

UC Davis

UC Davis Previously Published Works

Title

The physiological response of Populus tremula x alba leaves to the down-regulation of PIP1 aquaporin gene expression under no water stress

Permalink

<https://escholarship.org/uc/item/7tv6b5dk>

Journal

Frontiers in Plant Science, 4(DEC)

ISSN

1664-462X

Authors

Secchi, Francesca

Zwieniecki, Maciej A

Publication Date

2013

DOI

10.3389/fpls.2013.00507

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed



The physiological response of *Populus tremula x alba* leaves to the down-regulation of *PIP1* aquaporin gene expression under no water stress

Francesca Secchi* and Maciej A. Zwieniecki

Department of Plant Sciences, University of California Davis, Davis, CA, USA

Edited by:

Andrew J. McElrone, US
Department of Agriculture-
Agricultural Research Service, USA

Reviewed by:

Suleyman I. Allakhverdiev, Russian
Academy of Sciences, Russia
Xia Wu, University of Washington,
USA
Alicia Pou, University Catholique de
Louvain, Belgium

*Correspondence:

Francesca Secchi, Department of
Plant Science, University of
California Davis, One Shields
Avenue, Davis, CA 95616, USA
e-mail: fsecchi@ucdavis.edu

In order to study the role of *PIP1* aquaporins in leaf water and CO₂ transport, several lines of *PIP1*-deficient transgenic *Populus tremula x alba* were generated using a reverse genetic approach. These transgenic lines displayed no visible developmental or morphological phenotypes when grown under conditions of no water stress. Major photosynthetic parameters were also not affected by *PIP1* down regulation. However, low levels of *PIP1* expression resulted in greater leaf hydraulic resistance (an increase of 27%), which effectively implicated *PIP1* role in water transport. Additionally, the expression level of *PIP1* genes in the various transgenic lines was correlated with reductions in mesophyll conductance to CO₂ (g_m), suggesting that in poplar, these aquaporins influenced membrane permeability to CO₂. Overall, although analysis showed that *PIP1* genes contributed to the mass transfer of water and CO₂ in poplar leaves, their down-regulation did not dramatically impair the physiological needs of this fast growing tree when cultivated under conditions of no stress.

Keywords: transgenic poplar, aquaporin, leaf hydraulic resistance, mesophyll conductance, photosynthesis

INTRODUCTION

Water use efficiency (WUE) describes one of the major plant tradeoffs between rates of photosynthesis and transpiration (Gilbert et al., 2011). In general, increases in transpiration rates lead to higher photosynthetic activity. However, leaf hydraulic resistance often determines leaf transpiration capacity and is thus, directly related to carbon dioxide uptake (Brodribb et al., 2005). What is even more important, both water and CO₂ share important parts of their transport paths in leaves, including not only the gas phase but also the liquid phase where their movement through cellular membranes is facilitated by water channels (aquaporins) (Heinen et al., 2009). The investment in the abundance of water channels would provide two major benefits. (1) It would facilitate water fluxes across the plant by reducing hydraulic resistance for water uptake in roots and improving water distribution in leaves allowing for the maintenance of relatively high stomatal conductance (g_s) and so facilitating CO₂ diffusion into leaves. (2) Investment in aquaporins would increase mesophyll conductance (g_m) facilitating passage of CO₂ from air to chloroplasts. Thus, improving our understanding of the role of aquaporins in leaves remains an important topic of research agenda.

The plasma membrane proteins (PIP) family is divided into two subfamilies: PIP2 and PIP1. PIP2 proteins have been shown to be major channels for water (Chrispeels et al., 2001), PIP1 proteins have been shown to facilitate both, the diffusion of CO₂ (Uehlein et al., 2003) and supplement water permeability in conjunction with PIP2 proteins (Fetter et al., 2004; Temmei et al., 2005). Analysis of aquaporin contribution to membrane permeability to water or CO₂ usually comes from studies aiming at blocking aquaporins using heavy metals (Barone et al.,

1997; Hukin et al., 2002; Niemietz and Tyerman, 2002), ATP (Alves et al., 2001; Martinez-Ballesta et al., 2003) or anoxia (Tournaire-Roux et al., 2003; Frick et al., 2013). Most of these studies are related to analysis of water uptake by roots as infiltration of leaves with chemical compounds is much more difficult to achieve. However, a few reports showed that leaves of *Vicia faba* and *Phaseolus vulgaris* had limited mesophyll CO₂ conductance when treated with mercury chloride, suggesting an involvement of aquaporins in CO₂ diffusion across plasma membranes (Terashima and Ono, 2002). The problems of using chemical treatments to block aquaporins in leaves can be overcome with the use of transgenic lines with down or up-regulated expression of genes from the aquaporin's family (Hanba et al., 2004; Heckwolf et al., 2011; Uehlein et al., 2012). The majority of information we have up to date comes from analysis of the AQP1 water channel isolated from tobacco (Siefritz et al., 2001; Flexas et al., 2006) and PIP1;2 from *Arabidopsis thaliana* (Heckwolf et al., 2011). It was shown through functional assay and *in vivo* studies that NtAQP1 was associated with facilitating the diffusion of CO₂ as well as water (Siefritz et al., 2001; Uehlein et al., 2003, 2008; Flexas et al., 2006) and AtPIP1;2 was a relevant facilitator of CO₂ diffusion (Heckwolf et al., 2011; Uehlein et al., 2012). As both genes are members of the PIP1 subfamily, one can expect that a reduction in the expression level of *PIP1* genes should influence both photosynthesis via a reduction in g_m, and water transport via an increase of leaf hydraulic resistance. If the role of PIP1s is truly important then a reduction in the level of their gene expression should result in detrimental effects related to photosynthetic parameters and growth especially in productive species like poplar.

Species from the genus *Populus* are generally fast growing, short lived trees with high photosynthetic and transpiration rates making them a valuable species for biofuel production, wood for pulp, and residential areas that are in need of fast shade (Isebrands and Richardson, 2013). These physiological characters are necessary from the perspective of natural history of the genus as it evolved to often be one of the first trees in succession series on land opened following natural disturbance, where competitive success depends on fast growth rates but not necessarily on efficiencies of resource use. In nature, most of the species from that genus grow in relatively mesic habitats with easy access to water and nutrients (Isebrands and Richardson, 2013). High availability of light, water, and nutrients in naturally disturbed areas (fire, hurricane etc.) is only beneficial to species that can capture the resource faster than their competitors. This strategy does not promote efficiency in resource utilization but relies on a maximization of physiological traits that result in fast growth. When compared with other tree species *Populus* might look almost wasteful with its low WUE (Brueck, 2008) and relatively high nitrogen content (Niinemets, 1997). Thus, fast growing hybrid poplar might be a good subject to determine the role of *PIP1* aquaporins in the physiology of both CO₂ uptake and leaf hydraulics. We used transgenic plants with down-regulated expression of *PIP1* genes, with an expectation that performance of this fast growing species would be significantly impaired by a reduction in its capacity to transport water and CO₂ across membranes.

RESULTS

POPULUS TRANSFORMATION

Down-regulation of *PIP1* gene expression in hybrid poplar (*Populus tremula x alba*) was achieved by using a ~270-bp hairpin RNAi construct designed from sequences in the poplar *PIP1* subfamily. Primers were specifically designed for alignment on *PIP1* subfamily conserved domains (on five *PIP1* isoforms from *P. trichocarpa*). Agrobacterium-mediated transformation yielded numerous independent transformants. We analyzed 22 propagated lines and PCR revealed that all contained the transgene product of the expected size for the kanamycin resistance marker *nptII* gene. From those, six different transformed lines were randomly chosen for molecular and physiological analyses. To estimate levels of *PIP1* subfamily down-regulation in the resulting transgenic plants the qRT-PCR analyses were performed on leaves of mature plants. The target of the RNAi construct clearly showed significant down-regulation in all of the tested transgenic lines. *PIP1* transcription levels were reduced by 93–85% among lines (Figure 1; ANOVA $p < 0.001$). Transgenic line 5 was characterized by lower expression levels of *PIP1* genes and was also significantly different from lines 3 and 4.

In order to confirm the down regulation of different isoforms belonging to the same subfamily the expression level of two isoforms *PIP1.1* and *PIP1.3* were monitored. These genes were chosen because they were the most highly expressed genes in *P. trichocarpa* leaf tissue (Secchi et al., 2009). Both genes were strongly down-regulated in all transgenic lines (Figure 2A); however, we

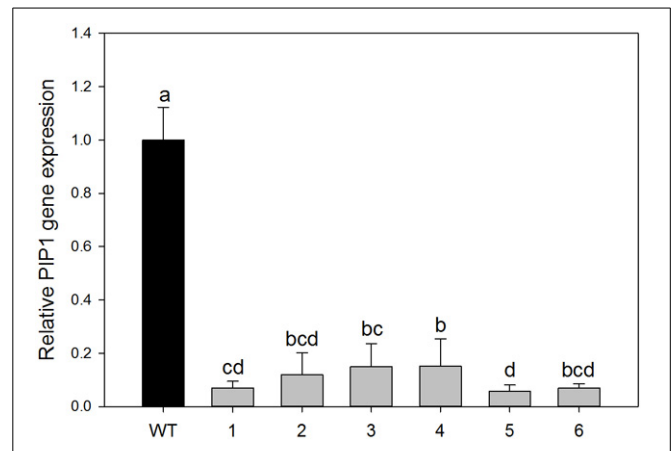


FIGURE 1 | Relative expression of *PIP1* genes in the leaves of wild-type and transgenic lines. Each histogram is the average of three independent biological samples with two technical replicates; error bars represent SE. Letters denote homogeneous groups based on the Fisher LSD method corrected for the multiple comparisons. There is a significant difference between relative gene expression in the seven different plant groups (wt and the six transgenic lines, 1–6), ($P < 0.001$, One Way ANOVA test).

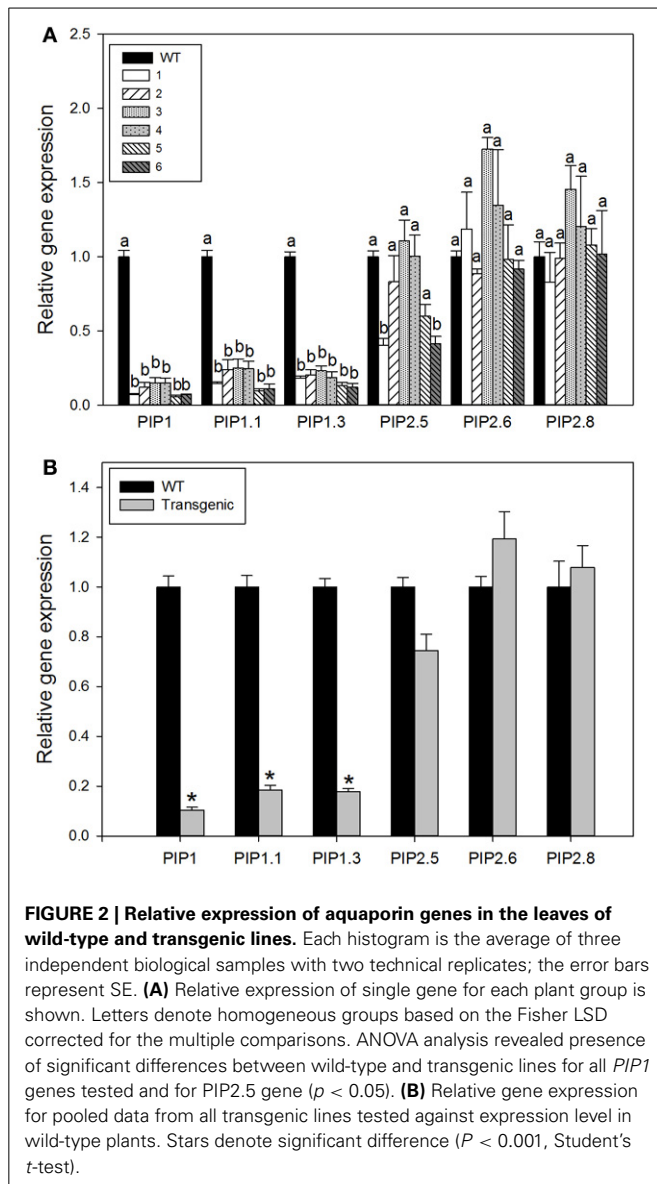
did not measure their protein abundance and thus, strong down gene regulation could be not coincided to the same strong protein suppression, although there are evidences were reduced abundance of protein was consistent with the previous analysis of RNA expression (Flexas et al., 2006).

To compare the effect of our construct on expression of the closely related *PIP2* subfamily, we measured the expression of three different members; *PIP2.5*, *PIP2.6*, and *PIP2.8*. In general, their transcript levels were not significantly different among lines. The expression was in some cases higher and some cases lower than in wild-type plants (Figure 2A). Thus, no compensatory responses occurred in members of *PIP2* gene subfamily in response to *PIP1* down-regulation (Figure 2B).

EFFECT OF DOWN-REGULATION OF *PIP1* EXPRESSION ON LEAF GAS EXCHANGE

Transgenic lines and wild-type plants were grown under the same conditions in the greenhouse (for details see methods). They were phenotypically undistinguishable. No differences were found in the specific leaf area (SLA) and chlorophyll content (Table 1). In addition, the maturation process of the leaves, estimated from the changes in chlorophyll content in growing leaves, was not different among lines (Supplemental Figure 1). There were no statistical differences in basic photosynthetic parameters (net CO₂ assimilation [A_N], stomatal conductance to water [g_s], RuBP regeneration rate [J], maximum rate of Rubisco carboxylation [$V_{c_{max}}$], or mitochondrial respiration in the light [R_d]; Table 1).

However, significant differences were found in mesophyll conductance to CO₂ (g_m) estimated using CO₂ curve-fitting method in combination with chlorophyll fluorescence to better estimate limitation states. As estimation of g_m using this method might be influenced by curve parameterization, we provide raw data for each plant such that reader can build their



own feel for the validity of the conclusion (SOM). In four different transgenic lines (1, 2, 5, and 6), g_m was less than half of that determined for wild-type plants. In the two remaining lines (3 and 4) there was a statistically significant reduction of g_m compared with the wild-type plants although it was physiologically less pronounced (Figure 3). Estimated mesophyll conductance (g_m) was positively correlated with the level of down-regulation of *PIP1* expression ($R^2 = 0.655$, $p = 0.05$ and $df = 1,4$) even when analyzed only for the transgenic lines. This suggests that the relation between *PIP1* down regulation and the observed reduction of mesophyll conductance to CO_2 might be causative and that estimation of the g_m using CO_2 curve fitting might reflect a real trend. An analysis of carbon isotopic composition ($\delta^{13}C$) showed no significant differences between wild-type and transgenic lines (Figure 4A, ANOVA $p = 0.74$ or Kruskal–Wallis ANOVA non-parametric test $p = 0.36$) decreasing the physiological importance of the observed

g_m reduction. A non-parametric analysis showed significant increases of nitrogen isotopic composition ($\delta^{15}N$) in transgenic plants (Figure 4B, Kruskal–Wallis ANOVA non-parametric test $p = 0.023$) although this increase was very small and not picked up by regular analysis of variance ($p = 0.23$). *Post-hoc* LSD test showed that only transgenic line 3 had a significant increase in $\delta^{15}N$.

EFFECT OF DOWN-REGULATION OF PIP1 EXPRESSION ON LEAF REHYDRATION KINETICS

Analysis of the leaf rehydration kinetics of wild type *Populus tremula x alba* leaves shows a pattern similar to that of leaf rehydration kinetics in other angiosperm trees (Zwieniecki et al., 2007), with two distinct phases: (1) a fast phase that accounts for approximately 50% of water uptake during rehydration and (2) a slow phase accounting for the remaining observed water uptake. The time constant of the fast phase in wild-type plants was determined to be only 10.8 s. Five out of six transgenic lines showed a significant increase in the time constant of the fast phase by an average of 27% to approximately 15 s (Figure 5; ANOVA $p = 0.0007$). Increase of the time constant was relatively small in relation to relative drop in expression level of *PIP1* aquaporins (Figure 5 inset) No significant changes in time constant of the second (slow) phase were observed between species (wild-type average was 238 s while average of for all transgenic plants was 245 s).

DISCUSSION

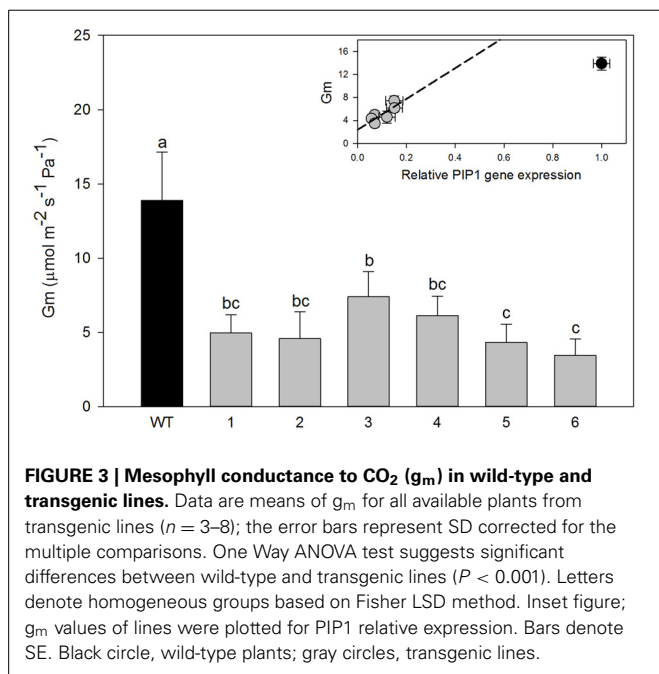
Leaves play a key role in plant growth and development, and they need to be continuously supplied with water and carbon dioxide in order to fulfill their photosynthetic functions (Heinen et al., 2009). Although the role of aquaporins as water channels is well accepted, and AtPIP1;2 and NtAQP1 have been confirmed to be CO_2 transport facilitators (Uehlein et al., 2003; Heckwolf et al., 2011), their involvement in facilitating CO_2 diffusion in woody plants is still under debate. To elucidate the dual role of aquaporins as facilitators of water and CO_2 exchange in leaves, we adopted a reverse genetics approach to investigate the effect of the RNAi-mediated suppression of *PIP1* genes on leaf hydraulic and photosynthesis characteristics. Since one aquaporin subfamily is composed of multiple genes with overlapping functions [*Populus trichocarpa PIP1* subfamily consists of five different genes; (Secchi et al., 2009; Almeida-Rodriguez et al., 2010)], we decided that a feasible approach to test their role was to down regulate more genes belonging to the same subfamily in order to reduce the potential for substitution of function or expression compensation within the family. We succeeded in generating several transgenic lines with substantial reductions in *PIP1* gene expression levels of which six randomly selected lines were characterized in detail. These lines did not show a significant effect on *PIP2* expression as compensation for *PIP1* down-regulation, despite that fact that all tested isoforms belonging to the *PIP1* subfamily were strongly down-regulated. Accordingly, we believe that the observed phenotypic differences between wild type and transgenic plants can be accounted for by the reduced expression of *PIP1* family members, which specify the role of these genes.

Despite a significant reduction in *PIP1* gene expression level, no morphological differences were observed between wild-type

Table 1 | Photosynthetic parameters, physiological characteristics and isotope analysis of the studied genotypes.

	WT	1	2	3	4	5	6	<i>p</i> -value
A_{max} ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$)	16.24 ± 0.52	17.75 ± 2.19	18.33 ± 0.82	18.19 ± 1.19	19.61 ± 1.17	20.39 ± 0.98	15.35 ± 1.34	0.098
A_N ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$)	13.17 ± 0.49	14.23 ± 1.32	13.66 ± 1.16	14.50 ± 0.82	15.08 ± 0.59	15.07 ± 0.89	11.95 ± 0.98	0.23
$g_s \text{ max}$ ($\text{mol m}^{-2} \text{ s}^{-1}$)	0.292 ± 0.007	0.329 ± 0.020	0.313 ± 0.033	0.296 ± 0.013	0.291 ± 0.012	0.300 ± 0.019	0.316 ± 0.009	0.517
g_s ($\text{mol m}^{-2} \text{ s}^{-1}$)	0.183 ± 0.017	0.269 ± 0.028	0.207 ± 0.061	0.211 ± 0.016	0.198 ± 0.017	0.205 ± 0.038	0.168 ± 0.038	0.316
g_m ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1} \text{ Pa}^{-1}$)	13.90 ± 1.15	4.97* ± 0.54	4.59* ± 1.05	7.42* ± 0.84	6.15* ± 0.58	4.33* ± 0.54	3.47* ± 0.63	<0.001
J ($\mu\text{mol m}^{-2} \text{ s}^{-1}$)	86.75 ± 2.04	92.20 ± 9.76	94 ± 9.77	97.25 ± 6.61	102.80 ± 5.27	111.60* ± 5.05	82.00 ± 1.18	0.02
$V_c \text{ max}$ ($\mu\text{mol m}^{-2} \text{ s}^{-1}$)	62.75 ± 2.20	68.20 ± 8.99	71.33 ± 3.84	71.75 ± 7.05	78.00 ± 4.64	89.00* ± 4.99	62.33 ± 1.86	0.016
R_d ($\mu\text{mol m}^{-2} \text{ s}^{-1}$)	1.13 ± 2.20	1.22 ± 0.10	1.16 ± 0.17	1.48 ± 0.09	1.14 ± 0.12	1.38 ± 0.13	1.13 ± 0.34	0.405
Chlorophyll (SPAD values)	41.40 ± 1.62	37.42 ± 2.45	39.30 ± 0.58	39.93 ± 2.23	40.06 ± 1.17	41.84 ± 2.18	35.47 ± 4.21	0.451
SLA ($\text{m}^2 \text{ g}^{-1}$)	0.0192 ± 0.001	0.0198 ± 0.002	0.0198 ± 0.002	0.0202 ± 0.001	0.0196 ± 0.001	0.0184 ± 0.001	0.0202 ± 0.003	0.977
$\delta^{13}\text{C}$	-30.52 ± 0.43	-31.27 ± 0.78	-30.81 ± 0.72	-30.39 ± 0.46	-30.62 ± 0.55	-30.02 ± 0.44	-30.34 ± 0.07	0.758
C Amount ($\mu\text{g mg}^{-1}$)	460.98 ± 4.45	456.95 ± 3.78	470.10 ± 10.64	468.61 ± 4.02	467.93 ± 1.59	463.93 ± 3.72	464.00 ± 5.78	0.475
$\delta^{15}\text{N}$	5.84 ± 0.13	6.05 ± 0.42	6.45 ± 0.17	6.55* ± 0.10	6.24 ± 0.14	5.88 ± 0.10	6.35 ± 0.15	0.227
N Amount ($\mu\text{g mg}^{-1}$)	30.93 ± 1.81	37.90 ± 2.77	41.26 ± 5.94	40.23 ± 1.88	39.83 ± 2.14	36.07 ± 3.50	32.59 ± 4.71	0.206

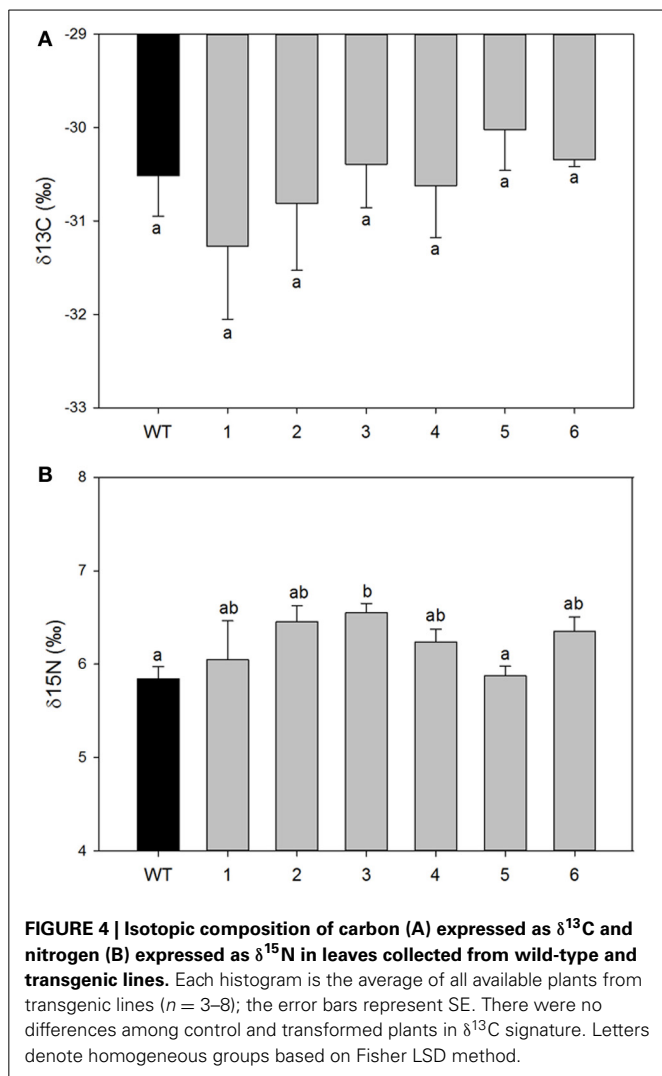
Values are means ± SE. *Statistically significant differences from post-hoc Fisher least significant difference (LSD with $\alpha = 0.05$) corrected for the multiple comparisons. *P*-value represents result of test for significant differences from ANOVA analysis. A_{max} , maximum photosynthesis; A_N , net photosynthesis; g_{max} , maximum stomatal conductance; g_s , stomatal conductance; g_m , mesophyll conductance; J , electron transport rate; $V_c \text{ max}$, maximum rate of Rubisco carboxylation; R_d , mitochondrial respiration in the light; SLA, specific leaf area; $\delta^{13}\text{C}$, isotopic composition of carbon; C, carbon; $\delta^{15}\text{N}$, isotopic composition of nitrogen; N, nitrogen.



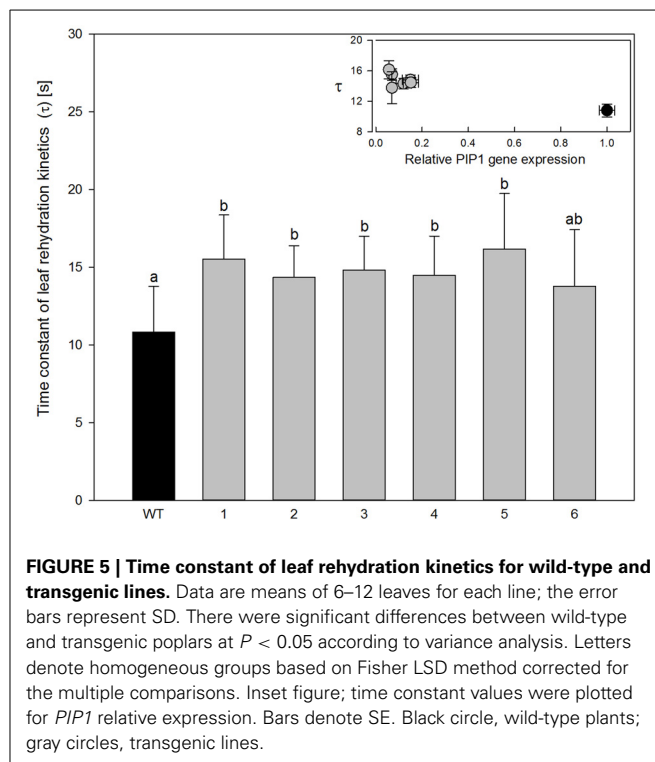
and transgenic plants. This result comes with some level of surprise although previous studies show no strong phenotypes for plants with modified expression of one or more specific aquaporin (Siefritz et al., 2002; Flexas et al., 2006; Heckwolf et al., 2011; Perrone et al., 2012). However, our general expectation was that transgenic plants with reductions of *PIP1* expression level would grow slower due to limitations related to CO₂ uptake and/or would compensate for higher hydraulic resistance with changes in leaf morphology similar to traits associated with drought stress tolerance. Neither of these expectations was met for plants

growing in the greenhouse environment, which were characterized by an ample supply of water, nutrients, and light. It seemed that a massive reduction of *PIP1* aquaporin gene expression was irrelevant to general plant performance under no-stress conditions. This would suggest that increasing *PIP1* transcript level is not strictly related to poplar performance but that these genes might play a secondary role or be important only under stress conditions. A similar conclusion was reported by Siefritz et al. (2002) showing that, despite a strong reduced in *NtAQP1* gene expression and root hydraulic conductivity, tobacco plants grown under optimal conditions in the greenhouse did not show morphological changes and that changes in expression did not seem to be important for water uptake. However, in a water limited environment antisense plants were not able to maintain turgor and seemed to be less water stress tolerant (Siefritz et al., 2002).

Even with a lack of any obvious morphological differences between wild-type and transgenic plants, we found specific physiological phenotypes consistent with reduced expression of *PIP1* genes. For example, all transgenic lines showed reductions in mesophyll conductance to CO₂. This g_m reduction was correlated with the expression level of *PIP1*, suggesting a causal relationship between *PIP1*s and g_m . This finding supports the role of PIP1s in the facilitation of diffusion of CO₂ across membranes. However, unlike in previous reports (Flexas et al., 2006), we did not observe any significant change in leaf photosynthetic capacity or in levels of carbon isotopic discrimination ($\delta^{13}\text{C}$). This would suggest that limitations imposed by g_m to CO₂ uptake in poplar were not a limiting factor. In fact, measured g_m in wild-type plants was high ($\sim 14 \mu\text{mol m}^{-2} \text{ s}^{-1} \text{ Pa}^{-1}$) and reductions by even 50% and higher still resulted in levels of g_m at the level of high performing coniferous leaves (Bickford et al., 2010; Silim et al., 2010; Zhu et al., 2011). It is, however, imperative to add here that CO₂ curve fitting method used here to estimate g_m is not the most reliable and future research with the inline ¹³C should be performed.



Similarly, we observed changes in the hydraulic properties of leaves that were also consistent with expectations, as reduced expression level resulted in a significant increase of the rehydration time constant for the fast water compartment in leaves. Although fast phase in both *wt* and transgenic plants was characterized by one of the fastest time constants among angiosperm species measured using this technique, the relative significance is related to change of the constant that reflects an increase of leaf hydraulic resistance directly involved in the transpiration path (Zwieniecki et al., 2007). As leaf hydraulic resistance was shown to correlate with the stomatal conductance and maximum photosynthetic capacity of leaves (Brodribb et al., 2005), we expected to observe some effect of increased leaf hydraulic resistance on photosynthesis in transgenic lines. However, this was not the case. Neither maximum g_s , nor maximum photosynthetic capacity was affected, and based on $\delta^{13}\text{C}$ results there was no limit in CO_2 supply that could reflect some difficulty in access to water. However we observed a small increase of $\delta^{15}\text{N}$ in the leaf tissue. This increase could potentially reflect a higher availability of NADPH for N assimilation when



competition from CO_2 is reduced [i.e., lower concentration of CO_2 in chloroplasts; (Bloom et al., 2010)]. However, only one line showed a significant increase of $\delta^{15}\text{N}$ and ANOVA analysis did support any general conclusion regarding *PIP1* role in change of $\delta^{15}\text{N}$ in transgenic plants.

Overall, we found that although *PIP1* genes were shown to facilitate CO_2 uptake and contribute to plant hydraulic conductance; under no water stress and under controlled greenhouse conditions they do not have a critical role for the well-being of poplar trees. It seemed that a strong reduction in *PIP1* gene expression has some effects on g_m but in a range where it has no physiologically meaningful effect on plant performance despite the fact that reduction of g_m should be most relevant under well watered conditions when stomata resistance is at its minimum (i.e., relative contribution of g_m to CO_2 flux is biggest since g_s is at its maximum). The same is true for the hydraulic path in the leaf where changes in leaf hydraulic resistance did not produce any detrimental effect on leaf physiology.

Thus, we can conclude that although *PIP1* genes were contributing to the mass transfer of water and CO_2 in *Populus tremula x alba* leaves, their down-regulation did not dramatically impair the physiological needs of this fast growing tree cultivated under a lack of stress. However, it is possible that *PIP1* aquaporins are relevant in the context of water stress via their role in the maintenance of plant water transport capacity (Secchi and Zwieniecki, 2010).

MATERIALS AND METHODS

PLANT GENOTYPES AND CONSTRUCT DEVELOPMENT

Hybrid white poplar *Populus tremula x Populus Alba* (INRA-France 717-1B4) was chosen for all experimental

procedures. An RNAi construct was used to down-regulate *PIP1* expression in poplar and was created in the pHannibal vector (Wesley et al., 2001). A *PIP1* cDNA fragment with introduced XhoI and KpnI restriction enzyme sites was amplified by PCR with oligonucleotides designed on conserved *PIP1* domains (listed in **Supplemental Table 1**) to facilitate directional cloning into the pHannibal vector. Primers were designed in order to down regulate more isoforms belonging to the same subfamily. A second set of PCR primers was used to amplify the same cDNA sequences but with BamHI and ClaI restriction sites to clone the same sequence in the reverse direction into the pHannibal vector. The final pHannibal vector containing both *PIP1* fragments was digested with NotI enzyme and cloned into NotI-digested pART27 binary vector (Gleave, 1992).

PLANT TRANSFORMATION

The vector harboring the insert was inserted into *Agrobacterium tumefaciens* C58/pMP90 (GV3101) strain through freeze/thaw shock transformation (Holsters et al., 1978). Shoot organogenesis was induced directly on mature leaf explants following the protocol by Meilan and Ma (Meilan and Ma, 2006). Briefly, leaf discs (4 mm in diameter) were incubated in *Agrobacterium* suspension with slow agitation for 1 h. The inoculated explants were co-cultivated in callus-induction medium (CIM) [MS supplemented with 10 μ M naphthaleneacetic acid (NAA) (Sigma, St. Louis, MO) and 5 μ M N6-(2-isopentenyl)adenine, (2iP) (Sigma)] at 22°C in the dark for 2 days. Explants were then washed five times in sterile deionized water and once in wash solution [1/2-strength MS medium, MS vitamins, 1 μ M NAA, 1 μ M 6-benzylaminopurine (BA; *PhytoTechnology* Lab., Shawnee Mission, KS), 1 μ M 2iP, 250 mg/L ascorbic acid, and 400 mg/L timentin]. For selection of transformed calli, explants were transferred to CIM containing 25 mg/l kanamycin and 200 mg/l timentin for 21 days. Shoots were induced by culturing explants on SIM medium [MS containing 0.2 μ M Thidiazuron (TDZ; *PhytoTechnology* Lab.), 100 mg/l kanamycin, and 200 mg/l timentin] sub-culturing every 3 week until shoots formed. For shoot elongation, explants were transferred onto SEM medium containing 0.1 μ M BA, 100 mg/l kanamycin, and 200 mg/l timentin. The regenerated shoots were rooted on half-strength MS medium supplemented 0.5 μ M indole-3-butyric acid (IBA; *PhytoTechnology* Lab.) and 25 mg/l kanamycin.

To ensure that transformation events were independent, a single clone per individual explant was selected for further propagation. Plantlets were confirmed as transgenic by PCR using *nptII* primers (**Supplemental Table 1**). Leaf genomic DNA isolation and PCR amplification were achieved using the Extraction-N-Amp Plant PCR Kits according to the manufacturer's instructions (Sigma, Saint Louis, MO, USA). After confirmation of transgene presence, the single plantlets were micro-propagated by repeated subcultural apical cutting on propagation media (half-strength MS medium with no antibiotics).

ESTIMATION OF AQUAPORIN EXPRESSION LEVELS IN TRANSGENIC EVENTS

Total RNA was isolated from leaf according to the protocol of Chang, (Chang et al., 1993). Contaminant genomic DNA was removed from the samples by digestion with RNase-free

DNase I (Fermentas), following the manufacturer's instructions. cDNA was synthesized using SuperScript II Reverse Transcriptase (Invitrogen) according to the supplier's instructions using oligo(dT)_{12–18} as a primer (Fermentas). Primers designed on *Populus trichocarpa* aquaporin sequences and previously reported (Secchi et al., 2009, 2011) were used to detect the expression level of PIP1.1; PIP1.3; PIP2.5, PIP2.6, and PIP2.8 genes in leaves of hybrid poplars. The oligonucleotide sequences are listed in **Supplemental Table 1**. Primers were tested on cDNA of hybrid poplar through PCR with RED Taq DNA Polymerase (Sigma) according to the manufacturer's instructions. The expected products were gel purified, inserted into the pGEM-T Easy vector (Promega) and sequenced using M13 forward and reverse primers. The isolated sequences were aligned with the corresponding sequences deposited on Genome Institute's (JGI) *P. trichocarpa* v.1.1 database, (ID numbers are provided in **Supplemental Table 1**) showing a nucleotide overall identity bigger than 95%. Gene transcript abundance was quantified with SYBR Green JumpStart Taq Ready Mix (Sigma) on an Eco Real-Time PCR System (Illumina, San Diego, USA). Thermocycler conditions for all real-time analyses were: 95°C for 5 min, followed by 40 cycles of 95°C for 15 s, 60°C for 30 s, and 72°C for 30 s.

Data were analyzed using Eco software (Illumina), and the expression values were normalized to the geometric mean of two housekeeping genes (ubiquitin and actin; **Supplemental Table 1**). These genes were found to have, for the same poplar species, the highest amplification efficiency and most stable expression across different tissues (Carraro et al., 2012). Real-time PCR was carried out using three biological replicates per transformed line. Two technical replicates were performed for each of the three biological replicates.

PLANT GROWTH AND PHYSIOLOGICAL MEASUREMENTS

Two months post-propagation plants (about 6 cm tall) were placed into moist potting mix in a 5.7 × 8.3-cm Rose Pot and transferred to a growth room (25 ± 1°C and 16-h photoperiod) for 3 weeks. After acclimation to ambient conditions, plants were transferred to a greenhouse and moved into 1-gallon pots filled with potting mix. Poplars were grown in a greenhouse for 4–6 months. Ambient conditions in the greenhouse were characterized by a temperature maintained in the range of 17–25°C, and the natural daylight was supplemented with light from metal halogen lamps (500–600 μ mol photons $m^{-2} s^{-1}$) to maintain a 12/12-h light/dark cycle. Plants were approximately 1.5 m tall at the onset of the experiments.

Six transgenic lines plus *wt* plants were chosen for analysis, each line contained from three to eight plants. Changes in chlorophyll concentration during the leaf maturation process was measured using SPAD meter (SPAD 502 Plus Chlorophyll Meter, Spectrum, Plainfield, IL, USA) on five consecutive leaves with the youngest being approximately 1 cm long. Measurements were repeated twice weekly for 8 weeks till no further increase in chlorophyll concentration was observed (SOM Graph). At that time leaves were fully expanded.

One of these mature leaves was then used for determination of gas exchange parameters. Measurements were made using

two LI-COR6400 gas exchange systems, one equipped with the fluorescence chamber (LI-COR, Lincoln, NE, USA). Before measurements, the leaf was acclimated in the gas exchange system at $400 \mu\text{mol mol}^{-1}$ reference CO_2 until stomata conductance was stable; this step took about 20 min. Light curves were measured in order to establish the appropriate photosynthetic photon flux density (PPFD) for use in CO_2 curve responses, 70% of saturating light i.e., $400 \mu\text{mol m}^{-2} \text{s}^{-1}$ as suggested by Gu et al. (2010). General measurement conditions were: PPFD, $400 \mu\text{mol m}^{-2} \text{s}^{-1}$ with 10% blue light component; leaf temperature, 25°C ; flow, $200 \mu\text{mol s}^{-1}$. For the response curves, CO_2 was reduced from 400 to $40 \mu\text{mol mol}^{-1}$ in decrements of $40 \mu\text{mol mol}^{-1}$. After a 9 min re-acclimation at $400 \mu\text{mol mol}^{-1}$ CO_2 was increased to 640 (increases of $40 \mu\text{mol mol}^{-1}$), and then to 700, 800, 900, 1000–2000. Gas exchange was measured at each CO_2 concentration after the cuvette CO_2 concentration was stable for 240 s. All measurements were empirically corrected for leaks by evaluating dry poplar leaves under measurement conditions. Standard chlorophyll fluorescence measurement (quantum efficiency of PSII) was made on half of the leaves after recording each gas exchange variable. The fluorescence data were used to better estimate the limiting factors of the curve, and the parameters chosen to fit it were then used for fitting the curve with data collected from gas exchange system (All CO_2 curve responses are available in SOM materials).

Collected data were then used to estimate A/C_i curve parameters using a previously published fitting procedure (Sharkey et al., 2007). Leaves used for CO_2 responses were then collected and cut in two along the mid-vein so all leaf parts (tip and base) was included in further analysis; half the leaf was immediately frozen in liquid nitrogen and kept at -80°C until molecular analyses were performed, the other half was dried in an oven and used for isotope analyses.

REHYDRATION KINETICS

Leaf rehydration kinetic analysis followed the procedure described in an earlier study (Zwieniecki et al., 2007). The “reverse Polish guillotine” was redesigned such that it allowed cutting of a leaf directly off the tree while simultaneously connecting its petiole to water-filled tube in one single motion. Only mature, fully expanded leaves were selected for rehydration measurements. Trees were well-watered with an average water potential below -0.5 MPa . A small portion of the target leaf petiole was secured in the guillotine holder using foam. A layer of high vacuum silicon grease (Dow Corning, Midland, MI, USA) was added to the petiole to help form a seal with the o-ring. Once system was ready for the cut, water was allowed to flow from the tube such that petiole cut was made in wet conditions. After the entire setup was prepared, the guillotine was deployed such that the leaf was severed from the branch and simultaneously connected to a tube linked to a water reservoir seated on an analytic balance (Sartorius 0.01 mg, Gottingen, Germany). Immediately after the cut, the leaf was placed under water at 20°C . Water uptake was recorded at 1 s intervals for ~ 1000 [s]. 4–12 leaves per transgenic line were measured for a total of 74 leaves. Leaves

were illuminated with $30 \mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD throughout the measurement period, a rate sufficient to saturate any light-induced changes in leaf hydraulic conductance using this measurement technique (Rockwell et al., 2011). After accounting for the evaporation from the measurement system itself, a linear uptake was still observed in all leaves. This residual uptake, which was due to capillary infiltration into leaf intercellular spaces, was also accounted for before further analysis by subtracting it from the generated curve (Zwieniecki et al., 2007).

The weight–time function of water flow from the balance provides a continuous record of leaf rehydration. Following the analysis of a previous study used a double exponential model:

$$f(t) = a * (1 - e^{-1/b * t}) + c * (1 - e^{-1/d * t}) \quad (1)$$

to describe water uptake by dehydrated leaves. In this model, parameters a and c describe the volume of the respective two rehydrating phases and b , d their corresponding time constants.. The phases were previously associated with two potential leaf compartments characterized by two different rates of rehydration (Zwieniecki et al., 2007). SLA of each leaf used to determine rehydration kinetic was calculated from dry weight and leaf area as $\text{SLA} = \text{leaf area/dry weight}$.

ISOTOPES ABUNDANCE ANALYSIS

Isotope analysis was performed in the UC Davis isotope facility. Samples were selected from mid part of the leaf with attention to avoid collecting low order veins from several mature leaves used in the photosynthesis measurements. Samples were then dried at 65°C for several days. Dry material was prepared following online protocol provided by the UC Davis isotope facility (<http://stableisotopefacility.ucdavis.edu/13cand15n.html>). Briefly leaf tissue was grounded to powder using pestle and mortar. 3–4 mg of material was weighed and packed in aluminum caps and sent for concentration analysis of ^{12}C , ^{13}C , and ^{14}N and ^{15}N .

STATISTICAL ANALYSIS

All statistical analysis was performed using Statistica (SoftStat, Inc.).

ACKNOWLEDGMENTS

We would like to thank CSIRO for providing the pHannibal vector. The authors also thank Nicola Carraro, Irene Perrone, Tracy Tisdale-Orr and Rachel Spicer for technical support, helpful discussion and access to Agrobacterium strains. Matthew Gilbert provided extended help in collecting and discussion of photosynthetic data as well editorial help to the manuscript. This work was supported by National Science Foundation Award No. IOS-0919729.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://www.frontiersin.org/journal/10.3389/fpls.2013.00507/abstract>

Supplemental File 1 | Raw data used for the CO₂ curve-fitting method in the format used by Sharkey et al. (2007)

Supplemental Figure 1 | Temporal changes of chlorophyll content in wild-type (black circles and line) and in transgenic plants. Measurements were repeated in the greenhouse twice weekly for 8 weeks till no further increase in chlorophyll concentration was observed. Data were collected on five consecutive leaves; here we reported values from the youngest leaf being approximately 1 cm long (**A**) and the oldest one (**B**). Data are means of all available plants from transgenic lines, the error bars represent *SD*.

Supplemental Table 1 | Sequences of primers used for quantitative real time PCR.

REFERENCES

- Almeida-Rodriguez, A. M., Cooke, J. E. K., Yeh, F., and Zwiazek, J. J. (2010). Functional characterization of drought-responsive aquaporins in *Populus balsamifera* and *Populus simonii x balsamifera* clones with different drought resistance strategies. *Physiol. Plant.* 140, 321–333. doi: 10.1111/j.1399-3054.2010.01405.x
- Alves, G., Sauter, J. J., Julien, J. L., Fleurat-Lessard, P., Ameglio, T., Guillot, A., et al. (2011). Plasma membrane H(+)-ATPase, succinate and isocitrate dehydrogenases activities of vessel-associated cells in walnut trees. *J. Plant Physiol.* 158, 1263–1271. doi: 10.1078/0176-1617-00503
- Barone, L., Shih, C., and Wasserman, P. (1997). Mercury-induced conformational changes and identification of conserved surface loops in plasma membrane aquaporins from higher plants. *J. Biol. Chem.* 272, 30672–30677. doi: 10.1074/jbc.272.49.30672
- Bickford, C. P., Hanson, D. T., and McDowell, N. G. (2010). Influence of diurnal variation in mesophyll conductance on modelled C-13 discrimination: results from a field study. *J. Exp. Bot.* 61, 3223–3233. doi: 10.1093/jxb/erq137
- Bloom, A. J., Burger, M., Rubio-Asensio, J. S., and Cousins, A. B. (2010). Carbon dioxide enrichment inhibits nitrate assimilation in wheat and arabidopsis. *Science* 328, 899–903. doi: 10.1126/science.1186440
- Brodribb, T. J., Holbrook, N. M., Zwieniecki, M. A., and Palma, B. (2005). Leaf hydraulic capacity in ferns, conifers and angiosperms: impacts on photosynthetic maxima. *New Phytol.* 165, 839–846. doi: 10.1111/j.1469-8137.2004.01259.x
- Brueck, H. (2008). Effects of nitrogen supply on water-use efficiency of higher plants. *J. Plant Nutr.* 31, 210–219. doi: 10.1002/jpln.200700080
- Carraro, N., Tisdale-Orr, T. E., Clouse, R. M., Knoller, A. S., and Spicer, R. (2012). Diversification and Expression of the PIN, AUX/LAX, and ABCB Families of Putative Auxin Transporters in *Populus*. *Front. Plant Sci.* 3:17. doi: 10.3389/fpls.2012.00017
- Chang, S., Puryear, J., and Cairney, J. (1993). A simple and efficient method for isolating RNA from pine tree. *Plant Mol. Biol. Rep.* 11, 113–116. doi: 10.1007/BF02670468
- Chrispeels, M. J., Morillon, R., Maurel, C., Gerbeau, P., Kjellbom, P., and Johansson, I. (2001). Aquaporins of plants: structure, function, regulation, and role in plant water relations. *Curr. Top. Membr.* 51, 277–334. doi: 10.1016/S1063-5823(01)51009-1
- Fetter, K., van Wilder, V., Moshelion, M., and Chaumont, F. (2004). Interaction between plasma membrane aquaporins modulate their water channel activity. *Plant Cell* 16, 215–228. doi: 10.1105/tpc.017194
- Flexas, J., Ribas-Carbo, M., Hanson, D. T., Bota, J., Otto, B., Cifre, J., et al. (2006). Tobacco aquaporin NTAQP1 is involved in mesophyll conductance to CO₂ in vivo. *Plant J.* 48, 427–439. doi: 10.1111/j.1365-313X.2006.02879.x
- Frick, A., Järvå, M., and Törnroth-Horsefield, S. (2013). Structural basis for pH gating of plant aquaporins. *FEBS Lett.* 7, 989–993. doi: 10.1016/j.febslet.2013.02.038
- Gilbert, M. E., Zwieniecki, M. A., and Holbrook, N. M. (2011). Independent variation in photosynthetic capacity and stomatal conductance leads to differences in intrinsic water use efficiency in 11 soybean genotypes before and during mild drought. *J. Exp. Bot.* 62, 2875–2887. doi: 10.1093/jxb/erq461
- Gleave, A. P. (1992). A versatile binary vector system with a T-DNA organizational-structure conducive to efficient integration of cloned DNA into the plant genome. *Plant Mol. Biol.* 20, 1203–1207. doi: 10.1007/BF00028910
- Gu, L. H., Pallardy, S. G., Tu, K., Law, B. E., and Wullschlegel, S. D. (2010). Reliable estimation of biochemical parameters from C-3 leaf photosynthesis-intercellular carbon dioxide response curves. *Plant Cell Environ.* 33, 1852–1874. doi: 10.1111/j.1365-3040.2010.02192.x
- Hanba, Y. T., Shibusaka, M., Hayashi, Y., Hayakawa, T., Kasamo, K., Terashima, I., et al. (2004). Overexpression of the barley aquaporin HvPIP2;1 increases internal CO₂ conductance and CO₂ assimilation in the leaves of transgenic rice plants. *Plant Cell Physiol.* 45, 521–529. doi: 10.1093/pcp/pch070
- Heckwolf, M., Pater, D., Hanson, D. T., and Kaldenhoff, R. (2011). The *Arabidopsis thaliana* aquaporin AtPIP1;2 is a physiologically relevant CO₂ transport facilitator. *Plant J.* 67, 795–804. doi: 10.1111/j.1365-313X.2011.04634.x
- Heinen, R. B., Ye, Q., and Chaumont, F. (2009). Role of aquaporins in leaf physiology. *J. Exp. Bot.* 60, 2971–2985. doi: 10.1093/jxb/erp171
- Holsters, M., Dewaele, D., Depicker, A., Messens, E., Vanmontagu, M., and Schell, J. (1978). Transfection and transformation of *Agrobacterium tumefaciens*. *Mol. Gen. Genet.* 163, 181–187. doi: 10.1007/BF00267408
- Hukin, D., Doering-Saad, C., Thomas, C., and Pritchard, J. (2002). Sensitivity of cell hydraulic conductivity to mercury is coincident with symplasmic isolation and expression of plasmalemma aquaporin genes in growing maize roots. *Planta* 215, 1047–1056. doi: 10.1007/s00425-002-0841-2
- Isebrands, J. G., and Richardson, J. (2013). *Poplars and Willows: Trees for Society and The Environment*. Boston, MA; Rome: CABI: FAO.
- Martinez-Ballesta, M. C., Martinez, V., and Carvajal, M. (2003). Aquaporin functionality in relation to H⁺-ATPase activity in root cells of *Capsicum annuum* grown under salinity. *Physiol. Plant.* 117, 413–420. doi: 10.1034/j.1399-3054.2003.00044.x
- Meilan, R., and Ma, C. (2006). Poplar (*Populus* spp.). *Methods Mol. Biol.* 344, 143–151. doi: 10.1385/1-59745-131-2:143
- Niemietz, C. M., and Tyerman, S. D. (2002). New potent inhibitors of aquaporins: silver and gold compounds inhibit aquaporins of plant and human origin. *FEBS Lett.* 531, 443–447. doi: 10.1016/S0014-5793(02)03581-0
- Niinemetts, U. (1997). Role of foliar nitrogen in light harvesting and shade tolerance of four temperate deciduous woody species. *Funct. Ecol.* 11, 518–531. doi: 10.1046/j.1365-2435.1997.00109.x
- Perrone, I., Gambino, G., Chitarra, W., Vitali, M., Pagliarini, C., Riccomagno, N., et al. (2012). The grapevine root-specific aquaporin VvPIP2;4N controls root hydraulic conductance and leaf gas exchange under well-watered conditions but not under water stress. *Plant Physiol.* 160, 965–977. doi: 10.1104/pp.112.203455
- Rockwell, F. E., Holbrook, N. M., and Zwieniecki, M. A. (2011). Hydraulic conductivity of red oak (*Quercus rubra* L.) leaf tissue does not respond to light. *Plant Cell Environ.* 34, 565–579. doi: 10.1111/j.1365-3040.2011.02263.x
- Secchi, F., Gilbert, M. E., and Zwieniecki, M. A. (2011). Transcriptome response to embolism formation in stems of *Populus trichocarpa* provides insight into signaling and the biology of refilling. *Plant Physiol.* 157, 1419–1429. doi: 10.1104/pp.111.185124
- Secchi, F., Matic, B., Zeidel, M. L., and Zwieniecki, M. A. (2009). Functional analysis of putative genes encoding the PIP2 water channel subfamily in *Populus trichocarpa*. *Tree Physiol.* 29, 1467–1477. doi: 10.1093/treephys/tp060
- Secchi, F., and Zwieniecki, M. A. (2010). Patterns of PIP gene expression in *Populus trichocarpa* during recovery from xylem embolism suggest a major role for the PIP1 aquaporin subfamily as moderators of refilling process. *Plant Cell Environ.* 33, 1285–1297. doi: 10.1111/j.1365-3040.2010.02147.x
- Sharkey, T. D., Bernacchi, C. J., Farquhar, G. D., and Singaas, E. L. (2007). Fitting photosynthetic carbon dioxide response curves for C-3 leaves. *Plant Cell Environ.* 30, 1035–1040. doi: 10.1111/j.1365-3040.2007.01710.x
- Siefridt, F., Biela, A., Eckert, M., Otto, B., Uehlein, N., and Kaldenhoff, R. (2001). The tobacco plasma membrane aquaporin NtAQP1. *J. Exp. Bot.* 52, 1953–1957. doi: 10.1093/jexbot/52.363.1953
- Siefridt, F., Tyree, M. T., Lovisolo, C., Schubert, A., and Kaldenhoff, R. (2002). PIP1 plasma membrane aquaporins in tobacco: from cellular effects to function in plants. *Plant Cell* 14, 869–876. doi: 10.1105/tpc.000901
- Silim, S. N., Ryan, N., and Kubien, D. S. (2010). Temperature responses of photosynthesis and respiration in *Populus balsamifera* L.: acclimation vs. adaptation. *Photosynth. Res.* 104, 19–30. doi: 10.1007/s11120-010-9527-y
- Temmei, Y., Uchida, S., Hoshino, D., Kanzawa, N., Kuwahara, K., Sasaki, S., et al. (2005). Water channel activities of *Mimosa pudica* plasma membrane intrinsic proteins are regulated by direct interaction and phosphorylation. *FEBS Lett.* 579, 4417–4422. doi: 10.1016/j.febslet.2005.06.082

- Terashima, I., and Ono, K. (2002). Effects of HgCl₂ on CO₂ dependence of leaf photosynthesis: evidence indicating involvement of aquaporins in CO₂ diffusion across the plasma membrane. *Plant Cell Physiol.* 43, 70–78. doi: 10.1093/pcp/pcf001
- Tournaire-Roux, C., Sutka, M., Javot, H., Gout, E., Gerbeau, P., Luu, D. T., et al. (2003). Cytosolic pH regulates root water transport during anoxic stress through gating of aquaporins. *Nature* 425, 393–397. doi: 10.1038/nature01853
- Uehlein, N., Lovisolo, C., Siefritz, F., and Kaldenhoff, R. (2003). The tobacco aquaporin NtAQP1 is a membrane CO₂ pore with physiological functions. *Nature* 425, 734–737. doi: 10.1038/nature02027
- Uehlein, N., Otto, B., Hanson, D. T., Fischer, M., McDowell, N., and Kaldenhoff, R. (2008). Function of *Nicotiana tabacum* aquaporins as chloroplast gas pores challenges the concept of membrane CO₂ permeability. *Plant Cell* 20, 648–657. doi: 10.1105/tpc.107.054023
- Uehlein, N., Sperling, H., Heckwolf, M., and Kaldenhoff, R. (2012). The *Arabidopsis* aquaporin PIP1;2 rules cellular CO₂ uptake. *Plant Cell Environ.* 35, 1077–1083. doi: 10.1111/j.1365-3040.2011.02473.x
- Wesley, S. V., Helliwell, C. A., Smith, N. A., Wang, M. B., Rouse, D. T., Liu, Q., et al. (2001). Construct design for efficient, effective and high-throughput gene silencing in plants. *Plant J.* 27, 581–590. doi: 10.1046/j.1365-313X.2001.01105.x
- Zhu, G. F., Li, X., Su, Y. H., Lu, L., and Huang, C. L. (2011). Seasonal fluctuations and temperature dependence in photosynthetic parameters and stomatal conductance at the leaf scale of *Populus euphratica* Oliv. *Tree Physiol.* 31, 178–195. doi: 10.1093/treephys/tptr005
- Zwieniecki, M. A., Brodribb, T. J., and Holbrook, N. M. (2007). Hydraulic design of leaves: insights from rehydration kinetics. *Plant Cell Environ.* 30, 910–921. doi: 10.1111/j.1365-3040.2007.001681.x
- Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
- Received: 11 June 2013; accepted: 26 November 2013; published online: 13 December 2013.
- Citation: Secchi F and Zwieniecki MA (2013) The physiological response of *Populus tremula x alba* leaves to the down-regulation of PIP1 aquaporin gene expression under no water stress. *Front. Plant Sci.* 4:507. doi: 10.3389/fpls.2013.00507
- This article was submitted to *Plant Physiology*, a section of the journal *Frontiers in Plant Science*.
- Copyright © 2013 Secchi and Zwieniecki. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.