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### **International Organization of Citrus Virologists Conference Proceedings (1957-2010)**

#### **Title**

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#### **Permalink**

<https://escholarship.org/uc/item/7gq3t4dm>

#### **Journal**

International Organization of Citrus Virologists Conference Proceedings (1957-2010), 18(18)

#### **ISSN**

2313-5123

#### **Authors**

Liu, K. H.  
Song, Z.  
Zhou, Y.  
et al.

#### **Publication Date**

2011

Peer reviewed

## Detection of Citrus tatter leaf virus by Real-Time RT-PCR

K. H. Liu<sup>1,2</sup>, Z. Song<sup>1,2</sup>, Y. Zhou<sup>1,2</sup>, Z. A. Li<sup>1,2</sup>, and C. Y. Zhou<sup>1,2</sup>

<sup>1</sup>Citrus Research Institute of Chinese Academy of Agricultural Sciences

<sup>2</sup>National Citrus Engineering Research Centre, Beibei, Chongqing 400712, P. R. China

**ABSTRACT.** Citrus tatter leaf disease caused by Citrus tatter leaf virus (CTLV) is a major disease leading to serious economic losses in the People's Republic of China due to the common usage of trifoliolate rootstock. A real-time RT-PCR assay using SYBR Green I was developed to detect and quantify CTLV RNA-targets in our laboratory. A pair of primers, ASG-Pf and ASG-Pr, were designed within the conserved sequence of coat protein gene. The assay had specificity of detecting CTLV in different citrus hosts, and its sensitivity was 100 times higher than that of the conventional RT-PCR. It is rapid and reliable, and SYBR Green I is inexpensive, and thus this is a valuable tool to detect CTLV.

Citrus tatter leaf disease caused by Citrus tatter leaf virus (CTLV) is a major systemic disease of citrus in the world, which may lead to severe economic losses, and it was first discovered in latently infected Meyer lemon trees in the United States in 1962, (8). Most citrus cultivars are latent hosts for CTLV, while CTLV-infected cultivars grafted on trifoliolate orange or its hybrids are usually stunted and chlorotic, and they also show a pronounced virus-induced bud union incompatibility (6, 7). It is known that CTLV is widespread in China, and it has caused tremendous economic loss in Zhejiang, Hunan, Fujian and Guangxi Provinces.

Although biological indexing, serology and reverse transcriptase polymerase chain reaction (RT-PCR) can all be used to detect CTLV (1, 2, 4, 5, 9), these methods either have limited requiring specialized facilities such as a greenhouse, or are less sensitive or costly. For rapid and reliable detection of CTLV, we established a system based on SYBR Green I dye.

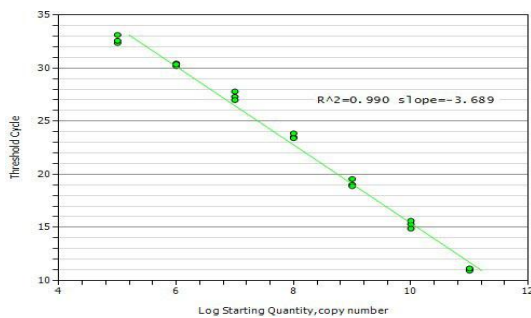
Total nucleic acid extraction was carried out following the method described by Zhou et al. (10). Primers ASG-Pf (5' GAG TTT GGA AGA CGT GCT TCA 3')

and ASG-Pr (5' TTG CAG AGA AGA AGG TAA AGC TC 3') were used to amplify a 165 bp region (3). For reverse transcription, 2 µL of nucleic acid extracts and 3 µL of water denatured at 95 °C for 3 min were used as templates. A 20 µL reaction mixture containing 5 µL of templates, 0.25 µM ASG-Pr, 4 µL of 5 × reaction buffer (TaKaRa), 0.5 mM dNTPs (TaKaRa), 0.5 µL RNAsin (Toyobo), and 0.5 µL of RTase (Toyobo). Reactions were incubated at 42 °C for 30 min. For real-time PCR, 25 µL reaction containing 1 µL of cDNA, 12.5 µL of SYBR Premix Ex Taq II (TaKaRa), and 0.08 µM of each primer were used. The real-time PCR program was performed as follows: at a temperature of 95°C for 3 min, followed by 40 cycles at 95°C for 20 s and then at 60°C for 30 s.

Real-time RT-PCR can recognize all tested CTLV isolates, and no amplification was obtained from healthy plants or samples infected with other citrus viruses or viroids such as *Citrus tristeza virus* (CTV), *Satsuma dwarf virus* (SDV), *Citrus psorosis virus* (CPsV) and *Citrus exocortis viroid* (CEVd).

Two copies per µL CTLV recombinant plasmids were detected by real-time PCR,

and  $2 \times 10^2$  copies per  $\mu\text{L}$  CTLV recombinant plasmids were detected by conventional PCR. The results showed that real-time RT-PCR was 100 times more sensitive than conventional PCR. Although it can detect as few as two copies per  $\mu\text{L}$  CTLV recombinant plasmids, the quantification range was established from  $2 \times 10^2$  to  $2 \times 10^8$  copies per  $\mu\text{L}$  recombinant plasmids due to the reliability of three repetitions (Fig. 1).



CTLV isolates from seven cultivars were successfully detected by real-time RT-PCR. The detectable  $C_t$  values of CTLV RNA-targets ranged from 8.95 to 30.71 (Table 1). Great differences of the CTLV titer were found among several citrus

species, with the lowest in grapefruit, and the highest in citrus hybrid.

TABLE 1  
 $C_t$  VALUE OF CITRUS TATTER LEAF VIRUS IN DIFFERENT CULTIVARS

Cultivars	Threshold cycle ( $C_t$ )
Citrus hybrid	8.95
Lemon	11.75
Jincheng	12.97
Navel orange	18.16
Ponkan	23.16
Huyou	28.90
Grapefruit	30.71

#### ACKNOWLEDGEMENTS

This study was supported by the Natural Science Foundation Project of CQ CSTC (2010BB1149), the Fundamental Research Funds for the Central Universities (XDJK2012C078) and the Program for Changjiang Scholars and Innovative Research Team in University (PCSIRT, IRT0976).

#### LITERATURE CITED

- Broadbent, P., C. M. Dephoff, and C. Gilkeson  
1994. Detection of citrus tatter leaf virus in Australia. *Austral. Plant Pathol.* 23: 20-24.
- Garnsey, S. M. and J. W. Jones  
1968. Relationship of symptoms to the presence of tatter leaf virus in several citrus hosts. In: *Proc. 4<sup>th</sup> Conf. IOCV*, 206-212. IOCV, Riverside CA.
- Guo, L. X., B. C. Xiang, H. Y. Chen, W. J. Duan, H. J. Chen, and S. F. Zhu  
2006. Detection of *Apple stem grooving virus* by real-time fluorescent RT-PCR one step assay. *Acta Phytopathol. Sinica* 36: 57-61.
- Kawai, A. and T. Nishio  
1990. Detection of Citrus tatter leaf virus by enzyme-linked immunosorbent assay (ELISA). *Ann. Phytopathol. Soc. Jap.* 56: 342-345.
- Magome, H., N. Yoshikawa, T. Takahashi, T. Ito, and T. Miyakawa  
1997. Molecular variability of the genomes of *Capilloviruses* from apple, Japanese pear, European pear, and citrus trees. *Mol. Plant Pathol.* 87: 389-396.

6. Miyakawa, T. and T. Masato  
1976. A bud-union abnormality of Satsuma mandarin on *Poncirus trifoliata* rootstock in Japan. In: *Proc. 7<sup>th</sup> Conf. IOCV*, 125-131. IOCV, Riverside CA.
7. Roistacher, C. N.  
1988. Citrus tatter leaf virus: further evidence for a single virus complex. In: *Proc. 10<sup>th</sup> Conf. IOCV*, 353-359. IOCV, Riverside CA.
8. Wallace, J. M. and R. J. Drake  
1962. Tatter leaf, a previously undescribed virus effect in citrus. *Plant Dis. Repr.* 46: 211-212.
9. Zhang, T. M., X. Y. Liang, and C. N. Roistacher  
1988. Occurrence and detection of *Citrus tatter leaf virus* (CTLV) in Huangyan, Zhejiang Province, China. *Plant Dis.* 72: 543-545.
10. Zhou, C.Y., H. Deborah, C. Rachael, P. Barkley, and J. Bowyer  
2001. A micro and rapid nucleotide acid extraction method of *Citrus tristeza virus* for amplification by RT-PCR. *J. Fujian Agric. Univ.* 30 (Suppl.): 200.