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Highlights

- Males of some biparental species eat placenta when their young are born.
- We studied the effects of placentophagia in sexually naive male California mice.
- Placenta treatment had no effect on paternal behaviors.
- Placenta treatment decreased latencies to approach pups and novel objects.
- Placenta treatment decreased fos expression in bed nucleus of the stria terminalis.

Consequences of Placentophagia by Adult Virgin Male California Mice (*Peromyscus californicus*)

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1. Introduction

Placentophagia, or ingestion of the afterbirth, is commonly performed by parturient females of most eutherian species, with some exceptions (e.g., pinnipeds, cetaceans, humans: Kristal, 1980; Young & Benyshek, 2010). The functional significance of placentophagia is unclear, but proposed explanations include avoiding predators or pathogens and meeting general or specific nutritional demands (reviewed by Kristal, 1980; Kristal et al., 2012). Studies on the effects of maternal placentophagia in several mammalian species have revealed that this behavior can modulate pain sensitivity and maternal motivation (reviewed by Kristal, 1991). For example, in rats (*Rattus norvegicus*) and cows (*Bos spp.*), placentophagia enhances opioid-mediated analgesia through an opioid-enhancing factor (POEF) produced by and found in the placenta (Hoey et al., 2011; Kristal, 1991; Kristal et al., 2012; Pinheiro-Machado et al., 1997). This hypoalgesic effect is mediated by the vagus nerve, may occur as soon as 5 minutes after ingestion, and can last for approximately one hour (Doer & Kristal, 1989; Tarapacki et al., 1992). Placentophagia-induced hypoalgesia was recently identified as being potentially mediated by δ -opioid receptor activation (Thompson et al., 2018). Decreased pain sensitivity during parturition may facilitate labor, as neonates are expelled more quickly (Kristal, 1991). Interestingly, POEF is found in placental tissues even of species that typically do not ingest placenta (i.e., dolphins, humans), suggesting that this substance is highly conserved among placental mammals (Abbott et al., 1991).

The placenta is an endocrine organ that produces many of the protein and steroid hormones involved in the onset and maintenance of maternal and paternal care in mammals (e.g., progestogens, estrogens, lactogens: Malassine et al., 2003). Although adult, sexually inexperienced female rats do not express high levels of spontaneous maternal-like behavior (i.e.,

aloparental behavior), this can be modified with exposure to pups and placenta, or with oral administration of placenta. For example, exposure of adult female virgin rats to pups smeared with placenta and amniotic fluid shortens the latency for the expression of aloparental care (i.e., maternal sensitization) (Kristal et al., 1981). Additionally, ingestion of placenta and amniotic fluid by adult virgin female rats enhances the stimulatory effect of intracerebroventricular morphine treatment on pup-induced maternal behavior (Neumann et al., 2009). Thus, placentophagia by some female mammals may induce physiological and behavioral changes that promote maternal care and, as a result, offspring survival.

Males of some mammal species, too, ingest placenta at the birth of their young. In the uniparental (i.e., only one parent, the mother, provides offspring care) Siberian hamster (*Phodopus sungorus*), males ingest experimentally presented placenta only if they are present at the birth of their pups (Gregg & Wynne-Edwards, 2006). Similarly, male rats, which are commonly averse towards the afterbirth, will begin to eat placenta after continuous exposure to it (Abbott et al., 1991). In several biparental (i.e., both males and females care for their young) mammals, males, in addition to females, sometimes ingest placenta during the birth of their offspring. Among primates, placentophagia by males has been observed in the common marmoset (*Callithrix jacchus*: T. E. Ziegler, pers. comm.), cotton-top tamarin (*Saguinus oedipus*: T. E. Ziegler, pers. comm.), and silvery marmoset (*C. argentata*: J. A. French, pers. comm.), as well as in some human populations (Coyle et al, 2015; Marraccini & Gorman, 2015). In biparental rodents, placentophagia by males has been reported in dwarf hamsters (*Phodopus campbelli*: Gregg & Wynne-Edwards, 2005; Jones & Wynne-Edwards, 2000), California mice (*Peromyscus californicus*: Lee & Brown, 2002; Perea-Rodriguez & Saltzman, 2014), and prairie voles (*Microtus ochrogaster*: K.L. Bales, pers. comm.) (but see McGuire et al., 2003).

Studies in dwarf hamsters and California mice indicate that adult males, similar to adult females, respond differently to placenta depending on their reproductive condition. In these two species, males are more likely to ingest placenta when housed with their pair-bonded, gestating mates and when they become fathers than when they are sexually inexperienced (Gregg & Wynne-Edwards, 2005; Perea-Rodriguez & Saltzman, 2014). These findings suggest that in at least some biparental mammals, males naturally become attracted to placenta during their mates' pregnancy and may commonly ingest placenta during the birth of their offspring. Still unknown, however, are the potential behavioral and/or physiological changes that males undergo as a consequence of ingesting placenta, and whether these changes influence the males' responses to their young.

In this study, we sought to characterize the behavioral and neural responses to an unfamiliar pup after oral administration of conspecific placenta to adult, virgin male California mice. We analyzed the presence of the protein Fos, the product of the c-Fos immediate-early gene that is commonly used as a marker of neuronal activity (Hoffman & Lyo, 2002), in key brain areas involved in paternal care in rodents. Adult virgin males were used because they are highly variable in their behavioral responses to pups, whereas virtually all California mouse fathers show pronounced, rapid-onset paternal care (de Jong et al., 2009; de Jong et al., 2012; Gubernick & Nelson, 1989; Horrell et al., 2017). We speculated that behavioral and neural effects of placentophagia were likely to be mediated by steroid hormones, and steroids can exert both rapid, transient effects via non-genomic mechanisms and delayed, more sustained effects via changes in gene expression (McEwen, 1991). Therefore, we analyzed responses to pups at three time points: 1, 7, and 24 h after placenta administration.

We hypothesized that the physiological changes resulting from ingestion of placenta lead to changes in both neural and behavioral responses to pup-related stimuli. We predicted that mice treated with placenta would approach pups more rapidly, would spend more time engaging in caretaking behaviors, and would express more Fos-immunoreactivity (Fos-ir) in brain areas positively linked to paternal care (ventral bed nucleus of the stria terminalis, medial preoptic area), as well as reduced Fos-ir in brain areas commonly activated by aversive stimuli (paraventricular nucleus of the hypothalamus, amygdala), compared to controls treated with oil vehicle only. Finally, we predicted that placenta ingestion would exert these behavioral and neural effects specifically in response to a pup as opposed to a neutral novel object.

2. Methods

2a. Animals

We used male California mice born and reared in our breeding colony at the University of California, Riverside that were descended from mice purchased from the Peromyscus Genetic Stock Center (University of South Carolina, Columbia, SC). Mice were housed in standard, shoebox-style, polycarbonate cages (44 x 24 x 20 cm) containing aspen shavings for bedding and cotton wool for nesting material, with *ad libitum* access to food (Purina Rodent Chow 5001) and water. Lighting was on a 14:10 light:dark cycle, with lights on from 05:00 until 19:00 h. Ambient temperature and humidity were kept at approximately 23°C and 70%, respectively. Mice were checked twice daily and weighed twice weekly, and cages were changed weekly.

Mice were weaned at 27-31 days of age and housed in same-sex groups of three or four age-matched individuals; these groups contained no more than two siblings from any one litter. As mice reached the age of sexual maturity (~90 days: Gubernick, 1988), male groups were

divided into pairs of unrelated males. We chose adult males specifically because we wanted to test animals at a stage when they would naturally search for mates, reproduce, and ingest placenta.

2b. Experimental Design

Virgin male California mice were treated with either placenta homogenized in sesame oil or oil alone via oral gavage (see below). Beginning 1, 7, or 24 h later, each mouse underwent a 1-h behavior test with either a 1- to 4-day-old pup or a control novel object - a pup-sized, oblong glass marble. Immediately following the behavior test (i.e., 2, 8 or 25 h after placenta or oil treatment), mice were euthanized and their brains were harvested for immunohistochemical analyses (see below). Each virgin male mouse was tested under a single treatment condition (placenta or oil), at a single time point (1, 7, or 24 h after gavage), and with a single test stimulus (pup or marble). At the time of testing, mice had never been exposed to pups (other than their own littermates) or marbles. The resulting sample sizes for each treatment, time point, and stimulus type are shown in Table 1.

Mice assigned to the *placenta* group were administered a single near-term placenta (from a gestating female no more closely related to the male than second cousin) homogenized in sesame oil. Mice in the *control* group were administered sesame oil alone. We administered placenta (or oil) via oral gavage because virgin male California mice are not likely to voluntarily ingest placenta (Perea-Rodriguez & Saltzman, 2014; Perea-Rodriguez & Saltzman, unpub. data). Mice from the two treatments did not differ in age at the time of testing (placenta: 158.9 ± 4.3 days, mean $\pm$ SEM; oil: 162.9 ± 5.2 days; $p=0.63$, $T=0.46$, $df=1$; unpaired T-Test).

2c. Placenta Collection

As previously described (Perea-Rodriguez & Saltzman, 2014; Perea-Rodriguez et al., 2018), placentas were collected from multiparous (2-7 previous litters) females 1-3 days prior to their estimated parturition date, determined by the date of their previous parturition and assessment of changes in female body mass based on measurements taken every 3-4 days. Fetuses were inspected visually to confirm that they were near-term and immediately euthanized with an intraperitoneal injection (0.1 mL) of pentobarbital sodium (Fatal-Plus: Vortech Pharmaceuticals, Dearborn, Michigan, USA). Placenta donors were euthanized using CO₂ inhalation, and placentas were removed and immediately stored at -70° C.

2d. Oral Gavage

Oral gavage was performed as previously described (Perea-Rodriguez et al., 2018) using a 5 cm length of Silastic® laboratory tubing (1.57 mm inside diameter x 2.41 mm outside diameter; Dow Corning, Copley, Ohio, USA) fitted onto an 18-gauge sterile needle; the needle's tip (~ 0.5 cm) had been filed off to avoid puncturing the tubing and injuring the animal. The needle was attached to a sterile 1 mL syringe containing either a single placenta (~0.4 g, and 0.1-0.2 mL in volume) homogenized in sesame oil (total volume: 0.5 mL) or 0.5 mL sesame oil alone. This volume was selected based on the size of the stomach and to minimize any discomfort to the mice. We used oil as a vehicle because we anticipated that hormonally mediated effects of placentophagia would likely be related to steroid hormones (Cornil & Charlier, 2010), as these hormones readily cross the blood-brain barrier and are biologically active follow ingestion; steroid hormones are hydrophobic and therefore oil-soluble.

160 Additionally, the sesame oil facilitated the passage of the placental tissues through the gavage
161 apparatus.

162 Mice underwent oral gavage between 08:30 and 09:30 h. We treated animals in the
163 morning because this is the time of day when California mice are most likely to give birth
164 (within a few hours after lights-on: Lee & Brown 2002; Perea-Rodriguez & Saltzman, unpub.
165 data) and therefore to ingest placenta. Each male mouse was first housed alone for 30 min in a
166 clean isolation cage containing fresh bedding, food, and water. Placentas were thawed on ice,
167 homogenized in 0.1-0.2 mL of sesame oil using a mortar and pestle, and collected using the
168 sterile syringe, which was then attached to the 18-gauge needle fitted with the Silastic tubing; air
169 bubbles were avoided as much as possible. Mice were lightly anesthetized using isoflurane
170 (Minrad, Orchard Park, NY, USA) and held vertically as the tubing was carefully inserted into
171 the esophagus and the contents of the syringe delivered over approximately 5-10 s. The recovery
172 time from anesthesia was between 60 and 180 s, at which point animals were observed in their
173 isolation cages for 10 min before being returned to the colony room.

174 175 2e. Behavior Testing

176 Each animal underwent a behavior test in the colony room during the lights-on phase of
177 the light:dark cycle, beginning at 09:30-10:30 h (1 h after oral gavage), 16:30-17:30 h (7 h after
178 gavage), or 09:30-10:30 h the next day (24 h after gavage). At the outset of each test, a 1- to 4-
179 day-old pup (no more closely related to the male than second cousin) or a clean, pup-sized,
180 oblong, glass marble was placed at the opposite end of the male's isolation cage from the focal
181 animal. Each mouse was exposed to its respective stimulus for 60 min before being euthanized
182 for tissue collection (see below). Behavior tests were videotaped, and the initial 20 minutes were

later scored using JWatcher software (Blumstein & Daniel, 2007). Behaviors scored were latency to approach the pup or marble, duration of investigating (i.e., sniffing) the pup, and duration of huddling + licking the pup (i.e., paternal behavior). All videos were scored by a single observer, who was blind to the animals' treatment.

2f. Brain Collection, Immunohistochemistry, and Fos-ir Quantification

Immediately after each hour-long behavior test, the focal mouse was deeply anesthetized with 10% pentobarbital (Vortech, Dearborn, Michigan, USA; 0.5 mL, i.p.) and perfused transcardially, first with 0.1M phosphate-buffered saline (PBS) and subsequently with 4% paraformaldehyde (PFA) (de Jong et al., 2009). Brains were placed in 4% PFA for 1 h immediately after perfusion to further increase tissue robustness. After the additional fixation period, brains were removed from PFA and stored in 0.1M PBS at 4°C until further processing. Brains were later cryoprotected in 30% phosphate-buffered sucrose for 2-4 days, embedded in optimal cutting temperature compound, frozen, and sliced into 30 µm sections on a cryostat set at -19°C. Five series of brain sections were collected sequentially and stored in 0.1M PBS with 0.01% sodium azide until staining occurred.

Fos immunohistochemistry was performed as previously described (de Jong et al., 2009). After pre-incubation with PBS containing 0.1% bovine serum albumin and 0.3% Triton-X-100 (i.e., PBS-BT), slices were incubated in a 1:10,000 dilution of rabbit-anti-c-Fos antibody (Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) in PBS-BT overnight. The next day, after removal of excess antibody through a series of PBS washes, the slices were incubated with donkey-anti-rabbit antibody (Jackson ImmunoResearch Laboratories, West Grove, PA, USA) in a 1:1,500 dilution with PBS-BT for 90 min. Signaling was enhanced using ABC-vector (1:800

dilution in PBS-BT, Vectastain Elite Kit, Vector Laboratories, Burlingame, CA, USA) before being stained with 3,3'-diaminobenzidine tetrahydrochloride (Sigma-Aldrich, St. Louis, MO, USA) in 0.6% Tris-buffer.

Using fine brushes, stained slices were mounted onto glass slides coated with gelatin and chrome alum. Mounted slices were air-dried overnight, cleared using a range of alcohols, and embedded in Entellan New (EMS, Hatfield, PA, USA) before being coverslipped. Micrographs of stained and mounted brain slices were taken using a digital camera (Canon EOS 40D) attached to a microscope (Leica Leitz DMRB). Micrographs of the medial preoptic area (MPOA), the dorsal (dBST) and ventral (vBST) regions of the bed nucleus of the stria terminalis, the paraventricular nucleus of the hypothalamus (PVN), and the central (CeA) and basolateral (BLA) nuclei of the amygdala were taken for each brain (Figure 1). Because no brain atlas was available for *Peromyscus* when the study was performed, brain regions/nuclei of interest were located based on a standard atlas of the mouse brain (Paxinos & Franklin, 2004), as in previous studies (de Jong et al., 2009; de Jong et al., 2012).

ImageJ software (1.46r; National Institutes of Health, USA) was used to count the number of Fos-ir neurons in a 200 x 200 μm square in a representative area of neurons in each region. The person counting was unaware of the treatment and stimulus condition of each animal. Some of the brain sections were not usable due to problems during the sectioning or staining process, so these were excluded from the analyses. The final sample sizes are presented in the results.

2g. Statistical Analyses

All statistical analyses were performed using R statistical software (R Core Team, 2014). Behavioral and immunohistochemical data were tested for normality using Shapiro-Wilk tests. Bartlett's tests were used to determine homogeneity of variance. Because data collection and immunohistochemical staining for the three time points were performed separately, data from each time point were analyzed independently. Normally distributed data (latency to approach stimuli, all Fos-ir data) were analyzed by 2-way ANOVAs, with treatment (placenta, oil) and stimulus (pup, marble) as factors. If a significant ($p \leq 0.05$) treatment x stimulus interaction was found, we performed post-hoc pairwise comparisons using Tukey's HSD tests. Tukey's HSD tests performs all pairwise comparisons while controlling the probability of making Type I errors. Non-normal data (duration of huddling + licking pup, duration of investigating pup) were analyzed using Mann-Whitney U tests to compare behavioral responses in placenta- vs. oil-treated mice within each stimulus condition.

3. Results

3a. Behavioral Responses to Stimuli

Among the mice tested with a pup at each time point, the proportion that showed paternal behavior (i.e., licking and/or huddling pup) did not differ between placenta- and oil-treated males (all p -values > 0.50 , Fisher's Exact test for each time point; Table 1). Additionally, placenta treatment did not affect the total duration of caretaking behavior (huddling + licking) that mice engaged in during the pup test at any time point (all p -values > 0.40 ; Mann-Whitney U test for each time point; Figure 2). At 7 h post-gavage, placenta-treated mice approached their assigned stimuli more quickly than oil-treated mice (main effect of treatment: $F_{1,25} = 4.22$, $p = 0.05$; 2-way ANOVA); however, this effect did not differ between males tested with pups and those tested

with marbles (main effect of stimulus: $p=0.15$; treatment x stimulus interaction: $p=0.43$). Latencies to approach pups or marbles did not differ significantly between placenta- and oil-treated mice at either of the other time points (1 h: main effect of treatment: $p=0.54$; main effect of stimulus: $p=0.50$; treatment x stimulus interaction: $p=0.66$; 24 h: main effect of treatment: $p=0.63$; main effect of stimulus: $p=0.71$; treatment x stimulus interaction: $p=0.56$; 2-way ANOVA for each time point; Figure 2). Finally, placenta treatment had no effect on the total duration of time mice spent sniffing pups at any of the time points (all p -values ≥ 0.3 ; Mann-Whitney U test for each time point).

3b. Neural Responses to Stimuli

In general, total Fos-ir in the brain areas investigated was lower in mice treated with placenta than in those treated with oil; however, most of our planned analyses did not reach statistical significance (Table 2). Treatment with placenta significantly altered neural responses to stimuli in the dBST at both the 1 h and 7 h time points. Placenta-treated mice tested 1 h after oral gavage had significantly lower Fos-ir in the dBST than oil-treated controls (main effect of treatment: $F_{1,20}=4.51$, $p=0.04$; 2-way ANOVA; Table 2, Figure 3). At this time point, Fos-ir in the dBST was not influenced by stimulus type (main effect of stimulus: $p=0.54$), nor by an interaction between treatment and stimulus ($p=0.87$). At the 7 h time point, placenta-treated mice still showed a reduction in Fos-ir in the dBST compared to oil-treated controls (main effect of treatment: $F_{1,18}=4.13$, $p=0.05$), and this effect differed between males exposed to a pup and those exposed to a marble (treatment x stimulus interaction: $F_{1,18}=7.33$, $p=0.01$; 2-way ANOVA). Among placenta-treated mice, those exposed to a pup 7 h after gavage tended to show a reduction in dBST Fos-ir compared to males exposed to a marble, but this reduction was not statistically significant ($p=0.06$, Tukey's HSD test); no such effect was seen in oil-treated

animals ($p=0.69$) nor did any additional pairwise comparison reach statistical significance. At 24 h post-treatment, Fos-ir in the dBST was not significantly influenced by a main effect of treatment or stimulus, or by an interaction between these two factors (all p -values >0.33).

Fos-ir in the MPOA, vBST, PVN, BLA, and CeA was not significantly affected by treatment (all p -values >0.07 ; Table 2, Figure 3). However, 1 h after gavage, Fos-ir in both the BLA and CeA was significantly higher in mice exposed to a pup than in those exposed to a marble (BLA: main effect of stimulus: $F_{1,20}=4.60$, $p=0.04$; CeA: main effect of stimulus: $F_{1,20}=5.71$, $p=0.02$; 2-way ANOVAs), compared to mice exposed to a marble (novel object). Neither of these effects differed between placenta- and oil-treated mice (p -values >0.12).

4. Discussion

In this study, we aimed to identify possible neural and behavioral consequences of placenta ingestion (i.e., placentophagia) by adult virgin males of a monogamous, biparental rodent species. Specifically, we sought to investigate the possible role placentophagia might play in facilitating pup-directed care in the California mouse, as males of this species ingest placenta during the birth of their offspring (Lee & Brown, 2002; Perea-Rodriguez & Saltzman, 2014) and engage in extensive paternal behavior (Gubernick & Alberts, 1987). We hypothesized that the physiological changes resulting from ingestion of placenta lead to changes in both neural and behavioral responses to pup-related stimuli.

The majority of our analyses found that placenta treatment had no effect on paternal behavior. The small number of statistically significant results indicate that 7 hours after treatment, placenta-treated virgin male mice showed reduced latencies to approach both pups and novel objects (marbles) compared to oil-treated mice. In addition, placenta treatment reduced

pup- and marble-induced activation (Fos-immunoreactivity) of the dorsal region of the bed nucleus of the stria terminalis (dBST) both 1 and 7 h after treatment. At the 1-h time point, placenta-treated mice had reduced Fos-ir in response to both pup and marble stimuli, compared to oil-treated mice. Taken together, these findings indicate that ingesting placenta does not produce any major effects on paternal care but may reduce responsiveness of the dBST as rapidly as within 1 h and for as long as at least 7 h. Ingestion of placenta did not alter pup-directed care or neural activity in other brain regions, including the PVN, BLA, CeA, vBST, and, most strikingly, the MPOA, which has been implicated in paternal behavior in California mice and other biparental mammals (Bales & Saltzman, 2016; Horrell et al., 2018; Saltzman & Ziegler, 2014).

In two biparental species, prairie voles and California mice, fatherhood modulates stress reactivity and anxiety-like behaviors, suggesting that males modify how they perceive potentially aversive or novel stimuli with changes in reproductive state or reproductive experience (Bardi et al., 2011; Chauke et al., 2012; Lieberwirth et al., 2013). In the same two species, paternally responsive males have increased Fos-ir in the medial posteromedial and medial BST after exposure to pups, compared to parentally unresponsive males (de Jong et al., 2009; Kirkpatrick et al., 1994). The BST is a limbic forebrain structure that has been linked to paternal care, stress, anxiety, and aggression in California mice and other species (Bester-Meredith & Marler, 2003; Davis & Marler, 2004; Davis et al., 2010; de Jong et al., 2009; Gungor & Paré, 2016; Trainor et al., 2010). Neurochemical changes in the BST can alter an animal's behavioral response to unpredictable, threatening, and aversive stimuli (i.e., unconditioned fear) (Walker & Davis, 1997). In rodents, the BST contains dorsal and ventral regions that differ in their electrophysiological properties (Egli & Winder, 2003; Frazier et al., 2006). The dorsal and

ventral BST also respond differentially to stressors, possibly due to their dissimilar inputs from other brain nuclei and to their sensitivity to certain neurotransmitters and neuropeptides (Daniel & Rannie, 2016); however, both regions show increased Fos-ir under stressful conditions (Di Bonaventura et al., 2014). Thus, the reduced activity in the dBST seen in placenta-treated mice, as well as the shorter latencies of these mice to approach pups and marbles, may be associated overall with increased motivation to interact with environmental stimuli, regardless of whether the stimuli are pup-related.

Studies on the consequences of placenta ingestion suggest that placentophagia by mothers may trigger behavioral and physiological changes that positively affect their offspring (e.g., Abbott et al., 1991; González-Mariscal et al., 1998). In the case of males, a study on rats, which are uniparental, showed that virgin males experience hypoalgesia after ingesting placenta (Abbott et al., 1991). Recently, we showed that oral administration of placenta to male California mice, irrespective of reproductive experience, increased exploration of a novel space (an open-field arena) but had no effect on paternal behaviors (Perea-Rodriguez et al., 2018). Similarly, in the present study, placentophagia did not enhance pup-directed care, but it decreased latencies to approach novel stimuli (pups and marbles) and led to changes in neural activity in a brain nucleus heavily involved in regulating responses to a variety of environmental stimuli, including pup-related and other social stimuli.

Some important caveats should be kept in mind when interpreting the results of this study. First, we evaluated effects of placentophagia only in virgin males, rather than in fathers, because fathers typically show maximum paternal care. In our recent study, however, behavioral effects of oral treatment with placenta did not differ among California mouse fathers, first-time expectant fathers, and virgin males (Perea-Rodriguez et al., 2018). Second, although Fos

expression has been linked to changes in neuronal activity, this is not always the case; Fos may or may not be expressed when neurons undergo changes in electrical activity or gene expression (Hoffman & Lyo, 2002). Third, the sample sizes in this study were relatively small. Fourth, the oral gavage procedure by which we administered placenta eliminated possible effects that placenta and amniotic fluid may have via olfactory or accessory olfactory pathways, and the oil preparation used may have limited absorption of some of the chemicals found in placenta and amniotic fluid, such as peptide hormones. Fifth, placentophagia may have affected how mice responded to pups and marbles through neural changes in brain nuclei that were not investigated in this study (e.g., subregions of the amygdala and BST). Finally, although the oral gavage procedure does not produce any significant changes in corticosterone secretion in California mice (unpub. data), the procedure itself could have produced or inhibited any effects of placenta ingestion, an issue that our experimental design was unable to address.

5. Conclusions

In conclusion, we found that placentophagia by adult, virgin male California mice did not lead to significant changes in paternal care. Placenta administration did, however, transiently reduce males' latencies to approach an unfamiliar pup or a novel object, and reduced Fos-immunoreactivity in the dorsal region of the bed nucleus of the stria terminalis after exposure to each of these stimuli. Thus, our results are consistent with findings from a previous study (Perea-Rodriguez et al., 2018) suggesting that ingestion of placenta may reduce neophobia and anxiety-related behavior in males, but not paternal behavior *per se*.

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7. References

- Abbott, P., Thompson, A. C., Ferguson, E. J., Doerr, J. C., Tarapacki, J. A., Kostyniak, P. J., Syracuse, J. A., Cartonia D. M., & Kristal, M. B. (1991). Placental opioid-enhancing factor (POEF): Generalizability of effects. *Physiol Behav.* 50, 933-940.
- Bales, K. L., & Saltzman, W. (2016). Fathering in rodents: neurobiological substrates and consequences for offspring. *Horm Behav.* 77, 249-59
- Bardi, M., Franssen, C. L., Hampton, J. E., Shea, E. A., Fanean, A. P., & Lambert, K. G. (2011). Paternal experience and stress responses in California mice (*Peromyscus californicus*). *Comp Med.* 61, 20-30.
- Bester-Meredith, J. K., & Marler, C. A. (2003). Vasopressin and the transmission of paternal behavior across generations in mated, cross-fostered *Peromyscus* mice. *Behav Neurosci.* 117, 455-463.
- Blumstein, D. T., & Daniel, J. C. (2007). *Quantifying Behavior the JWatcher Way*. Sinauer Associates, Inc., Sunderland, MA.
- Chauke, M., de Jong, T. R., Garland, T., & Saltzman, W. (2012). Paternal responsiveness is associated with, but not mediated by reduced neophobia in male California mice (*Peromyscus californicus*). *Physiol Behav.* 107, 65-75.

404 Cornil, C. A., & Charlier, T. D. (2010). Rapid behavioural effects of oestrogens and fast
 405 regulation of their local synthesis by brain aromatase. *J Neuroendocrinol.* 22, 664-673.
 406

407 Coyle, C. W., Hulse, K. E., Wisner, K. L., Driscoll, K. E., & Clark, C. T. (2015). Placentophagy:
 408 therapeutic miracle or myth? *Arch Womens Ment Health.* 18, 673-680.
 409

410 Daniel, S. E., & Rainnie, D. G. (2016). Stress modulation of opposing circuits in the bed nucleus
 411 of the stria terminalis. *Neuropsychopharmacology*, 41, 103.
 412

413 Davis, E. S., & Marler, C. A. (2004). C-fos changes following an aggressive encounter in female
 414 California mice: A synthesis of behavior, hormone changes and neural
 415 activity. *Neuroscience.* 127, 611-624.
 416

417 Davis, M., Walker, D. L., Miles, L., & Grillon, C. (2010). Phasic vs sustained fear in rats and
 418 humans: role of the extended amygdala in fear vs anxiety. *Neuropsychopharmacology.* 35, 105-
 419 135.
 420

421 de Jong, T. R., Chauke, M., Harris, B. N., & Saltzman, W. (2009). From here to paternity:
 422 Neural correlates of the onset of paternal behavior in California mice (*Peromyscus*
 423 *californicus*). *Horm Behav.* 56, 220-231.
 424

425 de Jong, T. R., Korosi, A., Harris, B. N., Perea-Rodriguez, J. P., & Saltzman, W. (2012).
 426 Individual variation in paternal responses of virgin male California mice (*Peromyscus*
 427 *californicus*): Behavioral and physiological correlates. *Physiol Biochem Zool.* 85, 740-751.
 428
 429 Di Bonaventura, M. V. M., Ciccocioppo, R., Romano, A., Bossert, J. M., Rice, K. C., Ubaldi,
 430 M., St Laurent, R., Gaetani, S., Massi, M., Shaham, S., & Cifani, C. (2014). Role of bed nucleus
 431 of the stria terminalis corticotrophin-releasing factor receptors in frustration stress-induced
 432 binge-like palatable food consumption in female rats with a history of food restriction. *J*
 433 *Neurosci.* 34, 11316-11324.
 434
 435 Doerr, J. C., & Kristal, M. B. (1989). Enhancement of opioid-mediated analgesia by ingestion of
 436 amniotic fluid: Onset latency and duration. *Physiol Behav.* 46, 913-915.
 437
 438 Egli, R. E., & Winder, D. G. (2003). Dorsal and ventral distribution of excitable and synaptic
 439 properties of neurons of the bed nucleus of the stria terminalis. *J Neurophysiol.* 90, 405-414.
 440
 441 Frazier, C. R., Trainor, B. C., Cravens, C. J., Whitney, T. K., & Marler, C. A. (2006). Paternal
 442 behavior influences development of aggression and vasopressin expression in male California
 443 mouse offspring. *Horm Behav.* 50, 699-707.
 444
 445 González-Mariscal, G., Melo, A. I., Chirino, R., Jiménez, P., Beyer, C., & Rosenblatt, J. S.
 446 (1998). Importance of mother/young contact at parturition and across lactation for the expression
 447 of maternal behavior in rabbits. *Dev Psychobiol.* 32, 101-111.

448

449 Gregg, J. K., & Wynne-Edwards, K. E. (2005). Placentophagia in naïve adults, new fathers, and
 450 new mothers in the biparental dwarf hamster, *Phodopus campbelli*. Dev Psychobiol. 47, 179-
 451 188.

452

453 Gregg, J. K., & Wynne-Edwards, K. E. (2006). In uniparental *Phodopus sungorus*, new mothers,
 454 and fathers present during the birth of their offspring, are the only hamsters that readily consume
 455 fresh placenta. Dev Psychobiol. 48, 528-536.

456

457 Gubernick, D. J. (1988). Reproduction in the California mouse, *Peromyscus californicus*. J
 458 Mammal. 69, 857-860.

459

460 Gubernick, D. J., & Alberts, J. R. (1987). "Resource" exchange in the biparental California
 461 mouse (*Peromyscus californicus*): Water transfer from pups to parents. J Comp Psychol. 101,
 462 328-324.

463

464 Gubernick, D. J., & Nelson, R. J. (1989). Prolactin and paternal behavior in the biparental
 465 California mouse, *Peromyscus californicus*. Horm Behav. 23, 203-210.

466

467 Gungor, N. Z., & Paré, D. (2016). Functional heterogeneity in the bed nucleus of the stria
 468 terminalis. J. Neurosci. 36, 8038-8049.

469

470 Hoey, R. F., Hurley, S. W., Daniels, D., & Kristal, M. B. (2011). Effect of amniotic-fluid

471 ingestion on vaginal–cervical-stimulation-induced Fos expression in female rats during
472 estrus. Brain Res. 1376, 51-59.

473

474 Hoffman, G. E., & Lyo, D. (2002). Anatomical markers of activity in neuroendocrine systems:
475 are we all ‘Fos-ed out’? J Neuroendocrinol. 14, 259-268.

476

477 Horrell, N. D., Perea-Rodriguez, J. P., Harris, B. N., & Saltzman, W. (2017). Effects of repeated
478 pup exposure on behavioral, neural, and adrenocortical responses to pups in male California
479 mice (*Peromyscus californicus*). Horm Behav. 90, 56-63.

480

481 Horrell, N. D., Hickmott, P. W. & Saltzman, W. (2018). Neural regulation of paternal behavior
482 in mammals: Sensory, neuroendocrine, and experiential influences on the paternal brain. Curr
483 Topics Behav Neurosci. DOI: 10.1007/7854_2018_55

484

485 Jones, J. S., & Wynne-Edwards, K. E. (2000). Paternal hamsters mechanically assist the delivery,
486 consume amniotic fluid and placenta, remove fetal membranes, and provide parental care during
487 the birth process. Horm Behav. 37, 116-125.

488

489 Kirkpatrick, B., Kim, J. W., & Insel, T. R. (1994). Limbic system fos expression associated with
490 paternal behavior. Brain Res. 658, 112-118.

491

Kristal, M. B. (1980). Placentophagia: A biobehavioral enigma (or *De gustibus non disputandum est*). *Neurosci Biobehav Rev.* 4, 141-150.

Kristal, M. B. (1991). Enhancement of opioid-mediated analgesia: a solution to the enigma of placentophagia. *Neurosci Biobehav Rev.* 15, 425-435.

Kristal, M. B., DiPirro, J. M., & Thompson, A. C. (2012). Placentophagia in humans and nonhuman mammals: Causes and consequences. *Ecol Food Nutr.* 51, 177-197.

Kristal, M. B., Whitney, J. F., & Peters, L. C. (1981). Placenta on pups' skin accelerates onset of maternal behaviour in non-pregnant rats. *Animal Beh.* 29, 81-85.

Lee, A. W., & Brown, R. E. (2002). The presence of the male facilitates parturition in California mice (*Peromyscus californicus*). *Can J Zool.* 80, 926-933.

Lieberwirth, C., Wang, Y., Jia, X., Liu, Y., & Wang, Z. (2013). Fatherhood reduces the survival of adult-generated cells and affects various types of behavior in the prairie vole (*Microtus ochrogaster*). *Eur J Neurosci.* 38, 3345-3355.

Malassine, A., Frendo, J. L., & Evain-Brion, D. (2003). A comparison of placental development and endocrine functions between the human and mouse model. *Hum Reprod Update.* 9, 531-539.

514 Marraccini, M. E., & Gorman, K. S. (2015). Exploring placentophagy in Humans: Problems and
515 recommendations. *J Midwifery Womens Health*. 60, 371-379.
516

517 McEwen, B. S. (1991). Non-genomic and genomic effects of steroids on neural activity. *Trends*
518 *Pharmacol Sci*. 12, 141-147.
519

520 McGuire, B., Henyey, E., McCue, E., & Bemis, W. E. (2003). Parental behavior at parturition in
521 prairie voles (*Microtus ochrogaster*). *J. Mammal*. 84, 513-523.
522

523 Neumann, A., Hoey, R. F., Daigler, L. B., Thompson, A. C., & Kristal, M. B. (2009). Ingestion
524 of amniotic fluid enhances the facilitative effect of VTA morphine on the onset of maternal
525 behavior in virgin rats. *Brain Res*. 1261, 29-36.
526

527 Paxinos, G., & Franklin, K. B. (2004). *The Mouse Brain in Stereotaxic Coordinates*. Gulf
528 Professional Publishing.
529

530 Perea-Rodriguez, J. P., & Saltzman, W. (2014). Differences in placentophagia in relation to
531 reproductive status in the California mouse (*Peromyscus californicus*). *Dev Psychobiol*. 56, 812-
532 820.
533

534 Perea-Rodriguez, J. P., Zhao, M., Harris, B. N., Raqueno, J., & Saltzman, W. (2018). Behavioral
535 and endocrine consequences of placentophagia in male California mice (*Peromyscus*
536 *californicus*). *Physiol Behav*. 188, 283-290.

537

538 Pinheiro Machado, L. C., Hurnik, J. F., & Burton, J. H. (1997). The effect of amniotic fluid
 539 ingestion on the nociception of cows. *Physiol Behav.* 62, 1339-1344.

540

541 Saltzman, W., & Ziegler, T. E. (2014). Functional significance of hormonal changes in
 542 mammalian fathers. *J Neuroendocrinol.* 26, 685-696.

543

544 Tarapacki, J. A., Thompson, A. C., & Kristal, M. B. (1992). Gastric vagotomy blocks opioid
 545 analgesia enhancement produced by placenta ingestion. *Physiol Behav.* 52, 179-182.

546 Team, R. C. (2014). *R: A Language and Environment for Statistical Computing.* R Foundation
 547 for Statistical Computing, Vienna, Austria.

548

549 Thompson, A. C., Feeney, C., & Kristal, M. B. (2018). Amniotic-fluid ingestion enhances central
 550 δ -opioid-induced hypoalgesia in rats in the cold-water tail-flick assay in a repeated-measures
 551 design. *Brain Res.* 1697, 53-58.

552

553 Trainor, B. C., Crean, K. K., Fry, W. H., & Sweeney, C. (2010). Activation of extracellular
 554 signal-regulated kinases in social behavior circuits during resident-intruder aggression
 555 tests. *Neuroscience.* 165, 325-336.

556

557 Walker, D. L., & Davis, M. (1997). Double dissociation between the involvement of the bed
 558 nucleus of the stria terminalis and the central nucleus of the amygdala in startle increases
 559 produced by conditioned versus unconditioned fear. *J Neurosci.* 17, 9375-9383.

560

561 Young, S. M., & Benyshek, D. C. (2010). In search of human placentophagy: a cross-cultural
562 survey of human placenta consumption, disposal practices, and cultural beliefs. *Ecol Food*
563 *Nutr.* 49, 467-484.

564

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576

577

578

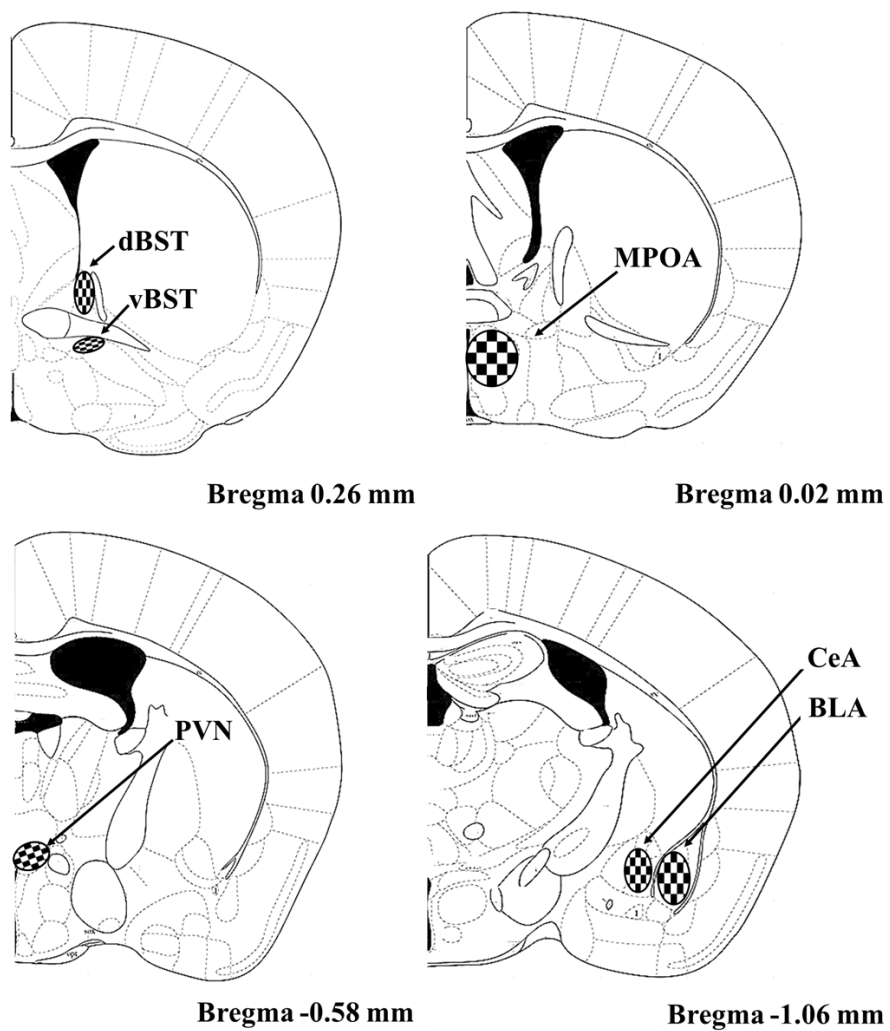
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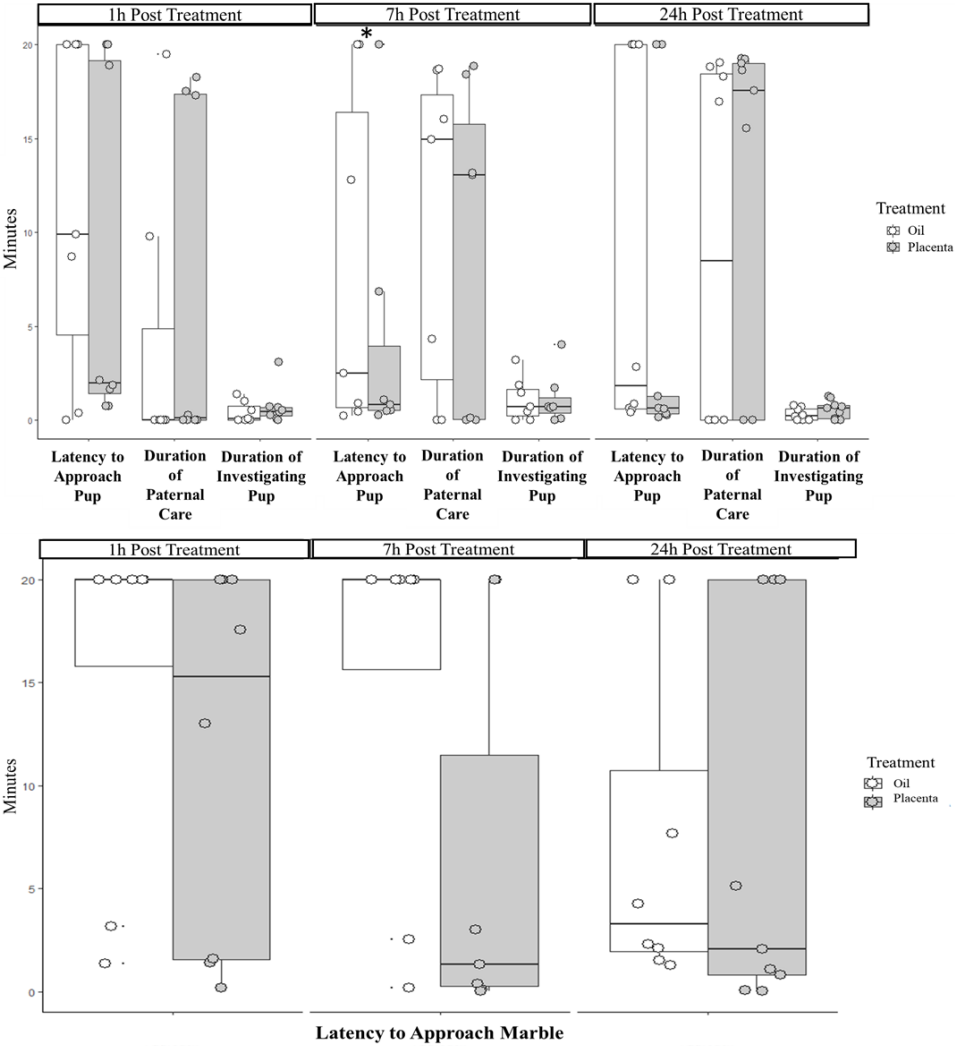
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583 **Figure 1:** Brain nuclei in which Fos-immunoreactivity was quantified. dBST: dorsal bed nucleus of the stria terminalis; vBST: ventral bed nucleus
584 of the stria terminalis; MPOA: medial preoptic area of the hypothalamus; PVN: paraventricular nucleus of the hypothalamus; BLA: basolateral
585 amygdala; CeA: central nucleus of the amygdala.
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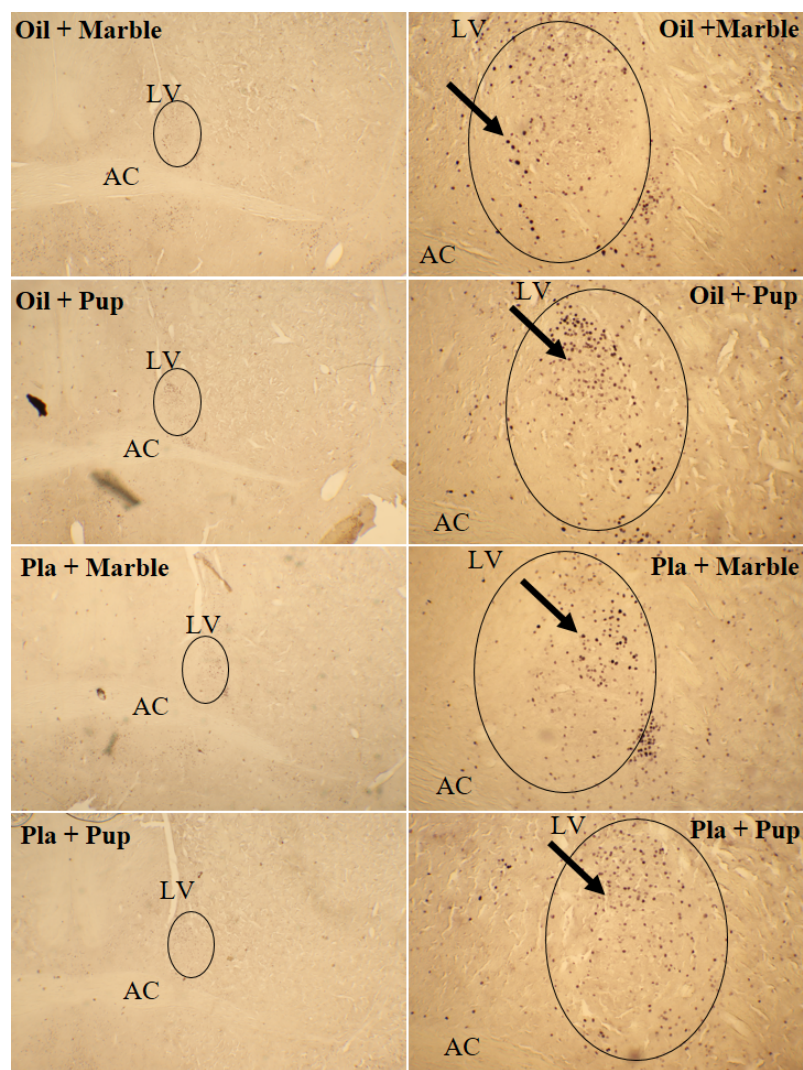
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589 **Figure 2:** Behavioral responses to a 1- to 4-day-old pup (top) and an oblong, pup-sized glass marble (bottom) by virgin male California mice 1, 7,
 590 or 24 after treatment with oil or placenta. Bars represent 1st quartiles, medians, and 3rd quartiles. The asterisk indicates a significant difference
 591 between treatments ($p \leq 0.05$).
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594 **Figure 3:** Representative photomicrographs of Fos labeling in the dorsal bed nucleus of the stria terminalis of virgin male California
595 mice exposed to a pup or a marble for 1 h, beginning 1 h following oral treatment with placenta or oil. Each image in the right-hand
596 column shows a higher magnification (100x) of the adjacent image (25x), with arrows indicating the black nuclear staining of Fos-
597 positive neurons. AC - anterior commissure; LV - lateral ventricle. Ovals indicate the area sampled.



599 **Table 1:** Sample sizes of placenta-treated and oil-treated virgin male California mice per time point and stimulus. Bold numbers
600 represent the number of mice tested with a pup that showed paternal behavior (huddling and licking pup).
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1 h Post-treatment		
	<u>Stimulus</u>	
Treatment	Pup	Marble
Placenta	8; 4	6
Oil	7; 2	6
7 h Post-treatment		
	<u>Stimulus</u>	
Treatment	Pup	Marble
Placenta	7; 5	6
Oil	7; 5	5
24 h Post-treatment		
	<u>Stimulus</u>	
Treatment	Pup	Marble
Placenta	9; 6	7
Oil	8; 4	7

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Table 2: Numbers of Fos-positive neurons following exposure to a pup or control object (marble) at each of three time points after treatment with placenta or oil. Data were analyzed using 2-way ANOVAs. Means, standard errors, and sample sizes are shown, as well as p-values for main effect of treatment, main effect of stimulus, and treatment x stimulus interaction. P-values ≤ 0.05 are shown in bold. MPOA – medial preoptic area of the hypothalamus, dBST – dorsal bed nucleus of the stria terminalis, vBST – ventral bed nucleus of the stria terminalis, PVN – paraventricular nucleus of the hypothalamus, BLA – basolateral amygdala, CeA – central nucleus of the amygdala.

Brain Area	Time Post-treatment	Marble		Pup		P-Value		
		Oil	Placenta	Oil	Placenta	Treatment	Stimulus	Treatment * Stimulus
MPOA	1h	14.75 $\pm$ 4.73 n=6	22.75 $\pm$ 6.10 n=6	27.83 $\pm$ 5.02 n=6	19.00 $\pm$ 3.25 n=6	0.93	0.35	0.10
	7h	10.10 $\pm$ 2.35 n=5	13.08 $\pm$ 3.21 n=6	7.33 $\pm$ 1.20 n=6	7.91 $\pm$ 2.85 n=6	0.46	0.13	0.64
	24h	15.60 $\pm$ 5.71 n=5	13.14 $\pm$ 2.81 n=7	17.62 $\pm$ 4.92 n=8	13.00 $\pm$ 2.49 n=8	0.65	0.85	0.78
dBST	1h	36.00 $\pm$ 5.48 n=6	27.58 $\pm$ 2.95 n=6	34.08 $\pm$ 4.40 n=6	24.25 $\pm$ 3.93 n=6	0.04	0.54	0.87
	7h	24.80 $\pm$ 5.42 n=6	26.91 $\pm$ 3.80 n=6	30.50 $\pm$ 2.86 n=6	12.80 $\pm$ 1.49 n=5	0.05	0.26	0.01
	24h	18.00 $\pm$ 5.83 n=5	27.57 $\pm$ 6.58 n=7	25.87 $\pm$ 5.56 n=8	24.5 $\pm$ 6.12 n=8	0.75	0.85	0.33
vBST	1h	12.33 $\pm$ 1.97 n=6	10.91 $\pm$ 1.43 n=6	11.33 $\pm$ 1.92 n=6	14.25 $\pm$ 1.27 n=6	0.78	0.40	0.27
	7h	8.60 $\pm$ 1.81 n=5	11.66 $\pm$ 2.11 n=6	10.08 $\pm$ 1.47 n=5	7.10 $\pm$ 0.92 n=5	0.49	0.80	0.93
	24h	9.87 $\pm$ 1.57 n=4	11.25 $\pm$ 1.25 n=2	10.33 $\pm$ 1.45 n=3	12.2 $\pm$ 3.10 n=5	0.49	0.80	0.93
PVN	1h	37.60 $\pm$ 4.61 n=5	27.7 $\pm$ 2.22 n=6	53.83 $\pm$ 11.03 n=6	34.54 $\pm$ 6.65 n=6	0.07	0.07	0.37
	7h	22.90 $\pm$ 2.14 n=5	27.36 $\pm$ 4.01 n=6	20.58 $\pm$ 6.56 n=6	24.16 $\pm$ 5.06 n=6	0.41	0.59	0.93

	24h	24.85 ± 3.54 n=7	23.50 ± 5.50 n=6	26.57 ± 5.69 n=8	35.42 ± 5.38 n=7	0.72	0.23	0.31
BLA	1h	24.83 ± 2.52 n=6	24.00 ± 4.52 n=6	39.91 ± 5.43 n=6	30.41 ± 6.63 n=6	0.31	0.04	0.39
	7h	21.90 ± 5.08 n=5	20.41 ± 3.23 n=6	21.08 ± 5.85 n=6	20.90 ± 1.17 n=5	0.85	0.99	0.88
	24h	21.33 ± 3.17 n=6	29.00 ± 5.95 n=7	33.60 ± 3.60 n=5	36.66 ± 11.02 n=6	0.60	0.21	0.69
	1h	28.41 ± 2.33 n=6	27.50 ± 2.50 n=6	41.41 ± 3.46 n=6	31.16 ± 4.98 n=6	0.12	0.02	0.19
CEA	7h	40.40 ± 6.23 n=5	43.83 ± 7.65 n=6	30.83 ± 4.20 n=6	28.00 ± 5.16 n=5	0.30	0.33	0.65
	24h	28.66 ± 4.40 n=6	30.28 ± 2.54 n=7	28.00 ± 5.16 n=5	33.16 ± 9.32 n=6	0.81	0.88	0.80

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