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Pharmacokinetics and Pharmacological Properties of Chloroquine and Hydroxychloroquine in the Context of COVID-19 Infection

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Key words: Azithromycin, Chloroquine, Hydroxychloroquine, COVID-19, SARS-CoV-2, Coronavirus, Pharmacokinetics, Safety.

Disclaimer: Abhay Joshi and Jenny H Zheng are employees of the Food and Drug Administration (FDA). This article reflects the views of the authors and should not be construed to represent FDA's views or policies.

#### CONFLICT OF INTEREST

Matthew Rizk and Philip Sabato are employees of Merck & Co Inc. David Wesche is an employee of Certara. Jack Cook is an employee of Pfizer Inc. Melanie Nicol and Rada Savic are co-investigators on separate clinical trials evaluating hydroxychloroquine. All other authors declared no competing interests for this work.

#### FUNDING

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#### ABSTRACT

Chloroquine and hydroxychloroquine are quinoline derivatives used to treat malaria. To date, these medications are not approved for the treatment of viral infections and there are no well-controlled, prospective, randomized clinical studies or evidence to support their use in patients with Coronavirus Infectious Disease-2019 (COVID-19). Nevertheless, chloroquine and hydroxychloroquine are being studied alone or in combination with other agents to assess their effectiveness in the treatment or prophylaxis for COVID-19. The effective use of any medication involves an understanding of its pharmacokinetics (PK), safety and mechanism of action. This work provides basic clinical pharmacology information relevant for planning and initiating COVID-19 clinical studies with chloroquine or hydroxychloroquine, summarizes safety data from healthy volunteer studies, and summarizes safety data from Phase 2 and Phase 2/3 clinical studies in patients with uncomplicated

malaria, including a Phase 2/3 study in pediatric patients following administration of azithromycin and chloroquine in combination. In addition, this work presents data describing the proposed mechanisms of action against the severe acute respiratory distress syndrome (ARDS) coronavirus–2 (SARS-CoV-2) and summarizes clinical efficacy to date.

Pfizer clinical trials cited: A0661139

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# INTRODUCTION

Since coronavirus disease 2019 (COVID-19), caused by the severe acute respiratory distress syndrome (ARDS) coronavirus–2 (SARS-CoV-2), first emerged in late 2019, there has been a widespread search for effective preventive and treatment options to ameliorate the devastating pandemic. As vaccine availability is likely 12 or more months away, there is an immediate need to identify therapeutic options. Early interest, experimental usage, clinical studies, and emergency use authorizations (EUAs) have focused on several potential therapies including antivirals such as remdesivir or antimalarial agents such as chloroquine (1) and hydroxychloroquine (2) with or without the antibacterial agent azithromycin (3), which has been previously used in the treatment of chloroquine-resistant malaria given reported in vitro synergy (4).

With the recent results from the RECOVERY trial (NCT04381936) as well as the announced suspension of hydroxychloroquine arms in several other trials, there is decreasing support for its use in hospitalized or severely ill patients. However, many trials, particularly in prevention or early treatment settings, or in specific subpopulations, are ongoing. This publication seeks to summarize the current state of knowledge for both chloroquine and hydroxychloroquine, including pharmacokinetic (PK) properties, known pharmacology and evidence related to use in treatment of COVID-19. Prior to the current pandemic, more studies had been reported with use of chloroquine than hydroxychloroquine, thus there is notably more information available for chloroquine regarding metabolism, safety, and overall use.

# CHEMICAL STRUCTURES

Both chloroquine and hydroxychloroquine are orally administered 4-aminoquinoline antimalarial compounds. The chemical structures of chloroquine and hydroxychloroquine are shown in Figure 1. Chloroquine is administered as chloroquine phosphate and has a molecular weight of 515.9 g/mol (Base: 319.9 g/mol). Hydroxychloroquine is administered as hydroxychloroquine sulfate and has a molecular weight of 433.95 g/mol (Base: 335.9 g/mol).

#### CLINICAL PHARMACOLOGY SUMMARY

Table S1 summarizes limited information on the clinical PK of chloroquine and hydroxychloroquine that is available from ARALEN® (CHLOROQUINE PHOSPHATE, USP) (5) and PLAQUENIL® (HYDROXYCHLOROQUINE SULFATE TABLETS, USP) (6) US product labels.

Information on the PK of pharmacologically active metabolites of chloroquine and hydroxychloroquine is discussed in multiple publications (7-21). A summary of the compiled information is provided below.

#### Absorption

# Chloroquine

The majority of the available PK data in literature are from healthy volunteers receiving a single oral dose containing 300 mg or 600 mg chloroquine base (Table 1). Cross study comparisons of exposure parameter estimates (Maximum concentration (Cmax), area under the concentration-time curve (AUC), and time to reach Cmax (Tmax)) were not pursued, as different formulations were used across the different studies. Gustafsson et. al. reported the mean ( $\pm$  SD) bioavailability of chloroquine from oral tablet as 89  $\pm$  16% (22). Tmax ranged from 1 to 6 hours across studies and this variability could be due to in vivo PK variability, formulation differences, and/or administration conditions. In addition, despite being in use for years, the available PK data on the effect of food on chloroquine absorption are limited.

# Hydroxychloroquine

Most of the available PK data in literature are from healthy volunteers receiving a single oral dose of 155 mg hydroxychloroquine base (Table 2). Cross study comparisons of exposure parameter estimates (Cmax, AUC, and Tmax) were not pursued, as the exact information on different formulations that were used across the different studies was not available. Tett et. al. reported the mean ( $\pm$  SD) fraction of the oral dose absorbed as 0.74 ( $\pm$  0.13) based on the blood and urine PK data (16). McLachlan et. al. reported similar findings with the reported mean fraction absorbed as 0.79 in nine patients with rheumatoid arthritis who received two doses of 155 mg racemic hydroxychloroquine base each, as a tablet and by intravenous infusion (25). In a separate study,

McLachlan et. al. reported the lack of food-effect on the extent of absorption of hydroxychloroquine, except that the absorption lag-time appeared to be significantly prolonged in the presence of food (26). Tett et. al. evaluated between and within subject PK variability in six healthy volunteers and reported that the between subject variability (BSV) in relative bioavailability was (27-38%) higher than the within subject variability (11-16%) (27). Based on these findings, Tett et. al. suggests a need for individualization of dosing to target concentrations associated with optimal outcomes and may minimize variability in response (27). The available PK data for hydroxychloroquine are limited to ascertain PK linearity.

McLachlan et. al. evaluated blood and plasma concentrations of the enantiomers of hydroxychloroquine in a separate study and reported that (R)-hydroxychloroquine had higher blood (Ratio= 2.2, Range= 1.6-2.9) and plasma (Ratio= 1.6, Range= 1.2-1.9) concentrations compared to the (S)-enantiomer (30). Ducharme et. al. noted similar findings with reported  $62 \pm 3\%$  (mean  $\pm$  SD) of the AUC of rac-hydroxychloroquine AUC for R(-)-hydroxychloroquine (17).

#### Distribution

#### Chloroquine

Estimates reported for apparent volume of distribution (V/F) for chloroquine are relatively large and with notable variation (Table 1). Edwards et. al. reported that chloroquine concentrations in packed cells were higher than the concentrations in plasma with a median ratio (cell:plasma) between 3 and 4. Similar estimates were reported by Gustafsson et. al. with a mean ratio (cell:plasma) of 4.8 (range: 2-5) (22). Preclinical studies indicate it is widely distributed in the liver, spleen, kidney and lung, with concentrations several hundred-fold above plasma concentrations (5). Walker et. al. evaluated plasma protein binding for chloroquine using equilibrium dialysis and reported mean protein binding as  $61 \pm 9\%$  (range 46-74%) in plasma from healthy subjects and  $64 \pm 7\%$  (range 55-79%) in plasma from patients with rheumatoid arthritis (31). Ofori-Adjei et. al. also evaluated protein binding estimates of 59% for chloroquine that is within the range reported by Walker et. al. (32). The findings reported by Ofori-Adjei et. al. suggest chloroquine protein binding is concentration independent over

the range of 25-400 ng/mL and (S)-chloroquine binds more to plasma (~67%) than (R)-chloroquine (~49%). Chloroquine is also reported to transfer via placenta and into milk (33, 34).

#### Hydroxychloroquine

The reported mean ( $\pm$  SD) blood to plasma hydroxychloroquine concentration ratio is 7.2 ( $\pm$ 4.2) (15). Estimated protein binding is between 30% and 50% and hydroxychloroquine is reported to bind to both albumin and alpha, glycoprotein (35, 36). The findings reported by McLachlan et. al. (35) suggest hydroxychloroquine demonstrates stereoselective protein binding, with (S)-hydroxychloroquine binding more to plasma (~64%) than (R)-hydroxychloroquine (~37%), resulting in approximately 52% average protein binding for racemic mixture. Hydroxychloroquine is reported to transfer via placenta and into milk (19, 37-39).

### **Metabolism and Drug Interaction**

#### Chloroquine

Information on the metabolism and excretion of chloroquine is scarce in the literature including information on specific metabolism pathways. Ducharme et. al. discussed the chloroquine metabolism data from literature and concluded that CYP3As and CYP2D6 are two enzymes affected or are involved in chloroquine metabolism (40). Projean et. al. concluded based on the investigations of chloroquine metabolism in human liver microsomes and recombinant human CYP450s that chloroquine would be metabolized into N-desethylchloroquine primarily via CYP2C8 and CYP3A4 (41). The same study noted that at low chloroquine concentrations, CYP2D6 may also play a significant role. Based on literature review, it is proposed that the co-administration of drugs that modulate CYP2C8, CYP3A4, and CYP2D6 could have potential effects on the PK of chloroquine (42). Based on in vitro studies, chloroquine is a weak inhibitor of CYP2D6 but it is unlikely to have a significant effect on the PK of other CYP2D6 substrates in human (42). Additional information on the potential of drug-drug interaction with chloroquine as object and precipitant is detailed by Kiang et. al. (43-45). Table 3 presents results of clinical drug interaction studies involving chloroquine as the precipitant and as the object.

#### Hydroxychloroquine

Similar to chloroquine, information on the metabolism and excretion of hydroxychloroquine is scarce in the literature, including information on specific metabolism pathways. Hydroxychloroquine demonstrates similar PK properties to chloroquine. Hydroxychloroquine's major metabolite is desethylhydroxychloroquine due to its metabolism by Cytochrome P450 enzymes CYP2D6, 2C8, 3A4 and 3A5 (53). Hydroxychloroquine is primarily eliminated through the kidneys. Rainsford, et. al. discusses the information on hydroxychloroquine metabolism pathway and potential drug interactions from the literature (19). In this review, the most significant drug interactions of relevance noted are with methotrexate. Based on the findings from the two studies (28, 50), Rainsford, et. al. concluded co-administration of hydroxychloroquine with methotrexate causes reduced Cmax while delaying Tmax and lack of any significant drug interactions related to effects on the PK of hydroxychloroquine by methotrexate. In addition, hydroxychloroquine is considered a weak inhibitor of CYP2D6 and 400 mg daily hydroxychloroquine dose increased metoprolol by 50.7% (54, 55). Table 4 presents results of clinical drug interaction studies involving hydroxychloroquine as the precipitant and the object, respectively.

#### Excretion

#### Chloroquine

The estimates for elimination half-life for chloroquine vary significantly (Table 1). Following multiple doses, chloroquine elimination half-life is reported to range from 30 to 60 days (57, 58). Krishna et. al. concluded that due to the large V/F (>100 l/kg), distribution rather than elimination processes determine the blood concentration profile of chloroquine in patients with acute malaria (57). Other potential reasons for high variability in elimination half-life for chloroquine could be differences in PK follow-up duration, improvement in sensitivity of analytical methods over time, and/or different methodologies of calculations. Regarding chloroquine apparent clearance (CL/F), the reported mean estimates ranged from 16 to 40 l/hr (Table 1). Estimated urinary recovery of chloroquine is reported to be 46% and 55% (20, 22). Augustijns et. al. evaluated whole blood PK of chloroquine enantiomers in humans after a single oral dose of the separate enantiomers and reported

that the total body clearance was lower for the (R)-enantiomer  $(8\pm2 \text{ l/h})$  than for the (S)-enantiomer  $(14\pm4 \text{ l/h})$  (10). The study also reported that terminal half-life and mean residence time (MRT) were longer for (R)-chloroquine (12 days and 16 days, respectively) than for (S)-chloroquine (10 days and 11 days, respectively).

#### Hydroxychloroquine

Similar to chloroquine, information on the metabolism and excretion of hydroxychloroquine is scarce in the literature including information on specific metabolism pathways. Fan et. al. reported the mean estimated elimination half-life for hydroxychloroquine to be 11 to 12 days when estimated with hydroxychloroquine plasma concentration over 62 days and 1 to 2 days when estimated with hydroxychloroquine plasma concentration over 72 hours (29). These findings suggest the impact of PK follow-up duration on the reported high variability in elimination half-life for hydroxychloroquine across the studies.

The reported mean amount of hydroxychloroquine excreted unchanged in the urine ranges between 23-27% (15, 16, 69). McLachlan et. al. evaluated renal clearance of the hydroxychloroquine enantiomers and reported that (S)-hydroxychloroquine had a mean ( $\pm$  SD) renal clearance from blood of 41  $\pm$  11 ml min, approximately twice that of (R)-hydroxychloroquine (30). Ducharme et. al. notes similar findings with the reported total urinary excretion of (S)-hydroxychloroquine to be higher than that of (R)-hydroxychloroquine (17). In the same study, the estimated elimination half-life of (S)-hydroxychloroquine (19  $\pm$  5 days) was significantly shorter than that of (R)-hydroxychloroquine (22  $\pm$  6 days), partly due to its faster urinary excretion and hepatic metabolism. Its renal clearance was twice that of (R)-hydroxychloroquine (4.61  $\pm$  4.01 vs 1.79  $\pm$  1.30 l/h).

# POPULATION PHARMACOKINETIC MODELS

With respect to the use of population PK (POP-PK) analysis approach, three studies for chloroquine (60-62) and four studies for hydroxychloroquine were identified during the literature search (18, 63-65). These studies are summarized in Table 5 and key information is summarized below. It is noteworthy that the intensity of the PK sampling, sensitivity of bio-analytical method used and/or sampling duration may affect the PK model selection process. In addition, the lack of variability in

intrinsic and extrinsic factors within the patient/subject population, whose PK data are being utilized for POP-PK analysis, may affect the ability of identifying relationship between covariates and PK. Obua et. al. reports that two-compartmental PK model best described the chloroquine pharmacokinetics based on a POP-PK analysis that utilized sparse PK data obtained from finger prick sampling in pediatric malaria patients (59). The same study reports that no correlation was identified between body weight or age with the PK model parameters. Höglund et al. reports that for adult malaria patients, the disposition of chloroquine was also adequately described by the two-compartment model without the use of any covariates on the model parameters (61). POP-PK approach was also used to assess differences in the PK of chloroquine in pregnant and non-pregnant women, and findings are discussed in Special Populations – Pregnancy section of this manuscript (60). Within the POP-PK analyses literature on chloroquine discussed here, the reported BSV (CV%) for CL/F estimate was ~30% (Relative standard error [RSE]: 24-35%). For the V/F estimates for central compartment, reported BSV (CV%) ranged from 40-57% (RSE: 22-67%).For hydroxychloroquine, Carmichael et. al. reports that one compartment PK model adequately described its pharmacokinetics based on pooled PK data from several pharmacokinetic studies in patients with Rheumatoid Arthritis (62). The same study notes that the limited amount of data per individual resulted in a one compartment PK model selection instead of a multi compartment model as reported in literature. Lim et. al. reports that in healthy adults and adult malaria patients, hydroxychloroquine pharmacokinetics were best described by a two-compartment PK model with first-order absorption with absorption lag time without any covariates (18). However, Morita et. al. reports that for patients with cutaneous lupus erythematosus (CLE) or systemic lupus erythematosus (SLE), a onecompartment model with first-order absorption and absorption lag time adequately described hydroxychloroquine PK and body weight was identified as a significant (P < 0.001) covariate (63). Balevic et. al. reports the use of POP-PK analysis to assess the effect of pregnancy on PK of hydroxychloroquine in patients with Rheumatic Diseases and notes lack of significant changes hydroxychloroquine exposure due to pregnancy in small cohort of patients (64). The study reports that a one-compartment PK model best described hydroxychloroquine PK and body weight was found to be a significant covariate on V/F. Within the POP-PK analyses literature on hydroxychloroquine

discussed above, the reported BSV (CV%) in CL/F estimates ranged from 16-44% (RSE: 4-41%). For V/F estimates for central compartment,BSV (CV%) ranged from 16-23% (RSE: ~58%).

# PHYSIOLOGIC-BASED PHARMACOKINETIC MODELING AND TISSUE PARTITIONING

SARS-CoV-2 is a virus with a predilection for the respiratory system resulting in COVID-19 in susceptible individuals, leading to ARDS. The target protein and cofactors for viral attachment and uptake are expressed in the lung and bronchial branches (66). Several researchers have proposed that lung tissue and/or lung-substructure concentrations of drugs may be used for dose selection in clinical trials for COVID-19 and ultimately understanding exposure-response against the virus or against the inflammatory response to the virus.

Physiologic-based pharmacokinetic (PBPK) models leverage characteristics of individual organs and tissues, physicochemical properties, absorption, distribution, metabolism and excretion, and tissue partition coefficients characteristics to allow prediction of systemic and tissue concentrations of parent and/or metabolite over time under various conditions and for various populations (67). This approach to model-informed drug development can provide greater insights into local concentration-effect relationships than can empiric-based PK modeling.

Several PBPK models for hydroxychloroquine have been developed. One was developed to better understand drug concentrations in the lysosome under varying pH conditions as well as various tissues (68). The other was developed to project optimized dosing of hydroxychloroquine for the treatment of SARS-CoV-2 by simulating plasma, blood and lung concentrations and comparing simulated concentrations with inhibitory concentrations determined in in-vitro SARS-CoV-2 growth assays (as described in subsequent section on the In Vitro data for SARS-COV-2) (2).

The same investigators have also modified a compound file of chloroquine to model systemic and pulmonary concentrations of chloroquine. The models were verified with observed PK data compared to concentrations predicted in silico in blood and plasma. The tissue partition coefficients were extrapolated from published tissue concentrations in rats.

The PBPK models and simulations for hydroxychloroquine and chloroquine reiterated prior knowledge that blood concentrations for both exceed that of plasma. More importantly, the full PBPK models indicated that hydroxychloroquine and chloroquine both have a predilection for lung. The model-predicted lung concentrations exceed plasma concentrations at steady state by more than 400fold. In addition, if anti-inflammatory effects and prevention of cytokine storm are instrumental in treating disease, then drug concentrations in the lung may provide beneficial effect. In addition, PBPK modeling and simulation have been utilized to inform dosing decisions for COVID-19 clinical trials. Ultimately, as efficacy and safety data from multiple COVID-19 prevention and treatment trials accrue, a more precise estimation across studies will help inform on effective doses and exposures over time.

# **TOXICITIES (ADVERSE EFFECTS)**

In both chloroquine and hydroxychloroquine use, gastrointestinal discomfort is the most commonly described side effect manifesting primarily as nausea, vomiting, diarrhea and stomach pain. These toxicities are more common at higher doses (69). Hypoglycemia may occur and can be enhanced in patients receiving concurrent hypoglycemic drugs. Rash and itching are well described and due to the long half-lives, they may last for prolonged periods after discontinuation. Hemolytic anemia is a serious adverse effect in individuals with glucose-6-phosphate dehydrogenase (G6PD) deficiency who receive chloroquine or hydroxychloroquine (5, 6). Retinal toxicity is a well-described adverse effect for hydroxychloroquine, however it is primarily a concern with chronic and cumulative doses of >1000 grams (70).

# Effects of Chloroquine and Hydroxychloroquine on QTc Prolongation

# Chloroquine

The effects of chloroquine alone as well as chloroquine and azithromycin on QTc prolongation were investigated in a previously unpublished, single center open-label, placebo-controlled, randomized, multiple-dose healthy volunteer study (N=119 total; 24 per group except for the chloroquine+1500mg Az group where 23 subjects were enrolled. Three subjects withdrew from the chloroquine+1500 mg Az group during the treatment phase of the study due to adverse events (1 due to diarrhea, 1 due to loss of appetite, and 1 due to nausea, diarrhea, and vomiting) and thus were not included in the

pharmacokinetic or QTc analyses). A parallel group design was utilized given the long half-lives of both chloroquine and azithromycin. Subjects were assigned to one of the following five treatment groups:

Placebo QD x 3 days

- 1000 mg chloroquine phosphate (600 mg base) QD x 3 days
- 1000 mg chloroquine phosphate plus 500 mg azithromycin QD x 3 days
- 1000 mg chloroquine phosphate plus 1000 mg azithromycin QD x 3 days
- 1000 mg chloroquine phosphate plus 1500 mg azithromycin QD x 3 days

Triplicate 12-lead ECGs were serially obtained on Day -1 (baseline), Day 1, Day 2 and Day 3 including at 0, 1, 2, 3, 4, 5, 6, 8, 10 and 12 hours on Day 3. Serial plasma samples were obtained for chloroquine and azithromycin quantitation on Days 1-3 including at times immediately following ECG assessments on Day 3. QT interval data were corrected for heart rate using Fridericia's method (QTcF). In all analyses, the means of the ECG parameters within a triplicate were used as the observations for a subject at a nominal time point. The primary endpoint was change of QTcF from baseline at each nominal time point on Day 3. Comparisons were made between treatments versus placebo and the chloroquine alone group using an ANOVA model. For each comparison the modelbased mean and 90% confidence interval were reported. No adjustments were made for multiplicity. The mean age, weight, and BMI were 35.5 years, 83.3 kg, and 26.6, respectively. In comparison to QTcF values on placebo, the chloroquine increased mean time matched QTcF values ranging from 18.4 to 35 msec on Day 3 in the chloroquine alone group (Table 6). The maximum time-matched difference in OTcF occurred at 10 hours post dose on Day 3. Maximum observed plasma chloroquine concentrations on Day 3 were similar for all groups [mean 0.335 (CV 29%) µg/mL] and generally occurred between 6- and 7-hours post dose. These results are consistent with the large magnitude of effect of chloroquine on QTcF in previous studies (49, 71). It should be noted that the high chloroquine dose (600 mg twice daily for 10 days or total dose 12g) of a recent phase IIb trial of patients with SARS in Manaus, Brazilian Amazon was stopped because of

safety. There was a trend for higher lethality compared with the lower dose and one quarter of the high dose patients developed QTc>500 msec (72). All patients in this study also received 5 days of azithromycin, also a QTc prolonging drug.

#### Hydroxychloroquine

The effect of hydroxychloroquine on QTc is less well studied than the effect of chloroquine. Case reports have noted prolonged QTc values with hydroxychloroquine toxicity (73-78). In a study of 85 patients with connective tissue disorders who were receiving hydroxychloroquine, the investigators concluded that QT intervals did not appear to be different from what would be considered normal in these patients (79).

Recently there are emerging QT data from 84 patients with SARS-CoV-2 infection treated with a combination of hydroxychloroquine and azithromycin. QTc values were determined for subjects at baseline (before administering hydroxychloroquine and azithromycin) and while on drug, but no reference was given regarding doses administered, the timing of QTc assessment with respect to dose nor the formula used to correct QT for heart rate. QTc values were prolonged maximally (28 msec) from baseline between Days 3 and 4. In 30% of patients, QTc increased by greater than 40 ms. In 11% of patients, QTc increased to >500 ms, representing high risk group for arrhythmia. Development of acute renal failure but not baseline QTc was a strong predictor of greater QTc prolongation (80).

#### SPECIAL POPULATIONS

#### **Special populations - Renal/Hepatic Impairment**

#### Chloroquine

No dose adjustment is suggested by the manufacturer for patients with renal impairment. No dose adjustment is suggested by the manufacturer for those with hepatic impairment; however, caution should be used in patients with hepatic disease and/or alcoholism, as the drug concentrates largely in the liver. Table S1 summarizes the language provided in both the chloroquine and hydroxychloroquine labels regarding both renal and hepatic impairment.

#### Hydroxychloroquine

No dose adjustment is suggested by the manufacturer for patients with renal impairment. Jallouli, et. al. evaluated 3 patients with systemic lupus erythematosus receiving long-term dialysis for hydroxychloroquine blood concentrations pre- and post-dialysis. Patients received either 200 mg/day (N=2) or 400 mg/day(N=1) of hydroxychloroquine, and blood concentrations did not change significantly pre- or post-dialysis (81). Hydroxychloroquine was not detected in the dialysis bath of all three patients (<50 ng/mL). This suggests hydroxychloroquine is not dialyzable. No dose adjustment is suggested by the manufacturer for those with hepatic impairment; however, caution should be used in patients with hepatic disease and/or alcoholism, as the drug concentrates largely in the liver. Table S1 summarizes the language provided in both the chloroquine and hydroxychloroquine labels regarding both renal and hepatic impairment.

#### **Special Populations - Age**

Geriatric Population: Typical adult dosing is recommended for patients greater than 65 years of age. Hydroxychloroquine and chloroquine clinical trials did not include a sufficient number of geriatric subjects to determine whether they respond differently than younger subjects. However, given both drugs are significantly renally cleared and elderly patients are more likely to have decreased renal function, consideration should be given for dose reduction in this population.

Pediatric Population: Both hydroxychloroquine and chloroquine are preferably dosed based on actual body weight in the pediatric population. The American Academy of Ophthalmology recommends keeping daily doses  $\leq 6.5$  mg/kg hydroxychloroquine-sulfate ( $\leq 5.0$  mg/kg base) and  $\leq 3.8$  mg/kg chloroquine-phosphate ( $\leq 2.3$  mg/kg base) for all patients to reduce the risk of retinopathy and permanent vision loss, though this risk is linked to duration of use over years (82). Approved pediatric doses are above this threshold for treatment of acute uncomplicated malaria.

#### Chloroquine

In infants, children and adolescents, the highest approved dose of chloroquine is for treatment of acute malarial attack. The highest recommended chloroquine dose for the treatment of acute, uncomplicated malarial attack is 10 mg base/kg PO followed by 5 mg base/kg at 6, 24, and 48 hours after initial dose for a total of 4 doses (83). Doses for chemoprophylaxis are suggested to be lower

(84). Several fatalities have been reported due to accidental ingestion at even lower doses (0.75-1 g chloroquine-phosphate in one 3-year-old child) (5). The injection label notes that in no instance should a single intramuscular or subcutaneous dose exceed 6.25 mg (5 mg base) per kg of body weight, since children are especially sensitive to the effects of the 4-aminoquinolines. Severe reactions and sudden death have been reported following parenteral administration in children (5, 85).

#### Hydroxychloroquine

In infants, children and adolescents, 10 mg base/kg followed by 5 mg base/kg hydroxychloroquine at 6, 24 and 48 hours after initial dose is the highest recommended dose for the treatment of acute uncomplicated malarial attack (83). Doses for chemoprophylaxis, juvenile rheumatoid arthritis and systemic lupus erythematosus are suggested to be lower (84).

#### **Special Populations - Pregnancy**

#### Chloroquine

Based on WHO Guidelines for the treatment of malaria, chloroquine is considered safe in the first trimester of pregnancy and has been studied in combination with azithromycin for intermittent preventive treatment in pregnancy (IPTp) as a fixed dose combination chloroquine and metabolites (desethylchloroquine) can be detected in the cord and urine of newborn infants (33, 86-89). Karunajeewa, et. Al. performed a population PK analyses of chloroquine and its active metabolite, monodesethylcholorquine in pregnancy. Pregnancy was found to have a significant effect on chloroquine and desethylchloroquine disposition after conventional doses of chloroquine. Exposures of both analytes were found to be significantly lower in pregnant patients (AUCs for chloroquine and desethylchloroquine saw reductions of 25% and 45%, respectively) in comparison to nonpregnant patients (61).

Ogunbona et. al. investigated the excretion of chloroquine and desethylchloroquine in breast milk in lactating mothers following a single 600 mg (chloroquine base) oral dose. The maximum daily dose an infant could receive from breastfeeding was  $\sim 0.7\%$  of maternal starting dose (90).

#### Hydroxychloroquine

American Academy of Pediatrics (AAP) considers use of hydroxychloroquine usually compatible with breastfeeding and hydroxychloroquine is considered appropriate treatment for SLE during pregnancy by the American College of Rheumatology (91, 92). Unless pregnancy is planned, fetal exposure cannot be avoided by discontinuing hydroxychloroquine at the time pregnancy is discovered, as hydroxychloroquine is stored in the liver with an extended half-life. Hydroxychloroquine can be detected in cord-blood in concentrations similar to those in post-partum maternal serum (93) and crosses the placenta. One study saw no differences when comparing patients with systemic lupus erythematosus with no hydroxychloroquine exposure (N=163), continuous hydroxychloroquine use (N=56) and those discontinuing hydroxychloroquine during the first trimester when assessing (N=38) for miscarriages, stillbirths, pregnancy losses and congenital abnormalities (94). Another study by Costedoat-Chalumeau et. al. demonstrated no difference in growth rate and no evidence of visual, hearing or developmental abnormalities in the mean 26-month follow-up of children between hydroxychloroquine and control group (95). Hydroxychloroquine has been shown to be transferred to human breast milk. Breastfed infants are exposed to ~2% of maternal dose on a body weight basis (37).

# **Special Populations - Critically ill**

Considering the adverse event profile and elimination pathway of both hydroxychloroquine and chloroquine, strong consideration to alternative therapies should be made in patients exhibiting these comorbidities. Arentz et al. recently published a case series report discussing characteristics and outcomes of 21 critically ill patients with COVID-19 in Washington State. Of the 21 patients with PCR confirmed SARS-CoV-2 infection by nasopharyngeal sample, 18 were identified to have comorbidities. Chronic kidney disease (N=10) and congestive heart failure (N=9) were the most common comorbidities (96).

# **PROPOSED MOA FOR COVID-19 DISEASE**

# **Direct Antiviral**

The presumed mechanism of antiviral activity of chloroquine and hydroxychloroquine is increasing the pH of the endosome that is required for viral/cellular fusion. Using flow cytometry and sorting of SARS--CoV spike glycoprotein and ACE2 on Vero E6 cells, CDC investigators also found that chloroquine impairs the glycosylation of ACE2, thereby impairing the binding of the virus to its cellular receptor (97). To date, there are no data on stereospecific antiviral effects of chloroquine or hydroxychloroquine.

#### Immunomodulatory

In addition to direct antiviral activity, chloroquine and hydroxychloroquine have known immunomodulatory and anti-inflammatory effects. The anti-inflammatory effects of hydroxychloroquine were first discovered through the serendipitous observation that WWII soldiers receiving chloroquine for malaria prophylaxis noted improvements in skin rashes and arthritis, ultimately leading to the development of this drug as a treatment for systemic lupus erythematosus and other immunologic conditions such as rheumatoid arthritis (98, 99). Immunomodulatory effects of chloroquine as well as hydroxychloroquine include inhibition of antigen presentation to dendritic cells, reduced cytokine production in macrophages and reduced signaling of both B and T cells. Therefore, in addition to direct antiviral effects, chloroquine and hydroxychloroquine may play a role in reducing the cytokine storm associated with COVID-19 progression to ARDS. Although these drugs are generally considered more immunomodulatory than immunosuppressive, it is worth pointing out that in early infection, the role of the innate immune system is a critical factor in preventing COVID-19 progression to serious disease. As our understanding of COVID-19 pathogenesis evolves, whether these immunomodulatory effects are beneficial or detrimental will need to be elucidated.

# **IN VITRO DATA FOR SARS-COV-2**

#### Chloroquine

Antiviral activity of chloroquine and hydroxychloroquine against COVID-19 has been mostly investigated in in vitro studies. Following the outbreak of the 2003 SARS virus, several compounds were screened for in vitro activity against SARS-CoV. In vitro hydroxychloroquine and chloroquine drug sensitivity data for SARS-CoV-2 are reported as percent inhibition and are most often obtained from 24- and 48-hour experiments in Vero or Vero E6 cells derived from African green monkey kidney epithelium (2, 100-102). CDC investigators reported that, when added post-infection,

chloroquine concentrations as little as 0.1  $\mu$ M (32 ng/mL) reduced viral spread by 50% while chloroquine concentrations of 100  $\mu$ M (32,000 ng/mL) reduced viral spread by up to 94% (97). The half-maximal inhibitory effect was estimated to occur at 4.4  $\mu$ M +/1.0  $\mu$ M (1408 +/320 ng/mL). Pretreatment with chloroquine was found to have an even more potent viral inhibition with 0.1, 1 and 10  $\mu$ M (32,320, and 3200 ng/mL), reducing infectivity by 28, 53 and 100% respectively, which may suggest lower drug exposures required for prophylaxis than treatment. Following the COVID-19 outbreak, Wang et al tested the in vitro inhibitory activity and cytotoxicity

of a number of potential therapeutics against SARS-CoV-2, including chloroquine in Vero E6 cells (100). Of the seven drugs tested, chloroquine (along with remdesivir) had one of the highest selectivity indexes with an EC<sub>50</sub> of 1.13  $\mu$ M (361 ng/mL) and CC<sub>50</sub> >100  $\mu$ M (>32,000 ng/mL). (Table 7)

#### Hydroxychloroquine

Similar to chloroquine, hydroxychloroquine has also been tested in a number of in vitro experiments for activity against SARS-CoV-2. A wide range of  $EC_{50}$  values have been reported under various conditions (Table 7), with some comparative studies finding greater potency with hydroxychloroquine (2) and some less potent (101). Below is a summary of the various experiments, including experimental characteristics and reported  $EC_{50}$  values is shown in Table 7.

#### **CLINICAL STUDIES FOR COVID-19**

For clinical use in COVID-19, the CDC provides anecdotal dosing suggestions, but explicitly states that optimal dosing and duration of hydroxychloroquine for COVID-19 are unknown (103). Translational pharmacokinetic/pharmacodynamic modeling is one approach to propose optimized hydroxychloroquine dosing regimens which ensure the highest likelihood of success as COVID-19 treatment. With lack of known exposure correlates for efficacy to date, several groups have attempted to integrate available pharmacological data and mechanistic knowledge related to COVID-19, including in vitro data for SARS-CoV-2, historical data on population pharmacokinetics, safety data of hydroxychloroquine from large patient cohorts, and newly emerging clinical PK/PD data from patients with COVID-19 (2, 104-106). Early characterization of the clinical exposure-response

relationship between hydroxychloroquine and SARS-CoV-2 viral load, suggests that the hydroxychloroquine doses needed to cause more rapid viral clearance compared to standard of care could be much higher than those currently being studied for COVID-19 patients in any setting and these doses may lead to substantial risk of cardiac toxicity (104-106). As these investigations have pointed out, when making in vitro-in vivo extrapolations to optimize dosing, there are many considerations to be made including 1) the compartment to be targeted (e.g. plasma, intracellular vs extracellular lung tissue, lung fluid, other organs), 2) free vs total concentrations, 3) plasma vs. serum vs whole blood, 4)  $EC_{50}$  vs  $EC_{90}$  or magnitude above these targets.

Initial support for the use of chloroquine and hydroxychloroquine came from a published report which states that results from 100 patients across multiple institutions found that for the treatment of COVID-19, chloroquine was superior to standard of care in preventing exacerbation of pneumonia, reducing days to conversion rate and shortening time to clinical recovery (1). To date, this study has not been published and data regarding magnitude of benefit, specific dosing or timing of treatment initiation, observed toxicities, and treatments received by the control group are not available. Other data with chloroquine, as mentioned above, is from one 81-person trial in Brazile compared 600 mg chloroquine base twice daily for 10 days (12 g total) to 450 mg x2 on day 1, then 450 mg daily for 4 additional days (2.7 g total). This study did not have a placebo arm and was halted prior to reaching enrollment goals because higher mortality was observed in the high dose chloroquine arm (72). One additional small study (n=22) showed no difference between chloroquine and lopinavir/ritonavir for viral clearance (107).

Clinical studies of hydroxychloroquine are heterogeneous in terms of hydroxychloroquine regimen, outcomes measurements (clinical, SARS-CoV-2 viral load), and severity of illness (108, 109). Only one study non-randomized study has reported a significant reduction in time to viral clearance among patients with mild disease who received hydroxychloroquine or hydroxychloroquine with azithromycin, and this small study had significant limitations including a control arm with significantly higher viral loads compared to those who received treatment (110). The largest well characterized observational cohorts of hospitalized patients, hydroxychloroquine with or without azithromycin were not associated with a mortality benefit (111-112).

Only 2 randomized clinical trials have been published in hospitalized COVID-19 patients, one of which found that high-dose hydroxychloroquine (1200 mg/day for three days followed by 800 mg/day for 2-3 weeks) given to mild or moderately ill patients did not improve time to negative PCR test nor time to alleviation of symptoms (113). A small randomized study in China reported no apparent clinical benefit of 400 mg daily for 5 days compared to placebo in COVID-19 patients, the majority of whom had mild disease (114). Recently, the RECOVERY trial (NCT04381936) stopped enrollment into the hydroxychloroquine arm of their trial in hospitalized COVID-19 patients, reporting no difference in mortality, hospital stay, or other outcomes, between 1542 subjects on hydroxychloroquine and 3132 subjects on standard care. Subsequently, the WHO-sponsored SOLIDARITY trial and an NIH-sponsored trial have suspended enrollment of their hydroxychloroquine arms, reporting no benefit observed although the data have not yet been released. Hydroxychloroquine has also been proposed for use in prophylactic settings. The first randomized clinical trial published in an outpatient setting found that high dose hydroxychloroquine (800 mg loading dose followed by 600 mg/day for 4 days) given within 4 days of a high-risk exposure did not reduce likelihood of acquiring COVID-19 (115). Studies in high risk health care workers evaluating a role for pre-exposure prophylaxis are currently underway.

#### DISCUSSION

In summary, to date, there is a lack of evidence from controlled, randomized clinical trials powered for efficacy endpoints (the gold standard for evidence-based medicine) for use of chloroquine or hydroxychloroquine to treat COVID-19. Increasingly, evidence points to a lack of benefit in hospitalized and severely ill patients; whether a role exists in early treatment or prevention remains to be determined. Although chloroquine and hydroxychloroquine are very similar in their pharmacologic properties, there are subtle differences of which clinicians and scientists should be aware. The Infectious Disease Society of America released guidelines for the treatment of COVID-19, recommending hydroxychloroquine only in the context of a clinical trial (116). While the NIH guidelines state there are insufficient clinical data for or against chloroquine or hydroxychloroquine, they do specifically recommend against the use of high dose hydroxychloroquine (defined as 600mg twice daily for 10 days) or hydroxychloroquine in combination with azithromycin.

#### REFERENCES

(4)

(7)

- Gao, J., Tian, Z. & Yang, X. Breakthrough: Chloroquine phosphate has shown apparent efficacy in treatment of COVID-19 associated pneumonia in clinical studies. *Biosci Trends* 14, 72-3 (2020).
- Yao, X. *et al.* In Vitro Antiviral Activity and Projection of Optimized Dosing Design of Hydroxychloroquine for the Treatment of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). *Clin Infect Dis*, (2020).
- (3) Gautret, P. *et al.* Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. *International journal of antimicrobial agents*, 105949 (2020).
  - Ohrt, C., Willingmyre, G.D., Lee, P., Knirsch, C. & Milhous, W. Assessment of azithromycin in combination with other antimalarial drugs against Plasmodium falciparum in vitro. *Antimicrob Agents Chemother* **46**, 2518-24 (2002).
- (5) Kent, G. & al-Abadie, M. Factors affecting responses on Dermatology Life Quality Index items among vitiligo sufferers. *Clin Exp Dermatol* **21**, 330-3 (1996).
- (6) Eleftheriadou, V., Thomas, K., Ravenscroft, J., Whitton, M., Batchelor, J. & Williams, H.
   Feasibility, double-blind, randomised, placebo-controlled, multi-centre trial of hand-held NB-UVB phototherapy for the treatment of vitiligo at home (HI-Light trial: Home Intervention of Light therapy). *Trials* **15**, 51 (2014).
  - Ette, E.I., Essien, E.E., Thomas, W.O. & Brown-Awala, E.A. Pharmacokinetics of chloroquine and some of its metabolites in healthy volunteers: a single dose study. *J Clin Pharmacol* **29**, 457-62 (1989).
- (8) Ette, E.I., Brown-Awala, E.A. & Essien, E.E. Chloroquine elimination in humans: effect of low-dose cimetidine. *J Clin Pharmacol* **27**, 813-6 (1987).
- (9) Essien, E., Ette, E., Thomas, W. & Brown-Awala, E. Chloroquine disposition in hypersensitive and non-hypersensitive subjects and its significance in chloroquine-induced pruritus. *Eur J Drug Metab Pharmacokinet* **14**, 71-7 (1989).
- Augustijns, P. & Verbeke, N. Stereoselective pharmacokinetic properties of chloroquine and de-ethyl-chloroquine in humans. *Clin Pharmacokinet* 24, 259-69 (1993).
- (11) De Vries, P., Oosterhuis, B. & Van Boxtel, C. Single-dose pharmacokinetics of chloroquine and its main metabolite in healthy volunteers. *Drug Investigation* 8, 143-9 (1994).

- (12) Nsimba, S. *et al.* Comparative in vitro and in vivo study of a sugar-coated chloroquine preparation marketed in Tanzania versus an ordinary brand. *J Clin Pharm Ther* **26**, 43-8 (2001).
- (13) Onyeji, C.O. & Ogunbona, F.A. Pharmacokinetic aspects of chloroquine-induced pruritus:
   influence of dose and evidence for varied extent of metabolism of the drug. *Eur J Pharm Sci* 13, 195-201 (2001).
- Pukrittayakamee, S. *et al.* Pharmacokinetic interactions between primaquine and chloroquine.
   Antimicrob Agents Chemother 58, 3354-9 (2014).
- (15) Tett, S., Cutler, D., Day, R. & Brown, K. A dose-ranging study of the pharmacokinetics of hydroxy-chloroquine following intravenous administration to healthy volunteers. *Br J Clin Pharmacol* **26**, 303-13 (1988).
- Tett, S., Cutler, D., Day, R. & Brown, K. Bioavailability of hydroxychloroquine tablets in healthy volunteers. *Br J Clin Pharmacol* 27, 771-9 (1989).
- (17) Ducharme, J., Fieger, H., Ducharme, M., Khalil, S. & Wainer, I. Enantioselective disposition of hydroxychloroquine after a single oral dose of the racemate to healthy subjects [see comments]. *Br J Clin Pharmacol* **40**, 127-33 (1995).
- (18) Lim, H.-S. *et al.* Pharmacokinetics of hydroxychloroquine and its clinical implications in chemoprophylaxis against malaria caused by Plasmodium vivax. *Antimicrob Agents Chemother* **53**, 1468-75 (2009).
- (19) Rainsford, K., Parke, A.L., Clifford-Rashotte, M. & Kean, W. Therapy and pharmacological properties of hydroxychloroquine and chloroquine in treatment of systemic lupus erythematosus, rheumatoid arthritis and related diseases. *Inflammopharamacology* 23, 231-69 (2015).
- (20) Frisk-Holmberg, M., Bergqvist, Y. & Domeij-Nyberg, B. Steady state disposition of chloroquine in patients with rheumatoid disease. *Eur J Clin Pharmacol* **24**, 837-9 (1983).
- (21) Frisk-Holmberg, M., Bergqvist, Y., Termond, E. & Domeij-Nyberg, B. The single dose kinetics of chloroquine and its major metabolite desethylchloroquine in healthy subjects. *Eur J Clin Pharmacol* **26**, 521-30 (1984).
- Gustafsson, L. *et al.* Disposition of chloroquine in man after single intravenous and oral doses. *Br J Clin Pharmacol* **15**, 471-9 (1983).
- (23) Edwards, G., Looareesuwan, S., Davies, A., Wattanagoon, Y., Phillips, R. & Warrell, D.Pharmacokinetics of chloroquine in Thais: plasma and red-cell concentrations following an

intravenous infusion to healthy subjects and patients with Plasmodium vivax malaria. *Br J Clin Pharmacol* **25**, 477-85 (1988).

- Wetsteyn, J., De Vries, P., Oosterhuis, B. & Van Boxtel, C. The pharmacokinetics of three multiple dose regimens of chloroquine: implications for malaria chemoprophylaxis. *Br J Clin Pharm* **39**, 696-9 (1995).
- McLachlan, A., Tett, S., Cutler, D. & Day, R. Bioavailability of hydroxychloroquine tablets in patients with rheumatoid arthritis. *Br J Rheumatol* **33**, 235-9 (1994).
- (26) McLachlan, A., Tett, S., Cutler, D. & Day, R. Absorption and in vivo dissolution of hydroxycholoroquine in fed subjects assessed using deconvolution techniques. *Br J Clin Pharmacol* **36**, 405-11 (1993).
- TETT, S., DAY, R. & CUTLER, D. Hydroxychloroquine relative bioavailability: within subject reproducibility. *Br J Clin Pharmacol* **41**, 244-6 (1996).
- (28) Carmichael, S.J., Beal, J., Day, R.O. & Tett, S.E. Combination therapy with methotrexate and hydroxychloroquine for rheumatoid arthritis increases exposure to methotrexate. *Rheumatol* 29, 2077-83 (2002).
- (29) Fan, H.-w., Ma, Z.-x., Chen, J., Yang, X.-y., Cheng, J.-I. & Li, Y.-b. Pharmacokinetics and bioequivalence study of hydroxychloroquine sulfate tablets in Chinese healthy volunteers by LC–MS/MS. *Rheumatol Ther* **2**, 183-95 (2015).
- (30) McLachlan, A., Tett, S., Cutler, D. & Day, R. Disposition of the enantiomers of hydroxychloroquine in patients with rheumatoid arthritis following multiple doses of the racemate. *Br J Clin Pharmacol* **36**, 78-81 (1993).
- (31) Walker, O., Birkett, D., Alvan, G., Gustafsson, L. & Sjoqvist, F. Characterization of chloroquine plasma protein binding in man. *Br J Clin Pharmacol* **15**, 375-7 (1983).
- (32) Ofori-Adjei, D., Ericsson, O., Lindstrom, B. & Sjoqvist, F. Protein binding of chloroquine enantiomers and desethylchloroquine. *Br J Clin Pharmacol* **22**, 356-8 (1986).
- (33) Akintonwa, A., Gbajumo, S. & Mabadeje, A. Placental and milk transfer of chloroquine in humans. *Ther Drug Mon* **10**, 147-9 (1988).
- (34) Ette, E.I., Essien, E.E., Ogonor, J.I. & Brown-Awala, E.A. Chloroquine in human milk. J Clin Pharmacol 27, 499-502 (1987).
- (35) McLachlan, A., Cutler, D. & Tett, S. Plasma protein binding of the enantiomers of hydroxychloroquine and metabolites. *Eur J Clin Pharmacol* **44**, 481-4 (1993).

- (36) Furst, D.E. Pharmacokinetics of hydroxychloroquine and chloroquine during treatment of rheumatic diseases. *Lupus* 5, 11-5 (1996).
- (37) Nation, R., Hackett, L., Dusci, L. & Ilett, K. Excretion of hydroxychloroquine in human milk. *Br J Clin Pharmacol* **17**, 368 (1984).
- (38) Østensen, M., Brown, N., Chiang, P. & Aarbakke. Hydroxychloroquine in human breast milk. *Eur J Clin Pharmacol* 28, 357- (1985).
- Peng, W., Liu, R., Zhang, L., Fu, Q., Mei, D. & Du, X. Breast milk concentration of hydroxychloroquine in Chinese lactating women with connective tissue diseases. *Eur J Clin Pharmacol* **75**, 1547-53 (2019).
- (40) Ducharme, J. & Farinotti, R. Clinical pharmacokinetics and metabolism of chloroquine. *Clin Pharmacokinetics* **31**, 257-74 (1996).
- Projean, D. *et al.* In vitro metabolism of chloroquine: identification of CYP2C8, CYP3A4, and CYP2D6 as the main isoforms catalyzing N-desethylchloroquine formation. *Drug Metab Dispos* 31, 748-54 (2003).
- (42) Kiang, T.K., Wilby, K.J. & Ensom, M.H. Drug Interaction Potential of Antimalarial Drugs
   Based on Known Metabolic Properties of Antimalarials. In: *Clinical Pharmacokinetic and Pharmacodynamic Drug Interactions Associated with Antimalarials* 17-25 (Springer, 2015).
- (43) Kiang, T.K., Wilby, K.J. & Ensom, M.H. Pharmacokinetic Drug Interactions Affecting Antimalarials. In: *Clinical Pharmacokinetic and Pharmacodynamic Drug Interactions Associated with Antimalarials* 27-55 (Springer, 2015).
- (44) Kiang, T.K., Wilby, K.J. & Ensom, M.H. Effects of Antimalarials on the Pharmacokinetics of Co-Administered Antimalarials. In: *Clinical Pharmacokinetic and Pharmacodynamic Drug Interactions Associated with Antimalarials* 87-117 (Springer, 2015).
- (45) Kiang, T.K., Wilby, K.J. & Ensom, M.H. Effects of Antimalarials on the Pharmacokinetics of Co-Administered Drugs. In: *Clinical Pharmacokinetic and Pharmacodynamic Drug Interactions Associated with Antimalarials* 57-85 (Springer, 2015).
- (46) Adjepon-Yamoah, K., Woolhouse, N. & Prescott, L. The effect of chloroquine on paracetamol disposition and kinetics. *Br J Clin Pharmacol* **21**, 322-4 (1986).
- (47) Back, D., Breckenridge, A., Grimmer, S., ML'E, O. & Purba, H. Pharmacokinetics of oral contraceptive steroids following the administration of the antimalarial drugs primaquine and chloroquine. *Contraception* **30**, 289-95 (1984).

- (48) Miller, A.K. *et al.* Pharmacokinetic interactions and safety evaluations of coadministered tafenoquine and chloroquine in healthy subjects. *Br J Clin Pharmacol* **76**, 858-67 (2013).
- (49) Cook, J.A., Randinitis, E.J., Bramson, C.R. & Wesche, D.L. Lack of a pharmacokinetic interaction between azithromycin and chloroquine. *Am J Trop Med Hyg* **74**, 407-12 (2006).
- Raina, R., Bano, G., Amla, V., Kapoor, V. & Gupta, K. The Effect of Aspirin Paracetamol and Analgin on Pharmacokinetics of Chloroquine. *Indian J Physiol Pharmacol* 37, 229- (1993).
- (51) Rengelshausen, J. *et al.* Pharmacokinetic interaction of chloroquine and methylene blue combination against malaria. *Eur J Clin Pharmacol* **60**, 709-15 (2004).
- (52) Mahmoud, B. Significant reduction in chloroquine bioavailablity following coadministration with the sudanese beverages aradaib, karkadi and lemon. *J Antimicrob Chemother* **33**, 1005-9 (1994).
- (53) Kalia, S. & Dutz, J.P. New concepts in antimalarial use and mode of action in dermatology.
   *Dermatol Ther* **20**, 160-74 (2007).
- (54) Masimirembwa, C.M., Gustafsson, L.L., Dahl, M.L., Abdi, Y.A. & Hasler, J.A. Lack of effect of chloroquine on the debrisoquine (CYP2D6 and S-mephenytoin (CYP2C19) hydroxylation phenotypes. *British journal of clinical pharmacology* **41**, 344-6 (1996).
- (55) Tfelt-Hansen, P., Ågesen, F.N., Pavbro, A. & Tfelt-Hansen, J. Pharmacokinetic variability of drugs used for prophylactic treatment of migraine. *CNS Drugs* **31**, 389-403 (2017).
- (56) Mehnert, J.M. *et al.* A phase I trial of MK-2206 and hydroxychloroquine in patients with advanced solid tumors. *Cancer Chemother Pharmacol* **84**, 899-907 (2019).
- (57) Krishna, S. & White, N.J. Pharmacokinetics of quinine, chloroquine and amodiaquine. *Clin Pharmacokinet* **30**, 263-99 (1996).
- (58) White, N. & Looareesuwan, S. Cerebral malaria. In: *Infections of the nervous system* 118-44 (Elsevier, 1987).
- (59) Miller, D.R., Khalil, S.K. & Nygard, G.A. Steady-state pharmacokinetics of hydroxychloroquine in rheumatoid arthritis patients. *DICP*, (1991).
- (60) Obua, C. *et al.* Population pharmacokinetics of chloroquine and sulfadoxine and treatment response in children with malaria: suggestions for an improved dose regimen. *br J Clin Pharm* **65**, 493-501 (2008).
- (61) Karunajeewa, H.A. *et al.* Pharmacokinetics of chloroquine and monodesethylchloroquine in pregnancy. *Antimicrob Agents Chemother* **54**, 1186-92 (2010).

- (62) Höglund, R., Moussavi, Y., Ruengweerayut, R., Cheomung, A., Äbelö, A. & Na-Bangchang,
   K. Population pharmacokinetics of a three-day chloroquine treatment in patients with
   Plasmodium vivax infection on the Thai-Myanmar border. *Malar J* 15, 129 (2016).
- (63) Carmichael, S.J., Charles, B. & Tett, S.E. Population pharmacokinetics of hydroxychloroquine in patients with rheumatoid arthritis. *Ther Drug Monit* **25**, 671-81 (2003).
- Morita, S., Takahashi, T., Yoshida, Y. & Yokota, N. Population pharmacokinetics of hydroxychloroquine in Japanese patients with cutaneous or systemic lupus erythematosus. *Ther Drug Monit* 38, 259-67 (2016).
- (65) Balevic, S.J., Green, T.P., Clowse, M.E., Eudy, A.M., Schanberg, L.E. & Cohen-Wolkowiez,
   M. Pharmacokinetics of Hydroxychloroquine in Pregnancies with Rheumatic Diseases. *Clin Pharmacokinet* 58, 525-33 (2019).
- (66) Lukassen, S. *et al.* SARS-CoV-2 receptor ACE2 and TMPRSS2 are primarily expressed in bronchial transient secretory cells. *EMBO J*, (2020).
- (67) Polak, S., Tylutki, Z., Holbrook, M. & Wiśniowska, B. Better prediction of the local concentration–effect relationship: the role of physiologically based pharmacokinetics and quantitative systems pharmacology and toxicology in the evolution of model-informed drug discovery and development. *Drug Discov Today*, (2019).
- (68) Collins, K.P., Jackson, K.M. & Gustafson, D.L. Hydroxychloroquine: a physiologically-based pharmacokinetic model in the context of cancer-related autophagy modulation. *J Pharmacol Exp Ther* **365**, 447-59 (2018).
  - 9) Munster, T. *et al.* Hydroxychloroquine concentration–response relationships in patients with rheumatoid arthritis. *Arthritis Rheum* **46**, 1460-9 (2002).
  - Ding, H.J., Denniston, A.K., Rao, V.K. & Gordon, C. Hydroxychloroquine-related retinal toxicity. *Rheumatology (Oxford)* 55, 957-67 (2016).
  - ) Vicente, J. *et al.* Assessment of Multi-Ion Channel Block in a Phase I Randomized Study Design: Results of the Ci PA Phase I ECG Biomarker Validation Study. *Clin Pharm Ther* **105**, 943-53 (2019).
  - Borba, M. *et al.* Chloroquine diphosphate in two different dosages as adjunctive therapy of hospitalized patients with severe respiratory syndrome in the context of coronavirus (SARS-CoV-2) infection: Preliminary safety results of a randomized, double-blinded, phase IIb clinical trial (CloroCovid-19 Study). *MedRXiv*, (2020).

(69) (70) (71) (72)

- (73) Newton-Cheh, C., Lin, A.E., Baggish, A.L. & Wang, H. Case 11-2011: A 47-Year-Old Man with Systemic Lupus Erythematosus and Heart Failure. *N Engl J Med* 364, 1450-60 (2011).
- (74) Radke, J.B., Kingery, J.M., Maakestad, J. & Krasowski, M.D. Diagnostic pitfalls and laboratory test interference after hydroxychloroquine intoxication: A case report. *Toxicol Rep* 6, 1040-6 (2019).
- (75) De Olano, J., Howland, M.A., Su, M.K., Hoffman, R.S. & Biary, R. Toxicokinetics of hydroxychloroquine following a massive overdose. *Am J Emerg Med* 37, 2264. e5-. e8 (2019).
- (76) Chen, C.-Y., Wang, F.-L. & Lin, C.-C. Chronic hydroxychloroquine use associated with QT prolongation and refractory ventricular arrhythmia. *Clin Toxicol (Phila)* **44**, 173-5 (2006).
- (77) Kandan, S.R. & Saha, M. Severe primary hypothyroidism presenting with torsades de pointes. *Case Reports* **2012**, bcr1220115306 (2012).
- (78) Gunja, N. *et al.* Survival after massive hydroxychloroquine overdose. *Anaesth Intensive Care* **37**, 130-3 (2009).
- (79) Costedoat-Chalumeau, N. *et al.* Heart conduction disorders related to antimalarials toxicity: an analysis of electrocardiograms in 85 patients treated with hydroxychloroquine for connective tissue diseases. *Rheumatology (Oxford)* **46**, 808-10 (2007).
- (80) Chorin, E. *et al.* The QT Interval in Patients with SARS-CoV-2 Infection Treated with Hydroxychloroquine/Azithromycin. *MedRXiv*, (2020).
- Jallouli, M. *et al.* Determinants of hydroxychloroquine blood concentration variations in systemic lupus erythematosus. *Arthritis Rheumatol* **67**, 2176-84 (2015).
- (82) Marmor, M.F., Kellner, U., Lai, T.Y., Melles, R.B. & Mieler, W.F. Recommendations on screening for chloroquine and hydroxychloroquine retinopathy (2016 revision). *Opthalmology* 123, 1386-94 (2016).
- (83) Schmid-Ott, G. *et al.* Stigmatization experience, coping and sense of coherence in vitiligo patients. *J Eur Acad Dermatol Venereol* **21**, 456-61 (2007).
- (84) Sehrawat, M., Arora, T.C., Chauhan, A., Kar, H.K., Poonia, A. & Jairath, V. Correlation of Vitamin D Levels with Pigmentation in Vitiligo Patients Treated with NBUVB Therapy. *ISRN dermatology* **2014**, 493213 (2014).
- (85) Saeki, H. *et al.* Guidelines for management of atopic dermatitis. *The Journal of dermatology* 36, 563-77 (2009).

- (86) Law, I. *et al.* Transfer of chloroquine and desethylchloroquine across the placenta and into milk in Melanesian mothers. *Br J Clin Pharm* **65**, 674-9 (2008).
- (87) Essien, E. & Afamefuna, G. Chloroquine and its metabolites in human cord blood, neonatal blood, and urine after maternal medication. *Clin Chem* 28, 1148-52 (1982).
- (88) ASTCT. ASTCT Response to Covid-19 < https://www.astct.org/connect/astct-response-tocovid-19> (2020). Accessed 22 Apr 2020.
- (89) WHO. *World Health Organization Guidelines for the treatment of malaria* (World Health Organization: 2015).
- (90) Ogunbona, F., Onyeji, C., Bolaji, O. & Torimiro, S. Excretion of chloroquine and desethylchloroquine in human milk. *Br J Clin Pharmacol* **23**, 473-6 (1987).
- (91) Transfer of drugs and other chemicals into human milk. *Pediatrics* **108**, 776-89 (2001).
- (92) Sammaritano, L.R. *et al.* 2020 American College of Rheumatology Guideline for the Management of Reproductive Health in Rheumatic and Musculoskeletal Diseases. *Arthritis Rheumatol* **72**, 461-88 (2020).
- (93) Costedoat-Chalumeau, N. *et al.* Evidence of transplacental passage of hydroxychloroquine in humans. *Arthritis Rheum* 46, 1123-4 (2002).
- (94) Clowse, M.E., Magder, L., Witter, F. & Petri, M. Hydroxychloroquine in lupus pregnancy. *Arthritis Rheum* 54, 3640-7 (2006).
- (95) Costedoat-Chalumeau, N. *et al.* Safety of hydroxychloroquine in pregnant patients with connective tissue diseases: a study of one hundred thirty-three cases compared with a control group. *Arthritis Rheum* **48**, 3207-11 (2003).
- (96) Arentz, M. *et al.* Characteristics and Outcomes of 21 Critically III Patients With COVID-19 in Washington State. *Jama*, (2020).
- (97) Vincent, M.J. *et al.* Chloroquine is a potent inhibitor of SARS coronavirus infection and spread. *Virology J* **2**, 69 (2005).
- (98) MA, A.-B. Chloroquine analogues in drug discovery: new directions of uses, mechanisms of actions and toxic manifestations from malaria to multifarious diseases. *J Antimicrob Chemother* **70**, 1608-21 (2015).
- (99) Ben-Zvi, I., Kivity, S., Langevitz, P. & Shoenfeld, Y. Hydroxychloroquine: from malaria to autoimmunity. *Clin Rev Allergy Immunol* **42**, 145-53 (2012).
- (100) Wang, M. *et al.* Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res* **30**, 269-71 (2020).

- (101) Liu, J. *et al.* Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting
   SARS-CoV-2 infection in vitro. *Cell Discov* 6, 1-4 (2020).
- (102) Touret, F. *et al.* In vitro screening of a FDA approved chemical library reveals potential inhibitors of SARS-CoV-2 replication. *BioRXiv*, (2020).
- (103) CDC.gov. Information for Clinicians on Investigational Therapeutics for Patients with Covid-19 (Clinical Care) [Internet]. 2020. https://www.cdc.gov/coronavirus/2019-ncov/hcp/therapeuticoptions.html. Accessed 20 Apr 2020.
- (104) Garcia-Cremades, M. *et al.* Optimizing hydroxychloroquine dosing for patients with COVID-19: An integrative modeling approach for effective drug repurposing. *Clin Pharmacol Ther.* (2020). doi 10.1002/cpt.1856
- (105) Fan, J. *et al.* Connecting hydroxychloroquine in vitro antiviral activity to in vivo concentration for prediction of antiviral effect: a critical step in treating COVID-19 patients. *Clin Infect Dis* (2020). Doi. 10.1093/cid/ciaa623
- (106) Perinel, S. *et al.* Towards Optimization of Hydroxychloroquine Dosing in Intensive Care Unit COVID-19 Patients. *Clin Infect Dis*, (2020). doi 10.1093/cid/ciaa394
- (107) Huang, M. *et al.* Treating COVID-19 with Chloroquine. *Journal of molecular cell biology* **12**, 322-5 (2020).
- (108) Hernandez, A.V., Roman, Y.M., Pasupuleti, V., Barboza, J.J. & White, C.M.J.A.o.I.M.
   Hydroxychloroquine or Chloroquine for Treatment or Prophylaxis of COVID-19: A Living Systematic Review. *Ann Intern med* (2020).doi 10.7326/M20-2496
- (109) Pastick, K.A. *et al.* Hydroxychloroquine and chloroquine for treatment of SARS-CoV-2 (COVID-19). *Open Forum Infectious Diseases*. **7**, ofaa130 (2020).
- (110) Gautret, P. *et al.* Clinical and microbiological effect of a combination of hydroxychloroquine and azithromycin in 80 COVID-19 patients with at least a six-day follow up: A pilot observational study. *Travel Med Infect Dis.* Mar-Apr 2020;34:101663 (2020).
- (111) Geleris, J. *et al.* Observational study of hydroxychloroquine in hospitalized patients with Covid-19. *N Engl J Med.* 2020 May 7; doi:10.1056/NEJMoa2012410. Online ahead or print. (2020).
- (112) Rosenberg, E.S. *et al.* Association of treatment with hydroxychloroquine or azithromycin with in-hospital mortality in patients with COVID-19 in New York state. *JAMA*. 2020 May 11;e208630. Doi: 10.1001/jama.2020.8630. Online ahead of print. (2020).

- (113) Tang, W. *et al.* Hydroxychloroquine in patients with mainly mild to moderate coronavirus disease 2019: open label, randomised controlled trial. *BMJ (Clinical research ed)* **369**, m1849 (2020).
- (114) Chen, J. *et al.* A pilot study of hydroxychloroquine in treatment of patients with common coronavirus disease-19 (COVID-19). *J Zhejiang Univ (Med Sci)* **49**, 0- (2020).
- Boulware, D.R. *et al.* A Randomized Trial of Hydroxychloroquine as Postexposure Prophylaxis for Covid-19. *N Engl J Med.* 2020 Jun 3 doi: 10.1056/NEJMoa2016638. Online ahead of print, (2020).
- (116) Bhimraj A, M.R., Shumaker AH, et al. Infectious Diseases Society of America Guidelines on the Treatment and Management of Patients with COVID-19. *IDSA*, (2020).

# Figure Legend

# Figure 1: Chemical structure of chloroquine and hydroxychloroquine

# Supplemental files

1. Table S1

ACCE

CQ Base Dose	Study Population [Age	N [nF]	Cmax, µg/mL	Tmax‡, hr	AUC (Time Duration <sup>§</sup>	T <sub>1/2</sub> , Days	CL	Vd	Bioanalytical method type:	Refer
	range in yearsj				),, h*µg/mL				Reported ELOQ	
Following Adm	inistration via Intrav	enous (IV)	) Infusion			I			1	
15 mg/kg as 4 hour IV	HV [18-46]	10 [2]	P:0.9±0.5	NA	NA	NA	P:36±14 l/hr	P:132±50 l/kg	HPLC:NA	(23)
infusion	Malaria Patients [15-35]	9 [2]	P:1.7±0.6	NA	NA	NA	P:32±15 l/hr	P:136±64 l/kg	HPLC:NA	
300 mg as 12- 24 mins IV infusion	HV [20-36]	11 [0]	P:0.8±0.3	NA	P:7.5±2.4 (168 hr)	P: 12	P:43±10 l/hr	P:204±86 1/kg	HPLC: 1 ng/mL	(22)
Following Sing	le Dose Administrati	on via Ora	l Route		1	1	I	1	1	1
150 mg tablet	HV [37-42]	5 [1]	NA	NA	B: 21.6 P: 1.9 (225 days)	B:27 P:33	B: 0.1 l/kg/hr P: 1.1 l/kg/hr	B: 79 l/kg P: 869 l/kg	HPLC: 1.5 Nmol/l in blood, 0.5 Nmol/l	(21)
300 mg tablet			NA	NA	B: 29.6 P: 5.2 (225 days)	B:53 P:63	B: 0.1 l/kg/hr P: 0.8 l/kg/hr	B: 185 l/kg P: 882 l/kg		
600 mg tablet			NA	NA	B: 92.1 P: 9.4 (225 days)	B:52 P:58	B: 0.1 l/kg/hr P: 0.9 l/kg/hr	B: 81 l/kg P: 710 l/kg		

Accepted

CQ Base Dose	Study Population [Age range in years]	N [nF]	Cmax, μg/mL	Tmax‡, hr	AUC (Time Duration <sup>§</sup> ),,	T <sub>1/2</sub> , Days	CL	Vd	Bioanalytical method type: Reported LLOQ	Refere nce
					h*µg/mL					
300 mg tablet	HV [20-36]	11	P:0.08±0.0 1	P:1-6	P: 6±1.3 (168 hr)	P:12	NA	NA	HPLC: 1 ng/mL	(22)
300 mg in solution	-		P:0.07±0.0 1	P:1-6	P: 5±0.9 (168 hr)	P: 9	NA	NA		
300 mg tablet	HV [23-30]	8 [3]	P:0.4±0.1	P:1-6	NA	P:4.4 ±0.4	P:5±0.4 l/kg/hr	P:16±3 l/kg	HPLC: 1 ng/mL	(8)
600 mg tablet	HV with history of CQ-induced pruritus [19-23]	8 [0]	P:0.4±0.2	P:4-6	P:19±2 (168 hr)	NA	P:16±5 l/hr <sup>¥</sup>	NA	HPLC: 10 ng/mL	(13)
	HV without history of CQ- induced pruritus [19-23]	6 [0]	P:0.2±0.06	P:2-6	P:17± 2 (168 hr)	NA	P:17±5 l/hr <sup>¥</sup>	NA		
600 mg tablet	HV [20-47]	16 [12]	P:0.3 (0.1- 0.5 <sup>‡</sup> )	P:1-6	P:15 (11– 25 <sup>‡</sup> ) (∞)	P:6 (3– 14)	P:40 (25–57 <sup>‡</sup> ) 1/h	P:7,600 (4,450– 12,400) 1	HPLC: 5 ng/mL	(14)

CQ Base	Study	N [nF]	Cmax,	Tmax‡,	AUC	T <sub>1/2</sub> ,	CL	Vd	Bioanalytical	Refere
Dose	Population [Age		µg/mL	hr	(Time	Days			method type:	nce
	range in years]				Duration <sup>§</sup>				Reported LLOQ	
					),,					
					h*µg/mL					
300 mg	HV [41 <sup>#</sup> ]	5 [2]	NA	NA	P:25	P:16±	P:0.4±0.1	P:250±116	HPLC:1 ng/mL	(24)
dose/week for					(42 Days)	5 $^{\Delta}$	l/kg/hr	l/kg		
3 weeks										
200 mg dose	HV [31 <sup>#</sup> ]	4 [0]	NA	NA	P:14	P:16±	P:0.6±0.1	P:302±102		
twice/week for					(42 Days)	$6^{\Delta}$	l/kg/hr	l/kg		
3 weeks										
50 mg	HV [30 <sup>#</sup> ]	5 [0]	NA	NA	P:13	P:20±	P:0.6±0.1	P: 283±112	-	
dose/day for 3					(42 Days)	$14^{\Delta}$	l/kg/hr	l/kg		
weeks										

# Table 1. Mean +/- SD Pharmacokinetic Estimates for Chloroquine (CQ)†

CQ Base	Study	N [nF]	Cmax,	Tmax‡,	AUC	T <sub>1/2</sub> ,	CL	Vd	Bioanalytical	Refere
Dose	Population [Age		µg/mL	hr	(Time	Days			method type:	nce
	range in years]				Duration <sup>§</sup>				Reported LLOQ	
					),,					
					h*µg/mL					

Abbreviations: AUC, Area under the concentration-time curve; B, Blood; CQ, Chloroquine; CL, Apparent clearance;  $C_{max}$ , Maximum plasma concentration; hr, hours; HV, healthy volunteers; IV, intravenous; LLOQ, Lower limit of quantification; NA, Information not available/reported; N, Number of subjects/patients; nF, Number of female subjects/patients; P, Plasma; SD, Standard deviation; Tmax, Time to reach Cmax;  $T_{1/2}$ , Elimination or terminal halflife; Vd, Apparent volume of distribution.

† Estimates were adjusted for unit uniformity and rounded.

# Mean age reported

‡ Range reported

§ If specific time duration for AUC is not reported, approximate time duration reported based on sampling duration or reported last concentrations

¥ Renal clearance estimates.

 $\Delta$  AUC estimates corrected to a single 600 mg dose

#### Table 2. Mean +/- SD Pharmacokinetic Parameter Estimates for Hydroxychloroquine (HCQ) † **HCQ Dose** Vd N [nF] AUC Study Tmax<sup>‡</sup>, $T_{1/2}$ , $\mathbf{CL}$ **Bioanalytical** Cmax, Population µg/mL (l/h) method type : hr (Time Days [Age range Reported **Duration** LLOQ in years] <sup>§</sup>), h\*µg/mL Following Administration via IV Infusion HV [19-27] 155 mg: IV P: 0.8 [0.4-P: HPLC: 1 5 [3] NA P: NA P: infusion over 0.5 h 1.1‡] 50±23 36757±11102 26±10 ng/mL B: 1.9 [1.1-B: 5791±2566 B: B: 6±1 2.4‡] 44±12

P: 1.7 [1.4-

B: 3.3 [2.3-

2.4‡]

4.2‡]

4[NA]

Refere

nce

(15)

Following Single Dose Administration via Oral Route

155 mg tablet	HV [19-27]	5 [3]	P: 0.05	2-4.5	NA	P: 32±9	NA	NA	HPLC: 1	(16)
			[0.03-0.08‡]						ng/mL	
			B: 0.2 [0.2-			B:				
			0.4‡]			50±16				
155 mg tablet with	HV [26.1 ±	9 [4]	B: 0.2 [0.1-	B:2.5-6	NA	NA	NA	NA	HPLC:NA	(26)
food	6.9 <sup>¥</sup> ]		0.3‡]							

P:

B:

53±22

43±22

P:

51757±30311

B: 5254±2021

P:

30±6

B: 6±1

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310 mg: IV

infusion over 0.5 h

Table 2.M	lean +/- SD F	harmac	okinetic Para	ameter Es	timates for ]	Hydroxyo	chloroqu	iine (HCQ) †		
HCQ Dose	Study Population [Age range in years]	N [nF]	Cmax, µg/mL	Tmax <sup>‡</sup> , hr	AUC (Time Duration <sup>§</sup> ), h*µg/mL	T <sub>1/2</sub> , Days	CL (1/h)	Vd	Bioanalytical method type : Reported LLOQ	Refere nce
155 mg in aqueous solution with food,			B: 0.2 [0.07- 0.4‡]	B:2.3-5.2						
155 mg tablet	HV [20-36]	24 [0]	B: 0.1 ± 0.04	B:3-4	$B:6.4 \pm 0.5$ (168 hr)	NA	B:2.7 ± 1.6¥	NA	HPLC: 5 ng/mL	(17)
155 mg tablet	HV [20-48]	10 [4]	B: 0.2±0.01	B:1.5-4.6	B: 1.8±0.9 (32 hr)	NA	NA	NA	HPLC: 1 ng/mL	(28)
155 mg in aqueous solution			B: 0.2±0.01	B:0.5-5.7	B: 1.7±1 (32 hr)	-				
155 mg two different tablet	HV [21-29]	20 [0]	P: 0.03 ± 0.001	P: 4 ±1¥	P: $1.8 \pm 0.4$ ( $\infty$ )	P:11±3	NA	NA	LC-MS/MS: 0.2 ng/mL	(29)
formulation			P: 0.04 ± 0.03	P: 4 ±1¥	$\begin{array}{c} P: 2 \pm 0.4 \\ (\infty) \end{array}$	P: 12±4				

# Table 2. Mean +/- SD Pharmacokinetic Parameter Estimates for Hydroxychloroquine (HCQ) †

HCQ Dose	Study	N [nF]	Cmax,	Tmax <sup>‡</sup> ,	AUC	T <sub>1/2</sub> ,	CL	Vd	Bioanalytical	Refere
	Population		μg/mL	hr	(Time	Days	(l/h)		method type :	nce
	[Age range				Duration				Reported	
	in years]				<sup>§</sup> ),				LLOQ	
					h*uɑ/mI					
				1	II µg/IIIL	1				

Abbreviations: AUC, Area under the concentration-time curve; B, Blood; CQ, Chloroquine; CL, Apparent clearance; C<sub>max</sub>, Maximum plasma concentration;

hr, hours; HV, healthy volunteers; IV, intravenous; LLOQ, Lower limit of quantification; NA, Information not available/reported; N, Number of

subjects/patients; nF, Number of female subjects/patients; P, Plasma; SD, Standard deviation; Tmax, Time to reach Cmax; T<sub>1/2</sub>, Elimination or terminal halflife; Vd, Apparent volume of distribution.

ine; vd, Apparent volume of distribution.

 $\ensuremath{^{\dagger}}$  Estimates were adjusted for unit uniformity and rounded.

# Mean age reported

‡ Range reported

§ If specific time duration for AUC is not reported, approximate time duration reported based on sampling duration or reported last concentrations

¥ Renal clearance estimates

Precipi	tant			
Object/Precipitant	Object/Precipi	Change in	Chloroquine	Reference
	tant Dose	AUC (%)	Dose	
Object			1	
Acetaminophen	1.5 g	21.7	250 mg chloroquine phosphate IM	(46)
(Paracetamol)			(150 mg chloroquine base) Single	
			Dose	
Levonorgestrel	150 µg	56.7	300 mg (formulation not mentioned)	(47)
			Single Dose	
Primaquine	30 mg (base)	21	1000 mg chloroquine phosphate	(14)
			(600 mg base) Single Dose	
Tafenoquine	450 mg	23.5	1000 mg chloroquine phosphate QD	(48)
			(600 mg base) for 2 days	
Ethinylestradiol	30 µg	-4.6	300 mg (formulation not mentioned)	(47)
			Single Dose	
Azithromycin	1000 mg	3.0	1000 mg chloroquine phosphate (600	(49)
			mg base) QD on Days 1 and 2, 500	
			mg chloroquine phosphate (300 mg	
			base) on Day 3	
Precipitant			1	
Acetaminophen	500 mg	23.9	600 mg (formulation not mentioned)	(50)
(Paracetamol)	Single Dose		Single Dose	
Cimetidine	400 mg QD	113	chloroquine sulfate (600 mg base)	(8)
	for 12 days			
Metamizole	500 mg	22.9	600 mg (formulation not mentioned)	(8)
(Dipyrone)	Single Dose		Single Dose	
Acetylsalicylic acid	325 mg	0.9	600 mg (formulation not mentioned)	(8)
(Aspirin)	Single Dose		Single Dose	
Azithromycin	1000 mg QD	-3.8	1000 mg chloroquine phosphate (600	(50)
	for 3 days		mg base) QD on Days 1 and 2,	
			500 mg chloroquine phosphate (300	
			mg base) on Day 3	

Table 3.DrugPrecent	g-Drug Interaction ipitant	n Studies Inv	volving Chloroquine as the Objec	et and the
Object/Precipitar	nt Object/Precipi tant Dose	Change in AUC (%)	Chloroquine Dose	Reference
Methylene blue	130 mg BID For 3 days	-17.9	Males: 1000 mg chloroquine phosphate QD (600 mg base) on Days 1 and 2 and 500 mg (300 mg base) on Day 3	(51)
			Females: 750 mg chloroquine phosphate (450 mg base) on Days 1 and 2 and 375 mg (225 mg base) on Day 3	
Primaquine	30 mg (base) Single Dose	6.7	1000 mg chloroquine phosphate (600 mg base) Single Dose	(48)
Tafenoquine	450 mg QD 2 days	-4	1000 mg chloroquine phosphate (600 mg base) QD on Days 1 and 2, 500 mg chloroquine phosphate (300 mg base) on Day 3	(49)
Tafenoquine	450 mg Single Dose	-3.7	1000 mg chloroquine phosphate (600 mg base) QD for 2 days	(49)
Tafenoquine	450 mg QD Days 2 and 3	5.6	1000 mg chloroquine phosphate (600 mg base) QD on Days 1 and 2, 500 mg chloroquine phosphate (300 mg base) on Day 3	(49)
Roselle (Hibiscus sabdariffa) Sudanese beverage	300 mL Single Dose	-71.3	600 mg (formulation not mentioned) Single Dose	(52)
Sweet lemon (Citrus limetta) Sudanese beverage	s 300 mL Single Dose	-68	600 mg (formulation not mentioned) Single Dose	(52)
Tamarind (Tamarindus indica) Sudanese beverage	300 mL Single Dose	-65.4	600 mg (formulation not mentioned) Single Dose	(52)

Table 3. Drug-D Precipit	orug Interaction tant	n Studies Inv	olving Chloroquine as the Objec	ct and the
Object/Precipitant	Object/Precipi	Change in	Chloroquine	Reference
	tant Dose	AUC (%)	Dose	
Source: UW Drug Intera	action Database (D	IDB) Copyright	University of Washington, accessed: Ap	ril 6-14, 2020
Note: All doses adminis	tered orally unless	otherwise stated	l	
Precipitant term refer	rs to the drug that	t causes an eff	fect on the substrate drug by inhibit	ing or
inducing enzymes				
Object term refers to	the drug whose	exposure may	or may not be changed by a precip	itant drug

Table 4 I	Drug-Drug Interaction	on Studies Involvi	ing Hydroxychloro	quine as the Objec	t and the
1	Precipitant				
Object/	Object/	Change in	HCQ Dose	HCQ Interval	Reference
Precipitant	Precipitant	AUC (%)			
	Dose				
Object				•	
Methotrexate	15 mg	51.8	200 mg	single dose	(28)
			400 mg		(55)
Metoprolol	100 mg	50.7	(8 days)	twice daily	
	200 mg		400 mg		(56)
MK-2206	(21-day cycles)	16.4	(21-day cycles)	twice daily	
	150 mg		200 mg		(56)
MK-2206	(21-day cycles)	31.7	(21-day cycles)	twice daily	
	200 mg		200 mg		(56)
MK-2206	(21-day cycles)	56.2	(21-day cycles)	twice daily	
	135 mg		400 mg		(56)
MK-2206	(21-day cycles)	92.2	(21-day cycles)	twice daily	
Precipitant		•		•	
Methotrexate	15 mg (Single	-6.8	200 mg (Single		(28)
	Dose)		Dose)		
Abbreviations:	AUC, area under the	concentration-time	e curve; HCQ, hydro	xychloroquine	•
Source: UW Dr	rug Interaction Databa	ase (DIDB) Copyri	ght University of W	ashington, accessed	: April 6-14, 2020
Precipitant te	rm refers to the dru	ig that causes ar	effect on the sub	strate drug by inl	nibiting or
inducing enz	ymes				
Object term r	efers to the drug w	hose exposure n	nay or may not be	changed by a pr	ecipitant drug

Study Population [Age]	N [nF]	Treatment	PK matrix	Refere ce
Chloroquine	1		I	1
Pediatric malaria	83	Age 6-24 months:	Finger	(60)
patients	[34]	CQ (75 mg base/day for three days) +	prick/dried	
[Range: 6 months-5		sulfadoxine/pyrimethamine (250 mg/12.5 mg single	blood spot	
years]		dose)	sampling	
		Age 25-60 months:		
		CQ (150 mg base/day for three days) +		
		sulfadoxine/pyrimethamine (500 mg/25 mg single		
		dose)		
Pregnant and non-	60	450 mg CQ base for three days +	Plasma	(61)
pregnant women	[60]	single dose of 1500 mg sulfadoxine and		
[Mean age: 26		75 mg pyrimethamine		
years]				
Malaria patients	75	Multiple doses:	Plasma	(62)
[Range: 17-52	[39]	10 and 5 mg/kg CQ base at 0 h and 6–12 h on day 0,		
years]		and 5 mg/kg each on day 1 and day 2 + 15		
		mg/kg/day primaquine base for 14 days starting		
		from the second day (day 1) of CQ		
Hydroxychloroquine			I	1
Rheumatoid	123	HCQ dosing varied:	Whole blood	(63)
arthritis patients	[88]	Single dose: 155 mg HCQ base orally or via 30 min		
[Range: 20-81		IV infusion		
years]		Multiple doses: 155 mg/day or 310 mg/day HCQ		
		base dose orally with or without methotrexate		
Healthy adults and	91	Healthy adults: Single oral dose of 310 mg HCQ	Plasma	(18)
malaria patients	[21]	base or 310 mg/day HCQ base dose/week		
[Approximate mean		Malaria patients: 620 mg HCQ base + 310 mg		
age: 27 years]		HCQ base at 6, 24, and 48 hours		
Patients with CLE	90	Multiple doses: 155-310 mg HCQ base/day	Blood and	(64)
or SLE [Mean age:	[66]		Plasma	
42.5 years]				

)	Table 5.Popula	tion Pha	rmacokinetic Studies for Chloroquine and Hydroxyc	chloroquine	
	Study Population [Age]	N [nF]	Treatment	PK matrix	Referen ce
	Pregnant women	50	Multiple doses: 310 mg /day HCQ base dose for	Serum	(65)
	with rheumatic	[50]	most subjects with or without other concomitant		
	diseases		prescription medications		
)	[Median age: 31				
	years]				
	Abbreviations: CLE,	cutaneous	lupus erythematosus; CQ, chloroquine; HCQ, hydroxy	chloroquine; N,	number
	of subjects/patients; n	subjects/patients; nF, number of female subjects/patients; NA, information not available/reported; PK,			
	pharmacokinetic; SLE	E, systemi	c lupus erythematosus		

Table 6Analysis of Mean Change from Time-Matched Baseline in QTcF Repeated<br/>Measures ANOVA: Comparisons of Chloroquine versus Placebo Cohorts on<br/>Day 3 of a 1000 mg Chloroquine Phosphate QD regimen.

	ΔQTcF (msec)				
Time		90% Confidence Interval			
(hr postdose)	Mean	Upper Limit	Lower Limit		
0	18.4	13.3	23.5		
1	22.2	17.0	27.3		
2	25.7	20.6	30.8		
3	27.6	22.5	32.7		
4	31.3	26.1	36.4		
5	29.2	24.1	34.3		
6	29.9	24.8	35.1		
8	31.5	26.3	36.6		
10	35.0	29.9	40.2		
12	32.4	27.3	37.6		

Calculation of  $\Delta QTcF$ : The primary endpoint was change of QTcF from Day 1 baseline at each nominal time point on Day 3, using Fridericia's method correcting for heart rate effects on the QT interval. Comparisons were made between chloroquine versus placebo using an ANOVA model.

Cell Type	Viral Input	Drug	CQ EC <sub>50</sub> µ	HCQ EC <sub>50</sub> µ	Reference
	MOI	Incubation (hr)	M(ng/mL	M (ng/mL	
Vero E6	0.05	48	1.13 (361)	N/A	(100)
Vero	0.01	24	23.90(7646)	6.14 (2062)	(2)
Vero	0.01	48	5.47(1750)	0.72 (242)	(2)
Vero E6	0.01	48	2.71 (867)	4.51 (1515)	(101)
Vero E6	0.02	48	3.81 (1219)	4.06 (1364)	(101)
Vero E6	0.2	48	7.14 (2284)	17.31 (5814)	(101)
Vero E6	0.8	48	7.36 (2354)	12.96 (4353)	(101)
Vero E6	0.001	48	N/A	4.17 (1401)	(102)

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