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Perlecan Domain V is Upregulated in Human Brain Arteriovenous Malformation (BAVM) and could Mediate VEGF Effect in Lesional Tissue

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Abstract

Brain arteriovenous malformation (BAVM), a rare but important cause of intracranial hemorrhage, has increased angiogenesis and inflammation as key components of the nidus of abnormal vessels and stroma that form the resected surgical specimen. Accordingly, vascular endothelial growth factor (VEGF) and transforming growth factor- β (TGF β) have both been implicated in BAVM pathology for their pro-angiogenic and vascular-regulating roles. The c-terminal fragment of the extracellular matrix component perlecan (domain V, DV) has been shown to be increased and to, via the α 5 β 1 integrin, increase VEGF levels in and around areas of cerebral ischemic injury, another pro-angiogenic condition. We sought to determine whether the concentrations of DV, DV's proangiogenic receptor α 5 β 1 integrin, or DV's anti-angiogenic receptor α 2 β 1 integrin are elevated in human BAVM tissue. DV levels were increased in BAVM compared to control brain tissue from epileptic resection, as was α 5 β 1 integrin. Additionally, α 5 β 1 integrin. VEGF and TGF β levels were also increased in BAVM compared to control tissue. Furthermore, increases in all components were strongly correlated. Excessive generation of pro-angiogenic DV in BAVM suggests that DV may participate in its pathology and may represent a future therapeutic target.

Keywords

Arteriovenous Malformation; AVM

Disclosures and Conflicts of Interest: none declared

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Introduction

A brain arteriovenous malformation (BAVM) is an abnormal tangled mass of arteries and veins, devoid of a true intervening capillary bed, that results in shunting of blood from the arterial to venous circulations. [1,2] These abnormal vessels, along with intervening stromal elements that are predominantly glial and inflammatory cells, is termed the BAVM nidus. It is this nidus that is the target for surgical resection. Traditionally viewed as some kind of embryological failure to develop normally, there is an emerging view that a post-natal event occurs which triggers BAVM formation and evolution into the lesion that is seen on imaging studies and that forms the bulk of the resected surgical specimen. This view has been termed the response-to-injury hypothesis, and presupposes that there is some kind of genetic variation that predisposes one to formation of a BAVM if some sort of local pro-angiogenic stimulation occurs. Such stimulation could be, for example, a minor degree of traumatic injury or localized microvascular thrombosis that otherwise escapes clinical detection. [2] In the case of Hereditary Hemorrhagic Telangiectasia, the genetic variation is known, e.g., defective endoglin or ALK1, both members of the TGF β superfamily pathways. In the case of sporadic BAVM, the existence of underlying genetic variation is not currently understood but is the subject of investigation. [3]

The association of vascular endothelial growth factor (VEGF) with abnormal vasculature of the BAVM nidus [4] led us to hypothesize that the extracellular matrix heparan sulfate proteoglycan perlecan's C-terminal domain V (DV), a protein fragment endogenously cleaved and solubilized [5] that increases VEGF production and secretion following stroke, might also play a role in the pathology of BAVM. [6] As BAVM is an important cause of intracranial hemorrhage, but the specific molecular mechanisms of its etiology, growth, and rupture remain unclear, further characterization of the abnormal signaling molecules within resected BAVM is a potentially important way to identify potential therapeutic targets. [4,7] Here we investigated the relationship between levels of DV, total and endothelial cell expressed levels of its pro-angiogenic $\alpha 5\beta 1$ integrin [6] and anti-angiogenic $\alpha 2\beta 1$ integrin (normally absent from brain microvasculature) receptors, [8] and total levels of VEGF and TGF β in human BAVM.

Materials and methods

Western blot analysis

After IRB approval and informed consent, surgical specimens from either BAVM resection of non-embolized nidal tissue (cases, n=6), or temporal lobe samples obtained from the surgical treatment of epilepsy (controls, n=3) were frozen and homogenized in RIPA buffer (G Biosciences). Western blots were performed with specific antibodies to DV (R&D systems, MAB2364), α 5 (Millipore, AB1928), α 2 (Chemicon International, AB1936), VEGF (Abcam, ab9570), TGF β (Abcam, ab66043), CD31 (Abcam, ab54211), and β -actin (Abcam, ab13822). Blots were quantified using ImageJ software.

Immunohistochemistry

Coimmunohistochemistry was performed on frozen, acetone fixed human brain slices from BAVM or control (epilepsy) tissue with antibodies for α 5 or α 2 and CD31 with the appropriate Alexa Fluor conjugated secondary antibodies. Tissue slices were imaged with a BD Biosystems Carv II spinning disk confocal mounted on a Zeiss Axioplan. Fluorescent images were obtained using the same exposure time to allow for comparison of relative fluorescent intensities between different images and were analyzed using iVision-MacTM Image Acquisition and Analysis Software and Image J software.

Statistical analysis

Western blot values were normalized by dividing by the average control value \times 100. Two sample t-tests were used to investigate differences in average protein expression levels between cases and controls. To account for possible differences in total protein loading and endothelial cell mass, we performed analysis of covariance of protein levels and case-control status, adjusting for both β -actin and CD31.

Results

Table 1 shows the demographic and BAVM characteristics of the study cohort. Protein levels of DV, α 5, α 2, VEGF, TGF β , and CD31 were all elevated (p < .01) in BAVM compared to controls by western blot analysis (Fig. 1). DV had the largest difference between BAVM cases and controls (~14 fold) and α 5 differed more between BAVM cases and controls than did α 2 (~6 and 2.5 fold, respectively). Furthermore, VEGF and TGF β were also elevated in BAVM compared to control tissue (both ~9 fold). DV, α 5, VEGF, and TGF β protein levels in both BAVM cases and controls were strongly correlated (r = DV:TGF β = 0.86; DV:VEGF = 0.79; DV: α 5 = 0.78; TGF β :VEGF = 0.92; TGF β : α 5 = 0.86; VEGF: α 5 = 0.76). After adjusting for β -actin and CD31, protein levels remained higher in BAVM cases compared to controls for all five proteins (p = .05). β -actin was not associated with protein expression in any of the models (p > .1), while CD31 was only associated with VEGF (p = .009).

In addition to the higher levels of α 5 over that of α 2 (Fig. 1), α 5 (Fig. 2A) also exhibited greater colocalization to CD31-labeled endothelial cells compared to α 2 (Fig. 2B) by immunohistochemistry. Although α 2 was increased in BAVM compared to controls, it was more diffusely distributed than α 5.

Discussion

We have demonstrated in BAVM tissue that (1) Total DV protein is increased; (2) DV's pro-angiogenic receptor $\alpha.5\beta1$ integrin, compared to DV's anti-angiogenic receptor $\alpha.2\beta1$ integrin, has higher protein expression and is more localized to endothelial cells; and (3) Total VEGF and TGF β levels are elevated and are both correlated with DV expression. Because DV is known to potentiate an angiogenic cascade involving VEGF, and VEGF appears to be an important part of the BAVM nidus, DV may contribute to AVM pathology and represent a potential therapeutic target. In this study, we employed resected brain tissue from epilepsy patients as our control human brain tissue. Although the optimal control would be completely unaffected tissue, as epilepsy pathology or drugs that epileptic individuals take could potentially confound our investigation, epilepsy tissue is the otherwise most appropriate and available "normal" human tissue control.

Our data are in agreement with the view that extracellular matrix undergoes active remodeling as part of the angiogenic and inflammatory vascular environment, likely producing soluble DV from full-length perlecan. It is unlikely that high circulating levels of DV are a sufficient condition for BAVM genesis, as I.P. administration of up to 2 mg/kg in rats or mice do not cause focal brain lesions to form. [6] Rather, it is more likely that the higher levels of DV in BAVM tissue contribute to the progression of BAVM rather than cause BAVM per se. However, while the known functions of DV are of high relevance to BAVM, the mechanistic role of DV in the pathology and progression of BAVM cannot be conclusively determined by this observational study, but warrants additional investigation. The role of increased tissue levels of TGF β in BAVM are less clear, but TGF β is involved in many pathways including both vascular stabilization and inflammation, although controversy exists on TGF β actions in the setting of increased angiogensis. [9] However, the

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strong correlation between BAVM cases and controls of DV, VEGF, and TGF β levels suggest that DV is associated with the phenotype of the resected lesion.

The pro-angiogenic brain DV receptor $\alpha.5\beta1$ integrin appeared to have greater localization to endothelial cells compared to the $\alpha 2\beta 1$ integrin, thereby promoting brain endothelial cell angiogenesis. The more diffuse staining of $\alpha 2\beta 1$ integrin likely represents integrin expression on surfaces of other cell types such as astrocytes or neurons, which are of interest but not the focus of our investigation centered on CD31-positive endothelial cells. It is possible, given the strong correlation between DV and $\alpha 5\beta 1$ integrin, that elevated $\alpha 5\beta 1$ levels in BAVM is partly due to the increased protein levels of DV, as perlecan has been found to increase $\alpha 5\beta 1$ expression in brain endothelial cells. [10] $\alpha 5\beta 1$ is normally absent from mature, non-angiogenic brain blood vessels. The expression pattern that we observed — greatly elevated, endothelial-localized α 5 β 1 and slightly elevated but diffusely expressed a2B1 (i.e. not prominently expressed on CD31-positive endothelial cells), an integrin normally absent from brain microvasculature, further underscores the abnormality of the molecular signature of BAVM vascular channels. Finally, as upregulated DV and $\alpha 5\beta 1$ have been identified following cerebral ischemic injury and here in BAVM, this angiogenic protein-receptor association may be of interest to other fields of CNS angiogenic diseases and insults.

Conclusion

Excessive generation of DV and its pro-angiogenic receptor, along with the potentiation of angiogenic and vascular regulatory molecules, suggest that DV could potentially play a role in BAVM pathology and could represent a future therapeutic target.

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Fig. 1.

Protein levels of DV, VEGF, TGF β , a5, a2, and CD31 are increased in human BAVM compared to control (epilepsy) tissue. (A) Representative western blots for each protein investigated. Note that recombinant human DV (rhDV) was used as a positive control for DV blots. (B) Bar graph displaying the mean amounts of DV (p < .001), VEGF (p < .001), TGF β (p < .001), a5 (p = .001), a2 (p = .006), and CD31 (p = .02) were increased in BAVM patients compared to controls, while β -actin was not (p = .649). These relationships remained the same after adjusting for CD31 and β -actin in the ANCOVA models for DV (p = .04), VEGF (p < .001), TGF β (p = .008), a5 (p = .05), and a2 (p = .03).

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Fig. 2.

 $\alpha.5$ integrin, compared to $\alpha.2$ integrin, appears more greatly increased and localized to endothelial cells in human BAVM compared to control (epilepsy) tissue. Coimmunohistochemistry of CD31 (Abcam) with A. $\alpha.5$ (Millipore) or B. $\alpha.2$ (Chemicon International). Original magnification $\times 200$. Note that the increased $\alpha.2$ in BAVM displays a more diffuse staining pattern than $\alpha.5$ in BAVM.

TABLE 1

Baseline Characteristics of Study Cohort

Characteristics	Cases (n = 6)	Controls $(n = 3)$	P Value
Age	43 ± 18	31 ± 8	0.200
Sex			0.524
Male	2 (33%)	2 (67%)	
Female	4 (67%)	1 (33%)	
Presentation			
Hemorrhage	2 (33%)	NA	
Headache	3 (50%)	NA	
Incidental	1 (17%)	NA	
Size (cm)	1.8 ± 0.8	NA	
Venous Drainage			
1	4 (66%)	NA	
2	1 (17%)	NA	
3	1 (17%)	NA	
Location			
Cortical	0 (0%)	NA	
Frontal	4 (66%)	NA	
Occipital	1 (17%)	NA	
Parietal	1 (17%)	NA	
Spetzler-Martin Score*			
1	2 (33%)	NA	
2	3 (50%)	NA	
3	1 (17%)	NA	
4	0 (0%)	NA	
5	0 (0%)	NA	

 $\frac{1}{2}$ age represents SD

 $^{\pm}$ size represents SD (cm)

* The Spetzler-Martin Score is an estimation of the risk of neurosurgery to an AVM patient based on the AVM size, venous drainage, and brain location. Higher values represent a more at-risk patient and less operable AVM. [11]