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Clinical features of cats with aqueous tear deficiency: a retrospective case series of 10 patients (17 eyes)

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Abstract

Objectives The aim of this study was to describe the clinical findings, diagnostic test results and response to therapy of cats with Schirmer tear test 1 (STT-1) values below the reference interval.

Methods The medical records of three institutions were searched for cats with ocular surface disease and STT-1 values <9 mm/min, confirmed at two or more separate visits.

Results Ten cats (17 eyes) were included. The mean ± SD (range) age and STT-1 values in affected eye(s) were 6.1 ± 5.7 (0.2–16) years and 2.4 ± 3.1 (0–8) mm/min, respectively. Concurrent ocular surface disease was bilateral in 5/10 cats. Clinical signs included conjunctivitis (14/17 eyes), corneal ulceration (6/17 eyes), non-ulcerative keratitis (4/17 eyes), symblepharon (4/17 eyes), eosinophilic keratitis (3/17 eyes), corneal sequestrum (3/17 eyes), corneal fibrosis (2/17 eyes) and meibomitis (2/17 eyes). Management included topically applied lacrimomimetics, antiviral drugs, corticosteroids or immunomodulatory drugs; orally administered famciclovir; or surgical procedures, in various combinations. Response to therapy (defined as an increase in STT-1 value of ≥5 mm/min) was transient (seen at a single reassessment) in 65% of eyes and sustained (seen at ≥2 consecutive reassessments) in 18% of eyes.

Conclusions and relevance Clinical features seen in cats with low STT-1 values are described, although the association between aqueous deficiency and the reported ocular changes is unknown at this time. We encourage clinicians to assess the tear film in cats with ocular surface disease, and initiate therapy with lacrimomimetics if STT-1 values are repeatedly below normal. Such information will further define aqueous tear deficiency in cats, providing a better understanding of disease prevalence, pathogenesis and treatment.

Keywords: Dry eye; keratoconjunctivitis sicca; tear film; lacrimal; keratitis; conjunctivitis

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Introduction

Ocular surface diseases such as conjunctivitis, corneal ulceration, corneal sequestrum and eosinophilic keratitis are among the most common disorders recognized in cats with ophthalmic complaints.1 Owing to the intimate relationship between the ocular surface and the tear film, it is possible that tear-film deficiency is a serious comorbidity in many of these clinical presentations, as is the case in dogs2 and humans.3 In fact, a qualitative tear deficiency – or alteration in the mucin and/or lipid component of the tears – is often recognized in cats infected with feline herpesvirus-1,4 and those with spontaneous ulcerative keratitis,5 corneal sequestrum6 or conjunctivitis.7

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In contrast, quantitative tear deficiency – ie, a reduction in the aqueous portion of the tears – is seldom reported in cats. Although the paucity of feline reports may be due to low disease prevalence, aqueous deficiency may be under-recognized in this species because veterinarians are primed to look for a syndrome similar to that seen in dogs with keratoconjunctivitis sicca or ‘dry eye’, or because testing the aqueous tear film was long assumed to be unreliable in cats because it could be artifactually lowered as a result of stress.\(^4\) Normative data for the Schirmer tear test 1 (STT-1) and other diagnostic tests for assessment of the tear film have been established in healthy cats,\(^8\) and a recent case report showed that aqueous tear dysfunction in a cat was associated with chronic keratitis, conjunctivitis and impaired healing of corneal ulcers.\(^9\)

The purpose of the present report, therefore, is to describe the clinical findings of a series of cats in which STT-1 values were below the reference interval (RI).\(^8\) In so doing we aim to encourage veterinarians to be more alert to the potential role of tear deficiency and the value of tear testing in cats with various ocular surface diseases. This, in turn, will enhance understanding of tear deficiency in cats and power prospective, case-controlled studies designed to better define the clinical appearance, establish the pathogenesis and assess therapeutic protocols for dry-eye disease in this species.

**Material and methods**

Medical records of Iowa State University’s Lloyd Veterinary Medical Center, the University of California–Davis Veterinary Medical Teaching Hospital and Triangle Animal Eye Clinic in Tokyo, Japan, were searched from 2006 to 2018 for cats with unilateral or bilateral ocular surface disease and STT-1 values <9 mm/min\(^8\) at presentation and on at least one subsequent examination. Signalment, ocular abnormalities, diagnostic test results and management strategies were retrieved from the medical records. Response to therapy was defined as an increase in STT-1 value of ≥5 mm/min,\(^10,11\) and was further defined as ‘transient’ or ‘sustained’ if the improvement was noted at one or two or more consecutive follow-up visits, respectively.

**Results**

**Animals**

Seventeen eyes of 10 cats met all inclusion criteria (Table 1). The mean ± SD (range) age, body weight and STT-1 values in affected eye(s) were 6.1 ± 5.7 (0.2–16) years, 4.3 ± 1.9 (0.6–6.7) kg and 2.4 ± 3.1 (0–8) mm/min, respectively. Ocular surface disease was bilateral in five cats, one of which (case 6) had an STT-1 value <9 mm/min in only one eye. Disease was unilateral in the other five cats, three of which (cases 4, 9 and 10) had STT-1 values <9 mm/min in both eyes.

**Clinical diagnoses**

Concurrent ocular surface pathology included conjunctivitis (14 eyes), corneal ulceration (six eyes), non-ulcerative keratitis (four eyes), sypmlepharon (four eyes), eosinophilic keratitis (three eyes), corneal seques- trum (three eyes), corneal fibrosis (two eyes) and meibomitis (two eyes). Other ocular abnormalities seen included uveitis (three eyes), glaucoma (one eye), iris hyperpigmentation (one eye) and retrobulbar abscess (one eye). The ocular surface was described as ‘lacklustre’ in 10/17 affected eyes, and three cats had mild crusted discharge at the medial canthus (Figure 1).

**Diagnostic testing**

The STT-1 was performed in all eyes; mean ± SD (range) value in affected eyes was 2.4 ± 3.1 (0–8) mm/min\(^8\). The phenol red thread test (PRTT) was performed in cases 1–4 and was within normal limits (≥15 mm/15 s)\(^8\) in all affected eyes but one (case 2), despite these eyes all having low STT-1 values (Table 2). Corneal sensitivity was estimated using a Cochet-Bonnet aesthesiometer in two cats (cases 5 and 10) and was markedly reduced in all three affected eyes (Table 2).\(^12\) Tear film break-up time (TFBUT) was performed in case 5 only and was decreased bilaterally at 5–7 s. Fluorescein and rose Bengal staining, corneal cytology, conjunctival histology and various infectious disease tests were completed in some cats (Tables 2 and 3).

**Therapy**

A wide range of management techniques was used throughout the clinical course of the 10 cats (Table 1). Topically applied lacrimomimetic agents were used in 15 eyes (10 cats) for between 14 and 628 days from 1–12 times daily, depending on the severity of clinical signs. Most lacrimomimetic agents prescribed contained sodium hyaluronate; five eyes (four cats) with severe aqueous deficiency also received autologous serum. Corticosteroids were applied topically in five eyes (four cats) 1–3 times daily for 9–309 days. Other topical immunomodulatory drugs were used in four eyes (three cats) for 35–244 days; these included 0.03% tacrolimus ophthalmic ointment (two eyes), 0.2% ciclosporin ophthalmic ointment (one eye), or 1% (one eye) or 2% (one eye) ciclosporin ophthalmic suspension compounded in corn oil. Both cats that received ciclosporin compounded in corn oil developed blepharitis. Famciclovir was prescribed for oral administration in seven cats for 22–294 days at doses of 35–50 mg/kg three times daily (four cats), 86–90 mg/kg twice daily (two cats) or 100 mg/kg once daily (one cat). Topical antiviral drugs were prescribed in four eyes (four cats) for 109–366 days; these included 0.5% cidofovir ophthalmic solution applied twice daily (three eyes) or 0.1% idoxuridine ophthalmic solution applied 4–6 times daily (one eye).
Table 1  Summary information for 10 cats with reduced aqueous tear film (Schirmer tear test 1 < 9 mm/min) and concurrent ocular surface disease seen at the Triangle Animal Eye Clinic (TAEC), University of California–Davis Veterinary Medical Teaching Hospital (UCD-VMTH) or Iowa State University’s Lloyd Veterinary Medical Center (ISU-LVMC)

<table>
<thead>
<tr>
<th>Cat</th>
<th>Signalment</th>
<th>Clinic</th>
<th>Concurrent ocular disease</th>
<th>All therapies used throughout clinical course</th>
<th>Follow-up (days)</th>
<th>Clinical outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2 mo MI DSH</td>
<td>TAEC</td>
<td>Symblepharon OU Conjunctivitis OU Non-ulcerative keratitis OU</td>
<td>Sodium hyaluronate ophthalmic solution; oral famciclovir</td>
<td>63</td>
<td>Improved corneal transparency OU Improved conjunctivitis OU</td>
</tr>
<tr>
<td>2</td>
<td>5 mo FI DSH</td>
<td>TAEC</td>
<td>Symblepharon OU Conjunctivitis OU Non-ulcerative keratitis OU</td>
<td>Sodium hyaluronate ophthalmic solution, erythromycin 0.5%; oral famciclovir; symblepharon dissection; warm compresses</td>
<td>135</td>
<td>Improved keratoconjunctivitis OU Static symblepharon OU</td>
</tr>
<tr>
<td>3</td>
<td>7 yo FS Abyssinian</td>
<td>TAEC</td>
<td>Corneal sequestrum OS Conjunctivitis OS</td>
<td>Sodium hyaluronate ophthalmic solution; erythromycin 0.5%, ofloxacin 0.3%; oral famciclovir, warm compresses</td>
<td>331</td>
<td>Improved conjunctivitis OS</td>
</tr>
<tr>
<td>4</td>
<td>4 yo MN Russian Blue</td>
<td>TAEC</td>
<td>Eosinophilic keratitis OS Conjunctivitis OS</td>
<td>Sodium hyaluronate ophthalmic solution, prednisolone acetate 1%, erythromycin 0.5%; oral famciclovir, doxycycline</td>
<td>309</td>
<td>Resolved keratitis OS</td>
</tr>
<tr>
<td>5</td>
<td>16 yo MN DSH</td>
<td>UCD-VMTH</td>
<td>Non-healing corneal ulcer OU Anterior uveitis OU Conjunctivitis OD Corneal fibrosis OD Glaucoma OD</td>
<td>Sodium hyaluronate ophthalmic solution, serum, dorzolamide 2%-timolol 0.5%, latanoprost 0.005%; oral famciclovir, prednisolone; corneal debridement, thermokeratoplasty</td>
<td>431</td>
<td>Resolved corneal ulceration OU Residual corneal fibrosis and neovascularization OD</td>
</tr>
<tr>
<td>6</td>
<td>9 yo FS Maine Coon</td>
<td>UCD-VMTH</td>
<td>Corneal sequestrum OU Conjunctivitis OU Meibomitis OU Eosinophilic keratitis OD Ulcerative keratitis OS Corneal fibrosis OS</td>
<td>Sodium hyaluronate ophthalmic solution, cidofovir 0.5%, ciprofloxacin 0.3%, ofloxacin 0.3%; oral famciclovir, L-lysine; corneocconjunctival transposition</td>
<td>994</td>
<td>Resolved ulcerative keratitis OS Recurrence of eosinophilic keratitis OD</td>
</tr>
<tr>
<td>7</td>
<td>18 mo MN DSH</td>
<td>UCD-VMTH</td>
<td>Descemetocele OU Conjunctivitis OU Corneal fibrosis OS</td>
<td>Sodium hyaluronate ophthalmic solution, serum, neomycin-polymyxin B-dexamethasone, idoxuridine 0.1%, ciprofloxacin 0.3%; oral prednisolone, azithromycin; warm compresses</td>
<td>922</td>
<td>Non-ulcerative keratitis OU</td>
</tr>
<tr>
<td>8</td>
<td>14 mo FI Maine Coon</td>
<td>UCD-VMTH</td>
<td>Retrobulbar abscess OD Conjunctivitis OD Ulcerative keratitis OD Anterior uveitis OD</td>
<td>Sodium hyaluronate ophthalmic solution, serum, ceftazolin 5.5%, ciprofloxacin 0.3%; oral amoxicillin-clavulanic acid, prednisolone</td>
<td>241</td>
<td>Resolved retrobulbar disease OD Resolved conjunctivitis OD Active keratitis OD</td>
</tr>
<tr>
<td>9</td>
<td>14 yo FS DSH</td>
<td>ISU-LVMC</td>
<td>Ulcerative keratitis OD Conjunctivitis OD</td>
<td>Sodium hyaluronate ophthalmic solution, serum, dexamethasone 0.1%, cidofovir 0.5%, ciprofloxacin 1%, tobramycin 0.3%, atropine 1%; oral famciclovir; corneal debridement, bandage contact lens</td>
<td>187</td>
<td>Resolved keratoconjunctivitis OD</td>
</tr>
<tr>
<td>10</td>
<td>8 yo FS DLH</td>
<td>ISU-LVMC</td>
<td>Eosinophilic keratitis OD Conjunctivitis OD Iris hyperpigmentation OD</td>
<td>Sodium hyaluronate ophthalmic solution, prednisolone acetate 1%, cidofovir 0.5%; oral L-lysine; corneal debridement</td>
<td>217</td>
<td>Active keratitis OD Static iris hyperpigmentation OD</td>
</tr>
</tbody>
</table>

mo = month old; yo = year old; MI = male intact; MN = male neutered; FI = female intact; FS = female spayed; DSH = domestic shorthair; DLH = domestic longhair; OD = right eye; OS = left eye; OU = both eyes
Surgical procedures included bilateral dissection of symblepharon (case 2), thermokeratoplasty for a non-healing ulcer (case 5), or corneoconjunctival transposition following a lamellar keratectomy for sequestrum removal (case 6). Other therapies included topically or systemically administered antibiotics, systemically administered corticosteroids or warm compresses applied to the eyelids (Table 1).
Response to therapy
Mean ± SD (range) follow-up time was 383 ± 320 (63–994) days. Complete resolution of clinical signs was documented in three cats (cases 4, 5 and 9), whereas partial clinical improvement was noted in five cats (cases 1, 2, 3, 7 and 8). At the last recheck examination, keratoconjunctivitis was still present in the remaining two cats (cases 6 and 10). Figure 2 depicts representative images from follow-up examinations of cases 2, 4, 5 and 8. The STT-1 value increased by ≥5 mm/min in 14/17 affected eyes; this increase was transient (seen at a single reassessment) in 11 (65%) affected eyes and sustained (seen at two or more consecutive reassessments) in three (18%) affected eyes.

Discussion
This report describes the clinical signs, diagnostic test results, treatment, and outcome of ocular surface disease in a series of 10 cats (17 eyes) with aqueous tear deficiency (STT-1 value < 9 mm/min). Although the association between aqueous tear deficiency and ocular surface disease is unknown, based on this case series tear film dysfunction would be expected to have a detrimental effect on the ocular surface health of cats, similar to dogs and humans. Conversely, ocular surface diseases may have a negative effect on tear production and perpetuate dry eye. An STT-1 value < 9 mm/min (especially when confirmed on two separate occasions, as required in this case series)
should be considered abnormal in cats as the occurrence of low STT-1 values in ophthalmically normal cats is rare. Indeed, Paepe et al found that only 2/100 normal cats had STT-1 values <5 mm/min, while Sebbag et al showed that 0/135 normal cats had STT-1 values <7 mm/min. In addition to STT-1, the PRTT was also measured in the present study in four cats (eight eyes) but was normal in all but one affected eye. This supports previous data suggesting that the PRTT is less reliable than the STT-1 in cats. For this reason, and because the STT-1 provides a measure of basal and reflex tearing, as well as a coarse assessment of the neurological function essential for a normal lacrimal functional unit, these authors prefer using the STT-1 than the PRTT to assess the aqueous tear film in cats. Once reduced aqueous tear film is documented, vital stains such as fluorescein, rose bengal or lissamine green can be particularly helpful in highlighting subtle changes in the corneal and/or conjunctival surfaces. Corneal aesthesiometry is an essential diagnostic tool in cats in which neurologic dysfunction is suspected, as exemplified by cases 5 and 10 of this report and a cat from a previous report in which corneal hypoesthesia likely contributed to their aqueous tear deficiency. By contrast, tear osmometry has not proven as reliable or diagnostically useful in dogs or cats as it has in humans with tear-film deficiency.

Impaired tear production or release, or hastened tear loss, especially through evaporation, should all be considered as potential causes of low STT-1 values. In some cats of the present series, impaired tear secretion onto the ocular surface because of obstructed lacrimal ductules seems likely to have contributed to the aqueous tear deficiency noted. This may be a transient consequence of conjunctival swelling as a result of conjunctivitis (as seen in many cases in the present study) or orbital cellulitis, or a more permanent consequence of adhesions from symblepharon or eyelid agenesis (as seen in cases 1 and 2 herein). Where possible, correction of these underlying causes may normalize STT-1 values. This is likely what happened in case 8 in which successful treatment of the retrobulbar abscess was associated with a sustained increase in STT values. A neurogenic cause of aqueous tear deficiency should also be considered in cats. This is a recognized etiology in dogs and humans, and has been reported in one cat. Humans with chronic inflammation from herpes simplex virus-1 infection experience permanent damage to the trigeminal nerve with subsequent corneal hypoesthesia and decreased reflex tearing. Although multiple causes of neurogenic dry eye exist, the high prevalence of herpetic disease in cats warrants consideration and further investigation of a meta-herpetic form of dry-eye disease, as described in humans.

In dry-eye disease, absence of a confirmed etiologic diagnosis renders therapy challenging – as evident in the present case series. In such cases, therapy is limited to hydrating and lubricating the ocular surface.
with lacrimomimetic agents, and controlling concurrent ocular surface inflammation. This likely explains the generally poor responses observed in the cases presented here and highlights the need for future studies to determine the etiopathogenesis of qualitative and quantitative tear-film deficiency in cats. We hope this report will heighten feline practitioners’ awareness of the importance of performing the STT-1 in this species so that future studies might establish a more comprehensive definition of feline dry eye that better elucidates disease prevalence, clinical features, pathogenesis and therapeutic strategies. Such studies should include larger cohorts than reported here, and also a control population in order to better understand the pathogenic role of tear-film deficiency in feline ocular surface disease.

Conclusions
Aqueous tear deficiency can occur concurrently with common ocular surface diseases in cats. Although the causative association between aqueous tear deficiency and these diseases is not known, experience in other species suggests that tear-film deficiencies would be, at the very least, contributory. We encourage clinicians to perform the STT-1 in cats with ocular surface disease, and to initiate therapy with lacrimomimetics if tear-film dysfunction is noted. Complementary tests such as vital stains, TFBUT and corneal aesthesiometry can provide valuable information about the nature and sometimes the cause of the tear-film deficiency, as well as the extent of ocular surface damage. Such data will facilitate a deeper understanding of the prevalence and pathogenesis of diversity of clinical signs associated with, and response to therapy in cats with qualitative and quantitative tear-film deficiencies.

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