

UCSF

UC San Francisco Previously Published Works

Title

Delayed Goblet Cell Hyperplasia, Acetylcholine Receptor Expression, and Worm Expulsion in SMC-Specific IL-4R α -Deficient Mice

Permalink

<https://escholarship.org/uc/item/8cd3t5pt>

Journal

PLOS Pathogens, 3(1)

ISSN

1553-7366

Authors

Horsnell, William GC

Cutler, Antony J

Hoving, Claire J

et al.

Publication Date

2007

DOI

10.1371/journal.ppat.0030001

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

Delayed Goblet Cell Hyperplasia, Acetylcholine Receptor Expression, and Worm Expulsion in SMC-Specific IL-4R α -Deficient Mice

William G. C. Horsnell¹, Antony J. Cutler¹, Claire J. Hoving¹, Helen Mearns¹, Elmarie Myburgh¹, Berenice Arendse¹, Fred D. Finkelman², Gary K. Owens³, Dave Erle⁴, Frank Brombacher^{1*}

1 Division of Immunology, Institute of Infectious Disease and Molecular Medicine, Faculty of Health Sciences, University of Cape Town, Cape Town, South Africa, **2** Department of Medicine, University of Cincinnati College of Medicine, Cincinnati, Ohio, United States of America, **3** Department of Molecular Physiology and Biological Physics, University of Virginia Health Sciences Center, Charlottesville, Virginia, United States of America, **4** Lung Biology Center, Department of Medicine, University of California San Francisco, San Francisco, California, United States of America

Interleukin 4 receptor α (IL-4R α) is essential for effective clearance of gastrointestinal nematode infections. Smooth muscle cells are considered to play a role in the type 2 immune response-driven expulsion of gastrointestinal nematodes. Previous studies have shown in vitro that signal transducer and activator of transcription 6 signaling in response to parasitic nematode infection significantly increases smooth muscle cell contractility. Inhibition of the IL-4R α pathway inhibits this response. How this response manifests itself in vivo is unknown. In this study, smooth muscle cell IL-4R α -deficient mice (SM-MHC^{Cre}IL-4R α ^{-lox}) were generated and characterized to uncover any role for IL-4/IL-13 in this non-immune cell type in response to *Nippostrongylus brasiliensis* infection. IL-4R α was absent from α -actin-positive smooth muscle cells, while other cell types showed normal IL-4R α expression, thus demonstrating efficient cell-type-specific deletion of the IL-4R α gene. *N. brasiliensis*-infected SM-MHC^{Cre}IL-4R α ^{-lox} mice showed delayed ability to resolve infection with significantly prolonged fecal egg recovery and delayed worm expulsion. The delayed expulsion was related to a delayed intestinal goblet cell hyperplasia, reduced T helper 2 cytokine production in the mesenteric lymph node, and reduced M3 muscarinic receptor expression during infection. Together, these results demonstrate that in vivo IL-4R α -responsive smooth muscle cells are beneficial for *N. brasiliensis* expulsion by coordinating T helper 2 cytokine responses, goblet hyperplasia, and acetylcholine responsiveness, which drive smooth muscle cell contractions.

Citation: Horsnell WGC, Cutler AJ, Hoving CJ, Mearns H, Myburgh E, et al. (2007) Delayed goblet cell hyperplasia, acetylcholine receptor expression, and worm expulsion in SMC-specific IL-4R α -deficient mice. PLoS Pathog 3(1): e1. doi:10.1371/journal.ppat.0030001

Introduction

The interleukin (IL)-13/IL-4 receptor α (IL-4R α)/signal transducer and activator of transcription 6 (STAT-6) signaling pathway is essential in the control of a number of infectious diseases as well as being a key factor in the induction of allergic responses. Signaling through this pathway can either confer protective immunity or mediate tissue damage depending on the antigenic stimuli and the cell-specific response [1]. Previously, our laboratory provided the first description of the effect of a cell-specific deletion of IL-4R α from macrophages and neutrophils on the host's ability to respond to two parasitic infections [2]. It was demonstrated that such a deletion failed to affect resolution of infection by the nematode *Nippostrongylus brasiliensis*, while mice demonstrated an increased susceptibility to infection by the trematode *Schistosoma mansoni*. The work presented here describes the effect of a smooth muscle-specific disruption of IL-4R α expression on the immune response to *N. brasiliensis*.

Murine infection with *N. brasiliensis* induces a strong protective host T helper 2 (T_H2) response for which IL-13 production and signaling through IL-4R α are essential for successful clearance of infection [3,4]. Infective third-stage *N. brasiliensis* larva penetrate the skin and migrate via the blood system, to the lungs. Larva emerge from blood vessels and

enter the airways, from which they are coughed up and swallowed. Upon reaching the intestine, larva develop into egg-producing adult worms that attach to the small intestine epithelium. BALB/c mice clear *N. brasiliensis* infection after approximately 9 d [5].

Although essential for expulsion of *N. brasiliensis* from the intestine, the precise role of IL-4R α in coordinating the immune and physiological response remains unclear [6]. IL-13/IL-4R α /STAT-6 signaling is required for the host to produce an effective goblet cell hyperplasia [7]. Disruption of this response impairs the host ability to resolve an *N. brasiliensis* infection. Additionally, acetylcholine-driven contractions of longitudinal smooth muscle in the intestine are

Editor: Alan Sher, National Institutes of Health, United States of America

Received: March 27, 2006; **Accepted:** November 22, 2006; **Published:** January 12, 2007

Copyright: © 2007 Horsnell et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abbreviations: FACS, fluorescence-activated cell sorting analysis; GMF, geometric mean fluorescence; IL-4R, interleukin 4 receptor; MLN, mesenteric lymph node; PI, postinfection; SM-MHC, smooth muscle myosin heavy chain; STAT-6, signal transducer and activator of transcription 6; T helper 2, T_H2

* To whom correspondence should be addressed. E-mail: fbrombac@uctgsh1.uct.ac.za

Author Summary

Intestinal parasitic worm infections are a major public health concern, with more than 1 billion people infected worldwide. Symptoms associated with these infections are similar to that of other intestinal illnesses, including irritable bowel syndrome. It is likely that the immune response required to expel the worm can also, when activated inappropriately, cause the symptoms of irritable bowel syndrome. This makes understanding parasitic worm infections important in their own right and also as a model for other intestinal illnesses. In previous studies, we demonstrated the crucial importance of interleukin 4 receptor α (IL-4R α) responsiveness for worm expulsion in global IL-4R α -deficient mice. In this study, we specifically addressed the role of IL-4R α responsiveness in a novel smooth muscle cell-specific IL-4R α -deficient mouse model. These mice showed decreased ability to control the worm infection, with delayed expulsion and reduced protective immune responses. These data provide compelling evidence for smooth muscle cell IL-4R α being an important coordinator of both the immune and physiological responses to intestinal worm infections. A proposed model is suggested with IL-4R α responsiveness on smooth muscle cells coordinating T helper 2 cytokine responses, goblet hyperplasia, and acetylcholine-driven smooth muscle contractions for optimal worm expulsion.

also implicated in playing a role in worm expulsion [8]. A number of in vitro studies have shown that intestinal segments and intestinal smooth muscle cells previously exposed to infection by murine nematode models have increased contractile ability. This contractile ability of intestinal segments and/or smooth muscle cells is abrogated in STAT-6 $^{-/-}$ mice. Therefore, the IL-13/IL-4R α /STAT-6 pathway is necessary for elevated smooth muscle cell contractility required to aid worm expulsion [6,9,10]. Additionally, IL-13/IL-4R α /STAT-6-dependent smooth muscle cell signaling can induce responses in surrounding tissues [11], as well as inducing smooth muscle cell release of chemokines, such as thymus- and activation-regulated chemokine [12], in order to coordinate early host responses to pathogens. From these studies, it is apparent that both goblet cell and smooth muscle cell responses to nematode infections are coordinated by the host immune response to infection and that this coordination is essential for optimal disease resolution [13].

To date, no studies have been able to demonstrate in vivo the effect of a cell-specific inhibition of the IL-13/IL-4R α /STAT-6 pathway in smooth muscle cells. Using smooth muscle myosin heavy chain (SM-MHC)^{Cre}IL-4R α ^{-lox} mice, we demonstrate that disrupted IL-4R α expression in smooth muscle cells influences host immunity to an intestinal nematode infection. The absence of smooth muscle IL-4R α delays worm expulsion and goblet cell hyperplasia. Furthermore, induction of T_H2 cytokines is delayed and/or reduced, as is intestinal expression of the M3 acetylcholine receptor, in response to infection with *N. brasiliensis*.

Results

Transgenic mice, expressing Cre recombinase under the control of the smooth muscle cell-specific myosin heavy chain promoter (SM-MHC^{Cre}), were backcrossed to the BALB/c genetic background for nine generations and then intercrossed with IL-4R α ^{-/-} and “floxed” IL-4R α ^{lox/lox} BALB/c mice

to establish smooth muscle cell-specific IL-4R α -deficient BALB/c mice (SM-MHC^{Cre}IL-4R α ^{-lox}), with one deleted and one floxed IL-4R α allele (SM-MHC^{Cre}IL-4R α ^{-lox}) to increase the efficiency of Cre-mediated site-specific recombination. Mutant mouse strains were identified by PCR genotyping (Figure 1A), and cell specificity of disrupted IL-4R α expression was confirmed by fluorescence-activated cell sorting analysis (FACS).

IL-4R α expression was analyzed on α -actin-positive cells derived from aortic cells (Figure 1B). Surface expression of IL-4R α on α -actin-positive cells was equivalent in SM-MHC^{Cre}IL-4R α ^{-lox} (geometric mean fluorescence [GMF]: 11.02) and global IL-4R α ^{-/-} (GMF: 11.2) mice (Figure 1B). Low levels of expression were present in IL-4R α ^{-lox} mice (GMF: 18.37). IL-4R α expression on α -actin-positive smooth muscle cells isolated from small intestine and lung was too low to detect using FACS analysis (unpublished data). However, Cre mRNA was highly expressed in tracheal and intestinal tissue in the SM-MHC^{Cre}IL-4R α ^{-lox} mice. As expected, IL-4R α ^{-lox} mice demonstrated no Cre expression. In agreement with the smooth muscle specificity of the deletion, IL-4R α mRNA expression was substantially depressed in both tracheal and intestinal tissue in SM-MHC^{Cre}IL-4R α ^{-lox} mice compared to IL-4R α ^{-lox} mice (Figure 1C). Importantly, IL-4R α expression was maintained on CD3⁺ T cells, CD19⁺ B cells (Figure 1D), and macrophages (unpublished data) in smooth muscle cell-specific IL-4R α knockout mice and equivalent to levels expressed on transgenic Cre-negative IL-4R α ^{-lox} control littermates. Functional analysis confirmed IL-4R α responsiveness in these cell types (unpublished data). Together, these results provide convincing support for the specificity of smooth muscle cell disruption of IL-4R α in SM-MHC^{Cre}IL-4R α ^{-lox} mice, in agreement with previously published data on the characterization of SM-MHC^{Cre} transgenic mice [14].

To investigate a possible role of IL-4/IL-13-stimulated smooth muscle cells in nematode infections, comparative infection studies with the gastrointestinal nematode *N. brasiliensis* were performed. Worm fecundity in the host was followed by determination of egg production in a time kinetic (Figure 2A). As previously demonstrated [2], control IL-4R α ^{-lox} mice behaved as BALB/c mice with peak fecal egg production found at day 7 and subsequently declining thereafter due to a functional host protective immune response [1,3]. Both the IL-4R α ^{-/-} and SM-MHC^{Cre}IL-4R α ^{-lox} mice demonstrated prolonged egg production, with SM-MHC^{Cre}IL-4R α ^{-lox} mice having eggs present in their feces until day 12 postinfection (PI). As expected, IL-4R α ^{-/-} mice demonstrated a chronic infection with eggs present in feces at day 14 PI. Determining the number of worms in the intestine at various time points following infection with *N. brasiliensis* resulted in comparable worm burdens between IL-4R α ^{-lox}, IL-4R α ^{-/-}, and SM-MHC^{Cre}IL-4R α ^{-lox} mice at days 4 and 7 PI. However, at day 10 PI, IL-4R α ^{-lox} control mice, but not SM-MHC^{Cre}IL-4R α ^{-lox} or IL-4R α ^{-/-} mice, had cleared the worm (Figure 2B), explaining the extended worm fecundity. SM-MHC^{Cre}IL-4R α ^{-lox} mice, but not IL-4R α ^{-/-} mice, showed complete worm expulsion at day 14 PI (Figure 2A). Examination of total serum IgE antibody (Figure 2C) levels showed that SM-MHC^{Cre}IL-4R α ^{-lox} mice responded like the IL-4R α ^{-lox} mice. Together, these results demonstrate increased susceptibility to *N. brasiliensis* in smooth muscle cell-

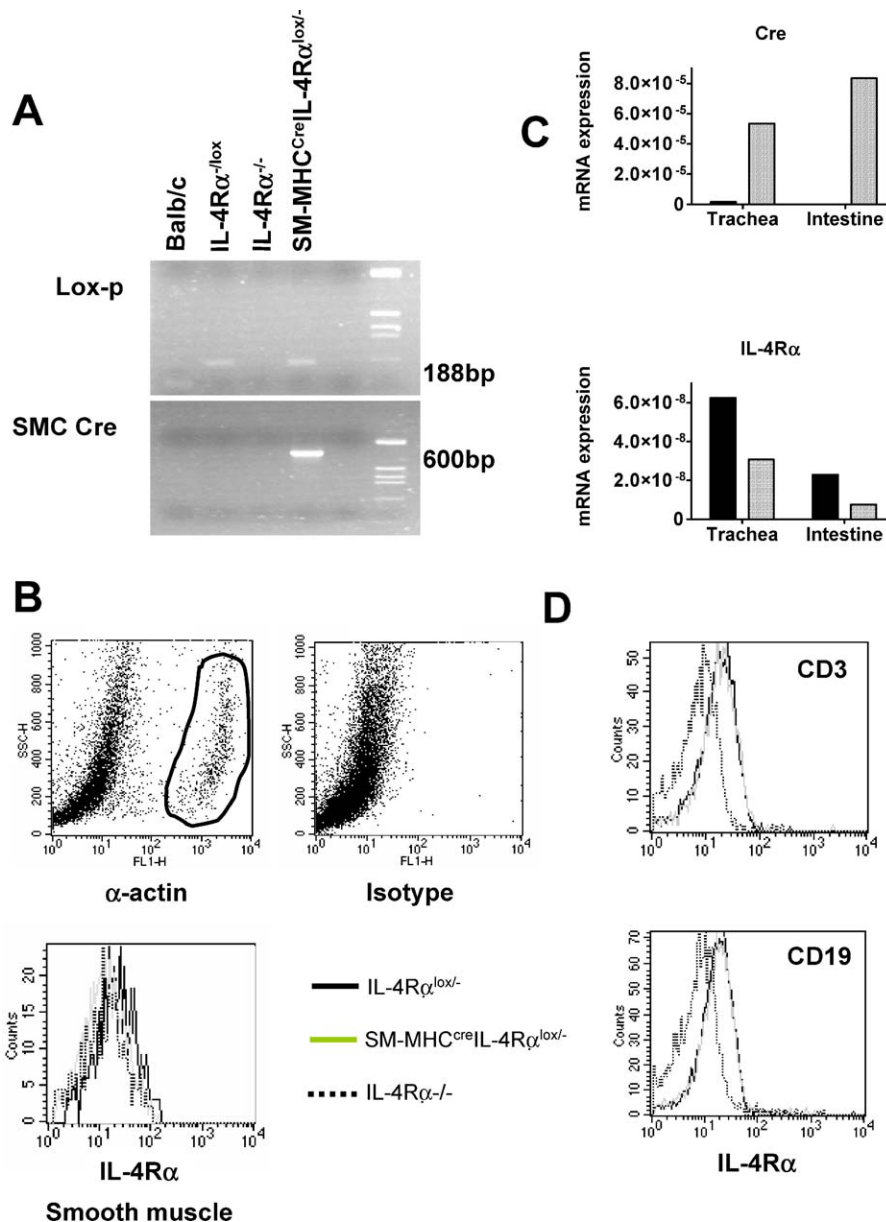


Figure 1. IL-4R α Expression Is Impaired on Smooth Muscle Cells in SM-MHC^{Cre}IL-4R α ^{-lox} Mice

(A) Genomic integrity of the SM-MHC^{Cre}IL-4R α ^{-lox} hemizygous mice was established by PCR.

(B) Smooth muscle cells were identified by intracellular α -actin (i) staining versus isotype control (ii). IL-4R α surface expression was analyzed on gated α -actin-positive cells (iii).

(C) cDNA levels in trachea and small intestine of IL-4R α ^{-lox} (black bars) and SM-MHC^{Cre}IL-4R α ^{-lox} (hatched bars). Data are derived from pooled tissue samples from three mice and are representative of two experiments.

(D) IL-4R α expression on lymphocyte subpopulations of T cells and B cells is unaffected in SM-MHC^{Cre}IL-4R α ^{-lox} mice.

doi:10.1371/journal.ppat.0030001.g001

specific IL-4R α -deficient mice with increased parasite burden and delayed worm expulsion.

T_H2 cytokines drive protective mechanisms following *N. brasiliensis* infection [4]. Therefore, cytokine production by anti-CD3-stimulated CD4⁺ T cells purified from mesenteric lymph nodes (MLNs) was analyzed at days 4, 7, and 10 PI. A reduction ($p < 0.05$) of T_H2 cytokine responses was observed from CD4⁺ T cells of SM-MHC^{Cre}IL-4R α ^{-lox} mice at all time points compared to IL-4R α ^{-lox} control mice, including IL-4, IL-5, IL-9, and IL-13 (Figure 3). Impairment was comparable to mesenteric CD4⁺ T cell from IL-4R α ^{-/-} mice at day 7 PI.

Whereas global IL-4R α ^{-/-} mice shifted to a polarized T_H1 cytokine response, indicated by the production of interferon γ , this was not observed in infected SM-MHC^{Cre}IL-4R α ^{-lox} mice, which had similar interferon γ levels as IL-4R α ^{-lox} control mice. In order to ascertain any compensatory cytokine production in the intestine, we examined IL-13 levels from small intestine tissue at days 4, 7, and 10 PI (Figure 4). At days 4 and 7 PI, IL-13 levels were significantly elevated in IL-4R α ^{-lox} mice compared to IL-4R α ^{-/-} and SM-MHC^{Cre}IL-4R α ^{-lox} mice ($p < 0.05$). By day 10 PI, intestinal IL-13 levels were reduced in IL-4R α ^{-lox} mice but still

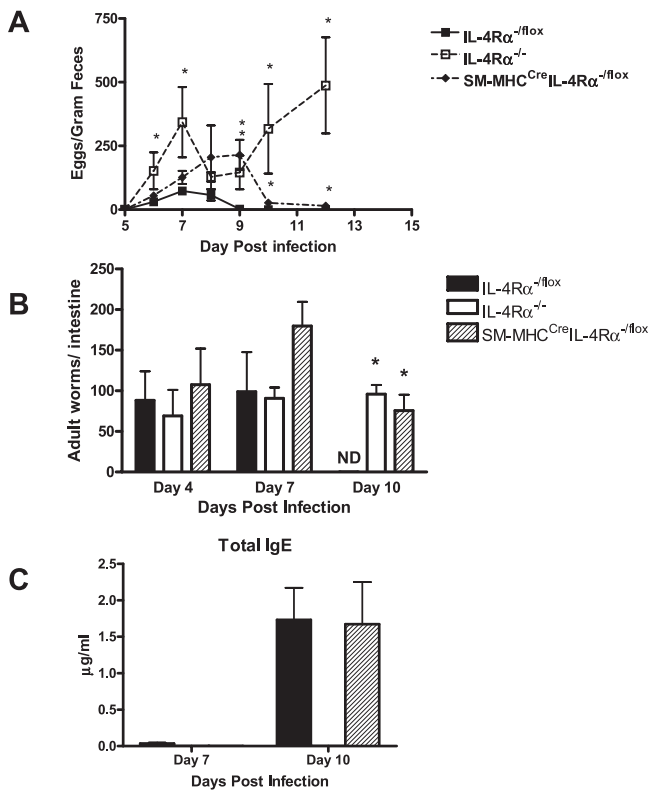


Figure 2. SM-MHC^{Cre}IL-4R α ^{-/-} Mice Have a Delayed Adult Worm Expulsion from the Intestine

(A) *N. brasiliensis* egg production in infected mice was assessed daily from day 5 PI using the modified McMaster technique. (B) Worm burden was established on days 4, 7, and 10 PI by counting worms in intestines removed from infected mice. (C) Serum IgE antibody responses in IL-4R α ^{-/-}lox and SM-MHC^{Cre}IL-4R α ^{-/-}lox mice are equivalent. Serum from infected mice was taken on days 7 and 10 PI and analyzed for antibody production by ELISA as described in Materials and Methods. *Significant differences from IL-4R α ^{-/-}lox mice ($p < 0.05$); data are representative of four separate experiments. doi:10.1371/journal.ppat.0030001.g002

significantly higher than those in IL-4R α ^{-/-} mice ($p < 0.05$). SM-MHC^{Cre}IL-4R α ^{-/-}lox mice, however, also showed significantly higher levels of IL-13 than did IL-4R α ^{-/-} mice at day 10 PI ($p < 0.05$) in accordance with the delayed worm expulsion.

Reduced T_H2 responses in the MLNs had no influence on systemic type 2 antibody responses, as there were similar total serum IgG₁ (unpublished data) and IgE (Figure 2) concentrations in infected SM-MHC^{Cre}IL-4R α ^{-/-}lox and IL-4R α ^{-/-}lox mice. Effective clearance of *N. brasiliensis* is associated with a CD4-driven T_H2 cytokine response with IL-13 playing an essential role [1]. In order to confirm a requirement for CD4⁺ T cells in conferring protection, we carried out a CD4⁺ antibody-driven depletion of these cells. Depletion was confirmed using FACS analysis (unpublished data). As expected [15], IL-4R α ^{-/-}lox-treated mice were unable to clear infection, and CD4⁺ T cells were also essential for clearance in SM-MHC^{Cre}IL-4R α ^{-/-}lox mice, as depletion resulted in increased adult worm burdens in SM-MHC^{Cre}IL-4R α ^{-/-}lox mice (Figure 5).

T_H2 cytokine-driven expulsion of *N. brasiliensis* infections is associated with a concomitant increase in IL-4R α -dependent intestinal goblet cell hyperplasia and mucus production, a process impaired in IL-4R α ^{-/-} mice [3]. Interestingly, impair-

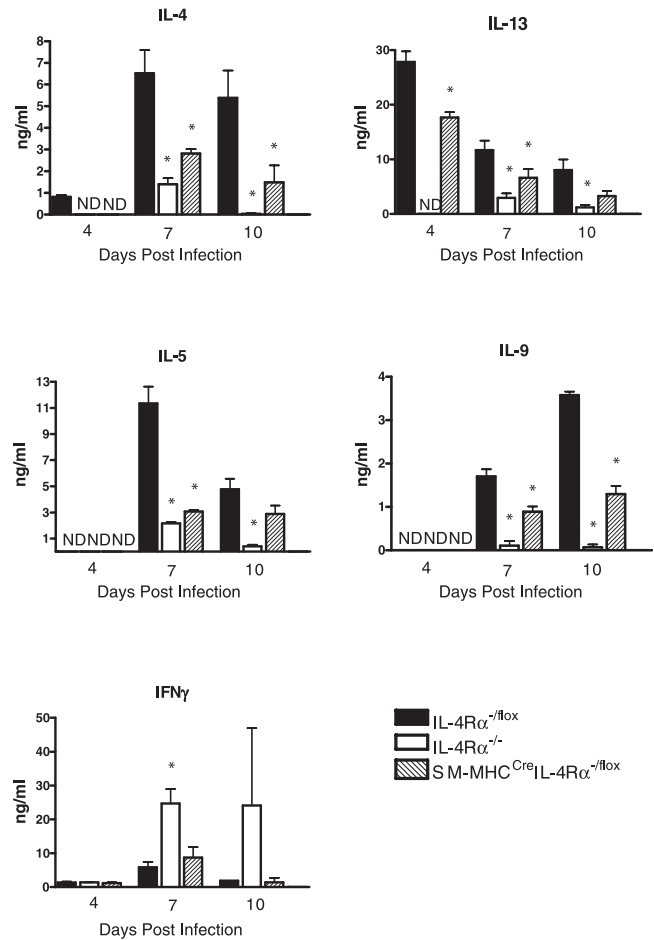


Figure 3. CD4⁺ Lymphocytes Are Essential for *N. brasiliensis* Clearance
MLNs were removed from infected mice on days 4, 7, and 10 PI. CD4⁺ cells were isolated and stimulated with CD3 for 72 h. Supernatants were then analyzed for cytokine production by ELISA as described in Materials and Methods. *Significant difference ($p < 0.05$) from IL-4R α ^{-/-}lox mice. Data are representative of four repeated experiments. doi:10.1371/journal.ppat.0030001.g003

ment of goblet cell hyperplasia was observed in SM-MHC^{Cre}IL-4R α ^{-/-}lox mice. At day 7 PI, where SM-MHC^{Cre}IL-4R α ^{-/-}lox mice showed comparable worm burdens and egg production as Cre-negative IL-4R α ^{-/-}lox control mice (Figure 2), qualitative analysis of intestine histology sections, stained with periodic-acid Schiff reagent to visualize goblet cell mucus production, indicated abrogated mucus production in global IL-4R α ^{-/-} mice and a transient reduction of goblet cell hyperplasia in SM-MHC^{Cre}IL-4R α ^{-/-}lox mice, compared to IL-4R α ^{-/-}lox control mice (Figure 5). The mucus production was delayed in SM-MHC^{Cre}IL-4R α ^{-/-}lox mice as by day 10 PI in goblet cell hyperplasia was comparable to levels observed in IL-4R α ^{-/-}lox control mice at day 7 PI (Figure 6).

In addition to goblet cell hyperplasia, another proposed mechanism of expulsion of *N. brasiliensis* from the host is an increased contractile ability of smooth muscle cells [6]. Induction of such contractility is primarily mediated through an acetylcholine-driven cholinergic response mediated by the M3 muscarinic receptor [6,16,17]. We examined mRNA expression levels of the M3 receptor in the intestine of mice at days 4, 7, and 10 PI (Figure 7). At day 4 PI, no significant difference was noted between groups, although a trend for

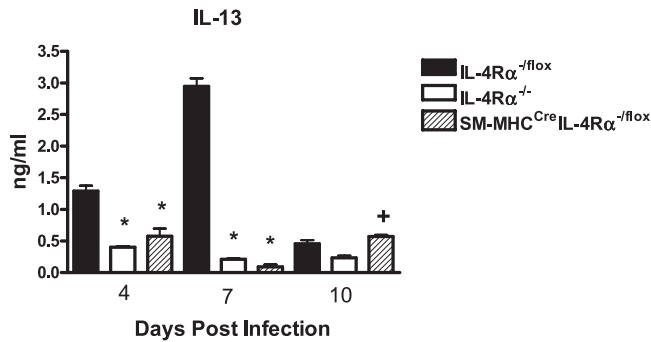


Figure 4. Intestinal IL-13 Levels Are Disrupted in SM-MHC^{Cre}IL-4R α ^{-lox} Mice

Intestinal supernatants were analyzed for IL-13 cytokine production by ELISA as described in Materials and Methods. *Significant difference ($p < 0.05$) from IL-4R α ^{-lox} mice; +significant difference from day 10 PI IL-4R α ^{-/-} mice. Data are representative of three repeated experiments. doi:10.1371/journal.ppat.0030001.g004

higher expression in IL-4R α ^{-lox} mice was noted. We found that at peak infection (day 7 PI), IL-4R α ^{-lox} mice had significantly higher ($p < 0.05$) expression levels of M3 than both IL-4R α ^{-/-} and SM-MHC^{Cre}IL-4R α ^{-lox} mice. By day 10 PI, IL-4R α ^{-/-} mice still showed a significantly lower level of M3 mRNA expression compared to IL-4R α ^{-lox} mice. However, SM-MHC^{Cre}IL-4R α ^{-lox} mice showed increased M3 expression compared to that on day 7 PI. This important result is the first report of IL-4R α expression having an effect on the expression of acetylcholine receptors in vivo.

Together, these results show smooth muscle IL-4R α plays an important role in the regulation of both draining lymph and intestinal cytokine production, goblet cell hyperplasia, and acetylcholine responsiveness. Disruption of these responses in the SM-MHC^{Cre}IL-4R α ^{-lox} mice results in delayed expulsion of the parasites.

Discussion

This work provides the first description of the generation, characterization, and functional analysis of a smooth muscle cell-specific IL-4R α -deficient mouse model. Disruption of IL-4R α expression in smooth muscle cells was applied to a disease model where smooth muscle cells are proposed to

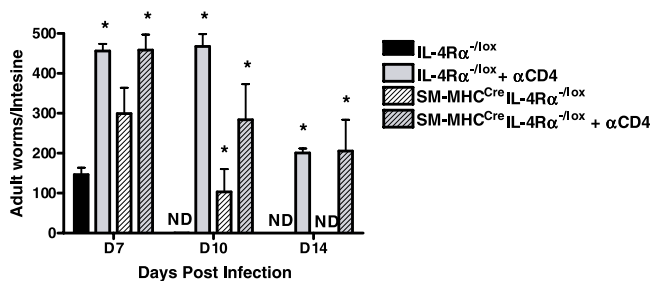


Figure 5. T_{H2} Cytokine Responses Are Impaired in SM-MHC^{Cre}IL-4R α ^{-lox} Mice

Mice were injected IP with depleting anti-CD4 antibody 3 d before infection and subsequently every 4 d. Intestinal worm counts were examined at days 7, 10, and 14 PI in IL-4R α ^{-lox} mice and SM-MHC^{Cre}IL-4R α ^{-lox} mice. *Significant differences compared to IL-4R α ^{-lox} mice ($p < 0.05$); data are representative of a single experiment. doi:10.1371/journal.ppat.0030001.g005

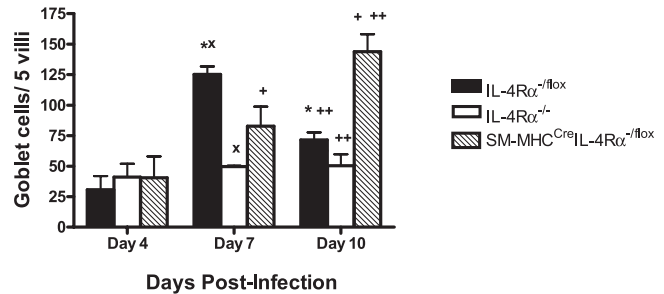


Figure 6. Intestinal Goblet Cell Hyperplasia Is Delayed in SM-MHC^{Cre}IL-4R α ^{-lox} Mice following Infection with *N. brasiliensis*

Mucus-producing goblet cells were visualized using periodic-acid Schiff reagent staining at days 4, 7, and 10 PI. The number of hyperplastic goblet cells per five villi was calculated. Values indicate mean \pm SD, with *, x, +, and ++ indicating significant differences between groups ($p < 0.05$). *Significant decrease in hyperplastic goblet cells in IL-4R α ^{-lox} mice at day 10 PI compared to IL-4R α ^{-lox} mice at day 7 PI. x, Significantly less hyperplastic goblet cells in IL-4R α ^{-/-} mice than in IL-4R α ^{-lox} mice at day 7 PI. +, SM-MHC^{Cre}IL-4R α ^{-lox} mice had significantly more hyperplastic goblet cells at day 10 PI than did SM-MHC^{Cre}IL-4R α ^{-lox} mice at day 7 PI. ++, SM-MHC^{Cre}IL-4R α ^{-lox} mice had significantly more hyperplastic goblet cells than did IL-4R α ^{-lox} mice at day 10 PI. Data are representative of four separate experiments. doi:10.1371/journal.ppat.0030001.g006

play an important role in the resolution of infection, namely, a gastrointestinal nematode infection [6].

Clearance of nematode pathogens from the intestine is considered to require a number of physiological and immunological responses by the host. Increased intestinal contractions [6], increased mucus production [18], and elevated levels of T_{H2}-associated antibodies and cytokines [3] are all mechanisms induced by nematode infection. Wild-type mice infected with *N. brasiliensis* cleared the infection at day 9 PI, while SM-MHC^{Cre}IL-4R α ^{-lox} mice had an impaired ability to clear the nematode until day 12 PI. We demonstrated this impairment to be associated with a delay in goblet cell hyperplasia and the subsequent influx of mucus into the lumen of the host intestine. These physiological disruptions were related to an inability of the host to amplify appropriate cytokine production both locally and by CD4⁺ T cells from the draining MLNs.

A number of authors have demonstrated nematode-induced amplification of intestinal smooth muscle contractions to be dependent on IL-13/IL-4R α /STAT-6 signaling. Isolated strips of smooth muscle from the small intestine of *N. brasiliensis*-infected STAT6^{-/-} mice have a significantly decreased tensile potential in vitro [6]. Depressed contractile ability was also observed in other nematode models in the absence of STAT-6 [6,9]. The significance of these nematode-induced contractions in the resolution of infection remains unclear. Recent work has demonstrated that the serotonin receptor 5-HT_{2a} is a potent inducer of IL-13- and *N. brasiliensis*-dependent intestinal contractions. However, specific inhibition of 5-HT_{2a} failed to affect the ability of the host to resolve infection [19]. We demonstrate a striking reduction in the expression of the acetylcholine M3 receptor in SM-MHC^{Cre}IL-4R α ^{-lox} and IL-4R α ^{-/-} mice following *N. brasiliensis* infection. The M3 expression data we present here

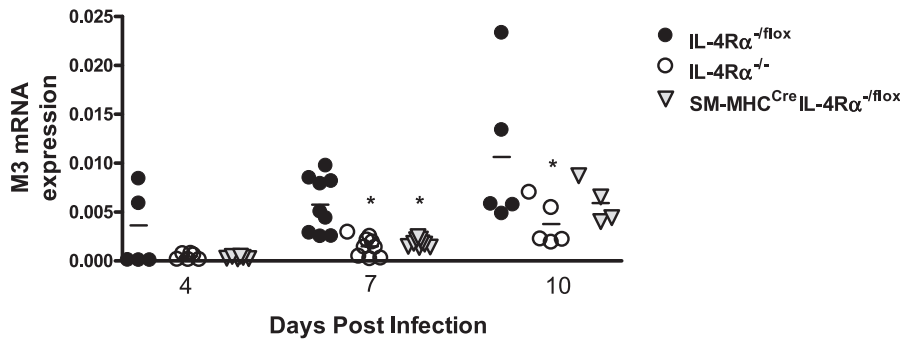


Figure 7. Intestinal Expression of M3 Receptor Is Inhibited by Disrupted Smooth Muscle Cell IL-4R α Expression

mRNA was extracted from intestines of *N. brasiliensis* infected mice at days 4, 7, and 10 PI. Synthesized cDNA was probed with primers to M3. Increases are normalized against α -actin. Data are representative of two to five experiments per time point. $n \geq 4$ mice per group. * $p < 0.05$. doi:10.1371/journal.ppat.0030001.g007

are similar to those of 5-HT $_2a$ in response to *N. brasiliensis* infection. However, the potential role of M3 in mediating expulsion of intestinal parasites is more compelling. M3 $^{-/-}$ mice are incapable of eliciting smooth muscle contractions [16]; this is not the case in 5-HT $_2a$ $^{-/-}$ mice [20]. Previous studies have demonstrated IL-13- and STAT-6-dependent increases in acetylcholine-induced smooth muscle contractions in tissue from *N. brasiliensis*-infected mice [6]. M3 is the principal acetylcholine receptor in smooth muscle and drives 75% of the contractile response in the small intestine [16]. As such, our demonstration of significant inhibition of M3 expression in IL-4R α -deficient mice is compelling in vivo evidence of IL-4R α -muscarinic receptor interactions contributing to proposed muscle hypercontractility-aided nematode expulsion [8].

In addition to contractile responses, host epithelial responses constitute a second major physiological response to the parasite. This response varies according to parasite; in the case of the intraepithelial nematode *Trichuris muris*, expulsion is driven by epithelial cell turnover [21]. The principal aspect of this response to the luminal dwelling *N. brasiliensis* is induction of goblet cell-driven mucus produc-

tion. Goblet cell-derived mucus is essential for clearance of *N. brasiliensis* infection [22,23]. Secreted mucus directly affects viability of the worms through inhibition of parasite motility [24,25] and ability to feed [26]. Pathogen-induced mucus production is strongly influenced by the host immune response. A deficiency in T $_H2$ polarization severely impairs the ability of goblet cells to secrete mucus and expel *N. brasiliensis* [18]. Mucus production is also modulated by the enteric nervous system via innervation of mucosal mast cells [27] and goblet cells [28]. Innervation of epithelial mucus-producing cells is also important for the host mucosal response to *N. brasiliensis* infection [29,30]. Previous studies have established the importance of this epithelial response, the most significant cells for effective expulsion being the mucus-producing goblet cells. This body of work combined with the data we present suggests that smooth muscle cells may represent an intermediate zone of signal transduction between the epithelium and MLNs. Disruption of the ability of the smooth muscle cells to respond to IL-4R α ultimately results in a delayed mucosal response and depressed MLN cytokine production.

Prolonged *N. brasiliensis* infection, due to a deficiency in smooth muscle cell expression of IL-4R α , may therefore be a result of the host's inability to mount an effective mucus response. Delayed mucus responses to *N. brasiliensis* infection are associated with an impaired T $_H2$ response [18]. The delayed expulsion we report here is then explained by the reduced MLN CD4 $^+$ T $_H2$ response (Figure 3), delaying mucus production (Figure 6) through inhibition of smooth muscle responsiveness to neurotransmitters (Figure 7) and cytokines. The depressed T $_H2$ response we suggest to be a result of smooth muscle cells being unable to react effectively to the key smooth muscle contraction amplifying cytokine IL-13 and the neurotransmitter acetylcholine [6] sufficiently to stimulate rapid cytokine production in the MLNs. Parasite clearance would then be more reliant on local effector lymphoid tissue responses [31]. The resulting recovery in response to infection and its eventual clearance in the SM-MHC Cre IL-4R α fllox mouse may then be explained by local responses in the intestine providing a sufficient, albeit delayed and reduced compensatory response which induces the eventual disease-resolving response (Figure 4).

In conclusion, we have demonstrated in vivo a significant role for smooth muscle cell IL-4R α in the optimal resolution of a gastrointestinal nematode infection. Deletion of smooth

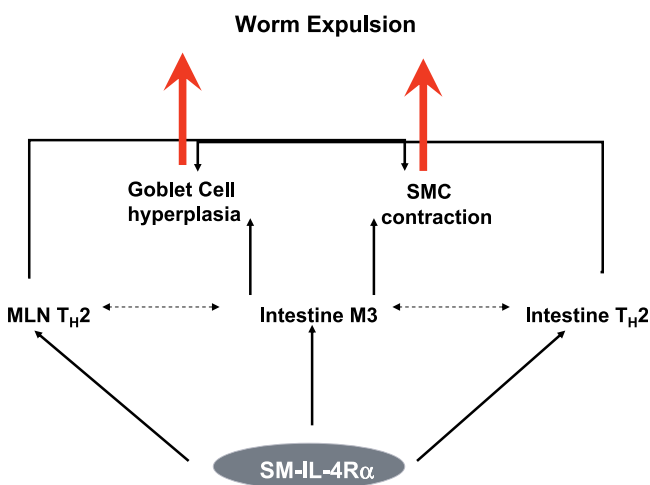


Figure 8. Role of Smooth Muscle IL-4R α in *N. brasiliensis* Infection

Solid arrows represent demonstrated effects of smooth muscle IL-4R α on the host response to *N. brasiliensis* infection. Dotted arrows indicate other potential and/or likely interactions. doi:10.1371/journal.ppat.0030001.g008

muscle IL-4R α significantly disrupts the host ability to resolve infection with *N. brasiliensis*. We demonstrate severe disruption of both known and proposed mediators of expulsion. Depressed M3 receptor expression, delayed goblet cell hyperplasia, disruption of CD4⁺ MLNs, and intestinal cytokine production provide compelling evidence for an important role in the induction of both physiological effector mechanisms and immunological mediators of expulsion. Together, these data are suggestive of smooth muscle IL-4R α being an important inducer of T_H2 cytokine signaling from the lymph node and tissue and goblet cell hyperplasia and having a striking effect on the key smooth muscle contraction-inducing M3 muscarinic receptor (Figure 8).

Materials and Methods

Generation and genotyping of conditional IL-4R α -deficient mice. SM-MHC^{Cre} mice were a kind gift from Gary K. Owens, Charlottesville, Virginia, United States [14,32]. SM-MHC^{Cre} mice were backcrossed to Balb/c for nine generations and then intercrossed with IL-4R α ^{-/-} mice ($n = 30$). These mice were then mated with IL-4R α ^{-lox} mice ($n = 2$) to generate SM-MHC^{Cre}IL-4R α ^{-lox} mice. Transgene negative littermates (IL-4R α ^{-lox}) were used as controls in all experiments. Mice were housed under specific pathogen-free barrier conditions in the University of Cape Town animal facility. All work was approved by the University of Cape Town animal ethics board.

Genotyping. Specific PCR primer pairs were for the IL-4R α , 5'-GTACAGCGCACATTGTTTT-3' and 5'-CTCGGCGCACTGACC CATCT-3'; deletion, 5'-GGCTGCCCTGGAATAACC-3' and 5'-CCTTTGAGAAGTGGCGGGCT-3'; LoxP, 5'-CCCTTCCTGG CCTGAATTT-3' and 5'-GTTTCCTCCTACCGCTGATT-3'; and Cre, 5'-ATGCCCAAGAAGAAGAGGAAGGT-3' and 5'-GAAAT-CAGTGGCTTCGAACGCTAGA-3'. PCR conditions were as follows: 94 °C for 1 min, 94 °C for 30 s, 57 °C for 30 s, and 72 °C for 1 min for 40 cycles on an MJ thermocycler (Biozym Diagnostik, <http://www.biozym.com>).

Analysis of IL-4R α expression by FACS. A single cell suspension of smooth muscle cells was prepared as previously described [33] along with lymph node cells. For the intracellular stain, cells in single cell suspension were incubated with 1% normal rat serum and stained with rat anti-mouse IL-4R α -PE (mIL-4RM-1; BD Biosciences, <http://www.bdbiosciences.com>). Stained cells were then washed, fixed in 2% paraformaldehyde, permeabilized with saponin, preblocked with 2% NRS and Fc block (2.4G2), and stained with anti- α -actin FITC (Abcam, <http://www.abcam.com>) or isotype control IgG2a-FITC (BD Biosciences). For lymphocyte staining, anti-CD3-FITC, anti-CD19-PE, and anti-IL-4R α biotin in combination with SA-APC were used to identify lymphocyte subsets (BD Pharmingen, <http://www.bdbiosciences.com>). Nonviable cells were stained with 7-AAD and excluded from analysis (Sigma, <http://www.sigmaaldrich.com>). Acquisition was performed using FACSCalibur and cells analyzed using Cellquest (Becton Dickinson, <http://www.bd.com>).

Infection studies. Mice were injected subcutaneously with 750 *N. brasiliensis* L3 larva (kindly provided by Klaus Erb, Wurzburg, Germany). Analysis of parasite eggs in feces was carried out using

the modified McMaster technique [34]. Adult worm burdens were determined as previously described [3].

CD4⁺ T-cell depletion. CD4⁺ T cells were depleted from mice by intraperitoneal injection of 0.5 mg of anti-CD4⁺ monoclonal antibody (clone GK1.5) 3 d prior to infection. Mice received booster injections every 3 d to maintain depletion. Effective depletion was confirmed by FACS analysis.

Ex vivo restimulation of lymphocytes. CD4⁺ T cells were purified from pooled MLNs at days 7 and 10 PI. Enrichment was carried with a negative selection. Briefly, cells in single cell suspension were stained with anti-CD8, CD11b, GR-1, B220, and CD16/32. Stained cells were depleted using goat anti-rat IgG-coated magnetic beads (Biomag Beads; Qiagen, <http://www.qiagen.com>). Cell purity was approximately greater than 98%. CD4⁺ T cells were restimulated for 72 h with anti-CD3 (clone 145-2C11; 20 μ g/ml). Supernatants were then collected and stored at -80 °C until analysis.

ELISA analysis. Cytokines in supernatants and serum antibody isotype levels from infected animals were determined as previously described [35].

Histology. Tissue samples were fixed in a neutral buffered formalin solution. Following embedding in paraffin, samples were cut into 5- μ m sections. Sections were stained with hematoxylin and eosin or periodic-acid Schiff reagent. The number of positively stained cells per five villi were counted by light microscopy. All samples were randomized and counted in a blinded fashion.

RT-PCR. RNA was extracted from the intestine of infected mice with the use of Tri-reagent (Sigma), and cDNA was synthesized using the ImProm-II Reverse Transcription System (Promega, <http://www.promega.com>). M3 cDNA was amplified using the following primers: 5'-CGG AAA AGG ATG TCG-3' and 5'-GGC ACT CGC TGA A-3'. Data were normalized using the α -actin housekeeping gene.

Statistics. Values are given as mean \pm SD, and significant differences were determined using the Mann-Whitney *U* test.

Supporting Information

Accession Numbers

The GenBank (<http://www.ncbi.nlm.nih.gov/Genbank>) accession numbers for the genes and gene products discussed in this paper are IL-13 (16163), IL-4R α (16190), STAT-6 (20852), M3 (12671), smooth muscle myosin heavy chain (17880), and α -actin (11475).

Acknowledgments

For technical assistance, we would like to thank Lizette Fick, Erica Smit, Wendy Green, and Reagan Peterson.

Author contributions. WGCH and FB conceived and designed the experiments. WGCH, AJC, CJH, HM, EM, and BA performed the experiments. WGCH, AJC, and CJH analyzed the data. FDF, GKO, and DE contributed reagents/materials/analysis tools. WGCH and AJC wrote the paper.

Funding. This work was supported by grants from the Wellcome Trust, National Research Foundation (South Africa), The Royal Society, and the Medical Research Council (South Africa).

Competing interests. The authors have declared that no competing interests exist.

References

- Brombacher F (2000) The role of interleukin-13 in infectious diseases and allergy. *Bioessays* 22: 646–656.
- Herbert DR, Holscher C, Mohrs M, Arendse B, Schwegmann A, et al. (2004) Alternative macrophage activation is essential for survival during schistosomiasis and downmodulates T helper 1 responses and immunopathology. *Immunity* 20: 623–635.
- Barner M, Mohrs M, Brombacher F, Kopf M (1998) Differences between IL-4R α -deficient and IL-4-deficient mice reveal a role for IL-13 in the regulation of Th2 responses. *Curr Biol* 8: 669–672.
- Urban JF Jr, Noben-Trauth N, Donaldson DD, Madden KB, Morris SC, et al. (1998) IL-13, IL-4R α , and Stat6 are required for the expulsion of the gastrointestinal nematode parasite *Nippostrongylus brasiliensis*. *Immunity* 8: 255–264.
- Ogilvie BM (1971) *Nippostrongylus brasiliensis* in mice: An explanation for the failure to induce worm expulsion from passively immunized animals. *Int J Parasitol* 1: 161–167.
- Zhao A, McDermott J, Urban JF Jr, Gause W, Madden KB, et al. (2003) Dependence of IL-4, IL-13, and nematode-induced alterations in murine

- small intestinal smooth muscle contractility on Stat6 and enteric nerves. *J Immunol* 171: 948–954.
- Khan WI, Motomura Y, Blennerhassett PA, Kanbayashi H, Varghese AK, et al. (2005) Disruption of CD40-CD40 ligand pathway inhibits the development of intestinal muscle hypercontractility and protective immunity in nematode infection. *Am J Physiol Gastrointest Liver Physiol* 288: G15–G22.
- Khan WI, Collins SM (2006) Gut motor function: Immunological control in enteric infection and inflammation. *Clin Exp Immunol* 143: 389–397.
- Akiho H, Lovato P, Deng Y, Ceponis PJ, Blennerhassett P, et al. (2005) Interleukin-4 and -13-induced hypercontractility of human intestinal muscle cells: Implication for motility changes in Crohn's disease. *Am J Physiol Gastrointest Liver Physiol* 288: G609–G615.
- Akiho H, Blennerhassett P, Deng Y, Collins SM (2002) Role of IL-4, IL-13, and STAT6 in inflammation-induced hypercontractility of murine smooth muscle cells. *Am J Physiol Gastrointest Liver Physiol* 282: G226–G232.
- Faffe DS, Flynt L, Bourgeois K, Panettieri RA Jr, Shore SA (2005) IL-13 and IL-4 induce VEGF release from airway smooth muscle cells: Role of VEGF genotype. *Am J Respir Cell Mol Biol*: 213–218.
- Faffe DS, Whitehead T, Moore PE, Baraldo S, Flynt L, et al. (2003) IL-13 and

- IL-4 promote TARC release in human airway smooth muscle cells: Role of IL-4 receptor genotype. *Am J Physiol Lung Cell Mol Physiol* 285: L907–L914.
13. Khan WI, Collins SM (2004) Immune-mediated alteration in gut physiology and its role in host defence in nematode infection. *Parasite Immunol* 26: 319–326.
 14. Regan CP, Manabe I, Owens GK (2000) Development of a smooth muscle-targeted cre recombinase mouse reveals novel insights regarding smooth muscle myosin heavy chain promoter regulation. *Circ Res* 87: 363–369.
 15. Mohrs M, Shinkai K, Mohrs K, Locksley RM (2001) Analysis of type 2 immunity in vivo with a bicistronic IL-4 reporter. *Immunity* 15: 303–311.
 16. Matsui M, Motomura D, Fujikawa T, Jiang J, Takahashi S, et al. (2002) Mice lacking M2 and M3 muscarinic acetylcholine receptors are devoid of cholinergic smooth muscle contractions but still viable. *J Neurosci* 22: 10627–10632.
 17. Stengel PW, Yamada M, Wess J, Cohen ML (2002) M(3)-receptor knockout mice: muscarinic receptor function in atria, stomach fundus, urinary bladder, and trachea. *Am J Physiol Regul Integr Comp Physiol* 282: R1443–R1449.
 18. McKenzie GJ, Bancroft A, Grecis RK, McKenzie AN (1998) A distinct role for interleukin-13 in Th2-cell-mediated immune responses. *Curr Biol* 8: 339–342.
 19. Zhao A, Urban JF Jr, Morimoto M, Elfrey JE, Madden KB, et al. (2006) Contribution of 5-HT(2A) receptor in nematode infection-induced murine intestinal smooth muscle hypercontractility. *Gastroenterology* 131: 568–578.
 20. Fiorica-Howells E, Hen R, Gingrich J, Li Z, Gershon MD (2002) 5-HT(2A) receptors: Location and functional analysis in intestines of wild-type and 5-HT(2A) knockout mice. *Am J Physiol Gastrointest Liver Physiol* 282: G877–G893.
 21. Cliffe LJ, Humphreys NE, Lane TE, Potten CS, Booth C, et al. (2005) Accelerated intestinal epithelial cell turnover: A new mechanism of parasite expulsion. *Science* 308: 1463–1465.
 22. Khan WI, Abe T, Ishikawa N, Nawa Y, Yoshimura K (1995) Reduced amount of intestinal mucus by treatment with anti-CD4 antibody interferes with the spontaneous cure of *Nippostrongylus brasiliensis*-infection in mice. *Parasite Immunol* 17: 485–491.
 23. Miller HR, Huntley JF, Wallace GR (1981) Immune exclusion and mucus trapping during the rapid expulsion of *Nippostrongylus brasiliensis* from primed rats. *Immunology* 44: 419–429.
 24. Lee DL, Biggs WD (1990) Two- and three-dimensional locomotion of the nematode *Nippostrongylus brasiliensis*. *Parasitology* 101 (Part 2): 301–308.
 25. Artis D, Wang ML, Keilbaugh SA, He W, Brenes M, et al. (2004) RELM β /FIZZ2 is a goblet cell-specific immune-effector molecule in the gastrointestinal tract. *Proc Natl Acad Sci U S A* 101: 13596–13600.
 26. Rothwell TL (1989) Immune expulsion of parasitic nematodes from the alimentary tract. *Int J Parasitol* 19: 139–168.
 27. Stead RH, Tomioka M, Quinonez G, Simon GT, Felten SY, et al. (1987) Intestinal mucosal mast cells in normal and nematode-infected rat intestines are in intimate contact with peptidergic nerves. *Proc Natl Acad Sci U S A* 84: 2975–2979.
 28. Neutra MR, Phillips TL, Phillips TE (1984) Regulation of intestinal goblet cells in situ, in mucosal explants and in the isolated epithelium. *Ciba Found Symp* 109: 20–39.
 29. Stead RH, Kosecka-Janiszewska U, Oestreicher AB, Dixon MF, Bienenstock J (1991) Remodeling of B-50 (GAP-43)- and NSE-immunoreactive mucosal nerves in the intestines of rats infected with *Nippostrongylus brasiliensis*. *J Neurosci* 11: 3809–3821.
 30. Masson SD, McKay DM, Stead RH, Agro A, Stanis A, et al. (1996) *Nippostrongylus brasiliensis* infection evokes neuronal abnormalities and alterations in neurally regulated electrolyte transport in rat jejunum. *Parasitology* 113 (Part 2): 173–182.
 31. van Panhuys N, Perret R, Prout M, Ronchese F, Le Gros G (2005) Effector lymphoid tissue and its crucial role in protective immunity. *Trends Immunol* 26: 242–247.
 32. Madsen CS, Regan CP, Hungerford JE, White SL, Manabe I, et al. (1998) Smooth muscle-specific expression of the smooth muscle myosin heavy chain gene in transgenic mice requires 5'-flanking and first intronic DNA sequence. *Circ Res* 82: 908–917.
 33. Ray JL, Leach R, Herbert JM, Benson M (2001) Isolation of vascular smooth muscle cells from a single murine aorta. *Methods Cell Sci* 23: 185–188.
 34. Dunn A, Keymer A (1986) Factors affecting the reliability of the McMaster technique. *J Helminthol* 60: 260–262.
 35. Mohrs M, Ledermann B, Kohler G, Dorfmueller A, Gessner A, et al. (1999) Differences between IL-4- and IL-4 receptor alpha-deficient mice in chronic leishmaniasis reveal a protective role for IL-13 receptor signaling. *J Immunol* 162: 7302–7308.