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Draft genome sequence of the Tremellomycetes yeast *Papiliotrema laurentii* 5307AH, isolated from aircraft

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ABSTRACT *Papiliotrema laurentii* 5307AH was isolated from an aircraft polymer-coated surface. The genome size is 19,510,785 bp with a G + C content of 56%. The genome harbors genes encoding oxygenases, cutinases, lipases, and enzymes for styrene degradation, all of which could play a critical role in survival on xenobiotic surfaces.

KEYWORDS biodegradation, aircraft, *Papiliotrema laurentii*, polymers

Papiliotrema laurentii strain 5307AH was isolated from an environmental consortium collected inside an aircraft (1). *P. laurentii* species have been found inhabiting a variety of habitats such as natural, agricultural, and human-made environments (2). For instance, *P. laurentii* has been found in hydrocarbon-contaminated soils (3), inside aircrafts (1), and on surfaces within the International Space Station (4). The aircraft-derived *P. laurentii* strain 5307AH was maintained in glycerol stocks at -80°C , and overnight cultures were inoculated from single colonies on tryptic soy agar plates. Overnight broth cultures were grown on tryptic soy broth at 27°C with shaking (200 rpm). A 1 mL aliquot of overnight culture was centrifuged, and genomic DNA was extracted from the pellet using the Qiagen PowerMicrobiome kit. The DNA was quantified using the Qubit dsDNA Quantification Assay Kit (Life Technologies) following the manufacturer's instructions (5).

The *P. laurentii* 5307AH genome was sequenced using PacBio technology. Five micrograms of genomic DNA was sheared to >10 kb using Covaris g-Tubes. The sheared DNA was treated with exonuclease to remove single-stranded ends and DNA damage repair mix, followed by end repair and ligation of blunt adapters using SMRTbell Template Prep Kit 1.0 (Pacific Biosciences). The library was purified with AMPure PB beads. PacBio Sequencing primer was then annealed to the SMRTbell template library, and sequencing polymerase was bound using Sequel Binding kit 2.0. The prepared SMRTbell template libraries were then sequenced on a Pacific Biosciences Sequel sequencer using v3 sequencing primer, 1M v2 SMRT cells, and version 2.1 sequencing chemistry with 1×360 and 1×600 sequencing movie run times. The resulting 2,143,392 reads totaling 8.04 Gb were assembled with Falcon version 1.8.8 (6), improved with finisherSC version 2.0 (7), and polished with Arrow version SMRTLink version 5.1.0.26412, yielding 24 scaffolds, $364.38\times$ sequencing read coverage depth, and an N_{50} value of 1.59 Mb with a G + C content of 56%. The assembled genome was annotated using the JGI Annotation pipeline (8). There were 7,537 predicted protein-coding genes with a median length of 422 amino acids, of which 5,474 (72.63%) had Pfam domains, 252 (3.35%) had CAZyme annotations, 1,417 (18.80%) had signal peptides, and 1,497 (19.86%) had transmembrane domains. Mitochondria were assembled separately from the Falcon pre-assembled reads (preads) using an in-house tool (assemblemito.sh), used to filter the preads, and polished with Arrow version SMRTLink version 5.1.0.26412 (9).

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A secondary Falcon assembly was generated using the mitochondria-filtered preads, improved with finisherSC version 2.0 (10), and polished with Arrow version SMRTLink version 5.1.0.26412 (9). Contigs less than 1,000 bp were excluded.

Furthermore, a search of the KEGG Metabolic Pathway database showed that the *P. laurentii* 5307AH genome encodes enzymes from the family of cutinases, lipases, hydroxylases, oxygenases, among others, which could provide an advantage to this fungal strain for survival in xenobiotic environments.

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Sajeet Haridas, Data curation, Formal analysis, Methodology, Software | Chia S. Hung, Conceptualization, Investigation, Methodology | Jasmyn Pangilinan, Formal analysis, Methodology, Software | Anna Lipzen, Formal analysis, Methodology, Software | Hyunsoo Na, Formal analysis, Methodology, Software | Mi Yan, Formal analysis, Methodology, Software | Vivian Ng, Formal analysis, Methodology, Software | Igor V. Grigoriev, Funding acquisition, Software, Supervision | Justin Biffinger, Funding acquisition, Project administration, Writing – review and editing | Daniel Barlow, Funding acquisition, Project administration | Nancy Kelley-Loughnane, Funding acquisition, Project administration, Resources | Wendy J. Crookes-Goodson, Conceptualization, Funding acquisition, Project administration, Resources | Vanessa A. Varaljay, Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Project administration, Supervision, Writing – review and editing.

DATA AVAILABILITY

The raw sequencing reads and draft genome with annotation have been deposited in GenBank under BioProject accession number [PRJNA500119](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA500119). The Sequence Read Archive (SRA) accession number for the raw sequencing reads is [SRS4101967](https://www.ncbi.nlm.nih.gov/sra/SRS4101967). The draft annotated genome assembly accession number is [JAODAN0000000000](https://www.ncbi.nlm.nih.gov/genbank/JAODAN0000000000). Mitochondrial reads can be accessed using accession number [PP706695](https://www.ncbi.nlm.nih.gov/genbank/PP706695).

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