | 61 | 60 | Hydrograph separation program (PULSE) combined with water chemistry monitoring | Kim et al. (2010) |
| The Shaying river basin, China | 39 | 40 | Groundwater flow model (MODFLOW) combined with a numerical simulation software (MT3DMS) | Chen (2013) |
| Seven USGS sites in wide range of BFI values, USA | 12–84 | 19–97 | The “BFI Program” combined with LOADEST model | Tesoriero et al. (2013) |
| The Yongan River watershed, southeastern China | 82 | 67 | The revised Regional Nutrient Management model was used to estimate groundwater and N contribution | Hu et al. (2018) |
| Subwatershed of Mahantango Creek, Susquehanna River, USA | 64 | 65 | Hydrograph separation methods based on recession analysis combined with the FLUXMASTER and LOADEST model | Zhu et al. (2011) |
| Two rice agriculture catchment, subtropical central China | 28–42 | 27–37 | HYSEP and recursive digital filter methods combined with the ESTIMATOR program | Wang et al. (2015) |
| Raccoon River watershed, west central Iowa, USA | 54 | 67 | HYSEP and recursive digital filter methods combined with the ESTIMATOR | Schilling and Zhang (2004) |
| Sub-watersheds of the Capitol Region Watershed, Minnesota, USA | 27–66 | 31–68 | Baseflow separated from comparison by hydrographs to baseflow rates during known dry periods combined with water chemistry monitoring | Janke et al. (2014) |
| The Yongan River watershed, southeastern China | 79 | 76 | Stable isotopes of water ($^2\text{H-H}_2\text{O}/^{18}\text{O-H}_2\text{O}$) combined with N stable isotopes ($^{15}\text{N-NO}_3^-/^{18}\text{O-NO}_3^-$) | This study |

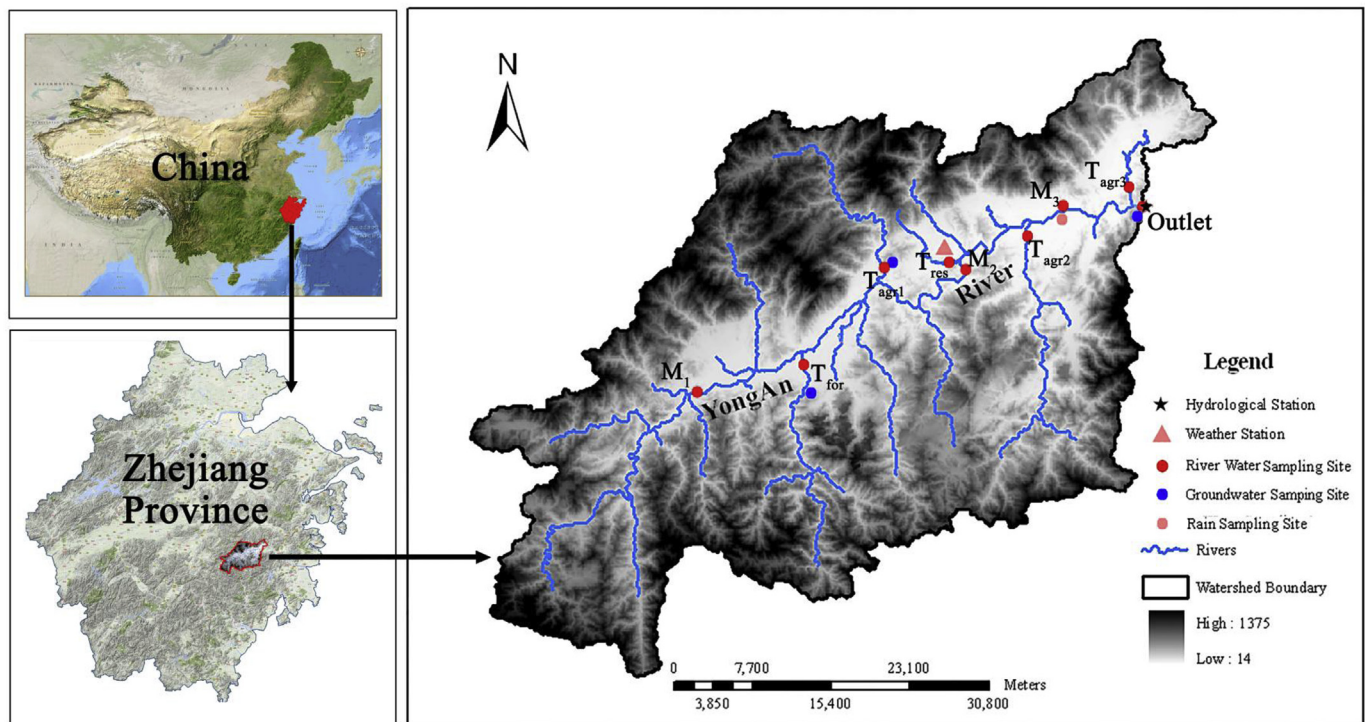


Fig. 1. Location of the Yongan watershed in China and Zhejiang Province and the sampling sites for river water, groundwater and rainfall. “M” represents mainstem, “T” represents tributary. Subscript “agr” represents agricultural tributary, “for” represents forest tributary and “res” represents residential tributary.

single depth-width integrated sample. The Weather Bureau collected and preserved (filtered + refrigeration) rainfall samples on an event basis to provide a monthly composite sample. The composite rainfall sample was a volume-weighted mixture of all rainfall events and was used for water chemistry and stable isotope analysis. Three wells were established in the agricultural tributary (GW_{agr} , 6 m depth), forest tributary (GW_{for} , 6 m depth), and at the

watershed outlet ($\text{GW}_{\text{outlet}}$, 15 m depth) (Fig. 1). Wells were located 100–200 m from the river channel and screened at depths of 3–6 m for GW_{agr} and GW_{for} and at depths of 5–8 m for $\text{GW}_{\text{outlet}}$. Groundwater samples were collected using a peristaltic pump to purge a minimum of 3 well volumes of water before collecting a representative sample for water quality analysis. All water samples were immediately sealed and stored at 4 °C.

Table 2
Land-use distribution for sub-catchments and the entire Yongan watershed.

| Catchments | Drainage area (km ²) | A (%) | D (%) | F (%) | N (%) | PF (%) | PD | AD |
|----------------------|----------------------------------|-------|-------|-------|-------|--------|-----|-----|
| Trib _{agr2} | 357 | 11 | 2 | 74 | 23 | 6% | 226 | 88 |
| Trib _{agr3} | 163 | 44 | 1 | 54 | 1 | 20 | 287 | 55 |
| Trib _{agr1} | 202 | 14 | 1 | 71 | 14 | 6 | 274 | 110 |
| Trib _{res} | 35 | 21 | 9 | 49 | 21 | 10 | 828 | 136 |
| Trib _{for} | 218 | 3 | 1 | 76 | 20 | 2 | 79 | 35 |
| Entire watershed | 2474 | 13 | 3 | 67 | 17 | 6 | 288 | 99 |

A - agricultural lands, D - developed lands, F - forest lands, N - non-utilized lands, PF → paddy field, PD → population density (capita km⁻²); AD → domestic animal density (capita km⁻²).

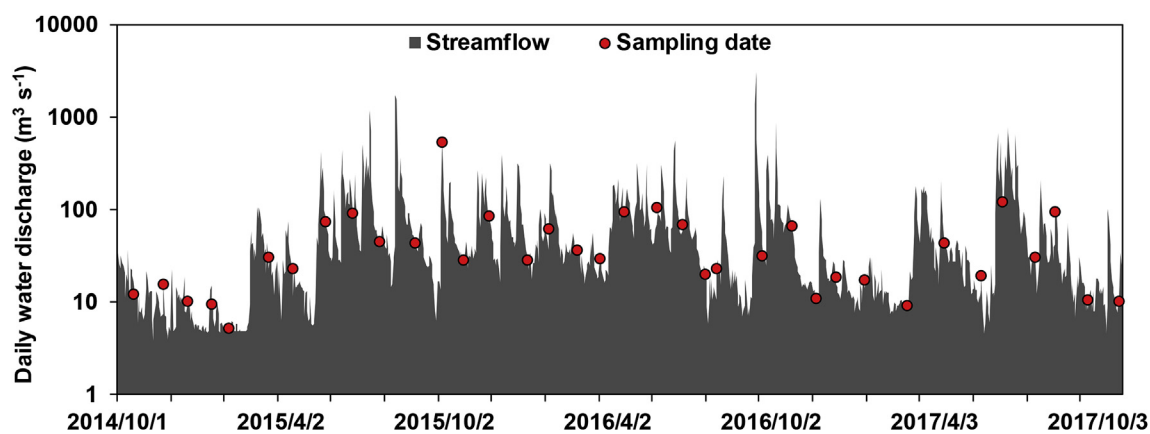


Fig. 2. Daily discharge and sampling dates in the Yongan watershed in 2014–2017.

2.3. Water quality and stable isotope analysis

Rainfall, groundwater and river water were analyzed for temperature (°C), pH, dissolved oxygen (DO), dissolved organic carbon (DOC), dissolved N₂O, total nitrogen (TN), total dissolved nitrogen, nitrate (NO₃⁻) and ammonium (NH₄⁺). Temperature, pH and DO were measured in the field using YSI electrodes (Xylem, New York, USA). All water samples were analyzed within 24 h; additional details on analytical methods are provided in Section 2 of SI.

Rainfall, groundwater and river water samples at the watershed outlet site were collected between October 2014 and October 2016 for isotopic analysis of δ¹⁸O-H₂O and δ²H-H₂O. Water isotopes (i.e., δ¹⁸O-H₂O and δ²H-H₂O) were determined using an isotope-ratio mass spectrometer (Delta V Advantage, Thermo Scientific). Two internal standards calibrated against VSMOW (Biscayne Aquifer Drinking Water (USGS45) and Lake Louise Drinking Water (USGS47)) were used to calibrate raw data and ensure long-term stability of analyses. The analytical precision was ±0.2‰ for δ¹⁸O-H₂O and ±1‰ for δ²H-H₂O.

Rainfall, groundwater and river water samples for δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻ stable isotope analysis were collected from all catchments. Samples were frozen at -20 °C, and subsequently transported to the Environmental Stable Isotope Lab (Institute of Soil Science, Chinese Academy of Sciences, Nanjing, China) for analysis. Water samples were pre-processed by the chemical method of McIlvin and Altabet (2005) (See Section 2 of SI) to convert NO₃⁻ to gaseous N₂O for detection using a continuous-flow isotope ratio mass spectrometer (IRAM, Isoprime100). Isotope ratio values are reported in parts per thousand (‰) relative to atmospheric N₂ and Vienna Standard Mean Ocean water (VSMOW) for δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻, respectively (Kendall et al., 2007). Sample analysis had an average precision of 0.31‰ for δ¹⁵N-NO₃⁻ and 0.55‰ for δ¹⁸O-NO₃⁻. The load-weighted mean (M_L) values for δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻ were used to remove the NO₃⁻ load variability and emphasize

biogeochemical influences (Voss et al., 2006). Although no groundwater sample was collected in the residential tributary (T_{res}), δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻ values were estimated based on observed significant relationships between stable isotope values and area percentages of agricultural and residential land use across the three groundwater sampling wells (for ¹⁵N-NO₃⁻, R² = 0.998, p < 0.05; for ¹⁸O-NO₃⁻, R² = 0.997, p < 0.05, See Section 2 of SI). A similar approach was adopted in previous studies (Voss et al., 2006; Xue et al., 2009).

The combined isotopic signals of δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻ were used to assess nitrification and denitrification processes. For denitrification, microbes utilize NO₃⁻ with lighter isotopes leading to enrichment of δ¹⁸O-NO₃⁻ and δ¹⁵N-NO₃⁻ at a ratio between 0.43 and 1.0 (Kendall et al., 2007; Xue et al., 2009; Ji et al., 2017). For nitrification, 2/3 of the O in the NO₃⁻ product is derived from ambient soil water and 1/3 from atmospheric O₂ (Kendall, 1998). As a result, the δ¹⁸O values of NO₃⁻ derived from nitrification should range between -1.92‰ and 7.44‰, based on a δ¹⁸O value of +23.5‰ for atmospheric O₂ and δ¹⁸O values of -14.63‰ to -0.59‰ for precipitation (Yang et al., 2013).

2.4. River water source identification based on stable isotopes of water

Stable isotopes of water (²H-H₂O and ¹⁸O-H₂O) were examined to identify river water sources and better understand hydrological processes. Physically based stable isotopes serve as effective tracers with isotope fractionation producing a natural labeling effect that provides information on hydro-climate processes (McGuire and McDonnell, 2006). The sine wave regression method was conducted to estimate the young water fractions (F_{yw}). F_{yw} is defined as the proportion of the transit-time distribution younger than a threshold age (-0.2 year) (Kirchner, 2016). We calculated F_{yw} using sine wave regressions on the δ¹⁸O (‰) time series (McGuire and

McDonnell, 2006) to determine the cosine and sine coefficients for precipitation and streamflow (See Section 3 of SI).

2.5. Riverine nitrogen source identification with stable isotope analysis in R (SIAR)

The Bayesian model (SIAR) was applied to estimate contributions of multiple NO_3^- sources in individual water samples (Parnell et al., 2010). Nitrogen pollution in rivers is mainly derived from point source (e.g., domestic, industry, and animal wastewaters) and non-point source (NPS) pollution pathways (Ongley et al., 2010). Driven by watershed hydrological processes, NPS pollution derived from various sources (e.g., chemical fertilizer, manure/sewage, and atmospheric deposition) will be transported to rivers through soil and groundwater (Griffiths et al., 2016). Due to hydrological and biogeochemical N legacy effects, it may take years to decades for some N sources to be transported to rivers through soil and groundwater in watersheds (Meals et al., 2010; Hamilton, 2012). Some long-term field experiments and watershed modeling results indicated that a very limited proportion (<2%) of annual N inputs was delivered to groundwater and surface water in the current year (Chen et al., 2018). Therefore, this study apportioned riverine N to groundwater, soil and wastewater to evaluate the efficacy of previous modeling results. Riverine N source apportionment analysis was conducted by SIAR at the outlet of Yongan watershed, agricultural tributary (T_{agr1}), forest tributary (T_{for}) and residential tributary (T_{res}). Groundwater and river water N isotope values were directly measured, while isotopic values for soil and wastewater N were obtained from published values for a nearby watershed (Ji et al., 2017). (See SIAR details in Section 3 of SI).

2.6. Statistical analysis

All correlation, regression and nonparametric analyses were performed using SPSS (version 17.0, SPSS, Chicago, IL, USA). One-way ANOVA with a LSD multiple comparisons test was used to examine differences in stable isotope values ($\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$) among sites. Nonparametric tests were conducted to compare N concentrations among sites, since these values did not follow a normal distribution according to the Kolmogorov–Smirnov test.

3. Results and discussion

3.1. Spatial and temporal patterns in N dynamics

Across mainstem ($n = 143$) and tributary ($n = 164$) river waters, NO_3^- (Mean \pm SD: $67 \pm 18\%$) was the major component of TN and was significantly higher than NH_4^+ ($15 \pm 12\%$) and organic N ($18 \pm 15\%$). This pattern indicates the widespread role of NPS pollution and highlights the importance of NO_3^- source identification (Ongley et al., 2010). Longitudinally, NO_3^- ($p < 0.001$) and NH_4^+ ($p < 0.001$) concentrations increased from upstream to downstream along the mainstem, with NO_3^- increasing by 60% and NH_4^+ by 300% (Fig. 3a), due to increasing N loads discharged from tributaries. Observed TN concentrations in tributaries dominated by residential and agricultural land use were significantly ($p < 0.01$) higher than those in the mainstem (Table 3), potentially caused by dilution effects. With respect to land-use, NO_3^- and NH_4^+ concentrations were significantly ($p < 0.001$) higher in tributaries dominated by residential and agricultural land use than those dominated by forests (Fig. 3c), which is consistent with previous studies in eastern and northern China (Sun et al., 2013; Yi et al., 2017).

There were no significant differences ($p > 0.05$) in NO_3^- concentrations among seasons in the mainstem; however, NH_4^+

concentrations were lower ($p < 0.05$) in summer (Fig. 3b). Similar seasonal patterns were observed in tributaries and groundwater (Fig. 3d and f). These results may be due to higher biological uptake and N transformation rates during the summer growing season (Sudduth et al., 2013; Griffiths et al., 2016). There was a significant positive relationship ($r = 0.25$, $p = 0.004$, $n = 131$) between water discharge and NO_3^- concentration in the mainstem, which became stronger ($r = 0.37$, $p < 0.001$, $n = 128$) when the three highest water discharge points (at least 2 times higher than other records) were removed (Fig. 4a). The trend of increasing NO_3^- concentration with increasing discharge further indicates the widespread role of NPS pollution (Lai et al., 2011). There was no significant relationship observed between water discharge and NO_3^- concentration in agricultural tributaries ($p = 0.296$, $n = 92$), which may suggest offsetting inputs of NPS N pollution load and dilution effects with increasing runoff/discharge (Fig. 4c). A significant negative relationship was found between water discharge and NO_3^- concentration in residential ($r = 0.25$, $p = 0.004$, $n = 131$) and forest ($r = 0.54$, $p = 0.01$, $n = 22$) tributaries (Fig. 4c), which may indicate a prominent dilution effect and less NPS pollution from residential and forest catchments (Kaushal et al., 2011). There were no significant differences in NH_4^+ concentrations between the low flow regime (30% lower bound of flow regime) and high flow regime (30% upper bound of flow regime) in mainstem ($p = 0.264$) or tributary ($p = 0.198$) sites (Fig. 4b and d, Table 3). This may be explained by enhanced biogeochemical processes during the low flow regime (Chen et al., 2013), which will lead to similarly low NH_4^+ concentrations to that in the high flow regime caused by dilution effects.

Observed N concentrations in groundwater samples were similar to those in river water (Fig. 3), suggesting the potential for relatively high groundwater contributions to rivers (Griffiths et al., 2016). As expected, NO_3^- concentrations in groundwater were significantly ($p < 0.001$) higher in catchments dominated by agriculture than at the watershed outlet and forest-dominated catchments. In contrast, NH_4^+ concentrations in groundwater were ~8–20 times lower than NO_3^- (Table 3). This is mainly due to cation exchange reactions in the soil, which limit NH_4^+ leaching mobility to groundwater as compared to NO_3^- (Swaney et al., 2012). Higher NH_4^+ concentrations in groundwater ($p < 0.05$) were observed at the outlet site and lower concentrations observed in catchments dominated by agriculture and forests (Fig. 3e). Comparing N concentrations between precipitation and river water (Fig. 3a and c), precipitation produced a dilution effect for NO_3^- while contributing an NH_4^+ source to rivers. Although no detailed spatial data were available in this study, high NH_4^+ concentrations in precipitation are generally derived from local fertilizer and manure volatilization (Kang et al., 2016) associated with excessive anthropogenic N inputs to lands in the Yongan watershed (Chen et al., 2016).

3.2. Investigating water and N cycling processes with stable isotopes

3.2.1. Water cycling characteristics and water source identification

Slopes of $\delta^2\text{H}-\text{H}_2\text{O}$ versus $\delta^{18}\text{O}-\text{H}_2\text{O}$ for river water (5.98) and groundwater (4.71) were lower than the local meteoric water line (8.23; Fig. 5). This result is associated with evapotranspiration, since ^{18}O will be more enriched than deuterium during evaporation (Gonfiantini, 1986), thereby increasing oxygen and hydrogen isotopic values generating a systematic deviation from LMWL (Sánchez-Murillo et al., 2015). The isotopic composition of river water (CV: 10.5–13.7% for $\delta^2\text{H}-\text{H}_2\text{O}$; 8.9–11.8% for $\delta^{18}\text{O}-\text{H}_2\text{O}$) was less variable than that of precipitation (CV: 91.4% for $\delta^2\text{H}-\text{H}_2\text{O}$; 53.4% for $\delta^{18}\text{O}-\text{H}_2\text{O}$), suggesting a substantial portion of river water may originate from groundwater and subsurface water sources. River water is a mixture of surface runoff, subsurface water and

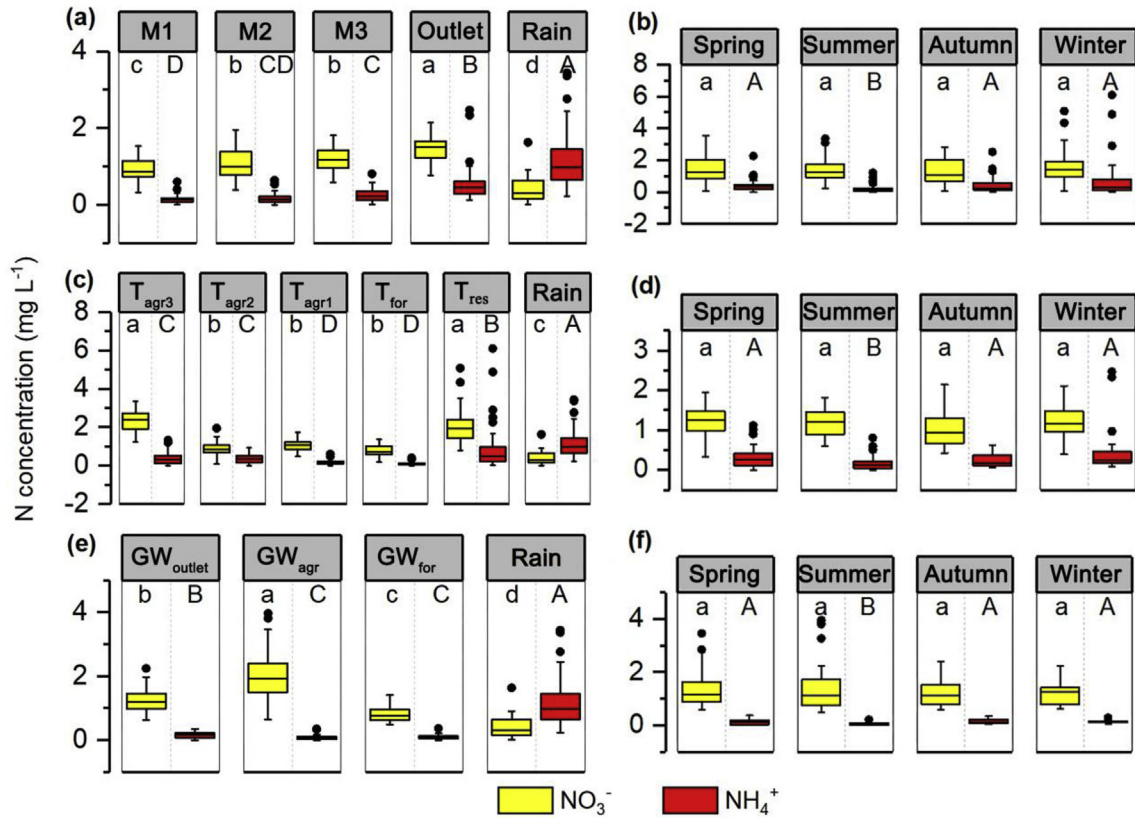


Fig. 3. Box plots for NO_3^- and NH_4^+ in (a) different mainstem sites and (b) in different seasons, (c) in different tributaries sites and (d) in different seasons, (e) in different groundwater sites, and (f) in different seasons. “M” represents the mainstem, “T” represents the tributary and “GW” represents groundwater. Subscript “agr” represents agricultural tributary, “for” represents forest tributary and “res” represents residential tributary. Box plots illustrate the 25th, 50th and 75th percentile; the whiskers indicate the 2.5th and 97.5th percentiles; and the points represent data outliers.

Table 3
Nitrogen concentrations and NO_3^- isotopic composition in the Yongan watershed during different flow regimes.

| | TN (mg N L^{-1}) | NO_3^- (mg N L^{-1}) | NH_4^+ (mg N L^{-1}) | $\delta^{15}\text{N-NO}_3^-$ (‰) | $\delta^{18}\text{O-NO}_3^-$ (‰) |
|---------------------------|-----------------------------|--|--|----------------------------------|----------------------------------|
| Low flow regime | | | | | |
| Mainstem | 1.75 ± 0.80 | 1.07 ± 0.38 | 0.35 ± 0.48 | 7.35 ^a | 0.85 ^a |
| Tributary _{agr} | 1.98 ± 1.39 | 1.20 ± 0.67 | 0.36 ± 0.35 | 8.27 ^a | 2.76 ^a |
| Tributary _{res} | 4.94 ± 3.09 | 2.78 ± 1.25 | 1.29 ± 1.33 | 9.03 ^a | 0.48 ^a |
| Tributary _{for} | 1.29 ± 0.35 | 0.99 ± 0.37 | 0.16 ± 0.14 | 2.25 ^a | -1.32 ^a |
| Medium flow regime | | | | | |
| Mainstem | 1.67 ± 0.55 | 1.23 ± 0.42 | 0.24 ± 0.26 | 9.29 ^a | 1.17 ^a |
| Tributary _{agr} | 1.92 ± 0.81 | 1.47 ± 0.85 | 0.27 ± 0.24 | 9.05 ^a | 2.22 ^a |
| Tributary _{res} | 3.62 ± 2.00 | 2.15 ± 0.71 | 1.08 ± 1.70 | 12.66 ^a | 1.03 ^a |
| Tributary _{for} | 1.00 ± 0.22 | 0.68 ± 0.28 | 0.07 ± 0.05 | 4.54 ^a | 1.49 ^a |
| High flow regime | | | | | |
| Mainstem | 1.93 ± 1.58 | 1.18 ± 0.33 | 0.27 ± 0.19 | 6.00 ^a | 0.81 ^a |
| Tributary _{agr} | 2.38 ± 1.03 | 1.43 ± 0.71 | 0.33 ± 0.25 | 6.00 ^a | 1.15 ^a |
| Tributary _{res} | 2.69 ± 1.21 | 1.51 ± 0.32 | 0.56 ± 0.69 | 7.94 ^a | 0.28 ^a |
| Tributary _{for} | 1.20 ± 0.37 | 0.70 ± 0.27 | 0.11 ± 0.09 | 4.88 ^a | -0.76 ^a |
| Whole period | | | | | |
| Mainstem | 1.78 ± 0.65 | 1.33 ± 0.70 | 0.30 ± 0.53 | 6.91 ^a | 0.90 ^a |
| Tributary _{agr} | 2.09 ± 1.07 | 1.39 ± 0.77 | 0.31 ± 0.27 | 6.87 ^a | 1.49 ^a |
| Tributary _{res} | 3.74 ± 2.74 | 2.11 ± 0.94 | 0.44 ± 0.75 | 9.36 ^a | 0.51 ^a |
| Tributary _{for} | 1.34 ± 0.32 | 0.77 ± 0.32 | 0.12 ± 0.10 | 4.64 ^a | -0.25 ^a |
| GW _{outlet} | 1.69 ± 0.48 | 1.26 ± 0.41 | 0.15 ± 0.10 | 9.89 ± 5.57 | 2.45 ± 2.92 |
| GW _{agr} | 2.37 ± 0.89 | 2.01 ± 0.81 | 0.09 ± 0.08 | 8.16 ± 3.64 | 1.38 ± 2.73 |
| GW _{for} | 1.19 ± 0.30 | 0.80 ± 0.22 | 0.09 ± 0.08 | 5.97 ± 3.75 | 0.85 ± 3.68 |

Superscript “a” denotes the load-weighted mean (M_L) values for $\delta^{15}\text{N-NO}_3^-$ and $\delta^{18}\text{O-NO}_3^-$.

groundwater inputs having different ages (Howden et al., 2011). The overlapping water isotopic compositions between river water and groundwater (Fig. 5) indicate that groundwater (having a

relatively older age) is an appreciable source of river water (Ji et al., 2017). Estimated baseflow (i.e., groundwater and slow subsurface flow) contributions based on the young water fractions (F_{yw}) for the

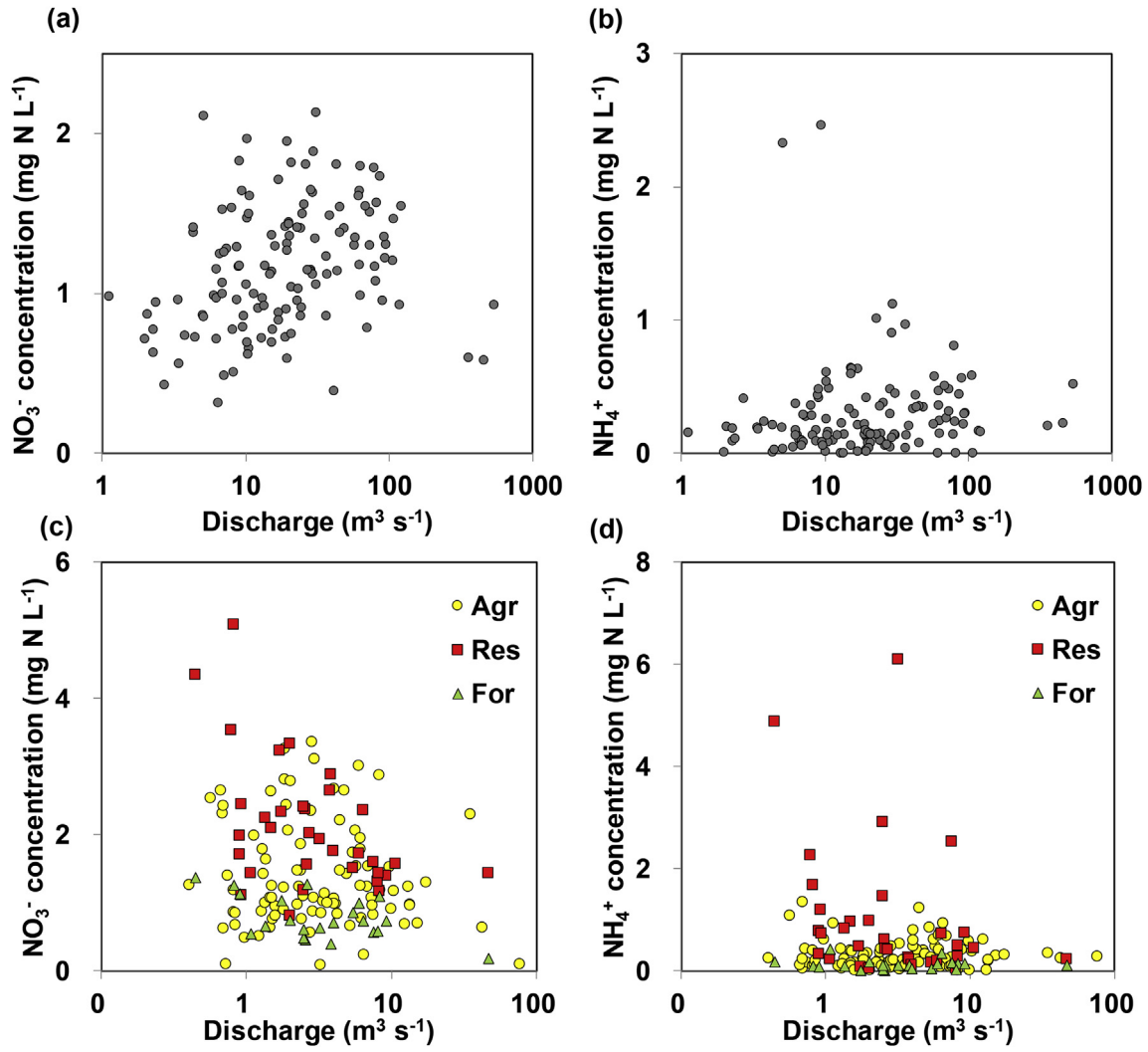


Fig. 4. Scatter plots for (a) NO_3^- and (b) NH_4^+ in mainstem, (c) NO_3^- and (d) NH_4^+ in agricultural (agr), residential (res) and forest (for) tributaries.

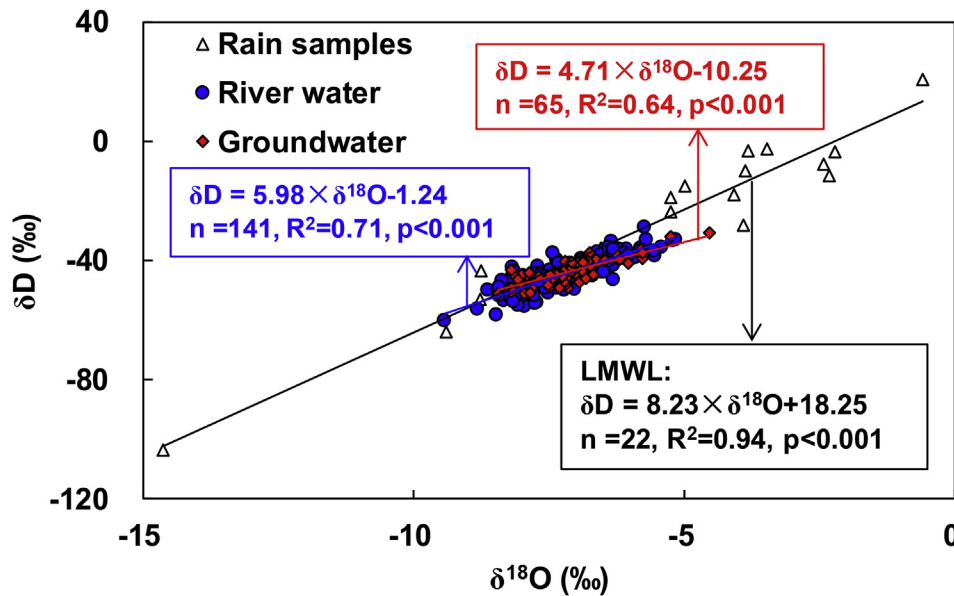


Fig. 5. Scatter plots for $^2\text{H}\text{-H}_2\text{O}$ and $^{18}\text{O}\text{-H}_2\text{O}$ in river water, groundwater and rainfall samples. Local meteoric water line (LMWL) is a regression line of measured stable isotopic composition from local rainfall samples.

sub-catchments and the entire Yongan watershed ranged from 79 to 85% of total streamflow (Fig. 8d). Such a high water contribution from baseflow to streamflow is consistent with ReNuMa model results for the Yongan watershed and has been commonly observed in many watersheds worldwide (Table 1). However, the use of watershed hydrological models in previous studies to quantify the fraction of different streamflow components has many challenges resulting from the high level of uncertainty associated with inherent limitations and assumptions of these methods (He and Lu, 2016). Instead, the F_{yw} method can accurately estimate streamflow components from the amplitude ratio of tracer cycles in precipitation and streamflow with a reported precision of a few percent (McGuire and McDonnell, 2006; Kirchner, 2016). Subsurface hydrological processes, with a particular focus on groundwater, could lead to a considerable lag effect in delivery of watershed nutrients to rivers (Meals et al., 2010). The lag effect from groundwater transport of NO_3^- derived years or decades ago could lead to an appreciable delay in the response of watershed N management activities (Chen et al., 2018; Van Meter et al., 2018).

3.2.2. N cycling process identification using NO_3^- stable isotopes

The ranges of $\delta^{18}\text{O}-\text{NO}_3^-$ and $\delta^{15}\text{N}-\text{NO}_3^-$ signatures between groundwater and river water were overlapping (Fig. 6), suggesting that groundwater may be a source of surface water NO_3^- (Griffiths et al., 2016). The $\delta^{18}\text{O}-\text{NO}_3^-$ and $\delta^{15}\text{N}-\text{NO}_3^-$ source signatures indicated (Fig. 6d) that direct N inputs from soil and chemical fertilizer were potentially limited in agricultural tributaries, but soil N was a potentially major N source in the forest tributary. In contrast, wastewater was the potentially dominant N contributor in the residential tributary (Fig. 6d). These results are similar to previous studies and emphasize human impacts on N dynamics (Xia et al., 2017; Yi et al., 2017). Greater than 80% of river water (including mainstem and tributaries) $\delta^{18}\text{O}$ values for NO_3^- (Fig. 6b and d) fall within the range expected for NO_3^- formed as a product of nitrification (Yang et al., 2013). Among different tributaries, $\delta^{15}\text{N}-\text{NO}_3^-$ values for residential land were significantly higher than those for agricultural ($p < 0.001$) and forested tributaries ($p = 0.002$) (Table 3). Estimated slopes between $\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}$ in the mainstem (slope = 0.23; $R^2 = 0.09$, $p = 0.001$, $n = 131$) and tributaries (slope = 0.24; $R^2 = 0.12$, $p < 0.001$, $n = 147$) were considerably lower than slopes expected for waters experiencing considerable denitrification (slopes between 0.43 and 1.0) (Xue et al., 2009; Ji et al., 2017; Xia et al., 2017). Although there was no significant decrease in N concentrations during spring and winter compared to other seasons (Fig. 3b), a positive relationship ($R^2 = 0.39$, $p = 0.003$, $n = 20$) with a slope of 0.85 was found during the low flow regime in spring and winter (Fig. 6a). This isotopic relationship may suggest the potential for denitrification (Kendall et al., 2007). However, no significant relationship ($p = 0.814$, $n = 22$) was observed between $\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$ in the forest tributary (Fig. 6b). Although a positive relationship between $\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$ in agricultural ($R^2 = 0.15$, $p < 0.001$, slope = 0.28, $n = 92$) and residential ($R^2 = 0.16$, $p = 0.020$, slope = 0.29, $n = 33$) tributaries was observed (Fig. 6b), the slopes were too low to provide strong evidence for denitrification. The weak denitrification signatures may result from high DO concentrations ($6.76 \pm 1.70 \text{ mg L}^{-1}$) in river waters, which were much higher than expected to support denitrification ($< 2 \text{ mg L}^{-1}$, Rivett et al., 2008). Overall, these results suggest that nitrification rather than denitrification is the likely dominant N transformation process in Yongan watershed (Yang et al., 2013; Ji et al., 2017).

For groundwater samples, the slope (0.14) for the $\delta^{15}\text{N}-\text{NO}_3^-$ versus $\delta^{18}\text{O}-\text{NO}_3^-$ relationship was lower than the expected denitrification slopes (Fig. 6f). NO_3^- produced through nitrification combines with relatively low $\delta^{18}\text{O}$ from water that could conceal

the effects of denitrification on the $\delta^{15}\text{N}-\text{NO}_3^-$ versus $\delta^{18}\text{O}-\text{NO}_3^-$ relationship (Osaka et al., 2010). There was also no significant negative relationship between $\ln[\text{NO}_3^-]$ and $\delta^{15}\text{N}-\text{NO}_3^-$, which would be expected due to the NO_3^- loss that accompanies the increase of $\delta^{15}\text{N}-\text{NO}_3^-$ during denitrification (Kendall et al., 2007). Notably, mixed N sources might obscure the groundwater nitrate concentration vs $\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{15}\text{N}-\text{NO}_3^-$ vs $\delta^{18}\text{O}-\text{NO}_3^-$ relationships. In China, N leaching from land-applied fertilizer and manure (e.g., domestic and animal wastes) and the prevalence of leaky septic systems are considered major sources of groundwater NO_3^- in agricultural and residential areas (Lu et al., 2008). In addition, the legacy effect may also have a considerable influence on isotopic signals, as well as the groundwater nitrate concentration vs $\delta^{15}\text{N}-\text{NO}_3^-$ relationship and $\delta^{15}\text{N}-\text{NO}_3^-$ vs $\delta^{18}\text{O}-\text{NO}_3^-$ relationship, since N in water samples is a mixture from different sources and times (Howden et al., 2011).

Measured dissolved N_2O concentrations in groundwater ($3.51 \pm 3.17 \mu\text{g L}^{-1}$) were much higher ($p < 0.001$) than surface waters ($0.58 \pm 0.35 \mu\text{g L}^{-1}$) (Fig. 7). Higher dissolved N_2O concentrations in the groundwater of outlet and T_{agr1} sites than for T_{for} ($p < 0.001$, Fig. 7) suggest that N_2O production (and thereby nitrification and denitrification) is more prevalent in agricultural lands and near the outlet site than for forested sites. Comparatively lower DO concentrations ($3.71 \pm 1.46 \text{ mg L}^{-1}$) measured in $\text{Outlet}_{\text{gw}}$ and GW_{agr} were more favorable for denitrification than those measured in surface waters (Rivett et al., 2008). Two observations during the low flow regime (January in 2015 and 2016) had very high $\delta^{15}\text{N}-\text{NO}_3^-$ values [$> 20\%$, exceeded the maximum groundwater $\delta^{15}\text{N}-\text{NO}_3^-$ observed in nearby regions by Xue et al. (2009), Fig. 6e], which may be influenced by random human activities or analytical errors. When these two high $\delta^{15}\text{N}-\text{NO}_3^-$ values were removed, a significant relationship ($R^2 = 0.33$, $p = 0.001$, slope = 0.48, $n = 31$) between $\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$ was observed for $\text{Outlet}_{\text{gw}}$ (Fig. 6e), implying a potential for denitrification. Since denitrification is strongly influenced by DOC availability (Ben Maamar et al., 2015; Pinay et al., 2015), $\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$ data for $\text{GW}_{\text{outlet}}$ and GW_{agr} were separated by DOC concentrations for further analysis. The upper 50% DOC concentration group showed a significant relationship between $\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$ ($R^2 = 0.71$, $p = 0.001$, slope = 0.62, $n = 12$). These results suggest that denitrification is likely occurring along the groundwater flowpath. Additional studies are warranted to assess the importance of denitrification in groundwater across various land uses at the watershed scale.

3.3. N source identification in river water

For the entire Yongan watershed, SIAR estimates indicated that groundwater contributed $43 \pm 17\%$ of the river NO_3^- load, followed by soil N ($33 \pm 8\%$) and wastewater N ($25 \pm 15\%$) (Fig. 8a). The high contributions from soil and groundwater confirm that NPS pollution was the dominant contributor to riverine N load. The sum of estimated N contributions from groundwater ($43 \pm 17\%$) and soil ($33 \pm 8\%$) was virtually identical to the 79% baseflow contribution for the entire watershed (Fig. 8d). Corresponding to estimated baseflow contributions across the three sub-catchments (i.e., 85%, 84%, and 81%, Fig. 8d), groundwater N contributed to $52 \pm 17\%$, $41 \pm 18\%$ and $37 \pm 16\%$ of riverine NO_3^- loads in agricultural residential and forest tributaries, respectively (Fig. 8a). These results imply that slow flow through the soil zone and groundwater system is an important source of both riverine NO_3^- and water. These estimates are consistent with a previous modeling study that predicted NPS and point sources contributed ~72% and ~18% of riverine N load during the 2000s in the Yongan watershed, respectively (Hu et al., 2018). Additional modeling results from the Yongan watershed estimated that $> 70\%$ of the N exported by rivers was derived

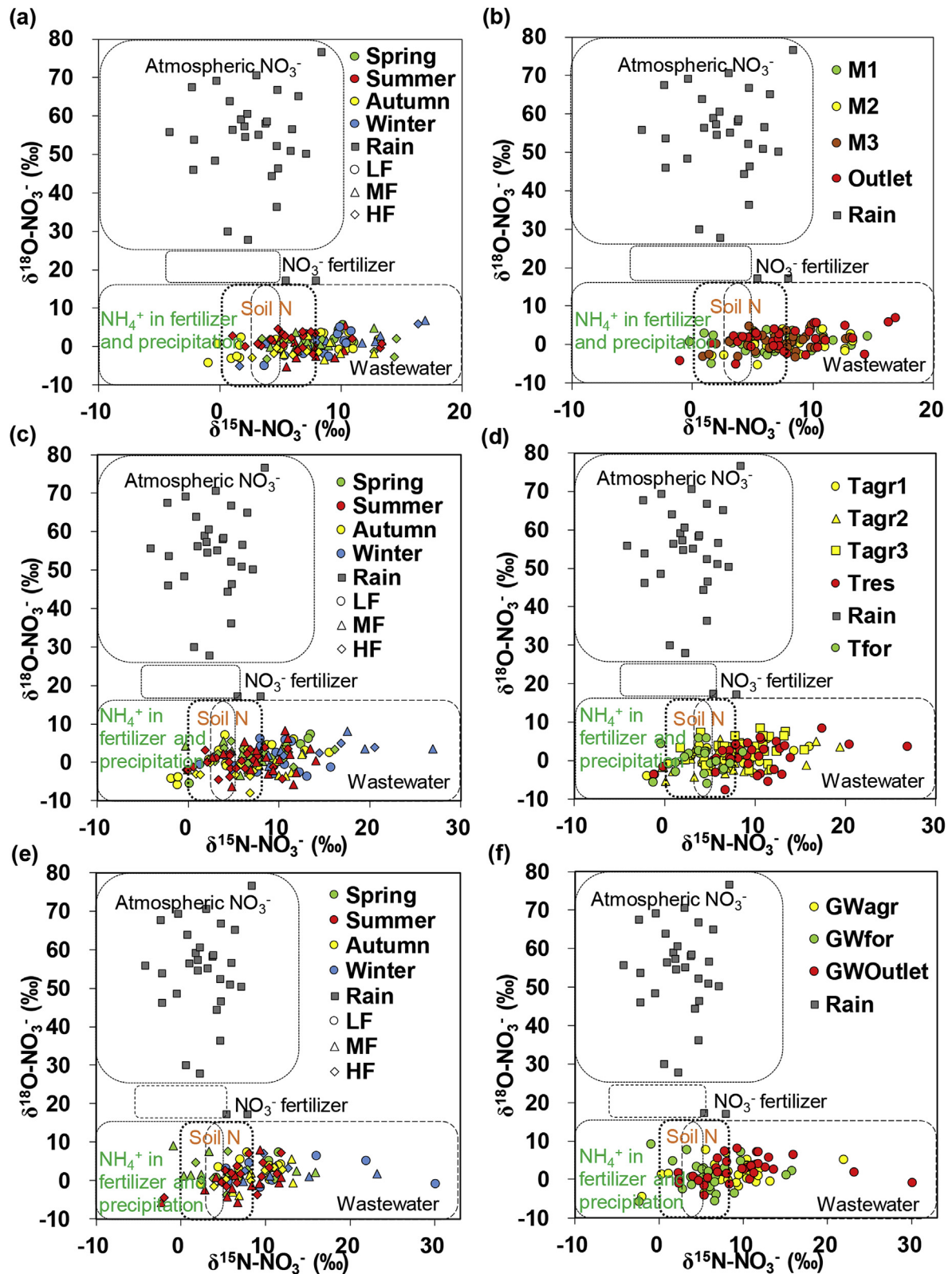


Fig. 6. Dual $\delta^{15}\text{N-NO}_3^-$ and $\delta^{18}\text{O-NO}_3^-$ in rainfall and (a) mainstem water samples in different seasons and flow regimes, (b) mainstem water samples at different sampling sites, (c) tributaries water samples in different seasons and flow regimes, (d) tributaries water samples at different sampling sites, (e) groundwater in different seasons and flow regimes and (f) groundwater at different sampling sites. Area shows the range of the $\delta^{15}\text{N-NO}_3^-$ and $\delta^{18}\text{O-NO}_3^-$ values from Xue et al. (2009), Kaushal et al. (2011) and Yi et al. (2017). Circle in (a), (c) and (e) represents low flow regime (LF = lowest 30% of flows), triangle represents medium flow regime (MF) and rhombus represents high flow regime (HF = upper 30% of flows). T_{agr} is agricultural tributary, T_{res} is residential tributary, T_{for} is forest tributary and GW is groundwater.

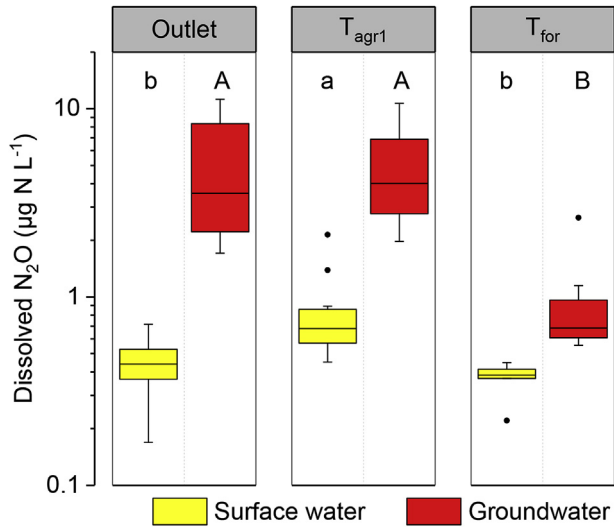


Fig. 7. Box plots for dissolved N₂O-N concentrations in surface water and groundwater at outlet site of Yongan watershed, agricultural tributary (T_{agr1}) and forest tributary (T_{for}).

from soil and groundwater N pools (Chen et al., 2014a, 2016, 2014b). High soil and groundwater N contributions to riverine NO₃⁻ loads are widely reported in watersheds worldwide from models and water quality monitoring (Table 1). This study integrated information from multiple isotope tracers, providing independent methods for verifying and understanding N transformation and transport pathways. Future measurements of ³H, CFCs, and sulfur hexafluoride (SF₆) (Sanford and Pope, 2013; Tesoriero et al., 2013) may further refine estimates for the ages of groundwater and

surface water providing additional verification.

The dominant contributions of soil and groundwater N to riverine N implies the potential for a considerable N leaching lag effect at the watershed scale (Meals et al., 2010; Sanford and Pope, 2013; Hamilton, 2012; Van Meter et al., 2017, 2018). In terms of previous modeling studies, there was ~10 years of lag time between N inputs from various NPS inputs (e.g., fertilizer, biological fixation, atmospheric deposition) and riverine N export in Yongan watershed due to long residence times for hydrologic flowpaths and biogeochemical transformations (Chen et al., 2014b, 2016, 2014a). Previous studies reported relatively long lag times for nitrate transport due to hydrological (~1 to >50 years) and biogeochemical processes (<10–~100 years) in watersheds worldwide (Meals et al., 2010; Hamilton, 2012; Chen et al., 2018; Dupas et al., 2018). These N-leaching lag effects introduce considerable uncertainty in apportioning riverine nitrate to conventional sources (e.g., atmospheric deposition, chemical fertilizer, soil N, manure/sewage).

As expected, there were considerable differences in N source components across different catchments (Fig. 8). For the residential tributary, wastewater N was estimated to contribute the highest riverine NO₃⁻ proportion (~35%), which is consistent with the higher population density (828 capita km⁻²) and domestic animal numbers (136 capita km⁻²) observed in the residential tributary catchment compared to the other catchments (<300 capita km⁻² and <120 capita km⁻²) (Table 2). For the agricultural tributary, groundwater N was estimated to contribute nearly 60% of the riverine NO₃⁻ load, with soil and wastewater contributing ~35% and ~5%, respectively. In contrast, N sources in the forest tributary were dominated by soil N (>50%) and groundwater (~40%). The lower groundwater N contribution in the forest catchment can be explained by significantly lower groundwater NO₃⁻ concentrations in forest-dominated catchments (0.80 ± 0.22 mg N L⁻¹) compared to agricultural catchments (2.01 ± 0.81 mg N L⁻¹) (Fig. 3e). The

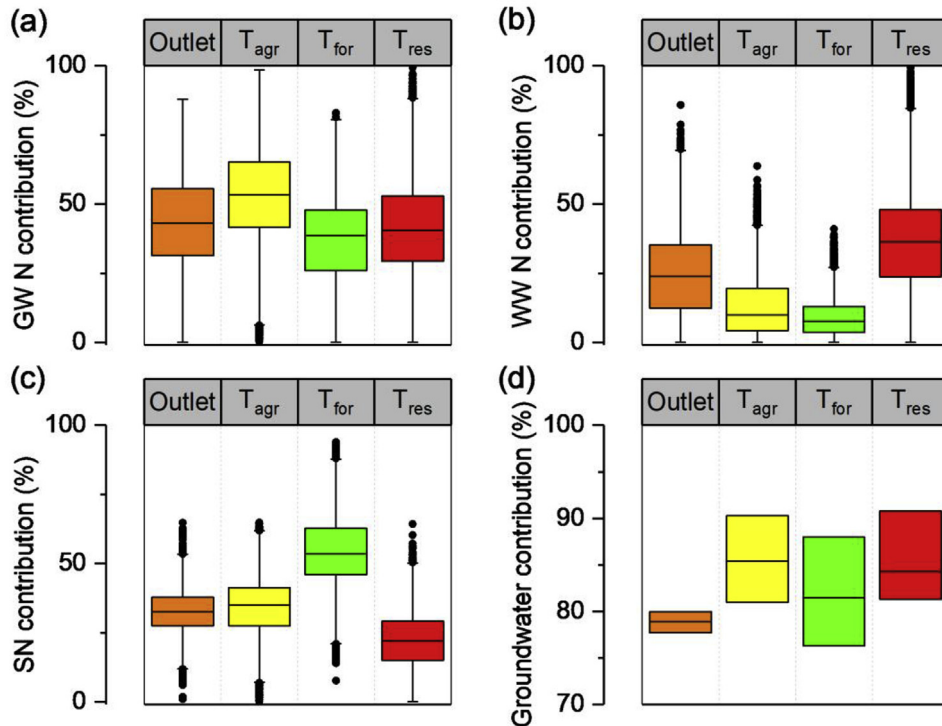


Fig. 8. Box plots for (a) the contribution of groundwater, (b) wastewater N and (c) soil N to riverine NO₃⁻ in at the Yongan watershed outlet (Outlet), agricultural tributary (T_{agr}) and forest tributary (T_{for}); (d) water contribution from groundwater in different catchments based on sine wave regressions. Box plots in (a), (b), (c) illustrate the 25th, 50th and 75th percentile; the whiskers indicate the 2.5th and 97.5th percentiles; and the points represent data outliers. Box plots in (d) illustrate the 2.5th, mean values and 97.5th percentiles.

paddy field proportion in T_{agr1} (5.6%) was much higher than that in T_{for} (1.5%), which lead to higher fertilizer application and higher water infiltration into subsurface flow and/or groundwater hydrologic pathways (Table 2). For example, $\sim 380 \text{ mm yr}^{-1}$ of water infiltration combined with high soil N levels contributed considerable NO_3^- leaching to groundwater in paddy fields of the nearby Taihu region (Xu et al., 2017). The different N source (e.g., soil, groundwater and wastewater) contributions among different sub-catchments are consistent with results observed in nearby watersheds in eastern China (Xing and Liu, 2016; Xia et al., 2017). Overall, the differences distinguished for N contributions among tributaries are strongly related to the influence of land use on N fate and transport.

3.4. Implications for river N pollution control

Results of this study were mutually supported by hydro-chemistry, multiple stable isotopes ($^{15}\text{N-NO}_3^-/^{18}\text{O-NO}_3^-$ and $^2\text{H-H}_2\text{O}/^{18}\text{O-H}_2\text{O}$), dissolved N_2O concentrations, and previous modeling results, thereby providing an integrated approach for exploring and quantifying watershed N transformation and transport pathways. This study provides watershed managers with important information for making science-based decisions on N pollution control strategies. Since NPS pollution dominated in Yongan watershed, management strategies for controlling riverine N pollution should be focused on reducing NPS pollution (Fig. 8a and c). NPS N pollution can be controlled by several management strategies, such as improved irrigation and nutrient management, reduced-tillage with enhanced crop residue cover, seasonal cover crops, and vegetative buffer zones between croplands and rivers (Ji et al., 2017). To address high wastewater N contributions in residential catchments, centralized collection of wastes from individual households, currently having marginal septic systems, is critical for reducing N pollution and several other human-health hazards. For example, efficient wastewater collection and centralized treatment systems and/or artificial wetlands should be implemented to efficiently reduce N pollution loads from domestic and animal wastes (Chen et al., 2013).

Results of this study combined with previous modeling studies (Chen et al., 2014a, 2014b; Hu et al., 2018) confirm the potential for high NO_3^- contributions from groundwater and soil N pools (Fig. 8), implying a N-leaching legacy effect in the Yongan watershed. Therefore, reduction of N inputs from various sources (e.g., atmospheric deposition, chemical fertilizer, biological fixation) may not be immediately recognized as decreased riverine N loads (Meals et al., 2010). Instead, measures aimed at reducing soil and groundwater N pools that contribute appreciable NO_3^- loads should be considered, in addition to measures to reduce anthropogenic input sources. Since riparian and hyporheic zones are recognized as hot spots for NO_3^- removal, restoration of stream and riparian zones is a compelling strategy for enhancing groundwater NO_3^- removal by denitrification (Boano et al., 2014). Possible strategies for enhancing in-stream denitrification include increasing river water residence times, restoring floodplains and increasing microbially available carbon (Hester and Gooseff, 2010). Permeable reactive barrier technology also provides a promising way for sustainable *in situ* groundwater restoration, which makes use of *in situ* groundwater flow to bring N pollutants in contact with reactive materials for NO_3^- removal by microbial denitrification (Liu et al., 2013; Obiri-Nyarko et al., 2014). To reduce the soil N pool contribution to legacy N in agricultural soils, sustainable nutrient management measures should be integrated to optimize uptake of legacy nutrients and reduce new fertilizer inputs (Chen et al., 2018) and incorporate perennial vegetation to more efficiently sequester nutrients while also providing a source of forage or biofuels (Van Meter et al., 2017;

Thomas and Abbott, 2018).

4. Conclusion

This study combined 3-years of multiple stable isotopes, hydrologic, water chemistry and nitrous oxide measurements as well as N sources identified in previous modeling studies to assess N dynamics and riverine N sources in the Yongan Watershed. This investigation provides a reliable and integrated approach for verifying and understanding N pollution legacy effects observed in many watersheds worldwide. Information from $\delta^{15}\text{N-NO}_3^-$ and $\delta^{18}\text{O-NO}_3^-$ indicated that riverine N dynamics were regulated by contributing sources, nitrification and denitrification, as well as hydrological processes. Information from $^2\text{H-H}_2\text{O}$ and $^{18}\text{O-H}_2\text{O}$ indicated that slow subsurface and groundwater flows, accounting for >75% of river water discharge, were potentially major hydrologic pathways for N delivery to the river. Riverine NO_3^- source contributions varied with dominant land-use types, with the highest contributions from groundwater (60%), wastewater (35%), and soil (50%) occurring in agricultural, residential and forest catchments, respectively. High N contributions from groundwater and soil N pools imply considerable potential for nutrient leaching legacy effects. Results highlight that river N pollution control requires particular attention to groundwater restoration and soil N management in addition to N input control strategies. Several future research needs were identified to better understand the fate and transport of N through the soil and groundwater systems, as these are the dominant N pools regulating river N pollution.

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.watres.2018.11.082>.

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