UCSF

UC San Francisco Previously Published Works

Title

Phase I Study of CTT1057, an 18F-Labeled Imaging Agent with Phosphoramidate Core Targeting Prostate-Specific Membrane Antigen in Prostate Cancer

Permalink

https://escholarship.org/uc/item/7dw436r8

Journal

Journal of Nuclear Medicine, 60(7)

ISSN

0161-5505

Authors

Behr, Spencer C Aggarwal, Rahul VanBrocklin, Henry F et al.

Publication Date

2019-07-01

DOI

10.2967/jnumed.118.220715

Peer reviewed

Phase I Study of CTT1057, an ¹⁸F-Labeled Imaging Agent with Phosphoramidate Core Targeting Prostate-Specific Membrane Antigen in Prostate Cancer

Spencer C. Behr*¹, Rahul Aggarwal*², Henry F. VanBrocklin¹, Robert R. Flavell¹, Kenneth Gao¹, Eric J. Small^{2,3}, Joseph Blecha¹, Salma Jivan¹, Thomas A. Hope¹, Jeffry P. Simko⁴, John Kurhanewicz¹, Susan M. Noworolski¹, Natalie J. Korn¹, Romelyn De Los Santos⁴, Matthew R. Cooperberg³, Peter R. Carroll³, Hao G. Nguyen³, Kirsten L. Greene³, Beatrice Langton-Webster⁵, Clifford E. Berkman⁵, and Youngho Seo¹

¹Department of Radiology and Biomedical Imaging, University of California, San Francisco, San Francisco, California; ²Department of Medicine, University of California, San Francisco, San Francisco, California; ³Department of Urology, University of California, San Francisco, San Francisco, California; ⁴Department of Pathology, University of California, San Francisco, San Francisco, California; and ⁵Cancer Targeted Technology, Woodinville, Washington

Agents targeting prostate-specific membrane antigen (PSMA) comprise a rapidly emerging class of radiopharmaceuticals for diagnostic imaging of prostate cancer. Unlike most other PSMA agents with a urea backbone. CTT1057 is based on a phosphoramidate scaffold that irreversibly binds to PSMA. We conducted a first-in-humans phase I study of CTT1057 in patients with localized and metastatic prostate cancer. Methods: Two patient cohorts were recruited. Cohort A patients had biopsy-proven localized prostate cancer preceding radical prostatectomy, and cohort B patients had metastatic castration-resistant prostate cancer. Cohort A patients were imaged at multiple time points after intravenous injection with 362 ± 8 MBg of CTT1057 to evaluate the kinetics of CTT1057 and estimate radiation dose profiles. Mean organ-absorbed doses and effective doses were calculated. CTT1057 uptake in the prostate gland and regional lymph nodes was correlated with pathology, PSMA staining, and the results of conventional imaging. In cohort B, patients were imaged 60-120 min after injection of CTT1057. PET images were assessed for overall image quality, and areas of abnormal uptake were contrasted with conventional imaging. Results: In cohort A (n = 5), the average total effective dose was 0.023 mSv/MBq. The kidneys exhibited the highest absorbed dose, 0.067 mGy/MBq. The absorbed dose of the salivary glands was 0.015 mGy/MBq. For cohort B (n = 15), CTT1057 PET detected 97 metastatic lesions, and 44 of 56 bone metastases detected on CTT1057 PET (78.5%) were also detectable on bone scanning. Eight of 32 lymph nodes positive on CTT1057 PET (25%) were enlarged by size criteria on CT. Conclusion: CTT1057 is a promising novel phosphoramidate PSMA-targeting ¹⁸F-labeled PET radiopharmaceutical that demonstrates similar biodistribution to urea-based PSMA-targeted agents, with lower exposure to the kidneys and salivary glands. Metastatic lesions are detected with higher sensitivity on CTT1057 imaging than on conventional imaging. Further prospective studies with CTT1057 are warranted to elucidate its role in cancer imaging.

Received Sep. 20, 2018; revision accepted Nov. 13, 2018.

Key Words: prostate cancer; PSMA; PET; dosimetry; CTT1057

J Nucl Med 2019; 60:910-916 DOI: 10.2967/jnumed.118.220715

Early and accurate diagnosis of prostate cancer is essential for proper patient management. Contrast-enhanced CT and radionuclide bone scanning are used for staging and restaging. However, more sensitive and accurate imaging tests are needed than are available with current standard-of-care examinations (1-4).

PET imaging is an enticing choice as it offers the potential to both stage patients and provide insight into tumor biology. Prostate-specific membrane antigen (PSMA) is a promising target because it is overexpressed in both the primary tumor and the metastatic lesions. Recently, urea-based PSMA-targeting agents such as ⁶⁸Ga-PSMA-11 have shown promise in prostate cancer imaging (5-7).

¹⁸F-labeled agents such as ¹⁸F-DCFPyL (8) and ¹⁸F-PSMA-1007 (9) have also entered clinical development. ¹⁸F offers several advantages over ⁶⁸Ga. First, cyclotron production of ¹⁸F enables large-scale batches. Second, ¹⁸F has a longer half-life than ⁶⁸Ga (110 min vs. 68 min), simplifying transport and potentially allowing for a greater delay in imaging. Third, the shorter positron range of ¹⁸F than of ⁶⁸Ga provides improved image quality.

CTT1057 is a peptidomimetic incorporating a phosphoramidate scaffold (Fig. 1), resulting in irreversible binding to PSMA and improved tumor uptake as shown in preclinical models (10). Hence, the different biodistribution of CTT1057 from that of other urea-based agents is worth careful investigation, particularly for developing a corresponding PSMA-targeted therapeutic radiopharmaceutical. Salivary glands and kidneys are organs at risk when a urea-based PSMA-targeting radiopharmaceutical therapy is applied; thus, dose profiles in these organs are of great importance for developing a new PSMA imaging agent such as CTT1057.

In this first-in-humans phase I study, the safety, biodistribution, radiation dosimetry, and histopathologic correlation were evaluated in men before prostatectomy, and the utility of CTT1057 in detecting PSMA-avid metastases was assessed in patients with

For correspondence or reprints contact: Spencer C. Behr, Department of Radiology and Biomedical Imaging, University of California, San Francisco, 505 Parnassus Ave., Box 0628, San Francisco, CA 94143.

E-mail: spencer.behr@ucsf.edu

^{*}Contributed equally to this work.

Published online Nov. 21, 2018.

COPYRIGHT © 2019 by the Society of Nuclear Medicine and Molecular Imaging.

FIGURE 1. Chemical structure of CTT1057 with comparison to other urea-based PSMA PET agents: ¹⁸F-DCFPyL, ¹⁸F-PSMA-1007, and ⁶⁸Ga-PSMA-11. Orange area highlights phosphoramidate backbone in CTT1057, and blue area highlights urea backbone in other 3 agents.

metastatic castration-resistant prostate cancer (mCRPC). Furthermore, a comparison of different absorbed doses in organs between CTT1057 and several other PSMA imaging agents was made.

MATERIALS AND METHODS

This prospective study was performed after approval by the local Institutional Review Board and by the Food and Drug Administration under an investigational new drug application (124021) held by Cancer Targeted Technology and registered as NCT02916537.

Preparation of CTT1057

CTT1057 was prepared under current good manufacturing practices at our institution. CTT1057 was produced on an ORA Neptis Perform synthesizer (Optimized Radiochemical Applications) coupling succinimidyl-¹⁸F-fluorobenzoate to the primary amine precursor CTT1298, as previously described (11).

Patient Characteristics and Study Design

Written informed consent was obtained from each patient before radiotracer administration. Key inclusion criteria were an Eastern Cooperative Oncology Group performance status of 0 or 1,

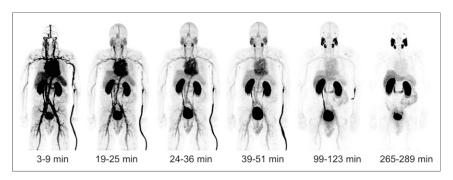


FIGURE 2. PET maximum-intensity projections from patient 3. This patient was 73-y-old preprostatectomy patient who had Gleason 3 + 4 prostate cancer and PSA of 6.7 ng/mL 2 wk before imaging. Patient did not have any PSMA-avid lymph nodes or metastases at time of imaging. Surgery 12 wk after imaging confirmed disease localized to prostate, with no lymph node involvement. Activity seen adjacent to left arm is due to radiotracer.

histologically confirmed adenocarcinoma of the prostate, and adequate hematologic and organ function. In cohort A, patients had to be deemed eligible for radical prostatectomy within 12 wk after CTT1057 PET. No systemic anticancer therapy was allowed before imaging, including androgen deprivation, chemotherapy, or investigational systemic therapy. In cohort B, patients were required to have mCRPC as defined by the prostate cancer working group 2 criteria, with at least 3 metastatic lesions seen on CT or MRI of the chest, abdomen, or pelvis or on radionuclide bone scanning performed within 12 wk of CTT1057 PET. Key exclusion criteria for both cohorts were any other radionuclide within 5 physical half-lives, histologic evidence of small cell or neuroendocrine prostate cancer in more than 50% of biopsy tissue, or a history of invasive secondary malignancies within 12 mo. Safety assessments included serial monitoring of vital signs and injection site, adverse event assessment on the day of scanning up to 6 h, and an assessment by telephone at 7 d after CTT1057 PET.

Imaging Protocol for Cohort A

All PET scans were performed on a 3-T time-of-flight PET/MR system (Signa PET/MR; GE Healthcare). Three-dimensional PET data were acquired in list mode with a varying time per bed position as time progressed for 6 bed positions. PET images were reconstructed with a matrix size of 192×192 , 28 subsets and 3 iterations of a timeof-flight ordered-subsets expectation maximization algorithm, an inplane gaussian postreconstruction filter of 5 mm in full width at half maximum, and a point-spread-function model. Attenuation correction for PET reconstruction was performed using the MR-based attenuation-correction technique provided by the scanner manufacturer. Fullbody MR images were axial MR-based attenuation-correction images, 2-point Dixon 3-dimensional fast spoiled gradient-recalled echo images, axial variable flip-angle single-shot fast spin echo T2-weighted images, and axial diffusion-weighted images. Patients without a clinical prostate MR examination within 12 wk of this study received a multiparametric MRI examination with an endorectal coil (12). Dedicated pelvic MR sequences included axial 3-dimensional, reduced-fieldof-view T2-weighted imaging, multiple diffusion-weighted imaging (b = 0, 600, and 1,400), and axial T1-weighted imaging.

The administered mass of CTT1057 was $4.01 \pm 3.20 \mu g$ (range, $1.49-8.83 \mu g$). The administered activity was $362 \pm 8 \mu g$ (range, $352-370 \mu g$).

Six PET bed positions were acquired to cover vertex to mid thighs. Six sequential PET examinations were acquired for each patient (i.e., PET1 through PET6). Acquisition durations were 1 min/bed position for PET1 and PET2, 2 min/bed position for PET3 and PET4, and 4 min/bed position for PET5 and PET6. PET1 through PET4 were acquired immediately after administration of CTT1057 without any interruptions. PET5 was started approximately 50–80 min after injection. PET6 was acquired approximately 100 min after PET5. Between PET5 and PET6, dedicated MR pelvic images were acquired as described

TABLE 1
Radiation Dose Estimates (OLINDA 1.1, ICRP60) of CTT1057

Site	Absorbed dose (mGy/MBq)
Adrenals	0.009 ± 0.001
Brain	0.006 ± 0.000
Breasts	0.005 ± 0.001
Gallbladder wall	0.014 ± 0.001
Lower large intestine wall	0.013 ± 0.003
Small intestine	0.010 ± 0.001
Stomach wall	0.007 ± 0.001
Upper large intestine wall	0.009 ± 0.001
Heart wall	0.018 ± 0.001
Kidneys	0.067 ± 0.001
Liver	0.016 ± 0.000
Lungs	0.013 ± 0.001
Muscle	0.007 ± 0.001
Pancreas	0.009 ± 0.001
Red marrow	0.007 ± 0.001
Osteogenic cells	0.009 ± 0.002
Skin	0.005 ± 0.001
Spleen	0.016 ± 0.001
Testes	0.010 ± 0.002
Thymus	0.007 ± 0.001
Thyroid	0.005 ± 0.001
Urinary bladder wall	0.259 ± 0.126
Effective dose (mSv/MBq)	0.023 ± 0.007

earlier. Patients were encouraged to void during breaks, but this was not mandated.

Imaging Protocol for Cohort B

For cohort B, only one static scan 60-120 min after administration of 369.83 ± 7 MBq of CTT1057 (range, 355.2-381.1 MBq) was acquired, at 3 min/bed position for 6 bed positions. The PET reconstruction parameters for cohort B were the same as for cohort A. The full-body MR examination was performed without intravenous contrast material using the same protocol as for cohort A.

Radiation Dosimetry for Cohort A

From PET1 through PET6, time-activity data were generated for brain, lungs, heart, liver, gallbladder, spleen, kidneys, and urinary bladder. Using the average male organ mass and density values, the mean activity concentrations multiplied by the volumes of each organ were used as the total activities. Residence times were derived by curve-fitting the time-activity data. The data fitting and dose calculations were performed on OLINDA|EXM (version 1.1, Vanderbilt University). The absorbed dose to each organ (mGy/MBq) and the ICRP 60 effective dose (mSv/MBq) were tabulated. Finally, organ-specific absorbed dose was estimated using a sphere model implemented in OLINDA 1.1 for salivary and lacrimal glands.

Radiology–Pathology Primary Tumor Assessment for Cohort A

Under the guidance of a pathologist, the resected prostates were weighed, measured, inked, infiltrated with 400 mL of 10% buffered

formalin using a syringe, and fixed overnight in a 10% neutral buffered formalin bath. The prostates were sliced at 3-mm intervals perpendicular to the urethra using a customized slicer that was designed for this purpose, the apical sides of all sections were placed down in their respective tissue cassettes (to preserve orientation) and processed and cut as whole-mount sections. Slides were evaluated using a conventional light microscope, with all tumors marked and characterized as to tumor Gleason score, percentage of various Gleason grades, and tumor growth patterns. The slides were subsequently digitized on a high-resolution flatbed scanner for digital overlapping with the radiographic images. For immunohistochemical assessment, tissues were sectioned at 4 µm; heat-induced epitope retrieval with a citrate buffer and streptavidin horseradish peroxidase were used. PSMA expression in the primary prostate tumors was assessed using a mouse monoclonal antibody to PSMA (clone 3E6; Dako).

Whole-Body Image Analysis

Two board-certified nuclear medicine physicians with 7 and 12 y of PET experience reviewed all PET images on an Advantage Workstation (GE Healthcare). MR-based attenuation correction, non-attenuation-corrected images, and MR images were reviewed.

For cohort A, each PET time point was assessed for focal radiotracer uptake in the prostate. Uptake was considered positive if it was clearly above the adjacent prostatic parenchyma. For each focal CTT1057-avid lesion, a region of interest (ROI) was drawn to encompass the lesion, with care to exclude the urinary bladder. This ROI was then propagated to the other PET time points, and the SUV_{max} was recorded. If no focal uptake was clearly seen above background, an ROI was drawn in the location of the tumor based on pathology and multiparametric MRI. Blood-pool uptake and muscle uptake were calculated by drawing a 1-cm ROI and recording the SUV_{mean} in the ascending aorta for blood pool and right gluteal musculature for muscle. The ratio of tumor to blood pool and tumor to musculature was calculated at each PET time point. PSMA-avid lymph nodes were considered positive if PSMA uptake was clearly above adjacent blood pool and in the expected location of a lymph node on cross-sectional imaging.

For cohort B, SUV_{mean} was calculated for multiple normal organs for each patient by drawing a 1-cm ROI. The anatomic location and SUV_{max} were determined for each identifiable metastatic lesion, with up to 5 lesions per organ. Lesion-to-background ratios (i.e., tumor SUV_{max} to SUV_{mean} for liver, blood pool, and muscle) were calculated. Additionally, each nuclear medicine physician qualitatively scored each study using a visual analog scale of 1 to 100 (1 = non-diagnostic, 100 = perfect study).

RESULTS

Patient Characteristics and Safety Results

In total, 20 patients were enrolled (5 in cohort A and 15 in cohort B). Supplemental Table 1 summarizes individual patient characteristics, surgical pathology, and CTT1057 imaging results for cohort A (supplemental materials are available at http://jnm.snmjournals.org). Supplemental Table 2 summarizes patient characteristics, prior treatment, and PSA levels for cohort B. Average age was similar between cohorts. In cohort A, patients had Gleason 3 + 4 (n = 3) or Gleason 4 + 3 (n = 2) patterns, with a mean PSA of 12.29 ng/mL (range, 4–38.76 ng/mL). In cohort B, the mean PSA was 49.2 ng/mL (range, 0.7–1,238.6 ng/mL), and all were receiving ongoing androgen deprivation therapy. Nine patients (60%) had received prior treatment with abiraterone or enzalutamide. All 15 patients had definitive local therapy: prior

TABLE 2

Comparison of Organ and Absorbed and Effective Dose Estimates for CTT1057 Compared with Those of ⁶⁸Ga-PSMA-11 (13), ¹⁸F-DCFPyL (8), and ¹⁸F-PSMA-1007 (9)

	Absorbed dose (mGy/MBq)					
Organ	¹⁸ F-CTT1057 (this work)	⁶⁸ Ga-PSMA-11 (Afshar-Oromieh et al.)	¹⁸ F-DCFPyL (Szabo et al.)	¹⁸ F-PSMA-1007 (Giesel et al.)		
Adrenals	9.32E-03	1.42E-02	3.11E-02	1.94E-02		
Brain	5.79E-03	9.00E-03	2.19E-02	7.20E-03		
Breasts	5.06E-03	8.80E-03	4.57E-03	8.06E-03		
Gallbladder wall	1.43E-02	1.44E-02	1.44E-02	2.22E-02		
Lower large intestine wall	1.35E-02	1.23E-02	1.05E-02	4.83E-02		
Small intestine	9.72E-03	1.63E-02	9.13E-03	1.56E-02		
Stomach wall	7.47E-03	1.20E-02	1.16E-02	1.42E-02		
Upper large intestine wall	9.08E-03	5.40E-02	1.67E-02	4.08E-02		
Heart wall	1.78E-02	1.09E-02	1.29E-02	2.51E-02		
Kidneys	6.74E-02	1.62E-01	9.45E-02	1.70E-01		
Liver	1.59E-02	3.09E-02	3.80E-02	6.05E-02		
Lungs	1.33E-02	1.02E-02	1.08E-02	1.11E-02		
Muscle	7.44E-03	1.05E-02	6.32E-03	1.00E-02		
Pancreas	9.10E-03	1.38E-02	2.44E-02	1.92E-02		
Red marrow	6.95E-03	9.20E-03	1.04E-02	1.33E-02		
Osteogenic cells	9.10E-03	1.42E-02	9.58E-03	1.55E-02		
Skin	4.94E-03	8.85E-02	4.05E-03	7.30E-03		
Spleen	1.61E-02	4.46E-02	1.85E-02	7.39E-02		
Testes	9.86E-03	1.04E-02	1.01E-02	8.37E-03		
Thymus	6.72E-03	9.90E-03	5.56E-03	9.90E-03		
Thyroid	5.47E-03	9.70E-03	8.56E-03	8.50E-03		
Urinary bladder wall	2.59E-01	1.30E-01	8.64E-02	1.87E-02		
Effective dose (mSv/MBq)	2.28E-02	2.36E-02	1.39E-02	2.20E-02		

radical prostatectomy in 5 (33%) and prostate radiation without or with pelvic radiation in 10 (67%).

No adverse events or changes in vitals were associated with CTT1057 injection in the study.

Biodistribution for Cohort A

PET images demonstrated uptake within the salivary glands, lacrimal glands, liver, spleen, and proximal small bowel (Fig. 2). Blood-pool activity progressively decreased at 90–120 min after injection, with rapid excretion through the kidneys into the urinary bladder. In 4 of the 5 patients, biliary excretion was noted.

Radiation Dosimetry for Cohort A

The effective dose was estimated at 0.023 \pm 0.007 mSv/MBq (Table 1). A factor for the variation in estimated effective doses between patients was the absorbed dose in the urinary bladder. Table 2 also shows the dose comparison to 3 other PSMA-targeted PET imaging agents, 68 Ga-PSMA-11 (*13*), 18 F-DCFPyL (*8*), and 18 F-PSMA-1007 (*9*). The estimated absorbed doses in salivary and lacrimal glands were 0.0146 and 0.00732 mGy/MBq, respectively.

Primary Tumor Analysis for Cohort A

Four patients had CTT1057-avid prostate lesions corresponding to the pathology-proven cancer. The one patient without focal prostatic PSMA uptake had a PSA of 4 ng/dL and Gleason 3 + 4. The highest uptake in the primary tumors was seen at PET5 and PET6 (Fig. 3).

In total, 68 lymph nodes were resected during surgery from the 5 patients. None of these lymph nodes was macroscopically positive; one node had microscopic metastatic disease (<0.2 mm). This patient had the highest PSA level (38.76 ng/mL) and Gleason 4+3.

Qualitative and Quantitative Image Analysis for Cohort B

The average visual analog scale score of CTT1057 images was 76 ± 5.4 . The normal-organ uptake of CTT1057 is shown in Supplemental Table 3.

All 15 patients had evidence of at least one PSMA-avid metastasis on CTT1057 PET. In total, 97 PSMA-avid lesions were evaluated, with an average of 6.5 lesions per patient. The median SUV_{max} of the lesions was 12.17 (interquartile range, 5.9–19.02). The distribution of metastatic sites detected by CTT1057 PET is shown in Table 3. CTT1057 demonstrated improved sensitivity for bone and lymph node detection compared with conventional bone scanning and CT of the chest, abdomen, or pelvis. This result is consistent with other reports that showed PSMA PET to have improved detection rates of bone metastases over bone

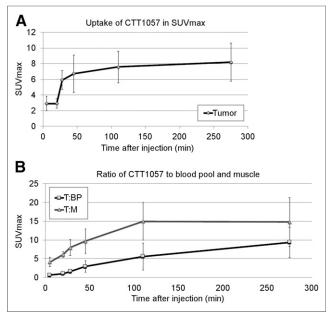


FIGURE 3. CTT1057 uptake in primary prostate tumors over time. (A) Comparison of average SUV_{max} over multiple PET time points for all tumors. (B) Graph of ratio of tumor to blood pool (T:BP) and tumor to muscle (T:M) over time of 6 CTT1057-avid tumors. Error bars indicate SD.

scanning (14). Of the 56 positive bone metastases detectable on CTT1057, 44 (78.5%) were detectable on bone scanning. Of the 32 positive lymph nodes on CTT1057, only 8 (25%) were enlarged by size criteria on baseline CT imaging. All CT enlarged lymph nodes (>1.5 cm in short axis) were positive on CTT1057. Three lesions were seen on bone scanning only, without corresponding CTT1057 uptake. Figure 4 shows examples of CTT1057-avid osseous and lymph node metastases. Two patients who were scanned with CTT1057 PET subsequently underwent metastatic tumor biopsy of a PSMA-avid lesion for purposes of clinical care. In both cases, the CT-guided core-needle biopsy was positive for prostate cancer and contained sufficient tumor material to permit histopathologic evaluation (Supplemental Fig. 1).

DISCUSSION

This first-in-humans study demonstrated that this novel, phosphoramidate-based PSMA-targeted imaging agent, CTT1057, is safe and has no adverse reactions. Furthermore, CTT1057 both

accurately identified the primary prostate cancer observed on pathologic examination of radical prostatectomy specimens and detected osseous and lymph node metastases with improved sensitivity compared with conventional imaging.

The biodistribution of CTT1057 showed characteristics similar to those of other PSMA PET agents. Overall, there was high salivary, renal, and urinary bladder uptake. The high renal uptake was likely a combination of the PSMA expression and excretion of radiotracer, which resulted in high accumulation of radiotracer in the urinary bladder—similar to that reported for both ⁶⁸Ga-based and ¹⁸F-based PSMA agents (8,15). Unlike the other urea-based PSMA agents, we observed only minimal radiotracer in the small bowel. Additionally, a high number of patients imaged with CTT1057 had biliary excretion of the radiotracer, which was rarely reported with the urea-based radiotracers until recently, with PSMA-1007 (9). Our data indicate that delayed imaging 90 min or later would be preferred, as is in line with other PSMA PET agents. However, unlike PSMA-1007 (9), CTT1057 is similar to other PSMA-based agents in the high accumulation within the urinary bladder, which is a potential limitation as it could obscure adjacent lesions in the prostate or regional pelvic lymph nodes.

As detailed in Table 4, the effective dose from CTT1057 of 0.0231 mGy/MBq was similar to effective doses reported for ⁶⁸Ga-PSMA-11 (*15*) and ¹⁸F-PSMA-1007 (*9*) and moderately higher than ¹⁸F-DFCPyL (*8*). When compared with radiation-sensitive organs such as red marrow and breast tissue, the absorbed dose was less than ⁶⁸Ga-PSMA-11 and ¹⁸F-PSMA-1007 and similar to ¹⁸F-DFCPyL. The effective dose of CTT1057 was increased by the significant urinary accumulation. For example, when the urinary bladder dose was simulated to be the same as from ¹⁸F-DCFPyL, the effective dose scaled down to 0.014 mSv/MBq. Future studies with oral hydration, intravenous furosemide, and voiding before imaging could decrease the overall effective dose and the absorbed doses to the testes and urinary bladder.

In comparison to the data reported only for salivary glands, the dose (0.0146 mGy/MBq) from CTT1057 was significantly lower than the dose (0.0610 mGy/MBq) from ⁶⁸Ga-PSMA I&T (16). Xerostomia has also been shown to be the dose-limiting toxicity in clinical trials with urea-based PSMA-617 (17,18). Since the same phosphoramidate binding scaffold is being pursued as a therapeutic PSMA-targeted agent, this is an important finding that could potentially reduce complications to salivary and lacrimal glands during radionuclide therapy. Additionally, the absorbed dose of CTT1057 in the kidneys was less than reported for the other 3 PSMA-based agents (8,9,19).

TABLE 3
Metastatic Sites of Uptake on CTT1057 and Conventional Imaging

	CTT1057 PET		Whole-body bone scan	n CT chest/abdomen/pelvis		
Site	n	Median SUV _{max}	IQR	n	size (cm)*	
Lymph node	32	11.28	5.98–23.99	NA	0.9 (range, 0.4-2.8)	
Bone	56	12.96	7.32-18.52	44 (78.5%)	NA	
Prostate bed	3	14.09	13.64–25.8	NA	1.43 (range, 1.2-1.7)	
Lung	6	3.16	2.65-5.45	NA	1.2 (range, 0.8 -2.1)	

^{*}Measurements were performed in long-axis measurements except for lymph nodes, which were measured along short axis. IQR = interquartile range; NA = not applicable.

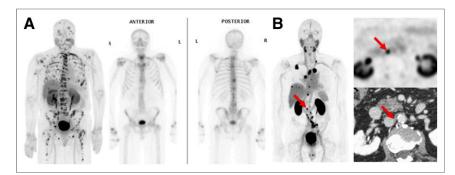


FIGURE 4. (A) Example of CTT1057-avid osseous and lymph lesions from cohort B, showing CTT1057 PET maximum-intensity projection (left) and anterior (middle) and posterior (right) ^{99m}Tc-HDP planar bone scan. (B) Example of patient with extensive CTT1057-avid osseous metastatic disease and only minimal uptake on standard-of-care bone scan. CTT1057 MIP PET (left), axial PET (top right), and axial CT (bottom right) highlights patient with 4-mm short lymph node (arrows) that is not enlarged by size criteria on conventional CT but displays marked CTT1057 uptake indicative of high likelihood of metastatic involvement.

All 5 enrolled preprostatectomy patients had Gleason patterns of 7. In 4 patients, PSMA uptake was clearly seen above background in the primary tumors (Supplemental Fig. 2) and PSA levels were less than 10 ng/mL The most intense uptake in the primary tumor by CTT1057 also had the highest PSA levels (38.8) and a Gleason pattern of 4 + 3 (Supplemental Fig. 3). These findings are consistent with those recently reported by Uprimny et al. (20). The overall assessment for the sensitivity for lymph node detection was limited because this study was performed primarily for dosimetry and safety. Of the 68 lymph nodes resected, there was no macroscopic disease, and only one node had microscopic metastases (<0.2 mm of disease). This lymph node did not demonstrate any significant uptake of CTT1057, a finding that might have been due to a minimal amount of disease present, below the threshold for PET imaging (21–23).

In the mCRPC cohort, more lesions were detected with CTT1057 PET-based imaging than with conventional CT or bone scanning. Despite the limitation of pathologic confirmation of PSMA-avid lesions, the results support future studies with CTT1057 for routine clinical care in this setting. In patients with oligometastatic CRPC, when consideration is frequently made for the focal targeting of metastases with radiation, improved sensitivity of lesion detection permits more comprehensive and accurate targeting of detectable metastases. In addition, the choice of systemic therapy can differ in nonmetastatic versus mCRPC, and earlier detection of metastases enables the earlier institution of recommended therapies for mCRPC, including abiraterone, enzalutamide, and other agents in clinical development. Conversely, the absence of PSMA uptake in osseous lesions detectable on conventional bone scanning may signify a metabolically inactive tumor indicative of therapeutic response. These hypotheses warrant evaluation in future prospective studies in the mCRPC setting.

The degree of intratumoral uptake of CTT1057 appears to be comparable to other PSMA-targeted tracers in clinical development, including ⁶⁸Ga-PSMA-11 (24–26). The high uptake of CTT1057 in both osseous and soft-tissue metastases, at significantly higher levels than observed in normal organs and blood pool, provides a strong justification for the development of a therapeutic PSMA-targeted radiopharmaceutical based on a similar chemical scaffold, for use as a theranostic pair. ¹⁷⁷Lu-labeled CTT1403, based on the same PSMA-binding scaffold as CTT1057, is currently in preclinical testing (27) and is slated to begin clinical trials shortly.

CONCLUSION

Overall, CTT1057 is a promising novel PSMA-targeting ¹⁸F-labeled PET imaging agent based on a phosphoramidate core that is safe, without any radiotracerrelated adverse reactions. The biodistribution of CTT1057 is similar to that of other PSMA-targeted agent, and exposure rates of CTT1057 are also similar to those of the urea-based PET compounds, with the exception of lower exposure to kidneys and salivary glands, which may be significant for future therapeutic considerations. Future larger-cohort studies are needed to determine the sensitivity and specificity of CTT1057 for detection of metastases.

DISCLOSURE

This work was supported in part by the Assistant Secretary of Defense for Health Affairs, through the Prostate Cancer Research Program under Award No. W81XWH-14-1-0603 (HFV). Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the Department of Defense or U.S. Army. Research was supported by National Institutes of Health grant P41EB013598 from Cancer Targeted Technology (CTT). Spencer Behr and Thomas Hope have received a research grant from GE Healthcare. Beatrice Langton-Webster is the CEO of CTT, and Clifford Berkman is the CSO of CTT. No other potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

We acknowledge Vahid Ravanfar for his time and effort in the study.

REFERENCES

- Blomqvist L, Carlsson S, Gjertsson P, et al. Limited evidence for the use of imaging to detect prostate cancer: a systematic review. Eur J Radiol. 2014;83:1601–1606.
- Hricak H, Dooms GC, Jeffrey RB, et al. Prostatic carcinoma: staging by clinical assessment, CT, and MR imaging. *Radiology*. 1987;162:331–336.
- Scheidler J, Hricak H, Vigneron DB, et al. Prostate cancer: localization with three-dimensional proton MR spectroscopic imaging—clinicopathologic study. *Radiology*. 1999;213:473–480.
- Shinohara K, Wheeler TM, Scardino PT. The appearance of prostate cancer on transrectal ultrasonography: correlation of imaging and pathological examinations. J Urol. 1989:142:76–82.
- Udovicich C, Perera M, Hofman MS, et al. ⁶⁸Ga-prostate-specific membrane antigen-positron emission tomography/computed tomography in advanced prostate cancer: current state and future trends. *Prostate Int.* 2017;5:125–129.
- Schmidt-Hegemann NS, Fendler WP, Buchner A, et al. Detection level and pattern of positive lesions using PSMA PET/CT for staging prior to radiation therapy. *Radiat Oncol.* 2017;12:176.
- Chang SS. Overview of prostate-specific membrane antigen. Rev Urol. 2004; 6(suppl 10):S13–S18.
- Szabo Z, Mena E, Rowe SP, et al. Initial evaluation of [18F]DCFPyL for prostatespecific membrane antigen (PSMA)-targeted PET imaging of prostate cancer. *Mol Imaging Biol.* 2015;17:565–574.
- Giesel FL, Hadaschik B, Cardinale J, et al. F-18 labelled PSMA-1007: biodistribution, radiation dosimetry and histopathological validation of tumor lesions in prostate cancer patients. Eur J Nucl Med Mol Imaging. 2017;44:678–688.

- Ganguly T, Dannoon S, Hopkins MR, et al. A high-affinity [18F]-labeled phosphoramidate peptidomimetic PSMA-targeted inhibitor for PET imaging of prostate cancer. *Nucl Med Biol.* 2015;42:780–787.
- Jivan S, Neuman K, Villeret G, et al. Fully automated preparation of [18F]CTT1057, a new prostate cancer imaging agent, prepared using the ORA Neptis® Perform Synthesizer. J Labelled Comp Radiopharm. 2017;60:1.
- Starobinets O, Korn N, Iqbal S, et al. Practical aspects of prostate MRI: hardware and software considerations, protocols, and patient preparation. *Abdom Radiol* (NY), 2016;41:817–830.
- Eder M, Neels O, Muller M, et al. Novel preclinical and radiopharmaceutical aspects of [⁶⁸Ga]Ga-PSMA-HBED-CC: a new PET tracer for imaging of prostate cancer. *Pharmaceuticals (Basel)*. 2014;7:779–796.
- 14. Zacho HD, Nielsen JB, Haberkorn U, Stenholt L, Petersen LJ. ⁶⁸Ga-PSMA PET/ CT for the detection of bone metastases in prostate cancer: a systematic review of the published literature. *Clin Physiol Funct Imaging*. October 29, 2017 [Epub ahead of print].
- Afshar-Oromieh A, Zechmann CM, Malcher A, et al. Comparison of PET imaging with a ⁶⁸Ga-labelled PSMA ligand and ¹⁸F-choline-based PET/CT for the diagnosis of recurrent prostate cancer. Eur J Nucl Med Mol Imaging. 2014;41:11–20.
- Herrmann K, Bluemel C, Weineisen M, et al. Biodistribution and radiation dosimetry for a probe targeting prostate-specific membrane antigen for imaging and therapy. J Nucl Med. 2015;56:855–861.
- Kratochwil C, Bruchertseifer F, Rathke H, et al. Targeted alpha-therapy of metastatic castration-resistant prostate cancer with ²²⁵Ac-PSMA-617: dosimetry estimate and empiric dose finding. *J Nucl Med.* 2017;58:1624–1631.
- Hofman MS, Violet J, Hicks RJ, et al. [¹⁷⁷Lu]-PSMA-617 radionuclide treatment in patients with metastatic castration-resistant prostate cancer (LuP-SMA trial): a single-centre, single-arm, phase 2 study. *Lancet Oncol.* 2018; 19:825–833.

- Afshar-Oromieh A, Hetzheim H, Kubler W, et al. Radiation dosimetry of ⁶⁸Ga-PSMA-11 (HBED-CC) and preliminary evaluation of optimal imaging timing. *Eur J Nucl Med Mol Imaging*. 2016;43:1611–1620.
- Uprimny C, Kroiss AS, Decristoforo C, et al. (68)Ga-PSMA-11 PET/CT in primary staging of prostate cancer: PSA and Gleason score predict the intensity of tracer accumulation in the primary tumour. Eur J Nucl Med Mol Imaging. 2017;44:941–949.
- Park SY, Zacharias C, Harrison C, et al. Gallium 68 PSMA-11 PET/MR imaging in patients with intermediate- or high-risk prostate cancer. Radiology. 2018;288:495–505.
- Mottaghy FM, Heinzel A, Verburg FA. Molecular imaging using PSMA PET/CT versus multiparametric MRI for initial staging of prostate cancer: comparing apples with oranges? Eur J Nucl Med Mol Imaging. 2016;43:1397–1399.
- Grubmuller B, Baltzer PA, Hartenbach S, et al. PSMA ligand PET/MRI for primary prostate cancer: staging performance and clinical impact. *Clin Cancer Res.* 2018;24:6300–6307.
- Afshar-Oromieh A, Malcher A, Eder M, et al. PET imaging with a [68Ga]gallium-labelled PSMA ligand for the diagnosis of prostate cancer: biodistribution in humans and first evaluation of tumour lesions. Eur J Nucl Med Mol Imaging. 2013;40:486–495.
- Cho SY, Gage KL, Mease RC, et al. Biodistribution, tumor detection, and radiation dosimetry of ¹⁸F-DCFBC, a low-molecular-weight inhibitor of prostate-specific membrane antigen, in patients with metastatic prostate cancer. *J Nucl Med.* 2012;53:1883–1891.
- Morigi JJ, Stricker PD, van Leeuwen PJ, et al. Prospective comparison of ¹⁸F-fluoromethylcholine versus ⁶⁸Ga-PSMA PET/CT in prostate cancer patients who have rising PSA after curative treatment and are being considered for targeted therapy. *J Nucl Med.* 2015;56:1185–1190.
- Choy CJ, Ling X, Geruntho JJ, et al. ¹⁷⁷Lu-labeled phosphoramidate-based PSMA inhibitors: the effect of an albumin binder on biodistribution and therapeutic efficacy in prostate tumor-bearing mice. *Theranostics*. 2017;7:1928–1939.