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# **T cell behavior at the maternal-fetal interface**

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#### **Abstract**

Understanding the function of T cells at the maternal-fetal interface remains one of the most difficult problems in reproductive immunology. A great deal of work over the last two decades has led to the view that the T cells that populate the decidua have important roles in both normal and pathological pregnancies, but the exact nature of these roles has remained unclear. Indeed, the old assumption that decidual T cells are uniformly threatening to fetal survival because the placenta is fundamentally an 'allograft' has given way to the idea that different T cells subsets contribute in different ways to pregnancy success or failure. Accordingly, some T cells are thought to protect the placenta from immune rejection and facilitate embryo implantation, while others are thought to contribute to pregnancy pathologies such as preeclampsia and spontaneous abortion. Here, we review the current state of information on the behavior of decidual T cells with a focus on both mouse and human studies, and with an emphasis on the many unresolved areas within this overall emerging framework.

#### **Introduction**

Many different kinds of maternal leukocytes populate the maternal-fetal interface (i.e. the decidua), each with diverse functions in implantation, placental development, parturition and infectious disease control (for review, see (Bulmer *et al.*, 2010, Erlebacher, 2013a, Trundley and Moffett, 2004)). For example, natural killer cells (NK cells) are well known as critical regulators of spiral artery remodeling in both mice and humans. Here we will discuss the role of decidual αβ T cells (henceforth referred to as simply T cells), whose functions remain particularly poorly understood. The difficulty on working on these cells in part comes from the existence of multiple T cell subsets possessing diverse sets of functions. In addition, each T cell possesses a distinct antigen specificity as determined by its uniquely expressed T cell receptor (TCR) generated during T cell development in the thymus as the result of TCR α and TCR β gene rearrangements. Nonetheless, subset differentiation and antigen specificity will ultimately be central to understanding the function and significance of decidual T cells. Due to the complexity of these issues, we begin with a broad introduction into T cell biology, along with a brief historical perspective on the role of T cell subsets in pregnancy. We then move on to the specifics of T cell biology at the maternal-

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fetal interface, with an emphasis on the many unresolved issues concerning these cells. Several of the pathways we cover are diagrammed in Figure 1. An introduction to natural killer T (NKT) cells, CD4<sup>-</sup> CD8<sup>-</sup> T cells, and  $\gamma\delta$  T cells, which are relatively rare in the pregnant uterus, can be found elsewhere (Tilburgs et al., 2010a); other reviews of decidual αβ T cells, with additional perspectives, can also be found elsewhere (Ernerudh *et al.*, 2011, Saito et al., 2010, Teles et al., 2013b, Tilburgs and Strominger, 2013).

#### **T cell activation and subsets – a primer**

Following their development in the thymus, T cells are considered to be naïve until they first encounter cognate peptide antigen. This encounter takes place in the secondary lymphoid organs – i.e. the spleen for blood-borne antigens and the regional lymph nodes for bloodborne antigens and antigens that drain via lymphatic vessels from peripheral tissues – and entails interactions with dendritic cells (DCs) that present the cognate antigen on their cell surface in association with major histocompatibility complex (MHC) molecules. For CD4 T cells, TCR signaling is stimulated upon engagement of peptide antigen:MHC class II complexes; for CD8 T cells, TCR signaling is stimulated upon engagement of peptide antigen:MHC class I complexes. If the DC also supplies sufficient costimulation, which includes critical signals generated from the surface molecules CD80 and CD86, the responding T cell proliferates (i.e. undergoes clonal expansion) and then differentiates into one of several T cells subsets as specified by the set of cytokines encountered during antigen exposure. As discussed below, these subsets are  $T_H1$ ,  $T_H2$ ,  $T_H17$  effector CD4 T cells, regulatory CD4 T ( $T_{\text{Reg}}$ ) cells, and cytotoxic T lymphocytes (CTLs), which are effector CD8 T cells. Once activated, an effector T cell exits the spleen or lymph node and then homes via the blood to the peripheral tissues. Although a relatively open area of research, T cell behavior within peripheral tissues is also thought to be actively regulated through pathways that include local antigen presentation and cytokine production by DCs and macrophages, as well as cross-inhibition of the different  $T_H$  cell subsets via their own production of cytokines.

#### **Effector CD4 T cells**

 $T_H1$ ,  $T_H2$ , and  $T_H17$  cells are the three main effector CD4 T cell subsets currently known (for review, see (Zhu *et al.*, 2010)). They are defined by the set of transcription factors they respectively express in order to maintain their differentiated state, as well as by the set of cytokines they express that mediate their effector functions. In addition, each subset expresses a characteristic set of chemokine receptors that directs recruitment to sites of inflammation by promoting the extravasation of the cells across the endothelium. Consequently, the set of chemokine ligands expressed by an inflamed tissue will determine to a large extent the effector T cell subsets that can gain access to this tissue.

 $T<sub>H</sub>1$  cells primarily function within peripheral tissues to promote the eradication of virusinfected cells and intracellular pathogens. The cells express the transcription factors T-bet and STAT4, and secrete interferon- $\gamma$  (IFN $\gamma$ ) as their signature cytokine. They also express the chemokine receptors CXCR3, which is the receptor for CXCL9 (MIG), CXCL10 (IP-10), CXCL11 (I-TAC), and CCR5, which is a receptor for CCL5 (RANTES). IFNγ

produced by  $T_H1$  cells promotes macrophage activation as well as stromal and endothelial cell expression of the CXCL9, CXCL10, and CXCL11, which means that activated  $T_H1$ cells promote both their own recruitment as well as the recruitment of CTLs, which also express CXCR3 (Nakanishi et al., 2009). T<sub>H</sub>1 cells also produce tumor necrosis factor-α (TNF $\alpha$ ), which serves to promote inflammation in a variety of ways. Importantly,  $T_H1$  cells are the primary CD4 T cells that drive surgical allograft rejection and therefore have long been considered major threats to fetal survival and potential contributors to pregnancy pathologies.

 $T_H2$  cells primarily function in allergic reactions through their regulation of antibody isotype switching, and in the eradication of helminths. The cells express the transcription factors GATA3 and STAT6, secrete the cytokines IL-4, IL-5 and IL-10, and preferentially express the chemokine receptor CCR4, which is required for migration to sites of allergic inflammation such as the airways. Historically, interest in  $T_H2$  cells in pregnancy has come from the fact that these cells provided an alternative, less potentially embryotoxic differentiation state for CD4 T cells in comparison to  $T_H1$  cells, as well as the ability of  $T_H2$ cytokines to repress  $T_H1$  cell differentiation and function *in trans*. Thus, it had been thought that pregnancy would involve a general Th2-skewing of the T cell response in order to minimize the generation of  $T_H1$  cells. A great deal of data, however, has rendered this idea overly simplistic (discussed further in (Saito et al., 2010)).

 $T_H$ 17 cells augment acute inflammatory responses and mediate host immunity against extracellular bacteria and fungi. The cells express the transcription factors RORγt, STAT3 and IRF4, and secrete members of the IL-17 family of pro-inflammatory cytokines, most notably IL-17A. IL-17A acts on epithelial cells to induce neutrophil chemoattractants such as CXCL1 and neutrophil survival factors such as G-CSF. The predominant chemokine receptor of  $T_H$ 17 cells is CCR6, which attracts them to epithelial surfaces expressing their ligand CCL20. As the most recently described  $T_H$  cell subset, their role in reproduction is only now beginning to be elucidated.

### **CTLs**

CTLs are the CD8 T cell counterparts of effector  $T_H1$  cells, and like  $T_H1$  cells play a major role in virus and intracellular pathogen clearance. They express the same set of transcription factors (T-bet and STAT4) and cytokines (IFN $\gamma$  and TNF $\alpha$ ) as T<sub>H</sub>1 cells, and their recruitment to peripheral tissues is similarly governed by localized expression of CXCR3 and CCR5 ligands. In contrast to  $T_H1$  cells, however, CTLs have the capacity to directly kill target cells by virtue of their expression of the cytolytic molecules perforin and granzyme. These molecules are released upon TCR-mediated interactions with target cells expressing cognate peptide:MHC class I complexes. As with  $T_H1$  cells, CTLs play a major role in graft rejection and so too have been considered direct threats to fetal survival. This view has been tempered, however, by the realization that invasive trophoblasts, in both mice an humans, express a limited set of classical MHC class I molecules (King *et al.*, 2000, Madeja *et al.*, 2011), which means that there is less opportunity for CTLs to directly attack the placenta as they might a surgical organ transplant. On the other hand, the cytokines IL-12 and IL-18 produced by DCs and macrophages can induce CD8 T cells to produce IFNγ in a TCR-

independent manner (Freeman *et al.*, 2012). Thus, the mere presence of these cells at the maternal-fetal interface has, in principle, the potential to augment decidual inflammation to the detriment of pregnancy success.

#### **Regulatory T cells**

 $T_{\text{Reg}}$  cells suppress the activity of other immune cell types and are involved in down regulating immune responses after the elimination of invading organisms, in preventing autoimmunity, and in minimizing pathogenic responses to commensals (for a review, see (Campbell and Koch, 2011)). The cells are defined by their expression of the FOXP3 transcription factor, and are identified as such using intracellular staining protocols in conjunction with the cells'  $CD4^+$   $CD25^{\text{hi}}$  surface phenotype. The cells express a wide variety of different chemokines receptors, and so have a rather promiscuous homing capacity.  $T_{\text{Reg}}$  cells are thought to primarily mediate their immunosuppressive effects though secretion of the cytokines IL-10 and TGF-β, as well as by acting as a sink for IL-2, which is a T cell mitogen and the ligand for CD25.

 $T_{Reg}$  cells are further classified in two groups based upon their origin. Natural  $T_{Reg}$  (nT<sub>Reg</sub>) cells are generated in the thymus directly from T cell precursors and have reactivity towards self antigen. As a result, these cells are primarily thought to be involved in preventing the development of autoimmune reactions. In contrast, induced  $T_{\text{Reg}}$  (i $T_{\text{Reg}}$ ) cells are generated in secondary lymphoid organs from naïve CD4 T cells following their simultaneous exposure to innocuous foreign antigen and TGF- $\beta$ . iT<sub>Reg</sub> cells are therefore thought to play a specialized role in mitigating pathogenic responses to commensal flora. Recently,  $nT_{Reg}$ cells and  $i_{\text{Reg}}$  cells, including those within the human decidua, have been distinguished by virtue of their differential expression of the Helios transcription factor, but the use of this marker has been challenged (Akimova *et al.*, 2011, Gottschalk *et al.*, 2012, Verhagen and Wraith, 2010).

Interest in the role of  $T_{\text{Reg}}$  cells in pregnancy, which is currently very intense, was initially stimulated by the dual findings that the cells increased in frequency in blood during both human and mouse pregnancy (Aluvihare *et al.*, 2004, Somerset *et al.*, 2004), and were required to prevent pregnancy failure in mice in allogeneic mating combinations (Aluvihare *et al.*, 2004) (of note, the finding of  $T_{\text{Reg}}$  cell expansion during human pregnancy has since become controversial (Ernerudh et al., 2011)). More recently, the CNS1 enhancer element of the FOXP3 gene, which is specifically required for  $iT_{Reg}$  cell generation, was found to exist only in placental mammals, being absent from even monotremes and marsupials (Samstein *et al.*, 2012). This result suggested a highly specific role for  $i_{\text{Reg}}$  cells in pregnancy, potentially both systemically and at the maternal-fetal interface. We discuss the possible role of  $T_{\text{Reg}}$  cells within the decidua further below (for an additional review, see (Ernerudh *et al.*, 2011)); their potential systemic function during pregnancy has been discussed elsewhere (Erlebacher, 2013b).

#### **Memory and tissue resident memory T cells**

Virtually all effector T cells generated in response to an immunogenic antigen undergo apoptosis once antigen clearance is complete. However, a few of these antigen-experienced

cells survive and differentiate into memory T cells, which are capable of mounting rapid responses after a second contact with antigen. Classically, memory cells have been divided into two subsets based upon their homing patterns (for review, see (Mueller *et al.*, 2013)). Central memory T cells express CD62L (L-selectin) and the chemokine receptor CCR7 and recirculate throughout the blood and secondary lymphoid organs like naïve T cells. In contrast, effector memory T cells are CD62L− CCR7− and recirculate throughout peripheral tissues patrolling for antigen. More recently, a third class of memory T cells has been described, termed resident memory  $T$  cells ( $T_{RM}$  cells) that, as implied by their name, remain stationary within the tissues where they are thought to provide an immediate, yet antigen-specific, defense against a re-infecting pathogen (Mueller  $et al., 2013$ ). These cells have recently been identified in the mouse reproductive tract (Schenkel *et al.*, 2013), but their role in pregnancy has not yet been investigated.

#### **T cell composition in the decidua**

A thorough analysis of the T cell subset composition of the mouse decidua has not been published. Compared to other leukocyte subtypes such as NK cells and monocytes, T cells are quite rare, and comprise only about 3% of total decidual leukocytes on embryonic day (E) 8.5 ((Croy et al., 2012) and Nancy and Erlebacher, unpublished data). The cells are split about 50:50 between CD4 and CD8 T cells (Croy et al., 2012), and approximately 15% of the CD4 T cells are FOXP3<sup>+</sup> T<sub>Reg</sub> cells on E13.5-14.5 (Samstein *et al.*, 2012). The proportion of these  $T_{Reg}$  cells that are  $nT_{Reg}$  versus  $iT_{Reg}$  cells is unknown. In contrast, a great deal more work has been performed on the human decidua, where T cells comprise a much greater proportion of total leukocytes (10-20%; for review see (Erlebacher, 2013a)). About ~30-45% of the cells are CD4 T cells and 45-75% are CD8 T cells (Bulmer *et al.*, 1991, Marlin et al., 2011, Mjosberg et al., 2010, Tilburgs et al., 2009b, Vassiliadou and Bulmer, 1996), with the large majority of both subsets being antigen-experienced (CD45RA− or CD45RO+) (Saito et al., 1994, Slukvin et al., 1996, Tilburgs et al., 2010b). Based upon their chemokine expression profile, putative  $T_H2$  and  $T_H17$  cells comprise only ~5% and 2% of first trimester decidual CD4 T cells, respectively, while  $T_H1$  (CCR4<sup>-</sup> CXCR3+ CCR6−) cells surprisingly comprise ~5-30% of the cells (Mjosberg et al., 2010). About 5% of the CD4 T cells are CD25<sup>hi</sup> FOXP3<sup>+</sup> T<sub>Reg</sub> cells (Mjosberg *et al.*, 2010, Tilburgs et al., 2008), with about 55% of these cells at term in turn being putative Helios<sup>+</sup> nT<sub>Reg</sub> cells, and 45% being putative Helios<sup>-</sup> iT<sub>Reg</sub> cells (Hsu *et al.*, 2012). In humans, T<sub>Reg</sub> cell frequencies have also been noted to be higher in the decidua than in the blood (Dimova et al., 2011, Mjosberg et al., 2010, Nagamatsu et al., 2011, Sasaki et al., 2004, Schumacher et al., 2009, Tilburgs et al., 2008, Tilburgs et al., 2006).

These data have an important caveat, however, since they were generated via flow cytometric analyses of disaggregated tissues. Namely, it is currently unknown how many of the T cells visualized in this matter are present within the tissue parenchyma versus the blood that is flowing through the tissue. In mice, this issue has been highlighted by our recent data discussed below demonstrating that TH1 cells and CTLs are actively prevented from extravasating into the decidua (Nancy *et al.*, 2012). Indeed, using a technique we developed for discriminating intravascular from extravascular leukocytes in the decidua (Tagliani et al., 2011), we have found that the majority of the CD4 and CD8 T cells isolated

from non-perfused decidua on E8.5 are actually intravascular under steady state conditions (Nancy and Erlebacher, unpublished data). It is unclear whether vascular perfusion is able to remedy this problem, as many leukocytes within the mouse decidua are adherent to the endothelium (Kruse et al., 1999). Judging from work in rats, these leukocytes are likely to remain even after vascular perfusion (Welsh and Enders, 1985).

A related interpretative issue concerns the comparison made frequently between the lymphocyte subset composition of peripheral blood of a pregnant woman and the composition of the woman's decidua. This comparison, in which the over-representation of a given subset is sometimes taken to imply the existence of local mechanisms that directly enhance that subset's recruitment or survival have not sufficiently considered the fact that naïve T cells, which lack tissue-homing chemokine receptors (e.g. CXCR3, CCR4, CCR5, CCR6), enter non-lymphoid peripheral tissues relatively poorly. For example, in a recent study on pregnant women at term, naïve (CD45RA+) CD8 T cells were shown to comprise ~50% of all CD8 T cells in the blood, but only ~5% of CD8 T cells in the decidua, with these latter cells including ones that might actually be intravascular (Tilburgs *et al.*, 2010b). Similar results have been obtained for both CD4 and CD8 T cells in studies of the first trimester (Saito et al., 1994, Slukvin et al., 1996). These data imply that that all differentiated T cell subsets will, without any specific recruitment mechanism, appear enriched within the decidua compared to the blood when cell numbers are expressed as proportions of respective total CD4 or CD8 T cells. Despite some evidence pointing to the involvement of specific chemoattractants (Schumacher et al., 2009), this issue is particularly relevant to suggestions that  $T_{\text{Reg}}$  cells are actively recruited to the human maternal-fetal interface (Schumacher et al., 2009, Tilburgs et al., 2008, Tilburgs et al., 2006).

#### **Antigen specificity and locations of antigen presentation**

Two of the most persistent and difficult questions regarding the biology of decidual T cells are whether the cells have specificity for placental antigens, and if so, where they encounter these antigens. The former question is far from trivial given the existence of resident memory cells and the well-known property of effector memory T cells to continuously traffic through peripheral tissues even in the absence of local inflammation or infection.  $T_{\text{Reg}}$  cells also have promiscuous homing properties and are thought to populate all tissues to some extent at steady state (Campbell and Koch, 2011). Thus, it is entirely conceivable that a large fraction of decidual T cells do not have placental specificity but rather populate the decidua either because they just happen to be migrating through this tissue in an antigen non-specific fashion, or because they are the same cells that were present in the endometrium at the time of implantation. These possibilities are also supported by the lack of data showing that maternal T cells are robustly activated to placental antigens, as discussed further below, as well as data indicating that certain T cell subsets are actively prevented from extravasating into the decidua, also discussed below.

The question of where T cells encounter placental antigens, while understood to a much greater extent, still has unresolved aspects. In mice, the uterine draining lymph nodes (LN) are sites of placental antigen presentation, but this is surprisingly not the result of DC migration from the decidua. Rather, decidual DC become trapped within the tissue, and

placental antigens instead are disseminated in cell-free form both via the regional lymphatics as well as through the blood, which gives them access to the spleen and all LN throughout the body (Collins et al., 2009, Erlebacher et al., 2007, Moldenhauer et al., 2009, Rowe et al., 2012). Ensuing antigen presentation by the antigen presenting cells (APCs) that reside within these secondary lymphoid organs is ultimately non-immunogenic, however, as effector T cells are not generated from naïve precursors. Instead, the responding T cells are largely deleted as they simultaneously undergo multiple rounds of cell division. This work has been based upon the use of transgenic mice that express model antigens as surrogate fetal/placental antigens in order to clearly detect antigen-specific T cell responses, but it is assumed that findings also apply to endogenous placental antigens (for review, see (Moldenhauer  $et al., 2010$ ). In the case of at least one model antigen, it has recently been shown that the systemic CD4 T cell response can also involve conversion into  $iT_{\text{Re}g}$  cells, but the number of such cells generated in this matter is at maximum on the order of 1000 cells per mouse (Rowe et al., 2012).

The upshot of this work is that it might be incredibly difficult to detect maternal T cells specifically responding to placental antigens in humans, especially if blood alone is analyzed. In one recent attempt, CD8 T cells specific for male HY minor histocompatibility antigen were indeed detected in the blood of pregnant women bearing male concepti, but the cells were only detectable in half of the relevant pregnancies and their percentage (of total CD8 T cells) was on average only 0.043% after 10 days of in vitro peptide stimulation (Lissauer et al., 2012). Interestingly, the cells that grew out had an effector memory phenotype. In another study, a mother/child mismatch in human leukocyte antigen- (HLA-) C, which is one of the classical MHC class I molecules in humans, was found to enrich the frequency of CD25dim (i.e. activated or memory) decidual CD4 T cells by about 10%, an observation that was attributed to the presentation of paternal HLA-C-derived peptides by maternal APCs (Tilburgs *et al.*, 2009a). HLA-C-specific CD4 T cells were not directly identified, however, and no change was observed in the frequency of decidual CD28<sup>−</sup> (effector or effector/memory) CD8 T cells.

Indeed, recent work in mice has complicated even the idea that the expanded numbers of T<sub>Reg</sub> cells observed systemically during both mouse and human gestation represent the aggregate response of a large number of CD4 T cell clones synchronously converting to  $i<sub>Reg</sub>$  cells in response to shed placental antigen. Specifically, it was found that treatment of non-pregnant mice with the pregnancy hormone progesterone, given at doses to achieve serum levels similar to what occur at midgestation, induced the same systemic ~2-4-fold expansion of  $T_{\text{Reg}}$  cells typically seen during mouse pregnancy (Mao *et al.*, 2010). Conversely, this study also revealed that  $T_{\text{Reg}}$  cells expand in a progesterone receptordependent fashion in mice rendered "pseudopregnant," which is a temporary hormonal state induced by the act of copulation alone (without actual embryo implantation) and associated with elevated progesterone production by the ovary. Thus, expanded systemic  $T_{\text{Reg}}$  cell numbers during both mouse and human pregnancy might largely reflect an antigen nonspecific effect of progesterone rather than specific induction by placental antigens. Of note, however, another study has shown no effect of progesterone on  $T_{\text{Reg}}$  cell numbers in mice (Zhao et al., 2007).

Importantly, however, it remains possible that decidual APCs present placental antigen within the decidua itself. Indeed, CD83<sup>+</sup> DCs in the first trimester human decidua form clusters with decidual T cells (Kammerer *et al.*, 2000), while CD14<sup>+</sup> DC-SIGN<sup>+</sup> cells, which are thought to be a specialized macrophage subset with antigen presenting capacity (Erlebacher, 2013a, Nagamatsu and Schust, 2010), have been found in close association with decidual  $T_{\text{Reg}}$  cells (Hsu *et al.*, 2012). If these spatial proximities reflect ongoing antigen presentation, however, it remains unclear the extent to which the responding T cells are naïve given the exclusion of naïve T cells from peripheral tissues and the low proportion of naïve T cells in the human decidua (Saito et al., 1994, Slukvin et al., 1996, Tilburgs et al., 2010b). Indeed, the current set of data in mice strongly argues against the intra-decidual presentation of placental antigens to at least naïve CD8 T cells, since such cells can be seen responding to a surrogate placental antigen in the spleen and uterine draining LN without any concurrent response within the decidua itself (Erlebacher *et al.*, 2007). Thus, it is important to consider the alternative possibility that the intra-decidual presentation of placental Ag serves to reinforce the activation of placenta-specific T cells that were first exposed to antigen elsewhere. This possibility is consistent with the demonstrated systemic generation of iT<sub>Reg</sub> cells specific for shed placental antigens (Rowe *et al.*, 2012) followed by the homing of these cells to the decidua. Given recent data that the maternal-fetal interface is normally colonized at low levels with a variety of microbes (Stout et al., 2013), it is also possible that decidual APCs present microbe-derived peptides, perhaps as part of a mechanism to combat decidual infection.

#### **Recruitment**

Another dimension to the issue of the composition and function of decidual T cells comes from recent work from our laboratory showing that activated  $T_H1$  cells and CTLs are actively excluded from the mouse decidua (Nancy et al., 2012). This exclusion prevented antigen-specific fetal loss but itself was not antigen-specific. Rather, it was linked to an epigenetic program activated in differentiating decidual stromal cells (DSCs) that silenced expression of the Cxcl9, Cxcl10, Cxcl11 and Ccl5 genes that encode the key  $T_H1$  cell- and CTL-attracting chemokines CXCL9, CXCL10, CXLC11, and CCL5 discussed above.

In addition to suggesting a major reason why the fetus is not rejected by the maternal immune system, these results demonstrated that epigenetic pathways active in the stromal compartment of the mouse decidua play a major role in controlling T cell subset population dynamics at the maternal-fetal interface. Two pressing questions, therefore, are whether similar pathways are active in humans, and, if so, the extent to which they influence T cell composition of the human decidua in various physiological and pathophysiological settings. In particular, the existence of chemokine gene silencing pathways raises the possibility that the apparent relative proportions of T cell subsets in the decidua in normal pregnancy might not be the consequence of the active, specific recruitment of certain T cell subsets over others, or the selective proliferation, survival or retention of these subsets, but rather because T cell subsets strictly dependent upon CXCL9, CXCL10, CXCL11 and CCL5 for peripheral tissue recruitment are excluded. Indeed, demonstration that CXCL9, CXCL10, CXCL11 and CCL5 are silenced in human decidual stromal cells would suggest that the  $T_H1$  cells that populate the human decidua at relatively high numbers are present from the time of

#### **Regulation**

The decidua-specific factors and pathways that regulate decidual T cell behavior are very poorly understood, and several promising ideas have not panned out in a robust fashion due to the absence of pregnancy phenotypes in gene-deficient mouse models. These include the proposals that the death ligand FasL, the negative costimulatory molecule PD-L1, and the tryptophan catabolizing enzyme idoleamine 2,3-dioxygenase, which are all expressed at the maternal-fetal interface would respectively kill, render inactive, or starve infiltrating effector T cells (Baban et al., 2004, Chaouat and Clark, 2001, Taglauer et al., 2009). In contrast, there is mounting evidence that the multifunctional carbohydrate-binding protein Galectin-1 may locally regulate decidual T cell behavior to promote a uterine environment supportive of pregnancy (for review see (Barrientos et al., 2013)). Consistent with this possibility, female mice deficient in Galectin-1 show higher rates of fetal loss in allogeneic but not syngeneic pregnancies (Blois et al., 2007). Galectin-1 has also been shown to be colocalized to clusters of T cells within the human decidua that are undergoing apoptosis (Kopcow et al., 2008), suggesting a possible role in regulating intradecidual T cell survival. As discussed further below, decidual NK cells have also recently been shown to repress T<sub>H</sub>17 cells through their production of IFN<sub>Y</sub> (Fu et al., 2013). Lastly, as also discussed further below, decidual  $T_{\text{Reg}}$  cells are thought to attenuate the activity of decidual effector  $T$ cells.

#### **Functions and Pathological associations**

#### **Implantation**

At present, it is technically difficult to perform intrauterine T cell ablations in mice, and ethical concerns preclude experimentation on pregnant women. Consequently, our current understanding of the function of decidual T cell subsets during pregnancy has largely been inferential, as it derives from knowledge about the function of each subset in non-uterine tissues and discoveries of associations between human pregnancy pathologies and changes in a subset's prevalence within the decidua. Examples of this approach will be discussed further below. One emerging exception to the current state of ambiguity, however, comes from work suggesting that  $T_{Reg}$  cells play an important role in preparing the endometrium for implantation (for review, see (Robertson et al., 2013)). This idea has been gaining support from multiple studies in mice showing that  $T_{\text{Reg}}$  cell deficiency in the periimplantation period, induced experimentally via a variety of means, causes either implantation failure or embryo resorption soon after implantation. In some studies (Aluvihare et al., 2004, Shima et al., 2010) but not others (Teles et al., 2013a), this failure occurs in allogeneic but not syngeneic mating combinations, suggesting that T cell responses to alloantigens present in semen might be important for implantation. Consistent with this possibility, maternal CD8 T cells undergo a transient wave of proliferation in the uterine LN following copulation in response to a surrogate seminal antigen (Moldenhauer *et al.*, 2009). Interestingly, in this model, seminal fluid rather than spermatozoa was the antigencontaining material.

A specific, intrauterine requirement for  $T_{Reg}$  cells in implantation was first suggested by the observation that  $CD4^+$   $CD25^+$   $FOXP3^+$   $T_{Reg}$  cell frequencies also transiently increase in the uterine LN on E3.5 (but not in the non-uterine LN), concurrent with the wave of antigeninduced T cell proliferation described above (Guerin et al., 2011). In addition,  $FOXP3^+$ cells, which are presumably also  $T_{\text{Reg}}$  cells, were found to be greater in number in the uteri of mice on E3.5 of pregnancy as compared to the uteri of mice in estrous. Both of these phenomena required seminal plasma, which, provocatively, is rich in the  $T_{\text{Reg}}$  cell-inducing cytokine TGF-β (Sharkey *et al.*, 2012). Copulation also induced expression of the CCL19 chemokine by uterine epithelial cells, which was suggested to be a recruitment factor for uterine  $T_{\text{Reg}}$  cells (Guerin *et al.*, 2011). In another study, mice deficient in CCR7, the receptor for CCL19, both lacked uterine Tregs and also showed reduced fertility (Teles et al., 2013a). This study also showed evidence of uterine fibrosis in mice lacking  $T_{\text{Re} \rho}$  cells.

Together these data suggest a scenario whereby  $T_{\text{Reg}}$  cells, induced in the uterine LN in response to semen and seminal antigens, home back to the uterus where they prepare the endometrium for implantation (Figure 2) (for further discussion, see (Robertson et al., 2013)). Presumably, the  $T_{\text{Reg}}$  cells in part act to dampen uterine inflammation, which is induced by semen (De et al., 1991). However, several issues still need to be addressed before the existence of this pathway can be firmly established. First, the extent to which antigen recognition is important for post-copulation  $T_{Reg}$  cell expansion and ultimate  $T_{Reg}$ cell function in the uterus needs to be determined. This is a key issue because postulating a specific requirement for  $i_{\text{Reg}}$  cells in the peri-implantation period would need to be reconciled with the absence of a major implantation defect in mice that are specifically deficient in these cells (Samstein et al., 2012). Second, it will be important to determine whether the  $T_{\text{Reg}}$  cells that expand in the uterine LN following insemination selectively home to the uterus, as this would imply the existence of specific, and currently unappreciated, uterine-homing pathways. Indeed, it remains possible that the requirement for CCR7 in generating the cells actually reflects the CCR7 dependence of DC emigration from the reproductive tract (Collins *et al.*, 2009), which is expected to be necessary for the presentation of seminal antigens in the draining LN. Third, the idea that the antiinflammatory effects of  $T_{\text{Reg}}$  cells are important for preparing the uterus for implantation needs to be reconciled with the growing sense that a modest amount of uterine inflammation promotes implantation (Dekel et al., 2010).

Lastly, and most importantly, the current set of data does not resolve whether  $T_{\text{Reg}}$  cells act locally within the uterus, or systemically, or both. Indeed,  $T_{Reg}$  depletion induces systemic inflammation (Kim et al., 2007), and the peri-implantation period is particularly sensitive to inflammation-induced ovarian insufficiency (Erlebacher et al., 2004). Thus, it remains possible that implantation failure in the absence of sufficient  $T_{\text{Reg}}$  cell function systemically is due to impaired progesterone production by the corpus luteum. In fact, the requirement for  $T_{\text{Reg}}$  cells during the peri-implantation period might even reflect a crucial role for  $T_{\text{Reg}}$  cells within the ovary itself, a possibility suggested by work in cows showing that the luteolytic effect of prostaglandin PGF<sub>2 $\alpha$ </sub> is associated with reductions in ovarian T<sub>Reg</sub> cell numbers (Poole and Pate, 2012).

#### **Preeclampsia and spontaneous abortion**

Potential roles for decidual  $T_{\text{Reg}}$  cells have also emerged from studies on the pathogenesis of preeclampsia and spontaneous abortion (for review, see (Cerdeira et al., 2012, Ernerudh et al., 2011)). Stimulated by reports of reduced  $T_{\text{Reg}}$  cell frequencies in the peripheral blood of pregnant women with preeclampsia compared to control pregnant women (Darmochwal-Kolarz et al., 2007, Prins et al., 2009, Santner-Nanan et al., 2009, Sasaki et al., 2007, Toldi et al., 2012) (a finding not replicated with other cohorts (Hu et al., 2008, Paeschke et al., 2005)), it was initially suggested that  $T_{\text{Reg}}$  frequencies at the maternal-fetal interface were also reduced in preeclampsia (Quinn et al., 2011, Sasaki et al., 2007). A recent report failed to reproduce this observation, but instead revealed that preeclamptic patients showed specific reductions decidual frequencies of Helios<sup>−</sup> FOXP3<sup>+</sup> CD25<sup>+</sup> CD4<sup>+</sup> cells, which presumably represent iT<sub>Reg</sub> cells (Hsu *et al.*, 2012). This defect was furthermore linked to an altered phenotype of decidual DC-SIGN+ APCs, which were found to be located in proximity to decidual  $T_{\text{Reg}}$  cells. Reduced decidual  $T_{\text{Reg}}$  cell frequency or function has also been reported in cases of spontaneous abortion (Inada et al., 2013, Sasaki et al., 2004, Schumacher et al., 2009, Wang et al., 2010), sometimes in association with increased  $T_H$ 17 cell frequencies (Wang et al., 2010).

Given the well-known immunosuppressive functions of  $T_{\text{Reg}}$  cells, these data together suggest a scenario in which  $T_{Reg}$  cells expand within the decidua, potentially as the result of the local presentation of placental antigens by decidual APCs, in order to limit effector T cell activity at the maternal-fetal interface (Ernerudh *et al.*, 2011). Without such a limit, effector T cells (i.e.  $T_H1$  cells,  $T_H17$  cells and CTLs) are freer to induce pregnancy pathologies ranging from first trimester fetal loss to inadequate spiral artery remodeling. Exact effector mechanisms for a given pathology, however, are unknown and an important area of research. In addition,  $T_{\text{Reg}}$  cells might also generally limit decidual inflammation through antigen non-specific mechanisms. This latter possibility is consistent with the cells' secretion of anti-inflammatory factors such as IL-10 and TGF-β, which will have broad effects, as well as with emerging data in mice discussed above that uterine  $T_{\text{Reg}}$  cells play an anti-inflammatory role in preparing the uterus for embryo implantation. Such a latter role would obviously be independent of placental antigen exposure. Importantly, elevated proportions of decidual  $T_H$ 17 cells *per se* have been linked to the pathogenesis of spontaneous abortion (Nakashima *et al.*, 2010, Wang *et al.*, 2010), and recent work in both mice and humans (Fu *et al.*, 2013) suggests that decidual NK cells, through their production of IFN $\gamma$ , are also capable of suppressing decidual T<sub>H</sub>17 cell accumulation. Thus, elevated decidual  $T_H$ 17 cell frequencies in spontaneous abortion have also been associated with low NK cell numbers (Fu et al., 2013). Together, these data build upon older work and have led to a revised Th1/Th2/Th17/T<sub>Reg</sub> cell paradigm for pregnancy success (Saito *et al.*, 2010).

#### **VUE and chronic deciduitis**

Overt, histologically apparent T cell accumulations, akin to those seen with organ transplant rejection, are not findings typically associated with preeclampsia or spontaneous abortion. Thus, the contribution of T cells to these pathologies is thought to be mainly due to changes in T cell subset proportions and altered T cell functions. In contrast, villitis of unknown etiology (VUE), chronic deciduitis, and chronic chorioamnionitis are three histological

findings of the third trimester placenta where maternal T cells do accumulate at the maternal-fetal interface (Erlebacher, 2013a). Given the greater amount of relevant literature, we will discuss the pathogenesis and significance of VUE, with the recognition that the set of questions this lesion raises also applies to chronic deciduitis and chronic chorioamnionitis.

VUE affects 5-15% of all births and is characterized by focal accumulations of maternal T cells, comprised of both CD4 and CD8 T cells, as well as fetal macrophages (i.e. Hofbauer cells) (for reviews, see (Redline, 2007, Tamblyn et al., 2013)). The lesions are primarily located in the distal placental villi, but it has been suggested that they arise from leukocytes that have first infiltrated into the decidua before spreading vertically via the anchoring villi to reach the villus tree. Its origin is not thought to be infectious. From a clinical viewpoint, VUE is an interesting lesion since its incidence is increased in several complications of human pregnancy, including idiopathic preterm birth and intrauterine growth restriction. Its causal relationship with these pregnancy complications is at present unclear. The lesion is associated with increased placental mRNA expression of CXCL9, CXCL10, CXCL11 and  $CL5$  (Freitag *et al.*, 2013, Kim *et al.*, 2009), as well as focal expression of CXCL9, CXCL10, and CCL5 protein at the sites of leukocyte accumulation (Kim *et al.*, 2009). T<sub>H</sub>1 cells are thought to primarily comprise the CD4 T cell component of the response, while the CD8 T cells are thought to be CTLs.

From a basic viewpoint, VUE is also interesting because its histological features are what might be expected if maternal T cells were to be rejecting the placenta. Considering this possibility raises the interesting question of the antigen specificity of the infiltrating T cells, which remains unknown. This is a key question since it speaks to underlying pathogenic mechanisms. If the T cells are specific for placental antigens, then there must have been at some point a breakdown in those pathways that normally prevent the immunogenic presentation of placental antigens to maternal T cells. In addition, there must have been a breakdown in mechanisms that prevent  $T_H1$  cell and CTL infiltration into the decidua and placenta. Indeed, the focal nature of VUE in itself points to the potential existence of such mechanisms, since otherwise such activated T cells should show a diffuse pattern of infiltration. Indeed, we have previously conjectured that focal dysregulation of the epigenetic pathways that silence Cxcl9, Cxcl10, and Ccl5 expression in mouse DSCs, if these pathways are also active in human DSCs, might underlie at least the decidual component of VUE (Erlebacher, 2013a). It also is possible that maternal T cell directly enter the placental tree as the result of a focal damage or because syncytiotrophoblasts (which form the cellular layer of the villus tree in contact with maternal blood) have upregulated certain key adhesion molecules (Redline, 2007, Tamblyn et al., 2013)).

If, on the other hand, the infiltrating T cells in VUE are not specific for placental antigens, then work on underlying pathogenic mechanisms needs to consider only how these T cells gain access to the decidua and placenta, and could consider the same set of hypotheses as if they did have placental specificity. Interestingly, the possibility that the infiltrating T cells are bystanders (with respect to fetal and placental antigens) raises the possibility that they were generated at distal sites, perhaps in response to infection.

Importantly, although indirect presentation pathways are thought to mediate T cell recognition of placental antigens under physiological conditions in both mice and humans, as discussed above, it remains possible that the T cells within VUE lesions are specific for intact paternal MHC molecules. In the case of CD8 T cells, the cells might be interacting directly with paternal HLA-C molecules expressed not only by extravillous trophoblasts (for those areas of the lesion within the decidua) but also with all MHC class I molecules expressed by the fetus-derived non-trophoblastic stromal constituents of the villi including fibroblasts, endothelial cells and Hofbauer cells. In the case of CD4 T cells, the cells might be interacting directly with Hofbauer cells, which express MHC class II molecules in VUE lesions. If such directly alloreactive maternal T cells are indeed constituents of VUE lesions, the question again arises of how these cells first become activated.

#### **Conclusions**

Importantly, the above pathological scenarios view effector T cells within the decidua as having a negative impact upon pregnancy success. This idea is certainly more likely if a large proportion of these T cells turn out to have specificity for placental antigens. However, recent work suggesting that the human maternal-fetal interface is normally colonized with a wide variety of bacteria (Stout et al., 2013) suggests that effector  $T$  cells within the decidua might perform a positive function in controlling infection. If this is true, then this function must be very finely tuned so that the response, on the one hand, is sufficient to prevent progression to pregnancy endpoints such as acute infectious chorioamnionitis, while on the other hand is not so robust that the attendant inflammation in fighting the infection leads to pregnancy complications such as preterm labor secondary to non-infectious chorioamnionitis. As discussed further elsewhere (Erlebacher, 2013a, Tilburgs and Strominger, 2013), the existence of intra-decidual mechanisms to limit maternal T cell responses against the placenta might also explain why the decidua is susceptible to infection by certain organisms such as Listeria monocytogenes and cytomegalovirus.

Indeed, how intradecidual T cell behavior is balanced in order to accommodate the competing demands of reproduction and host defense can now be viewed as a key question facing research on the biology of decidual T cells. As discussed above, other important unresolved questions include the pathways that regulate intradecidual T cell function, the antigen specificity of decidual T cells, and the degree to which dysregulation of decidual T cells function or migration are causative pathogenic factors in human pregnancy complications. It is hoped that the current surge in interest in this critical area of reproductive immunology will lead to insights with potential impact on human reproductive health.

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#### **Abbreviations**

**NK cell** natural killer cell



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#### **Figure 1. T cell behavior at the maternal-fetal interface**

A schematized representation of the pregnant uterus, illustrating several of the regulatory pathways discussed in this review. (A) CD83+ DCs form clusters with decidual T cells, presumably as a consequence of ongoing antigen presentation. The significance of this observation is not currently known, nor are the T cell subsets involved (naïve, regulatory, effector, memory). (B) The potential role of decidual CD14+ DC-SIGN+ APCs in converting naïve decidual T cells into  $T_{Reg}$  cells. (C, D) The potential role of NK cells and  $T_{Reg}$  cells in repressing decidual CTLs,  $T_H1$  and  $T_H17$  cells. (E) Impeded  $T_H1$  cell and CTL recruitment to the decidua from the blood as a result of Cxcl9, Cxcl10, Cxcl11, and Ccl5 chemokine gene silencing in decidual stromal cells. Naïve  $T$  cells and  $T_{Reg}$  cells are shown entering the decidua from maternal blood, however there is currently no direct evidence that this happens to a major extent. The idea that naïve T cells enter into the decidua from the blood is particularly unlikely.



**Figure 2. TReg cell generation in response to semen in the pre-implantation mouse uterus** Two complementary mechanisms are shown; the evidence for the first is discussed in the text. (A-E) Under the influence of seminal fluid rich in TGF- $\beta$  (A), DCs ingest and process seminal fluid antigens and then migrate via lymphatic vessels (B) to the uterine LN. In these LN, they present seminal fluid antigens and convert responding naïve  $T$  cells into  $T_{Reg}$  cells due to the influence of TGF-β (C). Of note, it is unclear whether the TGF-β that acts during this phase of the response is derived from seminal fluid, or is produced by maternal cells. Once generated, the  $T_{Reg}$  cells migrate via the blood back to the uterus (D), possibly attracted by CCL19 induced by uterine epithelial cells in response to semen (E). (A, F) Alternatively, uterine DCs might present seminal fluid antigens to naïve T cells within the uterus itself, which locally induces iT<sub>Reg</sub> cell generation due to the high levels of TGF-β in seminal fluid.