# **UC Berkeley**

## **UC Berkeley Previously Published Works**

## **Title**

Draft Genome Sequences of Two Janthinobacterium lividum Strains, Isolated from Pristine Groundwater Collected from the Oak Ridge Field Research Center

#### **Permalink**

https://escholarship.org/uc/item/83v40767

### **Journal**

Microbiology Resource Announcements, 5(26)

#### **ISSN**

2169-8287

#### **Authors**

Wu, Xiaoqin Deutschbauer, Adam M Kazakov, Alexey E et al.

#### **Publication Date**

2017-06-29

#### DOI

10.1128/genomea.00582-17

Peer reviewed







# Draft Genome Sequences of Two Janthinobacterium lividum Strains, Isolated from Pristine Groundwater Collected from the Oak Ridge Field Research Center

Xiaoqin Wu,<sup>a</sup> Adam M. Deutschbauer,<sup>b</sup> Alexey E. Kazakov,<sup>b</sup> Kelly M. Wetmore,<sup>b,c</sup> Bryson A. Cwick,<sup>a</sup> Robert M. Walker,<sup>a</sup> Pavel S. Novichkov,<sup>b</sup> Adam P. Arkin,<sup>b,d</sup> Romy Chakraborty<sup>a</sup>

Earth and Environmental Sciences, Lawrence Berkeley National Laboratory, Berkeley, California, USAa; Environmental Genomics and Systems Biology Division, Lawrence Berkeley National Laboratory, Berkeley, California, USAb; Program in Comparative Biochemistry, University of California, Berkeley, California, USAc; Department of Bioengineering, University of California, Berkeley, California, USAd

**ABSTRACT** We present here the draft genome sequences of two *Janthinobacterium lividum* strains, GW456P and GW458P, isolated from groundwater samples collected from a background site at the Oak Ridge Field Research Center. Production of a purple pigment by these two strains was observed when grown on diluted (1/10) LB agar plates.

The genus *Janthinobacterium* includes rod-shaped, Gram-negative, motile, aerobic bacteria that are known to produce the water-insoluble purple pigment violacein (1, 2). Violacein has been known to have antimicrobial and antiviral properties (1, 3, 4), inhibiting the growth of bacteria (5, 6) and providing bacteria protection against predators (7, 8). To date, only a few environmental isolates of *Janthinobacterium* spp. have been sequenced (2), and these were isolated from lake sediments (9) and soils (10, 11), mostly in cold habitats (12–16). Until now, no sequenced representatives of *Janthinobacterium* spp. have been available from the subsurface aquatic environment. In this study, two violet-pigmented *J. lividum* strains were isolated on diluted (1/10) LB agar plates, from groundwater samples collected from two wells (GW456 and GW458) located at the background area of the Oak Ridge Field Research Center, Oak Ridge, Tennessee, USA.

Genomic DNA was extracted using the PureLink Genomic DNA mini kit (Invitrogen). Genomic sequencing libraries of the two *J. lividum* strains, GW456P and GW458P, were prepared using the NEBNext DNA library prep kit for Illumina (New England Biolabs). Briefly, 1  $\mu$ g of genomic DNA was fragmented by ultrasonication to an average size of 800 bp with a Covaris S220 focused ultrasonicator. After end-repair, A-tailing, and ligation of the adapter, we size-selected 800-bp products with AMPure XP beads. The sequencing libraries were quantified on a bioanalyzer with a DNA1000 chip (Agilent). We performed paired-end sequencing (2  $\times$  150 bp) on an Illumina MiSeq using a MiSeq version 2 reagent kit (300 cycles).

The raw reads were assembled using the A5 microbial assembly pipeline version 0.0.4 (17), implemented within the U.S. Department of Energy's Systems Knowledge Database (Kbase; http://www.kbase.us) (18). This pipeline automates the processes of read cleaning, error correction, contig assembly, crude scaffolding, misassembly correction, and final scaffolding with stringent parameters repairing previously broken contigs. The draft genome sequence of strain GW456P was 6.27 Mb in length, in 78

**Received** 4 May 2017 **Accepted** 15 May 2017 **Published** 29 June 2017

**Citation** Wu X, Deutschbauer AM, Kazakov AE, Wetmore KM, Cwick BA, Walker RM, Novichkov PS, Arkin AP, Chakraborty R. 2017. Draft genome sequences of two *Janthinobacterium lividum* strains, isolated from pristine groundwater collected from the Oak Ridge Field Research Center. Genome Announc 5: e00582-17. https://doi.org/10.1128/genomeA

**Copyright** © 2017 Wu et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Romy Chakraborty, rchakraborty@lbl.gov.

Wu et al. genameAnnouncements"

contigs, and the total G+C content was 62.89%. For strain GW458P, the genome size was 6.29 Mb, consisting of 156 contigs with a total G+C content of 63.29%.

Automated annotation for each genome was performed using the Joint Genome Institute's Integrated Microbial Genomics (IMG) version 4.12.0 annotation pipeline (19). Proteins related to violacein biosynthesis were identified in both strains by comparison with the VioA, VioB, VioC, VioD, and VioE proteins from *Janthinobacterium* sp. HH01 (2) using the GenomeExplorer program (20). We also identified three genes known to regulate violacein biosynthesis (2): the sensor kinase *jqsS*, the response regulator *jqsR*, and the autoinducer synthase *jqsA*. Upstream regions of the *vioABCDE* operons in both strains contain the inverted repeat TTGATATTTATCAA, which coincides with the published JqsR binding motif (21). Thus, we propose that the expression of violacein biosynthesis operons in strains GW456P and GW458P is dependent on quorum sensing. These genomes are useful for exploring the physiology and regulation of violacein pigment production by these two *J. lividum* strains.

**Accession number(s).** Genomes of *J. lividum* strain GW456P and *J. lividum* strain GW458P have been deposited at DDBJ/ENA/GenBank under the accession numbers NEHB00000000 and NEGZ00000000, respectively.

#### **ACKNOWLEDGMENT**

This material by ENIGMA-Ecosystems and Networks Integrated with Genes and Molecular Assemblies (http://enigma.lbl.gov), a Scientific Focus Area Program at Lawrence Berkeley National Laboratory, is based upon work supported by the U.S. Department of Energy, Office of Science, Office of Biological and Environmental Research under contract number DE-AC02-05CH11231.

#### **REFERENCES**

- Rodrigues AL, Göcke Y, Bolten C, Brock NL, Dickschat JS, Wittmann C. 2012. Microbial production of the drugs violacein and deoxyviolacein: analytical development and strain comparison. Biotechnol Lett 34: 717–720. https://doi.org/10.1007/s10529-011-0827-x.
- Hornung C, Poehlein A, Haack FS, Schmidt M, Dierking K, Pohlen A, Schulenburg H, Blokesch M, Plener L, Jung K, Bonge A, Krohn-Molt I, Utpatel C, Timmermann G, Spieck E, Pommerening-Röser A, Bode E, Bode HB, Daniel R, Schmeisser C, Streit WR. 2013. The *Janthinobacterium* sp. HH01 genome encodes a homologue of the *V. cholerae* CqsA and *L. pneumophila* LqsA autoinducer synthases. PLoS One 8:e55045. https://doi.org/10.1371/journal.pone.0055045.
- Durán N, Justo GZ, Ferreira CV, Melo PS, Cordi L, Martins D. 2007. Violacein: properties and biological activities. Biotechnol Appl Biochem 48:127–133. https://doi.org/10.1042/BA20070115.
- Andrighetti-Fröhner CR, Antonio RV, Creczynski-Pasa TB, Barardi CR, Simões CM. 2003. Cytotoxicity and potential antiviral evaluation of violacein produced by *Chromobacterium violaceum*. Mem Inst Oswaldo Cruz 98:843–848. https://doi.org/10.1590/S0074-02762003000600023.
- 5. Durán N. 1990. Violaceina: a descoberta de um antibiótico. Ciencia Hoje 11:58 80.
- Durán N, Erazo S, Campos V. 1983. Bacterial chemistry-II: antimicrobial photoproduct from pigment of *Chromobacterium violaceum*. Anais Acad Brasil Cienc 55:231–234.
- Matz C, Deines P, Boenigk J, Arndt H, Eberl L, Kjelleberg S, Jürgens K. 2004. Impact of violacein-producing bacteria on survival and feeding of bacterivorous nanoflagellates. Appl Environ Microbiol 70:1593–1599. https://doi.org/10.1128/AEM.70.3.1593-1599.2004.
- Matz C, Webb JS, Schupp PJ, Phang SY, Penesyan A, Egan S, Steinberg P, Kjelleberg S. 2008. Marine biofilm bacteria evade eukaryotic predation by targeted chemical defense. PLoS One 3:e2744. https://doi.org/10 .1371/journal.pone.0002744.
- McTaggart TL, Shapiro N, Woyke T, Chistoserdova L. 2015. Draft genome of *Janthinobacterium* sp. RA13 isolated from Lake Washington sediment. Genome Announc 3(1):e01588-14. https://doi.org/10.1128/genomeA .01588-14.
- Shoemaker WR, Muscarella ME, Lennon JT. 2015. Genome sequence of the soil bacterium *Janthinobacterium* sp. KBS0711. Genome Announc 3(3):e00689-15. https://doi.org/10.1128/genomeA.00689-15.

- Valdes N, Soto P, Cottet L, Alarcon P, Gonzalez A, Castillo A, Corsini G, Tello M. 2015. Draft genome sequence of *Janthinobacterium lividum* strain MTR reveals its mechanism of capnophilic behavior. Stand Genomic Sci 10:110 https://doi.org/10.1186/s40793-015-0104-z.
- Shivaji S, Ray MK, Kumar GS, Reddy GSN, Saisree L, Wynn-Williams DD. 1991. Identification of *Janthinobacterium lividum* from the soils of the islands of Scotia Ridge and from Antarctic peninsula. Polar Biol 11: 267–271. https://doi.org/10.1007/BF00238461.
- Kim SJ, Shin SC, Hong SG, Lee YM, Lee H, Lee J, Choi IG, Park H. 2012. Genome sequence of *Janthinobacterium* sp. strain PAMC 25724, isolated from alpine glacier cryoconite. J Bacteriol 194:2096. https://doi.org/10 .1128/JB.00096-12.
- Gong X, Skrivergaard S, Korsgaard BS, Schreiber L, Marshall IPG, Finster K, Schramm A. 2017. High quality draft genome sequence of Janthinobacterium psychrotolerans sp. nov., isolated from a frozen freshwater pond. Stand Genomic Sci 12:8. https://doi.org/10.1186/ s40793-017-0230-x.
- Smith HJ, Foreman CM, Akiyama T, Franklin MJ, Devitt NP, Ramaraj T. 2016. Genome sequence of *Janthinobacterium* sp. CG23\_2, a violaceinproducing isolate from an Antarctic supraglacial stream. Genome Announc 4(1):e01468-15. https://doi.org/10.1128/genomeA.01468-15.
- Koo H, Strope BM, Kim EH, Shabani AM, Kumar R, Crowley MR, Andersen DT, Bej AK. 2016. Draft Genome Sequence of *Janthinobacterium* sp. Ant5-2-1, isolated from proglacial Lake Podprudnoye in the Schirmacher Oasis of east Antarctica. Genome Announc 4(1):e01600-15. https://doi.org/10.1128/genomeA.01600-15.
- Coil D, Jospin G, Darling AE. 2015. A5-miseq: an updated pipeline to assemble microbial genomes from Illumina MiSeq data. Bioinformatics 31:587–589. https://doi.org/10.1093/bioinformatics/btu661.
- 18. Arkin AP, Stevens RL, Cottingham RW, Maslov S, Henry CS, Dehal P, Ware D, Perez F, Harris NL, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Gunter D, Murphy-Olson D, Chan S, Kamimura RT, Brettin TS, Meyer F, Chivian D, Weston DJ, Glass EM, Davison BH, Kumari S, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia J, Chia J, Colasanti R, Conrad N, Davis JJ, DeJongh M, Devoid S, Dietrich E, Drake MM, Dubchak I, Edirisinghe JN, Fang G, Faria JP, Frybarger PM, Gerlach W, Gerstein M, Gurtowski J, Haun HL, He F, Jain R, Joachimiak MP, Keegan KP, Kondo S, Kumar V, Land ML, Mills M, Novichkov P, Oh T,

genomea.asm.org 2

Volume 5 lssue 26 e00582-17

Genome Announcement gen∎meAnnouncements™

- Olsen GJ, Olson B, Parrello B, Pasternak S, Pearson E, Poon SS, Price G, Ramakrishnan S, Ranjan P, Ronald PC, Schatz MC, Seaver SMD, Shukla M, Sutormin RA, Syed MH, Thomason J, Tintle NL, Wang D, Xia F, Yoo H, Yoo S. 2016. The DOE systems biology Knowledgebase (KBase). bioRxiv 096354. https://doi.org/10.1101/096354.
- Markowitz VM, Chen IM, Palaniappan K, Chu K, Szeto E, Pillay M, Ratner A, Huang J, Woyke T, Huntemann M, Anderson I, Billis K, Varghese N, Mavromatis K, Pati A, Ivanova NN, Kyrpides NC. 2014. IMG 4 version of the integrated microbial genomes comparative analysis system. Nucleic Acids Res 42:D560–D567. https://doi.org/10.1093/nar/gkt963.
- Mironov AA, Vinokurova NP, Gelfand MS. 2000. Software for analysis of bacterial genomes. Mol Biol 34:222–231. https://doi.org/10.1007/ BF02759643.

 Haack FS, Poehlein A, Kröger C, Voigt CA, Piepenbring M, Bode HB, Daniel R, Schäfer W, Streit WR. 2016. Molecular keys to the *Janthinobacterium* and *Duganella* spp. interaction with the plant pathogen *Fusarium graminearum*. Front Microbiol 7:1668 https://doi.org/10.3389/fmicb.2016.01668.

Volume 5 Issue 26 e00582-17 genomea.asm.org **3**