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Crypt Organoid Culture as an in Vitro Model in Drug Metabolism and Cytotoxicity Studies

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ABSTRACT

The gastrointestinal tract is enriched with xenobiotic processing proteins that play important roles in xenobiotic bioactivation, metabolism, and detoxification. The application of genetically modified mouse models has been instrumental in characterizing the function of xenobiotic processing genes (XPG) and their proteins in drug metabolism. Here, we report the utilization of three-dimensional crypt organoid cultures from these animal models to study intestinal drug metabolism and toxicity. With the successful culturing of crypt organoids, we profiled the abundance of Phase I and Phase II XPG expression, drug transporter gene expression, and xenobiotic nuclear receptor (XNR) gene expression. Functions of XNRs were examined by treating crypt cells with XNR prototypical agonists. Real-time quantitative polymerase chain reaction demonstrated that the representative downstream target genes were induced.

These findings were validated from cultures developed from XNR-null mice. In crypt cultures isolated from *Pxr*^{-/-} mice, pregnenolone 16 α -carbonitrile failed to induce *Cyp3a11* gene expression; similarly, WY14643 failed to induce *Cyp4a10* in the *Ppara*^{-/-} crypts. Crypt cultures from control (*Ugt1*^{F/F}) and intestinal epithelial cell (IEC) specific *Ugt1* null mice (*Ugt1* ^{Δ IEC}) were treated with camptothecin-11, an anticancer prodrug with severe intestinal toxicity that originates from insufficient UGT1A1-dependent glucuronidation of its active metabolite SN-38. In the absence of *Ugt1* gene expression, *Ugt1* ^{Δ IEC} crypt cultures exhibit very limited production of SN-38 glucuronide, concordant with increased apoptosis in comparison with *Ugt1*^{F/F} crypt cultures. This study suggests crypt organoid cultures as an effective in vitro model for studying intestinal drug metabolism and toxicity.

Introduction

The gastrointestinal (GI) tract is lined with epithelial cells that express an abundance of xenobiotic or drug metabolizing enzymes (DMEs) in addition to efflux and influx transporters. Thus, the absorption of xenobiotics and drugs are subjected to metabolism by DMEs localized in the mature enterocyte. This process contributes significantly to bioactivation, metabolism, and detoxification of drugs, all of which are associated with the outcome of drug efficacy and safety. A greater appreciation attributed to the GI tract and drug metabolism has been achieved by examining drug metabolism and toxicity in knockout animal models or fully humanized animal models. For example, human CYP3A4 is considered by many to be one of the most versatile of the CYPs involved in drug metabolism because it accounts for the metabolism of almost half of all prescribed medications (Zanger et al., 2008). Van Herwaarden AE et al. (van Herwaarden et al., 2007) and Van

Waterschoot RA et al. (van Waterschoot et al., 2008, 2009) generated intestine- and liver-specific expression of human CYP3A4 in a *Cyp3a*-null background, demonstrating intestinal CYP3A4 activity had a profound impact on systemic exposure of the anticancer drug docetaxel following its oral administration. Camptothecin (CPT)-11 (Irinotecan), is a prodrug that is hydrolyzed by carboxylesterase (CES) in different tissues to form the active topoisomerase inhibitory metabolite SN-38 (Kaneda et al., 1990; Sugimoto et al., 1990), which is further metabolized by UGT1A1-dependent glucuronidation to form an SN-38 glucuronide (SN-38G) (Iyer et al., 1998). Classically, the bioactivation and detoxification of CPT-11 have been credited primarily to hepatic metabolism (Alimonti et al., 2004; Michael et al., 2006). However, recent studies have confirmed that intestinal CES2 has a greater affinity toward CPT-11 in comparison with liver CES1 (Hatfield et al., 2011). With the development of *Ugt1*^{F/F} mice, deletion of the *Ugt1* locus in intestinal epithelial cells (*Ugt1* ^{Δ IEC}) produced severe CPT-11-induced intestinal toxicity in comparison with *Ugt1*^{F/F} mice or mice with a liver-specific *Ugt1* deletion (*Ugt1* ^{Δ HEP}) (Chen et al., 2013). This result highlighted that UGT1A1 expression in intestinal tissue is critical in preventing SN-38-induced toxicity.

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ABBREVIATIONS: AhR, aryl hydrocarbon receptor; BNF, β -naphthoflavone; CAR, constitutive androstane receptor; CES, carboxylesterase; CYP, cytochrome P450; DHA, docosahexaenoic acid; DMEs, drug metabolism enzymes; GI, gastrointestinal; IEC, intestinal epithelial cell; ISC, intestinal stem cell; KO, knockout; PCN, pregnenolone 16 α -carbonitrile; PPAR, peroxisome proliferator-activated receptor; PXR, pregnane X receptor; RT-qPCR, reverse transcription quantitative polymerase chain reaction; SN-38G, SN-38 glucuronide; TCDD, 2,3,7,8-tetrachlorodibenzo-*p*-dioxin; TCPOBOP, 4-Bis-[2-(3,5-dichloropyridyloxy)]benzene; UGT, UDP-glucuronosyltransferase; XNR, xenobiotic nuclear receptors; XPG, xenobiotic processing gene.

Given the importance of the GI tract and the beneficial application of genetically modified mouse models to examine drug metabolism and toxicity, the development of primary cells cultured from these mice to examine gene expression and toxicity will provide an important tool to complement *in vivo* animal experiments. Sato et al. (2009) established long-term culture conditions in which one single *Lgr5+* stem cell embedded in matrix gel can independently generate villus-like epithelial domains with the presence of all differentiated IECs. The three-dimensional culture is a continuously expanding, self-organizing structure that is reminiscent of normal gut. This result has been popularly applied in the study of crypt-villus biology, regenerative medicine, and gene therapy (Sato and Clevers, 2013; Grabinger et al., 2014; Ranga et al., 2014), but its applications to investigate intestinal epithelial cell-specific xenobiotic metabolism has been lacking. In this article, we report the application of crypt organoid cultures from genetically modified mice as a model to examine gene expression and drug metabolism.

Materials and Methods

Reagents. WY14643 [4-chloro-6-(2,3-xylylidino)-2-pyrimidinylthioacetic acid], pregnenolone 16 α -carbonitrile (PCN), 1,4-Bis-[2-(3,5-dichloropyridyloxy)]benzene (TCPOBOP), phenobarbital, docosahexaenoic acid (DHA), and β -naphthoflavone (BNF), *N*-acetylcysteine, and CPT-11 standard were from Sigma-Aldrich (St. Louis, MO). SN-38 and T0901317 were purchased from Cayman Chemical Company (Ann Arbor, MI). SN-38 glucuronide was from Santa Cruz Biotechnology (Santa Cruz, CA). Internal standards deuterated d10-CPT-11 and d3-SN-38 were purchased from Toronto Research Chemicals (Toronto, Ontario, Canada). 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin (TCDD) was obtained from the National Institutes of Health National Cancer Institute, Chemical Carcinogen Reference Standard Repository (Bethesda, MD). Matrigel basement membrane matrix was purchased from BD Biosciences (Bedford, MA). Advanced DMEM/F12 medium and the supplements, including GlutaMax, Hepes, penicillin streptomycin, mouse EGF, N2, and B27 supplements, were purchased from Thermo Fisher Scientific (Waltham, MA); mouse Noggin was from Peprotech (Rocky Hill, NJ). HEK-293 cells producing R-Spondin 1 protein (293T-HA-Raspol_Fc cell line) was a generous gift from the Dr. Calvin Kuo laboratory at Stanford University (Ootani et al., 2009). The cells were cultured and passaged per standard protocol. For collecting R-Spondin 1 conditioned media, 293T-HA-Raspol_Fc cells at approximately 75% confluency were changed to advanced DMEM/F12 medium and cultured for another week. Medium was then collected by passing through 0.22 μ m sterile filter.

Animals. C57BL/6J mice were purchased from Charles River Laboratories (Raleigh, NC). Intestinal-specific *Ugt1* deletion mice (*Ugt1^{ΔIEC}*) were generated in our laboratory (Chen et al., 2013). *Ppara*^{-/-} mice were purchased from Jackson Laboratories (Bar Harbor, ME) (Lee et al., 1995). *Pxr*^{-/-} and *Car*^{-/-} mouse models were a kind gift from Drs. Ron Evans (Laboratory at Salk Institute, La Jolla, CA) (Xie et al., 2000) and Masahiko Negishi (Ueda et al., 2002; Yamazaki et al., 2005) (National Institute of Environmental Health Sciences), respectively. The proposed mouse experiments have been approved per the procedures as set by the University of California San Diego Institutional Animal Care and Use Committee. All the proposed mouse strains were housed in a pathogen-free facility with automated temperature control and a 12/12-hour light/dark cycle.

Isolation of Crypt Cells and Organoid Culture. Intestinal crypt cell isolation and organoid culturing were carried out according to previous publications (Sato et al., 2009; Sato and Clevers, 2013) with several modifications. Briefly, mouse small intestine was dissected, cut longitudinally, and washed briefly by ice-cold Dulbecco's phosphate-buffered saline (DPBS). The tissue was further dissected into small pieces and incubated in DPBS containing 2 mM EDTA at 4°C with gentle shaking. One hour later, the EDTA solution was removed, and 10% fetal bovine serum in DPBS buffer added followed by vigorously shaking to release villi and crypt cells. The cell solution was passed through tea filter to remove tissue debris, and then further filtered through a 70 μ m cell strainer. The filtrate was centrifuged at 1000 *g* for 10 minutes at 4°C, and the cell pellet washed twice by DPBS. Crypt cells were counted. Approximately

1000 crypts were suspended into 50 μ l ice-cold Matrigel and plated into prewarmed 24-well tissue culture plates. Ten minutes after the incubation, 0.5 ml of complete growth medium (advanced DMEM with GlutaMax, Hepes, penicillin/streptomycin, N2 and B27 supplements, with 2.5 mM *N*-acetylcysteine, 0.1 μ g/ml mouse Noggin, 0.05 μ g/ml mouse EGF, and 10% of R-Spondin 1-conditioned medium) was added. The growth of crypt organoids was monitored. Mouse EGF was added every other day. Fresh medium was added every 4 days. Cells were normally ready for passage every 7 days. Crypt cells were exposed to various chemicals on day 4 and, after 24 hours, the cells were collected in triplicate for further analysis. The results were described as the average \pm S.E.M., which represent multiple independent experiments with compatible data. Student's *t* test was used for statistical analysis.

RNA Preparation and Reverse Transcription Quantitative Polymerase Chain Reaction Analysis. Culture medium was removed, and the plates were kept on ice. Ice-cold DPBS was added to break up the Matrigel and the organoids released by pipetting several times through P1000 tips. Crypt cells were collected by centrifugation. Cell pellets were then incubated with 1 ml of TRIzol reagent and RNA isolated as outlined by TRIzol kit. RNA concentrations were determined with a Nanodrop Lite spectrophotometer (Thermo Scientific). Reverse transcription was performed by using 1 μ g of RNA and following instructions from the iScript cDNA Synthesis Kit (BioRad, Hercules, CA). Real-time quantitative polymerase chain reaction (RT-qPCR) experiments were carried out on a CFX96 quantitative polymerase chain reaction (qPCR) system (BioRad) by using Ssoadvanced SYBR Green reagent (BioRad). Primers were designed through mouse primer depot (<https://mouseprimerdepot.nci.nih.gov/>). Transcription levels were quantified with the Ct value normalized to mouse cyclophilin (*Cph*).

Protein Preparation and Western Blots. Cells were released from Matrigel. Cell lysates were prepared in a RIPA buffer. After protein concentration determination, protein samples were subjected to gel electrophoresis by using Nupage 4–12% (wt/vol) Bis-Tris gradient gel (ThermoFisher). Western blots were developed and imaged using a ChemiDoc Touch Imaging System (BioRad). The following antibodies were used: goat anti UGT1A (Santa Cruz, sc-27418), rabbit anti-cleaved caspase 3 (Cell Signaling, #9661; Danvers, MA), and anti- α -tubulin (Sigma, T9026).

Liquid Chromatography/Mass Spectrometry Studies of CPT-11, SN-38, and SN-38G. Sample preparation was performed as previously described (Marangon et al., 2015). Media or cell lysates were centrifuged, and the supernatants were added with 0.1% acetic acid (v/v 1:3) containing the internal standards deuterated d10-CPT-11 and d3-SN-38 at 15 ng/ml. Standards were prepared by spiking 10 μ l of working solutions in 90 μ l of blank medium, which sustains subsequent treatments as samples. Samples were centrifuged and 1 μ l of the supernatants was injected for analysis by a Prominence HPLC (Shimadzu, Tokyo, Japan) coupled with an API4000 mass spectrometer (Sciex, Framingham, MA). Separation was achieved using a Gemini C18 column (100 \times 4.6 mm, 3 μ M; Phenomenex, Torrance, CA), and mass spectrometry analysis was achieved using positive mode with ion spray voltage set at 5000 V and a source temperature of 600°C. The following transitions were used for quantification in multiple reaction monitoring (MRM) mode: CPT-11: *m/z* 587.4 \rightarrow 124.2; SN-38: *m/z* 393.3 \rightarrow 349.3; SN-38G: *m/z* 569.3 \rightarrow 392.3; and the internal standards d10-CPT-11: *m/z* 597.4 \rightarrow 133.3; and d3-SN-38: *m/z* 396.2 \rightarrow 352.2.

Results

Development of Crypt Organoids and the Expression of Drug Metabolism Enzymes. Each crypt culture contained multiple cells and normally formed spherical structures on the first day after the incubation (Fig. 1A), followed by budding. Small buds usually could be visualized in 2–4 days after the incubation. The crypt region underwent continuous budding events; by day 5 after incubation, the lumen of organoids became distorted and dark, which were filled with matured enterocytes or apoptotic cells (Fig. 1A). Well-developed organoids were collected between days 7 and 10 after cells were passaged, and RT-qPCR was conducted to investigate gene expression patterns. As demonstrated in Fig. 1B, Phase I and Phase II XPGs along with transporter genes were identified in mature crypt organoids. Cytochrome P450 (*Cyp*) gene

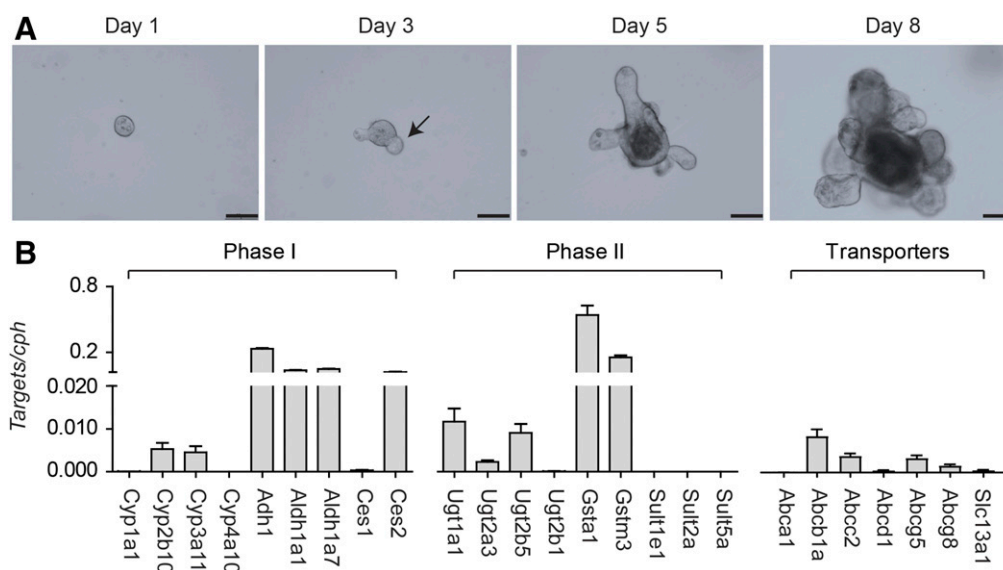


Fig. 1. The growth of small intestinal crypt organoids and the expressions of XPGs in mature crypt organoids. (A) Shown are mouse small intestinal crypt organoid cultures at various times after primary culture. Images were taken with a Nikon Eclipse Ts2 inverted microscope, with the scale bar shown at 90 μm . (B) Mature crypt organoids (in triplicate) were collected 8 days after plating and RNA prepared for RT-qPCR analysis. Gene expression of Phase I and Phase II drug metabolism genes and transporters are expressed as *Target genes/mouse Cph* and are described as average \pm S.E.M. ($n = 3$). Data represent multiple independent experiments with similar results.

expression for *Cyp2b10* and *Cyp3a11* along with other XPGs including alcohol dehydrogenase *Adh1*, aldehyde dehydrogenase *Aldh1a1*, and *Aldh1a7* were actively expressed, all of which have been known to exhibit high expression levels in intestine (Vaglenova et al., 2003; Huang et al., 2009; Renaud et al., 2011). In comparison with carboxylesterase (*Ces*) 1, *Ces2* is known as the intestinal specific isoform (Hatfield et al., 2011), which is consistent with our results, although RT-qPCR results cannot provide absolute gene expression levels. Phase II XPGs were highlighted by gene expression of *Gst* in addition to the *Ugts* encoded by the *Ugt1* and *Ugt2* gene families, both of which are intestinal enriched genes (Gibbs et al., 1998; Strassburg et al., 1999; Shin et al., 2009). Efflux drug transporters were also identified in crypt organoids, such as *P-glycoprotein* (*Pgp*, *Abcb1a*) and *Mrp2* (*Abcc2*), both of which are important transporters of many drugs and xenobiotics in enterocytes (Sparreboom et al., 1997; Mottino et al., 2000), along with intestinal specific transporters *Abcg5/8* (Berge et al., 2000), which are important sterol efflux pumps.

Functional Nuclear Receptors Observed in Organoids that Respond to Xenobiotic Treatment. Fundamental to understanding the regulatory events associated with DMEs has been a clear vision characterizing the role of the xenobiotic nuclear receptors (XNRs) in control and expression of the xenobiotic processing genes (XPGs) (Rushmore and Kong, 2002; Staudinger et al., 2010, 2013; Chai et al., 2013; Hoffmann and Partridge, 2015). Xenobiotic and environmental toxicant exposure has the potential to modify XPG expression through interactions with XNRs. Altered gene expression through activation of XNRs affects drug metabolism as well as drug-drug interactions, leading to changes in drug efficacy and safety. The gene expression levels of XNRs in crypt organoids were determined, as shown in Fig. 2A. We examined expression of the genes encoding the aryl hydrocarbon receptor (*Ahr*), pregnane X receptor (*Pxr*), constitutive androstane receptor (*Car*), peroxisome proliferator-activated receptors (*Ppar*), retinoid X receptor (*Rxr*), and the liver X receptor (*Lxr*), and RNA encoded by each gene was detectable. These gene expression patterns are relative to *Cph* gene expression and do not reflect comparative expression values of RNA between the genes, because the primers for each gene may have differences in their annealing properties. To

determine their functionality, crypt organoids were then treated with specific XNR ligands and downstream target gene expression evaluated. TCDD treatment, the prototypical agonist of the aryl hydrocarbon receptor (AhR) (Quattrochi and Tukey, 1993), induced *Cyp1a1* gene expression (Fig. 2B); BNF, an AhR agonist (Sinal et al., 1999), also induced *Cyp1a1* (Fig. 2B). Similarly, when crypt organoids were treated with TCPOBOP, a mouse-specific CAR agonist (Tzamelis et al., 2000), *Cyp2b10* was induced (Fig. 2C). *Cyp3a11* gene expression was induced by PCN (Fig. 2D), a well-studied PXR ligand (Guo et al., 2002). We observed that *Cyp3a11* was also induced by TCPOBOP and the LXR agonist T0901317 (N-(2,2,2-Trifluoroethyl)-N-[4-[2,2,2-trifluoro-1-hydroxy-1-(trifluoromethyl)ethyl]phenyl]benzenesulfonamide), implying that *Cyp3a11* gene expression can also be regulated by activated CAR and LXR. *Cyp4a10* was largely induced by the PPAR α agonist WY14643 (Fig. 2E). Drug transporter genes were also inducible, as shown in Fig. 2F, with induction of *Abca1* gene expression by the LXR agonist T0901317.

Crypt Organoids Isolated from XNR-knockout Mice for XPG Expression Studies. Knockout (KO) mouse models have been used to understand the physiological and pharmacological role of a specific gene. Knockout mouse models have also been exploited in drug metabolism studies, particularly in elucidating XNR-mediated gene regulation of XPGs. Here we have used both global and intestinal-specific KO mouse models to investigate the use of organoid cultures in drug metabolism studies. Crypt cells were isolated from both wild-type and *Ppara*^{-/-} mice, and after 5 days the organoids were treated with breast milk (1:100 dilution), the fatty acid DHA at 5 μM , and the PPAR α agonist WY14643 at 100 μM . Twenty-four hours after treatment, cells were collected for RT-qPCR analysis. In wild-type cells, breast milk, DHA, and WY14643 activated gene expression of the PPAR α -targeted gene *Cyp4a10*; however, these inductions were completely diminished in crypt organoids isolated from *Ppara*^{-/-} mice (Fig. 3A). Similarly, PCN also lost its capability in mediating the *Cyp3a11* induction in *Pxr*^{-/-} organoids (Fig. 3B). These results confirm that crypt organoids maintain the genetic characteristics that are exhibited in KO mice following activation of XNRs.

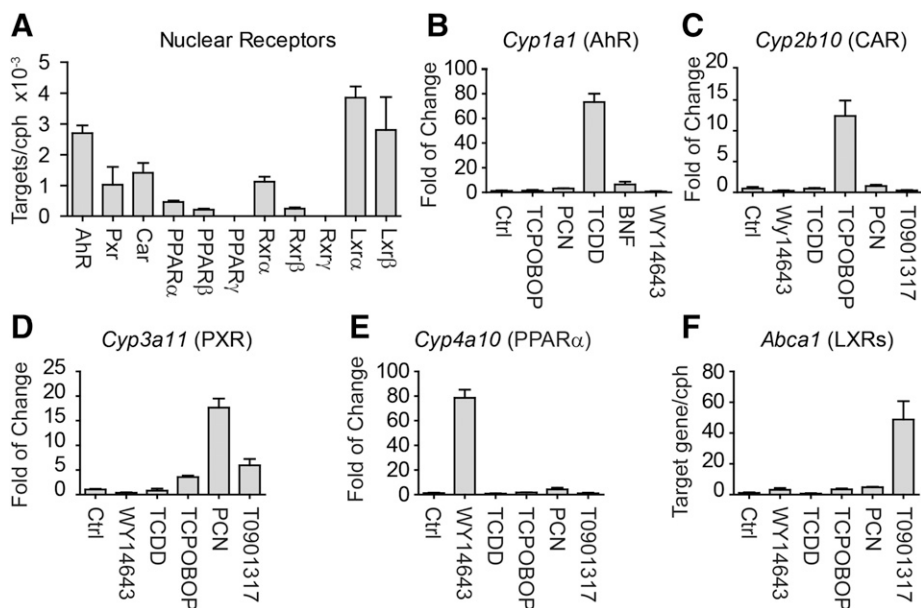


Fig. 2. Gene expression analysis of NRs and their function in mature crypt cells. (A) Shown are nuclear receptor expression levels from mature crypt cells conducted by RT-qPCR analysis, average \pm S.E.M. ($n = 3$). (B–F) Crypts in exponential growth period were treated with different nuclear receptor agonists, including TCDD (50 nM), β -Naphthoflavone (BNF) (20 μ M), TCPOBOP (10 μ M), PCN (30 μ M), T0901317 (100 μ M), and WY14643 (100 μ M). Twenty-four hours after agonist exposure, crypt cells were collected for RT-qPCR analysis (average \pm S.E.M., experiments carried out in triplicate).

Intestinal Crypt Cultures Metabolize CPT-11. Stable crypt organoids express CES1 and 2, UGT1A1, and the transporters ABCC2 and ABCB1A based upon gene expression analysis. CPT-11 enters intestinal epithelial cells and is cleaved by CES to the active topoisomerase inhibitor SN-38, a product that is toxic and damaging to tissues if not metabolized further by glucuronidation. To determine if intestinal crypt cultures could metabolize CPT-11 and culminate with the formation of the SN-38 glucuronide (SN-38G), cells were first treated with a range of CPT-11 concentrations (Fig. 4A). After exposure for 24 hours, culture media (extracellular) and cells (intracellular) were harvested for quantitation of SN-38 and SN-38G by HPLC-MS analysis. In culture media, as the concentration of CPT-11 was increased there was a concordant increase in SN-38 and SN-38G (Fig. 4A). The concentrations of SN-38 fit a linear model with R-squared at 0.99, but the SN-38G levels showed poor linearity with R-squared at 0.84 indicating enzymatic saturation (Fig. 4A). As the concentration of CPT-11 is increased, there is the rapid appearance of intracellular SN-38 at 10 μ M CPT-11 that is nearly triple the concentration of intracellular SN-38G (Fig. 4B), supporting the extracellular values that implicate UGT1A1 saturation. When the ratios of SN-38G/SN-38 were determined in both compartments, the ratios were much higher in the

extracellular compartment, indicating that both SN-38 and SN-38G are removed from the cell by efflux transporters (Fig. 4C). The highest concentration of CPT-11 used resulted in a sharp decline in both the extracellular and intracellular SN-38G/SN-38 ratios, indicating that enzymatic cleavage of CPT-11 to SN-38 and the resulting glucuronidation of SN-38 to SN-38G were declining. Because SN-38 is toxic, this result corresponded to an increase in cell death, as demonstrated by induction of cleaved caspase 3, a marker of cell apoptosis (Fig. 4D).

In the absence of glucuronidation, the intestinal epithelium is sensitive to the toxic actions of SN-38 (Chen et al., 2013). When we targeted the deletion the *Ugt1* locus and the *Ugt1a1* gene in *Ugt1^{ΔIEC}* mice and challenged *Ugt1^{ΔIEC}* mice with CPT-11, they were highly susceptible to diarrhea in comparison with wild-type *Ugt1^{F/F}* mice (Chen et al., 2013). Apoptosis was not observed in *Ugt1^{F/F}* crypt cells when treated with CPT-11 at 1 μ M, but apoptosis was apparent in crypt organoids from *Ugt1^{ΔIEC}* mice at this concentration (Fig. 4E), in which cells showed no expression of UGT1 protein (Fig. 4E) and failed to form the metabolite SN-38G (Fig. 4F). Therefore, crypt organoids from *Ugt1^{ΔIEC}* mice were at least 10-fold more sensitive to the toxic actions of CPT-11 than *Ugt1^{F/F}* mice. Crypt organoid cultures can be employed to examine the metabolic outcome of drug and xenobiotic metabolism, the outcome of metabolism

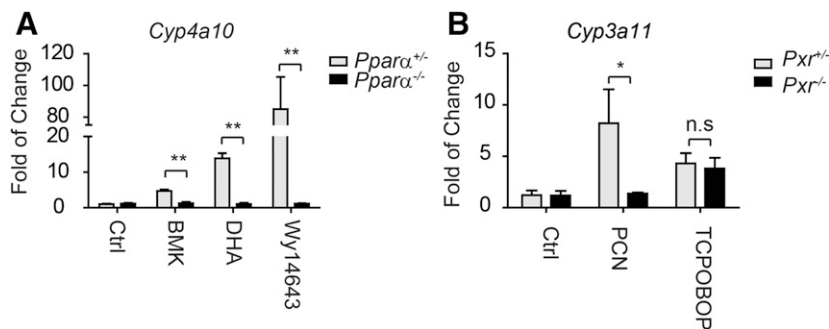


Fig. 3. RT-qPCR results of mature crypt cells from both wildtype and genetically modified mice. (A) Crypt cells were isolated from *Ppara*^{+/+} (wild-type) and *Ppara*^{-/-} (*Ppara* deletion) mice. After several passages, crypt cells were treated with vehicle control (Ctrl, DMSO at 0.1%), breast milk (BMK) at 1:100 (v/v) dilution into the culture medium, DHA (5 μ M), and WY14643 (100 μ M) for 24 hours. Cells were collected for RT-qPCR analysis to measure *Cyp4a10* gene expression. Results were normalized with vehicle-treated wild-type crypts and described as fold change (average \pm S.E.M., $n = 3$). (B) Crypt cells were isolated from wild-type *Pxr*^{+/+} and *Pxr*^{-/-} knockout mice. Cells were treated with PCN at 30 μ M and TCPOBOP at 10 μ M for 24 hours followed by *Cyp3a11* gene expression analyzed by RT-qPCR ($n = 3$). n.s., Nonsignificant, * $P < 0.05$, ** $P < 0.01$, Student's *t* test.

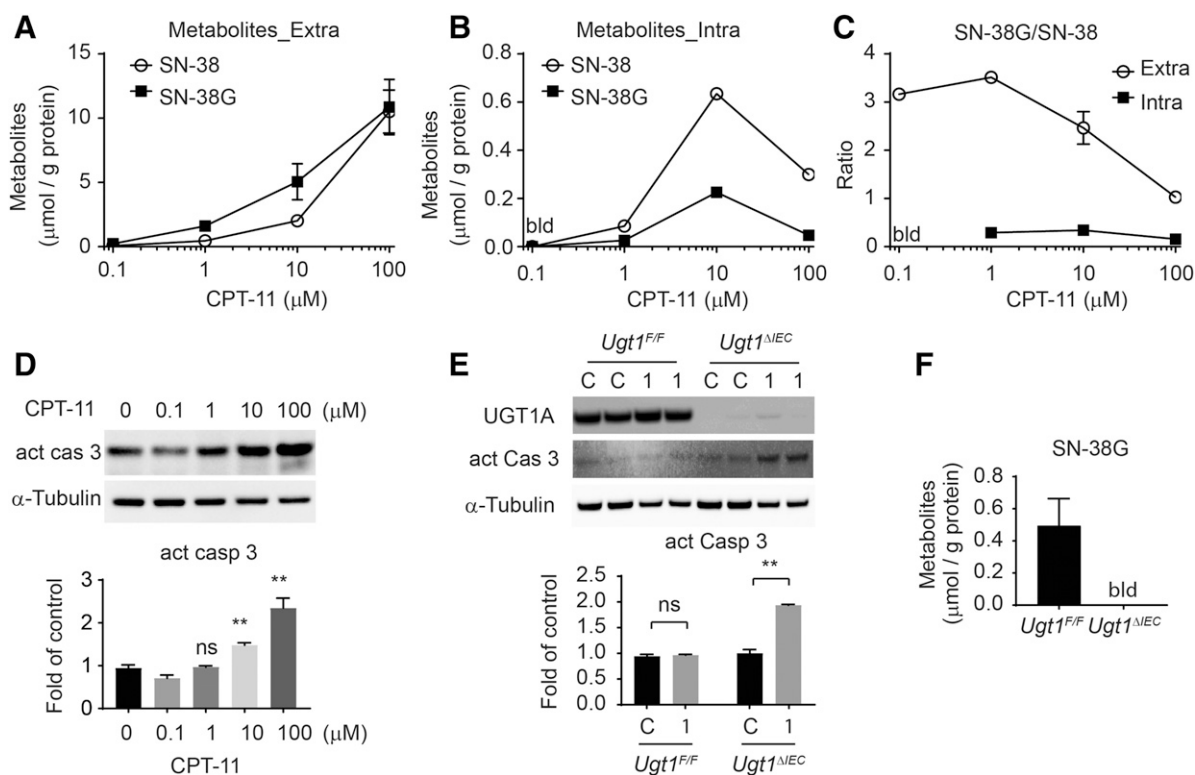


Fig. 4. Irinotecan metabolism and toxicity studies using crypt organoids from both *Ugt1^{F/F}* and *Ugt1^{ΔIEC}* mice. (A–D) Crypts were isolated from *Ugt1^{F/F}* mice. Cells were treated with CPT-11 at 0, 0.1, 1, 10, and 100 μM (in triplicate). After 24 hours, both media and cells were collected for HPLC-MS analysis. SN-38 and SN-38G were determined in both extracellular compartment (A) and intracellular compartment (B) (cells from three wells were combined and then subjected to the bioanalysis), and the ratio of SN-38G/SN-38 (C). (D) Apoptosis was analyzed by Western blot of cleaved caspase 3, which was further quantified by using the ChemiDoc Touch Imaging System and Image Laboratory software (BioRad), and the data were calculated based on independent Western blots (average ± S.E.M., *n* = 3). (E and F) Crypts were isolated from both *Ugt1^{F/F}* and *Ugt1^{ΔIEC}* mice. After 3 days, cells were treated with control or CPT-11 at 1 μM. (E) Twenty-four hours later, whole cell extracts were prepared for Western blot analysis using an anti-UGT1A, antiactive caspase 3, and anti-α-tubulin antibodies. Western blots of cleaved caspase 3 were further quantified. (F) Cell culture media were collected for bioanalysis of SN-38G (average ± S.E.M., *n* = 3). bld, Below limit of detection.

following XNR activation, and the disposition of metabolism under conditions of altered pharmacogenetics.

Discussion

Recent advances in establishing long-term culture conditions of crypt-villi that propagate into functional organoids with intact luminal surfaces have made it possible to investigate intestinal epithelial cell functionality in tissue culture (Sato et al., 2009; Sato and Clevers, 2013). Because intestinal epithelial cells turn over in adults, for example, every 5 days in mice, new epithelial cells are formed from long-lived intestinal stem cells (ISCs) that sit at the base of the crypts (Barker et al., 2008). These stem cells produce the rapidly proliferating progenitor cells called transit-amplifying cells that migrate up the crypt-villus axis and eventually differentiate into mature epithelial cells. With discovery of the signaling pathways that are necessary to maintain stemness leading to epithelial cell proliferation, environmental cues have been employed to promote the crypt ISCs to form mini-gut cultures that are composed of budding epithelial cells (Sato et al., 2009,2011; Sato and Clevers, 2013). These three-dimensional mini-gut organoid cultures resemble in vivo intestinal epithelial structures, with a single ISC producing the different types of differentiated cells needed to support the development of the mature enterocytes (Sato et al., 2009,2011). It has also been demonstrated that the organoids maintain absorptive and digestive functions, making the tissue an excellent model to examine drug uptake and metabolism and the impact of xenobiotic exposure on epithelial cell function. Taking advantage of this unique physiology, we explored the

use of three-dimensional mini-gut organoids to examine XPG expression, inducibility, and metabolism.

Within 8 days of culturing crypt stem cells, the expanding organoids show well-structured epithelial cell projections. Constitutive expression of XPGs was abundant, with those encoding the classically identified Phase I, Phase II, and Phase III (transporters) genes well represented. The culture medium contains high concentrations of *N*-acetylcysteine, which is a substrate in the synthesis of glutathione (Weinander et al., 1994). Elevated glutathione concentrations provide a rich reductive capacity for the cell, indicating that organoid cultures may be an excellent model to examine receptor-mediated induction of XPGs by drugs and xenobiotics without producing excessive oxidative stress. The major XNRs that are targeted for activation by drugs and xenobiotics are also expressed and include the AhR, PXR, CAR, PPARα, and LXR, which are key modulators of gene expression that often control the balance between exposures, resulting in a toxic or efficacious outcome. Expression of the XNRs is functional in the proliferating organoids, as we demonstrated by receptor-mediated induction of key genes linked to drug transport and metabolism. Similar results were recently demonstrated (Bijmans et al., 2017). There was high specificity for activation of the AhR by TCDD and induction of *Cyp1a1* gene expression (Strom et al., 1992). Although BNF is an agonist of the AhR, it also induces *Cyp1a1*, but the response is not as potent as TCDD (Sinal et al., 1999). Interestingly, we did not notice any unusual growth inhibition properties of our proliferating organoids following TCDD treatment as was noted in a recent publication examining growth properties following activation of the AhR (Park et al., 2016). These differences may result from the

concentrations of TCDD used; although we found excellent induction using 50 nM, Park et al. (2016) treated their organoid preparations with 0.1 to 1.0 μ M. The mouse *Cyp2b10* gene is selectively induced by mouse-specific CAR ligands (Honkakoski et al., 1998). Activation of PXR by PCN generates robust induction of *Cyp3a11* gene expression. Activation of CAR has been shown previously to induce *Cyp3a11* (Hernandez et al., 2009), corroborating our results with crypt organoids demonstrating induction of *Cyp3a11* gene expression by the CAR activator TCPOBOP. This is further supported when crypt organoids deficient in PXR exhibit elevated *Cyp3a11* gene expression when treated with TCPOBOP. In addition, activation of LXR by T0901317, which targets activation of *Abcal* gene expression (Wagner et al., 2003), can also support induction of *Cyp3a11* gene expression. These findings indicate that *Cyp3a11* can be regulated following activation of PXR, CAR, and LXR. We showed previously that oral administration to mice with the PPAR α agonist WY14643 induces intestinal CYP4A protein expression (Senekeo-Effenberger et al., 2007). Significant activation of *Cyp4a10* gene expression in organoid cultures is also documented following exposure with WY14643, a response that is eliminated in cultures derived from PPAR α -null mice.

The functionality of the crypt organoid cultures can be exploited for metabolism and toxicology studies by implementing cultures derived from XNR and XPG knockout mice. We demonstrated this capability using *Ugt1^{ΔIEC}* mice, where we have targeted the specific deletion of the *Ugt1* locus in intestinal epithelial cells. These mice are deficient in the UGT1A proteins, including UGT1A1. Characteristic of crypt organoids and intestinal epithelial cells, they express CES2, necessary to cleave the prodrug CPT-11 to SN-38, the active topoisomerase 1 inhibitor. The high expression levels of CES2 are an excellent example that documents the maintenance of tissue-specific expression markers that can be exploited to examine drug metabolism. It supports previous reports from Clevers' laboratory (Sato et al., 2009; Sato and Clevers, 2013) and more recently the van Mills group (Bijmans et al., 2017) that gene expression profiles in intestinal organoids resemble patterns observed in normal tissue. We showed previously in *Ugt1^{ΔIEC}* mice that treatment with CPT-11 led to severe intestinal damage, resulting from the efficient generation of SN-38 coupled with the inability to generate UGT1A1-dependent SN-38G (Chen et al., 2013). Organoids generated from *Ugt1^{FF}* mice efficiently generated SN-38 and SN-38G, with cell apoptosis being evident at 10 to 100 μ M CPT-11. When organoids from *Ugt1^{ΔIEC}* mice were treated with the same concentrations of CPT-11, remarkable cell apoptosis was evident at 1 μ M CPT-11. These findings provide evidence that examining drug metabolism in crypt organoids can be efficiently linked to specific XPGs while confirming a mechanistic link between gene expression and toxicity.

In conclusion, crypt organoids maintain functional, tissue-specific gene expression encoding DMEs and can be further regulated following activation of XNRs. Crypt organoids developed from stem cells isolated from genetically modified mice maintain their engineered phenotypical features, as determined at both the gene expression and functional levels. It is argued that the use of organoid cultures may eventually reduce the need for animal models in scientific research, because the cultures maintain tissue-specific gene expression that can be regulated following activation of XNRs (Clevers, 2013; Bijmans et al., 2017). Although the organoid cultures are a superior model when compared with primary cells or long-term differentiated cells, there will always be the need for drug testing in vivo. An example of this need is to define the mechanisms leading to regulation of intestinal *UGT1A1* gene expression during development. Neonatal jaundice and severe forms of neonatal hyperbilirubinemia develop because of delayed expression of the *UGT1A1* gene during development, and its expression in the GI tract is critical in preventing bilirubin toxicity. Agents that activate many of the XNRs when

given orally to neonatal *hUGT1* mice induce intestinal UGT1A1 and reduce serum bilirubin. These same agents can induce the *UGT1A1* gene in tissue culture. However, we discovered that intestinal *UGT1A1* gene expression is very responsive to agents that induce mild oxidative stress (Liu et al., 2016). This new mechanism, which can be replicated only in vivo, may lead to the identification of new agents or therapeutics that may be considered as new therapy in the future to prevent the substantial mortality and permanent morbidities resulting from severe neonatal hyperbilirubinemia, which is estimated to impact over 1 million newborns every year (Wong et al., 2011; Bhutani et al., 2013). Thus, crypt organoids could become a valuable tool in drug metabolism and toxicity studies to complement those efforts being conducted with humanized and genetically modified mice.

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Authorship Contributions

Participate in research design: Chen.
Conduct experiments: Lu, Rettenmeier, Paszek, Trottier, Barbier, and Chen.
Contributed new reagents or analytic tools: Tukey, Trottier, Barbier, and Chen.
Performed data analysis: Lu, Rettenmeier, Trottier, Barbier, and Chen.
Wrote and contributed to the writing of the manuscript: Lu, Rettenmeier, Paszek, Yueh, Tukey, Trottier, Barbier, and Chen.

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