

UC Riverside

UC Riverside Previously Published Works

Title

Efficient Dicer processing of virus-derived double-stranded RNAs and its modulation by RIG-I-like receptor LGP2

Permalink

<https://escholarship.org/uc/item/7nb8m3xs>

Journal

PLOS Pathogens, 17(8)

ISSN

1553-7366

Authors

Zhang, Yuqiang

Xu, Yan

Dai, Yunpeng

et al.

Publication Date

2021

DOI

10.1371/journal.ppat.1009790

Peer reviewed

Identification of positive and negative regulators of antiviral RNA interference in *Arabidopsis thaliana*

Si Liu¹, Meijuan Chen ¹, Ruidong Li², Wan-Xiang Li¹, Amit Gal-On³, Zhenyu Jia ²✉ & Shou-Wei Ding ¹✉

Virus-host coevolution often drives virus immune escape. However, it remains unknown whether natural variations of plant virus resistance are enriched in genes of RNA interference (RNAi) pathway known to confer essential antiviral defense in plants. Here, we report two genome-wide association study screens to interrogate natural variation among wild-collected *Arabidopsis thaliana* accessions in quantitative resistance to the endemic cucumber mosaic virus (CMV). We demonstrate that the highest-ranked gene significantly associated with resistance from both screens acts to regulate antiviral RNAi in ecotype Columbia-0. One gene, corresponding to *Reduced Dormancy 5 (RDO5)*, enhances resistance by promoting amplification of the virus-derived small interfering RNAs (vsiRNAs). Interestingly, the second gene, designated *Antiviral RNAi Regulator 1 (VIR1)*, dampens antiviral RNAi so its genetic inactivation by CRISPR/Cas9 editing enhances both vsiRNA production and CMV resistance. Our findings identify positive and negative regulators of the antiviral RNAi defense that may play important roles in virus-host coevolution.

¹Department of Microbiology & Plant Pathology, University of California, Riverside, CA, USA. ²Department of Botany & Plant Sciences, University of California, Riverside, CA, USA. ³Department of Plant Pathology and Weed Science, Volcani Center, Rishon LeZion 7528809, Israel. ✉email: arthur.jia@ucr.edu; shou-wei.ding@ucr.edu

Diverse antiviral defense mechanisms have been documented in higher plants^{1–4}. Several lines of evidence show that the RNA interference (RNAi) pathway mediates an essential antiviral immunity mechanism in plants^{1,2}. In antiviral RNAi, host cells process viral long dsRNA into small interfering RNAs (siRNAs) by a Dicer nuclease to trigger specific virus clearance in RNA-induced silencing complex (RISC) by an Argonaute (AGO) protein^{1,2,5–7}. Both Dicer and AGO gene families appear to have expanded in plants for the control of virus infection. In the model plant species *Arabidopsis thaliana*, for example, 3 of the 4 Dicer-like genes (DCLs) and 7 of the 10 AGO genes participate in the RNAi-mediated antiviral immunity^{1,2,8–17}. Moreover, among the 6 RNA-dependent RNA polymerase (RdRP) genes of *A. thaliana*, RdRP 1 (RDR1), RDR2 and RDR6 have all been shown to direct viral siRNA amplification^{1,2,18,19}. Interestingly, the 19-member AGO family of rice plants includes a hormone-inducible AGO18 that promotes antiviral RNAi by enhancing the expression of AGO1, which is necessary for vsiRNA-RISC assembly^{20,21}. Notably, viral suppressors of RNAi (VSRs) are essential virulence proteins of plant RNA and DNA viruses^{22,23}. Studies on the origins and variability of VSR genes have shown that antiviral RNAi exerts selection pressure against plant virus genomes^{24–27}.

Viruses with an RNA genome possess extraordinary adaptive abilities because of their error-prone replication mechanisms. Less is known about the natural variation that wild host plants in ecosystems with little human intervention accumulate and select to inherit in their progeny in response to virus infection^{1–4}. Recently, whole genome information has become available for 1135 natural inbred lines from worldwide wild-collected accessions of *A. thaliana*²⁸. Cucumber mosaic virus (CMV) is a natural pathogen of *A. thaliana* with incidence reaching up to 80% in some localities²⁹. CMV contains a tripartite positive-strand RNA genome coding for 5 proteins required for genome replication, virion assembly, in-planta movement and RNAi suppression³⁰. VSR protein 2b is encoded by an ‘out-of-frame’ gene that overlaps the more ancient viral RdRP gene and is the most variable viral protein among CMV strains^{31–33}. CMV is transmissible by mechanical contacts and many species of aphids as well as less efficiently through the seed³⁰.

In this work, we investigate whether single nucleotide polymorphisms (SNPs) significantly associated with virus resistance among wild plant populations are enriched in specific pathways known to confer antiviral protection in plants. We use genome-wide association studies (GWAS) approach³⁴ to interrogate natural variation among re-sequenced wild *A. thaliana* populations in quantitative resistance³⁵ to CMV. For a better survey of host natural variation in antiviral responses, we conduct independent GWAS screens with two distantly related CMV strains, one unmodified and the other rendered defective in counter-defense against antiviral RNAi. We show that the highest-ranked gene significantly associated with quantitative virus resistance identified from each of two independent GWAS screens functions in antiviral RNAi. Notably, we demonstrate opposing roles in antiviral RNAi for the two genes evolved in ecotype Columbia-0, indicating that these regulatory genes of antiviral RNAi defense may play important roles in virus-host coevolution.

Results

Mapping host natural variation identifies a quantitative virus resistance gene. We first assessed natural variation among the fully re-sequenced *A. thaliana* accessions in quantitative resistance to CMV- Δ 2b, a previously characterized mutant of the highly virulent subgroup I CMV strain Fny that is rendered susceptible to antiviral RNAi by introducing nucleotide substitutions to prevent VSR-2b expression^{13,36}. Virus accumulation levels in the upper non-

inoculated leaves were measured by enzyme-linked immunosorbent assay (ELISA) of the viral coat protein (CP) two weeks after mechanical inoculation of *A. thaliana* seedlings.

The virus titers in 496 accessions displayed a nearly normal distribution after log-transformation (Fig. 1a and Supplementary Data 1). Analyzing the data with the easyGWAS pipeline³⁴ found that none of the SNPs significantly associated with quantitative virus resistance mapped to any of the known antiviral RNAi genes. The most significantly associated SNP resided in a region of chromosome 4 that codes for the gene At4g11040 (Fig. 1b, c and Supplementary Fig. 1a), corresponding to the previously reported *Reduced Dormancy 5 (RDO5)/Delay of Germination 18 (DOG18)*^{37–39}. We found that CMV- Δ 2b titers were significantly different between the accessions classified as haplotypes T and G according to the SNP (Supplementary Fig. 1b). Northern blot analysis further verified that CMV- Δ 2b replicated to lower levels in 4 selected accessions of haplotype T including Col-0 than the 3 accessions of haplotype G (Supplementary Fig. 1c). We then compared CMV- Δ 2b infection in accession Antwerpen-1 (An-1) carrying a single base-pair (bp) frameshifting deletion³⁹ in *RDO5* with accession Columbia-0 (Col-0), classified in the resistant haplotype along with 380 additional accessions among those examined (Fig. 1c and Supplementary Fig. 1b). Western blot analysis of the viral CP showed that CMV- Δ 2b accumulated to much higher levels in An-1 plants than either Col-0 plants or the two independent lines of An-1 transgenically complemented with the *RDO5* gene from Col-0 (Fig. 1d). These results suggest suppression of CMV- Δ 2b accumulation by *RDO5* from Col-0 plants.

To verify the role of *RDO5* in Col-0 plants, we obtained two Col-0 mutants carrying a T-DNA insertion at different positions in the second exon of *RDO5*, designated *rdo5-4* and *rdo5-5* (Fig. 1c). We also generated two independent transgene complementation lines of *rdo5-4* with the same *RDO5* gene driven by its native promoter described above. ELISA detection of the viral CP revealed significantly enhanced accumulation of CMV- Δ 2b in both *rdo5-4* and *rdo5-5* mutant plants compared to either wild-type Col-0 plants or either of the two *RDO5*-complemented lines of *rdo5-4* plants (Fig. 1e). Moreover, both Western blotting detection of CP and Northern blotting detection of the viral genomic RNAs showed that CMV- Δ 2b accumulated to higher levels in *rdo5-4* and *rdo5-5* mutant plants than wild-type Col-0 plants and the complemented lines of *rdo5-4* mutant (Fig. 1e, f). By comparison, CMV- Δ 2b replicated to higher levels in *rdr6* mutant plants than *rdo5-4* and *rdo5-5* mutant plants (Fig. 1e, f), which may explain the absence of clear symptomatic differences between the infected Col-0 and *rdo5* mutant plants (Supplementary Fig. 2a).

To further verify the role of *RDO5* by an independent approach, we performed gene knockout via CRISPR/Cas9 in Col-0 plants and obtained another homozygous *rdo5* mutant containing a deletion of 344 bp starting from the 12th codon of *RDO5*, designated *cr15* (Fig. 1c and Supplementary Fig. 2b). We found that *cr15* plants also supported significantly enhanced replication of CMV- Δ 2b compared to its wild-type Col-0 and the two transgene complemented lines of *rdo5-4* mutant (Fig. 1e, f). Together, our findings show that the highest-ranked gene in the GWAS screen confers quantitative virus resistance in Col-0 accession.

RDO5 enhances antiviral RNAi by promoting viral siRNA amplification. The seed-specific *RDO5* codes for a nuclear pseudophosphatase that promotes seed dormancy in a manner independent of the biosynthesis of phytohormone abscisic acid^{37–39}. Consistently, *rdo5-4*, *rdo5-5* and *cr15* mutants exhibited

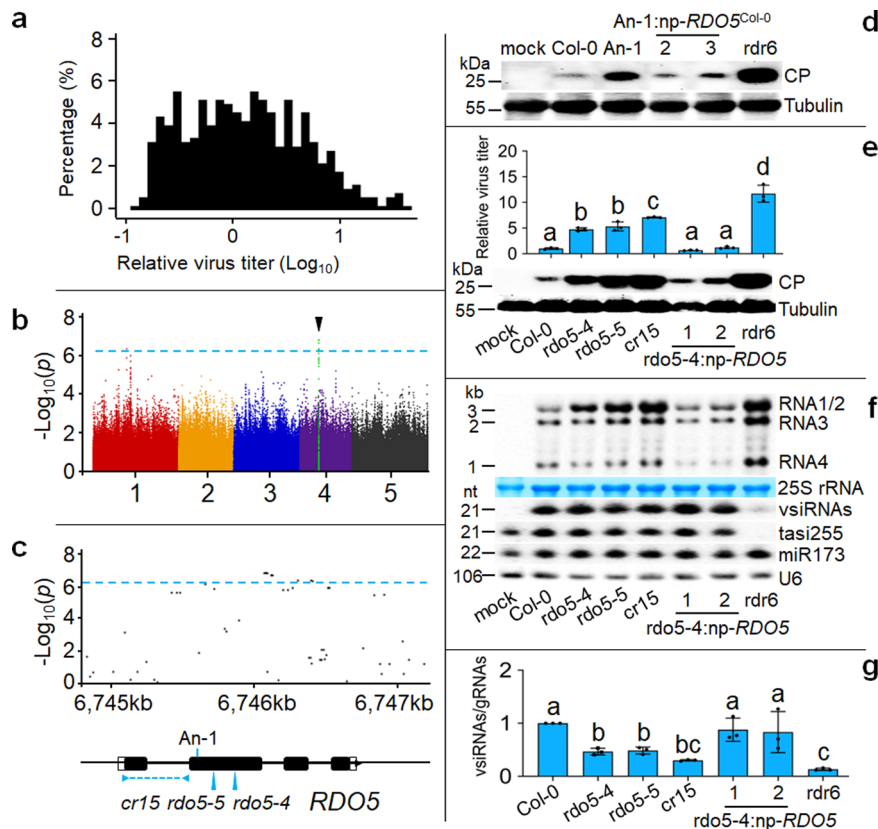


Fig. 1 *RDO5* confers quantitative virus resistance. **a** A nearly normal distribution of CMV- Δ 2b accumulation levels in 496 Arabidopsis accessions after ELISA readings at 2 weeks post-infection were log-transformed. **b** Manhattan plot for GWAS mapping of the quantitative resistance phenotype to CMV- Δ 2b infection. Five *A. thaliana* chromosomes were depicted in different colors. The horizontal dash-dot line corresponds to the significance threshold ($p = 5.48 \times 10^{-7}$). The black triangle above the threshold indicates the most significantly associated locus. **c** Regional Manhattan plot (from 6745 kb to 6747 kb), structure of *RDO5*, and positions of T-DNA insertion alleles (*rdo5-4* and *rdo5-5*), CRISPR/Cas9 deletion allele (*cr15*) and the single base pair deletion in accession An-1. **d** Western detection of viral coat protein (CP) accumulation in An-1 and two *RDO5*-complemented lines. Detection of tubulin alpha chain was shown as loading control. **e, f** ELISA and Western detection (**e**) of viral CP accumulation with the titer in Col-0 set as 1 and Northern detection (**f**) of CMV- Δ 2b genomic RNAs 1-3 (gRNAs), subgenomic RNA4 and vsiRNAs as well as endogenous miRNA 173 (miR173) and trans-acting siRNA 255 (tasi255) in wild-type (Col-0), *rdo5* mutants and complemented lines. 25 S rRNA was stained and U6 RNA probed on the same membrane as loading controls. **g** Ratios of vsiRNAs/gRNAs were calculated from Phosphor-imager readings of Northern hybridization signals in (**f**) with the ratio in Col-0 set as 1. The experiments in (**d**) and (**f**) were repeated three times independently with similar results. Data presented are means \pm SEM from three replicates (**e**) or independent experiments (**g**), letters indicate significant differences (one-way ANOVA, Duncan, $p < 0.05$) and black dots represent the individual values. The source data underlying blots in (**d**), (**e**) and (**f**), ELISA data in (**e**) and ratio data in (**g**) are provided as a Source Data file.

significantly enhanced seed germination rates than Col-0 and the complementation lines of *rdo5-4* and An-1 plants carrying the wild-type *RDO5* transgene derived from Col-0 (Supplementary Fig. 2c), providing evidence for functional rescue by an additional phenotype. Notably, we found that CMV- Δ 2b infection induced expression of *RDO5* in leaves, suggesting a different function of *RDO5* in vegetative tissues in response to virus infection (Supplementary Fig. 2d).

A. thaliana resistance to CMV- Δ 2b is mediated by antiviral RNAi dependent mainly on the DCL4 and RDR6 pathway^{13,19,33}. To investigate the mechanism of *RDO5*, we first compared the accumulation levels of the vsiRNAs in the same panel of wild-type and mutant infected plants used above for determining viral accumulation by Northern blot analysis (Fig. 1f). We found that the vsiRNAs accumulated to similar levels in wild-type Col-0 plants and the three *rdo5* mutants (*rdo5-4*, *rdo5-5* and *cr15*) despite the fact that CMV- Δ 2b replicated to significantly higher levels in the *rdo5* mutant plants (Fig. 1e, f) and were thus expected to generate more abundant vsiRNA precursors for Dicer processing, suggesting deficiency of the mutant plants in the vsiRNA biogenesis. No marked differences were found in the

accumulation of endogenous siRNA or microRNAs between wild-type and *rdo5* mutant plants with or without CMV- Δ 2b infection (Fig. 1f and Supplementary Fig. 2e). In *A. thaliana* plants infected with 2b-deficient mutants of CMV, the vsiRNAs are amplified by the host RDR1 and/or RDR6 so that loss of vsiRNA amplification in mutant plants leads to decreased ratios of vsiRNAs and viral genomic RNAs (gRNAs) although viral gRNAs accumulate to higher levels compared to the wild-type plants^{19,40}. Thus, we measured the ratios of vsiRNAs and viral gRNAs detected in the infected wild-type and mutant plants. As expected, loss of RDR6-dependent vsiRNA amplification in the control *rdr6* mutant plants was associated with \sim 7-fold decrease of the vsiRNAs/gRNAs ratio compared to the resistant Col-0 plants (Fig. 1g). We observed significantly decreased vsiRNAs/gRNAs ratios in *rdo5-4*, *rdo5-5* and *cr15* mutant plants than those in Col-0 plants and the two *RDO5*-complemented lines of *rdo5-4* plants (Fig. 1g), suggesting a role of *RDO5* in the amplification of vsiRNAs.

To further investigate the mechanism of *RDO5*, we generated homozygous double or triple mutant plants by genetic crosses of *rdo5-5* plants with *rdr1* and *rdr6* single or double mutants.

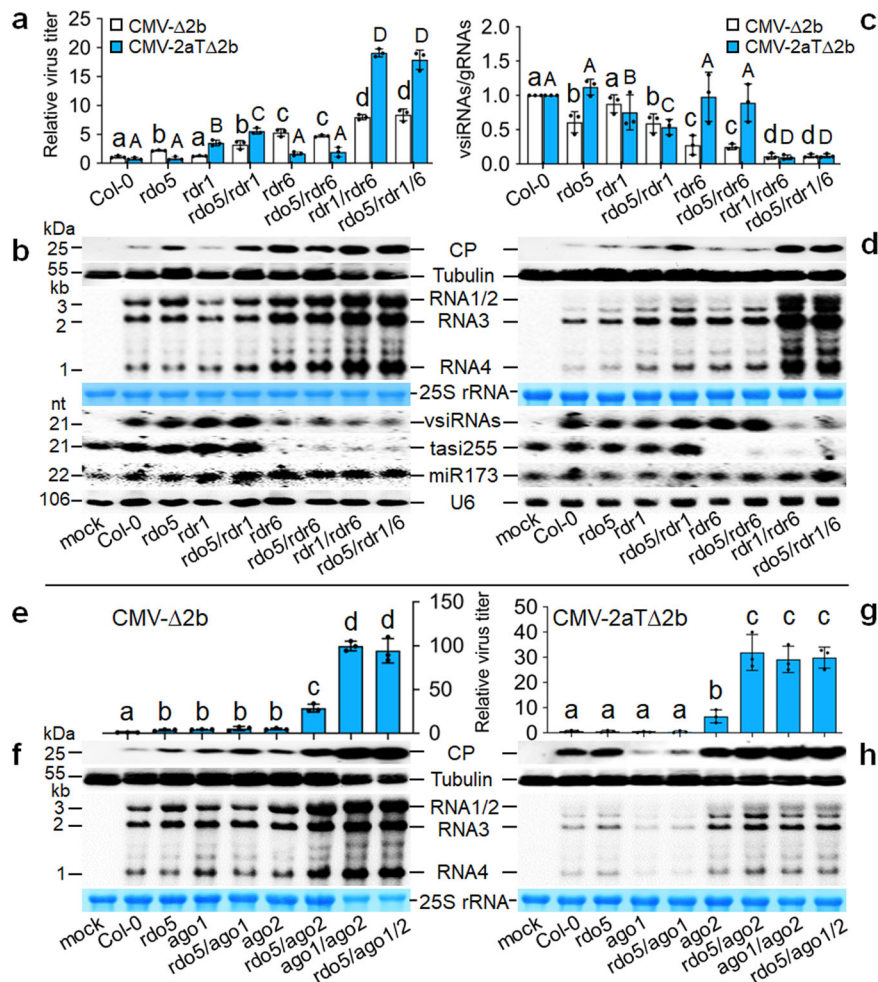


Fig. 2 RDO5 promotes antiviral RNAi. **a–h** Accumulation of CMV-Δ2b (**a, b, e, f**) or CMV-2aTΔ2b (**a, d, g, h**) detected in wild-type (Col-0), single, double or triple mutant plants as indicated at 2 weeks post-infection by ELISA (**a, e, g**) and Western blot analysis of the viral coat protein (CP) or Northern blot analysis of the viral RNAs 1–4 (**b, d, f, h**). Detection of the vsRNAs and plant endogenous small RNAs (**b, d**), and the calculation of vsRNAs/gRNAs ratios (**c**) were as described in the legend to Fig. 1. Note the reduced sample loading (to ½) for total proteins and total RNAs (**f**) from *ago1/2* double mutant plants and *rdo5/ago1/2* triple mutant plants infected with CMV-Δ2b. Data presented are means ± SEM from three replicates (**a, e, g**) or independent experiments (**c**), letters indicate groups with significant differences (one-way ANOVA, Duncan, $p < 0.05$) and black dots represent the individual values. The experiments in (**b**), (**d**), (**f**) and (**h**) were repeated three times independently with similar results. The source data underlying blots in (**b**), (**d**), (**f**) and (**h**), ELISA data in (**a**), (**e**) and (**g**), and ratio data in (**c**) are provided as a Source Data file.

Consistent with a predominantly RDR6-dependent antiviral RNAi against CMV-Δ2b described previously^{13,36}, CMV-Δ2b replicated to significantly lower levels in both wild-type Col-0 and *rdr1* plants than *rdr6* or *rdr1 rdr6* plants (Fig. 2a, b) and vsRNA amplification in *rdr1* plants was as efficient as in Col-0 plants in contrast to defective vsRNA amplification in *rdr6* or *rdr1 rdr6* plants (Fig. 2b, c). We found that CMV-Δ2b accumulation was significantly enhanced and the vsRNAs/gRNAs ratio was significantly decreased in *rdo5 rdr1* double mutant plants compared to *rdr1* single mutant plants (Fig. 2a–c). By contrast, no significant differences in either CMV-Δ2b accumulation or the vsRNAs/gRNAs ratio were observed between *rdr6* and *rdo5 rdr6* plants (Fig. 2a–c). Moreover, neither CMV-Δ2b accumulation nor the vsRNAs/gRNAs ratio was significantly different between *rdr1 rdr6* double mutant plants and *rdo5 rdr1 rdr6* triple mutant plants (Fig. 2a–c). These findings indicate that RDO5 acts specifically in the antiviral RNAi defense mechanism by enhancing vsRNA amplification in an RDR6-dependent pathway.

We performed an additional set of infection experiments to verify the proposed RDR6-dependent antiviral activity of RDO5

using CMV-2aTΔ2b, which contains a 295-nt deletion in RNA 2 of Fny-CMV resulting in both the loss of VSR-2b expression and a C-terminal truncation of the viral RdRP protein¹⁹. Unlike CMV-Δ2b, efficient CMV-2aTΔ2b infection occurs only in *rdr1 rdr6* double mutant plants because it triggers potent amplification of vsRNAs by both RDR1 and RDR6 pathways^{13,19}. ELISA and Western blotting detection of the viral CP as well as Northern blotting detection of the viral RNAs found no significant differences in the accumulation of CMV-2aTΔ2b and the vsRNAs/gRNAs ratios between either wild-type plants (Col-0) or *rdr6* mutant plants with *rdo5* mutant plants (*rdo5-4*, *rdo5-5* and *cr15*) or the two complemented lines of *rdo5-4* plants (Supplementary Fig. 3). These results indicate that RDO5 is dispensable for RDR1-dependent amplification of the vsRNAs. Consistently, enhanced accumulation of CMV-2aTΔ2b and decreased vsRNAs/gRNAs ratio were observed in *rdo5 rdr1* plants compared to *rdr1* plants, but no significant differences in either CMV-2aTΔ2b accumulation or the vsRNAs/gRNAs ratio were observed between *rdr6* and *rdo5 rdr6* plants (Fig. 2a, c, d). Moreover, we detected no significant differences in either CMV-2aTΔ2b accumulation or the vsRNAs/gRNAs ratio between *rdr1*

rdr6 plants and *rdo5 rdr1 rdr6* plants (Fig. 2a, c, d). These findings show that *RDO5* confers virus resistance in the antiviral RNAi pathway dependent on *RDR6*, but independent of *RDR1*.

It is known that *AGO1* and *AGO2* act cooperatively to direct antiviral RNAi against CMV- $\Delta 2b$ by associating with two distinct sets of vsRNAs predominantly with 5'-terminal U and A, respectively¹³. We generated an additional set of homozygous double or triple mutants by combining *rdo5-5* with *ago1-27* (hypomorphic allele) and/or *ago2-1* (null allele). We found that CMV- $\Delta 2b$ replicated to similar levels in *ago1 ago2* and *rdo5 ago1 ago2* plants (Fig. 2e, f). Similarly, no significant differences in virus accumulation were detected between the double and triple mutant plants after infection with CMV-2aT $\Delta 2b$ (Fig. 2g, h). Thus, *RDO5* exhibited no antiviral activity against either CMV- $\Delta 2b$ or CMV-2aT $\Delta 2b$ when both *AGO1* and *AGO2* were not functional, demonstrating that the antiviral activity of *RDO5* is completely dependent on *AGO1* and *AGO2*. Interestingly, whereas both CMV- $\Delta 2b$ and CMV-2aT $\Delta 2b$ replicated to significantly higher levels in *rdo5 ago2* plants than *ago2* plants (Fig. 2e–h), combining *rdo5-5* with *ago1-27* did not enhance the accumulation of neither CMV- $\Delta 2b$ nor CMV-2aT $\Delta 2b$ (Fig. 2e–h), suggesting that *RDO5* was involved in the *AGO1*, but not the *AGO2* pathway.

Together, our mechanistic studies show that identified by GWAS mapping as the highest-ranked gene with natural variation significantly associated with resistance, *RDO5* confers virus resistance in Col-0 plants in the antiviral RNAi pathway by promoting *RDR6*-dependent vsRNA amplification.

Natural variation identifies a host gene that inhibits antiviral defense.

In a parallel GWAS screen, we used a wild-type isolate of subgroup II CMV strain Q (Q-CMV) that is not modified in counter-defense against antiviral RNAi, but causes much weaker disease symptoms than Fny-CMV in Col-0 plants. We measured the accumulation levels of Q-CMV in 500 accessions of *A. thaliana* by detecting the viral genomic RNA3 with quantitative reverse transcription-polymerase chain reaction (RT-qPCR) (Supplementary Fig. 4a and Supplementary Data 1). Using the easyGWAS pipeline³⁴, we did not identify SNPs significantly associated with quantitative resistance in any of the known antiviral RNAi genes as found in the above GWAS screen. The SNP associated most significantly with quantitative resistance against Q-CMV resided in a region of chromosome 5 between genes At5g05130 and At5g05140 (Fig. 3a, b and Supplementary Fig. 4b), neither of which was characterized previously. We found that Q-CMV accumulation levels as measured by RT-qPCR were significantly different between the accessions classified as haplotypes A and G according to the SNP (Supplementary Fig. 4c). Northern blot analysis further verified that Q-CMV replicated to lower levels in 3 selected accessions of haplotype G than the 4 accessions of haplotype A including Col-0 (Supplementary Fig. 4d).

Functional studies in Col-0 plants identified At5g05140 as a regulator of antiviral RNAi, designated *Antiviral RNAi Regulator 1* (*VIR1*). Firstly, we found that expression of *VIR1*, but not At5g05130, was induced in Col-0 plants by infection with Q-CMV, Q-CMV- $\Delta 2b$ or CMV-2aT $\Delta 2b$ (Supplementary Fig. 5a). *VIR1* expression levels were also significantly higher in the selected accessions of haplotype A than those from haplotype G after Q-CMV infection (Supplementary Fig. 5b). As shown in Fig. 3b, we obtained *VIR1* and At5g05130 knockout mutants of Col-0 plants. *vir1-1* mutant contained a T-DNA insertion in the eighth exon of *VIR1*. *cr5* mutant was generated by CRISPR/Cas9 genome editing to delete a 215-bp fragment in the first exon of At5g05130. Neither *vir1-1* nor *cr5* exhibited visible developmental defects (Supplementary Fig. 6a). RT-qPCR analysis showed that

Q-CMV accumulated to significantly lower levels in *vir1-1* mutant plants than Col-0 (Fig. 3c). In contrast, no statistically significant differences in Q-CMV accumulation were found between Col-0 and At5g05130-*cr5* mutant plants or two independent *VIR1*-transgene complementation lines of *vir1-1* (Fig. 3c). Moreover, *vir1-1* plants, but not *cr5* plants or the transgene-complemented lines of *vir1-1*, supported significantly reduced replication of Q-CMV- $\Delta 2b$ (Fig. 3c), a 2b-deletion mutant of Q-CMV defective in suppressing antiviral RNAi initiated by either *DCL2* or *DCL4* shown by a previous study³³. We next calculated and compared the ratio of the vsRNAs/gRNAs detected by Northern blotting (Fig. 3d–f). The results indicated that production of the vsRNAs triggered by either Q-CMV or Q-CMV- $\Delta 2b$ was significantly enhanced in *vir1-1* mutant plants compared to Col-0 plants, *cr5* mutant plants or the two *VIR1*-complemented lines of *vir1-1* (Fig. 3d–f).

To further verify the role of *VIR1*, we generated two additional *vir1* mutants by CRISPR/Cas9 genome editing, designated *cr6-14* and *cr6-31*, which respectively contained an insertion and deletion of single cytosine nucleotide (C) in the 15th codon of *VIR1* (Supplementary Fig. 6b). Similar to *vir1-1*, both *cr6-14* and *cr6-31* mutants exhibited enhanced resistance to either Q-CMV or Q-CMV- $\Delta 2b$ since both viruses accumulated to lower levels and triggered production of more abundant vsRNAs in *cr6-14* and *cr6-31* mutants than Col-0 plants (Fig. 3c–f). However, we observed no obvious differences in the accumulation of endogenous siRNA and miRNAs between wild-type and *vir1* mutant plants either with or without CMV infection (Fig. 3d, f and Supplementary Fig. 6c). These findings indicate that the highest-ranked gene in the independent GWAS screen dampens antiviral RNAi in Col-0 accession against both Q-CMV and Q-CMV- $\Delta 2b$ by inhibiting production of the vsRNAs.

VIR1 is a negative regulator of antiviral RNAi.

The N-terminal region of *VIR1* protein shares strong homology with Arabidopsis transcript elongation factor IIS (*AtTFIIS*) conserved broadly in eukaryotes (Supplementary Fig. 5c). However, most of the critical residues in the C-terminal domain of *TFIIS* essential to facilitate mRNA synthesis by RNA polymerase II complex are not conserved in *VIR1* protein⁴¹. Similar to *rdo5* mutant plants, *rdo2* mutant plants that lack *AtTFIIS* display clearly reduced seed dormancy⁴². We found that *vir1-1* mutant plants exhibited significantly enhanced seed dormancy and that *VIR1* expression was detectable in seeds (Supplementary Fig. 6d, e), suggesting *VIR1* as a negative regulator of seed dormancy.

To determine whether *VIR1* interferes with vsRNA biogenesis, we constructed double and triple mutants by genetic crosses of *vir1-1* plants with *dcl2-1* and/or *dcl4-2* mutant plants characterized previously³³. We challenged this panel of mutant plants with CMV-2aT $\Delta 2b$ and CMV- $\Delta 2b$ derived from Fny-CMV since our previous studies have defined the genetic pathways in the biogenesis of vsRNAs triggered by these mutant viruses^{13,19}. RT-qPCR and Northern blotting analysis showed that CMV-2aT $\Delta 2b$ replicated to significantly lower levels in *vir1-1* mutant plants than Col-0 plants (Fig. 4a, b), as did Q-CMV and Q-CMV- $\Delta 2b$ (Fig. 3c–f). Notably, whereas no significant differences were detected in virus accumulation between *dcl2* and *vir1 dcl2* mutant plants, CMV-2aT $\Delta 2b$ replicated to significantly higher levels in *vir1 dcl4* double mutant plants than *dcl4* single mutant plants (Fig. 4a, b). Consistent with the known dominant role of *DCL4* over *DCL2* in the biogenesis of vsRNAs^{9–11,19,33}, 21-nt vsRNAs made by *DCL4* were undetectable whereas 22-nt vsRNAs by *DCL2* accumulated to high levels in either *dcl4* plants or *vir1 dcl4* plants (Fig. 4b). By comparison, however, 22-nt vsRNAs accumulated to markedly lower levels in *vir1 dcl4* plants than

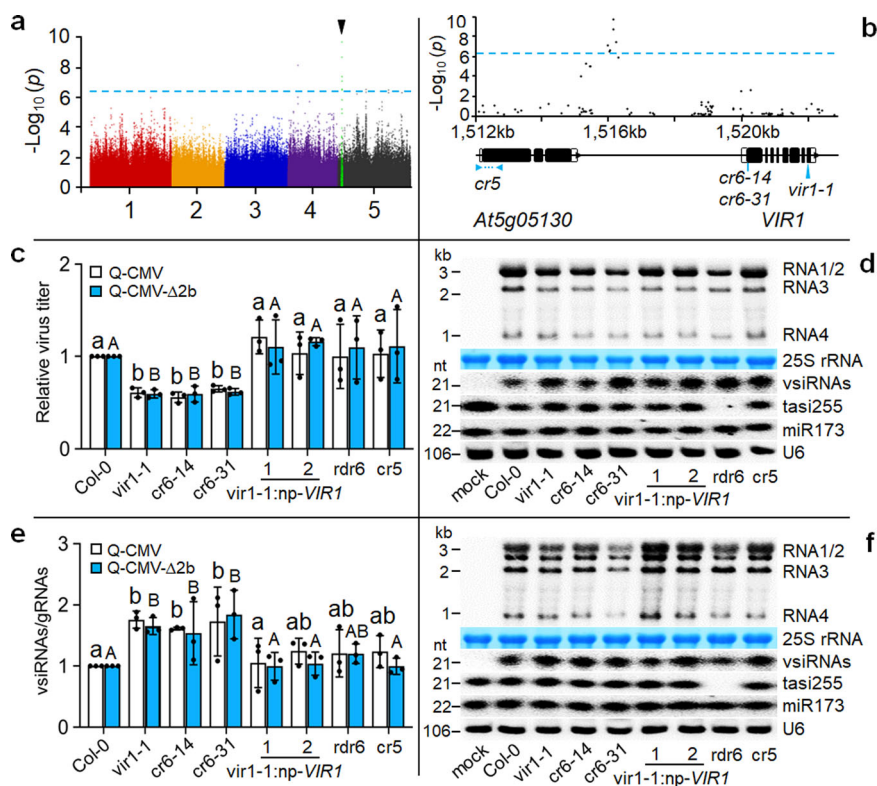


Fig. 3 Identification of *VIR1*. **a** Manhattan plot for GWAS mapping of the susceptibility phenotype to Q-CMV infection. The horizontal dash-dot line corresponds to the significance threshold ($p = 5.45 \times 10^{-7}$). The black triangle above the threshold indicates the most significantly associated locus. **b** Regional Manhattan plot (from 1512 kb to 1522 kb), structure of *VIR1*, and positions of T-DNA insertion allele *vir1-1* and CRISPR/Cas9 genome edited alleles (*cr5* in At5g05130 and *cr6-14* and *cr6-31* in *VIR1*). **c, d, f**, Accumulation of Q-CMV (**c, d**) or Q-CMV- Δ 2b (**c, f**) detected in wild-type (Col-0), *cr5* mutant, *vir1* mutants (*vir1-1*, *cr6-14* and *cr6-31*), or two independent *VIR1*-complemented lines of *vir1-1* at 2 weeks post-infection by RT-qPCR analysis of the viral RNA3 (**c**) or Northern blotting of the viral RNAs 1-4 (**d, f**). The vsRNAs and plant endogenous small RNAs were also detected by Northern blotting (**d, f**). **e** The vsRNAs/gRNAs ratios were calculated as described in the legend to Fig. 1. Data in (**c**) and (**e**) are means \pm SEM from three independent experiments, letters indicate groups with significant differences (one-way ANOVA, Duncan, $p < 0.05$) and black dots represent the individual values. The source data underlying blots in (**d**) and (**f**), qRT-PCR data in (**c**) and ratio data in (**e**) are provided as a Source Data file.

dcl4 plants (Fig. 4b), indicating that activation of DCL2-dependent antiviral RNAi by 22-nt vsRNAs in the absence of DCL4 is upregulated by *VIR1*.

RT-qPCR analysis revealed significant suppression of *DCL4* induction by CMV-2aT Δ 2b in Col-0 plants compared to *vir1-1* mutant plants (Fig. 4e). By contrast, *DCL2* expression was induced by CMV-2aT Δ 2b in *dcl4* single mutant plants, but not in *vir1 dcl4* double mutant plants or any other wild-type and mutant plants examined (Fig. 4e). Reduced induction of *DCL4* in Col-0 plants and strong induction of *DCL2* in *dcl4* mutant plants were both observed after infection with Q-CMV (Supplementary Fig. 7). These findings together suggest a model in which *VIR1* negatively regulates antiviral RNAi in wild-type plants by restricting transcriptional induction of *DCL4*, but upregulates 22-nt vsRNA-directed antiviral RNAi by transcriptional induction of *DCL2* in the absence of *DCL4*.

We observed no statistically significant differences in CMV-2aT Δ 2b accumulation between *dcl2 dcl4* and *vir1 dcl2 dcl4* mutant plants (Fig. 4a, b). This result indicates that although *VIR1* enhanced antiviral defense in *dcl4* single mutant, it failed to do so in *dcl2 dcl4* double mutant, consistent with the proposed role of *DCL2* in the upregulation of antiviral RNAi by *VIR1*. As reported previously³³, neither of the double and triple mutant plants produced 21- or 22-nt vsRNAs. The results from CMV- Δ 2b infection in the same panel of mutants (Fig. 4c, d) essentially reproduced those from CMV-2aT Δ 2b infection (Fig. 4a, b). For example, genetic inactivation of *VIR1* enhanced the resistance to

CMV- Δ 2b in Col-0 plants, but promoted virus susceptibility and reduced DCL2-dependent production of 22-nt vsRNAs in *dcl4* plants (Fig. 4c, d). However, CMV- Δ 2b replicated to the same levels in *dcl2 dcl4* and *vir1 dcl2 dcl4* mutant plants (Fig. 4c, d). Together, our findings indicate that in addition to a role in seed dormancy, *VIR1* negatively regulates antiviral RNAi in Col-0 plants by reducing vsRNA production possibly by restricting transcriptional induction of *DCL4*.

Discussion

In this work, we searched for the natural variation among wild *A. thaliana* populations that is most significantly associated with quantitative resistance to an endemic RNA virus. We conducted independent GWAS screens with wild-type Q-CMV and the VSR-deficient CMV- Δ 2b, respectively. Unlike Fny-CMV, from which CMV- Δ 2b was derived, Q-CMV replicates to higher levels in *dcl2 dcl4* mutant plants than wild-type Col-0 plants^{33,36}, indicating incomplete suppression of antiviral RNAi by Q-CMV. Surprisingly, none of the SNPs significantly associated with quantitative virus resistance from both of our GWAS screens and an additional GWAS screen reported recently by others⁴³ mapped to any of the known antiviral RNAi genes, including AGO2 shown recently to exhibit natural variation in non-host virus resistance⁴⁴. However, genetic studies show that both of the highest-ranked gene significantly associated with quantitative virus resistance identified from each of our GWAS screens

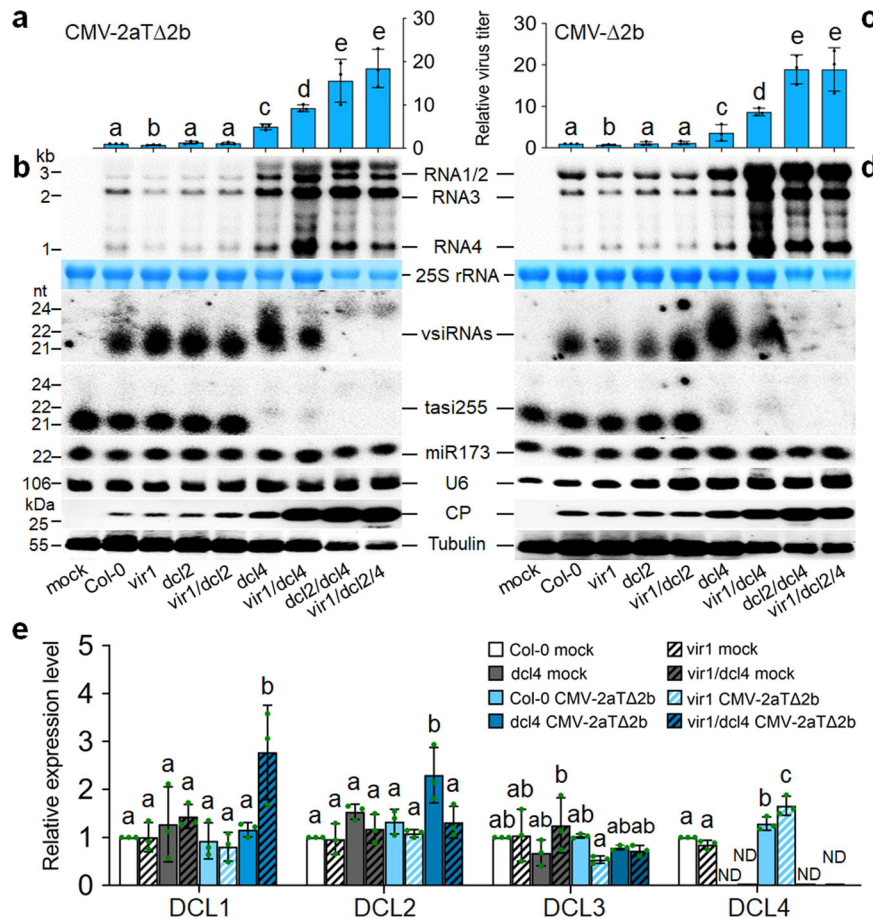


Fig. 4 VIR1 is a negative regulator of antiviral RNAi. **a-d** Accumulation of CMV-2aTΔ2b (**a, b**) or CMV-Δ2b (**c, d**) detected in wild-type (Col-0), single, double or triple mutant plants as indicated at 2 weeks post-infection by RT-qPCR analysis of the viral RNA3 (**a, c**), Northern blotting of the viral RNAs 1-4 (**b, d**), or Western blotting of the viral CP (**b, d**). The vsRNAs and plant endogenous small RNAs were also detected by Northern blotting (**b, d**). Note the reduced sample loading (to ½) for total proteins and total RNAs from *dcl2/4* double mutant plants and *vir1/dcl2/4* triple mutant plants (**b, d**). **e** RT-qPCR analysis of *DCL1*, *DCL2*, *DCL3* and *DCL4* mRNA levels in wild-type (Col-0), single or double mutant plants without infection or one week post-infection with CMV-2aTΔ2b as indicated. *A. thaliana* *EF1α* (*At5g60390*) mRNA was used as internal control. Data in (**a**), (**c**) and (**e**) are means ± SEM from three independent experiments, letters indicate groups with significant differences (one-way ANOVA, Duncan, $p < 0.05$) and green dots represent the individual values. ND not determined. The source data underlying blots in (**b**) and (**d**), and qRT-PCR data in (**a**), (**c**) and (**e**) are provided as a Source Data file.

function in antiviral RNAi. Further studies on the two identified genes evolved in Columbia-0 accession demonstrate opposing roles in antiviral RNAi. Our findings provide direct evidence to support antiviral RNAi as a dominant defense mechanism in virus-host coevolution, which is consistent with previous genetic studies that identify antiviral RNAi as an essential antiviral defense in plants^{1,2,5,6,8-17,20,21}.

Most of the antiviral RNAi pathway genes characterized to date have been identified by their known activity in experimentally induced RNAi to target mRNAs transcribed in the nucleus^{2,7,8,18,19,45-49}. Recently, mutant *Arabidopsis thaliana* and *Caenorhabditis elegans* defective in antiviral RNAi have been isolated by sensitized genetic screens using viruses or viral RNA replicons rendered inactive in RNAi suppression^{36,50-52}. In this work, we showed that antiviral RNAi pathway genes can be identified by GWAS mapping of natural variation in quantitative virus resistance among wild-collected *A. thaliana* accessions. Our findings illustrated the technical feasibility of using GWAS mapping for unbiased identification of host genes critical for virus resistance. Given the increasing availability of genomic sequences for crops^{53,54}, this approach may allow for identification of key plant genes essential for regulating crop resistance to important viral pathogens.

Whereas *VIR1* has not been characterized before this work, *RDO5* has a known function to promote seed dormancy with the mechanism yet to be defined. Our mechanistic studies demonstrate that *RDO5* specifically enhances antiviral RNAi by promoting amplification of the vsRNAs in a pathway dependent on RDR6, but independent of RDR1. By contrast, *VIR1* dampens antiviral RNAi by restricting production of the vsRNAs and suppresses seed dormancy in wild-type Col-0 plants. Our results suggest that *VIR1* may act by blocking viral induction of *DCL4* in a manner similar to the dominant negative mutant version of AtTFIIS shown recently to modify the transcriptome in *tflis* mutant plants⁴¹. Interestingly, *VIR1* is necessary for the transcriptional induction of *DCL2* and the upregulation of *DCL2*-dependent antiviral RNAi by 22-nt vsRNAs in the absence of *DCL4*, which may explain at least in part the dominant role of *DCL4* over *DCL2* in the biogenesis of vsRNAs known since 2006^{9-11,19,33}. However, it is unknown whether *VIR1* modulates the functional roles of *DCL2* that are active in the presence or absence of *DCL4* in uninfected plants^{9,55-58}. Notably, we show that *VIR1* inactivation by CRISPR/Cas9 genome editing confers resistance to CMV either active or defective in RNAi suppression, providing a strategy to generate transgene-free virus resistant plants. In summary, our findings indicate that *RDO5* and *VIR1*

have opposing roles in both antiviral RNAi and seed dormancy. We propose that a shared mechanism is under natural selection to regulate antiviral RNAi and seed dormancy.

Methods

Viruses and plant materials. Mutant viruses CMV- Δ 2b and CMV-2aT Δ 2b were derived from the subgroup I strain Fny-CMV isolated and cloned in New York from a muskmelon farm⁵⁹. In CMV- Δ 2b, three AUG codons at the first (start codon), 8th, and 18th positions of 2b ORF encoded by wild-type RNA2 of Fny-CMV were mutated to ACG so that the amino acids encoded in the +1 overlapping 2a ORF were not altered¹³. CMV-2aT Δ 2b contained a 295-nt deletion in the 2b coding sequence, which also removed the C-terminal 80 amino acids of the viral RdRP 2a protein^{13,19,60}. Q-CMV is a subgroup II strain isolated in Australia and molecularly cloned in 1995 following passages since 1964 in laboratory host plants⁶¹. In Q-CMV- Δ 2b, the 2b coding sequence of RNA2 was deleted and replaced with CCCGGG, which also removed the C-terminal 68 amino acids of the 2a protein^{7,33}. All viruses were purified after propagation in *Nicotiana glauca*. Virion concentration used for mechanical inoculation of *A. thaliana* was 10 μ g/mL for both CMV- Δ 2b and Q-CMV and 20 μ g/mL for both CMV-2aT Δ 2b and Q-CMV- Δ 2b.

A. thaliana natural accessions of CS78942, T-DNA insertion mutants *rdos-4* (SALK_206727), *rdos-5* (SALK_120098) and *vir1-1* (SALK_020870) were obtained from the Arabidopsis Biological Resource Center (<https://abrc.osu.edu>). Single and double mutants in *DCL*, *RDR* and *AGO* families were as described^{9-11,13,19,33,49,62,63}. Additional double and triple mutants of *rdos-4* or *vir1-1* were generated by genetic crosses. Seeds were vernalized at 4 °C in dark for 3 days and seedlings grown in growth room with 10 h light - 14 h dark cycle at 23 °C.

RDO5, *VIR1* and At5g05130 knockout mutants were generated in Col-0 background by CRISPR/Cas9 genome editing as described⁶⁴ with the sequences of guide RNAs (gRNAs) listed in Supplementary Data 2. Briefly, transgenic seeds were screened on MS plates containing 20 μ g/mL hygromycin B (Invitrogen) and T1 plants grown in soil for genotyping by PCR and sequencing with primer pairs listed in Supplementary Data 2. T1 plants with DNA editing in the target gene were backcrossed with Col-0 to identify mutants homozygous for the edited allele without Cas9 transgene. The *RDO5* and *VIR1* genomic fragments (gRDO5 and gVIR1) including 2 kb native promoter region upstream of the start codon, were PCR amplified from Col-0 plants by Phusion DNA polymerase (NEB) with the primer pairs listed in Supplementary Data 2 and verified by sequencing after cloning into pENTR/D-Topo. The resulting pENTR/D-gRDO5 and pENTR/D-gVIR1 were then transferred to PGWB616 by Gateway LR recombination enzyme mix (Invitrogen), yielding PGWB616-gRDO5 and PGWB616-gVIR1, respectively. These constructs were used in transgene complementation for *RDO5* in An-1 and *rdos-4* plants and for *VIR1* in *vir1-1* plants by *Agrobacterium*-mediated plant transformation using the floral dipping method⁶⁵. Transgenic plants were selected with 0.1% basta spraying and self-pollinated to generate two independent lines for each transgene. T3 seedlings were used for virus inoculation.

Genome-wide association studies mapping. Seedlings of 496 and 500 *A. thaliana* accessions (Supplementary Data 1) from the 1001 genome project²⁸ were infected by mechanical inoculation with CMV- Δ 2b and Q-CMV, respectively. The upper systematically infected leaves from 16 plants inoculated per accession were harvested at two weeks post-inoculation and pooled for viral accumulation measurements in three technical replicates. The accumulation level of CMV- Δ 2b was determined by ELISA as described⁶⁶ using the mouse monoclonal antibody (1:2000 dilution) specific to the coat protein (CP) of the CMV subgroup I strain provided kindly by Dr. Xueping Zhou. Briefly, 0.3 g pooled leaves freshly harvested from each accession were ground in 1 mL 0.05 M NaHCO₃ (pH 9.6) with cocktails protease inhibitor (Complete). After centrifugation, the supernatant was diluted with sample loading buffer and used for ELISA detection in triplicate with the CP antibody, alkaline phosphatase-conjugated secondary antibody (Invitrogen, WP20006, 1:2000 dilution), and P-nitrophenyl phosphate (Thermo) using an infinite 200 Pro plate reader (Tecan i-control). The supernatant from mock-inoculated Col-0 plants was included as negative control. Virus titer in each sample was calculated according to the standard curve generated using a serial dilution of CMV- Δ 2b virions (0, 0.1, 0.5, 1, 5, 10 and 20 μ g/mL). The accumulation level of Q-CMV was determined by RT-qPCR detection in triplicate of the viral RNA3 in the total RNA extracted from the pooled leaves of each accession. 1 μ g total RNA was used for reverse transcription (RT) and cDNAs diluted 10 times for subsequent quantitative PCR analysis by CFX96 Real-Time System with SYBR Green Supermix (Bio-Rad). mRNA of *EF1 α* gene (At5g60390) was used as internal reference as reported previously⁶⁷. Primer pairs for the detection of Q-CMV RNA3 and *EF1 α* mRNA were listed in Supplementary Data 2.

Association mapping was conducted based on the 1001 genomes data²⁸ using the easyGWAS pipeline³⁴ (<https://easygwas.ethz.ch/>), TAIR10 gene annotations and the EMMAX algorithms⁶⁸. The minor allele frequency cutoff was set at 5% such that 1,823,394 and 1,833,408 SNPs were used for GWAS mapping of CMV- Δ 2b and Q-CMV data from 496 and 500 accessions, respectively. Based on the calculation using the ANOVA approach, the heritability was 0.9727 for the CMV- Δ 2b screen trait and 0.9059 for the Q-CMV screen trait, respectively. The pairwise Pearson correlations calculated between the three replicates of virus detection were

0.976, 0.970 and 0.972 in CMV- Δ 2b screen, and 0.909, 0.905 and 0.925 in Q-CMV screen. The mapping trait was the mean value of three replicates of the qPCR on the viral RNA3 for Q-CMV screen and the log-transformed mean value of three replicates of ELISA on the viral CP for CMV- Δ 2b screen. We applied a *p* value threshold of 1/*m* to GWAS mapping, where *m* is the number of SNPs used for the analysis⁶⁹⁻⁷². The R program “qqman” was used to create Manhattan plots and quantile-quantile plots based on the results of the GWAS mapping. Two-sample *t* test was performed to compare virus titers of different haplotypes using GraphPad Prism 7, with *p* = 0.05 as the significance level.

Characterization of virus infection. To characterize virus infection, upper systematically infected leaves from 16 plants were harvested at two weeks post-inoculation and pooled for total RNA and protein extraction⁷³. Five and 20 μ g total RNAs were loaded each lane for Northern detection of the viral genomic and vsiRNAs by α -³²P- and γ -³²P-labelled probes, respectively⁷³. Hybridization signals were detected by phosphor imager Typhoon 9410 and analyzed by ImageQuant TL 7.0 (GE Healthcare). Ratios of vsiRNAs vs viral genomic RNAs (gRNAs) were calculated from Phosphor-imager readings of Northern hybridization signals^{19,40}. With the primer pairs listed in Supplementary Data 2 and *EF1 α* mRNA as the internal reference⁶⁷, RT-qPCR was used to detect the accumulation of *RDO5*, *VIR1*, *DCL1*, *DCL2*, *DCL3* or *DCL4* mRNA in upper non-inoculated leaves of plants one week after mock or virus inoculation. For Western blotting, total proteins were separated on 12% polyacrylamide gel before transferred to 0.45 μ m nitrocellulose membrane (GVS North America). Monoclonal antibody (1:5000 dilution) specific to Fny-CMV CP⁶⁶ was used for viral CP detection and probing with *A. thaliana* tubulin alpha chain specific antibody (Agrisera, AS20 4483, 1:5000 dilution) as loading control, Goat anti-mouse IgG (H + L) (Invitrogen, G-21040, 1:2000 dilution) was used as secondary antibody. All experiments were biologically repeated at least three times. We examined the expression pattern of *VIR1* in *A. thaliana* from the Arabidopsis RNA-Seq Database (<http://ipf.sustech.edu.cn/pub/athrna/>)⁷⁴ using 960 libraries generated from Col-0.

Seed dormancy assay. For seed dormancy assays³⁷, 50 newly harvested seeds were sown on wet filter paper in 9 cm diameter culture dishes, and incubated in growth room with 16 h light - 8 h dark cycle at 23 °C. Seed germination rates for each of the genotypes were determined 7 days after incubation with 4 independent repeats.

Reporting summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability

Data supporting the findings of this work are available within the paper and its Supplementary Information files. A reporting summary for this Article is available as a Supplementary Information file. Source data are provided with this paper.

Received: 30 September 2021; Accepted: 18 May 2022;

Published online: 30 May 2022

References

- Baulcombe, D. RNA silencing in plants. *Nature* **431**, 356–363 (2004).
- Guo, Z., Li, Y. & Ding, S. W. Small RNA-based antimicrobial immunity. *Nat. Rev. Immunol.* **19**, 31–44 (2019).
- Li, F. & Wang, A. RNA-targeted antiviral immunity: more than just RNA silencing. *Trends Microbiol.* **27**, 792–805 (2019).
- Meier, N., Hatch, C., Nagalakshmi, U. & Dinesh-Kumar, S. P. Perspectives on intracellular perception of plant viruses. *Mol. Plant Pathol.* **20**, 1185–1190 (2019).
- Ratcliff, F., Harrison, B. D. & Baulcombe, D. C. A similarity between viral defense and gene silencing in plants. *Science* **276**, 1558–1560 (1997).
- Hamilton, A. J. & Baulcombe, D. C. A species of small antisense RNA in posttranscriptional gene silencing in plants. *Science* **286**, 950–952 (1999).
- Li, H. W., Li, W. X. & Ding, S. W. Induction and suppression of RNA silencing by an animal virus. *Science* **296**, 1319–1321 (2002).
- Morel, J. B. et al. Fertile hypomorphic ARGONAUTE (*ago1*) mutants impaired in post-transcriptional gene silencing and virus resistance. *Plant Cell* **14**, 629–639 (2002).
- Bouche, N., Laussergues, D., Gascioli, V. & Vaucheret, H. An antagonistic function for Arabidopsis DCL2 in development and a new function for DCL4 in generating viral siRNAs. *EMBO J.* **25**, 3347–3356 (2006).
- Fusaro, A. F. et al. RNA interference-inducing hairpin RNAs in plants act through the viral defence pathway. *EMBO Rep.* **7**, 1168–1175 (2006).
- Deleris, A. et al. Hierarchical action and inhibition of plant Dicer-like proteins in antiviral defense. *Science* **313**, 68–71 (2006).

12. Raja, P., Jackel, J. N., Li, S., Heard, I. M. & Bisaro, D. M. Arabidopsis double-stranded RNA binding protein DRB3 participates in methylation-mediated defense against geminiviruses. *J. Virol.* **88**, 2611–2622 (2014).
13. Wang, X. B. et al. The 21-nucleotide, but not 22-nucleotide, viral secondary small interfering RNAs direct potent antiviral defense by two cooperative argonautes in *Arabidopsis thaliana*. *Plant Cell* **23**, 1625–1638 (2011).
14. Garcia-Ruiz, H. et al. Roles and programming of Arabidopsis ARGONAUTE proteins during Turnip mosaic virus infection. *PLoS Pathog.* **11**, e1004755 (2015).
15. Harvey, J. J. et al. An antiviral defense role of AGO2 in plants. *PLoS ONE* **6**, e14639 (2011).
16. Brosseau, C. & Moffett, P. Functional and genetic analysis identify a role for Arabidopsis ARGONAUTE5 in antiviral RNA silencing. *Plant Cell* **27**, 1742–1754 (2015).
17. Silva-Martins, G., Bolaji, A. & Moffett, P. What does it take to be antiviral? An Argonaute-centered perspective on plant antiviral defense. *J. Exp. Bot.* **71**, 6197–6210 (2020).
18. Mourrain, P. et al. Arabidopsis SGS2 and SGS3 genes are required for posttranscriptional gene silencing and natural virus resistance. *Cell* **101**, 533–542 (2000).
19. Wang, X. B. et al. RNAi-mediated viral immunity requires amplification of virus-derived siRNAs in *Arabidopsis thaliana*. *Proc. Natl Acad. Sci. USA* **107**, 484–489, <https://doi.org/10.1073/pnas.0904086107> (2010).
20. Wu, J. et al. Viral-inducible Argonaute18 confers broad-spectrum virus resistance in rice by sequestering a host microRNA. *eLife* **4**, <https://doi.org/10.7554/eLife.05733> (2015).
21. Yang, Z. et al. Jasmonate signaling enhances RNA silencing and antiviral defense in rice. *Cell Host Microbe* **28**, 89–103.e108 (2020).
22. Csorba, T., Kontra, L. & Burgyan, J. Viral silencing suppressors: tools forged to fine-tune host-pathogen coexistence. *Virology* **479–480**, 85–103 (2015).
23. Li, F. & Ding, S. W. Virus counterdefense: diverse strategies for evading the RNA-silencing immunity. *Annu Rev. Microbiol.* **60**, 503–531 (2006).
24. Murray, G. G., Kosakovsky Pond, S. L. & Obbard, D. J. Suppressors of RNAi from plant viruses are subject to episodic positive selection. *Proc. Biol. Sci. / R. Soc.* **280**, 20130965 (2013).
25. Kalyandurg, P. B. et al. Efficient RNA silencing suppression activity of Potato Mop-Top Virus 8K protein is driven by variability and positive selection. *Virology* **535**, 111–121 (2019).
26. Pavesi, A. Asymmetric evolution in viral overlapping genes is a source of selective protein adaptation. *Virology* **532**, 39–47 (2019).
27. Obbard, D. J., Gordon, K. H., Buck, A. H. & Jiggins, F. M. The evolution of RNAi as a defence against viruses and transposable elements. *Philos. Trans. R. Soc. Lond. Ser. B, Biol. Sci.* **364**, 99–115 (2009).
28. Genomes Consortium. Electronic address, m. n. g. o. a. a. & Genomes, C. 1,135 Genomes Reveal the Global Pattern of Polymorphism in *Arabidopsis thaliana*. *Cell* **166**, 481–491 (2016).
29. Montes, N., Alonso-Blanco, C. & Garcia-Arenal, F. Cucumber mosaic virus infection as a potential selective pressure on *Arabidopsis thaliana* populations. *PLoS Pathog.* **15**, e1007810 (2019).
30. Palukaitis, P. & Garcia-Arenal, F. Cucumoviruses. *Adv. Virus Res.* **62**, 241–323 (2003).
31. Ding, S. W., Li, W. X. & Symons, R. H. A novel naturally-occurring hybrid gene encoded by a plant RNA virus facilitates long-distance virus movement. *EMBO J.* **14**, 5762–5772 (1995).
32. Li, H. W. et al. Strong host resistance targeted against a viral suppressor of the plant gene silencing defence mechanism. *EMBO J.* **18**, 2683–2691 (1999).
33. Diaz-Pendon, J. A., Li, F., Li, W. X. & Ding, S. W. Suppression of antiviral silencing by cucumber mosaic virus 2b protein in *Arabidopsis* is associated with drastically reduced accumulation of three classes of viral small interfering RNAs. *Plant Cell* **19**, 2053–2063 (2007).
34. Grimm, D. G. et al. easyGWAS: a cloud-based platform for comparing the results of genome-wide association studies. *Plant Cell* **29**, 5–19 (2017).
35. Corwin, J. A. & Kliebenstein, D. J. Quantitative resistance: more than just perception of a Pathogen. *Plant Cell* **29**, 655–665 (2017).
36. Guo, Z. et al. Lipid flippases promote antiviral silencing and the biogenesis of viral and host siRNAs in *Arabidopsis*. *Proc. Natl Acad. Sci. USA* **114**, 1377–1382 (2017).
37. Xiang, Y. et al. Reduced Dormancy5 encodes a protein phosphatase 2C that is required for seed dormancy in *Arabidopsis*. *Plant Cell* **26**, 4362–4375 (2014).
38. Liu, F., Zhang, H., Ding, L., Soppe, W. J. J. & Xiang, Y. Reversal of RDO5 1, a Homolog of rice seed dormancy4, interacts with bHLH57 and controls ABA Biosynthesis and Seed Dormancy in *Arabidopsis*. *Plant Cell* **32**, 1933–1948 (2020).
39. Xiang, Y. et al. Sequence polymorphisms at the REDUCED DORMANCY5 Pseudophosphatase underlie natural variation in *Arabidopsis* Dormancy. *Plant Physiol.* **171**, 2659–2670 (2016).
40. Tong, X. et al. A small peptide inhibits siRNA amplification in plants by mediating autophagic degradation of SGS3/RDR6 bodies. *EMBO J.* e108050, <https://doi.org/10.15252/embj.2021108050> (2021).
41. Antosz, W. et al. Critical role of transcript cleavage in *Arabidopsis* RNA polymerase II transcriptional elongation. *Plant Cell* **32**, 1449–1463 (2020).
42. Grasser, M. et al. Transcript elongation factor TFIIS is involved in *Arabidopsis* seed dormancy. *J. Mol. Biol.* **386**, 598–611 (2009).
43. Rubio, B. et al. Genome-wide association study reveals new loci involved in *Arabidopsis thaliana* and Turnip mosaic virus (TuMV) interactions in the field. *N. Phytol.* **221**, 2026–2038 (2019).
44. Brosseau, C. et al. Natural variation in the *Arabidopsis* AGO2 gene is associated with susceptibility to potato virus X. *N. Phytol.* **226**, 866–878 (2020).
45. Dalmay, T., Hamilton, A., Rudd, S., Angell, S. & Baulcombe, D. C. An RNA-dependent RNA polymerase gene in *Arabidopsis* is required for posttranscriptional gene silencing mediated by a transgene but not by a virus. *Cell* **101**, 543–553 (2000).
46. Dalmay, T., Horsefield, R., Braunstein, T. H. & Baulcombe, D. C. SDE3 encodes an RNA helicase required for post-transcriptional gene silencing in *Arabidopsis*. *EMBO J.* **20**, 2069–2078 (2001).
47. Li, Y., Lu, J., Han, Y., Fan, X. & Ding, S. W. RNA interference functions as an antiviral immunity mechanism in mammals. *Science* **342**, 231–234 (2013).
48. Maillard, P. V. et al. Antiviral RNA interference in mammalian cells. *Science* **342**, 235–238 (2013).
49. Xie, Z. et al. Genetic and functional diversification of small RNA pathways in plants. *PLoS Biol.* **2**, E104 (2004).
50. Coffman, S. R. et al. *Caenorhabditis elegans* RIG-I homolog mediates antiviral RNA interference downstream of Dicer-Dependent Biogenesis of viral small interfering RNAs. *mBio* **8**, <https://doi.org/10.1128/mBio.00264-17> (2017).
51. Long, T., Meng, F. & Lu, R. Transgene-assisted genetic screen identifies rsd-6 and novel genes as key components of antiviral RNA interference in *Caenorhabditis elegans*. *J. Virol.* **92**, <https://doi.org/10.1128/JVI.00416-18> (2018).
52. Guo, Z. et al. Identification of a new host factor required for antiviral RNAi and amplification of viral siRNAs. *Plant Physiol.* **176**, 1587–1597 (2018).
53. Li, N. et al. Natural variation in ZmFBL41 confers banded leaf and sheath blight resistance in maize. *Nat. Genet.* **51**, 1540–1548 (2019).
54. Liu, Y. et al. Genomic basis of geographical adaptation to soil nitrogen in rice. *Nature* **590**, 600–605 (2021).
55. Mlotshwa, S. et al. DICER-LIKE2 plays a primary role in transitive silencing of transgenes in *Arabidopsis*. *PLoS ONE* **3**, e1755 (2008).
56. Wu, H. et al. Plant 22-nt siRNAs mediate translational repression and stress adaptation. *Nature* **581**, 89–93 (2020).
57. Taochy, C. et al. A genetic screen for impaired systemic RNAi highlights the crucial role of DICER-LIKE 2. *Plant Physiol.* **175**, 1424–1437 (2017).
58. Qin, C. et al. Roles of Dicer-Like Proteins 2 and 4 in Intra- and intercellular antiviral silencing. *Plant Physiol.* **174**, 1067–1081 (2017).
59. Rizzo, T. M. & Palukaitis, P. Construction of full-length cDNA clones of cucumber mosaic virus RNAs 1, 2 and 3: generation of infectious RNA transcripts. *Mol. Gen. Genet.* **222**, 249–256 (1990).
60. Soards, A. J., Murphy, A. M., Palukaitis, P. & Carr, J. P. Virulence and differential local and systemic spread of cucumber mosaic virus in tobacco are affected by the CMV 2b protein. *Mol. Plant Microbe Interact.* **15**, 647–653 (2002).
61. Ding, S. W. et al. Efficient infection from cDNA clones of cucumber Mosaic Cucumovirus RNAs in a new plasmid vector. *J. Gen. Virol.* **76**, 459–464 (1995).
62. Xie, Z., Allen, E., Wilken, A. & Carrington, J. C. DICER-LIKE 4 functions in trans-acting small interfering RNA biogenesis and vegetative phase change in *Arabidopsis thaliana*. *Proc. Natl Acad. Sci. USA* **102**, 12984–12989 (2005).
63. Henderson, I. R. et al. Dissecting *Arabidopsis thaliana* DICER function in small RNA processing, gene silencing and DNA methylation patterning. *Nat. Genet.* **38**, 721–725 (2006).
64. Wang, Z. P. et al. Egg cell-specific promoter-controlled CRISPR/Cas9 efficiently generates homozygous mutants for multiple target genes in *Arabidopsis* in a single generation. *Genome Biol.* **16**, 144 (2015).
65. Zhang, X., Henriques, R., Lin, S. S., Niu, Q. W. & Chua, N. H. Agrobacterium-mediated transformation of *Arabidopsis thaliana* using the floral dip method. *Nat. Protoc.* **1**, 641–646 (2006).
66. Yu, C., Wu, J. & Zhou, X. Detection and subgrouping of Cucumber mosaic virus isolates by TAS-ELISA and immunocapture RT-PCR. *J. Virol. Methods* **123**, 155–161 (2005).
67. Lilly, S. T., Drummond, R. S., Pearson, M. N. & MacDiarmid, R. M. Identification and validation of reference genes for normalization of transcripts from virus-infected *Arabidopsis thaliana*. *Mol. Plant Microbe Interact.* **24**, 294–304 (2011).
68. Kang, H. M. et al. Variance component model to account for sample structure in genome-wide association studies. *Nat. Genet.* **42**, 348–354 (2010).
69. Zhang, Y. M., Jia, Z. & Dunwell, J. M. Editorial: The applications of new multi-locus GWAS methodologies in the Genetic Dissection of Complex Traits. *Front. plant Sci.* **10**, 100 (2019).

70. Li, H. et al. Genome-wide association study dissects the genetic architecture of oil biosynthesis in maize kernels. *Nat. Genet.* **45**, 43–50 (2013).
71. Liu, S. et al. A single-nucleotide mutation in a GLUTAMATE RECEPTOR-LIKE gene confers resistance to Fusarium Wilt in *Gossypium hirsutum*. *Adv. Sci. (Weinh.)* **8**, 2002723 (2021).
72. Gong, Q. et al. A Phi-Class Glutathione S-Transferase Gene for Vercillium Wilt Resistance in *Gossypium arboreum* Identified in a Genome-Wide Association Study. *Plant Cell Physiol.* **59**, 275–289 (2018).
73. Cao, M. et al. Virus infection triggers widespread silencing of host genes by a distinct class of endogenous siRNAs in *Arabidopsis*. *Proc. Natl Acad. Sci. USA* **111**, 14613–14618 (2014).
74. Zhang, H. et al. A comprehensive online database for exploring approximately 20,000 Public *Arabidopsis* RNA-Seq Libraries. *Mol. Plant* **13**, 1231–1233 (2020).

Acknowledgements

We wish to thank Drs. Daoxin Xie and Xueping Zhou for sharing materials. This project was supported by funding from the Agricultural Experimental Station and College of Natural and Agricultural Sciences, the University of California, Riverside (to S.-W.D.) and by grants from the US-Israel Binational Agricultural Research and Development Fund (no. IS-5027-17C to A.G.O. and S.-W.D.) and Office of Sponsored Research of the King Abdullah University of Science and Technology, Saudi Arabia (no. OSR-2015-CRG4-2647 to Drs. Magdy Mahfouz and S.-W.D.).

Author contributions

S.L. designed and performed experiments, analyzed data, and wrote the paper. S.-W.D. conceived, designed, and supervised the study, and wrote the paper. Z.J. designed and supervised the GWAS screens and statistical analysis, and wrote the paper. M.C., R.L., W.-X.L. performed experiments or analyzed data. A.G.-O. provided intellectual input to the study. All authors revised and provided feedback for the final version of the paper.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41467-022-30771-0>.

Correspondence and requests for materials should be addressed to Zhenyu Jia or Shou-Wei Ding.

Peer review information *Nature Communications* thanks Yong Jiang and the other, anonymous, reviewer(s) for their contribution to the peer review of this work. Peer reviewer reports are available.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022