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Lasers, stem cells, and COPD

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Journal

Journal of Translational Medicine, 8(1)

ISSN

1479-5876

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Publication Date

2010-02-16

DOI

http://dx.doi.org/10.1186/1479-5876-8-16

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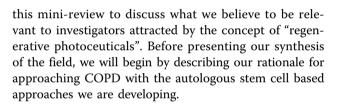
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Abstract

The medical use of low level laser (LLL) irradiation has been occurring for decades, primarily in the area of tissue healing and inflammatory conditions. Despite little mechanistic knowledge, the concept of a non-invasive, non-thermal intervention that has the potential to modulate regenerative processes is worthy of attention when searching for novel methods of augmenting stem cell-based therapies. Here we discuss the use of LLL irradiation as a "photoceutical" for enhancing production of stem cell growth/chemoattractant factors, stimulation of angiogenesis, and directly augmenting proliferation of stem cells. The combination of LLL together with allogeneic and autologous stem cells, as well as post-mobilization directing of stem cells will be discussed.

Introduction (Personal Perspective)

We came upon the field of low level laser (LLL) therapy by accident. One of our advisors read a press release about a company using this novel technology of specific light wavelengths to treat stroke. Given the possible role of stem cells in post-stroke regeneration, we decided to cautiously investigate. As a background, it should be said that our scientific team has been focusing on the area of cord blood banking and manufacturing of disposables for processing of adipose stem cells for the past 3 years. Our board has been interested in strategically refocusing the company from services-oriented into a more research-focused model. An unbiased exploration into the various degenerative conditions that may be addressed by our existing know-how led us to explore the condition of chronic obstructive pulmonary disease (COPD), an umbrella term covering chronic bronchitis and emphysema, which is the 4th largest cause of death in the United States. As a means of increasing our probability of success in treatment of this condition, the decision was made to develop an adjuvant therapy that would augment stem cell activity. The field of LLL therapy attracted us because it appeared to be relatively unexplored scientific territory for which large amounts of clinical experience exist. Unfortunately, it was difficult to obtain the cohesive "state-of-the-art" description of the molecular/cellular mechanisms of this therapy in reviews that we have searched. Therefore we sought in



COPD as an Indication for Stem Cell Therapy

COPD possesses several features making it ideal for stem cell based interventions: a) the quality of life and lack of progress demands the ethical exploration of novel approaches. For example, bone marrow stem cells have been used in over a thousand cardiac patients with some indication of efficacy [1,2]. Adipose-based stem cell therapies have been successfully used in thousands of race-horses and companion animals without adverse effects [3], as well as numerous clinical trials are ongoing and published human data reports no adverse effects (reviewed in ref [4]). Unfortunately, evaluation of stem cell therapy in COPD has lagged behind other areas of regenerative investigation; b) the underlying cause of COPD appears to be inflammatory and/or immunologically mediated. The destruction of alveolar tissue is associated with T cell reactivity [5,6], pathological pulmonary macrophage activation [7], and auto-antibody production [8]. Mesenchymal stem cells have been demonstrated to potently suppress autoreactive T cells [9,10], inhibit macrophage activation [11], and autoantibody responses [12]. Additionally, mesenchymal stem cells can be purified in high concentrations from adipose stromal vascular tissue together with high



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concentrations of T regulatory cells [4], which in animal models are approximately 100 more potent than peripheral T cells at secreting cytokines therapeutic for COPD such as IL-10 [13,14]. Additionally, use of adipose derived cells has yielded promising clinical results in autoimmune conditions such as multiple sclerosis [4]; and c) Pulmonary stem cells capable of regenerating damaged parenchymal tissue have been reported [15]. Administration of mesenchymal stem cells into neonatal oxygen-damaged lungs, which results in COPD-like alveoli dysplasia, has been demonstrated to yield improvements in two recent publications [16,17].

Based on the above rationale for stem cell-based COPD treatments, we began our exploration into this area by performing several preliminary experiments and filing patents covering combination uses of stem cells with various pharmacologically available antiinflammatories, as well as methods of immune modulation. These have served as the basis for two of our pipeline candidates, ENT-111, and ENT-894. As a commerciallyoriented organization, we needed to develop a therapeutic candidate that not only has a great potential for efficacy, but also can be easily implemented as part of the standard of care. Our search led us to the area of low level laser (LLL) therapy. From our initial perception as neophytes to this field, the area of LLL therapy has been somewhat of a medical mystery. A pubmed search for "low level laser therapy" yields more than 1700 results, yet before stumbling across this concept, none of us, or our advisors, have ever heard of this area of medicine.

On face value, this field appeared to be somewhat of a panacea: clinical trials claiming efficacy for conditions ranging from alcoholism [18], to sinusitis [19], to ischemic heart disease [20]. Further confusing was that many of the studies used different types of LLL-generating devices, with different parameters, in different model systems, making comparison of data almost impossible. Despite this initial impression, the possibility that a simple, non-invasive methodology could exist that augments regenerative potential in a tissue-focused manner became very enticing to us. Specific uses envisioned, for which intellectual property was filed included using light to concentrate stem cells to an area of need, to modulate effects of stem cells once they are in that specific area, or even to use light together with other agents to modulate endogenous stem cells.

The purpose of the current manuscript is to overview some of the previous work performed in this area that was of great interest to our ongoing work in regenerative medicine. We believe that greater integration of the area of LLL with current advancements in molecular and cellular biology will accelerate medical progress. Unfortunately, in our impression to date, this has been a very slow process.

What is Low Level Laser Irradiation?

Lasers (Light amplification by stimulated emission of radiation) are devices that typically generate electromagnetic radiation which is relatively uniform in wavelength, phase, and polarization, originally described by Theodore Maiman in 1960 in the form of a ruby laser [21]. These properties have allowed for numerous medical applications including uses in surgery, activation of photodynamic agents, and various ablative therapies in cosmetics that are based on heat/tissue destruction generated by the laser beam [22-24]. These applications of lasers are considered "high energy" because of their intensity, which ranges from about 10-100 Watts. The subject of the current paper will be another type of laser approach called low level lasers (LLL) that elicits effects through non-thermal means. This area of investigation started with the work of Mester et al who in 1967 reported non-thermal effects of lasers on mouse hair growth [25]. In a subsequent study [26], the same group reported acceleration of wound healing and improvement in regenerative ability of muscle fibers post wounding using a 1 J/cm² ruby laser. Since those early days, numerous in vitro and in vivo studies have been reported demonstrating a wide variety of therapeutic effects involving LLL, a selected sample of which will be discussed below. In order to narrow our focus of discussion, it is important to first begin by establishing the current definition of LLL therapy. According to Posten et al [27], there are several parameters of importance: a) Power output of laser being 10⁻³ to 10⁻¹ Watts; b) Wavelength in the range of 300-10,600 nm; c) Pulse rate from 0, meaning continuous to 5000 Hertz (cycles per second); d) intensity of 10⁻²-10 W/cm(2) and dose of 0.01 to 100 J/ cm². Most common methods of administering LLL radiation include lasers such as ruby (694 nm), Ar (488 and 514 nm), He-Ne (632.8 nm), Krypton (521, 530, 568, and 647 nm), Ga-Al-As (805 or 650 nm), and Ga-As (904 nm). Perhaps one of the most distinguishing features of LLL therapy as compared to other photoceutical modalities is that effects are mediated not through induction of thermal effects but rather through a process that is still not clearly defined called "photobiostimulation". It appears that this effect of LLL is not depend on coherence, and therefore allows for use of non-laser light generating devices such as inexpensive Light Emitting Diode (LED) technology [28].

To date several mechanisms of biological action have been proposed, although none are clearly established. These include augmentation of cellular ATP levels [29], manipulation of inducible nitric oxide synthase (iNOS) activity [30,31], suppression of inflammatory cytokines such as TNF-alpha, IL-1beta, IL-6 and IL-8 [32-36], upregulation of growth factor production such as PDGF, IGF-1, NGF and FGF-2 [36-39], alteration of mitochondrial membrane potential [29,40-42] due to chromophores found in the mitochondrial respiratory chain [43,44] as reviewed in [45], stimulation of protein kinase C (PKC) activation [46], manipulation of NF- κ B activation [47], direct bacteriotoxic effect mediated by induction of reactive oxygen species (ROS) [48], modification of extracellular matrix components [49], inhibition of apoptosis [29], stimulation of mast cell degranulation [50], and upregulation of heat shock proteins [51]. Unfortunately these effects have been demonstrated using a variety of LLL devices in noncomparable models. To add to confusion, dose-dependency seems to be confined to such a narrow range or does not seem to exist in that numerous systems therapeutic effects disappear with increased dose.

In vitro studies of LLL

In areas of potential phenomenology, it is important to begin by assessing in vitro studies reported in the literature in which reproducibility can be attained with some degree of confidence, and mechanistic dissection is simpler as compared with in vivo systems. In 1983, one of the first studies to demonstrate in vitro effects of LLL was published. The investigators used a helium neon (He-Ne) laser to generate a visible red light at 632.8 nm for treatment of porcine granulosa cells. The paper described upregulation of metabolic and hormone-producing activity of the cells when exposed for 60 seconds to pulsating low power (2.8 mW) irradiation [52]. The possibility of modulating biologically-relevant signaling proteins by LLL was further assessed in a study using an energy dose of 1.5 J/cm² in cultured keratinocytes. Administration of He-Ne laser emitted light resulted in upregulated gene expression of IL-1 and IL-8 [53]. Production of various growth factors in vitro suggests the possibility of enhanced cellular mitogenesis and mobility as a result of LLL treatment. Using a diode-based method to generate a similar wavelength to the He-Ne laser (363 nm), Mvula et al reported in two papers that irradiation at 5 J/cm² of adipose derived mesenchymal stem cells resulted in enhanced proliferation, viability and expression of the adhesion molecule beta-1 integrin as compared to control [54,55]. In agreement with possible regenerative activity based on activation of stem cells, other studies have used an in vitro injury model to examine possible therapeutic effects. Migration of fibroblasts was demonstrated to be enhanced in a "wound assay" in which cell monolayers are scraped with a pipette tip and amount of time needed to restore the monolayer is used as an indicator of "healing". The cells exposed to 5 J/cm² generated by an He-Ne laser migrated rapidly across the wound margin indicating a stimulatory or positive influence of phototherapy. Higher doses (10 and 16 J/cm²) caused a decrease in cell viability and proliferation with a significant amount of damage to the cell membrane and DNA [56]. In order to examine whether LLL may positively affect healing under non-optimal conditions that mimic clinical situations treatment of fibroblasts from diabetic animals was performed. It was demonstrated that with the He-Ne laser dosage of 5 J/cm² fibroblasts exhibited an enhanced migration activity, however at 16 J/cm² activity was negated and cellular damage observed [57]. Thus from these studies it appears that energy doses from 1.5 J/cm² to 5 J/cm² are capable of eliciting "biostimulatory effects" in vitro in the He-Ne-based laser for adherent cells that may be useful in regeneration such as fibroblasts and mesenchymal stem cells.

Studies have also been performed in vitro on immunological cells. High intensity He-Ne irradiation at 28 and 112 J/cm² of human peripheral blood mononuclear cells, a heterogeneous population of T cells, B cells, NK cells, and monocytes has been described to induce chromatin relaxation and to augment proliferative response to the T cell mitogen phytohemaglutin [58]. In human peripheral blood mononuclear cells (PBMC), another group reported in two papers that interleukin-1 alpha (IL-1 alpha), tumor necrosis factor-alpha (TNF-alpha), interleukin-2 (IL-2), and interferon-gamma (IFNgamma) at a protein and gene level in PBMC was increased after He-Ne irradiation at 18.9 J/cm² and decreased with 37.8 J/cm² [59,60]. Stimulation of human PBMC proliferation and murine splenic lymphocytes was also reported with He-Ne LLL [61,62]. In terms of innate immune cells, enhanced phagocytic activity of murine macrophages have been reported with energy densities ranging from 100 to 600 J/cm², with an optimal dose of 200 J/cm² [63]. Furthermore, LLL has been demonstrated to augment human monocyte killing mycobacterial cells at similar densities, providing a functional correlation [64].

Thus from the selected in vitro studies discussed, it appears that modulation of proliferation and soluble factor production by LLL can be reliably reproduced. However the data may be to some extent contradictory. For example, the over-arching clinical rationale for use of LLL in conditions such as sinusitis [65], arthritis [66,67], or wound healing [68] is that treatment is associated with anti-inflammatory effects. However the in vitro studies described above suggested LLL stimulates proinflammatory agents such as TNF-alpha or IL-1 [59,60]. This suggests the in vivo effects of LLL may be very complex, which to some extent should not be surprising. Factors affecting LLL in vivo actions would include degree of energy penetration through the tissue, the various absorption ability of cells in the various tissues, and complex chemical changes that maybe occurring in paracrine/autocrine manner. Perhaps an analogy to the possible discrepancy between LLL effects in vitro versus

in vivo may be made with the medical practice of extracorporeal ozonation of blood. This practice is similar to LLL therapy given that it is used in treatment of conditions such as atherosclerosis, non-healing ulcers, and various degenerative conditions, despite no clear mechanistic understanding [69-71]. In vitro studies have demonstrated that ozone is a potent oxidant and inducer of cell apoptosis and inflammatory signaling [72-74]. In contrast, in vivo systemic changes subsequent to administration of ozone or ozonized blood in animal models and patients are quite the opposite. Numerous investigators have published enhanced anti-oxidant enzyme activity such as elevations in Mg-SOD and glutathione-peroxidase levels, as well as diminishment of inflammation-associated pathology [75-78]. Regardless of the complexity of in vivo situations, the fact that reproducible, in vitro experiments, demonstrate a biological effect provided support for us that there is some basis for LLL and it is not strictly an area of phenomenology.

Animal Studies with LLL

As early as 1983, Surinchak et al reported in a rat skin incision healing model that wounds exposed He-Ne radiation of fluency 2.2 J/cm² for 3 min twice daily for 14 days demonstrated a 55% increase in breaking strength over control rats. Interestingly, higher doses yielded poorer healing [79]. This application of laser light was performed directly on shaved skin. In a contradictory experiment, it was reported that rats irradiated for 12 days with four levels of laser light (0.0, 0.47, 0.93, and 1.73 J/cm²) a possible strengthening of wounds tension was observed at the highest levels of irradiation (1.73 J/cm^2) , however it did not reach significance when analyzed by resampling statistics [80]. In another wound-healing study Ghamsari et al reported accelerated healing in the cranial surface of teats in dairy cows by administration of He-Ne irradiation at 3.64 J/cm² dose of low-level laser, using a helium-neon system with an output of 8.5 mW, continuous wave [81]. Collagen fibers in LLL groups were denser, thicker, better arranged and more continuous with existing collagen fibers than those in non-LLL groups. The mean tensile strength was significantly greater in LLL groups than in non-LLL groups [82]. In the random skin flap model, the use of He-Ne laser irradiation with 3 J/cm² energy density immediately after the surgery and for the four subsequent days was evaluated in 4 experimental groups: Group 1 (control) sham irradiation with He-Ne laser; Group 2 irradiation by punctual contact technique on the skin flap surface; Group 3 laser irradiation surrounding the skin flap; and Group 4 laser irradiation both on the skin flap surface and around it. The percentage of necrotic area of the four groups was determined on day 7-post injury. The control group had an average necrotic area of 48.86%; the group irradiated on the skin flap surface alone had 38.67%; the group irradiated around the skin flap had 35.34%; and the group irradiated one the skin flap surface and around it had 22.61%. All experimental groups reached statistically significant values when compared to control [83]. Quite striking results were obtained in an alloxan-induced diabetes wound healing model in which a circular 4 cm² excisional wound was created on the dorsum of the diabetic rats. Treatment with He-Ne irradiation at 4.8 J/ cm^2 was performed 5 days a week until the wound healed completely and compared to sham irradiated animals. The laser-treated group healed on average by the 18th day whereas, the control group healed on average by the 59th day [84].

In addition to mechanically-induced wounds, beneficial effects of LLL have been obtained in burn-wounds in which deep second-degree burn wounds were induced in rats and the effects of daily He-Ne irradiation at 1.2 and 2.4 J/cm² were assessed in comparison to 0.2% nitrofurazone cream. The number of macrophages at day 16, and the depth of new epidermis at day 30, was significantly less in the laser treated groups in comparison with control and nitrofurazone treated groups. Additionally, infections with S. epidermidis and S. aureus were significantly reduced [85].

While numerous studies have examined dermatological applications of LLL, which may conceptually be easier to perform due to ability to topically apply light, extensive investigation has also been made in the area of orthopedic applications. Healing acceleration has been observed in regeneration of the rat mid-cortical diaphysis of the tibiae, which is a model of post-injury bone healing. A small hole was surgically made with a dentistry burr in the tibia and the injured area and LLL was administered over a 7 or 14 day course transcutaneously starting 24 h from surgery. Incident energy density dosages of 31.5 and 94.5 J/cm² were applied during the period of the tibia wound healing. Increased angiogenesis was observed after 7 days irradiation at an energy density of 94.5 J/cm², but significantly decreased the number of vessels in the 14-day irradiated tibiae, independent of the dosage [86]. In an osteoarthritis model treatment with He-Ne resulted in augmentation of heat shock proteins and pathohistological improvement of arthritic cartilage [87]. The possibility that a type of preconditioning response is occurring, which would involve induction of genes such as hemoxygenase-1 [88], remains to be investigated. Effects of LLL therapy on articular cartilage were confirmed by another group. The experiment consisted of 42 young Wistar rats whose hind limbs were operated on in order to immobilize the knee joint. One week after operation they were assigned to three groups; irradiance 3.9 W/ cm2, 5.8 W/cm^2 , and sham treatment. After 6 times of treatment for another 2 weeks significant preservation of articular cartilage stiffness with 3.9 and 5.8 W/cm^2 therapy was observed [89].

Muscle regeneration by LLL was demonstrated in a rat model of disuse atrophy in which eight-week-old rats were subjected to hindlimb suspension for 2 weeks, after which they were released and recovered. During the recovery period, rats underwent daily LLL irradiation (Ga-Al-As laser; 830 nm; 60 mW; total, 180 s) to the right gastrocnemius muscle through the skin. After 2-weeks the number of capillaries and fibroblast growth factor levels exhibited significant elevation relative to those of the LLL-untreated muscles. LLL treatment induced proliferation in satellite cells as detected by BRdU [90].

Other animal studies of LLL have demonstrated effects in areas that appear unrelated such as suppression of snake venom induced muscle death [91], decreasing histamine-induced vasospasms [92], inhibition of post-injury restenosis [93], and immune stimulation by thymic irradiation [94].

Clinical Studies Using LLL

Growth factor secretion by LLL and its apparent regenerative activities have stimulated studies in radiationinduced mucositis. A 30 patient randomized trial of carcinoma patients treated by radiotherapy alone (65 Gy at a rate of 2 Gy/fraction, 5 fractions per week) without prior surgery or concomitant chemotherapy suffering from radiation-induced mucositis was performed using a He-Ne 60 mW laser. Grade 3 mucositis occured with a frequency of 35.2% in controls and at 7.6% of treated patients. Furthermore, a decrease in "severe pain" (grade 3) was observed in that 23.8% in the control group experienced this level of pain, as compared to 1.9% in the treatment group [95]. A subsequent study reported similar effects [96].

Healing ability of lasers was also observed in a study of patients with gingival flap incisions. Fifty-eight extraction patients had one of two gingival flap incisions lased with a 1.4 mW He-Ne (670 nm) at 0.34 J/cm². Healing rates were evaluated clinically and photographically. Sixty-nine percent of the irradiated incisions healed faster than the control incisions. No significant difference in healing was noted when patients were compared by age, gender, race, and anatomic location of the incision [97]. Another study evaluating healing effects of LLL in dental practice examined 48 patients subjected to surgical removal of their lower third molars. Treated patients were administered Ga-Al-As diode generated 808 nm at a dose of 12 J. The study demonstrated that extraoral LLL is more effective than intraoral LLL, which was more effective than control for the reduction of postoperative trismus and swelling after extraction of the lower third molar [98].

Given the predominance of data supporting fibroblast proliferative ability and animal wound healing effects of LLL therapy, a clinical trial was performed on healing of ulcers. In a double-blinded fashion 23 diabetic leg ulcers from 14 patients were divided into two groups. Phototherapy was applied (<1.0 J/cm²) twice per week, using a Dynatron Solaris 705(R) LED device that concurrently emits 660 and 890 nm energies. At days 15, 30, 45, 60, 75, and 90 mean ulcer granulation and healing rates were significantly higher for the treatment group as compared to control. By day 90, 58.3% of the ulcers in the LLL treated group were fully healed and 75% achieved 90-100% healing. In the placebo group only one ulcer healed fully [68].

As previously mentioned, LLL appears to have some angiogenic activity. One of the major problems in coronary artery disease is lack of collateralization. In a 39 patient study advanced CAD, two sessions of irradiation of low-energy laser light on skin in the chest area from helium-neon B1 lasers. The time of irradiation was 15 minutes while operations were performed 6 days a week for one month. Reduction in Canadian Cardiology Society (CCS) score, increased exercise capacity and time, less frequent angina symptoms during the treadmill test, longer distance of 6-minute walk test and a trend towards less frequent 1 mm ST depression lasting 1 min during Holter recordings was noted after therapy [99].

Perhaps one of the largest clinical trials with LLL was the NEST trial performed by Photothera. In this double blind trial 660 stroke patients were recruited and randomized: 331 received LLL and 327 received sham. No prespecified test achieved significance, but a post hoc analysis of patients with a baseline National Institutes of Health Stroke Scale score of <16 showed a favorable outcome at 90 days on the primary end point (P < 0.044) [100]. Currently Photothera is in the process of repeating this trial with modified parameters.

Relevance of LLL to COPD

A therapeutic intervention in COPD would require addressing the issues of inflammation and regeneration. Although approaches such as administration of bone marrow stem cells, or fat derived cellular components have both regenerative and anti-inflammatory activity in animal models, the need to enhance their potency for clinical applications can be seen in the recent Osiris's COPD trial interim data which reported no significant improvement in pulmonary function [101]. Accordingly, we sought to develop a possible rationale for how LLL may be useful as an adjunct to autologous stem cell therapy.

Table 1	Examples	of LLL	Properties	Relevant	to	COPD
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COPD Property	LLL Experiment	LLL Details R
Inflammation	In vivo. Decreased joint inflammation in zymosan-induced arthritis	Semiconductor laser (685 nm and 830 nm) at (2.5 J/cm ²)
	In vitro. Suppression of LPS-induced bronchial inflammation and TNF-alpha.	655 nm at of 2.6 J/cm ²
	In vivo. Carrageenan-induced pleurisy had decreased leukocyte infiltration and cytokine (TNF-alpha, IL-6, and MCP)	660 nm at 2.1 J/cm ²
	In vitro. LPS stimulated Raw 264.7 monocytes had reduced gene expression of MCP-1, IL-1 and IL-6	780 nm diode laser at 2.2 J/cm ²)
	In vivo. Suppression of LPS-stimulated neutrophil influx, myeloperoxidase activity and IL-1beta in bronchoalveolar lavage fluid.	660 nm diode laser at 7.5 J/cm ²
	In vitro. Inhibition of TNF-alpha induced IL-1, IL-8 and TNF-alpha mRNA in human synoviocytes	810 nm (5 J/cm ²) suppressed IL-1 and TNF, (25 J/cm ²) also suppressed IL-8
	In vivo. Reduction of TNF-alpha in diaphragm muscle after intravenous LPS injection.	4 sessions in 24 h with diode Ga-Asl-Al laser of 650 nm and a total dose of 5.2 J/cm ²
	In vivo. Inhibition of LPS induced peritonitis and neutrophil influx	3 J/cm ² and 7.5 J/cm ²
Growth Facto	r Production	
	In vivo. Upregulation of TGF- $\!\beta$ and PDGF in rat gingiva after incision.	He-Ne laser (632.8 nm) at a dose of 7.5 J/cm ²
	In vitro. Osteoblast-like cells were isolated from fetal rat calvariae had increased IGF-1	Ga-Al-As laser (830 nm) at (3.75 J/cm ²).
	In vitro. Upregulated production of IGF-1 and FGF-2 in human gingival fibroblasts.	685 nm, for 140 s, 2 J/cm ²
Angiogenesis		
	In vivo. Increased fiber to capillary ratio in rabbits with ligated femoral arteries.	Gallium-aluminum-arsenide (Ga-Al-As) diode laser, 904 nm and power of 10 mW
	In vitro. Stimulation of HUVEC proliferation by conditioned media from LLL-treated T cells	820 nm at 1.2 and 3.6 J/cm ² .
	In vitro. 7-fold increased production of VEGF by cardiomyocytes, 1.6-fold increase by smooth muscle cells (SMC) and fibroblasts. Supernatant of SMC had increased HUVEC-stimulating potential.	He:Ne continuous wave laser (632 nm). 0.5 J/cm ² for SMC, 2.1 J/cm ² for fibroblasts and 1.05 J/cm ² for cardiomyocytes.
	In vitro. Direct stimulation of HUVEC proliferation	670 nm diode device at 2 and 8 J/cm ²
Direct Stem C	ell Effects	
	In vivo. LLL precondition significantly enhanced early cell survival rate by 2-fold, decreased the apoptotic percentage of implanted BMSCs in infarcted myocardium and increased the number of newly formed capillaries.	635 nm at 0.96 J/cm ²
	In vitro. LLL stimulated MSC proliferation, VEGF and NGF production, and myogenic differentiation after 5-aza induction.	635 nm diode laser at 0.5 J/cm^2 for MSC proliferation, 5 J/cm^2 for VEGF and NGF production and for augmentation of induced myogenic differentiation
	In vitro. Increased proliferation of rat MSC.	Red light LED 630 nm at 2 and 4 J/cm(2)
	In vitro . Augmented proliferation of bone marrow and cardiac specific stem cells.	GA-AI-As 810 nm at 1 and 3 J/cm ²
	In vitro/In vivo. Administration of LLL-treated MSC resulted 53% reduction in infarct size, 5- and 6.3-fold significant increase in cell density that positively immunoreacted to BrdU and c-kit, respectively, and 1.4- and 2-fold higher level of angiogenesis and vascular endothelial growth factor, respectively, when compared to non-laser-treated implanted cells	Ga-Al-As laser (810 nm wavelength), 1 J/cm ²
	In vitro. Enhanced proliferation of adipose derived MSC in presence of EGF.	636 nm diode, 5 J/cm ²

In vitro . Enhanced proliferation and beta-1 integrin expression of a adipose derived MSC.	635 nm diode laser, at 5 J/cm ²
Clinical. 660 stroke patients: 331 received LLL and 327 received sham. No prespecified test achieved significance, but a post hoc analysis of patients with a baseline National Institutes of Health Stroke Scale score of <16 showed a favorable outcome at 90 days on the primary end point (P < 0.044).	808 nm. No density disclosed.

Table 1: Examples of LLL Properties Relevant to COPD (Continued)

Table 1 depicts some of the properties of LLL that provide a rationale for the combined use with stem cells. One of the basic properties of LLL seems to be ability to inhibit inflammation at the level of innate immune activation. Representative studies showed that LLL was capable of suppressing inflammatory genes and/or pathology after administration of lipopolysaccharide (LPS) as a stimulator of monocytes [102] and bronchial cells [34], in vitro, and leukocyte infiltration in vivo [103,104]. Inflammation induced by other stimulators such as zymosan, carrageenan, and TNF-alpha was also inhibited by LLL [32,105,106]. Growth factor stimulating activity of LLL was demonstrated in both in vitro and in vivo experiments in which augmentation of FGF-2, PDGF and IGF-1 was observed [36,37,107]. Endogenous production of these growth factors may be useful in regeneration based on activation of endogenous pulmonary stem cells [108,109]. Another aspect of LLL activities of relevance is ability to stimulate angiogenesis. In COPD, the constriction of blood vessels as a result of poor oxygen uptake is results in a feedback loop culminating in pulmonary hypertension. Administration of angiogenic factors has been demonstrated to be beneficial in several animal models of pulmonary pathology [110,111]. The ability of LLL to directly induce proliferation of HUVEC cells [112], as well as to augment production of angiogenic factors such as VEGF [113], supports the possibility of creation of an environment hospitable to neoangiogenesis which is optimal for stem cell growth. In fact, a study demonstrated in vivo induction of neocapillary formation subsequent to LLL administration in a hindlimb ischemia model [114]. The critical importance of angiogenesis in stem cell mediated regeneration has previously been demonstrated in the stroke model, where the major therapeutic activity of exogenous stem cells has been attributed to angiogenic as opposed to transdifferentiation effects [115].

Direct evidence of LLL stimulating stem cells has been obtained using mesenchymal stem cells derived both from the bone marrow and from the adipose tissue [116,117]. Interestingly in vivo administration of LLL stimulated MSC has resulted in 50% decrease in cardiac infarct size [118]. Clinical translation of LLL has been performed in the area of stroke, in which a 660 patient trial demonstrated statistically significant effects in post trial subset analysis [100].

Conclusions

Despite clinical use of LLL for decades, the field is still in its infancy. As is obvious from the wide variety of LLL sources, frequencies, and intensities used, no standard protocols exist. The ability of LLL to induce growth factor production, inhibition of inflammation, stimulation of angiogenesis, and direct effects on stem cells suggests the urgent need for combining this modality with regenerative medicine, giving birth to the new field of "regenerative photoceuticals". Development of a regenerative treatment for COPD as well as for other degenerative diseases would be of considerable benefit. Regarding COPD, such treatment would be life-saving/ life extending for thousands of affected individuals. Ceasing smoking or not starting to smoke would considerably impact this disease.

Acknowledgements

The authors thank Victoria Dardov and Matthew Gandjian for critical discussions and input.

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Authors' contributions

FL, SFJ, DTA, FR, VB, VG, CAD, RDNC, ANP, EC, DRK contributed to literature review, analysis and discussion, synthesis of concepts, writing of the manuscript and proof-reading of the final draft.

Competing interests

David R Koos is a shareholder, as well as Chairman and CEO of Entest Bio. Feng Lin is research director of Entest Bio. All other authors declare no competing interest.

Received: 7 January 2010

Accepted: 16 February 2010 Published: 16 February 2010

References

- Abdel-Latif A, Bolli R, Tleyjeh IM, Montori VM, Perin EC, Hornung CA, Zuba-Surma EK, Al-Mallah M, Dawn B: Adult bone marrow-derived cells for cardiac repair: a systematic review and meta-analysis. Arch Intern Med 2007, 167:989-997.
- Martin-Rendon E, Brunskill SJ, Hyde CJ, Stanworth SJ, Mathur A, Watt SM: Autologous bone marrow stem cells to treat acute myocardial infarction: a systematic review. Eur Heart J 2008, 29:1807-1818.
- 3. Vet-Stem Regenerative Veterinary Medicine. http://www.vet-stem.com.

- Riordan NH, Ichim TE, Min WP, Wang H, Solano F, Lara F, Alfaro M, Rodriguez JP, Harman RJ, Patel AN, Murphy MP, Lee RR, Minev B: Nonexpanded adipose stromal vascular fraction cell therapy for multiple sclerosis. J Transl Med 2009, 7:29.
- Motz GT, Eppert BL, Sun G, Wesselkamper SC, Linke MJ, Deka R, Borchers MT: Persistence of lung CD8 T cell oligoclonal expansions upon smoking cessation in a mouse model of cigarette smoke-induced emphysema. J Immunol 2008, 181:8036-8043.
- Maeno T, Houghton AM, Quintero PA, Grumelli S, Owen CA, Shapiro SD: CD8+ T Cells are required for inflammation and destruction in cigarette smoke-induced emphysema in mice. J Immunol 2007, 178:8090-8096.
- Woodruff PG, Koth LL, Yang YH, Rodriguez MW, Favoreto S, Dolganov GM, Paquet AC, Erle DJ: A distinctive alveolar macrophage activation state induced by cigarette smoking. Am J Respir Crit Care Med 2005, 172:1383-1392.
- 8. Stefanska AM, Walsh PT: Chronic obstructive pulmonary disease: evidence for an autoimmune component. *Cell Mol Immunol* 2009, **6**:81-86.
- Gonzalez-Rey E, Gonzalez MA, Varela N, O'Valle F, Hernandez-Cortes P, Rico L, Buscher D, Delgado M: Human adipose-derived mesenchymal stem cells reduce inflammatory and T cell responses and induce regulatory T cells in vitro in rheumatoid arthritis. *Ann Rheum Dis* 69:241-248.
- Lepelletier Y, Lecourt S, Arnulf B, Vanneaux V, Fermand JP, Menasche P, Domet T, Marolleau JP, Hermine O, Larghero J: Galectin-1 and Semaphorin-3A are two soluble factors conferring T cell immunosuppression to bone marrow mesenchymal stem cell. Stem Cells Dev 2009.
- Tsyb AF, Petrov VN, Konoplyannikov AG, Saypina EV, Lepechina LA, Kalsina S, Semenkova IV, Agaeva EV: In vitro inhibitory effect of mesenchymal stem cells on zymosan-induced production of reactive oxygen species. *Bull Exp Biol Med* 2008, 146:158-164.
- Sun L, Akiyama K, Zhang H, Yamaza T, Hou Y, Zhao S, Xu T, Le A, Shi S: Mesenchymal stem cell transplantation reverses multiorgan dysfunction in systemic lupus erythematosus mice and humans. *Stem Cells* 2009, 27:1421-1432.
- Feuerer M, Herrero L, Cipolletta D, Naaz A, Wong J, Nayer A, Lee J, Goldfine AB, Benoist C, Shoelson S, Mathis D: Lean, but not obese, fat is enriched for a unique population of regulatory T cells that affect metabolic parameters. *Nat Med* 2009, 15:930-939.
- Ogawa Y, Duru EA, Ameredes BT: Role of IL-10 in the resolution of airway inflammation. Curr Mol Med 2008, 8:437-445.
- Serrano-Mollar A, Nacher M, Gay-Jordi G, Closa D, Xaubet A, Bulbena O: Intratracheal transplantation of alveolar type II cells reverses bleomycininduced lung fibrosis. *Am J Respir Crit Care Med* 2007, **176**:1261-1268.
- Aslam M, Baveja R, Liang OD, Fernandez-Gonzalez A, Lee C, Mitsialis SA, Kourembanas S: Bone marrow stromal cells attenuate lung injury in a murine model of neonatal chronic lung disease. *Am J Respir Crit Care Med* 2009, 180:1122-1130.
- van Haaften T, Byrne R, Bonnet S, Rochefort GY, Akabutu J, Bouchentouf M, Rey-Parra GJ, Galipeau J, Haromy A, Eaton F, Chen M, Hashimoto K, Abley D, Korbutt G, Archer SL, Thébaud B: Airway delivery of mesenchymal stem cells prevents arrested alveolar growth in neonatal lung injury in rats. *Am J Respir Crit Care Med* 2009, 180:1131-1142.
- Zalewska-Kaszubska J, Obzejta D: Use of low-energy laser as adjunct treatment of alcohol addiction. *Lasers Med Sci* 2004, 19:100-104.
- Nikitin AV, Esaulenko IE, Shatalova OL: [Effectiveness of laser puncture in elderly patients with bronchial asthma accompanied by chronic rhinosinusitis]. Adv Gerontol 2008, 21:424-426.
- Vasil'ev AP, Strel'tsova NN, lu Senatorov N: [Laser irradiation in the treatment of ischemic heart disease]. Vopr Kurortol Fizioter Lech Fiz Kult 2001, 10-13.
- 21. Maiman TH: Stimulated optical radiation in Ruby. Nature 187:493.
- 22. Roy D: Ablative facial resurfacing. Dermatol Clin 2005, 23:549-559, viii.
- Brown MC: An evidence-based approach to patient selection for laser vision correction. J Refract Surg 2009, 25:S661-667.
- 24. Brancaleon L, Moseley H: Laser and non-laser light sources for photodynamic therapy. *Lasers Med Sci* 2002, **17**:173-186.
- Mester ESB, Tota JG: "Effect of laser on hair growth of mice". Kiserl Orvostud 1967, 19:628-631.

- Mester E, Korenyi-Both A, Spiry T, Tisza S: The effect of laser irradiation on the regeneration of muscle fibers (preliminary report). Z Exp Chir 1975, 8:258-262.
- Posten W, Wrone DA, Dover JS, Arndt KA, Silapunt S, Alam M: Low-level laser therapy for wound healing: mechanism and efficacy. *Dermatol Surg* 2005, 31:334-340.
- Vladimirov YA, Osipov AN, Klebanov Gl: Photobiological principles of therapeutic applications of laser radiation. *Biochemistry (Mosc)* 2004, 69:81-90.
- 29. Hu WP, Wang JJ, Yu CL, Lan CC, Chen GS, Yu HS: Helium-neon laser irradiation stimulates cell proliferation through photostimulatory effects in mitochondria. *J Invest Dermatol* 2007, **127**:2048-2057.
- Moriyama Y, Nguyen J, Akens M, Moriyama EH, Lilge L: In vivo effects of low level laser therapy on inducible nitric oxide synthase. *Lasers Surg Med* 2009, 41:227-231.
- Samoilova KA, Zhevago NA, Petrishchev NN, Zimin AA: Role of nitric oxide in the visible light-induced rapid increase of human skin microcirculation at the local and systemic levels: II. healthy volunteers. *Photomed Laser Surg* 2008, 26:443-449.
- 32. Yamaura M, Yao M, Yaroslavsky I, Cohen R, Smotrich M, Kochevar IE: Low level light effects on inflammatory cytokine production by rheumatoid arthritis synoviocytes. *Lasers Surg Med* 2009, **41**:282-290.
- 33. Shiba H, Tsuda H, Kajiya M, Fujita T, Takeda K, Hino T, Kawaguchi H, Kurihara H: Neodymium-doped yttrium-aluminium-garnet laser irradiation abolishes the increase in interleukin-6 levels caused by peptidoglycan through the p38 mitogen-activated protein kinase pathway in human pulp cells. J Endod 2009, 35:373-376.
- 34. de Lima Mafra F, Costa MS, Albertini R, Silva JA Jr, Aimbire F: Low level laser therapy (LLLT): attenuation of cholinergic hyperreactivity, beta(2)adrenergic hyporesponsiveness and TNF-alpha mRNA expression in rat bronchi segments in E. coli lipopolysaccharide-induced airway inflammation by a NF-kappaB dependent mechanism. *Lasers Surg Med* 2009, **41**:68-74.
- Aimbire F, de Oliveira Ligeiro AP, Albertini R, Correa JC, de Campos Ladeira CB, Lyon JP, Silva JA Jr, Costa MS: Low level laser therapy (LLLT) decreases pulmonary microvascular leakage, neutrophil influx and IL-1beta levels in airway and lung from rat subjected to LPS-induced inflammation. *Inflammation* 2008, 31:189-197.
- Safavi SM, Kazemi B, Esmaeili M, Fallah A, Modarresi A, Mir M: Effects of low-level He-Ne laser irradiation on the gene expression of IL-1beta, TNF-alpha, IFN-gamma, TGF-beta, bFGF, and PDGF in rat's gingiva. *Lasers Med Sci* 2008, 23:331-335.
- Saygun I, Karacay S, Serdar M, Ural AU, Sencimen M, Kurtis B: Effects of laser irradiation on the release of basic fibroblast growth factor (bFGF), insulin like growth factor-1 (IGF-1), and receptor of IGF-1 (IGFBP3) from gingival fibroblasts. *Lasers Med Sci* 2008, 23:211-215.
- Schwartz F, Brodie C, Appel E, Kazimirsky G, Shainberg A: Effect of helium/ neon laser irradiation on nerve growth factor synthesis and secretion in skeletal muscle cultures. J Photochem Photobiol B 2002, 66:195-200.
- Yu W, Naim JO, Lanzafame RJ: The effect of laser irradiation on the release of bFGF from 3T3 fibroblasts. *Photochem Photobiol* 1994, 59:167-170.
- 40. Zungu IL, Hawkins Evans D, Abrahamse H: Mitochondrial responses of normal and injured human skin fibroblasts following low level laser irradiation–an in vitro study. *Photochem Photobiol* 2009, **85**:987-996.
- Wu S, Xing D, Gao X, Chen WR: High fluence low-power laser irradiation induces mitochondrial permeability transition mediated by reactive oxygen species. J Cell Physiol 2009, 218:603-611.
- Lan CC, Wu CS, Chiou MH, Chiang TY, Yu HS: Low-energy helium-neon laser induces melanocyte proliferation via interaction with type IV collagen: visible light as a therapeutic option for vitiligo. Br J Dermatol 2009, 161:273-280.
- Karu T: Photobiology of low-power laser effects. *Health Phys* 1989, 56:691-704.
- Tiphlova O, Karu T: Role of primary photoacceptors in low-power laser effects: action of He-Ne laser radiation on bacteriophage T4-Escherichia coli interaction. *Lasers Surg Med* 1989, 9:67-69.
- Karu TI: Mitochondrial signaling in mammalian cells activated by red and near-IR radiation. Photochem Photobiol 2008, 84:1091-1099.

- Zhang L, Xing D, Zhu D, Chen Q: Low-power laser irradiation inhibiting Abeta25-35-induced PC12 cell apoptosis via PKC activation. *Cell Physiol Biochem* 2008, 22:215-222.
- Aimbire F, Santos FV, Albertini R, Castro-Faria-Neto HC, Mittmann J, Pacheco-Soares C: Low-level laser therapy decreases levels of lung neutrophils anti-apoptotic factors by a NF-kappaB dependent mechanism. Int Immunopharmacol 2008, 8:603-605.
- 48. Lipovsky A, Nitzan Y, Lubart R: A possible mechanism for visible lightinduced wound healing. *Lasers Surg Med* 2008, 40:509-514.
- Ignatieva N, Zakharkina O, Andreeva I, Sobol E, Kamensky V, Lunin V: Effects of laser irradiation on collagen organization in chemically induced degenerative annulus fibrosus of lumbar intervertebral disc. *Lasers Surg Med* 2008, 40:422-432.
- Silveira LB, Prates RA, Novelli MD, Marigo HA, Garrocho AA, Amorim JC, Sousa GR, Pinotti M, Ribeiro MS: Investigation of mast cells in human gingiva following low-intensity laser irradiation. *Photomed Laser Surg* 2008, 26:315-321.
- Coombe AR, Ho CT, Darendeliler MA, Hunter N, Philips JR, Chapple CC, Yum LW: The effects of low level laser irradiation on osteoblastic cells. *Clin Orthod Res* 2001, 4:3-14.
- 52. Gregoraszczuk E, Dobrowolski JW, Galas J: Effect of low intensity laser beam on steroid dehydrogenase activity and steroid hormone production in cultured porcine granulosa cells. *Folia Histochem Cytochem* (*Krakow*) 1983, **21**:87-92.
- Yu HS, Chang KL, Yu CL, Chen JW, Chen GS: Low-energy helium-neon laser irradiation stimulates interleukin-1 alpha and interleukin-8 release from cultured human keratinocytes. J Invest Dermatol 1996, 107:593-596.
- Mvula B, Mathope T, Moore T, Abrahamse H: The effect of low level laser irradiation on adult human adipose derived stem cells. *Lasers Med Sci* 2008, 23:277-282.
- Mvula B, Moore TJ, Abrahamse H: Effect of low-level laser irradiation and epidermal growth factor on adult human adipose-derived stem cells. *Lasers Med Sci* 25:33-39.
- Hawkins DH, Abrahamse H: The role of laser fluence in cell viability, proliferation, and membrane integrity of wounded human skin fibroblasts following helium-neon laser irradiation. *Lasers Surg Med* 2006, 38:74-83.
- Houreld N, Abrahamse H: In vitro exposure of wounded diabetic fibroblast cells to a helium-neon laser at 5 and 16 J/cm2. Photomed Laser Surg 2007, 25:78-84.
- Smol'yaninova NK, Karu TI, Fedoseeva GE, Zelenin AV: Effects of He-Ne laser irradiation on chromatin properties and synthesis of nucleic acids in human peripheral blood lymphocytes. *Biomed Sci* 1991, 2:121-126.
- Funk JO, Kruse A, Neustock P, Kirchner H: Helium-neon laser irradiation induces effects on cytokine production at the protein and the mRNA level. *Exp Dermatol* 1993, 2:75-83.
- Funk JO, Kruse A, Kirchner H: Cytokine production after helium-neon laser irradiation in cultures of human peripheral blood mononuclear cells. J Photochem Photobiol B 1992, 16:347-355.
- Gulsoy M, Ozer GH, Bozkulak O, Tabakoglu HO, Aktas E, Deniz G, Ertan C: The biological effects of 632.8-nm low energy He-Ne laser on peripheral blood mononuclear cells in vitro. J Photochem Photobiol B 2006, 82:199-202.
- Novoselova EG, Cherenkov DA, Glushkova OV, Novoselova TV, Chudnovskii VM, lusupov VI, Fesenko EE: [Effect of low-intensity laser radiation (632.8 nm) on immune cells isolated from mice]. *Biofizika* 2006, 51:509-518.
- Dube A, Bansal H, Gupta PK: Modulation of macrophage structure and function by low level He-Ne laser irradiation. *Photochem Photobiol Sci* 2003, 2:851-855.
- Hemvani N, Chitnis DS, Bhagwanani NS: Helium-neon and nitrogen laser irradiation accelerates the phagocytic activity of human monocytes. *Photomed Laser Surg* 2005, 23:571-574.
- Moustsen PA, Vinter N, Aas-Andersen L, Kragstrup J: [Laser treatment of sinusitis in general practice assessed by a double-blind controlled study]. Ugeskr Laeger 1991, 153:2232-2234.
- Shen X, Zhao L, Ding G, Tan M, Gao J, Wang L, Lao L: Effect of combined laser acupuncture on knee osteoarthritis: a pilot study. *Lasers Med Sci* 2009, 24:129-136.

- 67. Ekim A, Armagan O, Tascioglu F, Oner C, Colak M: Effect of low level laser therapy in rheumatoid arthritis patients with carpal tunnel syndrome. *Swiss Med Wkly* 2007, **137**:347-352.
- Minatel DG, Frade MA, Franca SC, Enwemeka CS: Phototherapy promotes healing of chronic diabetic leg ulcers that failed to respond to other therapies. *Lasers Surg Med* 2009, 41:433-441.
- Bocci V, Travagli V, Zanardi I: May oxygen-ozone therapy improves cardiovascular disorders?. Cardiovasc Hematol Disord Drug Targets 2009, 9:78-85.
- Bocci V, Borrelli E, Travagli V, Zanardi I: The ozone paradox: ozone is a strong oxidant as well as a medical drug. Med Res Rev 2009, 29:646-682.
- Re L, Mawsouf MN, Menendez S, Leon OS, Sanchez GM, Hernandez F: Ozone therapy: clinical and basic evidence of its therapeutic potential. *Arch Med Res* 2008, **39**:17-26.
- Damera G, Zhao H, Wang M, Smith M, Kirby C, Jester WF, Lawson JA, Panettieri RA Jr: Ozone modulates IL-6 secretion in human airway epithelial and smooth muscle cells. Am J Physiol Lung Cell Mol Physiol 2009, 296:L674-683.
- 73. Manzer R, Dinarello CA, McConville G, Mason RJ: Ozone exposure of macrophages induces an alveolar epithelial chemokine response through IL-1alpha. *Am J Respir Cell Mol Biol* 2008, **38**:318-323.
- McDonald RJ, Usachencko J: Neutrophils injure bronchial epithelium after ozone exposure. Inflammation 1999, 23:63-73.
- Rodriguez ZZ, Guanche D, Alvarez RG, Rosales FH, Alonso Y, Schulz S: Preconditioning with ozone/oxygen mixture induces reversion of some indicators of oxidative stress and prevents organic damage in rats with fecal peritonitis. *Inflamm Res* 2009.
- Zamora ZB, Borrego A, Lopez OY, Delgado R, Gonzalez R, Menendez S, Hernandez F, Schulz S: Effects of ozone oxidative preconditioning on TNF-alpha release and antioxidant-prooxidant intracellular balance in mice during endotoxic shock. *Mediators Inflamm* 2005, 2005:16-22.
- Borrego A, Zamora ZB, Gonzalez R, Romay C, Menendez S, Hernandez F, Montero T, Rojas E: Protection by ozone preconditioning is mediated by the antioxidant system in cisplatin-induced nephrotoxicity in rats. *Mediators Inflamm* 2004, 13:13-19.
- Martinez-Sanchez G, Al-Dalain SM, Menendez S, Re L, Giuliani A, Candelario-Jalil E, Alvarez H, Fernandez-Montequin JI, Leon OS: Therapeutic efficacy of ozone in patients with diabetic foot. *Eur J Pharmacol* 2005, 523:151-161.
- Surinchak JS, Alago ML, Bellamy RF, Stuck BE, Belkin M: Effects of low-level energy lasers on the healing of full-thickness skin defects. *Lasers Surg* Med 1983, 2:267-274.
- Broadley C, Broadley KN, Disimone G, Riensch L, Davidson JM: Low-energy helium-neon laser irradiation and the tensile strength of incisional wounds in the rat. Wound Repair Regen 1995, 3:512-517.
- Ghamsari SM, Taguchi K, Abe N, Acorda JA, Yamada H: Histopathological effect of low-level laser therapy on sutured wounds of the teat in dairy cattle. *Vet Q* 1996, 18:17-21.
- Ghamsari SM, Taguchi K, Abe N, Acorda JA, Sato M, Yamada H: Evaluation of low level laser therapy on primary healing of experimentally induced full thickness teat wounds in dairy cattle. *Vet Surg* 1997, 26:114-120.
- Pinfildi CE, Liebano RE, Hochman BS, Ferreira LM: Helium-neon laser in viability of random skin flap in rats. *Lasers Surg Med* 2005, 37:74-77.
- Maiya GA, Kumar P, Rao L: Effect of low intensity helium-neon (He-Ne) laser irradiation on diabetic wound healing dynamics. *Photomed Laser* Surg 2005, 23:187-190.
- Bayat M, Vasheghani MM, Razavi N, Taheri S, Rakhshan M: Effect of lowlevel laser therapy on the healing of second-degree burns in rats: a histological and microbiological study. J Photochem Photobiol B 2005, 78:171-177.
- Garavello I, Baranauskas V, da Cruz-Hofling MA: The effects of low laser irradiation on angiogenesis in injured rat tibiae. *Histol Histopathol* 2004, 19:43-48.
- Lin YS, Huang MH, Chai CY, Yang RC: Effects of helium-neon laser on levels of stress protein and arthritic histopathology in experimental osteoarthritis. Am J Phys Med Rehabil 2004, 83:758-765.
- Jamieson RW, Friend PJ: Organ reperfusion and preservation. Front Biosci 2008, 13:221-235.
- Akai M, Usuba M, Maeshima T, Shirasaki Y, Yasuoka S: Laser's effect on bone and cartilage change induced by joint immobilization: an experiment with animal model. *Lasers Surg Med* 1997, 21:480-484.

- Nakano J, Kataoka H, Sakamoto J, Origuchi T, Okita M, Yoshimura T: Lowlevel laser irradiation promotes the recovery of atrophied gastrocnemius skeletal muscle in rats. *Exp Physiol* 2009, 94:1005-1015.
- Doin-Silva R, Baranauskas V, Rodrigues-Simioni L, da Cruz-Hofling MA: The ability of low level laser therapy to prevent muscle tissue damage induced by snake venom. *Photochem Photobiol* 2009, 85:63-69.
- Gal D, Chokshi SK, Mosseri M, Clarke RH, Isner JM: Percutaneous delivery of low-level laser energy reverses histamine-induced spasm in atherosclerotic Yucatan microswine. *Circulation* 1992, 85:756-768.
- Kipshidze N, Sahota H, Komorowski R, Nikolaychik V, Keelan MH Jr: Photoremodeling of arterial wall reduces restenosis after balloon angioplasty in an atherosclerotic rabbit model. J Am Coll Cardiol 1998, 31:1152-1157.
- Novoselova EG, Glushkova OV, Cherenkov DA, Chudnovsky VM, Fesenko EE: Effects of low-power laser radiation on mice immunity. *Photodermatol Photoimmunol Photomed* 2006, 22:33-38.
- Bensadoun RJ, Franquin JC, Ciais G, Darcourt V, Schubert MM, Viot M, Dejou J, Tardieu C, Benezery K, Nguyen TD, Laudoyer Y, Dassonville O, Poissonnet G, Vallicioni J, Thyss A, Hamdi M, Chauvel P, Demard F: Lowenergy He/Ne laser in the prevention of radiation-induced mucositis. A multicenter phase III randomized study in patients with head and neck cancer. Support Care Cancer 1999, 7:244-252.
- Arun Maiya G, Sagar MS, Fernandes D: Effect of low level helium-neon (He-Ne) laser therapy in the prevention & treatment of radiation induced mucositis in head & neck cancer patients. *Indian J Med Res* 2006, 124:399-402.
- 97. Neiburger EJ: Rapid healing of gingival incisions by the helium-neon diode laser. J Mass Dent Soc 1999, **48**:8-13, 40..
- 98. Aras MH, Gungormus M: Placebo-controlled randomized clinical trial of the effect two different low-level laser therapies (LLLT)-intraoral and extraoral-on trismus and facial swelling following surgical extraction of the lower third molar. *Lasers Med Sci* 2009.
- Zycinski P, Krzeminska-Pakula M, Peszynski-Drews C, Kierus A, Trzos E, Rechcinski T, Figiel L, Kurpesa M, Plewka M, Chrzanowski L, Drozdz J: Laser biostimulation in end-stage multivessel coronary artery disease–a preliminary observational study. *Kardiol Pol* 2007, 65:13-21, discussion 22-13..
- 100. Zivin JA, Albers GW, Bornstein N, Chippendale T, Dahlof B, Devlin T, Fisher M, Hacke W, Holt W, Ilic S, Kasner S, Lew R, Nash M, Perez J, Rymer M, Schellinger P, Schneider D, Schwab S, Veltkamp R, Walker M, Streeter J, NeuroThera Effectiveness and Safety Trial-2 Investigators: Effectiveness and safety of transcranial laser therapy for acute ischemic stroke. Stroke 2009, 40:1359-1364.
- 101. Osiris Therapeutics Reports Interim Data for COPD Trial. http://www. medicalnewstoday.com/articles/155267.php.
- 102. Gavish L, Perez LS, Reissman P, Gertz SD: Irradiation with 780 nm diode laser attenuates inflammatory cytokines but upregulates nitric oxide in lipopolysaccharide-stimulated macrophages: implications for the prevention of aneurysm progression. *Lasers Surg Med* 2008, 40:371-378.
- 103. Correa F, Lopes Martins RA, Correa JC, Iversen W, Joenson J, Bjordal JM: Low-level laser therapy (GaAs lambda = 904 nm) reduces inflammatory cell migration in mice with lipopolysaccharide-induced peritonitis. *Photomed Laser Surg* 2007, 25:245-249.
- 104. Aimbire F, Lopes-Martins RA, Castro-Faria-Neto HC, Albertini R, Chavantes MC, Pacheco MT, Leonardo PS, Iversen W, Bjordal JM: Low-level laser therapy can reduce lipopolysaccharide-induced contractile force dysfunction and TNF-alpha levels in rat diaphragm muscle. Lasers Med Sci 2006, 21:238-244.
- 105. de Morais NC, Barbosa AM, Vale ML, Villaverde AB, de Lima CJ, Cogo JC, Zamuner SR: Anti-Inflammatory Effect of Low-Level Laser and Light-Emitting Diode in Zymosan-Induced Arthritis. Photomed Laser Surg 2009.
- 106. Boschi ES, Leite CE, Saciura VC, Caberlon E, Lunardelli A, Bitencourt S, Melo DA, Oliveira JR: Anti-Inflammatory effects of low-level laser therapy (660 nm) in the early phase in carrageenan-induced pleurisy in rat. *Lasers Surg Med* 2008, 40:500-508.
- 107. Shimizu N, Mayahara K, Kiyosaki T, Yamaguchi A, Ozawa Y, Abiko Y: Lowintensity laser irradiation stimulates bone nodule formation via insulinlike growth factor-I expression in rat calvarial cells. *Lasers Surg Med* 2007, 39:551-559.
- Hackett TL, Shaheen F, Johnson A, Wadsworth S, Pechkovsky DV, Jacoby DB, Kicic A, Stick SM, Knight DA: Characterization of side

population cells from human airway epithelium. Stem Cells 2008, 26:2576-2585.

- 109. Irwin D, Helm K, Campbell N, Imamura M, Fagan K, Harral J, Carr M, Young KA, Klemm D, Gebb S, Dempsey EC, West J, Majka S: Neonatal lung side population cells demonstrate endothelial potential and are altered in response to hyperoxia-induced lung simplification. Am J Physiol Lung Cell Mol Physiol 2007, 293:L941-951.
- 110. Thebaud B, Ladha F, Michelakis ED, Sawicka M, Thurston G, Eaton F, Hashimoto K, Harry G, Haromy A, Korbutt G, Archer SL: Vascular endothelial growth factor gene therapy increases survival, promotes lung angiogenesis, and prevents alveolar damage in hyperoxia-induced lung injury: evidence that angiogenesis participates in alveolarization. *Circulation* 2005, 112:2477-2486.
- 111. Thebaud B: Angiogenesis in lung development, injury and repair: implications for chronic lung disease of prematurity. *Neonatology* 2007, 91:291-297.
- Schindl A, Merwald H, Schindl L, Kaun C, Wojta J: Direct stimulatory effect of low-intensity 670 nm laser irradiation on human endothelial cell proliferation. Br J Dermatol 2003, 148:334-336.
- 113. Kipshidze N, Nikolaychik V, Keelan MH, Shankar LR, Khanna A, Kornowski R, Leon M, Moses J: Low-power helium: neon laser irradiation enhances production of vascular endothelial growth factor and promotes growth of endothelial cells in vitro. *Lasers Surg Med* 2001, **28**:355-364.
- 114. Ihsan FR: Low-level laser therapy accelerates collateral circulation and enhances microcirculation. *Photomed Laser Surg* 2005, 23:289-294.
- 115. Taguchi A, Soma T, Tanaka H, Kanda T, Nishimura H, Yoshikawa H, Tsukamoto Y, Iso H, Fujimori Y, Stern DM, Naritomi H, Matsuyama T: Administration of CD34+ cells after stroke enhances neurogenesis via angiogenesis in a mouse model. J Clin Invest 2004, 114:330-338.
- 116. Li WT, Leu YC: Effects of low level red-light irradiation on the proliferation of mesenchymal stem cells derived from rat bone marrow. Conf Proc IEEE Eng Med Biol Soc 2007, 2007;5830-5833.
- Tuby H, Maltz L, Oron U: Low-level laser irradiation (LLLI) promotes proliferation of mesenchymal and cardiac stem cells in culture. *Lasers* Surg Med 2007, 39:373-378.
- 118. Tuby H, Maltz L, Oron U: Implantation of low-level laser irradiated mesenchymal stem cells into the infarcted rat heart is associated with reduction in infarct size and enhanced angiogenesis. *Photomed Laser Surg* 2009, **27**:227-233.
- 119. Agaiby AD, Ghali LR, Wilson R, Dyson M: Laser modulation of angiogenic factor production by T-lymphocytes. *Lasers Surg Med* 2000, 26:357-363.
- 120. Zhang H, Hou JF, Shen Y, Wang W, Wei YJ, Hu S: Low Level Laser Irradiation Precondition to Create Friendly Milieu of Infarcted Myocardium and Enhance Early Survival of Transplanted Bone Marrow Cells. J Cell Mol Med 2009.
- 121. Hou JF, Zhang H, Yuan X, Li J, Wei YJ, Hu SS: In vitro effects of low-level laser irradiation for bone marrow mesenchymal stem cells: proliferation, growth factors secretion and myogenic differentiation. *Lasers Surg Med* 2008, **40**:726-733.

doi:10.1186/1479-5876-8-16

Cite this article as: Lin et al:: Lasers, stem cells, and COPD. Journal of Translational Medicine 2010 8:16.

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