

UCSF

UC San Francisco Previously Published Works

Title

Killer Cell Immunoglobulin-like Receptor Variants Are Associated with Protection from Symptoms Associated with More Severe Course in Parkinson Disease

Permalink

<https://escholarship.org/uc/item/8fb0q9n5>

Journal

The Journal of Immunology, 205(5)

ISSN

0022-1767

Authors

Anderson, Kirsten M
Augusto, Danillo G
Dandekar, Ravi
[et al.](#)

Publication Date

2020-09-01

DOI

10.4049/jimmunol.2000144

Peer reviewed



Published in final edited form as:

J Immunol. 2020 September 01; 205(5): 1323–1330. doi:10.4049/jimmunol.2000144.

Killer-cell Immunoglobulin-like Receptor Variants Are Associated with Protection from Symptoms Associated with More Severe Course in Parkinson's Disease

Kirsten M. Anderson^{*,||}, Danilo G. Augusto^{*,||}, Ravi Dandekar^{*}, Hengameh Shams^{*}, Chao Zhao^{*}, Tasneem Yusufali^{*}, Gonzalo Montero-Martín[†], Wesley M. Marin^{*}, Neda Nemat-Gorgani[‡], Lisa E. Creary[†], Stacy Caillier^{*}, Mohammad R. K. Mofrad[§], Peter Parham[‡], Marcelo Fernández-Viña[†], Jorge R. Oksenberg^{*}, Paul J. Norman[¶], Jill A. Hollenbach^{*}

^{*}UCSF Weill Institute for Neurosciences, University of California San Francisco, Department of Neurology, Sandler Neuroscience, 675 Nelson Rising Lane, San Francisco, CA 94107

[†]Department of Pathology, Stanford University School of Medicine, Palo Alto, CA 94304

[‡]Department of Structural Biology and Immunology, Stanford University, Palo Alto, CA 94305

[§]Molecular Cell Biomechanics Laboratory, Departments of Bioengineering and Mechanical Engineering, University of California, Berkeley, CA 94720, USA

[¶]Division of Biomedical Informatics and Personalized Medicine, and Department of Immunology and Microbiology, University of Colorado Anschutz Medical Campus, Aurora, CO, 80045

Abstract

Immune dysfunction plays a role in the development of Parkinson's disease (PD). Natural killer (NK) cells regulate immune functions and are modulated by killer cell immunoglobulin-like receptors (KIR). KIR are expressed on the surface of NK cells and interact with human leukocyte antigen (HLA) class I ligands on the surface of all nucleated cells. We investigated *KIR* allelic polymorphism to interrogate the role of NK cells in PD. We sequenced *KIR* genes from 1,314 PD patients and 1,978 controls using next generation methods and identified *KIR* genotypes using custom bioinformatics. We examined associations of *KIR* with PD susceptibility and disease features including age at disease onset and clinical symptoms. We identified two *KIR3DL1* alleles encoding highly expressed inhibitory receptors associated with protection from PD clinical features in the presence of their cognate ligand: *KIR3DL1*015/HLA-Bw4* from rigidity ($p_c = 0.02$, OR = 0.39, 95% CI 0.23 – 0.69) and *KIR3DL1*002/HLA-Bw4i* from gait difficulties ($p_c = 0.05$, OR = 0.62, 95% CI 0.44 – 0.88), as well as composite symptoms associated with more severe disease. We also developed a KIR3DL1/HLA interaction strength metric and found that weak KIR3DL1/HLA interactions were associated with rigidity ($p_c = 0.05$, OR = 9.73, 95% CI 2.13 – 172.5). Highly expressed *KIR3DL1* variants protect against more debilitating symptoms of PD, strongly implying a role of NK cells in PD progression and manifestation.

Corresponding Author: Jill A. Hollenbach, PhD, MPH, University of California, San Francisco, Department of Neurology, 675 Nelson Rising Lane, NS-221A, San Francisco, CA 94158, jill.hollenbach@ucsf.edu.

^{||}Co-first authors

Conflict of Interest Statement: The authors have no conflicts of interest to disclose.

Keywords

Parkinson's disease; KIR; expression; natural killer cells; neurological disorders; brain

Introduction

Parkinson's disease (PD) is a progressive neurodegenerative disease characterized by multiple symptoms including tremors, gait disturbances, postural instability, and rigidity, as well as numerous symptoms impacting sensory perception, cognition, and psychological function. PD is the second most common neurologic disease after Alzheimer's disease, affecting roughly 1% of adults over the age of 50 in the United States (1). The etiology of PD is thought to involve complex interactions between genetics and environmental risk factors (2–4), but it also shares many features of autoimmune neurological diseases, such as immune system dysfunction and disease progression over time. Our recent work (5) and that of others (1, 2, 6) have demonstrated association of PD risk and protection with polymorphism in *human leukocyte antigen (HLA)* genes and with other genes involved in the immune system. However, PD is unique in the environmental factors associated with risk and protection. In marked contrast to most other autoimmune diseases, cigarette smoking, coffee consumption and female sex are all factors associated with protection from PD (4). In addition to overall disease risk, clinical presentations of PD align with differences in disease prognosis. Symptoms impacting the ability to move (gait disturbances, postural instability, rigidity and bradykinesia) are all associated with a more severe PD course (7–10). Conversely, asymmetric onset of symptoms and tremor-dominant disease are associated with milder forms of PD and follow a less dramatic disease course (7, 11). Finally, contrasting with other degenerative brain disorders and dementias such as Alzheimer's, in PD later age of onset is correlated with more rapidly progressing disease (12–14). The heterogeneous presentation of PD is thus characterized by a varying constellation of disease features and complications (7–10).

The precise nature of immune regulatory dysfunction in PD is not known, although research has demonstrated a role for multiple immune cells in protection from and development of disease. For example, in PD patients, T cells respond to alpha-synuclein peptides and these responses drive T cell cytotoxic activity (15). B cells specific for alpha-synuclein are also found in PD. Some studies observed increased numbers of natural killer (NK) cells in the peripheral blood of PD patients (16, 17). NK cells are critical in protection from viral infection, but also have immunoregulatory functions and are implicated in multiple autoimmune diseases and cancers (18–24). NK cells are negative regulators of B cell somatic hypermutation, linking NK cell activity to regulation of antibody production (25). By contrast with PD, in multiple sclerosis (MS), an autoimmune disease of the central nervous system, NK cell numbers decrease during clinical relapse and increase as the patient's symptoms improve (26). Differences in NK activity and regulation suggest alternative roles for NK cells in neurological disease development.

Key modulators of NK cell functions are the killer immunoglobulin-like receptors (KIR) (19, 27, 28). KIR are present on the surface of NK cells and engage HLA class I ligands that

are present on the surface of all nucleated cells (27). These interactions modulate NK cell activity, including the cytotoxic killing of virus infected cells and tumors, or induction of cytokine secretion (29, 30). KIR are encoded by the fast evolving and highly polymorphic *KIR* gene family on chromosome region 19 (19q13.4) (31, 32). Individual *KIR* genes can be present or absent, with individual *KIR* gene-content haplotypes having 4-14 genes, and exhibiting considerable variation between individuals and populations (33, 34). KIR molecules are structurally and functionally distinct, interacting differently with specific sequence motifs of HLA class I allotypes (29).

Thus KIR3DL1 recognizes a sequence motif at positions 77-83 in the α_1 domain and forming the Bw4 epitope present on some HLA-A and HLA-B (35). An isoleucine at position 80 characterizes the HLA-Bw4i ligand, which confers a stronger interaction with KIR than the HLA-Bw4t ligand, which has threonine at position 80 (36, 37). KIR allotypes vary in their inhibitory capacity, which is governed by the strength of their binding to HLA class I and the magnitudes of their cell surface expression (38–41). Several studies report associations between *KIR* gene-content and the development of autoimmune disease, including neurological autoimmunity (24, 42–45). Given the evidence for disruption of NK cell homeostasis in PD, we investigated the possibility that *KIR* gene content and allelic variation play a role in PD. This is the first study to analyze *KIR* allele-level variation in any neurological autoimmune disease.

Using a recently developed sequencing method and custom bioinformatics pipeline (46), we determined and analyzed the association of individual *KIR* alleles with PD and as predictors of specific clinical symptoms. We found that variation associated with high levels of inhibitory KIR expression was associated with protection from specific disease symptoms and may promote a distinct disease course characterized by fewer symptoms impacting movement

Material and Methods

Study populations:

Cohorts of 1,314 Parkinson's disease patients and 1,978 healthy controls were analyzed. Patients and controls consisted of the National Institutes of Health (NIH)-GWAS and NIH-NeuroX datasets derived from the National Institute of Neurological Disorders and Stroke-funded Neurogenetics Repository at the Coriell Institute for Medical Research, as well as the UCSF-EPIC dataset (47). All patients were unrelated Euro-descendants from the United States with idiopathic PD. Controls were unrelated and self-reported Euro-descendants and free from neurologic disorders. Ancestral outliers were identified through analysis of genome-wide markers and removed from the cohorts prior to analysis, as described by Hollenbach et al. 2019 (5). Informed consent was obtained for each participant under locally approved protocols.

Clinical phenotype information.

Information pertaining to clinical phenotypes in PD were acquired from Coriell (48). We analyzed presence and absence of the following clinical phenotypes with respect to *KIR*-

HLA combinations: activation tremor, resting tremor, rigidity, bradykinesia, asymmetric onset, gait difficulties and postural instability.

KIR and HLA genotyping.

DNA samples were genotyped for all *KIR* genes according to Norman 2016 (46). To determine *KIR* gene content and allelic genotypes from NGS data, we used an extended version of our custom bioinformatics pipeline PING (Pushing Immunogenetics into the Next Generation) (46); which takes short-read sequences as input and through multiple alignment and filtration steps identifies *KIR* genotypes as previously described, with additional modifications (Marin et. al, in prep). The *HLA-A*, *-B*, *-C* genotypes for the study cohorts were reported previously (5).

Molecular Dynamics Simulations of KIR3DL1*015 and KIR3DL1*002.

The structure of KIR3DL1 was obtained from the protein data bank (PDB ID: 3vh8) and residue 47 was replaced by valine to model the 3DL1*015 allotype. For 3DL1*002, residue 238 was also changed to arginine using the mutate plugin in Visual Molecular Dynamics (VMD) package (49). Molecular models were solvated in the TIP3P explicit solvent and minimized for 100,000 steps and equilibrated for 10ns prior to the production run. All simulations were carried out using NAMD molecular dynamics package and CHARMM36 forcefield (50). The 3DL1*015 allotype was simulated for 90ns, while the 3DL1*002 simulation continued to 120ns. The NPT ensemble was used in which temperature and pressure were maintained at 310 K and 1 bar using Langevin thermostat and Langevin piston Nose-Hoover, respectively. Timestep of 2fs was used and periodic boundary condition was applied in all directions. Error bars in energy plots were obtained via dividing the standard deviation by \sqrt{n} where n is the number of frames. Visualization and post-processing analyses were performed using VMD and Bio3D packages (51).

KIR interaction scores.

We developed KIR-HLA interaction scores to distinguish within locus differences in KIR and HLA binding propensity. KIR3DL1 interaction scores were determined based on published data of KIR3DL1 allotype binding to HLA (41). We created three categorical measures of KIR3DL1 binding strength (strong, medium, weak) based on the interquartile ranges for the binding percentages determined by Saunders et. al (41). These categories were consistent with a previous analysis of KIR3DL1/HLA interaction strength (52). Our dataset of KIR and HLA genotypes included alleles that were not measured by Saunders et al 2016 (41). To include KIR3DL1-HLA pairs in our dataset for which there is no published functional data, we imputed the strength of the binding to KIR by taking into account 1) similar inhibitory capacity, 2) cell surface expression level and 3) sequence similarity to published allotypes (53, 54); for HLA, we considered sequence similarity to alleles with experimentally measured KIR binding data (39, 41). We identified the closest KIR protein sequence in regions known to interact with HLA and a composite score based on the full potential pairing possible between the KIR and HLA allotypes was given to each individual. All KIR/HLA pairs and their assigned strengths are given in Supplementary Table 1.

Statistical analyses.

Impact of genotype on disease and clinical outcomes were calculated by multivariate regression models using R version 3.6.0 (55). For discrete variables, we analyzed only *KIR* alleles with a frequency >1%, which were analyzed by logistic regression (presence/absence clinical phenotype). We adjusted the models for sex as a covariate. All *p*-values reported are two-tailed and were adjusted for multiple comparisons using the Bonferroni method (56).

Results

High expression *KIR3DL1* alleles in combination with HLA-Bw4/Bw4i reduce gait difficulties and rigidity in Parkinson's disease.

KIR allotypes differ in surface expression and strength of interaction with cognate HLA ligands (39, 52, 53). We analyzed *KIR* allelic polymorphism for all the *KIR* genes in combination with known HLA class I ligands for association with PD disease risk and clinical outcomes (57). No statistically significant *KIR-HLA* associations were observed with respect to disease predisposition in a case/control analysis (Supplemental Figure 1). In contrast, we found evidence that the high expression allele *KIR3DL1*015* in combination with HLA-Bw4 was associated with protection from rigidity, an important clinical feature of PD (Figure 1, $p=0.002$, $p_c=0.02$, OR = 0.39, 95% CI 0.23 – 0.69). Another highly expressed allele *KIR3DL1*002*, in combination with Bw4, was also strongly suggestive for protection from gait impairment ($p=0.006$, $p_c=0.06$, OR = 0.62, 95% CI 0.44 – 0.88).

Weak *KIR3DL1*/HLA interactions associate with higher risk of rigidity and lower risk of resting tremor.

We developed a scoring system to evaluate aggregate *KIR-HLA* interactions at the allele level and their potential inhibitory impact. Using the percent maximum binding values of Saunders et al. 2016 (41), we computed a range of *KIR3DL1*-HLA interaction values. Interaction strength is not associated with disease risk overall. On analyzing interaction strength with respect to PD symptoms, we found that strong *KIR3DL1*-HLA interactions are associated with the presence of resting tremors (Figure 2, $p_c=0.04$, OR = 1.52, 95% CI 1.07 – 2.20). By contrast, weak *KIR3DL1*-HLA interactions were negatively associated with resting tremors (Figure 2, $p_c=0.04$, OR = 0.56, 95% CI 0.35 - 0.92). We also found that weak *KIR3DL1*-HLA interactions are strongly associated with rigidity (Figure 2, $p_c=0.05$, OR = 9.73, 95% CI 2.13 – 172.5).

High-expressing *KIR3DL1*002* is at higher frequency in patients who present with symptoms related to movement.

Thus far we have identified a pattern in PD, whereby high expressing *KIR3DL1* alleles and strong *KIR3DL1*-HLA interactions are associated with protection from PD symptoms impacting movement. In contrast, low expressing *KIR3DL1* and weak *KIR3DL1*-HLA interactions are associated with risk of more debilitating symptoms. From these observations, we categorized the patients based upon their symptoms of PD.

Among the most debilitating PD symptoms are postural instability and gait difficulty, whereas resting tremors and asymmetric onset appear to correspond to a milder disease

course (8, 9). Postural instability and gait difficulty at disease onset and absence of resting tremors predicts poor survival in PD patients (58). Additionally, it has been shown that there are distinct differences in brain matter and morphology between subsets of patients with resting tremors and those with postural instability and gait difficulties (59, 60). Because postural instability and gait difficulty appear to distinguish subgroups of PD patients (8, 9, 60), we assigned the patients into two groups according to their symptoms: one consisting of patients having gait difficulties or postural instability, or both (PIGD+); the other group (PIGD-) comprised patients having neither symptom (Figure 3A). These two groups were then compared for other associations. In the PIGD+ group, the mean age of onset is 59, whereas in the PIGD- group it is 57 ($p = 0.005$). This difference suggests that this subdivision of PD patients has captured an additional factor important in disease prognosis. Furthermore, the PIGD+ group shows significantly increased symptoms of rigidity and bradykinesia, and decreased symptoms of asymmetric onset and resting tremors compared to the PIGD- group (Supplementary Figure 2, Supplementary Table 2).

We also examined *KIR3DL1* allelic variation, expression levels and strength of KIR3DL1-HLA interaction in the two groups of PD patients. The combination of *KIR3DL1*002* and HLA-Bw4 was associated with protection from the PIGD+ category of disease symptoms (Figure 3B, $p_c = 0.04$, OR = 0.57, 95% CI 0.39 – 0.85). Although no other alleles had significant associations after correction for multiple comparisons, we observed a strong trend for strong KIR3DL1 allotypes providing protection from the PIGD+ category of disease symptoms (Figure 3B).

Because KIR3DL1*015 and KIR3DL1*002 differ only at position G238R (Figure 4) and are both high expressing allotypes, we hypothesized that their association with protection from different PD symptoms is associated with conformational differences induced by position 238, which may in turn have important functional differences. To test this hypothesis, we performed all-atom molecular dynamics (MD) simulations of both KIR3DL1 allotypes and tracked conformational changes throughout the trajectories. In 3DL1*002, R238 anchors D2 to D1 and allows surface residues of the two domains to interact more effectively as shown in Figure 5A. Simulations indicated that the overall interaction energy between D1 and D2 was 50 kcal/mol higher in 3DL1*002 than 3DL1*015 (Figure 5B). The G to R substitution at position 238 resulted in 10-fold stronger association of this residue with D1 as shown in Figure 5C. We also observed a negative correlation between G238 and HLA binding residues E201, S227, S228, and D230, in 3DL1*015, which was diminished in 3DL1*002 (Figure 5D).

Discussion

Few studies have analyzed the impact of *KIR* allelic variation in immune-mediated diseases (61). To our knowledge this is the first study to examine the role of *KIR* allotypes and their functional differences in neurological disease. In the context of *KIR* gene content variation, a protective effect has been observed for the presence of *KIR2DS1* in Portuguese MS patients (42). In a study of Spanish MS patients, the activating *KIR3DS1* and inhibitory *KIR2DL5* were associated with disease (62). Our own work showed that the combination of KIR3DL1 and HLA-Bw4 protects against MS (24). While these studies indicate that *KIR*

polymorphism affects the course of neurological disorders, they also point to the value of comparing the allelic differences within each *KIR* gene for their functional differences and associations with disease. Importantly, different *KIR* alleles have known differences in levels of cell surface expression and NK cell inhibition (39, 52, 53, 63). Thus, it is critical to examine *KIR* associations at the allelic level.

While *KIR* allelic variation was not associated with overall disease risk in this study, we found that the *KIR3DL1*015* and *KIR3DL1*002*, in combination with HLA-Bw4, were associated with protection from rigidity and gait difficulties in PD patients, respectively, which are each associated with more severe disease course in PD. Further, *KIR3DL1*002* was shown to be protective from the more severe PIGD+ disease category. Both highly expressed (64), these two allotypes differ only by the amino acid at position 238 (41). Crystallography mutational analyses indicate that residue 238 does not interact directly with the HLA class I ligand (Figure 4) (41). This fact, however, cannot overrule the possibility that this residue causes differences in binding. Previous results suggested that position 238 may affect conformation of ligand binding loops as well as receptor oligomerization, which could partially explain the difference in strength observed for *3DL1*002* and *3DL1*007* molecules (39). In fact, binding differences have been observed for *KIR3DL1*002* and *KIR3DL1*015* (52). To further explore their functional differences, we studied conformational changes of *KIR3DL1*002* and *KIR3DL1*015* allotypes in silico. Simulations indicated that residue 238, located on the surface of the D2 domain, is crucial for adjusting the D1-D2 interaction and directly influences the D1-D1 angle, which is a major regulator of HLA binding. In addition, the coordination of movements between residues at the HLA binding site is affected by the type of residue at position 238 implying an allosteric effect of this residue on the *KIR3DL1* function. Altogether, these results strongly suggest that differential HLA binding is modulated by residue 238-induced structural differences in *KIR3DL1*. Therefore, we provide new insights of the role of position 238 in *KIR3DL1* and suggest the associations of these two allotypes with different PD symptoms could be the result of functional differences of these two molecules caused by the change G238R. These findings are in accordance with recent experimental work by Saunders et al. (65) that demonstrates that allelic variation in *KIR3DL1* results in functional differences impacting the ability of the receptor to interact with HLA.

At the same time, both *KIR3DL1*015* and *KIR3DL1*002* are highly expressed on the NK surface, which also suggests that high *KIR3DL1* expression contributes to protection from some clinical manifestations of disease. Our results suggest that high *KIR3DL1* expression has a protective effect in PD. Expression levels of *KIR3DL1* have significant functional consequences (63, 66) and NK cells with high-expressing *KIR* are correlated with greater NK cell reactivity (52). In human immunodeficiency virus infection (HIV), the combination of high expressing *KIR3DL1* and HLA-Bw4 results in reduced viral load and slower progression to acquired immunodeficiency syndrome (AIDS) (67). The combination of low-expressing *KIR3DL1* and HLA-Bw4 is associated with risk of psoriasis (68), whereas the combination of HLA-Bw4 and *KIR3DL1*, both high and low, provide protection from ankylosing spondylitis (69).

The overall inhibitory capacity of NK cells significantly impacts NK cell function, aiding to control viral replication and prevent tumor development (70). While KIR expression levels may be a useful measure of inhibitory capacity, quantitative data of HLA and KIR expression and their interaction should allow for more informative characterization of their inhibitory potential. Using a weighted score of inhibitory KIR-HLA interaction strength, higher KIR inhibition overall was shown to be associated with lower loads of HIV, improved spontaneous viral clearance of hepatitis C virus (HCV), and protection from human T lymphocyte virus 1 (HTLV-1) infection (70). Such consideration of specific interactions between KIR and HLA class I allotypes enables interpretation from a functional perspective that fully exploits high-resolution KIR and HLA class I sequencing data. For example, HLA-B*57:01 encodes a KIR3DL1 ligand that binds strongly to most KIR3DL1 allotypes (39, 41), whereas other HLA-B allotypes such as that encoded by HLA-B*44:03 have widely varying interactions that depend upon the particular KIR allotype (41). Based on the interaction scores that we calculated for KIR3DL1 with HLA-Bw4, we find that weak interaction scores are negatively associated with resting tremors, a characteristic symptom of a mild PD subtype (9). Weak interaction also positively associates with risk of rigidity, a symptom of a more severe PD (8, 9, 71). Thus, stronger inhibitory potential of KIR3DL1-expressing NK cells protects against the more severe symptoms of PD.

In conclusion, our finding that KIR3DL1 polymorphism is associated with specific combinations of PD symptoms suggests an impact of these NK cell receptors on disease progression. The high expression allele *KIR3DL1*002* combined with the HLA-Bw4 is associated with protection in the PIGD+ patient group, in which movement-associated symptoms predominate. A limitation to our analysis is that it examined disease symptoms at a single time-point, which does not take into account any changes in symptoms over time (72). Further studies examining KIR in PD would benefit from longitudinal analysis to determine whether specific KIR are associated with time to PIGD+ development.

One explanation for the associations observed in this study is related to the immunomodulatory role of NK cells. Key in PD development is the aggregation of alpha-synuclein in Lewy bodies (73). Recently it was shown that antibodies made against alpha-synuclein aid in the clearance of aggregates and prevent their formation, and PD patients have low levels of these protective antibodies (74). An effective B cell response could therefore be essential in preventing PD. NK cells impact B cell antibody responses in several ways: in germinal centers, NK cells may suppress affinity maturation and somatic hypermutation (25), and NK cells may enhance B cell antibody responses through direct interaction with B cells (20, 75). The cause of reduced anti- alpha-synuclein antibody levels in PD patients is unknown but may involve either impaired helper T cells or impaired B cells. In a meta-analysis of lymphocyte subsets in PD, NK cells were more abundant and helper T cells were less abundant than in controls, which could reflect a role for NK cells in regulating helper T cells (17). Thus, NK cell activity may hinder the formation of B cell antibody responses by reducing helper T cell numbers and dampening development of T and B cells.

Alternatively, our findings could point to a role for NK-mediated inflammation in PD. NK cells exert functions as immune regulators through production of cytokines and chemokines

that can either augment or activate inflammation (76). We found that strong KIR3DL1-HLA interactions and high KIR3DL1 expression were protective from disease features in PD. This suggests that more inhibition of NK cells results in reduced NK cell mediated inflammation, which would reduce the more severe symptoms of PD. In fact, a recent study clearly demonstrates that KIR3DL1 allotypes associated with higher cell surface expression form an increased number of receptor clusters and transduce more extensive signals (63). We speculate that our findings are consistent with a mechanism in which NK cells function in PD to inhibit B cell production of alpha-synuclein specific antibodies.

In summary, we have performed the first high-resolution analysis of *KIR* and their functional interactions with HLA class I ligands to identify associations with Parkinson's disease risk and clinical outcomes. Our results paint a consistent picture whereby compound *KIR3DL1-HLA* combinations are protective from more acute clinical features of PD and are consistent with a negative impact of NK cell activity in PD (16, 17). We found a distinct pattern of KIR mediated protection from PD whereby highly inhibitory and/or highly expressed alleles were associated with protection from the most severe symptoms of PD including rigidity, gait difficulties and postural instability. Our study is the first to demonstrate evidence of KIR in PD progression and further study could provide support for PD therapies targeting NK cells.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

We thank all members of the Parkinson Study Group who collected samples used in this study, as well as the patients and their families, whose help and participation made this work possible. The samples used in this work were obtained from the Coriell Biorepository, the Deprenyl and Tocopherol Antioxidative Therapy of Parkinsonism (DATATOP) trial, the UCSF-EPIC project, and the Fox Investigation for New Discovery of Biomarkers (BioFIND; biofind.ioni.usc.edu/). BioFIND is sponsored by the Michael J. Fox Foundation for Parkinson's Research (MJFF) with support from the National Institute for Neurological Disorders and Stroke (NINDS). DATATOP samples were provided for this research by the Indiana University Genetics Biobank with support from the MJFF.

Funding Sources: This study was supported by National Institutes of Health Grant U19NS095774.

REFERENCES

1. Polymeropoulos MH, Higgins JJ, Golbe LI, Johnson WG, Ide SE, Di Iorio G, Sanges G, Stenroos ES, Pho LT, Schaffer AA, Lazzarini AM, Nussbaum RL, and Duvoisin RC. 1996 Mapping of a gene for Parkinson's disease to chromosome 4q21-q23. *Science* 274: 1197–1199. [PubMed: 8895469]
2. Nussbaum RL, and Polymeropoulos MH. 1997 Genetics of Parkinson's disease. *Hum Mol Genet* 6: 1687–1691. [PubMed: 9300660]
3. Gorell JM, Johnson CC, Rybicki BA, Peterson EL, and Richardson RJ. 1998 The risk of Parkinson's disease with exposure to pesticides, farming, well water, and rural living. *Neurology* 50: 1346–1350. [PubMed: 9595985]
4. Hernan MA, Takkouche B, Caamano-Isorna F, and Gestal-Otero JJ. 2002 A meta-analysis of coffee drinking, cigarette smoking, and the risk of Parkinson's disease. *Ann Neurol* 52: 276–284. [PubMed: 12205639]
5. Hollenbach JA, Norman PJ, Creary LE, Damotte V, Montero-Martin G, Caillier S, Anderson KM, Misra MK, Nemat-Gorgani N, Osoegawa K, Santaniello A, Renschen A, Marin WM, Dandekar R, Parham P, Tanner CM, Hauser SL, Fernandez-Vina M, and Oksenberg JR. 2019 A specific amino

acid motif of HLA-DRB1 mediates risk and interacts with smoking history in Parkinson's disease. *Proc Natl Acad Sci U S A* 116: 7419–7424. [PubMed: 30910980]

6. Hamza TH, Zabetian CP, Tenesa A, Laederach A, Montimurro J, Yearout D, Kay DM, Doheny KF, Paschall J, Pugh E, Kusel VI, Collura R, Roberts J, Griffith A, Samii A, Scott WK, Nutt J, Factor SA, and Payami H. 2010 Common genetic variation in the HLA region is associated with late-onset sporadic Parkinson's disease. *Nat Genet* 42: 781–785. [PubMed: 20711177]
7. Geraghty JJ, Jankovic J, and Zetuský WJ. 1985 Association between essential tremor and Parkinson's disease. *Ann Neurol* 17: 329–333. [PubMed: 4004153]
8. Zetuský WJ, Jankovic J, and Pirozzolo FJ. 1985 The heterogeneity of Parkinson's disease: clinical and prognostic implications. *Neurology* 35: 522–526. [PubMed: 3982637]
9. Jankovic J, McDermott M, Carter J, Gauthier S, Goetz C, Golbe L, Huber S, Koller W, Olanow C, Shoulson I, and et al. 1990 Variable expression of Parkinson's disease: a base-line analysis of the DATATOP cohort. The Parkinson Study Group. *Neurology* 40: 1529–1534. [PubMed: 2215943]
10. Gasparoli E, Delibori D, Polesello G, Santelli L, Ermani M, Battistin L, and Bracco F. 2002 Clinical predictors in Parkinson's disease. *Neurol Sci* 23 Suppl 2: S77–78.
11. Zetuský WJ, and Jankovic J. 1985 Laterality and symptom association in Parkinson's disease. *Arch Neurol* 42: 1132–1133. [PubMed: 4062609]
12. Jacobs D, Sano M, Marder K, Bell K, Bylsma F, Lafleche G, Albert M, Brandt J, and Stern Y. 1994 Age at onset of Alzheimer's disease: relation to pattern of cognitive dysfunction and rate of decline. *Neurology* 44: 1215–1220. [PubMed: 8035918]
13. van der Vlies AE, Koedam EL, Pijnenburg YA, Twisk JW, Scheltens P, and van der Flier WM. 2009 Most rapid cognitive decline in APOE epsilon4 negative Alzheimer's disease with early onset. *Psychol Med* 39: 1907–1911. [PubMed: 19335933]
14. Pagano G, Ferrara N, Brooks DJ, and Pavese N. 2016 Age at onset and Parkinson disease phenotype. *Neurology* 86: 1400–1407. [PubMed: 26865518]
15. Sulzer D, Alcalay RN, Garretti F, Cote L, Kanter E, Agin-Liebes J, Liong C, McMurtrey C, Hildebrand WH, Mao X, Dawson VL, Dawson TM, Oseroff C, Pham J, Sidney J, Dillon MB, Carpenter C, Weiskopf D, Phillips E, Mallal S, Peters B, Frazier A, Lindestam Arlehamn CS, and Sette A. 2017 T cells from patients with Parkinson's disease recognize alpha-synuclein peptides. *Nature* 546: 656–661. [PubMed: 28636593]
16. Mihara T, Nakashima M, Kuroiwa A, Akitake Y, Ono K, Hosokawa M, Yamada T, and Takahashi M. 2008 Natural killer cells of Parkinson's disease patients are set up for activation: a possible role for innate immunity in the pathogenesis of this disease. *Parkinsonism Relat Disord* 14: 46–51. [PubMed: 17702627]
17. Jiang S, Gao H, Luo Q, Wang P, and Yang X. 2017 The correlation of lymphocyte subsets, natural killer cell, and Parkinson's disease: a meta-analysis. *Neurol Sci* 38: 1373–1380. [PubMed: 28497309]
18. Bottino C, Moretta L, and Moretta A. 2006 NK cell activating receptors and tumor recognition in humans. *Curr Top Microbiol Immunol* 298: 175–182. [PubMed: 16323416]
19. Boudreau JE, and Hsu KC. 2018 Natural killer cell education in human health and disease. *Curr Opin Immunol* 50: 102–111. [PubMed: 29413815]
20. Gao N, Jennings P, Guo Y, and Yuan D. 2011 Regulatory role of natural killer (NK) cells on antibody responses to *Brucella abortus*. *Innate Immun* 17: 152–163. [PubMed: 20418255]
21. James K, and Ritchie AW. 1984 Do natural killer cells regulate B-cell activity? *Immunol Today* 5: 193–194. [PubMed: 25289953]
22. Moretta L, Bottino C, Pende D, Castriconi R, Mingari MC, and Moretta A. 2006 Surface NK receptors and their ligands on tumor cells. *Semin Immunol* 18: 151–158. [PubMed: 16730454]
23. Hollenbach JA, Ladner MB, Saeteurn K, Taylor KD, Mei L, Haritunians T, McGovern DP, Erlich HA, Rotter JI, and Trachtenberg EA. 2009 Susceptibility to Crohn's disease is mediated by KIR2DL2/KIR2DL3 heterozygosity and the HLA-C ligand. *Immunogenetics* 61: 663–671. [PubMed: 19789864]
24. Hollenbach JA, Pando MJ, Caillier SJ, Gourraud PA, and Oksenberg JR. 2016 The killer immunoglobulin-like receptor KIR3DL1 in combination with HLA-Bw4 is protective against multiple sclerosis in African Americans. *Genes Immun* 17: 199–202. [PubMed: 26866467]

25. Rydzynski CE, Cranert SA, Zhou JQ, Xu H, Kleinstein SH, Singh H, and Waggoner SN. 2018 Affinity Maturation Is Impaired by Natural Killer Cell Suppression of Germinal Centers. *Cell Rep* 24: 3367–3373 e3364. [PubMed: 30257198]
26. Kastrukoff LF, Lau A, Wee R, Zecchini D, White R, and Paty DW. 2003 Clinical relapses of multiple sclerosis are associated with ‘novel’ valleys in natural killer cell functional activity. *J Neuroimmunol* 145: 103–114. [PubMed: 14644036]
27. Anfossi N, Andre P, Guia S, Falk CS, Roetynck S, Stewart CA, Breso V, Frassati C, Reviron D, Middleton D, Romagne F, Ugolini S, and Vivier E. 2006 Human NK cell education by inhibitory receptors for MHC class I. *Immunity* 25: 331–342. [PubMed: 16901727]
28. Hsu KC, Chida S, Geraghty DE, and Dupont B. 2002 The killer cell immunoglobulin-like receptor (KIR) genomic region: gene-order, haplotypes and allelic polymorphism. *Immunol Rev* 190: 40–52. [PubMed: 12493005]
29. Parham P, and Moffett A. 2013 Variable NK cell receptors and their MHC class I ligands in immunity, reproduction and human evolution. *Nat Rev Immunol* 13: 133–144. [PubMed: 23334245]
30. Cooper MA, Colonna M, and Yokoyama WM. 2009 Hidden talents of natural killers: NK cells in innate and adaptive immunity. *EMBO Rep* 10: 1103–1110. [PubMed: 19730434]
31. Guethlein LA, Norman PJ, Hilton HG, and Parham P. 2015 Co-evolution of MHC class I and variable NK cell receptors in placental mammals. *Immunol Rev* 267: 259–282. [PubMed: 26284483]
32. Wende H, Colonna M, Ziegler A, and Volz A. 1999 Organization of the leukocyte receptor cluster (LRC) on human chromosome 19q13.4. *Mamm Genome* 10: 154–160. [PubMed: 9922396]
33. JA Hollenbach IN, MBLadner RM Single, EA Trachtenberg 2012 Killer Cell Immunoglobulin-like Receptor (KIR) Gene-Content Variation in the HGDP-CEPH Populations Immunogenetics in press.
34. Hollenbach JA, Augusto DG, Alaez C, Bubnova L, Fae I, Fischer G, Gonzalez-Galarza FF, Gorodezky C, Karabon L, Kusnierczyk P, Noble J, Rickards O, Roberts C, Schaffer M, Shi L, Tavoularis S, Trachtenberg E, Yao Y, and Middleton D. 2013 16(th) IHIW: Population Global Distribution of Killer Immunoglobulin-like Receptor (KIR) and Ligands. *International journal of immunogenetics* 40: 39–45. [PubMed: 23280119]
35. Gumperz JE, Litwin V, Phillips JH, Lanier LL, and Parham P. 1995 The Bw4 public epitope of HLA-B molecules confers reactivity with natural killer cell clones that express NKB1, a putative HLA receptor. *J Exp Med* 181: 1133–1144. [PubMed: 7532677]
36. Cella M, Longo A, Ferrara GB, Strominger JL, and Colonna M. 1994 NK3-specific natural killer cells are selectively inhibited by Bw4-positive HLA alleles with isoleucine 80. *J Exp Med* 180: 1235–1242. [PubMed: 7931060]
37. Luque I, Solana R, Galiani MD, Gonzalez R, Garcia F, Lopez de Castro JA, and Pena J. 1996 Threonine 80 on HLA-B27 confers protection against lysis by a group of natural killer clones. *Eur J Immunol* 26: 1974–1977. [PubMed: 8765048]
38. Bari R, Bell T, Leung WH, Vong QP, Chan WK, Das Gupta N, Holladay M, Rooney B, and Leung W. 2009 Significant functional heterogeneity among KIR2DL1 alleles and a pivotal role of arginine 245. *Blood* 114: 5182–5190. [PubMed: 19828694]
39. Carr WH, Pando MJ, and Parham P. 2005 KIR3DL1 polymorphisms that affect NK cell inhibition by HLA-Bw4 ligand. *J Immunol* 175: 5222–5229. [PubMed: 16210627]
40. Kim S, Sunwoo JB, Yang L, Choi T, Song YJ, French AR, Vlahiotis A, Piccirillo JF, Cella M, Colonna M, Mohanakumar T, Hsu KC, Dupont B, and Yokoyama WM. 2008 HLA alleles determine differences in human natural killer cell responsiveness and potency. *Proc Natl Acad Sci U S A* 105: 3053–3058. [PubMed: 18287063]
41. Saunders PM, Pymm P, Pietra G, Hughes VA, Hitchen C, O’Connor GM, Loiacono F, Widjaja J, Price DA, Falco M, Mingari MC, Moretta L, McVicar DW, Rossjohn J, Brooks AG, and Vivian JP. 2016 Killer cell immunoglobulin-like receptor 3DL1 polymorphism defines distinct hierarchies of HLA class I recognition. *J Exp Med* 213: 791–807. [PubMed: 27045007]

42. Bettencourt A, Silva AM, Carvalho C, Leal B, Santos E, Costa PP, and Silva BM. 2014 The role of KIR2DS1 in multiple sclerosis--KIR in Portuguese MS patients. *J Neuroimmunol* 269: 52–55. [PubMed: 24529855]
43. Li X, Xia Q, Fan D, Cai G, Yang X, Wang L, Xin L, Ding N, Hu Y, Liu L, Xu S, Xu J, Wang K, and Pan F. 2015 Association between KIR gene polymorphisms and rheumatoid arthritis susceptibility: A meta-analysis. *Hum Immunol* 76: 565–570. [PubMed: 26187163]
44. Rizzo R, Bortolotti D, Gentili V, Rotola A, Bolzani S, Caselli E, Tola MR, and Di Luca D. 2019 KIR2DS2/KIR2DL2/HLA-C1 Haplotype Is Associated with Alzheimer's Disease: Implication for the Role of Herpesvirus Infections. *J Alzheimers Dis* 67: 1379–1389. [PubMed: 30689576]
45. Petrushkin H, Norman PJ, Lougee E, Parham P, Wallace GR, Stanford MR, and Fortune F. 2019 KIR3DL1/S1 Allotypes Contribute Differentially to the Development of Behcet Disease. *J Immunol* 203: 1629–1635. [PubMed: 31405953]
46. Norman PJ, Hollenbach JA, Nemat-Gorgani N, Marin WM, Norberg SJ, Ashouri E, Jayaraman J, Wroblewski EE, Trowsdale J, Rajalingam R, Oksenberg JR, Chiaroni J, Guethlein LA, Traherne JA, Ronaghi M, and Parham P. 2016 Defining KIR and HLA Class I Genotypes at Highest Resolution via High-Throughput Sequencing. *Am J Hum Genet* 99: 375–391. [PubMed: 27486779]
47. University of California, S. F. M SET., Cree BA, Gourraud PA, Oksenberg JR, Bevan C, Crabtree-Hartman E, Gelfand JM, Goodin DS, Graves J, Green AJ, Mowry E, Okuda DT, Pelletier D, von Budingen HC, Zamvil SS, Agrawal A, Caillier S, Ciocca C, Gomez R, Kanner R, Lincoln R, Lizee A, Qualley P, Santaniello A, Suleiman L, Bucci M, Panara V, Papinutto N, Stern WA, Zhu AH, Cutter GR, Baranzini S, Henry RG, and Hauser SL. 2016 Long-term evolution of multiple sclerosis disability in the treatment era. *Ann Neurol* 80: 499–510. [PubMed: 27464262]
48. Simon-Sanchez J, Scholz S, Matarin Mdel M, Fung HC, Hernandez D, Gibbs JR, Britton A, Hardy J, and Singleton A. 2008 Genomewide SNP assay reveals mutations underlying Parkinson disease. *Hum Mutat* 29: 315–322. [PubMed: 17994548]
49. Humphrey W, Dalke A, and Schulten K. 1996 VMD: visual molecular dynamics. *J Mol Graph* 14: 33–38, 27–38. [PubMed: 8744570]
50. Phillips JC, Braun R, Wang W, Gumbart J, Tajkhorshid E, Villa E, Chipot C, Skeel RD, Kale L, and Schulten K. 2005 Scalable molecular dynamics with NAMD. *J Comput Chem* 26: 1781–1802. [PubMed: 16222654]
51. Grant BJ, Rodrigues AP, ElSawy KM, McCammon JA, and Caves LS. 2006 Bio3d: an R package for the comparative analysis of protein structures. *Bioinformatics (Oxford, England)* 22: 2695–2696.
52. Boudreau JE, Mulrooney TJ, Le Ludec JB, Barker E, and Hsu KC. 2016 KIR3DL1 and HLA-B Density and Binding Calibrate NK Education and Response to HIV. *J Immunol* 196: 3398–3410. [PubMed: 26962229]
53. Yawata M, Yawata N, Draghi M, Little AM, Partheniou F, and Parham P. 2006 Roles for HLA and KIR polymorphisms in natural killer cell repertoire selection and modulation of effector function. *J Exp Med* 203: 633–645. [PubMed: 16533882]
54. Thomas R, Yamada E, Alter G, Martin MP, Bashirova AA, Norman PJ, Altfeld M, Parham P, Anderson SK, McVicar DW, and Carrington M. 2008 Novel KIR3DL1 alleles and their expression levels on NK cells: convergent evolution of KIR3DL1 phenotype variation? *J Immunol* 180: 6743–6750. [PubMed: 18453594]
55. Team RC 2019 R: A language and environment for statistical computing. R Foundation for Statistical Computing 3.6.0 ed, Vienna, Austria.
56. Hochberg Y, and Benjamini Y. 1990 More powerful procedures for multiple significance testing. *Stat Med* 9: 811–818. [PubMed: 2218183]
57. Augusto DG, and Petzl-Erler ML. 2015 KIR and HLA under pressure: evidences of coevolution across worldwide populations. *Hum Genet* 134: 929–940. [PubMed: 26099314]
58. Pinter B, Diem-Zangerl A, Wenning GK, Scherfler C, Oberaigner W, Seppi K, and Poewe W. 2015 Mortality in Parkinson's disease: a 38-year follow-up study. *Mov Disord* 30: 266–269. [PubMed: 25447933]

59. Benninger DH, Thees S, Kollias SS, Bassetti CL, and Waldvogel D. 2009 Morphological differences in Parkinson's disease with and without rest tremor. *J Neurol* 256: 256–263. [PubMed: 19219572]
60. Rosenberg-Katz K, Herman T, Jacob Y, Giladi N, Hendler T, and Hausdorff JM. 2013 Gray matter atrophy distinguishes between Parkinson disease motor subtypes. *Neurology* 80: 1476–1484. [PubMed: 23516323]
61. Takeshita LY G-GF, dos Santos EJ, Maia MHv, Rahman MM, Zain SM, Middleton D, Jones AR. 2013 A database for curating the associations between killer-cell immunoglobulin-like receptors and diseases in worldwide populations. Database.
62. Garcia-Leon JA, Pinto-Medel MJ, Garcia-Trujillo L, Lopez-Gomez C, Oliver-Martos B, Prat-Arrojo I, Marin-Banasco C, Suardiaz-Garcia M, Maldonado-Sanchez R, Fernandez-Fernandez O, and Leyva-Fernandez L. 2011 Killer cell immunoglobulin-like receptor genes in Spanish multiple sclerosis patients. *Mol Immunol* 48: 1896–1902. [PubMed: 21665278]
63. Kennedy PR, Barthen C, Williamson DJ, Pitkeathly WTE, Hazime KS, Cumming J, Stacey KB, Hilton HG, Carrington M, Parham P, and Davis DM. 2019 Genetic diversity affects the nanoscale membrane organization and signaling of natural killer cell receptors. *Sci Signal* 12.
64. Gardiner CM, Guethlein LA, Shilling HG, Pando M, Carr WH, Rajalingam R, Vilches C, and Parham P. 2001 Different NK cell surface phenotypes defined by the DX9 antibody are due to KIR3DL1 gene polymorphism. *J Immunol* 166: 2992–3001. [PubMed: 11207248]
65. Saunders PM, MacLachlan BJ, Pymm P, Illing PT, Deng Y, Wong SC, Oates CVL, Purcell AW, Rossjohn J, Vivian JP, and Brooks AG. 2020 The molecular basis of how buried human leukocyte antigen polymorphism modulates natural killer cell function. *Proc Natl Acad Sci U S A*.
66. Kennedy PR, Barthen C, Williamson DJ, and Davis DM. 2019 HLA-B and HLA-C Differ in Their Nanoscale Organization at Cell Surfaces. *Front Immunol* 10: 61. [PubMed: 30761133]
67. Martin MP, Qi Y, Gao X, Yamada E, Martin JN, Pereyra F, Colombo S, Brown EE, Shupert WL, Phair J, Goedert JJ, Buchbinder S, Kirk GD, Telenti A, Connors M, O'Brien SJ, Walker BD, Parham P, Deeks SG, McVicar DW, and Carrington M. 2007 Innate partnership of HLA-B and KIR3DL1 subtypes against HIV-1. *Nat Genet* 39: 733–740. [PubMed: 17496894]
68. Ahn RS, Moslehi H, Martin MP, Abad-Santos M, Bowcock AM, Carrington M, and Liao W. 2016 Inhibitory KIR3DL1 alleles are associated with psoriasis. *Br J Dermatol* 174: 449–451. [PubMed: 26286807]
69. Lopez-Larrea C, Blanco-Gelaz MA, Torre-Alonso JC, Bruges Armas J, Suarez-Alvarez B, Pruneda L, Couto AR, Gonzalez S, Lopez-Vazquez A, and Martinez-Borra J. 2006 Contribution of KIR3DL1/3DS1 to ankylosing spondylitis in human leukocyte antigen-B27 Caucasian populations. *Arthritis Res Ther* 8: R101. [PubMed: 16805919]
70. Boelen L, Debebe B, Silveira M, Salam A, Makinde J, Roberts CH, Wang ECY, Frater J, Gilmour J, Twigger K, Ladell K, Miners KL, Jayaraman J, Traherne JA, Price DA, Qi Y, Martin MP, Macallan DC, Thio CL, Astemborski J, Kirk G, Donfield SM, Buchbinder S, Khakoo SI, Goedert JJ, Trowsdale J, Carrington M, Kollnberger S, and Asquith B. 2018 Inhibitory killer cell immunoglobulin-like receptors strengthen CD8(+) T cell-mediated control of HIV-1, HCV, and HTLV-1. *Sci Immunol* 3.
71. Stebbins GT, Goetz CG, Burn DJ, Jankovic J, Khoo TK, and Tilley BC. 2013 How to identify tremor dominant and postural instability/gait difficulty groups with the movement disorder society unified Parkinson's disease rating scale: comparison with the unified Parkinson's disease rating scale. *Mov Disord* 28: 668–670. [PubMed: 23408503]
72. Kotagal V 2016 Is PIGD a legitimate motor subtype in Parkinson disease? *Ann Clin Transl Neurol* 3: 473–477. [PubMed: 27547776]
73. Baba M, Nakajo S, Tu PH, Tomita T, Nakaya K, Lee VM, Trojanowski JQ, and Iwatsubo T. 1998 Aggregation of alpha-synuclein in Lewy bodies of sporadic Parkinson's disease and dementia with Lewy bodies. *Am J Pathol* 152: 879–884. [PubMed: 9546347]
74. Li X, Koudstaal W, Fletcher L, Costa M, van Winsen M, Siregar B, Inganas H, Kim J, Keogh E, Macedo J, Holland T, Perry S, Bard F, Hoozemans JJ, Goudsmit J, Apetri A, and Pascual G. 2019 Naturally occurring antibodies isolated from PD patients inhibit synuclein seeding in vitro and recognize Lewy pathology. *Acta Neuropathol* 137: 825–836. [PubMed: 30805666]

75. Strowig T, Brilot F, Arrey F, Bougras G, Thomas D, Muller WA, and Munz C. 2008 Tonsilar NK cells restrict B cell transformation by the Epstein-Barr virus via IFN-gamma. *PLoS Pathog* 4: e27. [PubMed: 18266470]
76. Schuster IS, Coudert JD, Andoniou CE, and Degli-Esposti MA. 2016 “Natural Regulators”: NK Cells as Modulators of T Cell Immunity. *Front Immunol* 7: 235. [PubMed: 27379097]

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Key points:

- KIR3DL1 and HLA-Bw4 are associated with different PD clinical features
- Highly expressing KIR3DL1 variants protect against more debilitating symptoms of PD

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

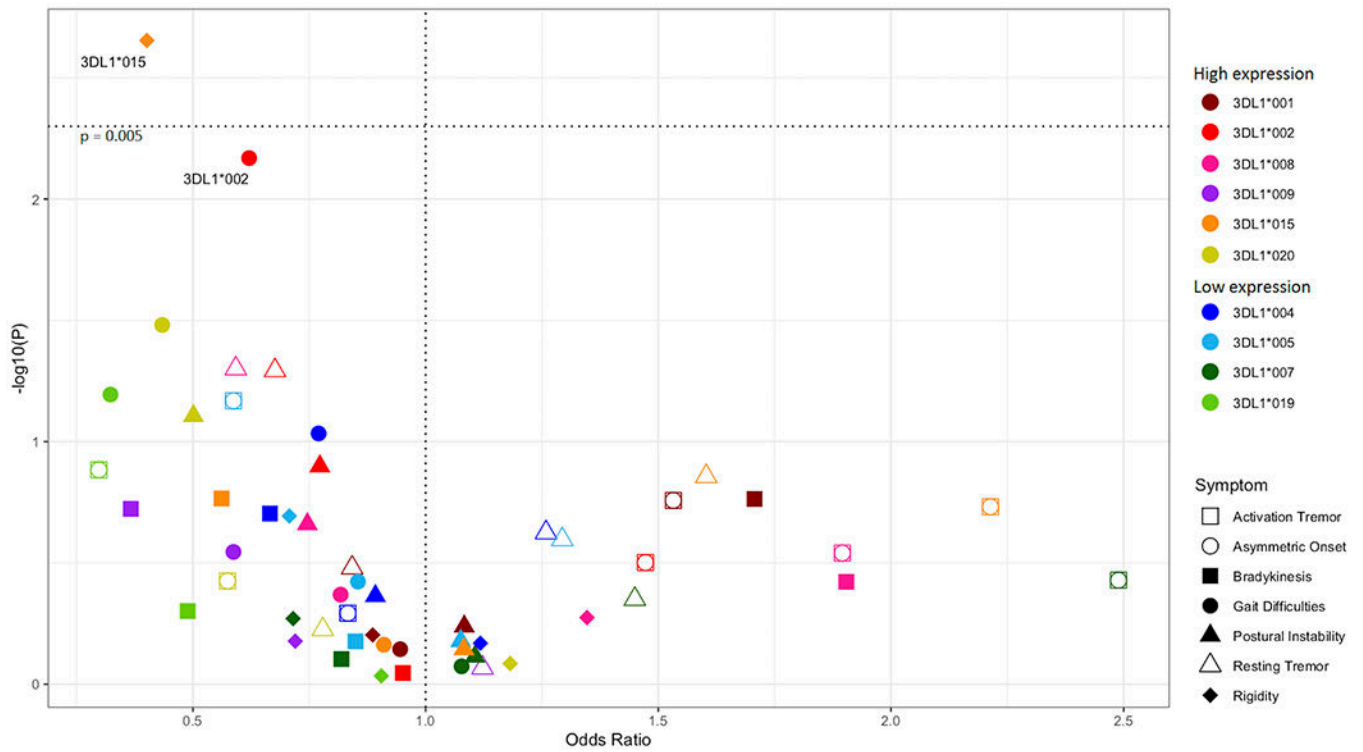


Figure 1. *KIR3DL1* alleles associated with protection from PD symptoms.

All *KIR3DL1* alleles were analyzed in combination with HLA-Bw4 ligands. Odds ratios (x – axis) by $-\log_{10}(p\text{-value})$ (y – axis). Each dot represents the odds ratio and corresponding $-\log_{10}(p\text{-value})$ for a KIR allele/symptom pair. PD symptoms are indicated by shapes and *KIR3DL1* alleles are indicated by color. The horizontal dotted line indicates the significant corrected p-value.

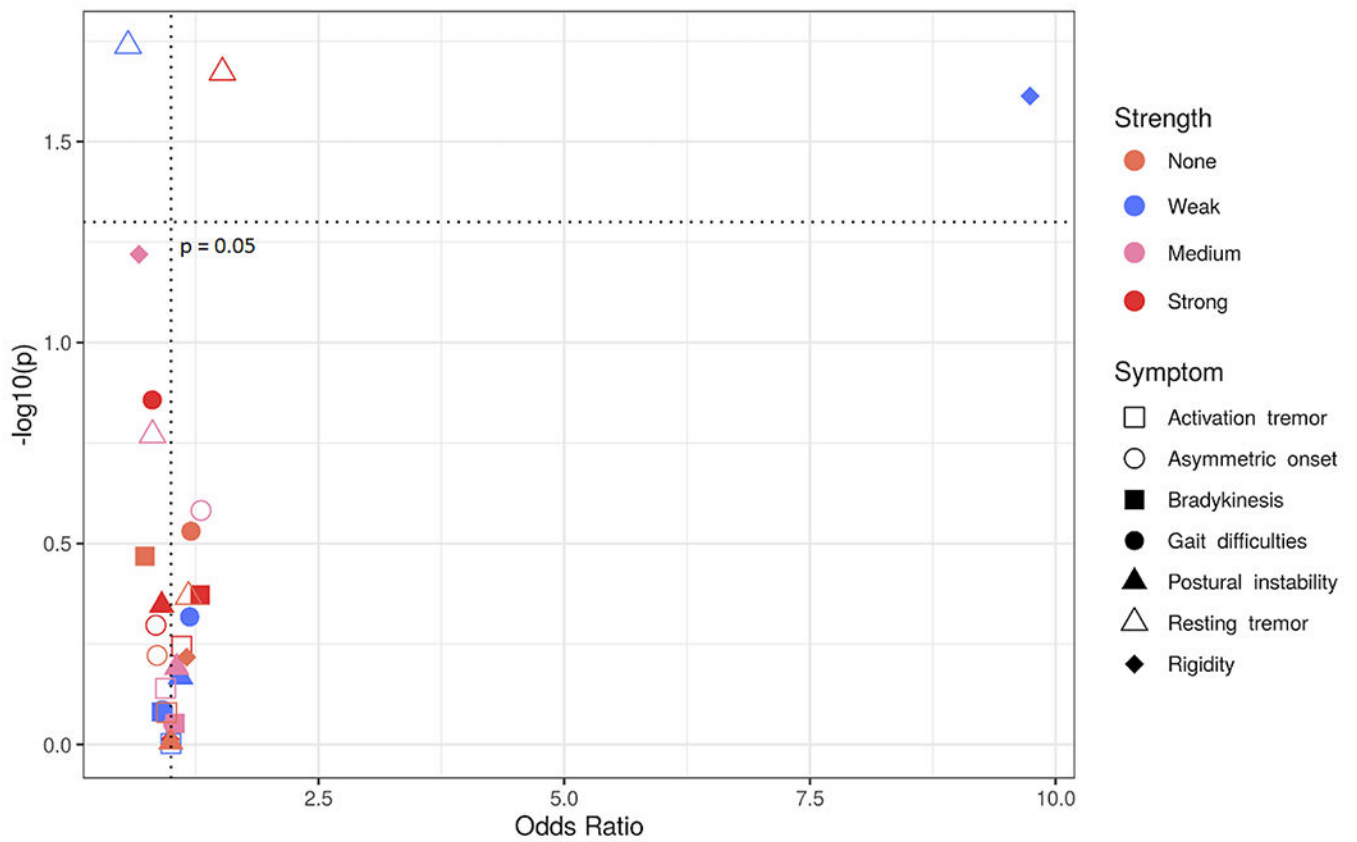


Figure 2. Strong interactions between KIR3DL1 and HLA-Bw4 alleles are associated with clinical features of PD.

Odds ratios (x – axis) by $-\log_{10}(p)$ -value (y – axis). Each dot represents the odds ratio and corresponding $-\log_{10}(p)$ -value for a KIR interaction strength/symptom pair. Different PD symptoms are indicated by shapes and KIR3DL1 interaction strengths are indicated by color.

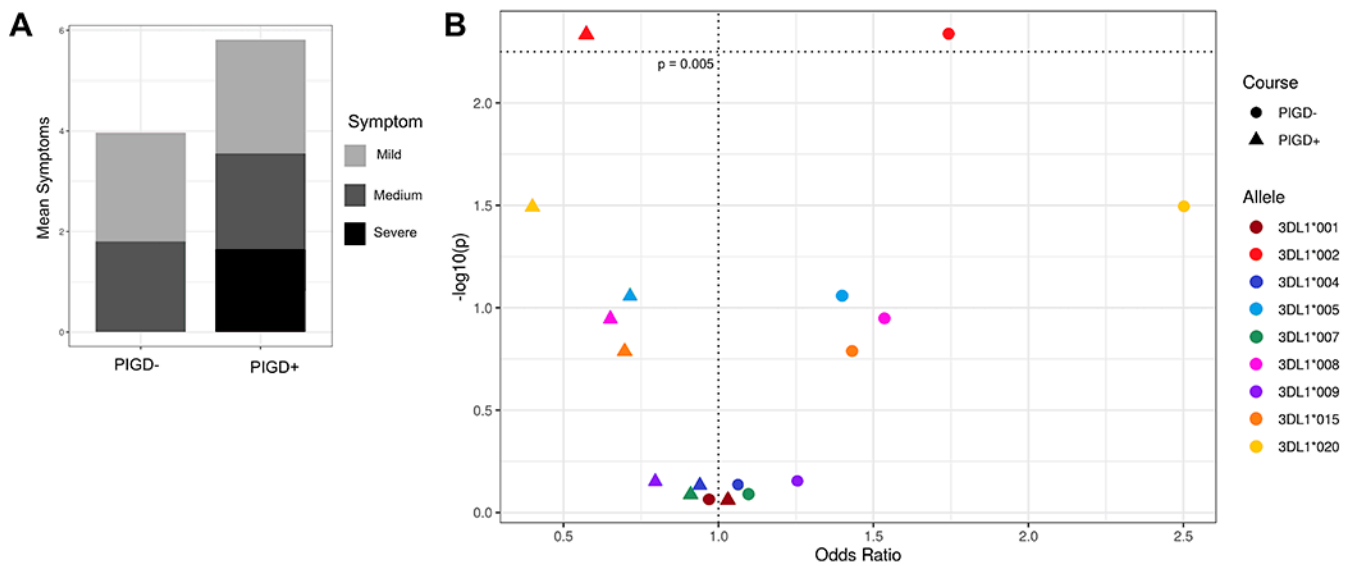


Figure 3. Stratification of PD patients by symptom type.

A) Mean number of symptoms (y – axis) per cluster (x – axis). Symptoms are broken down into three categories where red indicates the most debilitating (postural instability and gait difficulty), coral the moderate symptoms (rigidity and bradykinesia), and light pink the mild symptoms associated with less severe disease (resting and activation tremors and asymmetric onset). B) *KIR3DL1* allele-HLA-Bw4 associations with disease group. Odds ratio (x – axis) by $-\log_{10}(p)$ (y – axis). Each dot represents a *KIR3DL1* allele-HLA ligand pair, the different shapes indicate PIGD- (circle) or PIGD+ (triangle) disease. The horizontal dotted line indicates the value for a significant corrected p -value.

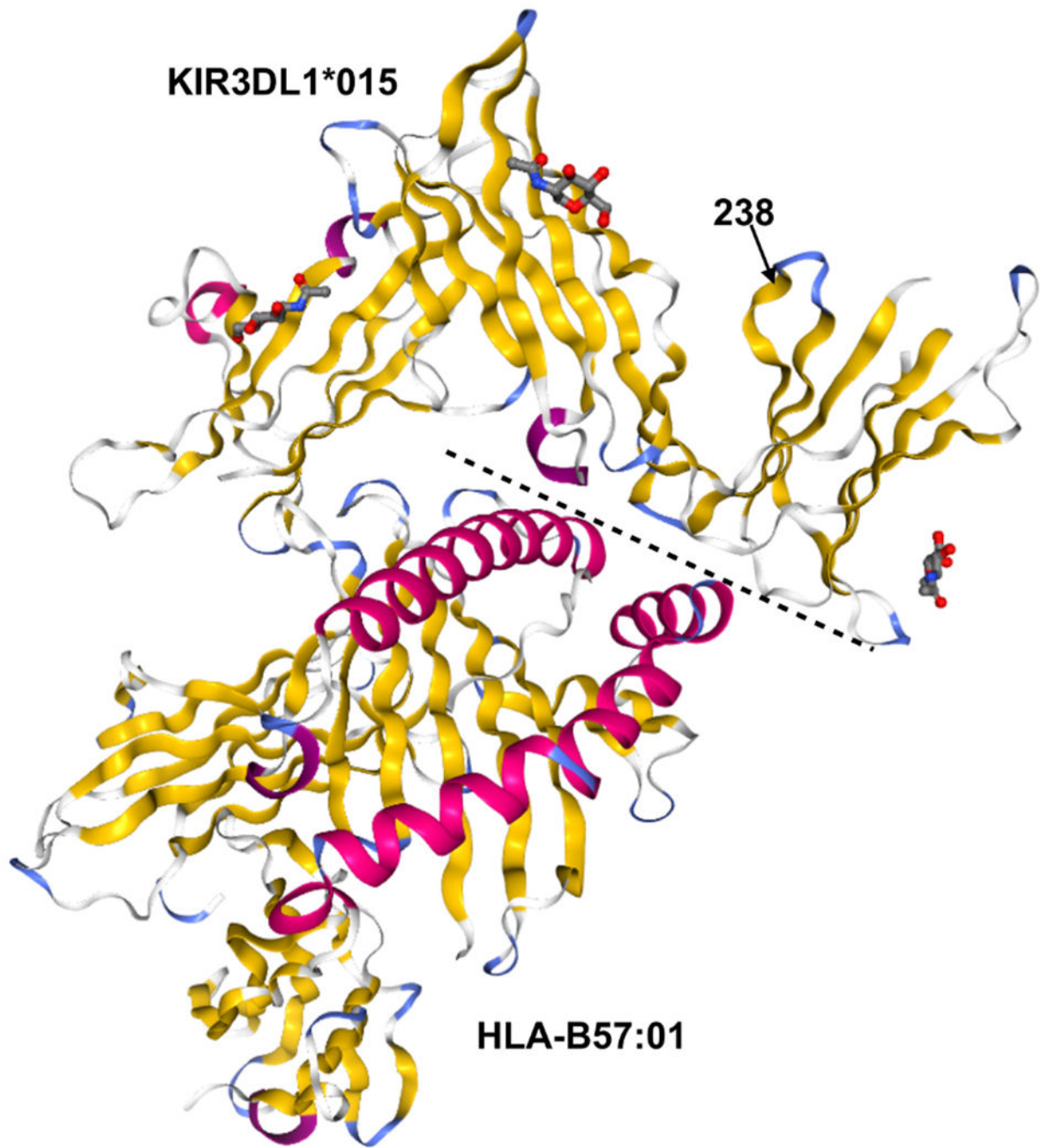


Figure 4. KIR3DL1*015 and KIR3DL1*002 Differ by A Single Amino Acid at Position 238. Crystal structures of KIR3DL1*015 (top) complexed with its ligand HLA-B*57:01 (bottom). Secondary structures are shown by color: alpha helices (pink) and beta sheets (yellow). The dashed line indicates the binding interface between KIR3DL1*015 and HLA-B*57:01. Position 238 is indicated by an arrow. Protein Data Bank accession no. 5B39 (41).

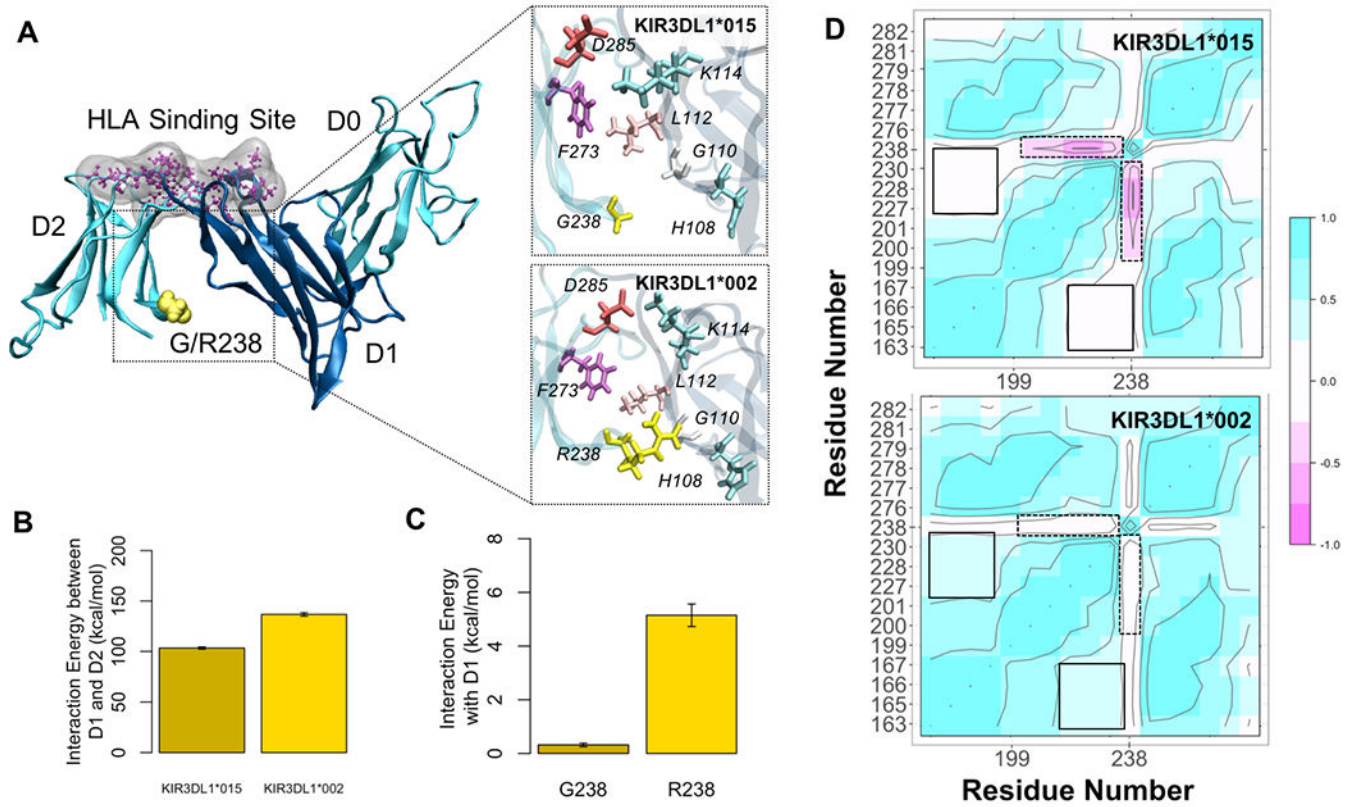


Figure 5. Molecular dynamics simulations of KIR3DL1*015 and KIR3DL1*002.

A) KIR3DL1 is composed of three extracellular domains (D0-D2). The HLA binding site (*purple*) is located at the hinge between D1 and D2. Residue 238 (*yellow*) is in the D2 domain facing towards D1 serving as an anchorage point. The G to R substitution in 3DL1*002 positions D2 closer to D1 allowing more contacts between surface residues of D1 and D2. **B)** This led to an increased D1-D2 interaction energy by 50 kcal/mol in 3DL1*002 compared to 3DL1*015. **C)** The association between residue 238 and D1 was also 10-fold higher in 3DL1*002 as opposed to 3DL1*015. **D)** Residue cross correlation (RCC) map depicts pairwise correlations between residue 238 and the HLA binding site, mapped on residues P163, M165, L166, A167, P199, Y200, E201, S227, S228, D230, F276, R277, H278, S279, Y281, and E282. The negative correlation between G238 and residues E201, S227, S228, and D230 in 3DL1*015 was eliminated upon G238R substitution in 3DL1*002 as shown by the dashed boxes. On the contrary, the positive correlation between HLA binding residues is enhanced in 3DL1*002 (solid boxes).