

UC Davis

UC Davis Previously Published Works

Title

The role of brown adipose tissue in branched-chain amino acid clearance in people

Permalink

<https://escholarship.org/uc/item/80m542jh>

Journal

iScience, 27(8)

ISSN

2589-0042

Authors

Abdelhafez, Yasser G

Wang, Guobao

Li, Siqi

et al.

Publication Date

2024-08-01

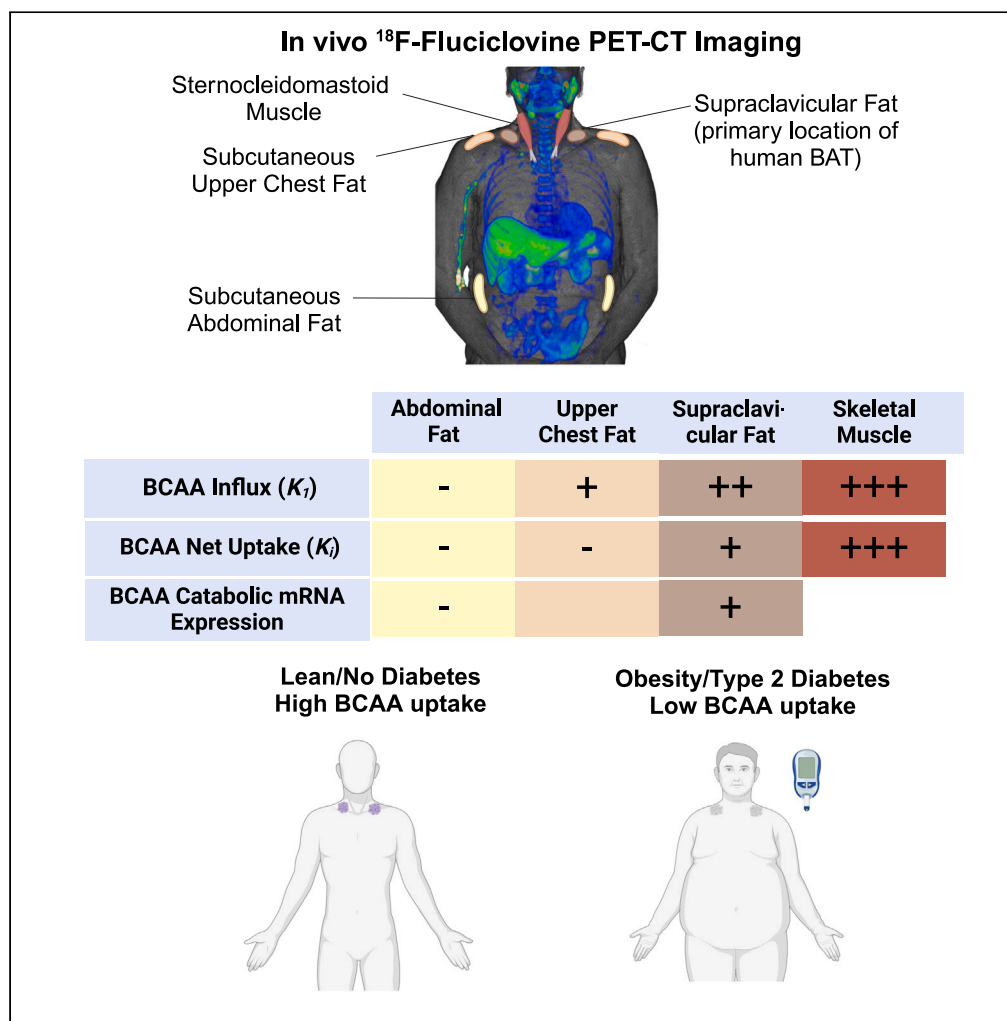
DOI

10.1016/j.isci.2024.110559

Peer reviewed

Article

The role of brown adipose tissue in branched-chain amino acid clearance in people



Yasser G. Abdelhafez, Guobao Wang, Siqi Li, ..., Samuel Klein, Ramsey D. Badawi, Maria Chondronikola

mc2425@medschl.cam.ac.uk

Highlights

Human BAT clears fluciclovine (BCAA analog for PET imaging) faster than abdominal AT

BCAA uptake in BAT is lower in people with obesity and type 2 diabetes

BCAA catabolic genes expression is higher in the BAT of people with high BAT volume



Article

The role of brown adipose tissue in branched-chain amino acid clearance in people

Yasser G. Abdelhafez,^{1,2} Guobao Wang,¹ Siqi Li,¹ Vanessa Pellegrinelli,³ Abhijit J. Chaudhari,¹ Anthony Ramirez,⁴ Fatma Sen,¹ Antonio Vidal-Puig,³ Labros S. Sidossis,⁵ Samuel Klein,⁶ Ramsey D. Badawi,¹ and Maria Chondronikola^{1,3,4,7,8,*}

SUMMARY

Brown adipose tissue (BAT) in rodents appears to be an important tissue for the clearance of plasma branched-chain amino acids (BCAAs) contributing to improved metabolic health. However, the role of human BAT in plasma BCAA clearance is poorly understood. Here, we evaluate patients with prostate cancer who underwent positron emission tomography-computed tomography imaging after an injection of ¹⁸F-fluciclovine (L-leucine analog). Supraclavicular adipose tissue (AT; primary location of human BAT) has a higher net uptake rate for ¹⁸F-fluciclovine compared to subcutaneous abdominal and upper chest AT. Supraclavicular AT ¹⁸F-fluciclovine net uptake rate is lower in patients with obesity and type 2 diabetes. Finally, the expression of genes involved in BCAA catabolism is higher in the supraclavicular AT of healthy people with high BAT volume compared to those with low BAT volume. These findings support the notion that BAT can potentially function as a metabolic sink for plasma BCAA clearance in people.

INTRODUCTION

Branched-chain amino acids (BCAAs: leucine, valine, and isoleucine) are essential amino acids that have a central carbon atom with an aliphatic side chain of three or more carbons (branch) attached to it. Apart from their role as structural building blocks of proteins, BCAA act as signaling molecules regulating numerous important metabolic processes (e.g., protein synthesis, insulin secretion, etc.).¹ High plasma BCAA concentration has been associated with insulin resistance and is thought to contribute to the pathogenesis of type 2 diabetes (T2D)^{2–5} and other cardiometabolic conditions.^{6,7} Although there has been intense scientific interest in understanding how BCAAs affect metabolic health, their mechanism of action is not completely understood.

Brown adipose tissue (BAT) has been proposed as a potential target for the prevention of cardiometabolic disease, due to its high capacity for substrate oxidation to produce heat.⁸ Although circulating lipids and glucose are the major substrates for BAT thermogenesis,⁹ more recent data suggest that BAT is also an avid consumer of plasma BCAA.^{10–13} Increased BCAA catabolism in BAT has been shown to increase insulin sensitivity and reduce adiposity in rodents.¹⁰ Early evidence from clinical investigations also supports the link between BAT and BCAA metabolism in people.^{10,14} However, these studies lack tissue-specific evidence supporting the direct involvement of BAT in the regulation of BCAA metabolism and its quantitative contribution in plasma BCAA clearance.

To this end, we conducted a proof-of-concept study aiming to determine whether human BAT has higher uptake rate for BCAA compared to other subcutaneous adipose tissue depots. We hypothesized that supraclavicular adipose tissue (the primary location of human BAT¹⁵) would demonstrate a higher BCAA uptake rate and a higher expression of genes involved in BCAA catabolism compared to subcutaneous abdominal and/or upper chest white adipose tissue. ¹⁸F-fluciclovine is a synthetic L-leucine analog positron emission tomography-computed tomography (PET-CT) radiotracer¹⁶ that has been developed and clinically used as cancer imaging agent.¹⁷ To this end, ¹⁸F-fluciclovine total-body PET/CT dynamic scans, conducted for clinical purposes, were analyzed using tracer kinetic modeling to evaluate BCAA kinetics in the supraclavicular, subcutaneous upper chest and abdominal adipose tissue. Furthermore, we assessed the expression of key genes involved in BCAA catabolism in supraclavicular and subcutaneous abdominal adipose tissue in a distinct cohort of healthy adults with overweight/obesity.

¹Department of Radiology, University of California Davis, Sacramento, CA 95817, USA

²Nuclear Medicine Unit, South Egypt Cancer Institute, Assiut University, El Fateh 71111, Egypt

³Institute of Metabolic Science-Metabolic Research Laboratories, Medical Research Council Metabolic Diseases Unit, University of Cambridge, Cambridge CB2 0QQ, UK

⁴Department of Nutrition, University of California Davis, Davis, CA 95616, USA

⁵Department of Kinesiology and Health, Rutgers University, New Brunswick, NJ 08901, USA

⁶Center for Human Nutrition, Washington University School of Medicine, Saint Louis, MO 63110, USA

⁷Department of Nutrition and Dietetics, Harokopio University of Athens, 17778 Athens, Greece

⁸Lead contact

*Correspondence: mc2425@medschl.cam.ac.uk

<https://doi.org/10.1016/j.isci.2024.110559>



Table 1. Patient demographic and anthropometric characteristics

Parameters	<i>n</i> = 36
Age (years)	72.2 ± 8.3
Race (White/unknown)	33/3
Ethnicity (non-Hispanic/Asian/unknown)	33/1/2
BMI (kg/m ²)	27.9 ± 5.2
Weight (kg)	88.4 ± 19.3
Height (cm)	178 ± 8

Data are means ± SD. Related also to [Table S1](#).

RESULTS

Participant characteristics

We identified 36 adult male patients with histopathologically proven prostate adenocarcinoma who underwent ¹⁸F-fluciclovine PET-CT imaging as part of their standard clinical care. The study participants were older, mostly white, and non-Hispanic ([Table 1](#)). The body mass index (BMI) of the participants ranged from the normal to the obese BMI category. The prevalence of T2D diagnosis in the study population was 13.8% (*n* = 5).

Tissue-specific metabolic assessment using ¹⁸F-fluciclovine PET-CT imaging

To evaluate whether human BAT has a high capacity for BCAA uptake, we first assessed the mean standardized uptake value for ¹⁸F-fluciclovine (standardized uptake value [SUV_{mean}]: index of ¹⁸F-fluciclovine uptake determined using static PET imaging of a single time frame 4–14 min post-injection) in supraclavicular adipose tissue (the primary location of human BAT¹⁵), the upper chest (upper body subcutaneous adipose tissue depot), and subcutaneous abdominal adipose tissue (classical white adipose tissue depot¹⁸) ([Figures 1A–1C](#)). The supraclavicular adipose tissue had the highest SUV_{mean} for ¹⁸F-fluciclovine compared to the subcutaneous upper chest and abdominal adipose tissue ([Figure 2A](#)). Additionally, the SUV_{mean} for ¹⁸F-fluciclovine was higher in the upper chest compared to the abdominal adipose tissue ([Figure 2A](#)). The radiodensity (which correlates negatively with lipid content and positively with arterial blood volume¹⁹) of the three adipose tissue depots was also assessed using the CT imaging data. Tissue radiodensity was (marginally) higher in the supraclavicular and upper chest adipose tissue compared to the abdominal adipose tissue, while it was similar between the upper chest and the supraclavicular adipose tissue ([Figure 2B](#)).

A two-tissue compartmental model ([Figure S1](#)) has been implemented to quantitatively describe ¹⁸F-fluciclovine kinetics in the three adipose tissue depots of interest. Considering that skeletal muscle is a major tissue responsible for the BCAA plasma clearance,¹¹ ¹⁸F-fluciclovine kinetics were also assessed in the sternocleidomastoid muscle as a reference tissue. [Figures 3A–3B](#) present the time-activity curves for ¹⁸F-fluciclovine in aorta, sternocleidomastoid muscle, and the three adipose tissue depots of interest. Supraclavicular adipose tissue had the highest net uptake rate *Ki* (which provides an assessment of the clearance of ¹⁸F-fluciclovine from blood to tissue) compared to the subcutaneous abdominal and upper chest adipose tissue ([Figure 3C](#)). Upper chest adipose tissue had a higher net uptake rate *Ki* for ¹⁸F-fluciclovine compared to the abdominal adipose tissue. *K1* (which provides an assessment of tracer delivery rate to the tissue and partially reflects tissue perfusion, [Figure 3D](#)) and *k3* (which provides an assessment of the flux rate from interstitial to intracellular compartment, [Figure 3E](#)) parameters were also higher in supraclavicular compared to the abdominal adipose tissue. Additionally, the supraclavicular adipose tissue had a higher *K1* compared to the upper chest adipose tissue, but the *k3* was similar between the two tissues ([Figures 3D](#) and [3E](#)). Skeletal muscle had higher ¹⁸F-fluciclovine delivery rate *K1*, net uptake rate *Ki*, and *k3* flux rate from interstitial to intracellular compartment compared to all adipose tissue depots assessed ([Figures 3C–3E](#)). Supraclavicular adipose tissue SUV_{mean} and net uptake rate *Ki* were not different among participants imaged during the four different seasons ([Figures S2A](#) and [S2B](#)). Subsequently, we investigated the interrelationships between supraclavicular adipose tissue ¹⁸F-fluciclovine kinetic parameters and radiodensity. Supraclavicular adipose tissue ¹⁸F-fluciclovine net uptake rate *Ki* positively correlated with radiodensity, SUV_{mean}, and ¹⁸F-fluciclovine delivery rate *K1* ([Figures S3A–S3C](#)). Supraclavicular adipose tissue SUV_{mean} for ¹⁸F-fluciclovine correlated with tissue radiodensity, while tissue radiodensity marginally and weakly correlated with ¹⁸F-fluciclovine delivery rate *K1* ([Figures S3D](#) and [S3E](#)).

Plasma BCAA clearance as a biomarker of adiposity and metabolic health

Considering that increased plasma BCAA catabolism in BAT has been implicated in the development of adiposity and insulin resistance,¹⁰ we evaluated the link between ¹⁸F-fluciclovine net uptake rate *Ki* in the supraclavicular adipose tissue with adiposity and T2D. There was a progressive decrease of ¹⁸F-fluciclovine net uptake rate *Ki* in the supraclavicular adipose tissue from the normal to the obese BMI category ([Figure 4A](#)), while BMI was inversely correlated with the ¹⁸F-fluciclovine net uptake rate in the supraclavicular adipose tissue ([Figure 4B](#)). Further, patients with diagnosis of T2D had significantly lower supraclavicular adipose tissue ¹⁸F-fluciclovine net uptake rate compared to participants without diagnosis of T2D ([Figure 4C](#)). *Post hoc* exploration of the study results revealed that BMI was negatively correlated with net uptake

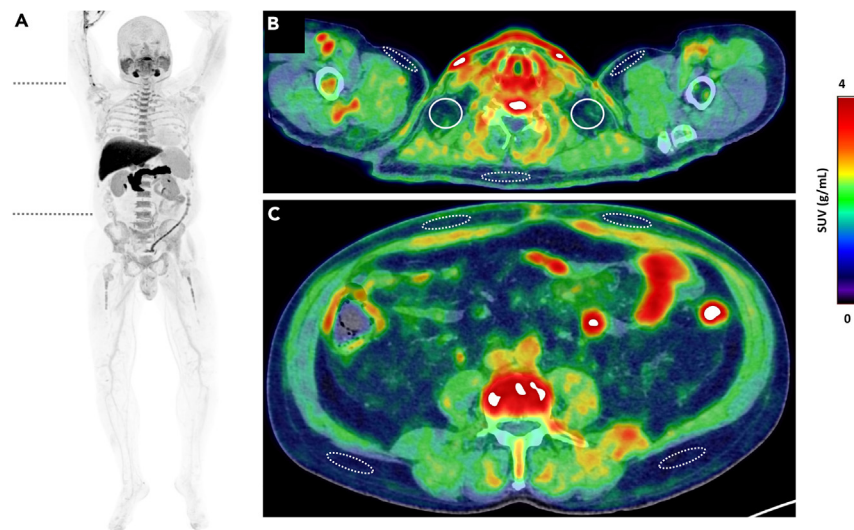


Figure 1. Normal ^{18}F -fluciclovine biodistribution

Maximum intensity projection image of a 61-year-old man (183 cm, 98 kg) with biochemical recurrence of prostate cancer, demonstrating physiologic ^{18}F -fluciclovine uptake, being most prominent in the pancreas, liver, salivary glands, and bone marrow. The two transverse dashed lines on (A) point to the level of cross-sectional axial fused PET/CT images (B) and (C), demonstrating supraclavicular adipose tissue depot (solid circles, B), upper chest subcutaneous adipose tissue (dashed ellipses, B), and abdominal subcutaneous adipose tissue depots at various locations opposite third lumbar vertebra (dashed ellipses, C). Visually, the supraclavicular regions show subtle higher ^{18}F -fluciclovine uptake compared to other regions shown in (B) and (C). The difference was more evident on quantification.

rate K_i for ^{18}F -fluciclovine in the upper chest adipose tissue ($r = -0.43$, $p = 0.01$) and the ^{18}F -fluciclovine delivery rate K_1 in the abdominal adipose tissue ($r = -0.43$, $p = 0.011$). BMI was not correlated with the net uptake rate for ^{18}F -fluciclovine in abdominal adipose tissue ($r = -0.17$, $p = 0.33$) and skeletal muscle ($r = -0.075$, $p = 0.67$). Similarly, BMI was not correlated with the ^{18}F -fluciclovine delivery rate K_1 in upper chest adipose tissue ($r = 0.13$, $p = 0.46$), supraclavicular adipose tissue ($r = -0.20$, $p = 0.25$), or skeletal muscle ($r = 0.16$, $p = 0.35$).

Expression of genes involved in BCAA metabolism in the supraclavicular and subcutaneous abdominal adipose tissue

To further assess the role of human BAT in BCAA metabolism, we evaluated the expression of key genes involved in BCAA mitochondrial uptake and breakdown in supraclavicular and subcutaneous abdominal adipose tissue in a different cohort of healthy men and women with overweight/obesity (Table S1). The expression of *Uncoupling Protein 1* (*UCP1*, which encodes the major thermogenic protein)⁹ and *Solute Carrier Family 25 Member 44* (*SLC24A44*, which encodes a key mitochondrial carrier protein involved in the transport of BCAA from the cytosol into the mitochondria¹⁰) were higher in the supraclavicular compared to the abdominal adipose tissue (Figures 5A and 5B). The expression of *Branched-Chain Keto Acid Dehydrogenase E1 Subunit Beta* (*BCKDHB*), which encodes a subunit of a multienzyme mitochondrial complex involved in the catabolism of BCAA,¹⁰ was about 50% (but not statistically significantly higher) in the supraclavicular compared to the subcutaneous abdominal adipose tissue (Figure 5C). *Branched-chain-amino-acid transaminase 1* (*BCAT1*, which encodes the cytosolic enzyme that converts BCAA to branched-chain keto acids in the cytosol²⁰) expression was similar in the supraclavicular and abdominal adipose tissue, while *branched-chain-amino-acid transaminase 2* (*BCAT2*, which encodes the enzyme that converts BCAA to branched-chain keto acids in the mitochondria²⁰) expression was higher in the abdominal adipose tissue compared to the supraclavicular adipose tissue depot (Figures 5D and 5E). Finally, we compared the expression of the previously mentioned genes in the supraclavicular adipose tissue of participants with high 2-Deoxy-2- ^{18}F -fluorodeoxyglucose PET-detectable BAT volume ≥ 20 mL (HBAT) and participants with low BAT volume < 20 mL (LBAT).^{21–23} We observed no statistically significant differences in terms of age and anthropometric and demographic characteristics between the two groups (Table S1). The expression of the *UCP1* gene along with the expression of *BCKDHB* and *SLC24A44* was higher in the supraclavicular adipose tissue of HBAT group compared to the LBAT group (Figures 5F–5H). The *BCAT1* and *BCAT2* gene expression was similar in the two groups (Figures 5I and 5J). *UCP1* expression in the supraclavicular adipose tissue depot was positively correlated with *SLC24A44* ($r = 0.38$, $p = 0.07$) and *BCKDHB* expression ($r = 0.44$, $p = 0.04$), but not with the expression of *BCAT1* ($r = 0.17$, $p = 0.44$) and *BCAT2* ($r = -0.04$, $p = 0.86$).

DISCUSSION

Increased plasma BCAA concentrations have been linked to the pathogenesis of T2D^{2–4}; hence understanding the mechanism(s) regulating BCAA metabolism can have important implications for the development of effective strategies for the prevention of T2D. Here, we report that the supraclavicular adipose tissue—the primary location of human BAT¹⁵—can more effectively remove BCAA from the circulation compared

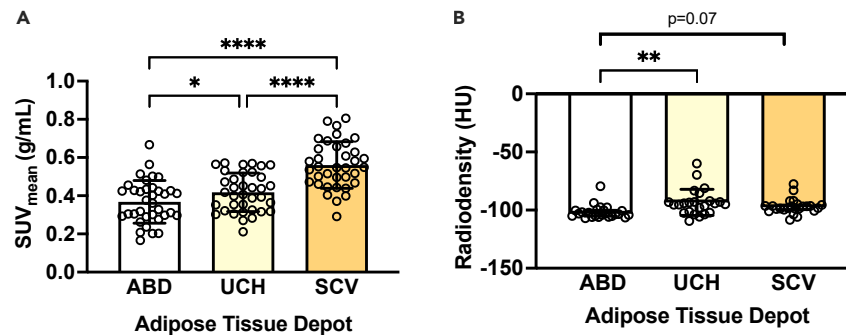


Figure 2. Tissue-specific ^{18}F -fluciclovine uptake assessment and CT density

(A) SUV_{mean} and (B) radiodensity of various tissues assessed by using ^{18}F -fluciclovine positron emission tomography-computed tomography ($n = 36$). Data are presented as mean \pm SD (for normally distributed data) or median (interquartile range) for skewed data. p values were determined by using analysis of variance (for normally distributed data) with Tukey correction for multiple comparisons or Friedman test (for skewed data) with Dunn correction for multiple comparisons. ABD,; abdominal; CT, computed tomography; HU, Hounsfield units; SCV, supraclavicular; UCH, upper chest; SUV, standardized uptake value. * $p < 0.05$, ** $p < 0.01$, and **** $p < 0.0001$.

to other subcutaneous adipose tissue depots investigated in this study. The higher BCAA net uptake rate in supraclavicular adipose tissue was attributable both to the increased perfusion rate of the tissue and to its intrinsic metabolic characteristics. The supraclavicular adipose tissue BCAA net uptake rate was inversely correlated with tissue radiodensity, while it was lower among people with increased adiposity and T2D. Consistent with these findings, transcriptional analysis of supraclavicular adipose tissue and subcutaneous adipose samples, collected from a different cohort of participants, further indicates that BAT is a tissue with a high capacity for BCAA catabolism compared to subcutaneous abdominal adipose tissue. Although BAT *per se* may have a higher capacity for plasma BCAA clearance compared to other subcutaneous adipose tissue depots, it is unlikely to have a major effect in regulating plasma BCAA concentrations due to its small measurable volume in people.

The findings of this study support the notion that human BAT may be an adipose tissue depot with increased ability to remove BCAA from the circulation compared to upper chest and subcutaneous abdominal adipose tissue depots. The analysis performed using dynamic ^{18}F -fluciclovine PET-CT imaging data indicates that the supraclavicular adipose tissue has the highest net uptake rate for ^{18}F -fluciclovine (K_i) compared to the subcutaneous upper chest and abdominal adipose tissue. The transcriptional data from a different cohort of healthy participants are consistent with ^{18}F -fluciclovine kinetic data. The expression of the *SLC25A44* (a key mitochondrial carrier protein involved in the transport of BCAA from the cytosol into the mitochondria¹⁰) and *UCP1* was higher in the supraclavicular compared to abdominal adipose tissue. Similarly, the expression of genes involved in thermogenesis and BCAA catabolism (i.e., *SLC25A44* and *BCKDHB*), but not *BCAT2*, was higher in the supraclavicular adipose tissue of participants with high BAT volume compared to those with low BAT volume. The expression of *BCAT2* was higher in the abdominal adipose tissue compared to the supraclavicular adipose tissue. *BCAT2* encodes the mitochondrial enzyme that catalyzes the reversible transamination of BCAA to branch-chained keto acids (first step in BCAA oxidation pathway), whereas *BCKDHB* encodes the subunit of a multienzyme mitochondrial complex that catalyzes the oxidative decarboxylation branched-chained keto acids (the second and rate-limiting step in BCAA catabolism).²⁰ Interestingly, *UCP1* expression in supraclavicular adipose tissue was positively correlated with *SLC25A44* and *BCKDHB* gene expression suggesting that BCKDH and *SLC25A44* may be involved in the regulation of thermogenesis in BAT as it has been previously reported in rodents.²⁴ Although decreased *BCAT2* activity in white adipose tissue rodents has been associated with the browning of white adipose tissue and prevention of obesity in rodents,²⁴ we found no relationship between *BCAT2* and *UCP1* expression in the supraclavicular adipose tissue depot.

To evaluate the physiological mechanisms mediating the high net uptake rate (K_i) for ^{18}F -fluciclovine observed in the supraclavicular adipose tissue compared to the other adipose tissue depots investigated, we assessed the K_1 tracer delivery rate (marker of perfusion) and k_3 flux rate from interstitial to intracellular compartment for target tissue. The supraclavicular adipose tissue has a higher K_1 tracer delivery rate and k_3 flux rate from interstitial to intracellular compartment compared to the abdominal adipose tissue, suggesting that both intrinsic tissue-specific metabolic characteristics and higher perfusion underlie the higher net uptake (K_i) for ^{18}F -fluciclovine reported in the supraclavicular adipose tissue compared the abdominal adipose tissue. Conversely, supraclavicular adipose tissue has a higher K_1 tracer delivery rate compared to the upper chest adipose tissue, but a similar ^{18}F -fluciclovine flux rate k_3 from interstitial to intracellular compartment, suggesting that increased perfusion may explain the higher net uptake rate detected in the supraclavicular adipose tissue depot compared to the upper chest adipose tissue.

The results of this study advance the current understanding on the role of BAT in the regulation of BCAA metabolism. BAT in rodents is thought to be responsible for $\sim 20\%$ of whole-body BCAA oxidation in room temperature conditions (mild cold for rodents) suggesting that BAT is the organ with the second highest contribution to whole-body BCAA oxidation after skeletal muscle¹¹; hence its contribution in thermoneutrality is estimated to be smaller. Consistent with these findings, Yoneshiro et al. reported that plasma BCAA catabolism in BAT is implicated in the BAT thermogenesis and the development of adiposity and insulin resistance.¹⁰ The role of human BAT in BCAA metabolism has

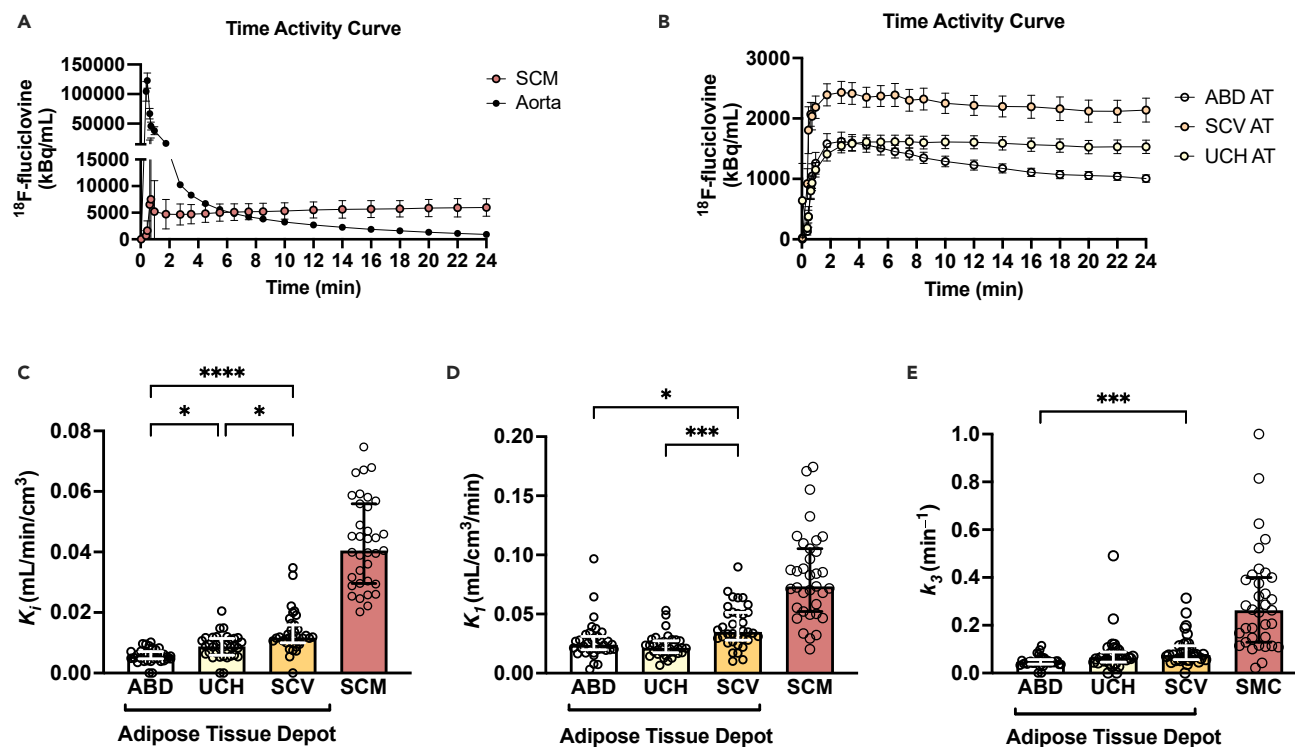


Figure 3. ^{18}F -fluciclovine kinetics in the different adipose tissue depots using PET imaging

(A and B) PET time-activity curves over the 24 min of acquisition after ^{18}F -fluciclovine injection in (A) blood pool in the ascending aorta (as input function) and sternocleidomastoid muscle as reference tissue as well as (B) in the adipose tissue depots of interest ($n = 35$). Each time point represents a mid-frame value. Data are means \pm SEM.

(C) Tracer delivery rate (K_1 ; index of tissue perfusion) in the different adipose tissue depots ($n = 35$).

(D) Net uptake rate (K) for ^{18}F -fluciclovine in the different adipose tissue depots ($n = 35$).

(E) Flux rate from interstitial to intracellular compartment (k_3) for ^{18}F -fluciclovine in the different adipose tissue depots ($n = 35$). Parametric analysis was not performed for one participant due to lack of imaging data for the first 4 min after injection. Data are median and interquartile range. p values were determined by using the Friedman test with Dunn correction for multiple comparisons. ABD, abdominal; SCM, sternocleidomastoid muscle; SCV, supraclavicular; UCH, upper chest; PET, positron emission tomography. * $p < 0.05$, *** $p < 0.001$, and **** $p < 0.0001$. Related also to [Figures S1–S3](#).

been studied only indirectly. Namely, acute cold exposure has been shown to decrease circulating BCAA levels in lean men with high BAT activity,¹⁰ while the ratio of fasting BCAA to total amino acid concentration has been correlated with total hemoglobin concentration in the supraclavicular adipose tissue (marker of BAT vascular density) in thermoneutrality ($\sim 24^\circ\text{C}$).¹⁴ Our findings are consistent with previous studies in rodents regarding the high capacity of BAT for BCAA uptake; hence we expect that the absolute contribution of BAT in the regulation of whole-body plasma BCAA clearance will be small considering that skeletal muscle has approximately a 3-fold higher uptake rate per volume of tissue compared to supraclavicular adipose tissue and is several fold larger in volume. In this study, we were unable to assess the total contribution of BAT in plasma BCAA clearance as the patients did not perform ^{18}F -FDG PET-CT imaging that would enable us to quantify BAT volume,²⁵ and arterial blood samples were not collected to allow the absolute quantification of leucine uptake. However, our results provide some estimates on the contribution of BAT in BCAA clearance in the absence of cold stimulation. Assuming BAT volume is $\sim 100\text{ cm}^3$ and a net uptake rate for ^{18}F -fluciclovine in the supraclavicular depot of $0.012\text{ mL/min per cm}^3$ of tissue, we estimate that BAT could clear about 1.2 mL of plasma from BCAA per minute, while skeletal muscle (assuming $\sim 30\text{ kg}$ muscle mass) could clear about 1.2 L of plasma from BCAA per minute (about 1,000-fold higher from BAT). The metabolic fate of BCAA in BAT was beyond the scope of this study, but BCAAs in BAT can be used for oxidation and TCA cycle anaplerosis or for the synthesis of intracellular lipids (TCA cycle cataplerosis).²⁶

This study also provides insights on the potential clinical significance of the BCAA clearance by BAT in the adiposity and metabolic health. Specifically, we report that the ^{18}F -fluciclovine net uptake rate in supraclavicular adipose tissue gradually decreases with adiposity, and it is inversely correlated with BMI. The correlation between BCAA uptake in supraclavicular adipose tissue and the radiodensity of the tissue—an index of the tissue lipid content¹⁹—further corroborates the relationship between BCAA clearance rate in BAT and lipid accumulation. BMI was not correlated with ^{18}F -fluciclovine delivery rate K_1 (index of tissue perfusion) in the supraclavicular adipose tissue, but it was correlated with ^{18}F -fluciclovine delivery rate in the abdominal adipose tissue, as it has been previously reported.^{27,28} These results suggest that the lower ^{18}F -fluciclovine net uptake rate in supraclavicular adipose tissue in people with high BMI is not attributable to an adiposity-related lower tissue perfusion. Further, we report that participants with diagnosis of T2D have lower BCAA clearance rate in supraclavicular adipose tissue

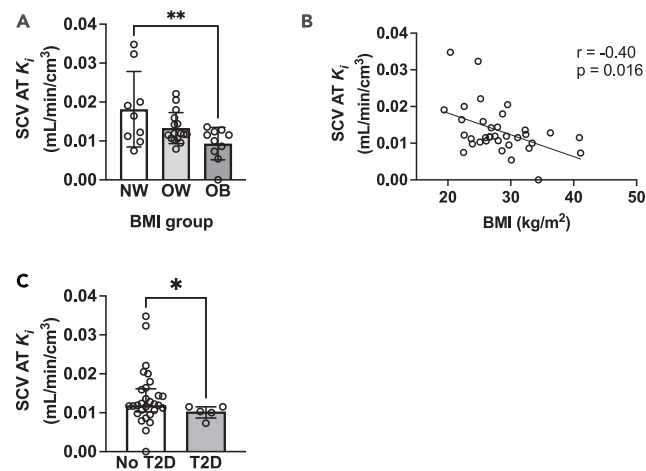


Figure 4. ¹⁸F-fluciclovine net uptake rate in supraclavicular adipose tissue as a marker of adiposity and metabolic health

(A) SCV adipose tissue net uptake rate for ¹⁸F-fluciclovine in patients in the normal (n = 9), overweight (n = 16), and obese (n = 10) BMI category. p values determined by using one-way analysis of variance with Tukey correction for multiple comparisons.

(B) Correlation between SCV adipose tissue net uptake rate for ¹⁸F-fluciclovine and BMI (n = 35). Spearman's rho was used to evaluate the correlation between the different variables of interest.

(C) SCV adipose tissue net uptake rate for ¹⁸F-fluciclovine in patients with (n = 5) and without diagnosis of T2D (n = 29). The medical record of one of the participants was not available. p value was determined by using Mann-Whitney U test. Data are means ± SD for normally distributed data or median (interquartile range) for skewed data. AT, adipose tissue; BMI, body mass index; K_i , net uptake rate reflecting clearance from blood; NW, normal weight; OB, obesity; OW, overweight; SCV, supraclavicular; T2D, type 2 diabetes. *p < 0.05 and **p < 0.01.

compared to those without diagnosis of T2D, suggesting that BCAA clearance rate in supraclavicular adipose tissue can be a marker of metabolic health, as it has been suggested for glucose.²⁹ Our findings are consistent with data from studies in rodents. Specifically, Yoneshiro et al. have previously reported that BCAA clearance in BAT is implicated in the development of adiposity and insulin resistance,¹⁰ while Samms et al. have recently reported that tirzepatide (a dual glucose-dependent insulinotropic polypeptide and glucagon-like peptide-1 receptor agonist known to improve glycemia leading to weight loss) also increases BCAA catabolism in BAT. The exact mechanism underlying the link between BCAA clearance in BAT and metabolic health remains to be clarified in light of the recent findings suggesting that leucine, isoleucine, and valine appear to have distinct metabolic roles in the regulation of metabolic processes relevant to the pathogenesis of T2D.^{30–32}

Our findings also indicate that the subcutaneous upper chest adipose tissue depot has a distinctive phenotype in terms of BCAA kinetics and radiodensity compared to the supraclavicular and abdominal adipose tissue depots. Specifically, subcutaneous upper chest adipose tissue has an intermediate phenotype between the subcutaneous abdominal and supraclavicular adipose tissue depot in terms of SUV_{mean} , K_1 (index of tissue perfusion) and K_i (net uptake rate) for ¹⁸F-fluciclovine. We also found that differences in tissue perfusion may explain the higher net tissue uptake rate K_i for ¹⁸F-fluciclovine reported in supraclavicular adipose tissue compared to upper chest adipose tissue, as the k_3 flux rate from interstitial to intracellular compartment for upper chest and supraclavicular adipose tissue depot were similar. The net uptake rate K_i for ¹⁸F-fluciclovine was also correlated with BMI, as it was in the supraclavicular adipose tissue. Upper chest adipose tissue radiodensity was higher than the subcutaneous abdominal adipose tissue depot, but not the supraclavicular adipose tissue depot. These findings provide insights into the heterogeneity of the adipose tissue organ³³ in people and suggest that different metabolic characteristics assessed in one adipose tissue depot do not necessarily extrapolate to the rest of the adipose tissue depots. Consistent with these results, Cypress et al. have previously reported on the variability in gene expression, differentiation capacity, and oxygen consumption among various adipose tissue depots in human neck.³⁴ Our results suggest that this may be true for other upper body adipose tissue depots, and, thus, further research is needed to characterize the heterogeneity between the different adipose tissue depots in humans.

In summary, we provided proof-of-concept *in vivo* functional and transcriptional evidence supporting the notion that supraclavicular adipose tissue—the primary BAT depot in humans—has a higher capacity for BCAA clearance rate in people compared to other subcutaneous adipose tissue depots. In addition, our findings support that supraclavicular adipose tissue BCAA net uptake in people constitutes a biomarker for lower adiposity and improved metabolic health. Further research is needed to better understand the contribution of BAT and the thermogenic adipocytes in adipose tissue depots in whole-body BCAA metabolism and its suggested role in the prevention of the obesity-related metabolic complications.

Limitations of the study

This study provides functional evidence of the ability of the supraclavicular adipose tissue depot to take up BCAA in people. However, it has limitations. ¹⁸F-fluciclovine is a leucine analog that is transported in the cells via transporters (i.e., L-type amino acid transporter 1 and the

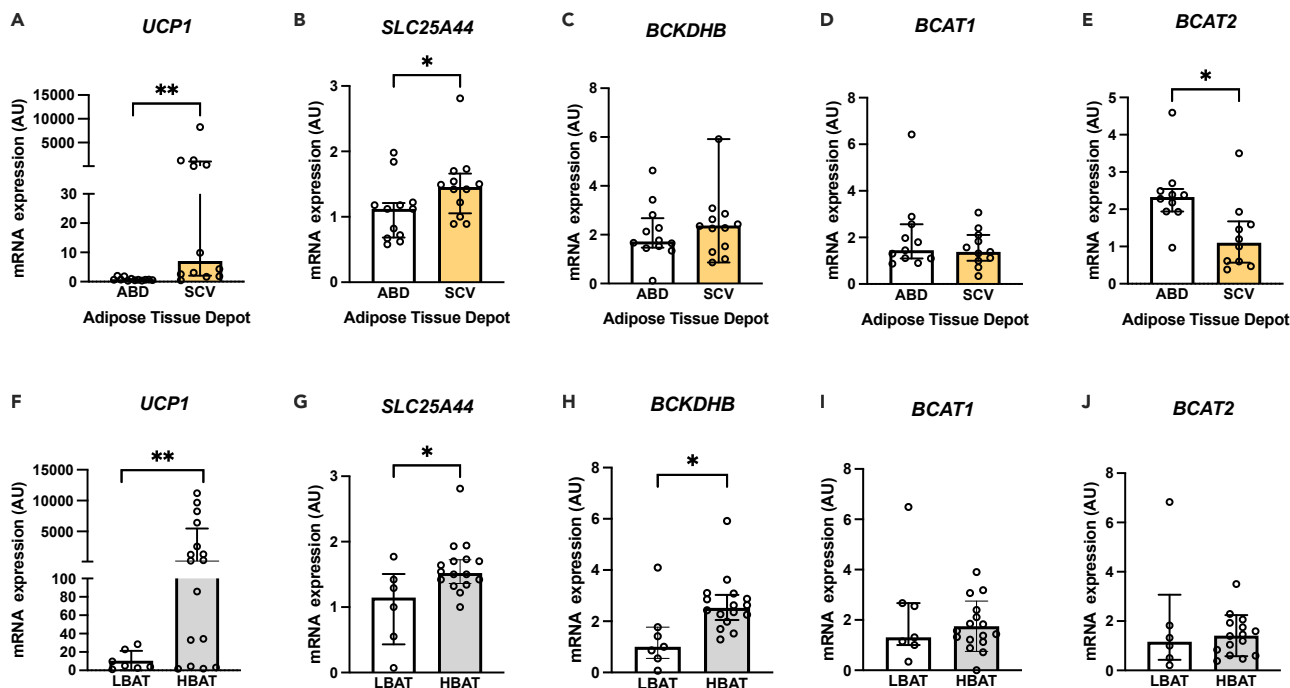


Figure 5. Expression of genes involved in BCAA metabolism: adipose tissue depot-specific variation and BAT status

(A–E) Expression of genes involved in BCAA metabolism and thermogenesis in paired supraclavicular and abdominal adipose tissue samples ($n = 10–12$). p values were determined by using the Wilcoxon signed-rank test.

(F–J) Expression of genes involved in BCAA catabolism and thermogenesis in the supraclavicular adipose tissue depot of people with low ($n = 6–7$) and high ($n = 15–16$) BAT volume (LBAT and HBAT, respectively). Data are median and interquartile range. p values were determined by using the Mann-Whitney test. The smaller number of observations in (D), (E), and (J) are due to limited sample availability. BAT, brown adipose tissue; BCAA, branched-chain amino acids; BCAT1/2, branched-chain amino acid aminotransferase 1 or 2; BCKDHB, branched-chain keto acid dehydrogenase E1 subunit beta; HBAT, group of participants with brown adipose tissue volume equal to or more than 20 mL; LBAT, group of participants with brown adipose tissue volume less than 20 mL; SLC25A44, solute carrier family 25 member 44; UCP1, uncoupling protein 1. * $p < 0.05$ and ** $p < 0.01$. Related to [Tables S1](#) and [S2](#).

alanine serine cysteine transporter^{235,36}) that are not exclusive to leucine. This suggests that ^{18}F -fluciclovine kinetics may potentially represent the kinetics of other amino acids as well. In contrast to leucine, ^{18}F -fluciclovine is not metabolized intracellularly. Additionally, ^{18}F -fluciclovine was originally developed as a cancer diagnostic imaging agent and its metabolism has been mostly, but not exclusively, investigated in the context of cancer using preclinical models.³⁷ The analysis of ^{18}F -fluciclovine kinetics was conducted in older men with prostate cancer receiving androgen deprivation therapy, and we cannot determine if the same results apply to healthy and/or younger men and women. BAT activity is known to decline with increasing age,^{38–40} and we expect that younger adults may have a higher net uptake for ^{18}F -fluciclovine. The effect of androgen deprivation treatment on BAT metabolism is currently unclear as it has not been investigated in people and results from preclinical models have been inconsistent.^{41,42} Considering that the results of the kinetics analysis are consistent with results of the gene expression analysis in samples collected from healthy adults, the directionality of the relationship reported in the kinetics analysis is bound to apply to healthy adults, but the magnitude may differ. The quantitative assessment performed in ^{18}F -fluciclovine imaging data may not accurately represent the maximal contribution of BAT in plasma BCAA clearance rate as the tracer injection and PET-CT imaging were performed during room temperature conditions ($\sim 20\text{--}21^\circ\text{C}$), regardless of the season, with full clothing, while cold stimulation has been reported to increase ^{18}F -fluciclovine in rodent BAT.¹⁰ Future studies, including cold stimulation, are needed to assess the role of BAT in BCAA metabolism in young lean individuals who are more likely to have high amount of active BAT.

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

- KEY RESOURCES TABLE
- RESOURCES AVAILABILITY
 - Lead Contact
 - Materials availability
 - Data and code availability

- **EXPERIMENTAL MODEL AND STUDY PARTICIPANT DETAILS**
 - Participants
 - Study approvals
- **METHOD DETAILS**
 - Total-body dynamic PET-CT imaging
 - ¹⁸F-fluciclovine-PET-CT imaging analysis
 - Adipose tissue biopsies from healthy participants and gene expression analysis
- **QUANTIFICATION AND STATISTICAL ANALYSIS**
 - Tracer kinetic modeling
 - Statistical analysis
 - Sample size estimation and statistical power
- **ADDITIONAL RESOURCES**

SUPPLEMENTAL INFORMATION

Supplemental information can be found online at <https://doi.org/10.1016/j.isci.2024.110559>.

ACKNOWLEDGMENTS

The authors thank the participants of the study for their time and effort; Drs. Simon Cherry, Ben Spencer, and the rest of the personnel of the EXPLORER Molecular Imaging Center at the University of California Davis; Dr. Palam Annamalai at the University of Texas Medical Branch and Drs. Raja Ramaswamy and Joseph Daniel Giardina at the Washington University School of Medicine at St. Louis for performing the supraclavicular adipose tissue biopsies; the Center for Human Nutrition and the Clinical Translational Research Unit at Washington University School of Medicine at St. Louis and the Clinical Research Unit at the University of Texas Medical Branch for their assistance in conducting the research studies; and the study subjects for their participation. This study was supported by grants from the National Institutes of Health (P30 DK05634, P30 DK020579, UL1 TR002345, R01 AR076088, R61 AT012187, and R01 CA206187), National Institute of Food and Agriculture (Hatch project number CA-D-NTR-2618-H), American Heart Association (17POST33060003), American Diabetes Association (1-14-TS-35), the Wellcome Trust (208363/A/17/Z), and the National Institute for Health Research (NIHR) Cambridge Biomedical Research Centre (NIHR203312*). V.P. and A.V.-P. were financially supported by a Wellcome strategic award (100574/Z/12/Z) and MRC MDU (MC_UU_12012/2). The views expressed are those of the authors and not necessarily those of the NIHR, the Department of Health and Social Care or other funders.

AUTHOR CONTRIBUTIONS

Y.G.A. and A.R. performed the analysis of the imaging data. S.L. and G.W. performed the kinetic analysis of the imaging data. M.C. performed the studies in healthy humans. M.C. and V.P. analyzed and prepared the biopsy samples for gene expression analysis. M.C. performed the statistical analyses of the data. M.C. and Y.G.A. wrote the first draft of the manuscript. M.C., Y.G.A., A.J.C., R.D.B., A.V.-P., L.S.S., and S.K. designed and supervised the studies and obtained funding for the work. M.C. is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis. All authors critically reviewed and edited the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

Received: September 29, 2023

Revised: June 24, 2024

Accepted: July 17, 2024

Published: July 20, 2024

REFERENCES

1. Neinast, M., Murashige, D., and Arany, Z. (2019). Branched Chain Amino Acids. *Annu. Rev. Physiol.* *81*, 139–164. <https://doi.org/10.1146/annurev-physiol-020518-114455>.
2. Vanweert, F., de Ligt, M., Hoeks, J., Hesselink, M.K.C., Schrauwen, P., and Phielix, E. (2021). Elevated Plasma Branched-Chain Amino Acid Levels Correlate With Type 2 Diabetes-Related Metabolic Disturbances. *J. Clin. Endocrinol. Metab.* *106*, e1827–e1836. <https://doi.org/10.1210/clinem/dgaa751>.
3. Würtz, P., Soininen, P., Kangas, A.J., Rönnemaa, T., Lehtimäki, T., Kähönen, M., Viikari, J.S., Raitakari, O.T., and Ala-Korpela, M. (2013). Branched-chain and aromatic amino acids are predictors of insulin resistance in young adults. *Diabetes Care* *36*, 648–655. <https://doi.org/10.2337/dc12-0895>.
4. Newgard, C.B., An, J., Bain, J.R., Muehlbauer, M.J., Stevens, R.D., Lien, L.F., Haqq, A.M., Shah, S.H., Arlotto, M., Slentz, C.A., et al. (2009). A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and contributes to insulin resistance. *Cell Metabol.* *9*, 311–326. <https://doi.org/10.1016/j.cmet.2009.02.002>.
5. Vanweert, F., Neinast, M., Tapia, E.E., van de Weijer, T., Hoeks, J., Schrauwen-Hinderling, V.B., Blair, M.C., Bornstein, M.R., Hesselink, M.K.C., Schrauwen, P., et al. (2022). A randomized placebo-controlled clinical trial for pharmacological activation of BCAA catabolism in patients with type 2 diabetes. *Nat. Commun.* *13*, 3508. <https://doi.org/10.1038/s41467-022-31249-9>.

6. Guo, F., Chen, R., Kong, L., Wei, P., Liu, Z., Wang, X., Hao, H., Lu, Y., and Hu, W. (2022). Effects of serum branched-chain amino acids on nonalcoholic fatty liver disease and subsequent cardiovascular disease. *Hepatol. Int.* 16, 1424–1434. <https://doi.org/10.1007/s12072-022-10387-8>.
7. Zhao, E., Giamberardino, S.N., Pagidipati, N.J., Voora, D., Ginsburg, G.S., Hoffmann, U., Karády, J., Ferencik, M., Douglas, P.S., Foldyna, B., and Shah, S.H. (2023). Branched-Chain Amino Acids in Computed Tomography-Defined Adipose Depots and Coronary Artery Disease: A PROMISE Trial Biomarker Substudy. *J. Am. Heart Assoc.* 12, e028410. <https://doi.org/10.1161/jaha.122.028410>.
8. Chondronikola, M., and Sidossis, L.S. (2019). Brown and beige fat: From molecules to physiology. *Biochim. Biophys. Acta Mol. Cell Biol. Lipids* 1864, 91–103. <https://doi.org/10.1016/j.bbalip.2018.05.014>.
9. Cannon, B., and Nedergaard, J. (2004). Brown adipose tissue: function and physiological significance. *Physiol. Rev.* 84, 277–359. <https://doi.org/10.1152/physrev.00015.2003>.
10. Yoneshiro, T., Wang, Q., Tajima, K., Matsushita, M., Maki, H., Igarashi, K., Dai, Z., White, P.J., McGarrah, R.W., Ilkayeva, O.R., et al. (2019). BCAA catabolism in brown fat controls energy homeostasis through SLC25A44. *Nature* 572, 614–619. <https://doi.org/10.1038/s41586-019-1503-x>.
11. Neinast, M.D., Jang, C., Hui, S., Murashige, D.S., Chu, Q., Morscher, R.J., Li, X., Zhan, L., White, E., Anthony, T.G., et al. (2019). Quantitative Analysis of the Whole-Body Metabolic Fate of Branched-Chain Amino Acids. *Cell Metabol.* 29, 417–429.e4. <https://doi.org/10.1016/j.cmet.2018.10.013>.
12. Yoneshiro, T., Kataoka, N., Walejko, J.M., Ikeda, K., Brown, Z., Yoneshiro, M., Crown, S.B., Osawa, T., Sakai, J., McGarrah, R.W., et al. (2021). Metabolic flexibility via mitochondrial BCAA carrier SLC25A44 is required for optimal fever. *Elife* 10, e66865. <https://doi.org/10.7554/eLife.66865>.
13. López-Soriano, F.J., Fernández-López, J.A., Mampel, T., Villarroya, F., Iglesias, R., and Alemany, M. (1988). Amino acid and glucose uptake by rat brown adipose tissue. Effect of cold-exposure and acclimation. *Biochem. J.* 252, 843–849. <https://doi.org/10.1042/bj2520843>.
14. Kuroiwa, M., Hamaoka-Fuse, S., Sugimoto, M., Kurosawa, Y., Aita, Y., Tomita, A., Anjo, M., Tanaka, R., Endo, T., Kime, R., and Hamaoka, T. (2021). Correlation of Plasma Amino Acid and Anthropometric Profiles with Brown Adipose Tissue Density in Humans. *J. Clin. Med.* 10, 2339. <https://doi.org/10.3390/jcm10112339>.
15. Leitner, B.P., Huang, S., Brychta, R.J., Duckworth, C.J., Baskin, A.S., McGehee, S., Tal, I., Dieckmann, W., Gupta, G., Kolodny, G.M., et al. (2017). Mapping of human brown adipose tissue in lean and obese young men. *Proc. Natl. Acad. Sci. USA* 114, 8649–8654. <https://doi.org/10.1073/pnas.1705287114>.
16. PubChem [Internet]. Bethesda (MD): National Library of Medicine (US), N.C.f.B.I.-P.C.S.f.C., Fluciclovine (18F); https://pubchem.ncbi.nlm.nih.gov/compound/Fluciclovine_18F.
17. Nanni, C., Zanoni, L., Bach-Gansmo, T., Minn, H., Willoch, F., Bogsrud, T.V., Edward, E.P., Savir-Baruch, B., Teoh, E., Ingram, F., et al. (2020). [18F]Fluciclovine PET/CT: joint EANM and SNMMI procedure guideline for prostate cancer imaging—version 1.0. *Eur. J. Nucl. Med. Mol. Imag.* 47, 579–591. <https://doi.org/10.1007/s00259-019-04614-y>.
18. Hepler, C., and Gupta, R.K. (2017). The expanding problem of adipose depot remodeling and postnatal adipocyte progenitor recruitment. *Mol. Cell. Endocrinol.* 445, 95–108. <https://doi.org/10.1016/j.mce.2016.10.011>.
19. U Din, M., Raiko, J., Saari, T., Saunavaara, V., Kudomi, N., Solin, O., Parkkola, R., Nuutila, P., and Virtanen, K.A. (2017). Human brown fat radiodensity indicates underlying tissue composition and systemic metabolic health. *J. Clin. Endocrinol. Metab.* 102, 2258–2267. <https://doi.org/10.1210/clinem.2016.2698>.
20. Suryawan, A., Hawes, J.W., Harris, R.A., Shimomura, Y., Jenkins, A.E., and Hutson, S.M. (1998). A molecular model of human branched-chain amino acid metabolism. *Am. J. Clin. Nutr.* 68, 72–81.
21. Chondronikola, M., Volpi, E., Børsheim, E., Porter, C., Saraf, M.K., Annamalai, P., Yfanti, C., Chao, T., Wong, D., Shinoda, K., et al. (2016). Brown adipose tissue activation is linked to distinct systemic effects on lipid metabolism in humans. *Cell Metabol.* 23, 1200–1206. <https://doi.org/10.1016/j.cmet.2016.04.029>.
22. Chondronikola, M., Volpi, E., Børsheim, E., Porter, C., Annamalai, P., Enerbäck, S., Lidell, M.E., Saraf, M.K., Labbe, S.M., Hurren, N.M., et al. (2014). Brown adipose tissue improves whole-body glucose homeostasis and insulin sensitivity in humans. *Diabetes* 63, 4089–4099. <https://doi.org/10.2337/db14-0746>.
23. Yamaguchi, S., Franczyk, M.P., Chondronikola, M., Qi, N., Gunawardana, S.C., Stromsdorfer, K.L., Porter, L.C., Wozniak, D.F., Sasaki, Y., Rensing, N., et al. (2019). Adipose tissue NAD(+) biosynthesis is required for regulating adaptive thermogenesis and whole-body energy homeostasis in mice. *Proc. Natl. Acad. Sci. USA* 116, 23822–23828. <https://doi.org/10.1073/pnas.1909917116>.
24. Ma, Q.-X., Zhu, W.-Y., Lu, X.-C., Jiang, D., Xu, F., Li, J.-T., Zhang, L., Wu, Y.-L., Chen, Z.-J., Yin, M., et al. (2022). BCAA–BCKA axis regulates WAT browning through acetylation of PRDM16. *Nat. Metab.* 4, 106–122. <https://doi.org/10.1038/s42255-021-00520-6>.
25. Chen, K.Y., Cypess, A.M., Laughlin, M.R., Haft, C.R., Hu, H.H., Bredella, M.A., Enerbäck, S., Kinahan, P.E., Lichtenbelt, W.v.M., Lin, F.I., et al. (2016). Brown Adipose Reporting Criteria in Imaging Studies (BARCIST 1.0): Recommendations for standardized FDG-PET/CT experiments in humans. *Cell Metabol.* 24, 210–222. <https://doi.org/10.1016/j.cmet.2016.07.014>.
26. Carpentier, A.C. (2020). Branched-chain Amino Acid Catabolism by Brown Adipose Tissue. *Endocrinology* 161, bqaa060. <https://doi.org/10.1210/endo/bqaa060>.
27. Virtanen, K.A., Lönnroth, P., Parkkola, R., Peltoniemi, P., Asola, M., Viljanen, T., Tolvanen, T., Knuuti, J., Rönnemaa, T., Huupponen, R., and Nuutila, P. (2002). Glucose Uptake and Perfusion in Subcutaneous and Visceral Adipose Tissue during Insulin Stimulation in Nonobese and Obese Humans. *J. Clin. Endocrinol. Metab.* 87, 3902–3910. <https://doi.org/10.1210/jcem.87.8.8761>.
28. Koh, H.C.E., van Vliet, S., Pietka, T.A., Meyer, G.A., Razani, B., Laforest, R., Gropler, R.J., and Mittendorfer, B. (2021). Subcutaneous Adipose Tissue Metabolic Function and Insulin Sensitivity in People With Obesity. *Diabetes* 70, 2225–2236. <https://doi.org/10.2337/db21-0160>.
29. Richard, G., Blondin, D.P., Syed, S.A., Rossi, L., Fontes, M.E., Fortin, M., Phoenix, S., Frisch, F., Dubreuil, S., Guérin, B., et al. (2022). High-fructose feeding suppresses cold-stimulated brown adipose tissue glucose uptake independently of changes in thermogenesis and the gut microbiome. *Cell Rep. Med.* 3, 100742. <https://doi.org/10.1016/j.xcrm.2022.100742>.
30. Yu, D., Richardson, N.E., Green, C.L., Spicer, A.B., Murphy, M.E., Flores, V., Jang, C., Kasza, I., Nikodemova, M., Wakai, M.H., et al. (2021). The adverse metabolic effects of branched-chain amino acids are mediated by isoleucine and valine. *Cell Metabol.* 33, 905–922.e6. <https://doi.org/10.1016/j.cmet.2021.03.025>.
31. Jang, C., Oh, S.F., Wada, S., Rowe, G.C., Liu, L., Chan, M.C., Rhee, J., Hoshino, A., Kim, B., Ibrahim, A., et al. (2016). A branched-chain amino acid metabolite drives vascular fatty acid transport and causes insulin resistance. *Nat. Med.* 22, 421–426. <https://doi.org/10.1038/nm.4057>.
32. Deelen, J., Kettunen, J., Fischer, K., van der Spek, A., Trompet, S., Kastenmüller, G., Boyd, A., Zierer, J., van den Akker, E.B., Ala-Korpela, M., et al. (2019). A metabolic profile of all-cause mortality risk identified in an observational study of 44,168 individuals. *Nat. Commun.* 10, 3346.
33. Cinti, S. (2005). The adipose organ. *Prostaglandins Leukot. Essent. Fatty Acids* 73, 9–15. <https://doi.org/10.1016/j.plefa.2005.04.010>.
34. Cypess, A.M., White, A.P., Vernochet, C., Schulz, T.J., Xue, R., Sass, C.A., Huang, T.L., Roberts-Toler, C., Weiner, L.S., Sze, C., et al. (2013). Anatomical localization, gene expression profiling and functional characterization of adult human neck brown fat. *Nat. Med.* 19, 635–639. <https://doi.org/10.1038/nm.3112>.
35. Okudaira, H., Nakanishi, T., Oka, S., Kobayashi, M., Tamagami, H., Schuster, D.M., Goodman, M.M., Shirakami, Y., Tamai, I., and Kawai, K. (2013). Kinetic analyses of trans-1-amino-3-[18F]fluorocyclobutanecarboxylic acid transport in Xenopus laevis oocytes expressing human ASCT2 and SNAT2. *Nucl. Med. Biol.* 40, 670–675. <https://doi.org/10.1016/j.nucmedbio.2013.03.009>.
36. Martarello, L., McConathy, J., Camp, V.M., Malveaux, E.J., Simpson, N.E., Simpson, C.P., Olson, J.J., Bowers, G.D., and Goodman, M.M. (2002). Synthesis of syn- and anti-1-amino-3-[18F]fluoromethyl-cyclobutane-1-carboxylic acid (FMACBC), potential PET ligands for tumor detection. *J. Med. Chem.* 45, 2250–2259. <https://doi.org/10.1021/jm010242p>.
37. Kim, S.J., and Lee, S.W. (2019). The role of 18F-fluciclovine PET in the management of prostate cancer: a systematic review and meta-analysis. *Clin. Radiol.* 74, 886–892. <https://doi.org/10.1016/j.crad.2019.06.022>.
38. Graja, A., and Schulz, T.J. (2015). Mechanisms of aging-related impairment of brown adipocyte development and function. *Gerontology* 61, 211–217. <https://doi.org/10.1159/000366557>.
39. Yoneshiro, T., Aita, S., Matsushita, M., Okamatsu-Ogura, Y., Kameya, T., Kawai, Y., Miyagawa, M., Tsujisaki, M., and Saito, M. (2011). Age-related decrease in cold-activated brown adipose tissue and accumulation of body fat in healthy humans.

- Obesity 19, 1755–1760. <https://doi.org/10.1038/oby.2011.125>.
40. Becher, T., Palanisamy, S., Kramer, D.J., Eljalby, M., Marx, S.J., Wibmer, A.G., Butler, S.D., Jiang, C.S., Vaughan, R., Schöder, H., et al. (2021). Brown adipose tissue is associated with cardiometabolic health. *Nat. Med.* 27, 58–65. <https://doi.org/10.1038/s41591-020-1126-7>.
 41. Lantero Rodriguez, M., Schilperoort, M., Johansson, I., Svedlund Eriksson, E., Palsdottir, V., Kroon, J., Henricsson, M., Kooijman, S., Ericson, M., Borén, J., et al. (2021). Testosterone reduces metabolic brown fat activity in male mice. *J. Endocrinol.* 251, 83–96.
 42. Fan, W., Yanase, T., Nomura, M., Okabe, T., Goto, K., Sato, T., Kawano, H., Kato, S., and Nawata, H. (2005). Androgen Receptor Null Male Mice Develop Late-Onset Obesity Caused by Decreased Energy Expenditure and Lipolytic Activity but Show Normal Insulin Sensitivity With High Adiponectin Secretion. *Diabetes* 54, 1000–1008. <https://doi.org/10.2337/diabetes.54.4.1000>.
 43. Cookson, M.S., Aus, G., Burnett, A.L., Canby-Hagino, E.D., D'Amico, A.V., Dmochowski, R.R., Eton, D.T., Forman, J.D., Goldenberg, S.L., Hernandez, J., et al. (2007). Variation in the definition of biochemical recurrence in patients treated for localized prostate cancer: the American Urological Association Prostate Guidelines for Localized Prostate Cancer Update Panel report and recommendations for a standard in the reporting of surgical outcomes. *J. Urol.* 177, 540–545. <https://doi.org/10.1016/j.juro.2006.10.097>.
 44. Spencer, B.A., Berg, E., Schmall, J.P., Omidvari, N., Leung, E.K., Abdelhafez, Y.G., Tang, S., Deng, Z., Dong, Y., Lv, Y., et al. (2021). Performance Evaluation of the uEXPLORER Total-Body PET/CT Scanner Based on NEMA NU 2-2018 with Additional Tests to Characterize PET Scanners with a Long Axial Field of View. *J. Nucl. Med.* 62, 861–870. <https://doi.org/10.2967/jnumed.120.250597>.
 45. Nioche, C., Orhac, F., Boughdad, S., Reuzé, S., Goya-Outi, J., Robert, C., Pellot-Barakat, C., Soussan, M., Frouin, F., and Buvat, I. (2018). LIFE: A Freeware for Radiomic Feature Calculation in Multimodality Imaging to Accelerate Advances in the Characterization of Tumor Heterogeneity. *Cancer Res.* 78, 4786–4789. <https://doi.org/10.1158/0008-5472.Can-18-0125>.
 46. Chondronikola, M., Annamalai, P., Chao, T., Porter, C., Saraf, M.K., Cesani, F., and Sidossis, L.S. (2015). A percutaneous needle biopsy technique for sampling the supraclavicular brown adipose tissue depot of humans. *Int. J. Obes.* 39, 1561–1564. <https://doi.org/10.1038/ijo.2015.76>.
 47. Chondronikola, M., Yoshino, J., Ramaswamy, R., Giardina, J.D., Laforest, R., Wahl, R.L., Patterson, B.W., Mittendorfer, B., and Klein, S. (2024). Very-low-density lipoprotein triglyceride and free fatty acid plasma kinetics in women with high or low brown adipose tissue volume and overweight/obesity. *Cell Rep. Med.* 5, 101370. <https://doi.org/10.1016/j.xcrm.2023.101370>.
 48. Morris, E.D., Endres, C.J., Schmidt, K.C., Christian, B.T., Muzic, R.F., and Fisher, R.E. (2004). Kinetic modeling in positron emission tomography. In *Emission tomography, 46Emission tomography*, pp. 499–540.
 49. Sörensen, J., Owenius, R., Lax, M., and Johansson, S. (2013). Regional distribution and kinetics of [F] fluciclovine (anti-[F] FACBC), a tracer of amino acid transport, in subjects with primary prostate cancer. *Eur. J. Nucl. Med. Mol. Imag.* 40, 394–402.
 50. Wang, G., Nardo, L., Parikh, M., Abdelhafez, Y.G., Li, E., Spencer, B.A., Qi, J., Jones, T., Cherry, S.R., and Badawi, R.D. (2022). Total-body PET multiparametric imaging of cancer using a voxelwise strategy of compartmental modeling. *J. Nucl. Med.* 63, 1274–1281.
 51. Faul, F., Erdfelder, E., Lang, A.G., and Buchner, A. (2007). G*Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behav. Res. Methods* 39, 175–191. <https://doi.org/10.3758/bf03193146>.

STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Reagent		
¹⁸ F-fluciclovine	PETNET Solutions, Sacramento, CA, USA	NA
Software and algorithms		
SPSS version 29	IBM Corporation	NA
Graphpad Prism version 10	Graph Pad Software	NA
Other		
total-body PET-CT scanner (uEXPLORER)	United Imaging Healthcare	NA

RESOURCES AVAILABILITY

Lead Contact

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Maria Chondronikola (mc2425@medschl.cam.ac.uk).

Materials availability

This study did not generate new unique reagents.

Data and code availability

- All data reported in this paper will be shared by the [lead contact](#) upon reasonable request.
- This paper does not report additional code.
- Any additional information required to reanalyze the data reported in this work paper is available from the [lead contact](#) upon reasonable request.

EXPERIMENTAL MODEL AND STUDY PARTICIPANT DETAILS

Participants

This is a retrospective review of institutional review board-approved registry-type single-institution study. All patients provided their informed consent. Eligible patients ($n = 36$) were adult males with histopathologically proven prostate adenocarcinoma and documented biochemical recurrence after primary treatment with radical prostatectomy (Table 1). The definition of the American Urological Association was used to establish evidence of biochemical recurrence (i.e., prostate-specific antigen concentration of ≥ 0.2 ng/mL, with a second confirmatory level detected at 0.2 ng/mL or more).⁴³ All patients underwent ¹⁸F-fluciclovine total-body PET-CT scanning as part of their standard-of-care evaluation for biochemical recurrence of prostate cancer (BCRPC). Considering the participants were older adults and BAT volume/activity declines with increasing age,^{38–40} the findings of this investigation may underestimate the contribution of BAT in BCAA uptake. The potential effect of sex/gender, ancestry/race/ethnicity and clinical condition on the outcomes of this proof-of-concept study is currently unclear.

Study approvals

The study was approved by the Institutional Review Boards of the University of California Davis, the University of Texas Medical Branch in Galveston, the Washington University School of Medicine in St. Louis, MO. Written informed consent was obtained from all participants before their participation in this study.

METHOD DETAILS

Total-body dynamic PET-CT imaging

All participants underwent list-mode dynamic single-bed total-body PET-CT acquisition for 25 min in the supine positions with hands above the head. The scan started immediately before the injection of 312 ± 10 (range 287–347) MBq of ¹⁸F-fluciclovine on a total-body PET-CT scanner (uEXPLORER, United Imaging Healthcare). The scanner has an axial field of view of 194 cm, PET spatial resolution of ~ 3.0 mm in full-width at half-maximum near the center of axial field-of-view, and an 80-detector row CT with a minimum slice thickness of 0.5 mm.⁴⁴ Prior to the acquisition of dynamic PET data, a low-dose CT (tube current: ~ 50 mA, tube voltage: 140 kVp) was acquired for attenuation correction and anatomical localization. The tube current was automatically modulated by the scanner manufacturer's algorithm. The PET data were

reconstructed into multiple total-body temporal series, including 32 frames ($12 \times 5s$, $3 \times 10s$, $3 \times 30s$, $6 \times 60s$, and $8 \times 120s$), which were used for kinetic analysis, and a single 10-min frame between the 4th min post-injection till 14th min, which was used for assessment of mean standardized uptake value (SUV_{mean} , g/mL) and visualization. This time window was selected to avoid early phase with high vascular tracer content and to avoid relatively later frames where skeletal muscles tend to demonstrate increasing tracer uptake. All reconstructions were performed using the manufacturer-provided software, employing 3D ordered subset expectation maximization (OSEM) algorithm, with 4 iterations and 20 subsets into a 256×256 matrix, rendering an isotropic voxel size of 2.344 mm. All standard corrections were applied (scatter, randoms, dead-time and normalization) without point-spread function (PSF) modeling. No post-reconstruction smoothing was applied. CT images were reconstructed using the manufacturer-provided iterative method with a slice thickness of 2.344 mm to match those of PET and an in-plane voxel size of $\sim 0.49 \times 0.49$ mm.

¹⁸F-fluciclovine-PET-CT imaging analysis

The reconstructed PET and CT images were transferred to a standard analysis workstation running LifeX software (versions 7.2).⁴⁵ PET and CT registration was confirmed visually and any misregistration was corrected if needed. Three volumes of interest (VOIs) were drawn on the fused PET-CT using the brush tool. First VOI was drawn on the left and right supraclavicular areas immediately at the root of the neck corresponding to the known site of BAT in adult human, second VOI on the subcutaneous adipose tissue in the proximal arm/upper chest and back of the shoulder regions (upper chest adipose tissue), and the third VOI on the subcutaneous abdominal adipose tissue at the level of third lumbar vertebra. To ensure adequate circumferential representation, subcutaneous VOIs were placed to sample left and right parts on consistent anatomical regions across subjects as possible (two for supraclavicular, three for upper chest, and four for subcutaneous abdominal). VOIs were at least 5.0 mL in volume. Within each VOI, two sequential sets of thresholds were applied: first on the CT images to include only adipose tissue densities (-190 to -30 Hounsfield Unit [HU]) then on PET images to exclude any potential spill-over from adjacent skeletal muscles (and other soft tissues with high activity). The study personnel that drew the VOI were blinded to the hypothesis of this investigation.

Adipose tissue biopsies from healthy participants and gene expression analysis

Supraclavicular and abdominal adipose tissue samples were collected from a group of adults with overweight/obesity who participated in clinical trials aiming to examine the role of human BAT in metabolic function (ClinicalTrials.gov NCT02786251 and NCT01791114). The entire cohort included 23 participants with supraclavicular adipose tissue samples and 12 of them also had matched abdominal adipose tissue biopsies. Table S1 summarizes the demographic and anthropometric characteristics of participants that provided adipose tissue biopsies. Each participant performed a 6-h exposure to mild cold followed by a 2-deoxy-2-[¹⁸F]-fluoro-glucose PET-CT scan and a supraclavicular adipose tissue biopsy.^{21,22,46,47} An abdominal subcutaneous adipose tissue biopsy was collected from the same participant immediately after the supraclavicular adipose tissue biopsy or 1–4 weeks apart under the same environmental and fasting conditions. During cold exposure, participants wore shorts, a sports bra (females only), a T-shirt/hospital gown and a liquid-conditioned garments (ThermoWrap, Belmont Medical Technologies, Billerica, MA or Polar Products Inc, Stow, OH). The room and cooling equipment temperature were initially set to 22°C and decreased by 1°C–2°C in intervals of 30–60 min until shivering was induced. Immediately upon shivering, cooling equipment temperature was increased by 1°C–2°C until shivering stopped. BAT volume was quantified using the BARCIST 1.0 recommendations.²⁵ All tissue samples were rinsed with ice-cold saline immediately after collection, cleaned off connective tissue and blood and immediately frozen in liquid nitrogen. Approximately 100 mg of adipose tissue was used for extraction of RNA using RNeasy Mini Kit (74104; Qiagen, Chatsworth, California) including an on-column DNase digestion step. The expression of the various genes involved in BCAA metabolism (*BCAT1/2*, *BCKDHB* and *SLC25A44*) and *UCP1*, relative to the housekeeping control gene ribosomal protein lateral stalk subunit P0 (*RPLP0*), was determined by using an ABI 7500 Real-Time PCR System (Invitrogen, Carlsbad, California) and Fast SYBR Green Master Mix (4385618, Invitrogen). The primers used are shown in Table S2.

QUANTIFICATION AND STATISTICAL ANALYSIS

Tracer kinetic modeling

An irreversible two-tissue (2T) compartmental model,^{48,49} as illustrated in Figure S1, was used to describe the kinetics of ¹⁸F-fluciclovine PET data. C_p is the tracer concentration in plasma. The tracer enters and exits the free-state compartment (C_f) with the rate of K_1 (mL/min/cm³) and k_2 (min⁻¹). k_3 (min⁻¹) is the rate constant of the tracer from C_f to the contained compartment C_c which may represent the intracellular space.⁴⁹ This irreversible model assumes that there is no exit process from the intracellular space (i.e., $k_4 = 0$). The total activity that can be measured by PET is the sum of different compartments:

$$C_T(t) = v_b C_b(t) + (1 - v_b)(C_f(t) + C_c(t))$$

where v_b (mL/mL) is the fractional blood volume. $C_b(t)$ represents the tracer concentration in the whole blood. All the kinetic parameters were estimated by fitting a measured tissue time activity curve (TAC) following the approach we have previously described elsewhere.⁵⁰ An image-derived TAC in the ascending aorta was used as the plasma input function. The macro-parameter of interest is the net uptake rate $K_i = \frac{K_1 k_2}{k_2 + k_3}$ in the unit of mL/min/cm³, which reflects the clearance of ¹⁸F-fluciclovine from blood to tissue.

Statistical analysis

Results are presented as means \pm SD or SEM for normally distributed data and as median and interquartile range for skewed data. Tissue differences between paired observations were assessed by using repeated-measures ANOVA (for normally distributed data) with Tukey correction for multiple comparisons and the Friedman test (for skewed data) with Dunn correction for multiple comparisons. Group differences between independent observations were assessed using one-way ANOVA with Tukey correction for multiple comparisons. Within and between subject differences in gene expression were assessed by using the Wilcoxon signed-rank test and the Mann Whitney test respectively. Spearman's rho was used to evaluate the correlation between the different variables of interest. Statistical analyses were performed by using GraphPad v9 (GraphPad Software, San Diego, CA). All statistical tests assumed a 95% level of confidence as significant.

Sample size estimation and statistical power

Based on our preliminary data, we estimated that at minimum 8 subjects in each group would be needed to detect paired differences between supraclavicular and subcutaneous abdominal adipose tissue SUV_{mean} for ^{18}F -fluciclovine equal to 0.2 g/mL using a 2-sided test with a power of 0.95 and an α value of 0.05. These computations were performed using G*Power 3.1.9.6.⁵¹

ADDITIONAL RESOURCES

Clinical trial registration number NCT02786251, clinicaltrials.gov (<https://clinicaltrials.gov/study/NCT02786251>) and NCT01791114, clinicaltrials.gov (<https://clinicaltrials.gov/study/NCT01791114>).