

# Lawrence Berkeley National Laboratory

## Recent Work

### Title

EXCITED-STATE DYNAMICS OF NADH AND 1-N-PROPYL-1,4-DIHYDRONICOTINAMIDE

### Permalink

<https://escholarship.org/uc/item/8fx5n5ct>

### Author

Boldridge, D.W.

### Publication Date

1984-07-01

UC 4  
LBL-18151  
c-1



**Lawrence Berkeley Laboratory**  
UNIVERSITY OF CALIFORNIA

RECEIVED  
LAWRENCE  
BERKELEY  
UNIVERSITY  
AUG 20 1984

LIBRARY AND  
DOCUMENTS SECTION

# CHEMICAL BIODYNAMICS DIVISION

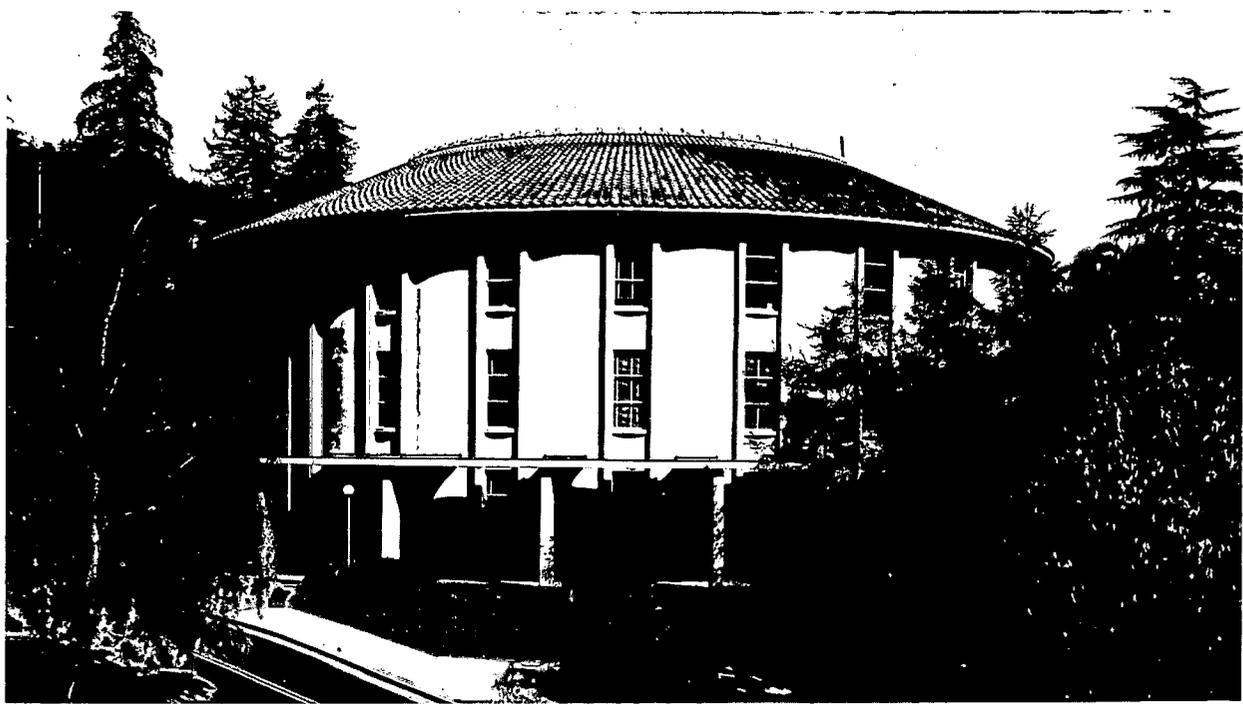
To be published as a chapter in Ultrafast Phenomena,  
D.H. Auston and K.B. Eisenthal, Springer-Verlag,  
Berlin, Heidelberg, New York, 1984

EXCITED-STATE DYNAMICS OF NADH AND  
1-N-PROPYL-1,4-DIHYDRONICOTINAMIDE

D.W. Boldridge, T.H. Morton, G.W. Scott,  
J.H. Clark, L.A. Philips, S.P. Webb,  
S.M. Yeh, and P. van Eikeren

July 1984

**For Reference**  
Not to be taken from this room



LBL-18151  
c-1

## **DISCLAIMER**

This document was prepared as an account of work sponsored by the United States Government. While this document is believed to contain correct information, neither the United States Government nor any agency thereof, nor the Regents of the University of California, nor any of their employees, makes any warranty, express or implied, or assumes any legal responsibility for the accuracy, completeness, or usefulness of any information, apparatus, product, or process disclosed, or represents that its use would not infringe privately owned rights. Reference herein to any specific commercial product, process, or service by its trade name, trademark, manufacturer, or otherwise, does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States Government or any agency thereof, or the Regents of the University of California. The views and opinions of authors expressed herein do not necessarily state or reflect those of the United States Government or any agency thereof or the Regents of the University of California.

# Excited-State Dynamics of NADH and 1-N-propyl-1,4-dihydronicotinamide

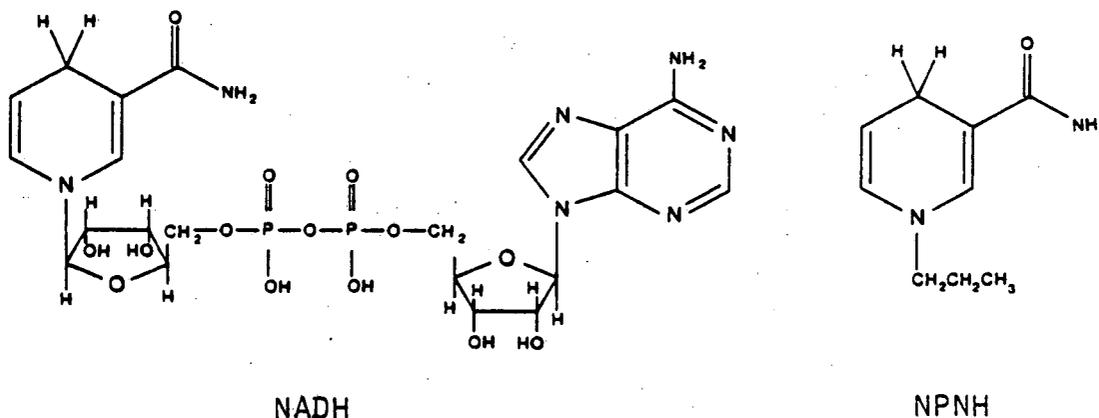
D. W. Boldridge, T. H. Morton, and G. W. Scott  
Department of Chemistry, University of California, Riverside, CA 92521

J. H. Clark, L. A. Philips, S. P. Webb, and S. M. Yeh  
Laboratory for Chemical Biodynamics, Lawrence Berkeley Laboratory,  
and Department of Chemistry, University of California, Berkeley, CA 94720

P. van Eikeren  
Department of Chemistry, Harvey Mudd College, Claremont, CA 91711

The reduced form of nicotinamide-adenine dinucleotide (NADH) is an important enzyme cofactor containing two chromophores - a dihydronicotinamide (lowest absorption band at 340 nm) and an adenine (absorption at 265 nm). Fluorescence of NADH has a  $\lambda_{\text{max}}$  at 460 nm and has been used for in vivo assays of NADH. The photophysics of NADH has been extensively studied [1-9]. In room temperature aqueous solution, the fluorescence quantum yield is 0.02 and the lifetime  $\sim 0.40$  ns [3-5]. Recently, biphotonic induced electron ejection by NADH has been reported [9]. NADH reportedly exists in aqueous solution in two conformations--extended and folded [3,5,10].

The present study reports time-resolved emission and transient absorption studies on the disodium salt of NADH and on a simple analog, 1-N-propyl-1,4-dihydronicotinamide (NPNH), (shown below) in several solvents.



Purity of NADH (Boehringer-Mannheim) and of NPNH (prepared according to ANDERSON and BERKELHAMMER [11]) was tested using a reverse phase HPLC analysis, which showed for each a single peak with a homogeneous uv absorption spectrum. The photophysics of directly excited dihydronicotinamide was observed following excitation by a 355-nm pulse from a modelocked neodymium laser. Fluorescence decay kinetics were obtained using Nd:YAG laser excitation and 2-ps resolution streak camera detection [12]. Transient absorption spectra and kinetics were obtained using Nd:glass laser excitation and a picosecond continuum probe with polychromator/vidicon detection [13]. Energy transfer from adenine to dihydronicotinamide in NADH was studied by fluorescence kinetics after excitation at 266 nm.

Following excitation of NADH, the growth of a broad, unstructured absorption spectrum was observed (see Fig. 1). The similarity of these spectra to that of the solvated electron,  $e_{\text{aq}}^-$  [14], suggests this identification, but does not exclude a solvated electron-ion pair. A deconvolution analysis indicates a buildup time of  $40 \pm 10$  ps for the transient

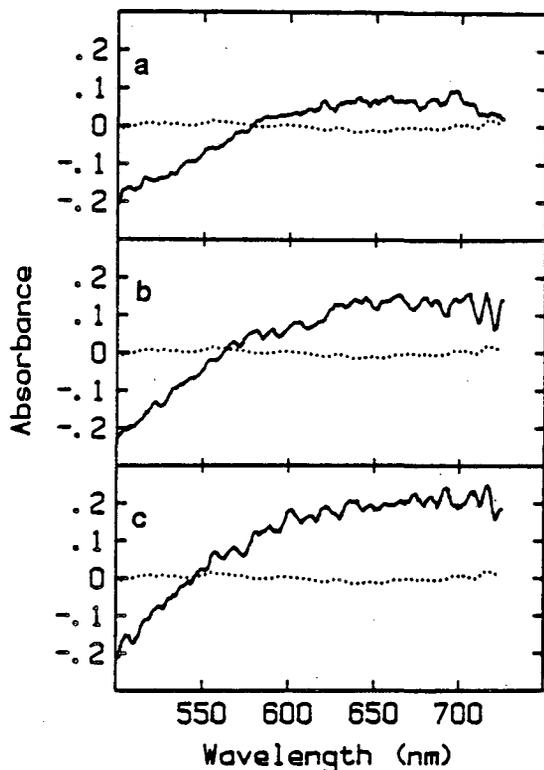


Figure 1. Transient absorption spectra of NADH in aqueous solution at room temperature taken at (a) 7 ps, (b) 20 ps, and (c) 376 ps after excitation at 355 nm. Apparent negative absorbance values at short wavelengths are due to sample fluorescence. Analysis shows that this spectrum grows linearly with intensity and not quadratically [9].

absorption spectrum. No perceptible decay occurs within 2 ns following excitation. Identical observations were made on NPNH. The spectral buildup rate is slower than electron solvation in water [14-16] and may be limited by electron ejection [17,18]. Since the electron ejection time does not match the known NADH fluorescence lifetime [3-5], multiple species and decay routes must be involved. Using the known extinction coefficient for the solvated electron [19], the transient absorbance observed upon excitation of NADH with an actinometrically calibrated pulse gave a quantum yield for photon-induced electron ejection from NADH of  $\approx 0.5$ . This yield is significantly higher than NADH photodecomposition [20], implying a high yield of ion recombination.

NADH fluorescence kinetics data with detection at 380 nm and 460 nm show a previously unreported, fast decaying emission component on the blue edge of the spectrum (see Fig. 2). Fluorescence kinetics detected at 420 to 700 nm yielded only longer fluorescence lifetimes and no risetime ( $<10$  ps). The decay time of the fast component ( $\approx 30$  ps) is similar to the absorption buildup. Fluorescence lifetimes of NADH and NPNH are summarized in the Table.

The fluorescence kinetics show these features: (1) The kinetics are relatively independent of excitation wavelength or the presence of buffer. (2) In  $D_2O$  and ethanol lifetimes are longer than in  $H_2O$ . Since the fluorescence decay kinetics following excitation at 266 nm and at 355 nm are similar, this fluorescence is ascribed to a "folded" conformation of NADH, which allows energy transfer from the adenine [3]. Therefore, folded forms of NADH exist in  $H_2O$  and  $D_2O$  and contribute to the longer-lived fluorescence. The results on NPNH suggest that open forms of NADH also contribute to this longer-lived fluorescence.



$S_1$  is the relaxed, fluorescent, excited singlet state of reduced nicotinamide,  $[D^+ \dots e^-]$  is a solvated, nonfluorescent ion pair, and  $D^+ + e_{aq}^-$  are a separated radical cation and a solvated electron. Initially fluorescence comes from "vertically" excited molecules,  $\{S_1\}$ , with, for example, ground-state solvent organization or hydrogen bonding.  $\{S_1\}$  could exhibit a slightly blue-shifted fluorescence, relative to  $S_1$ , with a lifetime determined by the sum of  $k_1$  and  $k_3$ . The ion pair and the solvated electron may well have similar absorption spectra. Whether the equilibrium with constant  $K$  would need to be established during the lifetimes of  $S_1$  and  $[D^+ \dots e^-]$  is unclear. However, this could explain the complex multiexponential decay reported for the longer lived fluorescence [4, 5]. If the lifetime of  $S_1$  were determined by  $k_2$ , then the rate of ion pair separation would have to depend on solvent deuteration (see table). A high probability of recombination of  $D^+$  with  $e_{aq}^-$  is consistent with its low reactivity observed in the gas phase. NPNH has an ionization potential  $< 8$  eV, and molecular ion dominates its mass spectrum at low ionization energies. In a Fourier Transform Mass Spectrometer,  $NPNH^+$  does not react with neutral NPNH or with  $10^{-6}$  torr of  $NH_3$  on the 0.1 s timescale (during which  $NPNH^+$  experiences  $\sim 50$  collisions with  $NH_3$ ). Further work to refine the kinetic model is in progress.

This work was supported by the Committee on Research, University of California, US Department of Energy Contract DE-AC03-76SF0098, and NIH grants NS 14992 and BRSG RR 07010-15. J. H. Clark is an Alfred P. Sloan Foundation Fellow and a Camille and Henry Dreyfus Foundation Teacher-Scholar. We thank Ms. Marian Hawkes for preparing the manuscript.

1. Dehydrogenases Requiring Nicotinamide Coenzymes, J. Jeffery ed., Birkhauser Verlag, 1980.
2. G. Blankenhorn in Pyridine Nucleotide Dependent Dehydrogenases, H. Sund, ed., De Gruyter, 1977, pp 185-205.
3. T.G. Scott, R.D. Spencer, N.J. Leonard, and G. Weber: J. Am. Chem. Soc. 92, 687 (1970).
4. A. Gafni and L. Brand: Biochem. 15, 3165 (1976).
5. A.J.W.G. Visser and A. van Hoek: Photochem. Photobiol. 33, 35 (1981).
6. M.F. Powell, W.H. Wong, and T.C. Bruice: Proc. Natl. Acad. Sci., USA 79, 4604 (1982).
7. F.M. Martens, J.W. Verhoeven, C.A.G.O. Varma, and P. Bergwerf: J. Photochem. 22, 99 (1983).
8. A. Ohno and N. Kito: Chem. Lett., 369 (1972).
9. B. Czochralska and L. Lindqvist: Chem. Phys. Lett. 101, 297 (1983).
10. N. J. Oppenheimer, L. J. Arnold, and N. O. Kaplan: Biochem. 17, 2613 (1978).
11. A. G. Anderson and G. Berkelhammer: J. Am. Chem. Soc. 80, 992 (1958).
12. S.P. Webb, S.W. Yeh, L.A. Philips, M.A. Tolbert, and J.H. Clark: "Excited-State Proton-Transfer Reactions in 1-Naphthol," in THIS VOLUME.
13. D.W. Boldridge and G.W. Scott: J. Chem. Phys. 79, 3639 (1983).
14. M.J. Bronskill, R.K. Wolff, and J.W. Hunt: J. Chem. Phys. 53, 4201 (1970).
15. G. A. Kenney-Wallace and D.C. Walker: J. Chem. Phys. 55, 447 (1971).
16. W.J. Chase and J.W. Hunt, J. Phys. Chem. 79, 2835 (1975).
17. Y. Wang, J.K. Crawford, M.J. McAuliffe, and K.B. Eisenthal: Chem. Phys. Lett. 74, 160 (1980).
18. G.A. Kenney-Wallace and C.D. Jonah: J. Phys. Chem. 86, 2572 (1982).
19. G.E. Hall and G.A. Kenney-Wallace: Chem. Phys. 32, 313 (1978).
20. V.V. Nikandrow, G.P. Brin, and A. Krasnovskii: Biokhimiya 43, 507 (1978).

This report was done with support from the Department of Energy. Any conclusions or opinions expressed in this report represent solely those of the author(s) and not necessarily those of The Regents of the University of California, the Lawrence Berkeley Laboratory or the Department of Energy.

Reference to a company or product name does not imply approval or recommendation of the product by the University of California or the U.S. Department of Energy to the exclusion of others that may be suitable.

TECHNICAL INFORMATION DEPARTMENT  
LAWRENCE BERKELEY LABORATORY  
UNIVERSITY OF CALIFORNIA  
BERKELEY, CALIFORNIA 94720