Assessments at multiple levels of biological organization allow for an integrative determination of physiological tolerances to turbidity in an endangered fish species
Assessments at multiple levels of biological organization allow for an integrative determination of physiological tolerances to turbidity in an endangered fish species

Matthias Hasenbein1,2,3, Nann A. Fangue2, Juergen Geist3, Lisa M. Komoroske1,2, Jennifer Truong1, Rina McPherson1 and Richard E. Connon1,*

1Department of Anatomy, Physiology and Cell Biology, School of Veterinary Medicine, University of California, Davis, CA 95616, USA
2Department of Wildlife, Fish & Conservation Biology, University of California, Davis, CA 95616, USA
3Chair of Aquatic Systems Biology, Department of Ecology and Ecosystem Management, Technische Universität München, Mühlenweg 22, Freising D-85354, Germany

*Corresponding author: Department of Anatomy, Physiology and Cell Biology, School of Veterinary Medicine, University of California, Davis, CA 95616, USA. Tel: +1 530 752 3141. Email: reconnon@ucdavis.edu

Turbidity can influence trophic levels by altering species composition and can potentially affect fish feeding strategies and predator–prey interactions. The estuarine turbidity maximum, described as an area of increased suspended particles, phytoplankton and zooplankton, generally represents a zone with higher turbidity and enhanced food sources important for successful feeding and growth in many fish species. The delta smelt (Hypomesus transpacificus) is an endangered, pelagic fish species endemic to the San Francisco Estuary and Sacramento–San Joaquin River Delta, USA, where it is associated with turbid waters. Turbidity is known to play an important role for the completion of the species’ life cycle; however, turbidity ranges in the Delta are broad, and specific requirements for this fish species are still unknown. To evaluate turbidity requirements for early life stages, late-larval delta smelt were maintained at environmentally relevant turbidity levels ranging from 5 to 250 nephelometric turbidity units (NTU) for 24 h, after which a combination of physiological endpoints (molecular biomarkers and cortisol), behavioural indices (feeding) and whole-organism measures (survival) were determined. All endpoints delivered consistent results and identified turbidities between 25 and 80 NTU as preferential. Delta smelt survival rates were highest between 12 and 80 NTU and feeding rates were highest between 25 and 80 NTU. Cortisol levels indicated minimal stress between 35 and 80 NTU and were elevated at low turbidities (5, 12 and 25 NTU). Expression of stress-related genes indicated significant responses for gst, hsp70 and glut2 in high turbidities (250 NTU), and principal component analysis on all measured genes revealed a clustering of 25, 35, 50 and 80 NTU separating the medium-turbidity treatments from low- and high-turbidity treatments. Taken together, these data demonstrate that turbidity levels that are either too low or too high affect delta smelt physiological performance, causing significant effects on overall stress, food intake and mortality. They also highlight the need for turbidity to be considered in habitat and water management decisions.

Key words: Delta smelt, fundamental niche, habitat preference, Hypomesus transpacificus, stress

Editor: Steven Cooke

Received 28 October 2015; Revised 19 January 2016; accepted 19 January 2016

Introduction

Estuaries are among the most threatened, anthropogenically modified and managed ecosystems worldwide (Kennish, 2002, Lotze et al., 2006). This is certainly the case in the San Francisco Estuary (herein referred to as ‘Estuary’) and Sacramento–San Joaquin River Delta (herein referred to as ‘Delta’), USA, representing a paragon for such heavily influenced ecosystems (Nichols et al., 1986). Estuaries are a unique ecosystem type denoted by the interface between coastal marine and riverine freshwater habitats (Wołowicz et al., 2007) and provide a dynamic habitat for highly specialized and adapted species. Characteristic of estuaries are mixing zones, which originate from freshwater outflow, saltwater intrusion and a distinctive recurring ebb tide–flood scenario. In these zones, environmental factors such as temperature, salinity, turbidity and flow can change quickly, posing additional challenges for organisms.

Environmental factors, whether abiotic or biotic, that cause stress in an organism evoke a stress response at multiple levels of biological organization, including the level of molecules, cells, organs, organ systems and organisms (Ricklefs and Wikelski, 2002; Kassahn et al., 2009; Schulte et al., 2011; Schulte, 2014). These physiological stress parameters can be used as proxies for fitness and performance, leading to performance curves that have been used to describe tolerances and niche dimensions (Shelford, 1931; Pörtner et al., 2010; Schulte, 2014). Precise information on tolerances and niche dimensions of a species help understand its distribution, which is one of the fundamental goals of ecology and physiology.

Stress is often described as a state of threatened or disturbed homeostasis, caused by either internal or external stressors, which can be restored by a complex repertoire of adaptations comprising physiological and behavioural responses (Chrousos and Gold, 1992; Chrousos, 1998; Schulte, 2014). The physiological stress responses in fish can be categorized as primary, secondary and tertiary stress responses (Iwama, 1998; Barton, 2002; Wendelaar Bonga, 2011). The primary stress response is characterized by immediate endocrine changes in stress hormones such as catecholamines, adrenocorticotropic hormone and cortisol (Iwama, 1998; Barton, 2002; Wendelaar Bonga, 2011). The secondary response is described by the physiological effects of the primary stress hormones, in particular the activation of metabolic pathways, hydromineral balance, immune and respiratory functions (Iwama, 1998; Barton, 2002; Wendelaar Bonga, 2011). The tertiary response encompasses changes in behaviour and physiology observed at the whole-animal level, such as growth, development, reproduction, disease resistance and survival (Iwama, 1998; Barton, 2002; Wendelaar Bonga, 2011). Recent reviews have highlighted the need for a mechanistic understanding of physiological responses of fish to environmental stressors for effective fisheries management and optimal conservation efforts (Geist, 2015; Horodysky et al., 2015). It is essential to use physiological approaches and mechanistic tools to understand how environmental stressors and alterations affect species at all levels of biological organization; organisms, populations and ecosystems (Wikelski and Cooke, 2006; Cooke et al., 2013).

In the Estuary and Delta, severe declines in the abundance of several fishes have been documented over the past decades; referred to as the pelagic organism decline (Sommer et al., 2007; Baxter et al., 2008, 2010; Thomson et al., 2010; Brooks et al., 2012). Most affected are two introduced species, striped bass (Morone saxatilis) and threadfin shad (Dorosoma petenense), and two native species, longfin smelt (Spirinchus thaleichthys) and delta smelt (Hypomesus transpacificus; Sommer et al., 2007; Thomson et al., 2010). The delta smelt is a pelagic fish species endemic to the Estuary and Delta and is listed as endangered and threatened under California State and Federal Endangered Species Acts, respectively (USFWS, 1993; CDFW, 2014). The observed decline in delta smelt abundance has led to the detection of genetic bottlenecks in years 2003, 2005 and 2007 (Fisch et al., 2011). Delta smelt abundance has been associated with the low salinity zone (Moyle et al., 1992) as well as with turbid water (Feyrer et al., 2007; Grimaldo et al., 2009; Bennett and Burau, 2015). The low salinity zone is centred around a salinity of 2 practical salinity units (PSU; Jassby et al., 1995; Kimmerer et al., 2013), with a variation in salinity of 0.5–6 PSU (Brown et al., 2014), and is also characterized by the estuarine turbidity maximum (ETM; Peterson et al., 1975; Schoellhamer, 2001). The ETM is described as an area of increased suspended particles, phytoplankton and zooplankton (Peterson et al., 1975; Roman et al., 2001; Sanford et al., 2001), thus representing a zone with higher turbidity and enhanced food sources, which is important for successful feeding and growth in many fish larvae (Dodson et al., 1989; North and Houde, 2001). The turbidity in an ETM can be extremely variable and change quickly, e.g. shifts in turbidity levels from 20 to >150 NTU within 1 h have been observed in the ETM of the St Lawrence Estuary, Canada (Dodson et al., 1989).

Turbidity, often described as the cloudiness or murkiness of water, is defined as an expression of the optical property causing light scattering and light absorption rather than direct light transmission through a water sample (Rice et al., 1994). Important factors contributing to the effect of turbidity are light intensity, suspended material and water depth (Lee and Rast, 1997), and these parameters interact with each other. In particular, the type of suspended material and its specific particle size, morphology, colour and refractive index influence the extent of light scattering and can affect vision and respiration at high densities (reviewed by Bruton, 1985). Along with other water physicochemical parameters, turbidity plays an important role in estuarine ecosystems. It is known, for example, to influence several trophic levels by altering species composition (Lunt and Smee, 2014) and can potentially affect the feeding strategy of fish (Hecht and Van der Lingen, 1992; De Robertis et al., 2003; Johansen and Jones, 2013), as well as predator–prey interactions. Much research has been
Turbidity is crucial to the completion of the delta smelt’s life cycle. Delta smelt have been reported to use turbid waters to hide from predators (Moyle, 2002), and it has been hypothesized that increased pulse turbidity from first flush events (first winter storm event of the year) might be a cue for the annual spawning migration of delta smelt to the northern Delta (Grimaldo et al., 2009; Sommer et al., 2011). Furthermore, laboratory studies on the feeding response of early life stages of delta smelt found a positive relationship between turbidity and feeding behaviour (Baskerville-Bridges et al., 2004, 2005), and for juveniles (120 days post-hatch; dp) constant feeding up to 120 NTU with a significant decrease in food intake at 250 NTU was observed (Hasenbein et al., 2013). However, there is still a lack of information on turbidity tolerance ranges for delta smelt, because postulated associations, as they relate to this species of concern, have not been fully evaluated. There is a great need to identify tolerance ranges of delta smelt for turbidity in order to manage water diversions and water pumping more efficiently. Knowing tolerance ranges for this sensitive species will help managers make informed decisions about delta smelt conservation. Moreover, knowledge of the turbidity preference is also crucial for conservation programmes that aim at captive breeding of this species, as well as for maintaining optimal holding conditions in ecological and ecotoxicological studies involving this species.

The aim of the present study was to evaluate short-term effects of environmentally relevant turbidities (5–250 NTU) on the physiological stress responses of late-larval delta smelt, with the goal of determining preferred turbidity levels and tolerance ranges under well-defined conditions. We focused on the larval life stage because it has been previously shown to be the most sensitive life stage (Connon et al., 2009, 2011a,b; Komoroske et al., 2014), dependent on turbidity for feeding (Baskerville-Bridges et al., 2004; Lindberg et al., 2013) and other aspects of life history (Lindberg et al., 2013), and tolerance to turbidity has not been tested. We hypothesized that different turbidity levels would affect overall physiological stress, which would in turn affect survival, but that maximal prey capture ability (feeding) would occur at specific turbidity requirements. Thus, in order to determine the preferred range of turbidities for late-larval delta smelt, we conducted integrative assessments using a combination of physiological endpoints (gene transcription, plasma hormones and metabolites) and contrasted these with ecological performance measures (food intake and survival).

Materials and methods

Study animals

Late-larval delta smelt (H. transpacifcicus; 60 dp) were provided by the Fish Conservation and Culture Laboratory (FCCL) UC Davis in Byron, CA, USA, where feeding ability and physiological response tests were conducted. Tests were conducted during 12–16 August 2013. The start and end of the test were mornings at 9.00 h. Fish were cultured according
to culture protocols described by Baskerville-Bridges et al. (2005) and Lindberg et al. (2013). In brief, fish were cultured in black tanks without substrate, plants or structure, because this resembles their natural habitat in the open pelagic zone. A recirculating system connected to biofilters was used to maintain larvae with a feeding regimen of six feedings per day. Larvae were fed with newly hatched Artemia franciscana at a volume in the tank of 1–3 nauplii ml⁻¹. The average rearing temperature throughout the rearing period was 17.4°C (±0.05 SE). The mean length (fork length) and weight of 60 dph delta smelt were 19.37 mm (±0.09 SE) and 0.0303 g (±0.00 SE), respectively. Fish were kept at turbidities of 9.94 NTU (±0.25 SE) for a period of 8 weeks before the test. The light intensities for larval and late-larval life stages are kept at low levels of 4–5 and 1–2 µmol m⁻² s⁻¹, respectively (Lindberg et al., 2013). The low light level accommodates for the increased light sensitivity of this fish species (Lindberg et al., 2013).

**Fish exposures**

Studies were set up to evaluate feeding success and physiologically stress responses to varying turbidities. Late-larval delta smelt (60 dph) were exposed, at a stocking density of 30 fish per vessel, to turbidities of 5, 12, 25, 35, 50, 80, 120 and 250 NTU (nominal values) in aerated facility water in 8 l black circular fish tanks (static water system; 2 Gallon Black Plastic Pail, Item # 3539; United States Plastic Corporation, USA), for a period of 24 h with a light–dark cycle of 16 h–8 h. Black exposure vessels were used because they are considered a necessary component for successful rearing and culture (Lindberg et al., 2013). Fish stocking density was determined in earlier studies (Hasenbein et al., 2016) to be in the optimal range of four to eight fish per litre. Salinity was kept constant at 2 PSU for the feeding test, whereas the physiology test had a salinity of 0.2 PSU. Nannochloropsis algae (Nanno 3600 – High yield grow out feed; Reed Mariculture Inc., USA) were spiked into exposure vessels to achieve desired turbidities. Light was provided by a Lithonia Lighting Fluorescent Luminaire (Bulb: Philips F40T12/DX, 40 W), and light intensity was kept constant at a low level of 48 lx (±1.13 SE). The exposure vessel and light intensity were chosen according to culture protocols (Lindberg et al., 2013). Fish tanks were aerated using air stones with a constantly low aeration rate to keep algae suspended. Aeration had to be limited because culture methods recommend a gentle aeration to account for early life-stage sensitivity (Baskerville-Bridges et al., 2005). Fish were fed Artemia in the facility tanks prior to test set-up, but were kept unfed throughout the duration of the test.

For both tests, physicochemical water parameters were monitored at test initiation and at termination. Dissolved oxygen was measured using a YSI Model 55 DO meter (YSI Inc., Xylem Inc., Yellow Springs, OH, USA). Salinity and specific conductance were measured using a YSI Model 63 Multimeter (YSI Inc., Xylem Inc., Yellow Springs, OH, USA). pH was measured using a waterproof portable pH meter (Hanna Instruments Model HI9124; Hanna Instruments Inc., Woonsocket, RI, USA). Turbidity was measured with a Hach 2100q portable turbidity meter (Hach Company, Loveland, CO, USA) that conducts a ratio turbidimetric determination using a primary nephelometric light scatter signal (90° angle) to the transmitted light scatter signal. A water sample was taken from the exposure vessel, shaken for 10 s and placed into the turbidity meter. Ammonia was measured using a Hach pocket colorimeter II Filter Photometer (Hach Company). Ammonia was determined in control treatments only, because the color of the Nannochloropsis algae interfered with the measurement method. Temperature was measured at 1 min intervals using iBCod submersible temperature loggers (Alpha Mach Inc., Ste-Julie, QC, Canada). Light levels were measured directly above the water surface of the exposure vessel using an Exttech instruments easy view 30 light meter (FLIR commercial system Inc., Nashua, NH, USA).

**Feeding test**

After 24 h, subsets of fish were fed with live newly hatched Artemia franciscana in solution [100 ml per bucket; density of 462 Artemia per ml (+21 SE); total food density in the vessel was five to six Artemia per millilitre of water] for a period of 7.5 min (duration determined in preliminary tests to result in 50% gut fullness; M. Hasenbein, R. E. Connon, N. A. Fangue and J. Geist, unpublished data). Artemia franciscana was chosen as a food item because this is the same as used in the culture procedures. At test termination, dead fish were identified, counted and discarded. Surviving fish were immediately euthanized with an overdose of tricaine methanesulfonate (MS-222; Finquel, USA) buffered with sodium bicarbonate. Specimens were transferred into 15 ml tubes filled with 70% ethanol and preserved until further processing for stomach content analysis. Fish were measured for weight and length and dissected for gut content (number of Artemia ingested) under a microscope (×40 resolution). Survival was recorded at test termination and the percentage of survival was calculated based on fish numbers at test set-up and test termination. Tests were conducted in quadruplicate, except for the 120 NTU treatment, which had only three replicates owing to vigorous aeration in one of the replicates resulting in elevated mortality.

**Physiology test**

At test termination, subsets of unfed fish were immediately euthanized as described in the previous subsection. Specimens were transferred into 1.5 ml tubes (Eppendorf) and subsequently snap frozen in liquid nitrogen. Samples were stored at −80°C for subsequent biochemical and molecular analyses.

**Cortisol assessments**

Methods for cortisol measurements are described by Hasenbein et al. (2016). In brief, whole-body cortisol was assessed in a total of four to seven fish from each treatment depending on survival, using modified methods established for zebrafish (Alsop and Vijayan, 2008; Cachat et al., 2010). Volumes of solutions were optimized for use in juvenile delta smelt (Hasenbein et al., 2013) and further enhanced for larval fish. Samples were defrosted on ice and homogenized in 1x phosphate-buffered saline (PBS) using a TissueLyzer LT. The
resulting homogenate was divided into equal amounts of 500 µl and used for cortisol and total protein determination. Cortisol was eluted from the homogenate using diethyl ether. Ether was then allowed to evaporate, and dried cortisol samples were resuspended in 1× PBS. Cortisol assessments were performed according to manufacturer’s instructions (Salivary Cortisol, Enzyme Immunoassay Kit; Salimetrics, Inc., State College, PA, USA), and levels (in milligrams per decilitre) were calculated with a four-parameter sigmoid standard curve (minus curve fit). Cortisol levels were normalized to total protein and denoted as cortisol concentration (picograms of cortisol per microgram of protein). The second half of the homogenate was used to determine protein content. After centrifugation at 16 500g for 30 min at 4°C, the supernatant of each sample was collected and used for total protein content determination following the manufacturer’s protocol (BCA Protein Assay Kit; Thermo Fisher Scientific Inc., Waltham, MA, USA).

**Quantitative polymerase chain reaction**

Total RNA was extracted from whole-body homogenates. RNA extractions were performed according to the manufacturer’s protocols using the RNeasy Mini Qiacube Kit (Qiagen®, Venlo, Limburg, The Netherlands). Qualitative and quantitative RNA determination was conducted using a NanoDrop ND1000 Spectrophotometer; 260/280 and 260/230 ratios ranged from 2.04 to 2.18 and from 1.94 to 2.35, respectively. Integrity of total RNA was assessed by electrophoresis on a 1% (w/v) agarose gel. Depending on survival, the stress response and physiology gene transcription of five to eight fish per treatment were assessed by quantitative polymerase chain reaction (qPCR). Complementary DNA (cDNA) synthesis was performed using Reverse Transcriptase III (SuperScript® III Reverse Transcriptase; InvitrogenTM, Carlsbad, CA, USA), and primers and probes for qPCR analyses were designed using Roche Universal Library Assay Design Center [https://www.roche-applied-science.com](https://www.roche-applied-science.com). Quantitation of transcription was performed using SDS 2.4 software (Applied Biosystems®, Life technologiesTM). Responding genes were normalized using a normalization factor calculated based on the geometric mean of two control genes, namely *glyceraldehyde-3-phosphate dehydrogenase* (*gapdh*) and *beta-actin* (*b-actin*). Normalization was performed according to the ‘geNorm’ algorithm version 3.5 as described by Vandesompele et al. (2002).

A total of 17 target genes were selected based on their involvement in the hypothalamic–pituitary–interrenal axis and in other important functions, such as energy metabolism, development and somatic growth, ion homeostasis, oxygen homeostasis, inflammatory response, osmoregulation and general stress. Selected genes (gene name and code), function, primer sequence and efficiency are listed in Table 1.

**Statistical analysis**

Data were analysed using the ‘stats’ package in R-project for statistical computing (version 3.0.2; [http://www.r-project.org; R-CoreTeam, 2014](http://www.r-project.org)). All data sets were tested for normal distribution and homoscedasticity using the Shapiro–Wilks normality test and the Fligner–Killeen test. Cortisol data were logarithmically transformed to meet normality criteria. Data were tested for effects of turbidity using a one-way ANOVA if normally distributed or a Kruskal–Wallis test if non-normally distributed. Turbidity was defined as the predictor (categorical variable) and the respective response variable (feeding, mortality, cortisol or qPCR) as the continuous variable. Where turbidity showed a significant effect, data were analysed for post hoc contrasts between treatments using Tukey’s HSD test or the multiple comparison test after Kruskal–Wallis test. Statistical decisions were based on an α level of 0.05. Further statistical information is presented in supplementary Table S1.

Principal component analysis (PCA) was carried out on the transcriptomic (qPCR) data set in order to analyse differences in transcription patterns between treatments and to determine to what extent these profiles were affected by the treatments. The PCA scores were calculated using the covariance matrix, and principal components 1, 2 and 3 (PC1, PC2 and PC3) were determined to explain the majority of the variation in the data, using a Scree test as described by D’Agostino and Russell (2005). PCA scores of PC1, PC2 and PC3 were plotted as a centroid graph to describe, interpret, visualize and support clustering of treatments with similar transcriptomic responses. In addition, the respective biplot was plotted to identify genes driving the two components and explain the percentage variation.

**Results**

**Physicochemical parameters**

Physicochemical water parameters for the feeding test and the physiology test are presented in Tables 2 and 3, respectively. Specific conductance, pH, salinity and temperature remained stable throughout the test duration. Dissolved oxygen declined slightly over time in all turbidity levels in the feeding test and in all but one turbidity level of the physiology test. Values ranged over time for the feeding test from 9.79 to 7.44 mg l⁻¹ and for the physiology test from 9.53 to 7.75 mg l⁻¹, respectively. At all turbidity levels of the feeding test and at turbidity levels 25, 35, 50, 80, 120 and 250 NTU of the physiology test, a decrease in turbidity was observed at the 24 h time point because delta smelt require only gentle aeration (Table 3). Ammonia concentration increased over time from 0.08 mg l⁻¹ (±0.01 SE) at test initiation to 0.15 mg l⁻¹ (±0.00 SE) at test termination.

**Feeding test**

**Survival**

Mean percentage survival was 81% (±3.43 SE) across all treatments (Fig. 1). Highest survival rates were observed in treatments 12, 25, 35, 50 and 80 NTU, with survival rates of 90 (±4.71 SE), 93 (±3.60 SE), 87 (±2.50 SE), 88 (±3.18 SE) and 88% (±3.69 SE), respectively. Survival decreased at low turbidities of 5 NTU, averaging 73% (±3.94 SE), as well as at
Table 1: Primer and probe sequences of genes used as molecular biomarkers to determine stress levels in late-larval delta smelt (Hypomesus transpacificus)

<table>
<thead>
<tr>
<th>Gene name</th>
<th>Gene code</th>
<th>Primer sequences</th>
<th>Function</th>
<th>Probe no.</th>
<th>Percentage efficiency</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glutathione-S-transferase</td>
<td>gst</td>
<td>5’→ 3’ AATCTCCGTCGACATGTTG</td>
<td>Cellular detoxification, osmotic stress</td>
<td>127</td>
<td>108</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ GGCGGCTCTTCAAAACAT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mineralocorticoid receptor 1</td>
<td>mr1</td>
<td>5’→ 3’ TTTCATCATTCCGGCGAGCTCA</td>
<td>Mediator for cortisol signalling; HPI axis</td>
<td>39</td>
<td>99</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ TGATGATCCTCCACGACATCCT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glucocorticoid receptor 2</td>
<td>gr2</td>
<td>5’→ 3’ CATCGTAAAGCTGAGGAGAA</td>
<td>Mediator for cortisol signalling; HPI axis</td>
<td>129</td>
<td>98</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ GCAGTTGAGCCCGGTTGTC</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pro-opiомelanocortin</td>
<td>pomc</td>
<td>5’→ 3’ TGCACCTGTCAGGGTCTGGA</td>
<td>Precursor of adrenocorticosteroid hormone, regulates glucose homeostasis</td>
<td>127</td>
<td>102</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ GAGAAGCTCTCCTCTGGGGA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>11-beta-hydroxyteroid-dehydrogenase type 1</td>
<td>11-beta-</td>
<td>5’→ 3’ CTTCTGACATGACTACAAGAC</td>
<td>Cortisol conversion; HPI axis</td>
<td>55</td>
<td>109</td>
</tr>
<tr>
<td>hsd-1</td>
<td></td>
<td>3’→ 5’ TCTGGACAGCGTGGTGAAGCT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>11-beta-hydroxyteroid-dehydrogenase type 2</td>
<td>11-beta-</td>
<td>5’→ 3’ TCTGGACATGACTACAAGAC</td>
<td>Cortisol conversion; HPI axis</td>
<td>14</td>
<td>106</td>
</tr>
<tr>
<td>hsd-2</td>
<td></td>
<td>3’→ 5’ TCTGGACAGCGTGGTGAAGCT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Beta actin</td>
<td>β-actin</td>
<td>5’→ 3’ TGCCACAGGACTCCATAC</td>
<td>Housekeeping gene, reference gene</td>
<td>11</td>
<td>107</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ CATCAGGCTCAGGAGGAGG</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glyceraldehyde 3-phosphatdehydrogenase</td>
<td>gapdh</td>
<td>5’→ 3’ TCCACGAGAAGACACCAA</td>
<td>Housekeeping gene, reference gene</td>
<td>159</td>
<td>95</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ CAGGAGTACACGACAA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Insulin like growth factor</td>
<td>igf</td>
<td>5’→ 3’ GACACCGTGCAAGTTGTATG</td>
<td>Development, somatic growth, interaction with growth hormone</td>
<td>110</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ CATAGGCTGCGGTTGTTGT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Serum/glucocorticoid regulated kinase</td>
<td>sgk3</td>
<td>5’→ 3’ TTATGAGATCAAAGGCAATGAC</td>
<td>Ion homeostasis, stimulates sodium transport via epithelial sodium channel</td>
<td>85</td>
<td>107</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ GGTGTGAAAGGAGGGAAGGTGA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hypoxia inducible factor 1 alpha</td>
<td>hif1a</td>
<td>5’→ 3’ GGCATGGCCAGGCTCCCTTA</td>
<td>Transcription factor, oxygen homeostasis</td>
<td>41</td>
<td>105</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ ATTAGCTGTTGGCACATAGC</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glucose transporter 2</td>
<td>glut2</td>
<td>5’→ 3’ GGCATGTCAGTTGCGCTCAT</td>
<td>Glucose homeostasis, energy metabolism</td>
<td>130</td>
<td>109</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ GACATGTCAGTGAGTAGTCATCC</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Heat shock protein 70 kD</td>
<td>hsp70</td>
<td>5’→ 3’ AAGATTCTGGGAAAGTGCAGACAGG</td>
<td>General stress, heat stress</td>
<td>20</td>
<td>107</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ CTTCTCTGAGCGCTGGTG</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Nuclear factor k-beta</td>
<td>nfkb</td>
<td>5’→ 3’ TGACAGGATGAAACATTTGTC</td>
<td>Transcription factor, host defense, chronic inflammatory diseases</td>
<td>123</td>
<td>103</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ CCAAAGTCCAGAGGCTTTGTA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ammonium transporter</td>
<td>NH4 trans</td>
<td>5’→ 3’ CAGGTCGTCTTATGCTGAGG</td>
<td>Excess ammonia elimination across gills and skin</td>
<td>61</td>
<td>95</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ CAGGTCGTCAAGTTACACAGG</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Catalase</td>
<td>catalase</td>
<td>5’→ 3’ GCACAGGAGCAATGAAACCTTTA</td>
<td>Antioxidant enzyme, protects cell from oxidative damage by reactive oxygen species</td>
<td>88</td>
<td>97</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ GGTGCGGAGTATCTGGTCA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sodium potassium ATPase</td>
<td>Na+K atp</td>
<td>5’→ 3’ GTTCATCCAAATCTAGCTCC</td>
<td>Ion transport during osmoregulation, osmoregulatory stress</td>
<td>88</td>
<td>109</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3’→ 5’ CATGATGTCGCAAATCTCT GC</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Abbreviations: HPI axis, hypothalamic–pituitary–interrenal axis.

high turbidities of 120 and 250 NTU, averaging 71 (±4.01 SE) and 58% (±6.16 SE), respectively.

Feeding

Highest feeding rates (number of Artemia ingested) were recorded at mid-range turbidities of 25, 35, 50 and 80 NTU (Fig. 2), where late-larval delta smelt ingested a mean of 17 (±4.40 SE) Artemia per fish at 25 NTU, 25 (±5.08 SE) at 35 NTU, 19 (±4.75 SE) at 50 NTU and 22 (±4.19 SE) at 80 NTU. Lower feeding rates were observed at both low and high turbidities, with prey ingestions of 8 (±2.68 SE) and 11 (±2.17 SE) Artemia per fish at turbidities of 5 and 12 NTU, respectively, and 14 (±4.81 SE) and 5 (±2.07 SE) Artemia per fish at turbidities of 120 and 250 NTU, respectively.
### Table 2: Physicochemical water parameters of the turbidity feeding test conducted on late-larval delta smelt 60 days post-hatch exposed to different levels of turbidity over 24 h

<table>
<thead>
<tr>
<th>Treatment turbidity (NTU) nominal concentration</th>
<th>Average DO (mg l(^{-1}))</th>
<th>Average Spec. Con. (µS cm(^{-1}))</th>
<th>Average pH</th>
<th>Average Salinity (PSU)</th>
<th>Average Turbidity (NTU) measured concentration</th>
<th>Average Ammonia (mg l(^{-1})) NH(_3)-N</th>
<th>Average Light intensity (lx)</th>
<th>Average Temperature (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5 NTU</td>
<td>Average 9.79 8.84</td>
<td>3738 3804</td>
<td>7.65 7.22</td>
<td>2.00 2.03</td>
<td>6.18 5.16</td>
<td>Average 0.08 ± 0.01 SE</td>
<td>Average 48.07 ± 1.13 SE</td>
<td>Average 17.52 ± 0.00 SE</td>
</tr>
<tr>
<td>SE</td>
<td>0.05 0.05</td>
<td>82 102</td>
<td>0.07 0.04</td>
<td>0.00 0.05</td>
<td>0.22 0.25</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>12 NTU</td>
<td>Average 9.48 8.88</td>
<td>3712 3852</td>
<td>7.82 7.32</td>
<td>2.05 2.05</td>
<td>11.70 10.6</td>
<td>Average 0.15 ± 0.00 SE</td>
<td>Average 10.00 ± 0.00 SE</td>
<td></td>
</tr>
<tr>
<td>SE</td>
<td>0.05 0.08</td>
<td>238 150</td>
<td>0.06 0.02</td>
<td>0.09 0.09</td>
<td>0.20 0.20</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>25 NTU</td>
<td>Average 9.55 8.64</td>
<td>3721 3732</td>
<td>7.89 7.48</td>
<td>1.98 1.98</td>
<td>24.20 19.90</td>
<td>Average 4.80 ± 0.80 SE</td>
<td>Average 0.80 ± 0.70</td>
<td></td>
</tr>
<tr>
<td>SE</td>
<td>0.03 0.15</td>
<td>129 141</td>
<td>0.03 0.04</td>
<td>0.08 0.08</td>
<td>1.00 0.80</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>35 NTU</td>
<td>Average 9.54 8.59</td>
<td>3941 3939</td>
<td>7.88 7.47</td>
<td>2.10 2.10</td>
<td>34.80 27.30</td>
<td>Average 34.80 ± 7.30 SE</td>
<td>Average 0.80 ± 0.70</td>
<td></td>
</tr>
<tr>
<td>SE</td>
<td>0.06 0.11</td>
<td>69 72</td>
<td>0.02 0.02</td>
<td>0.04 0.04</td>
<td>0.80 0.70</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>50 NTU</td>
<td>Average 9.45 8.42</td>
<td>3699 3743</td>
<td>7.87 7.47</td>
<td>1.93 2.00</td>
<td>51.30 43.80</td>
<td>Average 51.30 ± 8.30 SE</td>
<td>Average 0.80 ± 0.70</td>
<td></td>
</tr>
<tr>
<td>SE</td>
<td>0.15 0.11</td>
<td>24 45</td>
<td>0.04 0.01</td>
<td>0.03 0.00</td>
<td>1.00 0.80</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>80 NTU</td>
<td>Average 9.51 8.35</td>
<td>3932 3887</td>
<td>7.93 7.53</td>
<td>2.05 2.05</td>
<td>80.70 58.20</td>
<td>Average 80.70 ± 8.20 SE</td>
<td>Average 0.80 ± 0.70</td>
<td></td>
</tr>
<tr>
<td>SE</td>
<td>0.10 0.24</td>
<td>65 118</td>
<td>0.07 0.03</td>
<td>0.06 0.06</td>
<td>1.40 1.60</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>120 NTU</td>
<td>Average 9.57 8.44</td>
<td>3887 3845</td>
<td>7.90 7.52</td>
<td>2.07 2.07</td>
<td>120.97</td>
<td>Average 120.97 ± 9.70 SE</td>
<td>Average 2.5 ± 2.5</td>
<td></td>
</tr>
<tr>
<td>SE</td>
<td>0.04 0.14</td>
<td>129 174</td>
<td>0.01 0.05</td>
<td>0.07 0.09</td>
<td>2.5</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>250 NTU</td>
<td>Average 9.24 7.44</td>
<td>3760 3909</td>
<td>8.03 7.55</td>
<td>2.05 2.08</td>
<td>251.191</td>
<td>Average 251.19 ± 19.91 SE</td>
<td>Average 2.5 ± 2.5</td>
<td></td>
</tr>
<tr>
<td>SE</td>
<td>0.08 0.21</td>
<td>188 62</td>
<td>0.08 0.03</td>
<td>0.06 0.05</td>
<td>2.4</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Values are means with SE across replicates for each turbidity level taken at test initiation T0 and test termination T24.

Abbreviations: DO, dissolved oxygen; NTU, nephelometric turbidity unit; PSU, practical salinity unit; Spec. Con., specific conductance; T0, time point at test start; T24, time point at test termination.
Significant differences between feeding rates were detected between 35 and 250 NTU ($P < 0.05$). No ingested algae were found in fish guts.

**Physiology test**

**Cortisol**

No statistically significant differences in whole-body cortisol levels were observed in delta smelt maintained at turbidities ranging from 5 to 250 NTU (Fig. 3). However, cortisol values tended to be lowest in fish maintained at 50 NTU and above, whereas they were elevated and more variable in fish held at the lower turbidity levels of 5, 12 and 25 NTU.

**Quantitative PCR**

Changes in gene transcription for all 15 genes measured with qPCR are presented in Table 4 and in Supplementary Table S2. Four out of 15 genes, namely $gst$, $hsp70$, $glut2$ and $NH4\ trans$, responded significantly in delta smelt (Table 4). $gst$ was significantly differentially expressed at the highest turbidity (250 NTU), relative to 25 ($P < 0.05$) and 35 NTU ($P < 0.05$), suggesting oxidative stress at higher turbidity. $hsp70$ indicated highest general stress levels at 120 and 250 NTU, significantly different from the 5 ($P < 0.01$), 12 ($P < 0.001$) and 35 NTU ($P < 0.05$) groups. No significant differences were observed at 25 NTU, owing to high variability in responses. $glut2$ was also highest at high turbidity, significantly differentiating between 250 and 5 NTU ($P < 0.01$), 35 ($P < 0.05$) and 80 NTU.
(P < 0.05). Interestingly, NH4 trans, a gene coding for the ammonia transporter, also responded significantly in fish maintained at 12 NTU relative to 120 (P < 0.001) and 250 NTU (P < 0.05).

Principal component analysis

Principal component analysis was conducted on all gene transcription data (Table 4 and Fig. 4). Principal component 1 explained 40.40% of the variation, whereas PC2 and PC3 explained 19.60 and 10.80% of the variation, respectively, leading to a cumulative variation of 70.80% (Fig. 4). Plotting PC1, PC2 and PC3 against each other for visual evaluation revealed trends in differentiation and clustering of treatments with similar transcriptomic responses. The plot for PC1 vs. PC2 (Fig. 4a) clustered treatments 25, 35, 50 and 80 NTU more closely with lower turbidity treatments (5 and 12 NTU) than with higher turbidity treatments (120 and 250 NTU). The respective biplot (Fig. 4b) indicates that several different genes, such as gr2, mr1, 11-beta-hsd-1, glut2, gst and catalase, loaded heavily on PC1, whereas pome, bif1a, sgk3, hsp70 and NH4 trans determined the loading for PC2. Differentiation of the highest turbidities (120 and 250 NTU) from all others tested was more pronounced when plotting PC1 vs. PC3 (Fig. 4c). In contrast, the plot for PC2 vs. PC3 (Fig. 4d) highlighted a more pronounced differentiation of the lowest turbidity treatments from mid-range and high turbidity treatments.

Discussion

In this study, the effects of different levels of turbidity on late-larval delta smelt survival, feeding and physiological stress responses were quantified over a period of 24 h. Overall, turbidities from 25 to 80 NTU were determined to be the optimal range in the tested conditions as evident from the highest survival, feeding and changes in gene expression compared with other treatments. Detrimental effects occurred both at low turbidities of 5 and 12 NTU and at high turbidities of 120 and 250 NTU. Consequently, even short-term (< 24 h) exposure to such conditions is likely to have significant adverse effects on delta smelt.

How turbidity affects an organism is influenced by light intensity, water depth and the type of suspended material (Lee and Rast, 1997); these factors can vary in the field depending on local habitat conditions, but can be controlled in laboratory assessments. Standardized turbidity loading experiments in the laboratory are conducted using several different types of particles, e.g. soil (Mussen et al., 2012), bentonite (Vogel and Beauchamp, 1999; Quesenberry et al., 2007), red clay (Swenson et al., 2012) and sand (Beauchamp, 1999).
and Matson, 1976), clay (Ardjosedoiro and Rammarine, 2002), natural sediment (Sirois and Dodson, 2000) and planktonic algae (Ajemian et al., 2015). Although these experiments are not perfect analogues of field conditions, our data correspond well with field observations where delta smelt are associated with specific water turbidity levels (10–50 NTU; Feyrer et al., 2007; Grimaldo et al., 2009; Bennett and Burau, 2015).

Delta smelt survival was highest at mid-range turbidities, but reduced at very low and high turbidities. The reduced survival at low turbidities may be explained by elevated light intensity resulting in increased stress levels (supported by trends in corticosteroids, which is possible in natural environments. Turbidity and light intensity are therefore likely to be key factors determining their location in the water column. Another potential cause for the elevated stress and reduced survival observed at low turbidity levels could stem from the concept that elevated turbidity is beneficial for planktivorous fish, in order to hide from predators. In fact, delta smelt are postulated to use turbid waters to hide from predators (Moyle, 2002). The ‘turbidity as a cover’ hypothesis as described by Lehtiniemi et al. (2005, and references therein) might play an important role in explaining the results of the present study. Although turbid conditions might minimize the perceived risk of predation, some predators might be able to take advantage of reduced anti-predator behaviours in these conditions.

Reduced survival at high turbidities could be caused by altered gill function, reducing the fish’s capacity for respiration (Mallatt, 1985; Evans et al., 2005; Hess et al., 2015). Clogging of fish gill rakers and gill filaments as a result of excess suspended material is not uncommon (Bruton, 1985; Sutherland and Meyer, 2007; Wong et al., 2013). Although not determined in the present study, it is possible that algal material restricted respiration by filling the fish’s gill cavities or affecting gill morphology, influencing function. The increased expression of gat at high turbidity in delta smelt in this test is an indication of oxidative stress, which could also result in osmotic imbalance, as well as overall effects on respiration. Highest survival rate across mid-range turbidities (12–80 NTU) indicates the optimal range for late-larval delta smelt in test conditions. However, increased stress levels and reduced feeding were determined in treatment 12 NTU, where mortality was low, indicating sub-lethal stress levels in fish exposed to this treatment in test conditions.

Feeding responses corresponded to survival as expected. A constantly high feeding rate was observed between 25 and 80 NTU, with a peak at 35 NTU. A similar result was found in studies on European smelt that tested the feeding response...
to turbidities and light intensities; determining highest feeding rates at 20 NTU and a constant feeding between 20 and 50 NTU (Horppila et al., 2004). This feeding pattern is typical for planktivorous fish, which benefit from turbid waters by a contrast enhancement that helps them to detect prey items (Boehlert and Morgan, 1985; Utne-Palm, 2002). Studies on larvae of Pacific herring (Clupea harengus pallasi) that assessed the feeding response to different concentrations of suspended sediments, found increased feeding rates at intermediate concentrations compared with reduced feeding rates in control treatments (no suspended sediments) and at very high concentrations (Boehlert and Morgan, 1985). Increased stress can lead to reduced food intake and food conversion rate in fish (Wendelaar Bonga, 2011), thus the observed low feeding rates at low turbidity levels may be related to increased stress levels, indicated by trends in cortisol. However, low feeding rates could also be attributed to backscattering of light. Light intensities above the light saturation level of the fish’s physiological capacities (sensitivity and vision) cause negative effects on prey detection (Utne-Palm, 2002) by minimizing the contrast between the background and the prey item, thus making it less visible (Cerri, 1983; Guthrie, 1986; Loew and McFarland, 1990). This would reduce the ability of the delta smelt to find prey, leading to decreased feeding rates. At high turbidities, reduced feeding is also likely owing to elevated stress levels, and this is consistent with assessed molecular indices of oxidative and osmotic stress. As with backscattering of light at low turbidity, reduction in feeding at high turbidity may also be caused by an impaired field of vision. Even though larval and juvenile delta smelt are planktivorous feeders, it is highly likely that they cope with a wide range of turbidities; however, if turbidities reach levels where visibility is impaired, prey detection may become impossible. We demonstrated reduced feeding at high turbidities (250 NTU) in prior studies on juvenile delta smelt (Hasenbein et al., 2013). Furthermore, high turbidities can reduce the reactive distance of a fish (Hecht and Van der Lingen, 1992; Miner and Stein, 1996; Utne, 1997; Utne-Palm, 2002) and with that, limit the volume of water that a fish could search for prey. The observed feeding in 120 and 250 NTU would therefore be resultant of incidental prey encounters at close proximities. Feeding assessments were conducted using A. franciscana, which is used in the culture procedures, and fish were fed in abundance. It is likely that feeding results presented here would be qualitatively similar if natural prey items were used, but absolute feeding performance values may differ markedly as a result of factors such as differential prey mobility and escape responses. Larval fish are constantly swimming and do not perform a coordinated hunt, rather feeding when directly encountering prey items (Bennett, 2005). In future studies, feeding performance could be explored further by using numerous prey species, including numerous zooplankton species that are commonly consumed in their natural diets.

Trends in cortisol and molecular biomarker data corresponded to both survival and feeding. A non-significant but detectable elevation in cortisol levels at 5, 12 and 25 NTU suggests higher stress in these treatments. As discussed above, at the lower turbidity levels (5 and 12 NTU), light sensitivity along with the confined water depth of the exposure vessel may have played important roles. Turbidity ranges between 35 and 80 NTU resulted in lower levels of cortisol in late-larval delta smelt. These trends support what was observed in the survival and the feeding response, indicating a similar range as favourable. Reduced cortisol levels observed at the highest turbidity (250 NTU) are likely to be associated with impairment of respiration; elevated mortality resulted in these treatments. The highest turbidity level (250 NTU) unexpectedly resulted in low cortisol levels, but gene expression responded significantly [e.g. gst indicative of oxidative and osmotic stress (respiration), hsp70 involved in general stress and glut2 involved in energy metabolism]. Taken together, this suggests that turbidities above 120 NTU do not necessarily evoke a primary stress response after 24 h, but secondary stress responses are at play, as indicated by the molecular biomarkers.

Increased stress also affects a number of organismal parameters owing to reallocation of metabolic energy. When stressed organisms spend more energy on regaining homeostasis (e.g. respiration, locomotion, tissue repair and hydromineral regulation), less energy can be invested in development and growth. Reduction in growth can in turn lead to starvation, reduced fitness and reduced activity and, ultimately, death (Wendelaar Bonga, 2011). Reduced activity and fitness have further implications on predator avoidance, as well as the discovery of prey items. Stress is also known to affect the susceptibility to disease (Snieszko, 1974; Pickering and Pottinger, 1989), and cortisol has been demonstrated to function as a key mediator modulating the immune response (Tort, 2011). Reduced survival in larval life stages owing to elevated or decreased turbidities can have significant long-term impacts on reproduction of the declining delta smelt population. In particular, low numbers of young fish will lead to a diminished population size and low numbers of adults potentially migrating back to the spawning grounds to secure the continued existence of this species. In addition, this might further reduce the genetic diversity and exacerbate the documented population bottleneck for delta smelt (Fisch et al., 2011).

Compared with the juvenile life stage (Hasenbein et al., 2013), the larval life stage appears to have more limited tolerance ranges for turbidity. Previous studies with juvenile delta smelt (120 dph) showed enhanced feeding at low turbidities and constant feeding up to 120 NTU, with a strong decline in feeding at 250 NTU (Hasenbein et al., 2013). In juvenile fish, stress-related biomarkers were largely unresponsive to turbidities up to 120 NTU but showed very high expression at 250 NTU (Hasenbein et al., 2013). In contrast, for the larval life stage, significant adverse effects in reduced feeding and elevated expression below 12 NTU and above 80 NTU were determined. Taken together, these data indicate that tolerance ranges are life-stage dependent and that ontogenetic changes need to be taken into consideration in tolerance range and niche assessments. Differential tolerances to environmental
parameters, such as temperature and salinity, between life stages of delta smelt have also been observed in other studies (e.g. Komoroske et al., 2014, 2015). Thus, understanding the tolerance ranges and niche dimensions for each life stage of a species is of utmost importance in order to conduct and pursue effective conservation efforts.

All study endpoints (survival, feeding, biochemical and molecular stress indicators) delivered consistent results, providing confidence to our estimate of the larval delta smelt turbidity requirements. However, some could argue that measuring only a couple of endpoints, such as survival and food intake, is easier and more cost effective to assess, whereas recent reviews have highlighted the importance of a mechanistic understanding of physiological responses, in particular with respect to informed management decisions (Geist, 2015; Horodysky et al., 2015). In general, when measuring endpoints across different biological levels, the correspondence is not always certain. Thus it is important to determine multiple, integrative endpoints at different levels of biological organization first, in order to make meaningful interpretations before moving forward with use of simple or fewer metrics. The approach used in the present study expands on the approach used by Newcombe and Macdonald (1991), by also including the molecular level and by more generally considering turbidity as part of the ecological niche of a species (taking both positive and negative effects into account). For instance, measuring molecular biomarkers involved in several different metabolic functions, such as the stress response (hypothalamic–pituitary–interrenal axis), energy metabolism, generalized stress and osmotic stress, reveals underlying physiological mechanisms and helps to evaluate potential adverse outcomes caused by stressors. Together with cortisol, molecular biomarkers provide explanations for behavioural and whole-organism endpoints, such as feeding and survival. In addition, some adverse effects would not be detected when solely measuring whole-organism endpoints. The use of multiple-level endpoints is transferable to numerous fish species and stressor types, illustrating their potential for wide use in the field of conservation physiology. Even though these results clearly show that the early life stage of the delta smelt is affected by changes in turbidity, it should be emphasized that these experiments were relatively short term and tested a single turbidity material, light intensity, alga concentration, and prey item type and life stage. Further tests that integrate these variables to a broader extent are required to explore more comprehensively the effects of turbidity on the delta smelt.

Supplementary material

Supplementary material is available at Conservation Physiology online.

Acknowledgements

The authors thank Dr Joan Lindberg, Luke Ellison and Galen Tigan at the University of California Davis Fish Conservation and Culture Laboratory for supplying delta smelt along with invaluable knowledge on delta smelt rearing, Dennis Cocherell for advice on experimental set-up and gear selection for this study, and Rebecca Hudson Davis, Colin Vandergraaf, Michelle Holtz and Roan Chiong for technical assistance. The authors also thank Dr Bernhard Stoeckle for advice and support with statistical analysis.

Funding

Funding was provided by the US Department of Interior, Bureau of Reclamation (contract R12AP20018 to R.E.C. and N.A.F.), the California Delta Stewardship Council (contract 201015533 to R.E.C. and N.A.F.), the State and Federal Contractors Water Agency (contract no. 15-13 to R.E.C.) and the University of California Agricultural Experiment Station (grant number 2098-H to N.A.F.). Partial student funding was provided to M.H. by the Bavarian Elite Programme Universität Bayern e.V.—Scholarship for graduate and post-graduate students and to L.M.K. by the National Science Foundation Graduate-12 Fellowship Program (under the Division of Graduate Education grant number 0841297 to S. L. Williams and B. Ludaescher) and the California Sea Grant Delta Science Doctoral Fellowship (R/ SF-56). The authors acknowledge the support of the Technische Universität München Graduate School’s Faculty Graduate Center Weihenstephan at Technische Universität München, Germany.

References


