## **UC Irvine** UC Irvine Previously Published Works

## Title

Kidney in the net of acute and long-haul coronavirus disease 2019: a potential role for lipid mediators in causing renal injury and fibrosis.

## Permalink

https://escholarship.org/uc/item/8422c34c

## Journal

Current opinion in nephrology and hypertension, 31(1)

ISSN

1062-4821

## Authors

Chiang, Kate C Imig, John D Kalantar-Zadeh, Kamyar <u>et al.</u>

# **Publication Date**

2022

**DOI** 10.1097/mnh.000000000000750

## **Copyright Information**

This work is made available under the terms of a Creative Commons Attribution License, available at <a href="https://creativecommons.org/licenses/by/4.0/">https://creativecommons.org/licenses/by/4.0/</a>

Peer reviewed



# Kidney in the net of acute and long-haul coronavirus disease 2019: a potential role for lipid mediators in causing renal injury and fibrosis

Kate C. Chiang<sup>a</sup>, John D. Imig<sup>b</sup>, Kamyar Kalantar-Zadeh<sup>c</sup>, and Ajay Gupta<sup>a,c</sup>

#### **Purpose of review**

Severe COVID-19 disease is often complicated by acute kidney injury (AKI), which may transition to chronic kidney disease (CKD). Better understanding of underlying mechanisms is important in advancing therapeutic approaches.

#### **Recent findings**

SARS-CoV-2-induced endothelial injury initiates platelet activation, platelet–neutrophil partnership and release of neutrophil extracellular traps. The resulting thromboinflammation causes ischemia–reperfusion (I/R) injury to end organs. Severe COVID-19 induces a lipid-mediator storm with massive increases in thromboxane  $A_2$  (Tx $A_2$ ) and PGD<sub>2</sub>, which promote thromboinflammation and apoptosis of renal tubular cells, respectively, and thereby enhance renal fibrosis. COVID-19-associated AKI improves rapidly in the majority. However, 15-30% have protracted renal injury, raising the specter of transition from AKI to CKD.

#### Summary

In COVID-19, the lipid-mediator storm promotes thromboinflammation, ischemia-reperfusion injury and cytotoxicity. The thromboxane  $A_2$  and  $PGD_2$  signaling presents a therapeutic target with potential to mitigate AKI and transition to CKD. Ramatroban, the only dual antagonist of the thromboxane  $A_2$ /TPr and  $PGD_2/DPr2$  signaling could potentially mitigate renal injury in acute and long-haul COVID. Urgent studies targeting the lipid-mediator storm are needed to potentially reduce the heavy burden of kidney disease emerging in the wake of the current pandemic.

#### Keywords

acute kidney injury, coronavirus disease 2019, prostaglandin D<sub>2</sub>, thromboinflammation, thromboxane

#### INTRODUCTION

Amidst the coronavirus disease 2019 (COVID-19) pandemic, the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) virus has caused immense suffering while placing tremendous burden on healthcare systems worldwide. Although mild cases present with common cold symptoms, severe COVID-19 disease is associated with multiorgan failure and dysfunction of the lung, kidney, heart, liver and brain. Acute kidney injury (AKI) has emerged as a common complication in hospitalized patients with severe COVID-19 with acute tubular injury as the major histopathological finding [1,2]. The pathogenesis of COVID-19-induced AKI (CoV2-AKI) is poorly understood.

End organ damage in COVID-19 is fueled by thromboinflammation characterized by platelet and neutrophil activation, and release of neutrophil extracellular traps (NETs) [3<sup>•</sup>] (Fig. 1). It has been proposed that thromboinflammation in COVID-19 is mediated by a cytokine storm. However, cytokine release in COVID-19 is comparable to if not lower than the cytokine levels in influenza patients [4], and yet alveolar capillary microthrombi are nine

Curr Opin Nephrol Hypertens 2022, 31:36-46

DOI:10.1097/MNH.000000000000750

Volume 31 • Number 1 • January 2022

<sup>&</sup>lt;sup>a</sup>KARE Biosciences, Orange, California, <sup>b</sup>Drug Discovery Center and Cardiovascular Center, Medical College of Wisconsin, Milwaukee, Wisconsin, USA and <sup>c</sup>Division of Nephrology, Hypertension and Kidney Transplantation, Department of Medicine, University of California Irvine (UCI) School of Medicine, Orange, California, USA

Correspondence to Ajay Gupta, MBBS, MD, Division of Nephrology, Hypertension and Kidney Transplantation, Department of Medicine, University of California Irvine (UCI) School of Medicine, Orange, CA 92868, USA. Tel: +1 562 419 7029; e-mail: ajayg1@hs.uci.edu

## **KEY POINTS**

- Severe COVID-19-induced lipid mediator storm in the lungs is associated with marked increase in thromboxane A<sub>2</sub> (TxA<sub>2</sub>), PGD<sub>2</sub> and leukotoxin diols.
- Thromboxane A<sub>2</sub> promotes thromboinflammation and thereby ischemia-reperfusion injury.
- PGD<sub>2</sub>/DPr2 signaling promotes a Th2 immune response and apoptosis of renal tubular cells.
- Leukotoxin diols induce mitochondrial dysfunction and cytotoxicity.
- The lipid mediator storm presents a therapeutic target in acute and long-haul COVID-19 with potential to mitigate AKI and transition to CKD.

times more prevalent in COVID-19 compared with influenza [5<sup>••</sup>]. Emerging evidence supports a lipidmediator storm fueling maladaptive immune responses and thrombotic events, all of which play a role in thromboinflammation [6<sup>•</sup>,7<sup>••</sup>]. In 33 hospitalized COVID-19 patients requiring mechanical ventilation, lipid analysis of bronchoalveolar lavage fluid (BALF) revealed massive increase in thromboxane levels [7<sup>••</sup>]. Here, we postulate the possible mechanisms of AKI during acute SARS-CoV-2 infection in the context of the lipid mediator storm. We also speculate on the impact of prolonged lipid mediator storm in the progression of chronic kidney disease in long-haul COVID-19.

#### CLINICAL SYNDROME OF ACUTE KIDNEY INJURY AND CHRONIC KIDNEY DISEASE IN CORONAVIRUS DISEASE 2019

Although COVID-19 is predominantly a respiratory disorder, the kidney is one of the most common organs affected by COVID-19 second only to the lung [8]. AKI is emerging as a common and important sequelae of COVID-19, with rates as high as 33-43% in hospitalized patients [9–12]. CoV2-AKI may present as collapsing glomerulopathy, acute interstitial nephritis, de novo glomerular disease, prerenal azotemia or acute tubular injury [1,13]. In kidney biopsy, samples from 17 mild COVID-19 patients, acute tubular injury, collapsing glomerulopathy, endothelial injury or thrombotic microangiopathy were the most common histological findings [14]. Interestingly, virus detection was negative in the patient samples [14], and most kidney biopsies performed several weeks after the onset of COVID-19 symptoms failed to show notable SARS-CoV-2 infection [15]. Although some studies have reported SARS-CoV-2 particles in urine samples [16–18] or in kidney glomerular compartments [19], a direct role of the virus in the development of AKI remains to be demonstrated [15].

Most patients who survive CoV2-AKI regain kidney function but up to 30% may remain on dialysis at discharge [20]. Moreover, in a multicenter cohort study of 3099 hospitalized adult COVID-19 patients, one in five patients developed AKI requiring renal replacement therapy (RRT) [11]. Of those who survived and were discharged, one in three remained RRT-dependent at discharge, and one in six remained RRT-dependent 60 days after ICU admission [11]. This raises the specter of transition from acute to chronic kidney disease. Better understanding of mechanisms underlying COVID-19-induced AKI are critical to designing therapeutic strategies.

#### MECHANISMS OF CORONAVIRUS DISEASE 2019-INDUCED ACUTE KIDNEY INJURY

The mechanisms underlying the development of CoV2-AKI are likely multifactorial, including viral septicaemia, an enhanced inflammatory response, endothelial damage, hypercoagulability, myocardial dysfunction, drug nephrotoxicity and the effects of general hypoxia and dehydration on renal perfusion [21]. Although there is evidence of an association between proinflammatory cytokines and kidney injury [22,23], few patients with COVID-19 exhibit cytokine profiles indicative of cytokine storm syndrome [4]. In fact, patients with COVID-19 exhibited lower cytokine levels than patients with influenza [4]. Therefore, other underlying mechanisms may play a greater role in fueling the thromboinflammation and AKI in COVID-19 as discussed in this review (Table 1).

#### Endothelial injury and dysfunction

SARS-CoV-2 virus infects endothelial cells causing diffuse endothelialitis, intussusceptive angiogenesis and impaired microcirculation in vascular beds [5<sup>••</sup>,24,25]. Endothelialitis and pyroptosis lead to release of endothelial microvesicles, which activate leukocytes and platelets through surface interaction, receptor activation, cellular fusion and the delivery of intravesicular cargo [25,26]. Endothelial cell injury, platelet activation, platelet–leukocyte aggregates and thrombosis in severe COVID-19 disease is evidenced by elevated serum levels of soluble P-selectin, von Willebrand factor, soluble thrombomodulin and soluble CD40L [3<sup>•</sup>,27<sup>•</sup>–29<sup>•</sup>]. Endothelial cells and platelets generate TxA<sub>2</sub>, a key mediator of platelet activation and thrombosis.

1062-4821 Copyright  $\ensuremath{\mathbb{C}}$  2021 Wolters Kluwer Health, Inc. All rights reserved.



FIGURE 1. Potential mechanisms of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) induced acute kidney injury and transition to chronic kidney disease in long-haul coronavirus disease. SARS-CoV-2 virus binds complement MASP-2, thereby activating the lectin pathways and production of membrane attack complex (MAC) in renal vascular beds and tubuli. MAC synergizes with TLR signaling to promote inflammation, and contributes to chemotaxis, leukocyte adhesion and systemic thrombosis. MAC deposition on endothelial cells along with necrosis and pyroptosis of endothelial cells, platelets and monocytes, leads to IL-1 $\alpha$  production and release. In turn, IL-1 $\alpha$  stimulates the IL-1 receptor expressed on endothelial cells leading to COX-2 and TxAS expression. Subsequent thromboxane A<sub>2</sub> production induces endothelial and platelet activation via the TPr receptor. Activated platelets express P-selectin leading to platelet-neutrophil partnership, neutrophil activation and release of neutrophil extracellular traps (NETs), which fuel thromboinflammation in COVID-19. Renal ischemia-reperfusion injury as a result of renal thromboinflammation leads to NF-κB p65 phosphorylation. SARS-CoV-2 also directly induces NF-κB p65 phosphorylation via binding of S1 protein to ACE2 leading to ER stress and unfolded protein response (UPR) signaling constituting IRE1 and PERK protein activation, and downstream MAP kinase signaling. NF-κB p65 phosphorylation induces COX-2 expression and eicosanoid production, including upregulation of L-PGDS derived PGD<sub>2</sub>. Patients with COVID-19 also exhibit elevated levels of leukotoxin diols. Linoleate is converted into epoxyoctadecenoic acids (EpOMEs) by CYP450 epoxygenase enzymes, and further metabolized into leukotoxin diols by soluble epoxide hydrolase (sEH). PGD<sub>2</sub>/DPr2 signaling and leukotoxin diol-induced mitochondrial dysfunction causes renal proximal tubular epithelial cell apoptosis and

#### **Complement activation**

Endothelial dysfunction and thrombosis in COVID-19 is also mediated by complement activation (Fig. 1). Nucleocapsid protein of SARS-CoV-2 virus binds to the Mannan-binding lectin-associated serine protease-2 (MASP-2), the lectin pathway's effector enzyme, resulting in complement activation [30]. Kidney biopsies in nine COVID-19 patients reveal enhanced renal complement deposition in vascular beds and tubules, along with glomerular MASP-2 and tubular C5b-9 deposition in the majority of cases [8]. Complement activation and assembly of the MAC C5b-9 is known to promote endothelial damage, thrombosis and renal injury, such as in atypical hemolytic uremic syndrome [31].

In COVID-19 infection, IL-1 $\alpha$  release from necrotic and pyroptotic cells is strongly associated with the lung injury and disease severity [32,33]. MAC C5b-9 induces synthesis of IL-1 $\alpha$  from porcine endothelial cells, and direct treatment with IL-1 $\alpha$ leads to COX-2 but not COX-1 expression [34]. Therefore, complement activation and cellular necrosis-led expression of IL-1 $\alpha$  may lead to thromboinflammation via COX-2 mediated thromboxane generation as discussed below (Fig. 1).

#### Lipid mediator storm promoting thromboinflammation and maladaptive immune response

SARS-CoV-2-induced direct COX-2 expression has been observed in various cell lines and tissues *in vitro* and *in vivo* including cardiomyocytes, Calu-3 and A549 lung cancer cell lines, ciliated lung cells, primary human bronchial epithelial cells, human ACE2-expressing mouse lungs and living human lung slices [35–37]. COX-2 expression is elevated more than four-fold in SARS-CoV-2-infected Calu-3 cells and human lung slices [36] and more than 50fold in infected cardiomyocytes (Professor Srinivasa T. Reddy, UCLA, personal communication following analysis of the supplemental material in reference [36]) [37].

SARS-CoV-2-induced cardiomyocyte COX-2 expression is highly correlated with pro-inflammatory genes and signaling pathways including interferon, IL-1β and NF-κB [37,38]. Interestingly, SARS-CoV-2 infection-induced S1-ACE2 receptor interaction leads to NF-kB signaling in various cells including the kidney [39,40]. NF-κB activates COX-2 expression and thromboxane  $A_2$  generation [41], leading to renal ischemia-reperfusion injury (Fig. 1). Inflammation, old age and obesity are additional stimulators of COX-2 expression [42,43]. Canonical NF-*k*B signaling pathway regulates early phase inflammation in I/R injury [44,45]. In a mouse model of I/R injury induced AKI, widespread NF-кВ activation was associated with impaired renal function, tubular apoptosis and neutrophil and macrophage infiltration after I/R injury, all of which were ameliorated by inhibiting expression of NF- $\kappa$ B [45].

In addition to COX-2 expression, SARS-CoV-2 infection inhibits PG-degrading enzyme 15-hydroxyprostaglandin-dehydrogenase (15-PGDH) in Calu-3 cells by about 90% [36]. 15-PGDH is the main enzyme for lipid mediator catabolism [36]. 15-PGDH can accept a wide variety of prostaglandin substrates with a high affinity for PGE<sub>2</sub>, PGF<sub>2</sub>, PGI<sub>2</sub> and 6-keto PGF<sub>1</sub>, and low affinity for PGD<sub>2</sub> and TxB<sub>2</sub> [46,47].

COX-2 expression and 15-PGDH suppression appear to promote a lipid-mediator storm in severe COVID-19. Archambault and colleagues have measured eicosanoids in the BALF in 33 severely ill patients with COVID-19 within 2h of initiation of mechanical ventilation, compared with 25 healthy controls. Severe COVID-19 patients had marked increases in fatty acid levels as well as an accompanying inflammatory lipid storm with predominance of cyclooxygenase (COX) metabolites notably TxB<sub>2</sub>  $>> PGE_2 > PGD_2$  [7<sup>••</sup>]. Plasma TxB<sub>2</sub> levels were also markedly increased in severe COVID-19 patients [28<sup>•</sup>]. COX-2 mediated thromboxane A<sub>2</sub> generation is thought to be a key catalyst for the prothrombotic state in COVID-19 [27"]. The involvement of COX-2 derived lipid mediators in the pathogenesis of

death leading to common histological findings in SARS-CoV-2 infection including acute tubular injury. Persistent ischemiareperfusion injury and apoptosis may lead to tubulointerstitial fibrosis and progression from AKI to CKD. SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; ACE2, angiotensin-converting enzyme 2; AKI, acute kidney injury; CKD, chronic kidney disease; COX, cyclooxygenase; CYP450, cytochrome P450; EpOMEs, epoxyoctadecenoic acids; ER, endoplasmic reticulum; IFN, interferon; IL, interleukin; IRE1, inositol-requiring enzyme 1; IRF, interferon regulatory factor; L-PGDS, lipocalin prostaglandin D2 synthase; MAPK, mitogen-activated protein kinase; MASP, mannan-binding lectin-associated serine protease; NF-κB, nuclear factor-kappa B; PERK, protein kinase R-like ER kinase; sEH, soluble epoxide hydrolase; TPr, thromboxane A<sub>2</sub> receptor; TxAS, thromboxane A<sub>2</sub> synthase; UPR, unfolded protein response.

<b>Table 1.</b> Renal	involvement in coronavirus disease	2019: lipid mediato	rs, pre and inflammatory cyt	okine, main clinical consequen	ices and therapeuti	c options
Lipid mediators	Changes in COVID-19	Receptors	Postulated humoral and cellular mediators	Clinicopathologic outcomes <sup>a</sup>	Antagonists	Effects of antagonism
PGD2	PGD <sub>2</sub> synthose $\uparrow [67^{\bullet}]$ DPr2 $\uparrow [67^{\bullet}]$ Plasma PGD <sub>2</sub> $\uparrow [5T \text{ Reddy}, \text{personal communication, 2021}]BALF PGD2 \uparrow [7^{\bullet}]$	DPr1	pVascular/renal superoxide↓[127] Blood pressure elevation↓[127]	Renoprotective Protection against Age-related hypertension [128] Antifibrotic [129]	Asapiprant Laropiprant	Hypertension [128] Renal inflammation [128]
		DPr2 (CRTH2)	L4, IL-5, IL-13 $\uparrow$ [72 $-74^{\circ}$ , 108] MDSCs $\uparrow$ [130, 131] Th2 $>>$ Th1 Th2 $>>$ Th1 Resolvabilia $\uparrow$ Rend apoptosis and fbrosis $\uparrow$ [78] FNA $\downarrow$ [133]	AKI — CKD [79] Inflammation Collapsing FSGS Acute tubular necrosis Renal fibrosis [79]	Ramatroban Fevipiprant	Th2>>Th1 ↓ [79,134] Renal fibrosis ↓ [79] IFNA ↑ → antiviral/renal protection [133,135]
TxA2	Plasma TxA₂↑[28 <b>°</b> ] BALF TxA₂↑[7 <b>°</b> ] TxA₂→11-dehydro-TxB₂→ DPr2 [107] Rendl thrombosis (I/R injury)	TP,	$\label{eq:constraint} \begin{array}{c} \mbox{Plateler-neutrophil} \\ \mbox{interaction} \uparrow \\ \mbox{NETosis} \uparrow [105] \\ \mbox{Podoplanin-CEC2} \\ \mbox{signaling} \uparrow [136] \\ \mbox{Vasoconstriction} \uparrow \\ \mbox{NO} \downarrow [91] \\ \mbox{NO} \downarrow [91] \\ \mbox{NO} \uparrow [105] \\ \mbox{NCP-1} \uparrow [137, 138] \\ \mbox{MCP-1} \uparrow [137, 138] \\ \end{array}$	I/R injury [139] AKI → CKD Glomerular thrombosis [140] Hypertension [141] GFR J [140] Glomerulosclerosis [141] Cardiac hypertrophy [141]	Ramatroban Seratrodast Térbogrel Ozagrel	Bleeding [142] Thrombo-inflammation $\downarrow$ [105,143,144] Oxidative stress $\downarrow$ [139,145] Pertusion $\uparrow^{\rm b}$ [141] GFR $\uparrow^{\rm b}$ [141] BP $\downarrow^{\rm b}$ [141] BP $\downarrow^{\rm b}$ [141] Renoprotective <sup>b</sup> [141]
PGE2	BALF PGE2 ↑ [7 <sup>••</sup> ]	EPr1	ROS↑[146] Fibronectin↑[146]	I/R Injury Collapsing FSGS	SC-19220	Renal function ↑ [147] Creatinine/BUN ↓ [147] Glomerulosclerosis ↓ [147]
		EPr4	Podocyte integrity ↑ [148] Inflammatory +/- [149] Fibrosis +/- [149]	Renoprotective [150]	ASP7657	Renoprotective [150] Proteinuria ↓ [150] Antii-inflammatory [150] Antifibrotic [150]
Leukotoxin Diols	12,13-DiHOME ↑ [82 <sup>■</sup> ] 9,10-DiHOME ↑ [82 <sup>■</sup> ]	۸	IL-6 † [83] Rend proximal tubular cell toxicity [84] Mitochondrial dysfunction [84]	AKI Acute tubular necrosis	she inhibitors	Renoprotective [151] Anti-inflammatory [150,151] Antifibrotic [151]
LTC <sub>4</sub> , LTD <sub>4</sub> , LTB <sub>4</sub> , LTE <sub>4</sub>	BALF Leukotrienes ↑ [ <b>7</b> <sup>■</sup> ]	CysLTr1	MPO † [152] Kidney hemorrhages † [152] Glomerular degeneration † [152]	AKI [124] I/R injury [125]	Montelukast	Oxidative damage ↓ [152] Renoprotective [125]
<sup>a</sup> Proposed clinical or <sub>f</sub> <sup>b</sup> Administration of a th	bathologic outcomes based on the changes in uromboxane sunthase inhibitor	n cellular or humoral mec	diators and the disease setting.			

#### Novel therapeutic approaches in nephrology and hypertension

40 www.co-nephrolhypertens.com

Volume 31 • Number 1 • January 2022

TxA<sub>2</sub>, thromboxane A<sub>2</sub>; TxB<sub>2</sub>, thromboxane B<sub>2</sub>; VCAM-1, vascular cell adhesion molecule 1.

acute respiratory syndrome coronavirus 2; cysLTr, cysteine leukotriene receptor; DiHOMEs, dihydroxyoctadecenoic acids; DPr, prostaglandin D2 receptor; EPr, prostaglandin E2 receptor; GFR, glomerular filtration rate; I/ R, ischemia-reperfusion, ICAM-1, intercellular adhesion molecule 1; IL, interleukin; MCP-1, monocyté chemoattractant protein 1; MDŠC, myeloid derived suppressor čells; MPÕ, myeloperoxidase; NET, neutrophil extracellular trap; PG, prostaglandin; ROS, reactive oxygen species; sEH, soluble epoxide hydrolase; Th1 and -2, Thelper cell types 1 and 2; TNF-a, tumor necrosis factor alpha; Tpr, thromboxane prostanoid receptor;

AKI, acute kidney injury; BAIF, bronchaalveolar lavage fluid; BP, blood pressure; BUN, blood urea nitrogen, LT, leukotriene; CKD, chronic kidney disease; COVID-19, coronavirus disease 2019; SARS-CoV2, severe

COVID-19 associated AKI remains to be investigated.

#### Thromboxane A<sub>2</sub>, a potential promoter of thrombotic events and renal ischemiareperfusion injury

In addition to pneumonia, SARS-CoV-2 causes a vascular disease leading to thrombotic microangiopathy, pulmonary thrombosis, pedal acro-ischemia ('COVID-toes'), arterial clots, strokes, cardiomyopathy, coronary and systemic vasculitis, bleeding, deep venous thrombosis, pulmonary embolism; and microvascular thrombosis [3,5,28,48–50]. Necropsies have revealed inflammatory microvascular thrombi containing neutrophils, platelets and NETs in the pulmonary, hepatic, renal and cardiac microvasculature as the hallmark of severe COVID-19 disease leading to ischemia-reperfusion injury and subsequent multiorgan failure, including AKI [3<sup>•</sup>,51,52].

Thromboxane  $A_2$  (TxA<sub>2</sub>), a key mediator of thrombosis, is released by platelets, endothelial cells, macrophages and neutrophils [53]. TxA<sub>2</sub> binds to the thromboxane-prostanoid receptor (TPr) on platelets, thereby stimulating platelet activation and aggregation [53]. In patients with acute SARS-CoV-2 infection, elevated TxA<sub>2</sub> is strongly associated with lung lesions and lower peripheral oxygen saturation [54]. This is consistent with pulmonary microvascular thrombi in lung autopsies of severe COVID-19 patients [5<sup>\*\*</sup>].

In addition to platelet activation, TxA<sub>2</sub> stimulates expression of thrombotic markers including platelet P-selectin [55], endothelial tissue factor [56–58] and CD40L release [59]. In particular, P-selectin is positively associated with creatinine and BUN, and neutrophil infiltration and extracellular trap release during I/R-induced AKI in mice [60]. Urinary excretion of 11-dehydro-thromboxane B<sub>2</sub>, a stable metabolite of TxA<sub>2</sub> is markedly increased in recently hospitalized patients with COVID-19 and is predictive of plasma D-dimer levels, need for mechanical ventilation and mortality [61\*\*]. Additionally, there was a significant positive correlation between urinary 11-dehydro-thromboxane B<sub>2</sub>, and urinary liver-type fatty acid binding protein, a marker of renal ischemia and injury [61<sup>••</sup>]. This suggests that increased generation of thromboxane in COVID-19 is associated with microvascular thrombosis and renal ischemia. Therefore, TxA<sub>2</sub>/TPr signaling presents a therapeutic target in COVID-19-associated AKI. Moreover, protracted microvascular thrombosis, evidenced by asymptomatic elevation of D-dimers in 25% patients up to 4 months post-SARS-CoV-2 infection [62] raises the possibility of sustained thromboxane storm and persistent ischemia to the kidneys leading to progression from AKI to CKD (Fig. 1). Further studies are needed to analyze lipid mediators in long-haul COVID-19.

#### Prostaglandin D<sub>2</sub> signaling as a potential mediator of coronavirus disease 2019associated acute kidney injury and chronic kidney disease

Prostaglandin  $D_2$  is upregulated during states of ischemia [63,64], and plasma levels of PGD<sub>2</sub> are markedly elevated in severe COVID-19 [7<sup>••</sup>,36].  $PGD_2$  is the most abundant prostanoid in the mammalian brain [65] and exerts its main functions through two receptors, DPr1 and DPr2 (aka CRTh2) [66]. In fact, lipocalin PGD<sub>2</sub> synthase and DPr2 expression were elevated in the renal tubules of COVID-19 autopsies [67\*\*]. SARS-CoV-2 infection of Calu-3 lung cells increased PGD<sub>2</sub> secretion more than two-fold [36]. DPr2 expression is upregulated more than 50-fold by TGF- $\beta$  acting in conjunction with IL-4 [68]. In COVID-19, TGF- $\beta$  is markedly increased and is in fact a determinant of the maladaptive host immune response [69]. IL-4 levels are also markedly elevated in severe COVID-19 and post COVID where they correlate with the number of circulating endothelial cells [70]. Increased expression of DPr2 receptors in conjunction with local increase in PGD<sub>2</sub> has the potential to massively amplify PGD<sub>2</sub>/DPr2 signaling.

PGD<sub>2</sub> via DPr2 is known to induce Th2 cell activation and release of type 2 cytokines including IL-4, IL-5 and IL-13, which promote allergic inflammation [71], a characteristic feature of COVID-19 [72<sup>•</sup>]. IL-13 was universally correlated with ARDS, AKI and mortality in COVID-19 [73<sup>•</sup>]. Plasma IL-13 levels are associated with need for mechanical ventilation in COVID-19. IL-13 has been recently reported to stimulate hyaluronan accumulation in mouse lungs [74<sup>•</sup>].

PGD<sub>2</sub>/DPr2 signaling also exerts pro-apoptotic and profibrotic actions on a variety of cells including islet cells, cardiomyocytes and osteoclasts [75– 77]. PGD<sub>2</sub>/DPr2 signaling promotes apoptosis of cultured pig kidney cells [78]. PGD<sub>2</sub>/DPr2 signaling also contributes to tubulointersitial fibrosis following unilateral ureteral obstruction in mice, which was ameliorated either with a DPr2 antagonist or by ablation of IL-4 and IL-13 [79]. Therefore, PGD<sub>2</sub>/ DPr2 signaling merits examination as a mediator of COVID-19-associated acute kidney injury and progression to CKD (Fig. 1).

#### Leukotoxin diols as a potential mediator of coronavirus disease 2019-induced renal inflammation and injury

Leukotoxin diols are a metabolite of linoleic acid (18:2n6) that is an essential source for long-chain

n-6 polyunsaturated fatty acids. CYP450 epoxygenase enzymes convert linoleate to linoleic epoxides (epoxyoctadecenoic acids, EpOMEs) [80,81]. EpOMEs are further metabolized by soluble epoxide hydrolase (sEH) to leukotoxin diols (dihydroxyoctadecenoic acids, or DiHOMEs) [80,81]. Interestingly, COVID-19 patients have increases in leukotoxin diols, 12,13-DiHOME and 9,10-DiHOME when compared with healthy controls [82<sup>•</sup>].

Vascular permeability and stimulation of neutrophil chemotaxis are two vascular actions associated with leukotoxin diols [83]. Endothelial cell activation by leukotoxin diols enhanced nuclear translocation of the transcription factor NF-кВ and the inflammatory cytokine IL-6 [83]. Leukotoxin diols also cause mitochondrial dysfunction and extensive cell death at the level of the proximal tubule [84]. Alexander et al. [85] have demonstrated that the morphological and molecular profiles of severe COVID-19 renal injury resemble sepsisinduced renal injury including microvascular dysfunction, inflammation and molecular reprograming but mitochondrial injury is more severe. These actions on the vascular endothelium and renal proximal tubule epithelial cells are consistent with the pathophysiology of AKI (Fig. 1).

#### Leukotrienes as potential mediators of coronavirus disease 2019-induced oxidative stress and acute kidney injury

Leukotrienes, notably LTB<sub>4</sub>, LTE<sub>4</sub> and eoxin E<sub>4</sub> are significantly increased in the BALF of severe COVID-19 patients [7<sup>••</sup>]. Leukotrienes have been implicated in renal disease. In rat model of glomerular nephritis, administration of antirat glomerular basement membrane antibodies led to urinary excretion of LTC<sub>4</sub>, LTE<sub>4</sub> and acetylated LTE<sub>4</sub> concomitantly with increased renal LTC<sub>4</sub> synthase activity [86]. Moreover, in a mouse model of renal–ischemia reperfusion, inhibition of leukotriene biosynthesis attenuated oxidative stress, histopathological markers of tissue damage, cytokine release and damage to renal function [87]. Therefore, leukotrienes likely play a key role in renal ischemia–reperfusion-induced AKI.

#### Other lipid mediators of uncertain significance in causing renal injury in coronavirus disease 2019

Many other lipid mediators have been reported to be increased in the lungs and plasma of COVID-19 patients but their role in causing renal injury in acute and long-haul COVID remains to be defined. These mediators include PGE<sub>2</sub>, specialized proresolving mediators, D-series resolvins and protectin D1 [7<sup>••</sup>,36].

#### Nitric oxide deficiency

Nitric oxide (NO) has been implicated in many physiological functions in the kidney [88], and is known to ameliorate renal injury in ischemic acute renal failure [89]. On the other hand, NO deficiency characterizes transition to chronic kidney disease and adverse kidney events [88,90].

SARS-CoV-2 virus-induced endothelial injury and pyroptosis is associated with inhibition of nitric oxide synthesis. Furthermore, TxA<sub>2</sub> directly inhibits endothelial and inducible nitric oxide synthase [91,92]. Conversely, NO phosphorylates TxA<sub>2</sub> receptor (TPr) and thereby inhibits platelet activation [93] whereas seratrodast or ramatroban as TPr antagonists enhance NO generation [91]. Therefore, TPr antagonism could potentially increase NO production and thereby mitigate AKI in COVID-19.

#### THERAPIES TARGETING THE LIPID MEDIATOR STORM AND ACUTE KIDNEY INJURY IN CORONAVIRUS DISEASE 2019

Therapies targeting the lipid mediator storm have been proposed in COVID-19. In a prospective study, Hong *et al.* treated ordinary, severe and critically ill COVID-19 patients with the COX-2 inhibitor, Celecoxib [94]. In ordinary COVID-19 cases, patients taking Celebrex did not progress to severe disease compared with 15.7% of ordinary cases under routine treatments. Moreover, none of the severe COVID-19 cases with full dose Celebrex treatment progressed to critical illness. Concern has been raised that blocking upstream mediators including COX-2 and COX-1 may result in more challenges than cures because of their broad inhibition of several essential prostanoids including vasodilatory prostacyclin [95].

Dexamethasone is effective in lowering 28-day mortality and need for renal replacement therapy in hospitalized patients [96]. The efficacy of dexamethasone may stem from inhibition of the lipid mediator storm [7<sup>••</sup>]. Dexamethasone destabilizes  $\beta$ globin–Cox-2 reporter mRNAs [97] and upregulates annexin A1 expression [98], thereby inhibiting COX-2 and phospholipase A<sub>2</sub> expression, respectively, and thereby eicosanoid production.

In contrast to dexamethasone, low-dose aspirin failed to limit 28-day mortality, progression to invasive mechanical ventilation or need for renal dialysis in hospitalized COVID-19 patients [99]. This may be because of failure of low-dose aspirin in high thromboxane states, as has been reported in the obese and elderly [100,101]. NF-κB has also served as a therapeutic target in COVID-19, and NF-κB antagonism may ameliorate I/R by downregulating COX-2 expression and TxA<sub>2</sub> production [102].

It has been proposed that blocking the deleterious effects of PGD<sub>2</sub> and TxA<sub>2</sub> with the dual DPr2/TPr antagonist Ramatroban might be beneficial in COVID-19 [7<sup>••</sup>]. Ramatroban as a TPr antagonist is 100 times more potent than aspirin in inhibiting platelet aggregation and P-selectin expression [103,104]. In addition to its antiplatelet action, ramatroban also improves vascular responsiveness; while inhibiting endothelial surface expression of ICAM-1 and VCAM-1; inhibiting MCP-1 expression in response to TNF- $\alpha$  or platelet-activating factor; and inhibiting macrophage infiltration [103]. In a rat model of endotoxic shock, ramatroban prevented hypotension and reduced mortality by 45%; effects attributable to marked reduction in myeloperoxidase levels in lungs, ileum and heart and more than 90% reduction in plasma TNF- $\alpha$ levels [105]. It is notable that infliximab, a TNF inhibitor used in auto-immune conditions, such as Crohn's disease and rheumatoid arthritis is being tested in the WHO-sponsored SOLIDARITY trial [106].

Dual DPr2/TPr antagonism would not only block the activity of both TxA<sub>2</sub> and PGH<sub>2</sub> on platelets and vessels but abrogate the effects of 11-dehydro-thromboxane B<sub>2</sub>, a stable metabolite of TxA<sub>2</sub> and a full agonist of the DPr2 receptor for PGD<sub>2</sub> [107]. Ramatroban is a potent inhibitor of IL-4, IL-5 and IL-13 production induced by  $PGD_2$  [108], and therefore, a promising therapy for the maladaptive immune response in severe COVID-19. Ramatroban (Baynas, Bayer Yakuhin Ltd., Japan) has been safely administered for the treatment of allergic rhinitis in Japan since 2000 [103]. Ramatroban exhibits surmountable binding to TPr [109], and reversible antiplatelet action [103]. This is of advantage in the event of bleeding complications reported in 5.6% of critically ill COVID-19 patients [110].

There is increasing evidence of ongoing lung inflammation in patients with long-haul COVID, which may culminate in lung fibrosis [111,112]. There is no animal model of long-haul COVID to date. However, an animal model of silicosis exhibits massive increases in lung PGD<sub>2</sub> and TxA<sub>2</sub> associated with lung inflammation and fibrosis [113<sup>••</sup>]. In this model, ramatroban reduced macrophage, lymphocyte and neutrophil infiltration of the lungs while inhibiting TNF- $\alpha$ , IL-6, IL-1 $\beta$ , IL-18 and NLRP3 activation, thereby reducing inflammation, fibrosis and cardiopulmonary dysfunction [113<sup>••</sup>]. Therefore, ramatroban is a promising candidate for chemoprophylaxis and treatment of lung and cardiac injury in COVID-19 thereby mitigating secondary kidney injury.

Soluble epoxide hydrolase inhibitors (sEHIs) have two activities that could be beneficial in COVID-19-induced AKI. Inhibition of sEH can lower

linoleic-derived leukotoxin diols and increase arachidonic acid-derived epoxyeicosatrienoic acids (EETs) [81]. The therapeutic benefits for sEHIs have been demonstrated in several animal models of kidney disease including AKI [81,114]. Cisplatin-induced AKI was opposed by sEH inhibition with decreased NF-ĸB activation and TNF- $\alpha$  inflammation [115,116]. The conversion of AKI to chronic kidney disease can also be opposed by sEHIs. Renal fibrosis induced by unilateral ureter obstruction is prevented by sEH inhibitor administration, which was attributed to increased EET levels and decreased inflammation associated with decreased neutrophil influx [117]. Antifibrotic actions for sEHIs can be attributed to suppression of epithelial to mesenchymal transition by increasing E-cadherin, decreasing  $\alpha$ -SMA and preventing the phenotype transition [117,118]. These kidney animal model studies have demonstrated that sEHIs provide renal protection through anti-inflammatory and antifibrotic mechanisms.

Development of sEHIs have been tested in clinical trials and SEHIs did not demonstrate major side effects or toxicity [119,120]. Interestingly, a major finding for the human clinical trial with GSK2256294 found improved endothelial function in obese smokers assessed by forearm blood flow response to the vasodilator bradykinin [121]. These actions on endothelial function would be beneficial in COVID-19-induced AKI. Taken together, sEHIs ability to lower leukotoxin diols and increase EETs have vascular, inflammatory, and epithelial actions that could treat COVID-19-induced AKI and prevent the progression to CKD.

In view of the massive increase in leukotrienes in patients with severe COVID-19, Montelukast, a cysteinyl leukotriene receptor 1 antagonist has recently been shown to reduce clinical deterioration in hospitalized patients with COVID-19 in a small retrospective study [122]. In a rat model of chronic renal failure, montelukast attenuated oxidative damage, myeloperoxidase, LTB<sub>4</sub>, cytokine levels, reduced glutathione, and finally, damage to glomerular structure [123]. Moreover, in a rat model of lipopolysaccharide-induced sepsis, Montelukast reduced renal injury, and levels of inflammatory and oxidative stress markers [124]. Montelukast also reduced AKI induced by rhabdomyolysis or intestinal ischemia in rats [125,126].

#### CONCLUSION

SARS-CoV-2 infection induces a lipid mediator storm with massive increases in pulmonary and systemic thromboxane  $A_2$ , PGD<sub>2</sub> and leukotoxin diols. Thromboxane  $A_2$  and PGD<sub>2</sub> appear to play a

1062-4821 Copyright  $\ensuremath{\mathbb{C}}$  2021 Wolters Kluwer Health, Inc. All rights reserved.

key role in COVID-19-associated thromboinflammation and maladaptive immune response, respectively while leukotoxin diols induce mitochondrial dysfunction and cytotoxicity. The lipid mediators may promote AKI, renal fibrosis and development of CKD. Approaches targeting the lipid mediator storm merit further exploration in order to reduce the heavy burden of kidney disease emerging in the wake of the current pandemic.

#### Acknowledgements

None.

#### **Financial support and sponsorship**

None.

#### **Conflicts of interest**

A.G. and K.C.C. have filed a patent on the use of ramatroban for COVID-19. Other authors declare no conflict of interest. Thromboxane  $A_2$  and prostaglandin  $D_2$  receptor antagonists are under investigation and not approved for the treatment of COVID-19.

#### REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest
  - Nadim MK, Forni LG, Mehta RL, et al. COVID-19-associated acute kidney injury: consensus report of the 25th Acute Disease Quality Initiative (ADQI) Workgroup. Nat Rev Nephrol 2020; 16:747-764.
  - Nugent J, Aklilu A, Yamamoto Y, et al. Assessment of acute kidney injury and longitudinal kidney function after hospital discharge among patients with and without COVID-19. JAMA Netw Open 2021; 4:e211095.
  - 3. Nicolai L, Leunig A, Brambs S, *et al.* Immunothrombotic dysregulation in COVID-19 pneumonia is associated with respiratory failure and coagulo-

pathy. Circulation 2020; 142:1176-1189. This is the first study to characterize thromboinflammation in patients with COVID-19, and highlights the role of neutrophil and platelet activation in SARS-CoV-2 pneumonia and systemic hypercoagulability.

- Mudd PA, Crawford JC, Turner JS, et al. Distinct inflammatory profiles distinguish COVID-19 from influenza with limited contributions from cytokine storm. Sci Adv 2020; 6:eabe3024.
- 5. Ackermann M, Verleden SE, Kuehnel M, *et al.* Pulmonary vascular endothe-
- lialitis, thrombosis, and angiogenesis in Covid-19. New Engl J Med 2020; 383:120-128.

This was the first study to describe splitting angiogenesis as a result of endothelial injury in the lungs of severe COVID-19 patients.

6. Gupta A, Kalantar-Zadeh K, Reddy ST. Ramatroban as a novel immunotherapy for COVID-19. Mol Genet Med 2020; 14:.

This is the first report to postulate a lipid mediator storm underlying the thromboinflammation and maladaptive immune response in COVID-19.

Archambault AS, Zaid Y, Rakotoarivelo V, et al. High levels of eicosanoids
 and docosanoids in the lungs of intubated COVID-19 patients. FASEB J 2021; 35:e21666.

This is the first study highlighting a lipid mediator storm in the lungs of patients with severe COVID-19. This observation identifies new therapeutic targets for the treatment of COVID-19.

- Pfister F, Vonbrunn E, Ries T, et al. Complement activation in kidneys of patients with COVID-19. Front Immunol 2021; 11:594849.
- Hirsch JS, Ng JH, Ross DW, et al. Acute kidney injury in patients hospitalized with COVID-19. Kidney Int 2020; 98:209–218.
- Chan L, Coca SG. Acute kidney injury in the time of COVID-19. Kidney360 2020; 1:588-590.
- Gupta S, Coca SG, Chan L, et al., STOP-COVID Investigators. AKI treated with renal replacement therapy in critically ill patients with COVID-19. J Am Soc Nephrol 2021; 32:161–176.

- Lau WL, Zuckerman JE, Gupta A, Kalantar-Zadeh K. The COVID-kidney controversy: can SARS-CoV-2 cause direct renal infection? Nephron 2021; 145:275–279.
- Mohamed MMB, Lukitsch I, Torres-Ortiz AE, et al. Acute kidney injury associated with coronavirus disease 2019 in urban New Orleans. Kidney360 2020; 1:614–622.
- Akilesh S, Nast CC, Yamashita M, et al. Multicenter clinicopathologic correlation of kidney biopsies performed in COVID-19 patients presenting with acute kidney injury or proteinuria. Am J Kidney Dis 2021; 77:82.e1-93.e1.
- Legrand M, Bell S, Forni L, et al. Pathophysiology of COVID-19-associated acute kidney injury. Nat Rev Nephrol 2021; 17:751-764.
- Su H, Yang M, Wan C, et al. Renal histopathological analysis of 26 postmortem findings of patients with COVID-19 in China. Kidney Int 2020; 98:219-227.
- Wölfel R, Corman VM, Guggemos W, et al. Virological assessment of hospitalized patients with COVID-2019. Nature 2020; 581:465–469.
- Sun J, Zhu A, Li H, et al. Isolation of infectious SARS-CoV-2 from urine of a COVID-19 patient. Emerg Microbes Infect 2020; 9:991–993.
- Puelles VG, Lütgehetmann M, Lindenmeyer MT, et al. Multiorgan and renal tropism of SARS-CoV-2. New Engl J Med 2020; 383:590-592.
- Ng JH, Hirsch JS, Hazzan A, et al. Outcomes among patients hospitalized with COVID-19 and acute kidney injury. Am J Kidney Dis 2021; 77:204-215.
- Bruchfeld A. The COVID-19 pandemic: consequences for nephrology. Nat Rev Nephrol 2021; 17:81–82.
- Gradin A, Andersson H, Luther T, et al. Urinary cytokines correlate with acute kidney injury in critically ill COVID-19 patients. Cytokine 2021; 146:155589.
- Nechemia-Arbely Y, Barkan D, Pizov G, et al. IL-6/IL-6R axis plays a critical role in acute kidney injury. J Am Soc Nephrol 2008; 19:1106–1115.
- Bombeli T, Karsan A, Tait JF, Harlan JM. Apoptotic vascular endothelial cells become procoagulant. Blood 1997; 89:2429–2442.
- Jung F, Krüger-Genge A, Franke RP, et al. COVID-19 and the endothelium. Clin Hemorheol Microcirc 2020; 75:7–11.
- Vitkova V, Zivny J, Janota J. Endothelial cell-derived microvesicles: potential mediators and biomarkers of pathologic processes. Biomark Med 2018; 12:161–175.
- 27. Goshua G, Pine AB, Meizlish ML, et al. Endotheliopathy in COVID-19associated coagulopathy: evidence from a single-centre, cross-sectional study. Lancet Haematol 2020; 7:e575-e582.

This article highlights the role of microvascular thrombosis and endothelialitis in the pathogenesis of COVID-19.

Hottz ED, Azevedo-Quintanilha IG, Palhinha L, et al. Platelet activation and

 platelet-monocyte aggregate formation trigger tissue factor expression in patients with severe COVID-19. Blood 2020; 136:1330-1341.

This study describes platelet and monocyte activation and platelet-monocyte parternship as playing a role in thromboinflammation in COVID-19.

- **29.** Manne BK, Denorme F, Middleton EA, *et al.* Platelet gene expression and function in patients with COVID-19. Blood 2020; 136:1317-1329.
- This study highlights the contribution of platelet activation and platelet-neutrophil,

platelet-monocyte, and platelet-T-cell aggregates in COVID-19 thromboinflammation.

- Gao T, Hu M, Zhang X, et al. Highly pathogenic coronavirus N protein aggravates lung injury by MASP-2-mediated complement over-activation. medRxiv 2020. 2020.03.29.20041962. doi:10.1101/2020.03.29.20041962.
- Cofiell R, Kukreja A, Bedard K, et al. Eculizumab reduces complement activation, inflammation, endothelial damage, thrombosis, and renal injury markers in aHUS. Blood 2015; 125:3253–3262.
- 32. Liu Y, Zhang C, Huang F, et al. Elevated plasma levels of selective cytokines in COVID-19 patients reflect viral load and lung injury. Natl Sci Rev 2020; 7:1003–1011.
- England H, Summersgill HR, Edye ME, et al. Release of interleukin-1α or interleukin-1β depends on mechanism of cell death. J Biol Chem 2014; 289:15942-15950.
- Bustos M, Coffman TM, Saadi S, Platt JL. Modulation of eicosanoid metabolism in endothelial cells in a xenograft model. Role of cyclooxygenase-2. J Clin Invest 1997; 100:1150–1158.
- 35. Chen JS, Alfajaro MM, Wei J, et al. Cyclooxgenase-2 is induced by SARS-CoV-2 infection but does not affect viral entry or replication. bioRxiv 2020. 2020.09.24.312769. doi:10.1101/2020.09.24.312769.
- Ricke-Hoch M, Stelling E, Lasswitz L, et al. Impaired immune response mediated by prostaglandin E2 promotes severe COVID-19 disease. PLOS ONE 2021; 16:e0255335.
- Sharma A, Garcia G Jr, Wang Y, et al. Human iPSC-derived cardiomyocytes are susceptible to SARS-CoV-2 infection. Cell Rep Med 2020; 1:100052.
- Lee JS, Park S, Jeong HW, et al. Immunophenotyping of COVID-19 and influenza highlights the role of type I interferons in development of severe COVID-19. Sci Immunol 2020; 5:eabd1554.
- 39. Hsu AC-Y, Wang G, Reid AT, et al. SARS-CoV-2 Spike protein promotes hyper-inflammatory response that can be ameliorated by Spike-antagonistic peptide and FDA-approved ER stress and MAP kinase inhibitors *in vitro*. bioRxiv 2020. 2020.09.30.317818. doi:10.1101/2020.09.30.317818.
- Hariharan A, Hakeem AR, Radhakrishnan S, *et al.* The role and therapeutic potential of NF-kappa-B pathway in severe COVID-19 patients. Inflammopharmacology 2021; 29:91–100.

Volume 31 • Number 1 • January 2022

- Lim JW, Kim H, Kim KH. Nuclear factor-κB regulates cyclooxygenase-2 expression and cell proliferation in human gastric cancer cells. Lab Invest 2001; 81:349–360.
- Hsieh PS, Jin JS, Chiang CF, et al. COX-2-mediated inflammation in fat is crucial for obesity-linked insulin resistance and fatty liver. Obesity (Silver Spring) 2009; 17:1150–1157.
- Kim JW, Zou Y, Yoon S, et al. Vascular aging: molecular modulation of the prostanoid cascade by calorie restriction. J Gerontol A Biol Sci Med Sci 2004; 59:B876–B885.
- Reid S, Scholey J. Recent approaches to targeting canonical NFκB signalling in the early inflammatory response to renal IRI. J Am Soc Nephrol 2021; 32:2117-2124.
- Markó L, Vigolo E, Hinze C, et al. Tubular epithelial NF-B activity regulates ischemic AKI. J Am Soc Nephrol 2016; 27:2658–2669.
- Slater DM, Zervou S, Thornton S. Prostaglandins and prostanoid receptors in human pregnancy and parturition. J Soc Gynecol Investig 2002; 9:118–124.
- Narumiya Š, FitzGerald GA. Genetic and pharmacological analysis of prostanoid receptor function. J Clin Invest 2001; 108:25–30.
- Goyal P, Choi JJ, Pinheiro LC, *et al.* Clinical characteristics of Covid-19 in New York City. N Engl J Med 2020; 382:2372–2374.
- Guan W-J, Ni Z-Y, Hu Y, et al., China Medical Treatment Expert Group for Covid-19. Clinical characteristics of coronavirus disease 2019 in China. New Engl J Med 2020; 382:1708–1720.
- Song W-C, Fitzgerald GA. COVID-19, microangiopathy, hemostatic activation, and complement. J Clin Invest 2020; 130:3950–3953.
- Rapkiewicz AV, Mai X, Carsons SE, et al. Megakaryocytes and platelet-fibrin thrombi characterize multiorgan thrombosis at autopsy in COVID-19: A case series. EClinicalMedicine 2020; 24:100434.
- Barnes BJ, Adrover JM, Baxter-Stoltzfus A, et al. Targeting potential drivers of COVID-19: neutrophil extracellular traps. J Exp Med 2020; 217:e20200652.
- Rucker D, Dhamoon AS. Physiology, Thromboxane A2. StatPearls. Treasure Island (FL). 2020.
- 54. Al-Hakeim HK, Al-Hamami SA, Almulla AF, Maes M. Increased Serum Thromboxane A2 and Prostacyclin but Lower Complement C3 and C4 Levels in COVID-19: Associations with Chest CT Scan Anomalies and Lowered Peripheral Oxygen Saturation. COVID 2021; 1:489–502.
- 55. Matsui Y, Amano H, Ito Y, et al. Thromboxane A2 receptor signaling facilitates tumor colonization through P-selectin-mediated interaction of tumor cells with platelets and endothelial cells. Cancer Sci 2012; 103:700-707.
- Bode M, Mackman N. Regulation of tissue factor gene expression in monocytes and endothelial cells: thromboxane A2 as a new player. Vasc Pharmacol 2014; 62:57–62.
- Del Turco S, Basta G, Lazzerini G, et al. Involvement of the TP receptor in TNF-alpha-induced endothelial tissue factor expression. Vascul Pharmacol 2014; 62:49–56.
- Eligini S, Violi F, Banfi C, et al. Indobufen inhibits tissue factor in human monocytes through a thromboxane-mediated mechanism. Cardiovasc Res 2006; 69:218–226.
- Santilli F, Davi G, Consoli A, et al. Thromboxane-dependent CD40 ligand release in type 2 diabetes mellitus. J Am Coll Cardiol 2006; 47:391–397.
- Singbartl K, Green SA, Ley K. Blocking P-selectin protects from ischemia/ reperfusion-induced acute renal failure. FASEB J 2000; 14:48–54.
- 61. Tantry US, Bliden KP, Cho A, et al. First Experience Addressing the Prognostic Utility of Novel Urinary Biomarkers in Patients With COVID-19. Open Forum Infect Dis 2021; 8: doi:10.1093/ofid/ofab274.

This study provides the first evidence for early prognostic information on COVID-19 severity, including death, using a simple, noninvasive urine sample.

- Townsend L, Fogarty H, Dyer A, et al. Prolonged elevation of D-dimer levels in convalescent COVID-19 patients is independent of the acute phase response. J Thromb Haemost 2021; 19:1064–1070.
- 63. Tokudome S, Sano M, Shinmura K, et al. Glucocorticoid protects rodent hearts from ischemia/reperfusion injury by activating lipocalin-type prostaglandin D synthase-derived PGD2 biosynthesis. J Clin Invest 2009; 119:1477–1488.
- 64. Taniguchi H, Mohri I, Okabe-Arahori H, et al. Prostaglandin D2 protects neonatal mouse brain from hypoxic ischemic injury. J Neurosci 2007; 27:4303-4312.
- Alam N. NREM sleep: anatomy and physiology. In: Kushida CA, editor. Encyclopedia of sleep. Waltham: Academic Press; 2013. pp. 453–459.
- 66. Pettipher R, Hansel TT, Armer R. Antagonism of the prostaglandin D2 receptors DP1 and CRTH2 as an approach to treat allergic diseases. Nat Rev Drug Discov 2007; 6:313-325.
- 67. Diao B, Wang C, Wang R, *et al.* Human kidney is a target for novel severe ■ acute respiratory syndrome coronavirus 2 infection. Nat Commun 2021;
- 12:2506.

This study describes increased expression of prostaglandin  $D_2$  sythase and DPr2 in the kidneys of COVID-19 patients.

- Nakajima T, Asano K, Shiraishi Y, et al. Expression of PGD2 receptor CRTH2 (DP2) on human lung fibroblasts. J Allergy Clin Immunol 2005; 115:S141.
- 69. Ferreira-Gomes M, Kruglov A, Durek P, *et al.* SARS-CoV-2 in severe COVID-19 induces a TGF-β-dominated chronic immune response that does not target itself. Nat Commun 2021; 12:1961.
- Chioh FW, Fong S-W, Young BE, et al. Convalescent COVID-19 patients are susceptible to endothelial dysfunction due to persistent immune activation. eLife 2021; 10:e64909.

- Domingo C, Palomares O, Sandham DA, *et al.* The prostaglandin D2 receptor 2 pathway in asthma: a key player in airway inflammation. Respir Res 2018; 19:189.
- **72.** Lucas C, Wong P, Klein J, *et al.* Longitudinal analyses reveal immunological
   misfiring in severe COVID-19. Nature 2020; 584:463–469.
- This is the first study to identify a maladaptive type 2 immune response in COVID-19.
  73. Gómez-Escobar LG, Hoffman KL, Choi JJ, et al. Cytokine signatures of end
  organ injury in COVID-19. Sci Rep 2021; 11:12606.
- This is the first study to identify nasopharyngeal IL-13 levels as correlating best with ARDS, AKI and mortality in COVID-19.
- **74.** Donlan AN, Sutherland TE, Marie C, *et al.* IL-13 is a driver of COVID-19 severity. JCI Insight 2021; 6:e150107.
- This is the first study to demonstrate IL-13-mediated hyaluronan synthesis in
- pulmonary disorder and describes the immunopathogenic role of IL-13 in COVID-19. **75.** Abadpour S, Tyrberg B, Schive SW, *et al.* Inhibition of the prostaglandin D2–
- GPR44/DP2 axis improves human islet survival and function. Diabetologia 2020; 63:1355-1367.
- Zuo S, Kong D, Wang C, et al. CRTH2 promotes endoplasmic reticulum stress-induced cardiomyocyte apoptosis through m-calpain. EMBO Mol Med 2018; 10:e8237.
- 77. Yue L, Durand M, Lebeau Jacob MC, et al. Prostaglandin D2 induces apoptosis of human osteoclasts by activating the CRTH2 receptor and the intrinsic apoptosis pathway. Bone 2012; 51:338–346.
- Maesaka JK, Palaia T, Fishbane S, Ragolia L. Contribution of prostaglandin D2 synthase to progression of renal failure and dialysis dementia. Semin Nephrol 2002; 22:407–414.
- Ito H, Yan X, Nagata N, et al. PGD2-CRTH2 pathway promotes tubulointerstitial fibrosis. J Am Soc Nephrol 2012; 23:1797–1809.
- Moghaddam MF, Grant DF, Cheek JM, et al. Bioactivation of leukotoxins to their toxic diols by epoxide hydrolase. Nat Med 1997; 3:562–566.
- Imig JD. Prospective for cytochrome P450 epoxygenase cardiovascular and renal therapeutics. Pharmacol Ther 2018; 192:1–19.
- 82. McReynolds CB, Cortes-Puch I, Ravindran R, et al. Plasma linoleate diols are
- potential biomarkers for severe COVID-19 infections. Front Physiol 2021; 12:663869.

This is the first study describing elevated plasma levels of proinflammatory leukotoxin diols in COVID-19.

- 83. Slim R, Hammock BD, Toborek M, et al. The role of methyl-linoleic acid epoxide and diol metabolites in the amplified toxicity of linoleic acid and polychlorinated biphenyls to vascular endothelial cells. Toxicol Appl Pharmacol 2001; 171:184–193.
- Moran JH, Weise R, Schnellmann RG, et al. Cytotoxicity of linoleic acid diols to renal proximal tubular cells. Toxicol Appl Pharmacol 1997; 146:53–59.
- 85. Alexander MP, Mangalaparthi KK, Madugundu AK, et al. Acute kidney injury in severe COVID-19 has similarities to sepsis-associated kidney injury. Mayo Clin Proc 2021.
- Petric R, Ford-Hutchinson AW. Elevated cysteinyl leukotriene excretion in experimental glomerulonephritis. Kidney Int 1994; 46:1322–1329.
- Hadi NR, Al-amran FG, Hussein AA. Effects of thyroid hormone analogue and a leukotrienes pathway-blocker on renal ischemia/reperfusion injury in mice. BMC Nephrol 2011; 12:70.
- Lee J. Nitric oxide in the kidney: its physiological role and pathophysiological implications. Electrolyte Blood Press 2008; 6:27.
- Chander V, Chopra K. Renal protective effect of molsidomine and L-arginine in ischemia-reperfusion induced injury in rats. J Surg Res 2005; 128:132–139.
- 90. Lei C, Berra L, Rezoagli E, et al. Nitric oxide decreases acute kidney injury and stage 3 chronic kidney disease after cardiac surgery. Am J Respir Crit Care Med 2018; 198:1279–1287.
- 91. Shiokoshi T, Ohsaki Y, Kawabe J, et al. Downregulation of nitric oxide accumulation by cyclooxygenase-2 induction and thromboxane A2 production in interleukin-1β-stimulated rat aortic smooth muscle cells. J Hypertens 2002; 20:455-461.
- 92. Zhao Z, Hu J, Gao X, et al. Hyperglycemia via activation of thromboxane A2 receptor impairs the integrity and function of blood-brain barrier in micro-vascular endothelial cells. Oncotarget 2017; 8:30030–30038.
- 93. Wang GR, Zhu Y, Halushka PV, et al. Mechanism of platelet inhibition by nitric oxide: in vivo phosphorylation of thromboxane receptor by cyclic GMPdependent protein kinase. Proc Natl Acad Sci 1998; 95:4888–4893.
- Hong W, Chen Y, You K, et al. Celebrex adjuvant therapy on coronavirus disease 2019: an experimental study. Front Pharmacol 2020; 11:561674.
- Antman EM, DeMets D, Loscalzo J. Cyclooxygenase inhibition and cardiovascular risk. Circulation 2005; 112:759–770.
- RECOVERY Collaborative Group. Horby P, Lim WS, et al. Dexamethasone in hospitalized patients with covid-19. New Engl J Med 2021; 384:693-704.
- Lasa M, Brook M, Saklatvala J, Clark AR. Dexamethasone destabilizes cyclooxygenase 2 mRNA by inhibiting mitogen-activated protein kinase p38. Mol Cell Biol 2001; 21:771–780.
- Sheikh MH, Solito E. Annexin A1: uncovering the many talents of an old protein. Int J Mol Sci 2018; 19:1045.
- 99. RECOVERY. Aspirin to be investigated as a possible treatment for COVID-19 in the RECOVERY trial. 2020. Available at: https://www.recoverytrial.net/ news/aspirin-to-be-investigated-as-a-possible-treatment-for-covid-19-in-therecovery-trial. [Accessed 8 December 2020]

1062-4821 Copyright © 2021 Wolters Kluwer Health, Inc. All rights reserved.

- 100. Petrucci G, Zaccardi F, Giaretta A, et al. Obesity is associated with impaired responsiveness to once-daily low-dose aspirin and in vivo platelet activation. J Thromb Haemost 2019; 17:885–895.
- **101.** Liu X-F, Cao J, Fan L, *et al.* Prevalence of and risk factors for aspirin resistance in elderly patients with coronary artery disease. J Geriatr Cardiol 2013; 10:21.
- **102.** Kircheis R, Haasbach E, Lueftenegger D, *et al.* NF-κB pathway as a potential target for treatment of critical stage COVID-19 patients. Front Immunol 2020; 11:598444.
- 103. Ishizuka T, Matsui T, Okamoto Y, et al. Ramatroban (BAY u 3405): a novel dual antagonist of TXA2 receptor and CRTh2, a newly identified prostaglandin D2 receptor. Cardiovasc Drug Rev 2004; 22:71–90.
- 104. Kariyazono H, Nakamura K, Arima J, et al. Evaluation of antiplatelet aggregatory effects of aspirin, cilostazol and ramatroban on platelet-rich plasma and whole blood. Blood Coagul Fibrinolysis 2004; 15:157–167.
- 105. Altavilla D, Canale P, Squadrito F, *et al.* Protective effects of BAY U 3405, a thromboxane A2 receptor antagonist, in endotoxin shock. Pharmacol Res 1994; 30:137–151.
- 106. WHO. WHO COVID-19 Solidarity Therapeutics Trial. 2021. Available at: https://www.who.int/emergencies/diseases/novel-coronavirus-2019/global-research-on-novel-coronavirus-2019-ncov/solidarity-clinical-trial-for-covid-19-treatments. [Accessed 17 September 2021]
- 107. Böhm E, Sturm GJ, Weiglhofer I, et al. 11-Dehydro-thromboxane B2, a stable thromboxane metabolite, is a full agonist of chemoattractant receptor-homologous molecule expressed on TH2 cells (CRTH2) in human eosinophils and basophils. J Biol Chem 2004; 279:7663–7670.
- 108. Xue L, Gyles SL, Wettey FR, et al. Prostaglandin D2 causes preferential induction of proinflammatory Th2 cytokine production through an action on chemoattractant receptor-like molecule expressed on Th2 cells. J Immunol 2005; 175:6531–6536.
- 109. Mathiesen JM, Christopoulos A, Ulven T, et al. On the mechanism of interaction of potent surmountable and insurmountable antagonists with the prostaglandin D2 receptor CRTH2. Mol Pharmacol 2006; 69:1441–1453.
- **110.** Al-Samkari H, Karp Leaf RS, Dzik WH, *et al.* COVID-19 and coagulation: bleeding and thrombotic manifestations of SARS-CoV-2 infection. Blood 2020; 136:489–500.
- 111. Peluso MJ, Deitchman AN, Torres L, et al. Long-term SARS-CoV-2-specific immune and inflammatory responses in individuals recovering from COVID-19 with and without postacute symptoms. Cell Rep 2021; 36:109518.
- 112. Talla A, Vasaikar SV, Lemos MP, et al. Longitudinal immune dynamics of mild COVID-19 define signatures of recovery and persistence. bioRxiv 2021. 2021.05.26.442666. doi: 10.1101/2021.05.26.442666.
- 113. Pang J, Qi X, Luo Y, et al. Multiomics study of silicosis reveals the potential

■ therapeutic targets PGD(2) and TXA(2). Theranostics 2021; 11:2381–2394. This is the first study to describe a lipid mediator storm in the lungs of silicosis patients; and the role that lipid mediators play in inducing inflammation and lung fibrosis in a mouse model of silicosis. Furthermore, this is the first study demonstrating that dual receptor antagonism of the TxA<sub>2</sub>/TPr and PGD<sub>2</sub>/DPr2 receptors by ramatroban mitigates inflammation and lung fibrosis in silicosis mice.

- **114.** Imig JD, Jankiewicz WK, Khan AH. Epoxy fatty acids: from salt regulation to kidney and cardiovascular therapeutics. Hypertension 2020; 76:3–15.
- 115. Liu Y, Webb HK, Fukushima H, et al. Attenuation of cisplatin-induced renal injury by inhibition of soluble epoxide hydrolase involves nuclear factor κB signaling. J Pharmacol Exp Ther 2012; 341:725-734.
- 116. Parrish AR, Chen G, Burghardt RC, et al. Attenuation of cisplatin nephrotoxicity by inhibition of soluble epoxide hydrolase. Cell Biol Toxicol 2009; 25:217-225.
- 117. Kim J, Imig JD, Yang J, et al. Inhibition of soluble epoxide hydrolase prevents renal interstitial fibrosis and inflammation. Am J Physiol Renal Physiol 2014; 307:F971-F980.
- 118. Liang Y, Jing Z, Deng H, et al. Soluble epoxide hydrolase inhibition ameliorates proteinuria-induced epithelial-mesenchymal transition by regulating the PI3K-Akt-GSK-3β signaling pathway. Biochem Biophys Res Commun 2015; 463:70-75.
- 119. Chen D, Whitcomb R, MacIntyre E, et al. Pharmacokinetics and pharmacodynamics of AR9281, an inhibitor of soluble epoxide hydrolase, in single- and multiple-dose studies in healthy human subjects. J Clin Pharmacol 2012; 52:319–328.
- 120. Lazaar AL, Yang L, Boardley RL, et al. Pharmacokinetics, pharmacodynamics and adverse event profile of GSK2256294, a novel soluble epoxide hydrolase inhibitor. Br J Clin Pharmacol 2016; 81:971–979.
- 121. Yang L, Cheriyan J, Gutterman DD, et al. Mechanisms of vascular dysfunction in COPD and effects of a novel soluble epoxide hydrolase inhibitor in smokers. Chest 2017; 151:555-563.
- **122.** Khan AR, Misdary C, Yegya-Raman N, *et al.* Montelukast in hospitalized patients diagnosed with COVID-19. Journal of Asthma 2021; 1–7.
- 123. Sener G, Sakarcan A, Sehirli O, et al. Chronic renal failure-induced multipleorgan injury in rats is alleviated by the selective CysLT1 receptor antagonist montelukast. Prostaglandins Other Lipid Mediat 2007; 83:257–267.
- 124. Khodir AE, Ghoneim HA, Rahim MA, Suddek GM. Montelukast reduces sepsis-induced lung and renal injury in rats. Can J Physiol Pharmacol 2014; 92:839–847.
- 125. Helmy MM, El-Gowelli HM. Montelukast abrogates rhabdomyolysis-induced acute renal failure via rectifying detrimental changes in antioxidant profile and systemic cytokines and apoptotic factors production. Eur J Pharmacol 2012; 683:294–300.

- 126. Wu S, Zhu X, Jin Z, et al. The protective role of montelukast against intestinal ischemia-reperfusion injury in rats. Sci Rep 2015; 5:15787.
- 127. Kong D, Wan Q, Li J, et al. DP1 activation reverses age-related hypertension via NEDD4L-mediated T-Bet degradation in T cells. Circulation 2020; 141:655-666.
- 128. Cheng K, Wu TJ, Wu KK, et al. Antagonism of the prostaglandin D2 receptor 1 suppresses nicotinic acid-induced vasodilation in mice and humans. Proc Natl Acad Sci U S A 2006; 103:6682–6687.
- 129. van den Brule S, Wallemme L, Uwambayinema F, et al. The D prostanoid receptor agonist BW245C [(4S)-(3-[(3R,S)-3-cyclohexyl-3-hydroxypropyl]-2,5-dioxo)-4imidazolidineheptanoi c acid] inhibits fibroblast proliferation and bleomycininduced lung fibrosis in mice. J Pharmacol Exp Ther 2010; 335:472-479.
- Blanco-Melo D, Nilsson-Payant BE, Liu W-C, et al. Imbalanced host response to SARS-CoV-2 drives development of COVID-19. Cell 2020; 181:1036.e9-1045.e9.
- 131. Trabanelli S, Chevalier MF, Martinez-Usatorre A, et al. Tumour-derived PGD2 and NKp30-B7H6 engagement drives an immunosuppressive ILC2-MDSC axis. Nat Commun 2017; 8:593.
- 132. Zhang S, Wu X, Yu S. Prostaglandin D2 receptord-type prostanoid receptor 2 mediates eosinophil trafficking into the esophagus. Dis Esophagus 2014; 27:601–606.
- 133. Werder RB, Lynch JP, Simpson JC, et al. PGD2/DP2 receptor activation promotes severe viral bronchiolitis by suppressing IFN-λ production. Sci Transl Med 2018; 10:eaao0052.
- 134. Chen W, Hardman C, Batty P, et al. Fevipiprant inhibits prostaglandin D2 mediated activation of group 2 innate lymphoid cells (ILC2s). Eur Respir J 2018; 52(Suppl 62):A4401.
- 135. Omer D, Pleniceanu O, Gnatek Y, et al. Human kidney spheroids and monolayers provide insights into SARS-CoV-2 renal interactions. J Am Soc Nephrol 2021; 32:2242–2254.
- 136. Badolia R, Inamdar V, Manne BK, et al. G(q) pathway regulates proximal Ctype lectin-like receptor-2 (CLEC-2) signaling in platelets. J Biol Chem 2017; 292:14516-14531.
- 137. Ishizuka T, Sawada S, Sugama K, Kurita A. Thromboxane A2 (TXA2) receptor blockade suppresses monocyte chemoattractant protein-1 (MCP-1) expression by stimulated vascular endothelial cells. Clin Exp Immunol 2000; 120:71–78.
- 138. Ishizuka T, Suzuki K, Kawakami M, et al. Thromboxane A2 receptor blockade suppresses intercellular adhesion molecule-1 expression by stimulated vascular endothelial cells. Eur J Pharmacol 1996; 312:367–377.
- 139. Chueh TH, Cheng YH, Chen KH, Chien CT. Thromboxane A2 synthase and thromboxane receptor deletion reduces ischaemia/reperfusion-evoked inflammation, apoptosis, autophagy and pyroptosis. Thromb Haemost 2020; 120:329-343.
- 140. Okumura M, Imanishi M, Okamura M, et al. Role for thromboxane A2 from glomerular thrombi in nephropathy with type 2 diabetic rats. Life Sci 2003; 72:2695–2705.
- 141. Purkerson ML, Joist JH, Yates J, et al. Inhibition of thromboxane synthesis ameliorates the progressive kidney disease of rats with subtotal renal ablation. Proc Natl Acad Sci 1985; 82:193–197.
- 142. Ishizuka T, Matsui T, Okamoto Y, et al. Ramatroban (BAY u 3405): a novel dual antagonist of TXA2 receptor and CRTh2, a newly identified prostaglandin D2 receptor. Cardiovasc Drug Rev 2006; 22:71–90.
- 143. Canale P, Squadrito F, Altavilla D, et al. Beneficial effects of BAY u3405, a novel thromboxane A2 receptor antagonist, in splanchnic artery occlusion shock. Pharmacology 1994; 49:376–385.
- 144. Squadrito F, loculano M, Altavilla D, et al. Reduction of myocardial leukocyte accumulation and myocardial infarct size following administration of BAY u3405, a thromboxane A2 receptor antagonist, in myocardial ischaemiareperfusion injury. Agents Actions 1993; 39:143–149.
- 145. Xu S, Jiang B, Maitland KA, et al. The thromboxane receptor antagonist S18886 attenuates renal oxidant stress and proteinuria in diabetic apolipoprotein E-deficient mice. Diabetes 2006; 55:110–119.
- 146. Nasrallah R, Hassouneh R, Zimpelmann J, et al. Prostaglandin E2 increases proximal tubule fluid reabsorption, and modulates cultured proximal tubule cell responses via EP1 and EP4 receptors. Lab Invest 2015; 95:1044–1055.
- 147. Chen X, Yin J, Xu Y, et al. Effect of selective inhibition or activation of PGE2 EP1 receptor on glomerulosclerosis. Mol Med Rep 2020; 22:2887–2895.
- 148. Faour WH, Thibodeau JF, Kennedy CR. Mechanical stretch and prostaglandin E2 modulate critical signaling pathways in mouse podocytes. Cell Signal 2010; 22:1222–1230.
- 149. Nasrallah R, Hassouneh R, Hébert RL. PGE2, kidney disease, and cardiovascular risk: beyond hypertension and diabetes. J Am Soc Nephrol 2016; 27:666–676.
- 150. Mizukami K, Yoshida H, Nozawa E, *et al.* Renoprotective effects of the novel prostaglandin EP4 receptor-selective antagonist ASP7657 in 5/6 nephrectomized chronic kidney disease rats. Naunyn Schmiedebergs Arch Pharmacol 2019; 392:451–459.
- 151. Kim J, Imig JD, Yang J, et al. Inhibition of soluble epoxide hydrolase prevents renal interstitial fibrosis and inflammation. Am J Physiol-Renal Physiol 2014; 307:F971-F980.
- 152. Sener G, Kabasakal L, Cetinel S, *et al.* Leukotriene receptor blocker montelukast protects against burn-induced oxidative injury of the skin and remote organs. Burns 2005; 31:587–596.