

UC Davis

UC Davis Previously Published Works

Title

Issues related to development of new antiseizure treatments

Permalink

<https://escholarship.org/uc/item/8q66w6dc>

Journal

Epilepsia, 54(0 4)

ISSN

0013-9580

Authors

Wilcox, Karen S
Dixon-Salazar, Tracy
Sills, Graeme J
et al.

Publication Date

2013-08-01

DOI

10.1111/epi.12296

Peer reviewed



Published in final edited form as:

Epilepsia. 2013 August ; 54(0 4): 24–34. doi:10.1111/epi.12296.

Issues related to development of new anti-seizure treatments

Karen S. Wilcox¹, Tracy Dixon-Salazar², Graeme J. Sills³, Elinor Ben-Menachem⁴, H. Steve White¹, Roger J. Porter⁵, Marc A. Dichter⁶, Solomon L. Moshé⁷, Jeffery L. Noebels⁸, Michael D. Privitera⁹, and Michael A. Rogawski¹⁰

¹Anticonvulsant Drug Development Program, Department of Pharmacology & Toxicology, University of Utah, Salt Lake City, UT 84108

²Department of Neuroscience, Howard Hughes Medical Institute, University of California, San Diego, La Jolla, CA

³Department of Molecular and Clinical Pharmacology, University of Liverpool, UK

⁴Institution for Clinical Neuroscience, Sahlgrenska Academy, University of Goteborg, Goteborg, Sweden

⁵Epilepsy Therapy Project and Department of Neurology University of Pennsylvania School of Medicine, Philadelphia, PA

⁶Department of Neurology, University of Pennsylvania School of Medicine, Philadelphia, PA

⁷Saul R. Korey Department of Neurology, Dominick P. Purpura Department of Neuroscience, Department of Pediatrics, Laboratory of Developmental Epilepsy, Montefiore/Einstein Epilepsy Management Center, Albert Einstein College of Medicine and Montefiore Medical Center, Bronx, New York, USA

⁸Departments of Neurology, Neuroscience and Molecular and Human Genetics, Baylor College of Medicine, Houston, TX

⁹Department of Neurology, University of Cincinnati, Cincinnati, OH

¹⁰Department of Neurology, University of California, Davis School of Medicine, Sacramento, CA

Summary

This report represents a summary of the discussions led by the anti-seizure treatment working group of the ILAE/AES Working Groups joint meeting in London (London Meeting). We review here what is currently known about the pharmacological characteristics of current models of

Address for correspondence: Karen S. Wilcox, Ph.D., Anticonvulsant Drug Development Program, Department of Pharmacology & Toxicology, University of Utah, Salt Lake City, UT 84108, karen.wilcox@hsc.utah.edu.

Disclosures: KSW has received funding from the NIH. TD-S has received funding from the NIH. GJS has received speaker / consultancy fees and support for conference attendance from Desitin, Eisai Ltd, GlaxoSmithKline and UCB Pharma and has received research funding from BBSRC, Epilepsy Research UK, European Commission, NIHR, and the Wellcome Trust. EB-M has been a Consultant for Eisai, Bial, Lundbeck, Electrocore, Janssen Cilag and Biocontrol. She receives research support from UCB, Bial and Eisai Inc. and VästraGötaland Region. She is Chief Editor of *Acta Neurologica Scandinavica*. HSW has received grant funding from the NIH, served as a consultant for Upsher-Smith Laboratories, Inero Health, Janssen Pharmaceuticals and is scientific co-founder of NeuroAdjuvants, Inc., SLC, UT. HSW has also served on the UCB Pharma Speakers Bureau and is the Senior Research Advisor to Citizen's United for Research in Epilepsy (CURE). MAD has no disclosures. RJP has consulted for NeuroPace, Lilly, Upsher-Smith, Zalicus, Medivation, Astellas, SK Pharma and Civitas. RJP is also the Chief Scientific Officer of the Epilepsy Therapy Project. SLM has received funding from the NIH, DoD, CURE, the Heffer Family Medical Foundation and the Segal Family Foundation. JLN has received funding from the NIH and CURE. MP has received research funding from FDA, American Epilepsy Society, Epilepsy Foundation, Neuren Pharma, Eisai, and UCB; serves on DSMB for Upsher Smith and Lilly. MAR has received support from NINDS, the Epilepsy Therapy Project, Gilead Sciences, and Eisai Inc. We confirm that we have read the Journal's position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

refractory seizures, both for adult and pediatric epilepsy. In addition, we address how the NINDS-funded Anticonvulsant Screening Program (ASP) is evolving to incorporate appropriate animal models in the search for molecules that might be sufficiently novel to warrant further pharmacological development. We also briefly address what we believe is necessary, going forward, to achieve the goal of stopping seizures in all patients, with a call to arms for funding agencies, the pharmaceutical industry, and basic researchers.

Keywords

Anti-seizure drug; pharmaco-resistant epilepsy; animal models of epilepsy

Introduction

Despite the development and availability of more than 22 anti-seizure drugs (ASDs), most of which have been identified as efficacious based on large, double-blind, randomized clinical trials, it is estimated that at least 25-40% of newly diagnosed epilepsy patients will remain resistant to drug therapy and continue to have seizures (Schmidt & Sillanpaa, 2012). These continued seizures have considerable impact on the patients' quality of life and greatly increase the risks of injury, socioeconomic disadvantage, and even death. Continuing seizures can also interfere with memory, cognitive function, educational opportunities, and may produce ongoing endocrine dysfunction (2012a). Thus, the need for more effective therapies remains urgent. However, because the marketplace is already awash with ASDs, many pharmaceutical companies now refrain from the expensive enterprise of developing new compounds. Therefore, the ability of the epilepsy research community to convince a limited number of pharmaceutical and biotechnology companies to finance the development of promising new compounds is a growing concern. This paper summarizes the discussions that took place at the London Workshop to address the strategies of how best to develop new therapies for patients who are resistant, or refractory, to existing ASDs and how to engage a pharmaceutical industry with limited resources to participate.

Most ASDs have been developed with the primary goal of stopping seizures and current drug discovery is based on screening in animal models of acute seizures and epilepsy. Given the failure of new ASDs that help in controlling the refractory population to arise out of testing in these existing models, the field is now engaged in developing new models for preclinical testing. While development in this area has been moving forward with new animal models of drug resistant seizures, the efficacy of the limited number of ASDs that have been successful in those models is yet to be reflected clinically (Schmidt & Sillanpaa, 2012). Thus there was general agreement at the London meeting that there remains an urgent and continuing need to develop, pharmacologically characterize, and validate animal models that mimic the various human epilepsies and can therefore predict efficacy in patients with drug resistant seizures.

In order to aid discussions of this critical topic, definitions of drug resistance have been formulated. These definitions are a result of a consensus obtained at a workshop held at the NIH in 2002 (Stables et al., 2003), by the ILAE (Kwan et al., 2010), and by the authors and participants of the 2012 London Workshop. Drug resistance in humans is herein described as the failure of a patient's seizures to respond to at least two anti-seizure medications that are appropriately chosen, used for an adequate period, and taken with proper adherence to the prescribed regimen. A patient may experience either complete drug resistance or, if seizures are reduced in either frequency and/or intensity, a patient may be classified as exhibiting partial responsiveness. Likewise, drug resistance in animal models is defined as persistent seizure activity that does not respond to monotherapy with at least two appropriate

ASDs. In addition, *in vitro* drug resistance is defined as persistent epileptiform activity that correlates with seizures and is not abated by at least two appropriately chosen ASDs. The underlying mechanisms of pharmacoresistance in epilepsy remain mysterious, with several hypotheses proposed but not proven (Schmidt & Loscher, 2009). It is likely that genetic variability makes a significant contribution and ongoing advances in the pharmacogenomics field should improve our understanding of drug resistant epilepsy, help to identify novel targets for therapeutic intervention, and may ultimately lead to personalized prescribing of both existing ASDs and novel agents that optimizes therapy for the individual patient (Szoek et al., 2006).

The workshop goal was to elucidate strategies for the identification and development of novel therapies within the constraints of current knowledge. New drugs for patients with refractory epilepsy might be more easily identified if we understood why some patients respond well to ASDs while others do not. Sadly, our current level of understanding of the basic mechanisms underlying epilepsy and drug responsiveness is lacking. However, we hypothesize that progress can be made toward obtaining novel therapies with the use of relevant animal models. Therefore, we review here what is currently known about the pharmacological characteristics of current models of refractory seizures, both for adult and pediatric epilepsy. In addition, we address how the NINDS-funded Anticonvulsant Screening Program (ASP) is evolving to incorporate newer animal models in the search for molecules that might be sufficiently novel to warrant further pharmacological development. We also briefly address what we believe is necessary, going forward, to achieve the goal of stopping seizures in all patients, with a call to arms for funding agencies and basic researchers.

Animal models of drug resistant adult epilepsy

Many animal models of drug resistant epilepsy have been developed over the last several decades. These provide a platform for understanding the basic mechanisms of pharmacoresistance and an opportunity to use more clinically relevant models of chronic epilepsy in drug discovery. However, surprisingly little is known about their pharmacological profiles. This lack of information regarding pharmacological sensitivity may be an impediment to future drug discovery efforts, as it is not yet clear which animal models are optimal for screening of compounds. London meeting participants expressed concern that it is becoming increasingly difficult to convince funding agencies of the need for such characterization. Another important consideration is the length of time it would currently take to validate the utility of any given model (i.e. demonstrating the efficacy of compounds selected by the model in refractory epilepsy patients). Nevertheless, in this section, we summarize what is currently known about the pharmacological profiles of the more commonly used animal models of drug resistant epilepsy to help determine if these models share drug response phenotypes with the seizure disorders they supposedly recapitulate. We anticipate that this summary may be useful for the research community in determining where efforts should focus when attempting to identify the most robust animal models for future drug discovery.

In Vivo Models of Drug Resistant Acute Seizures and Chronic Epilepsy

A diverse group of animal models of acquired and genetic epilepsies now exists. The challenge now is to characterize the efficacy of existing ASDs in these models and to publish that data in a clear and consistent manner, even where it is negative. Table 1 summarizes the effects of currently licensed ASDs in the more commonly used models of acquired epilepsy or drug resistant acute seizures. Two specific cautions apply - doses used in animal models may result in concentrations that do not reflect therapeutic levels in humans and differences between specific strains of animals are not considered.

1. 6Hz Stimulation Model—When the MES test failed to identify levetiracetam (LEV) as a potential ASD, the ASP resurrected the 6Hz model in an effort to detect potentially useful ASDs that might otherwise have been overlooked (Brown et al., 1953). Seizures are induced by electrical stimulation of the cornea at a frequency of 6Hz and are characterized by an initial stun, followed by forelimb clonus, twitching of the vibrissae, and finally a Straub-tail (Barton et al., 2001). At low stimulus intensities there is little discrimination between ASDs, with all compounds tested conferring protection against seizures (Barton et al., 2001). However, at a stimulus intensity of 32mA, it was no longer possible to determine an ED₅₀ for either PHT or lamotrigine (LTG). Finally, at the highest stimulus intensity used (44mA), only valproic acid (VPA) and LEV were able to prevent seizures, albeit with higher ED₅₀s (Barton et al., 2001). The ASP and others now routinely employ the 6Hz test for screening due to the relative insensitivity of this test to standard ASDs (Byrtus et al., 2011; Gasior et al., 2010; Kaminski et al., 2011). The rationale for this is that efficacy in the 6Hz test may be indicative of a mechanism of action that is sufficiently unique to encourage the sponsor of the respective compound to continue development.

2. Kindling Models—The amygdala and hippocampal kindling models are proposed models of TLE and both are widely utilized in ASD screening efforts. Validation of these models for therapy discovery has occurred, with protection against fully-kindled seizures often predictive of efficacy in the symptomatic treatment of partial seizures in patients. For the most part, these models are relatively sensitive to existing ASDs, although a lack of efficacy has been reported with some compounds, e.g. topiramate (TPM). Thus, it is arguable whether standard kindling models could be considered models of drug resistant epilepsy. Several groups have, however, modified and further developed the traditional kindling protocol leading to greater therapy resistance. Work from Löscher's group has identified two distinct populations of kindled Wistar rats; one with fully-kindled seizures that are suppressed by PHT and the other in which seizures are resistant to PHT. When the PHT-resistant sub-group of kindled animals is profiled further (Table 1), it shows differential responsiveness to other ASDs, with sensitivity to the anti-seizure effects of LEV, LTG, felbamate (FBM) and gabapentin (GBP) and resistance to vigabatrin (VGB) (Cramer et al., 1998; Ebert et al., 2000; Loscher et al., 2000; Loscher et al., 1993; Reissmuller et al., 2000).

A LTG-resistant kindled rat has also been described (Postma et al., 2000). Animals are kindled by amygdala stimulation in the presence of low concentrations of LTG that do not interfere with the generation of an afterdischarge. Kindled rats are then unresponsive to a later challenge dose of LTG which would otherwise be effective in blocking fully-kindled seizures. This model shows resistance to carbamazepine (CBZ) but not VPA (Srivastava & White, 2012). These observations suggest that seizures induced in this model may be particularly insensitive to ASDs with sodium channel blocking activity. The ASP now frequently uses the LTG-resistant kindled rat as part of routine screening of compounds in an effort to differentiate the pharmacology of the most promising compounds. The clinical relevance of this model to human epilepsy is not known; nonetheless, it provides yet another chronic model wherein the anticonvulsant profile of investigational ASDs can be characterized and differentiated from established ASDs.

Chemoconvulsant and electrically-induced status epilepticus

The capacity to undertake long-term video-EEG recordings from experimental animals is now commonplace. A number of studies have accordingly evaluated the effect of ASDs on the spontaneous recurrent seizures that arise following an initial status epilepticus (SE) induced by either chemoconvulsant (i.e. kainic acid or pilocarpine) or electrical stimulation of the perforant path or other limbic regions. These are often lengthy and expensive studies

to perform, not least because the frequency of spontaneous seizures in rodents can be quite variable (Williams et al., 2009). As of this writing, a comprehensive evaluation of currently available ASDs has not yet occurred in these models. Interestingly, however, the Löscher group has shown that rats exhibiting spontaneous recurrent seizures, subsequent to SE induced by either pilocarpine or electrical stimulation, fall into distinct response groups (responders, non-responders, and partial responders) when treated with either PHT or phenobarbital (PB) (Bethmann et al., 2007). These groups mirror drug response phenotypes in human epilepsy and, together with PHT- and LTG-resistant kindled rats, may be a useful means of identifying ASDs with preferential efficacy against drug resistant seizures.

Another chemoconvulsant model with potential utility in the pre-clinical identification of ASDs with efficacy against drug resistant seizures is the intrahippocampal kainate model. Within a few weeks of injection, both rodents show focal epileptiform activity (measured by EEG) arising from the hippocampus and occasional generalized seizures. While the focal discharges are not associated with behavioral correlates, EEG can be used as a surrogate in efficacy evaluations and the focal discharges occur with sufficient frequency and duration that medium throughput drug studies can be performed with adequate statistical power (Maroso et al., 2011). This model was instrumental in identifying the pre-clinical efficacy of the interleukin converting enzyme inhibitor VX-765, which is currently in clinical trials for the treatment of refractory epilepsy (Ravizza et al., 2006). This model is currently used by a contract research organization, Synapcell, to evaluate traditional ASDs as well as investigational compounds. The electrographic seizures that occur in this model have been found to be responsive to pregabalin (PGB), LEV (at high doses), VGB, diazepam (DZP) and tiagabine (TGB), while VPA, LTG, PHT and CBZ are inactive (Riban et al., 2002). To date, however, these findings have only been published in abstract form (Langlois et al., 2012).

3. Traumatic brain injury—Traumatic brain injury (TBI) in rodents results in the development of spontaneous focal seizures with very occasional secondary generalization (Bolkvadze & Pitkanen, 2012; D'Ambrosio et al., 2009; Kharatishvili & Pitkanen, 2010). The unpredictable seizure frequency has meant that experiments to characterize the effect of ASDs on spontaneous seizures have been difficult to perform. Nevertheless, some data is beginning to emerge which suggests that established ASDs, including CBZ and VPA, are not able to block the focal neocortical seizures that arise as a result of TBI (Eastman et al., 2011; Eastman et al., 2010). While focal seizures in these models may be drug-resistant, from a practical standpoint the low frequency of tonic-clonic convulsions and technical demands of these models mean that they are unlikely to be used as routine screens for new ASDs, and might find their use in the follow-up differentiation process. The search is on for biomarkers, such as altered seizure threshold, which might be employed as indicators of efficacy of potentially novel therapies (Bolkvadze & Pitkanen, 2012).

In Vitro Models of Drug Resistant Seizure-Like Activity

Several *in vitro* models have been evaluated for the drug responsiveness of electrographic measures that are correlated with seizure-like activity (Table 2). The ASP has implemented an *in vitro* model using brain slices prepared from adult kainate-treated rats in which lowered Mg^{2+} (0.1 mM) and elevated K^+ (5mM) concentrations result in recurrent discharges in the entorhinal cortex. These discharges have been shown to be resistant to therapeutic concentrations of PHT and CBZ, but can be entirely suppressed by ezogabine (EZG) (Smith et al., 2007). Similar observations have been made by Heinemann's group (Zhang et al., 1995) in slices prepared from naive rats, although the bursts have a shorter latency to onset in slices from kainate treated animals, increasing the throughput of the assay. Brain slice preparations from human patients following surgical resection of the

seizure focus have also been used and activity seems to be resistant to CBZ in these slices, particularly from patients who were clinically unresponsive to the drug (Jandova et al., 2006; Remy et al., 2003). However, given the variable nature of human specimens and unpredictability of surgeries, this is not a practical drug development assay. Finally, recent work has focused on electrographic, seizure-like activity in organotypic hippocampal slice cultures as a model of drug resistance as well as epileptogenesis (Dyhrfjeld-Johnsen et al., 2010; Wahab et al., 2010b). Burst firing in the slice culture model is insensitive to a number of conventional ASDs and may thus be a useful medium-to-high throughput screen for identification of novel compounds. As was the case for *in vivo* models, it is unclear which, if any, of these *in vitro* models will ultimately prove to be predictive of efficacy against drug resistant seizures.

Animal models of drug resistant pediatric epilepsy

During brain maturation there is a succession of developmental milestones, supported by discrete neuronal processes, during several critical periods in life, starting before birth and lasting till adulthood. Disruptions may lead to epilepsy, which is often drug resistant; in some cases these are due to specific genetic disorders, while in others they may be the result of insults suffered during a critical period, with different underlying consequences and thereby different mechanisms contributing to drug responsiveness. During infancy, childhood and peri-adolescence, specific epileptic syndromes may occur associated with one or more neurodevelopmental processes and for each age-group there may be specific comorbidities, consequences and treatments (Coppola & Moshe, 2009; Coppola & Moshe, 2012; Nehlig, 2012). Accordingly, it would be desirable to develop and study models that mimic these age periods, and which take account of gender, when developing novel ASDs for drug resistant pediatric epilepsy.

Transgenic mouse and model organism technology, combined with advances in our understanding of the complexities of epilepsy genetics, have resulted in a wide array of exciting models of pediatric epilepsies. Mouse models (and in some cases zebra-fish models) for severe myoclonic epilepsy of infancy, benign familial neonatal convulsions, infantile spasms and tuberous sclerosis have all been generated (Chege et al., 2012; Oakley et al., 2013; Price et al., 2009; Singh et al., 2008; Yu et al., 2006). Many of these genetic models exhibit spontaneous recurrent seizures and provide the research community with an opportunity to identify rational therapies for specific pediatric epileptic syndromes. However, very few of these models have been characterized in terms of their responsiveness to existing ASDs (Oakley et al., 2013; Yu et al., 2006; Zhang et al., 2013). As a result, it is currently unknown if these genetic models recapitulate the pharmacoresistant seizure phenotype observed in the corresponding human disorder. In addition, as none of these models are routinely employed in drug discovery, it remains to be seen if they have validity for therapy development.

With respect to pediatric models of induced seizures, there are several that could be considered for use in drug discovery. There are two main concerns with many of these models; ASDs are typically administered prior to the epileptogenic insult and the seizures are induced on a substrate of a normal nervous system. Such models include febrile seizures induced by heating the animal, injection of kainate or NMDA, exposure to flurothyl, kindling, and exposure to hypoxic/ischemic conditions (Galanopoulou, 2013; Moshé & Ludvig, 1988; Noam et al., 2012). For the most part, the resulting seizures only occur in response to the insult and do not generally persist or recur spontaneously. Nevertheless, these models reveal discrete windows of responsiveness to various agents, as exemplified by the flurothyl-induced seizure studies performed by Velisek et al (Velisek et al., 1995a; Velisek et al., 1995b). Hasson et al (Hasson et al., 2008) have also shown that the ability to

control prolonged kainate- or pilocarpine-induced SE is age dependent, with total failure of the tested drugs in postnatal (P) day 9 rats, while the same treatments were effective in P15 and P21 animals. These models represent a viable screen for the acute effects of putative ASDs as a function of age. However, seizures in these models are neither spontaneous nor recurrent and as a consequence do not mimic the phenotype of children with catastrophic epilepsies.

Two models of pediatric epilepsy that result in spontaneous seizures are the tetrodotoxin (TTX) model and a multiple hit model of infantile spasms (Galanopoulou, 2013; Lee et al., 2008). Continuous infusion of TTX into the cortex of young rats results in spontaneous seizures. However, the pharmacological profile of this model has not been explored. In the multiple hit model of infantile spasms, young rats (typically P3) are injected i.c.v. with doxorubicin and lipopolysaccharide and then, two days later, with the serotonin depletion agent p-chlorophenylalanine (Scantlebury et al., 2010) (Galanopoulou, 2013). These animals subsequently develop spontaneous spasms and cognitive disabilities. The spasms meet the criteria for pharmacoresistance, as they are insensitive to both PHT and adrenocorticotrophic hormone (ACTH), a commonly used therapy in infantile spasms (Scantlebury et al., 2010). If the pharmacological characterization of these genetic and acquired models of catastrophic pediatric epilepsies proves to be consistent with the human phenotype, they should be incorporated into future drug screening approaches in an effort to identify therapies in these rare but nonetheless devastating conditions.

Available Infrastructure for Drug Development

The ASP was established in the mid-1970s as a contract between the NINDS and the University of Utah and is the last remaining branch of the Anticonvulsant Drug Development (ADD) program. The ASP has been successful in achieving its primary objective, which was to provide an incentive for drug development for the symptomatic treatment of seizures. The ASP directly contributed to the identification and characterization of nine compounds that are now licensed for the treatment of epilepsy and have provided important preclinical validation for several other approved agents as well as those in the pipeline. While generally better tolerated, it is important to note that the newer ASDs still produce adverse events which can contribute to a poor quality of life (Perucca & Gilliam, 2012) and there still remains a substantial population of pharmacoresistant patients. Thus, the need for novel therapies with better efficacy and improved side effect profiles remains. The ASP was recently reviewed by a group that provided a number of recommendations to enhance outcome in the treatment and prevention of epilepsy. The working group report is available online (http://www.ninds.nih.gov/research/asp/asp_working_group_report_022712.htm) and in *Epilepsia* (2012b). Discussion points are raised in several letters in response to the working group recommendations (Bialer, 2012; French, 2012; Ranganathan, 2012; Shinnar & Glauser, 2012). It is not our intention to revisit the report or the critiques but rather to discuss how the ASP is revising its protocols to increase the likelihood of identifying compounds that will offer significant improvements in treatment.

The issue at hand is defining a pathway that will lead to the identification of more efficacious therapies for the refractory patient. Since the current approach has been successful in its ability to identify mechanistically novel compounds, the currently available models can be used to define efficacy. The problem is that we don't, at this point, know which molecular mechanism(s), if appropriately engaged, will lead to improved seizure control. As such, there is no real guarantee that any of the models discussed above will be more or less likely to identify a highly effective therapy. In other words, is it the model or the mechanism that continues to elude us? In the absence of more insight into the

mechanistic basis of pharmacoresistance, the community needs to be circumspect in its approach to drug discovery and not be too eager to dismiss a process that has, for decades, been extremely successful in bringing new therapies to the patient. Instead, we need to continue to look for complementary approaches and novel models that can be used in parallel to more fully differentiate the anti-seizure profile of novel investigational drugs so that the most promising leads are advanced for further preclinical and clinical evaluation.

Screening Strategies for Future Success at the ASP

As part of the evolution of the ASP, approaches are being considered that will implement more stringent screening assays into its protocol. The idea is to build on the strengths of the MES, 6Hz and other protocols and add filters that will quickly advance the most promising, paradigm-shifting investigational drugs. Early screens in the discovery process must be robust enough to capture positive hits, limit false negatives, be efficient in terms of cost and compound availability, and be of medium-to-high throughput. There are currently no validated animal models of pharmacoresistant epilepsy that meet all of these criteria but those that are well-characterized and considered by the research community to offer predictive value for clinical efficacy are being incorporated and optimized.

Figure 1 illustrates the strategy that has been proposed for implementation of novel screening assays within the ASP. In addition to the MES test, the 6Hz test becomes part of the initial screening process. The corneal kindled mouse model will also be included at an early stage. Efficacy data in this model correlates exceptionally well to that obtained in the hippocampal kindling model (Rowley & White, 2010) and requires very little compound. Compounds that are effective in any of these initial screens will move on to a phase in which activity is differentiated across a range of models. Compounds will be evaluated in the LTG-resistant kindled rat and the *in vitro* slice model derived from kainate-treated rats. In addition, compounds will be evaluated for their ability to alter seizure threshold in the i.v. PTZ test. This test identifies compounds that may lower seizure threshold. A negative outcome in this test would not necessarily halt development; however, it does provide a cautionary signal that should be followed as the compound proceeds into IND-enabling toxicology studies and human development.

Upon meeting prescribed efficacy criteria, compounds will be further investigated in spontaneous seizures models, a pediatric seizure model, and in benzodiazepine-resistant SE. They will also be assessed for their *in vitro* neuroprotective effects and propensity for cognitive and neuropsychiatric adverse events using relevant models. The aim is to identify molecules that are anticonvulsant in the traditional sense, potentially effective against refractory seizures, are more tolerable, and devoid of the propensity to exacerbate comorbidities.

The ASP and NINDS are working closely with sponsors of compounds that might require more specialized testing, such as chronic dosing regimens or use in specific models. This allows sponsors to engage with and utilize the expertise of the ASP if the standard screening protocol is not appropriate. There may also be the flexibility to establish sub-contracts with external laboratories. For example, there is little point in importing genetically engineered models of human epilepsies that display spontaneous recurrent seizures into the ASP when the screening can be performed in their lab of origin. While no such sub-contracts currently exist, it is being explored as a way to leverage finite resources in a difficult drug development landscape.

Partnering with Regulators to Enable New Treatments for Drug Resistant Epilepsy

The preclinical development of treatments for drug resistant epilepsy must be conducted with an understanding of the regulatory requirements for marketing authorization. At the same time, regulators will benefit from an appreciation of developments in preclinical science as they implement changes in policy to enable novel medications to be registered for the treatment of drug resistant epilepsy. An ongoing dialogue between researchers and regulators is essential to ensure that those regulatory pathways exist and are based on sound evidence.

Marketing authorization of new ASDs is based on well-controlled clinical studies demonstrating safety and efficacy in groups of patients with drug resistant epilepsy. However, those trials are not powered or designed to assess seizure freedom or efficacy in patient sub-groups. Guidance on clinical trial designs to assess efficacy in treating drug resistant epilepsy does not exist, and we believe such guidance from regulatory agencies should address the following: (1) preclinical data (if any) to support regulatory decisions; (2) patient selection; (3) clinical endpoints; (4) study designs and duration. Some of these issues are considered below.

Preclinical studies

The EMA guidance acknowledges that studies on drug mechanism and characterization of drug action in diverse experimental models are important, but there is no mention of experimental studies aimed at assessing potential agents for utility in the treatment of drug resistant epilepsy. The FDA guidance does not discuss pre-clinical studies at all. Preclinical studies could provide useful information on issues such as sub-populations that might benefit from a specific treatment, dosing and duration of treatment, appropriate clinical endpoints, potential biomarkers, and expected adverse effects.

Patient selection

Guidance from both FDA and EMA allows studies of a heterogeneous group of patients with incomplete seizure control or who are experiencing excessive adverse effects from current medication. Unfortunately, studies of this kind ensure that a simple demonstration of efficacy will not provide information on the utility of a new drug in any specific subgroup of patients. When there is data to suggest selective efficacy against a specific seizure type or syndrome, this should guide the design of the clinical development program in terms of patient selection. This may also allow the sponsor a rapid and cost effective route to market by capitalizing on a designation of 'orphan drug status'. Under these circumstances, sponsors are incentivized and patients may benefit from a syndrome-specific compound.

Clinical endpoints

Clinical endpoints currently focus on seizure reduction in medication resistant patients using the ASD as adjunctive treatment compared to placebo without emphasis on seizure freedom, monotherapy, or the use of biomarkers. Monotherapy studies in newly diagnosed patients typically target seizure freedom, but are not useful in assessing efficacy in medication resistant epilepsy. Seizure freedom in pharmacoresistant patients is an important endpoint, but one that requires unrealistically large sample sizes. One monotherapy approach currently endorsed by both FDA and EMA is "conversion to monotherapy" where patients with drug resistant epilepsy are randomized to receive either a high or a low dose of the investigational drug (an established ASD may be used as the low dose comparator instead) (Arroyo & Perucca, 2003). The endpoint in this design is retention time following discontinuation of

baseline medications. This design is seldom used because of ethical considerations, although a trial design based on historical controls may be applicable in some instances (French et al., 2012). In addition to traditional efficacy outcomes, funding organizations should support the development of biomarkers. The accompanying manuscript by Engel *et al.*, (2013) discusses this issue.

Summary and Recommendations

The London meeting participants agree that there is a pressing need for better ASDs to address refractory seizures in patients who are not eligible for surgery. Quality of life for those patients is impacted dramatically by uncontrolled seizures and they are at a higher risk of SUDEP. What is not clear is the route to success for new therapy development for these patients in the current scientific and economic climate. One direction is to screen compounds that are rationally designed using a variety of *in vivo* and *in vitro* models that exhibit pharmacoresistant phenotypes. This is an approach that the ASP and the wider epilepsy research community are implementing. The last several decades have seen an enormous increase in the development of genetic and acquired models that recapitulate epilepsy throughout the lifespan. These models have generated new knowledge regarding the basic mechanisms underlying seizure disorders and, in many cases, have identified potential new therapeutic targets. However, there is often little information regarding the pharmacological profiles of existing ASDs in these models. This gap in our knowledge holds the field back, as it is not clear which models will be most useful in identifying novel therapies. Therefore, we recommend that funding be made available for the comparative characterization studies necessary to assess the suitability of these models for drug discovery. Sadly, grant review panels often regard such studies as lacking innovation and manuscript reviewers can be reluctant to accept negative findings for publication, even though negative data with existing ASDs will be the benchmark for models of pharmacoresistant epilepsy. It is essential that these characterization studies are communicated to the field and the use of searchable, open-access, online databases might be one way to address this concern.

A major concern is the issue of bringing new drugs to a saturated epilepsy therapy marketplace, with many pharmaceutical companies having suspended development of ASDs due to the perceived difficulties in recouping the investment required to obtain a new drug approval. However, there was a sense that commercial enthusiasm might be restored under specific circumstances, as follows: (1) it is likely that any compound with disease modifying properties would be sufficiently attractive to merit the necessary investment; this is discussed in more detail in other papers in this series; (2) if an ASD were a novel, first-in-class compound with better tolerability and safety profiles than existing agents, there would potentially be support for development. To that end, the Office of Translational Research at NINDS has several programs now in place that can support the early development of innovative new therapies and which limit the economic risks that companies face (http://www.ninds.nih.gov/funding/areas/translational_research/index.htm); (3) any new treatment that possessed a sufficiently broad anti-seizure profile and also mitigated some of the common co-morbidities associated with epilepsy would be of interest; (4) identification of niche compounds with efficacy against a specific seizure type or syndrome in limited patient populations could take advantage of ‘orphan drug’ status, giving sponsors a privileged route to market. Given recent dramatic developments in the genetics of epilepsy, it is not difficult to envision the reality and benefits of this approach; and (5) modifications to regulatory statutes and guidelines to incentivize companies to pursue the development of new ASDs; such changes could include increasing patent life, pathways to earlier monotherapy licenses for first-in-class compounds, and approval of a catch-all “epilepsy” indication without specifying age, seizure type, or adjunctive use restrictions.

In conclusion, the participants of the London meeting were enthusiastic about recent advances in basic research that have increased our understanding of the mechanisms underlying epilepsy. These have revealed a number of new directions for preclinical drug development and while there is much work still to be done, there is also hope that novel, more-effective and targeted therapies can be rapidly identified using animal models that are more reflective of the clinical condition. Such compounds have the potential to offer unprecedented opportunities for seizure freedom, improved quality of life, and a reduced risk of SUDEP for patients currently living with refractory epilepsy.

Acknowledgments

The authors wish to thank Rafal Kaminski and other meeting participants for helpful discussions.

References

- Epilepsy Across the Spectrum: Promoting Health and Understanding. The National Academies Press; 2012a.
- The NIH/NINDS Anticonvulsant Screening Program (ASP): recommendations from the working group's 2012 review of the Program. *Epilepsia*. 2012b; 53:1837–1839. [PubMed: 23030261]
- Albus K, Wahab A, Heinemann U. Standard antiepileptic drugs fail to block epileptiform activity in rat organotypic hippocampal slice cultures. *Br J Pharmacol*. 2008; 154:709–724. [PubMed: 18414393]
- Arroyo S, Perucca E. Translating monotherapy trials into clinical practice: a look into the abyss. *Epilepsy Behav*. 2003; 4:457–463. [PubMed: 14527485]
- Bankstahl M, Bankstahl JP, Loscher W. Inter-individual variation in the anticonvulsant effect of phenobarbital in the pilocarpine rat model of temporal lobe epilepsy. *Exp Neurol*. 2012; 234:70–84. [PubMed: 22201552]
- Barton ME, Klein BD, Wolf HH, White HS. Pharmacological characterization of the 6 Hz psychomotor seizure model of partial epilepsy. *Epilepsy Res*. 2001; 47:217–227. [PubMed: 11738929]
- Bethmann K, Brandt C, Loscher W. Resistance to phenobarbital extends to phenytoin in a rat model of temporal lobe epilepsy. *Epilepsia*. 2007; 48:816–826. [PubMed: 17319923]
- Bialer M. Comments on the working group's 2012 review of the Program. *Epilepsia*. 2012; 53:1844–1846. [PubMed: 23030267]
- Bolkvadze T, Pitkanen A. Development of post-traumatic epilepsy after controlled cortical impact and lateral fluid-percussion-induced brain injury in the mouse. *J Neurotrauma*. 2012; 29:789–812. [PubMed: 22023672]
- Brown WC, Schiffman DO, Swinyard EA, Goodman LS. Comparative assay of an antiepileptic drugs by psychomotor seizure test and minimal electroshock threshold test. *J Pharmacol Exp Ther*. 1953; 107:273–283. [PubMed: 13035666]
- Byrtus H, Obniska J, Czopek A, Kaminski K. Synthesis and anticonvulsant activity of new n-mannich bases derived from 5-cyclopropyl-5-phenyl-hydantoins. *Arch Pharm (Weinheim)*. 2011; 344:231–241. [PubMed: 21469172]
- Chege SW, Hortopan GA, M TD, Baraban SC. Expression and function of KCNQ channels in larval zebrafish. *Dev Neurobiol*. 2012; 72:186–198. [PubMed: 21692188]
- Coppola A, Moshe SL. Why is the developing brain more susceptible to status epilepticus? *Epilepsia*. 2009; 50(12):25–26. [PubMed: 19941516]
- Coppola A, Moshe SL. Animal models. *Handb Clin Neurol*. 2012; 107:63–98. [PubMed: 22938964]
- Cramer S, Ebert U, Loscher W. Characterization of phenytoin-resistant kindled rats, a new model of drug-resistant partial epilepsy: comparison of inbred strains. *Epilepsia*. 1998; 39:1046–1053. [PubMed: 9776324]
- D'Ambrosio R, Hakimian S, Stewart T, Verley DR, Fender JS, Eastman CL, Sheerin AH, Gupta P, Diaz-Arrastia R, Ojemann J, Miller JW. Functional definition of seizure provides new insight into post-traumatic epileptogenesis. *Brain*. 2009; 132:2805–2821. [PubMed: 19755519]

- Dyhrfjeld-Johnsen J, Berdichevsky Y, Swiercz W, Sabolek H, Staley KJ. Interictal spikes precede ictal discharges in an organotypic hippocampal slice culture model of epileptogenesis. *J Clin Neurophysiol.* 2010; 27:418–424. [PubMed: 21076333]
- Eastman CL, Verley DR, Fender JS, Stewart TH, Nov E, Curia G, D'Ambrosio R. Antiepileptic and antiepileptogenic performance of carisbamate after head injury in the rat: blind and randomized studies. *J Pharmacol Exp Ther.* 2011; 336:779–790. [PubMed: 21123672]
- Eastman CL, Verley DR, Fender JS, Temkin NR, D'Ambrosio R. ECoG studies of valproate, carbamazepine and halothane in frontal-lobe epilepsy induced by head injury in the rat. *Exp Neurol.* 2010; 224:369–388. [PubMed: 20420832]
- Ebert U, Reissmüller E, Loscher W. The new antiepileptic drugs lamotrigine and felbamate are effective in phenytoin-resistant kindled rats. *Neuropharmacology.* 2000; 39:1893–1903. [PubMed: 10884570]
- Ebert U, Rundfeldt C, Lehmann H, Loscher W. Characterization of phenytoin-resistant kindled rats, a new model of drug-resistant partial epilepsy: influence of experimental and environmental factors. *Epilepsy Res.* 1999; 33:199–215. [PubMed: 10094431]
- French J. The value of blind screening in the Anticonvulsant Screening Program. *Epilepsia.* 2012; 53:1842–1843. [PubMed: 23030265]
- French JA, Temkin NR, Shneker BF, Hammer AE, Caldwell PT, Messenheimer JA. Lamotrigine XR conversion to monotherapy: first study using a historical control group. *Neurotherapeutics.* 2012; 9:176–184. [PubMed: 22139591]
- Galanopoulou AS. Basic mechanisms of catastrophic epilepsy - Overview from animal models. *Brain Dev.* 2013
- Gasior M, Yankura J, Hartman AL, French A, Rogawski MA. Anticonvulsant and proconvulsant actions of 2-deoxy-D-glucose. *Epilepsia.* 2010; 51:1385–1394. [PubMed: 20491877]
- Glien M, Brandt C, Potschka H, Loscher W. Effects of the novel antiepileptic drug levetiracetam on spontaneous recurrent seizures in the rat pilocarpine model of temporal lobe epilepsy. *Epilepsia.* 2002; 43:350–357. [PubMed: 11952764]
- Grabenstatter HL, Clark S, Dudek FE. Anticonvulsant effects of carbamazepine on spontaneous seizures in rats with kainate-induced epilepsy: comparison of intraperitoneal injections with drug-in-food protocols. *Epilepsia.* 2007; 48:2287–2295. [PubMed: 17711461]
- Grabenstatter HL, Ferraro DJ, Williams PA, Chapman PL, Dudek FE. Use of chronic epilepsy models in antiepileptic drug discovery: the effect of topiramate on spontaneous motor seizures in rats with kainate-induced epilepsy. *Epilepsia.* 2005; 46:8–14. [PubMed: 15660763]
- Hasson H, Kim M, Moshe SL. Effective treatments of prolonged status epilepticus in developing rats. *Epilepsy Behav.* 2008; 13:62–69. [PubMed: 18337179]
- Jandova K, Pasler D, Antonio LL, Raue C, Ji S, Njunting M, Kann O, Kovacs R, Meencke HJ, Cavalheiro EA, Heinemann U, Gabriel S, Lehmann TN. Carbamazepine-resistance in the epileptic dentate gyrus of human hippocampal slices. *Brain.* 2006; 129:3290–3306. [PubMed: 16951410]
- Kaminski K, Rzepka S, Obniska J. Synthesis and anticonvulsant activity of new 1-[2-oxo-2-(4-phenylpiperazin-1-yl)ethyl]pyrrolidine-2,5-diones. *Bioorg Med Chem Lett.* 2011; 21:5800–5803. [PubMed: 21875804]
- Kharatishvili I, Pitkanen A. Association of the severity of cortical damage with the occurrence of spontaneous seizures and hyperexcitability in an animal model of posttraumatic epilepsy. *Epilepsy Res.* 2010; 90:47–59. [PubMed: 20435440]
- Kwan P, Arzimanoglou A, Berg AT, Brodie MJ, Allen Hauser W, Mathern G, Moshe SL, Perucca E, Wiebe S, French J. Definition of drug resistant epilepsy: consensus proposal by the ad hoc Task Force of the ILAE Commission on Therapeutic Strategies. *Epilepsia.* 2010; 51:1069–1077. [PubMed: 19889013]
- Langlois M, M.; Bouyssieres, CV.; Duveau, V.; Dumont, C.; Roucard, C. Improving the development of promising drug candidates: preclinical solutions developed in chronic models of epilepsy. Annual Meeting of the American Epilepsy Society; San Diego, CA. 2012.
- Lee CL, Frost JD Jr, Swann JW, Hrachovy RA. A new animal model of infantile spasms with unprovoked persistent seizures. *Epilepsia.* 2008; 49:298–307. [PubMed: 17941850]

- Leite JP, Cavalheiro EA. Effects of conventional antiepileptic drugs in a model of spontaneous recurrent seizures in rats. *Epilepsy Res.* 1995; 20:93–104. [PubMed: 7750514]
- Loscher W, Reissmuller E, Ebert U. Anticonvulsant efficacy of gabapentin and levetiracetam in phenytoin-resistant kindled rats. *Epilepsy Res.* 2000; 40:63–77. [PubMed: 10771259]
- Loscher W, Rundfeldt C, Honack D. Pharmacological characterization of phenytoin-resistant amygdala-kindled rats, a new model of drug-resistant partial epilepsy. *Epilepsy Res.* 1993; 15:207–219. [PubMed: 8223417]
- Maroso M, Balosso S, Ravizza T, Iori V, Wright CI, French J, Vezzani A. Interleukin-1beta biosynthesis inhibition reduces acute seizures and drug resistant chronic epileptic activity in mice. *Neurotherapeutics.* 2011; 8:304–315. [PubMed: 21431948]
- Moshé, SL.; Ludvig, N. Kindling. In: Pedley, TA.; Meldrum, BS., editors. *Recent Advances in Epilepsy.* Churchill Livingstone; 1988. p. 21-44.
- Nehlig, A. Syndromes, animal models, and brain development. In: Bureau, M.; Genton, P.; Dravet, C.; Delgado-Escueta, AV.; Tassinari, CA.; Thomas, P.; Wolf, P., editors. *Epileptic Syndromes in Infancy, Childhood and Adolescence.* John Libbey Eurotext Ltd; 2012.
- Noam Y, Raol YH, Holmes GL. Searching for new targets for treatment of pediatric epilepsy. *Epilepsy Behav.* 2012
- Oakley JC, Cho A, Cheah CS, Scheuer T, Catterall WA. Synergistic GABA-enhancing therapy against seizures in a mouse model of Dravet syndrome. *J Pharmacol Exp Ther.* 2013
- Perucca P, Gilliam FG. Adverse effects of antiepileptic drugs. *Lancet Neurol.* 2012; 11:792–802. [PubMed: 22832500]
- Postma T, Krupp E, Li XL, Post RM, Weiss SR. Lamotrigine treatment during amygdala-kindled seizure development fails to inhibit seizures and diminishes subsequent anticonvulsant efficacy. *Epilepsia.* 2000; 41:1514–1521. [PubMed: 11114208]
- Price MG, Yoo JW, Burgess DL, Deng F, Hrachovy RA, Frost JD Jr, Noebels JL. A triplet repeat expansion genetic mouse model of infantile spasms syndrome, Arx(GCG)₁₀₊₇, with interneuronopathy, spasms in infancy, persistent seizures, and adult cognitive and behavioral impairment. *J Neurosci.* 2009; 29:8752–8763. [PubMed: 19587282]
- Ranganathan R. The response of NINDS to recommendations from the working group's 2012 review of the Program. *Epilepsia.* 2012; 53:1839–1840. [PubMed: 23030262]
- Rattka M, Brandt C, Loscher W. The intrahippocampal kainate model of temporal lobe epilepsy revisited: Epileptogenesis, behavioral and cognitive alterations, pharmacological response, and hippocampal damage in epileptic rats. *Epilepsy Res.* 2013; 103:135–152. [PubMed: 23196211]
- Ravizza T, Lucas SM, Balosso S, Bernardino L, Ku G, Noe F, Malva J, Randle JC, Allan S, Vezzani A. Inactivation of caspase-1 in rodent brain: a novel anticonvulsive strategy. *Epilepsia.* 2006; 47:1160–1168. [PubMed: 16886979]
- Reissmuller E, Ebert U, Loscher W. Anticonvulsant efficacy of topiramate in phenytoin-resistant kindled rats. *Epilepsia.* 2000; 41:372–379. [PubMed: 10756400]
- Remy S, Gabriel S, Urban BW, Dietrich D, Lehmann TN, Elger CE, Heinemann U, Beck H. A novel mechanism underlying drug resistance in chronic epilepsy. *Annals of neurology.* 2003; 53:469–479. [PubMed: 12666114]
- Riban V, Bouilleret V, Pham-Le BT, Fritschy JM, Marescaux C, Depaulis A. Evolution of hippocampal epileptic activity during the development of hippocampal sclerosis in a mouse model of temporal lobe epilepsy. *Neuroscience.* 2002; 112:101–111. [PubMed: 12044475]
- Rowley NM, White HS. Comparative anticonvulsant efficacy in the corneal kindled mouse model of partial epilepsy: Correlation with other seizure and epilepsy models. *Epilepsy Res.* 2010; 92:163–169. [PubMed: 20951004]
- Scantlebury MH, Galanopoulou AS, Chudomelova L, Raffo E, Betancourth D, Moshe SL. A model of symptomatic infantile spasms syndrome. *Neurobiol Dis.* 2010; 37:604–612. [PubMed: 19945533]
- Schmidt D, Loscher W. New developments in antiepileptic drug resistance: an integrative view. *Epilepsy Curr.* 2009; 9:47–52. [PubMed: 19421380]
- Schmidt D, Sillanpaa M. Evidence-based review on the natural history of the epilepsies. *Curr Opin Neurol.* 2012; 25:159–163. [PubMed: 22274775]

- Shinnar S, Glauser T. Commentary on Anticonvulsant Screening Program update: children are not little adults. *Epilepsia*. 2012; 53:1843–1844. [PubMed: 23030266]
- Singh NA, Otto JF, Dahle EJ, Pappas C, Leslie JD, Vilaythong A, Noebels JL, White HS, Wilcox KS, Leppert MF. Mouse models of human KCNQ2 and KCNQ3 mutations for benign familial neonatal convulsions show seizures and neuronal plasticity without synaptic reorganization. *J Physiol*. 2008; 586:3405–3423. [PubMed: 18483067]
- Smith MD, Adams AC, Saunders GW, White HS, Wilcox KS. Phenytoin- and carbamazepine-resistant spontaneous bursting in rat entorhinal cortex is blocked by retigabine in vitro. *Epilepsy Res*. 2007; 74:97–106. [PubMed: 17395429]
- Srivastava AK, White HS. Carbamazepine, but not valproate, displays pharmacoresistance in lamotrigine-resistant amygdala kindled rats. *Epilepsy Res*. 2012
- Stables JP, Bertram E, Dudek FE, Holmes G, Mathern G, Pitkanen A, White HS. Therapy discovery for pharmacoresistant epilepsy and for disease-modifying therapeutics: summary of the NIH/NINDS/AES models II workshop. *Epilepsia*. 2003; 44:1472–1478. [PubMed: 14636315]
- Szoeke CE, Newton M, Wood JM, Goldstein D, Berkovic SF, TJ OB, Sheffield LJ. Update on pharmacogenetics in epilepsy: a brief review. *Lancet Neurol*. 2006; 5:189–196. [PubMed: 16426995]
- Velisek L, Veliskova J, Ptachewich Y, Ortiz J, Shinnar S, Moshe SL. Age-dependent effects of gamma-aminobutyric acid agents on flurothyl seizures. *Epilepsia*. 1995a; 36:636–643. [PubMed: 7555978]
- Velisek L, Veliskova J, Ptachewich Y, Shinnar S, Moshe SL. Effects of MK-801 and phenytoin on flurothyl-induced seizures during development. *Epilepsia*. 1995b; 36:179–185. [PubMed: 7821276]
- Wahab A, Albus K, Gabriel S, Heinemann U. In search of models of pharmacoresistant epilepsy. *Epilepsia*. 2010a; 51(3):154–159. [PubMed: 20618423]
- Wahab A, Albus K, Heinemann U. Drug refractoriness of epileptiform activity in organotypic hippocampal slice cultures depends on the mode of provocation. *Epilepsy Res*. 2010b; 90:304–308. [PubMed: 20599358]
- Williams PA, White AM, Clark S, Ferraro DJ, Swiercz W, Staley KJ, Dudek FE. Development of spontaneous recurrent seizures after kainate-induced status epilepticus. *J Neurosci*. 2009; 29:2103–2112. [PubMed: 19228963]
- Yu FH, Mantegazza M, Westenbroek RE, Robbins CA, Kalume F, Burton KA, Spain WJ, McKnight GS, Scheuer T, Catterall WA. Reduced sodium current in GABAergic interneurons in a mouse model of severe myoclonic epilepsy in infancy. *Nat Neurosci*. 2006; 9:1142–1149. [PubMed: 16921370]
- Zhang B, McDaniel S, Rensing NR, Wong M. Vigabatrin Inhibits Seizures and mTOR Pathway Activation in a Mouse Model of Tuberous Sclerosis Complex. *PLOS One*. 2013
- Zhang CL, Dreier JP, Heinemann U. Paroxysmal epileptiform discharges in temporal lobe slices after prolonged exposure to low magnesium are resistant to clinically used anticonvulsants. *Epilepsy Res*. 1995; 20:105–111. [PubMed: 7750506]

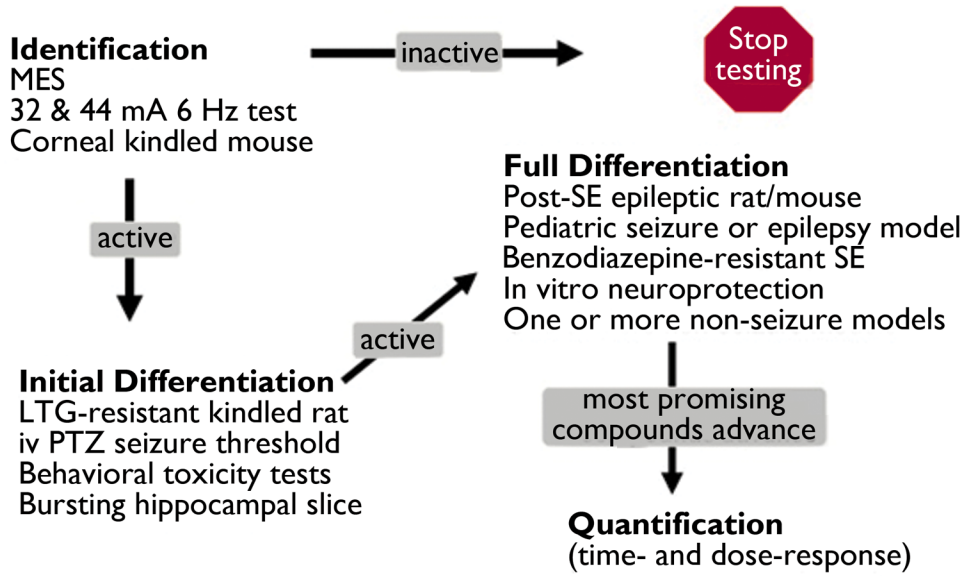


Figure 1. Proposed testing protocol of the ASP based on recommendations of the working group that was assembled to review the program in 2011. The inclusion of the corneal kindled mouse at the front end provides a chronic seizure model that was missing from the original screening mechanism.

Table 1
Pharmacological properties of animal models of acute and chronic drug resistant seizures

Animal Model	Pharmacosensitivity of Spontaneous Seizures		Seizure type	References
	Sensitive to ASD	Resistant to ASD		
6 Hz acute seizures (44 mA)	EZG, LEV, VPA	PHT, TPM, LTG	Complex partial seizures	(Barton et al., 2001)
Corneal kindled mouse	EZG, LEV, LTG, PHT, TGB, CBZ, VGB, VPA	TPM		(Rowley & White, 2010)
Amygdala kindled rat*	VPA, FBM, CZP, GBP, LTG, LEV	TPM	Focal seizures that secondarily generalize	
LTG-resistant kindled rat	FBM, EZG, VPA	LTG, CBZ, PHT, TPM	Focal seizures that secondarily generalize	(Postma et al., 2000; Srivastava & White, 2012)
PHT-resistant kindled rat	LEV, LTG, FBM, GBP, TPM	PHT, VGB	Focal seizures that secondarily generalize	(Ebert et al., 2000; Ebert et al., 1999; Loscher et al., 2000; Loscher et al., 1993)
KA-induced SRS	CBZ	TPM* (seizure freq reduced by 56%)	Focal seizures that secondarily generalize	(Grabenstatter et al., 2007; Grabenstatter et al., 2005)
PILO-induced SRS	LEV [§] , PB, CBZ, PHT	ESM	Focal seizures that secondarily generalize	(Glien et al., 2002; Leite & Cavalheiro, 1995)
PB-resistant PILO-induced SRS		PB, PHT	Focal seizures that secondarily generalize	(Bankstahl et al., 2012)
Intrahippocampal KA (mouse)	PGB, LEV, VGB, DZP, TGB	VPA, LTG, PHT, CBZ	Focal seizures	(Langlois et al., 2012; Riban et al., 2002)
Intrahippocampal KA (rat)	PB		Focal seizures that secondarily generalize	(Rattka et al., 2013)
Electrically-induced SRS	VPA, LTG, VGB, CZP	PB, PHT	Focal seizures that secondarily generalize	(Bethmann et al., 2007)
TBI		CBZ, carisbamate	Focal seizures	(Eastman et al., 2011)
TBI		CBZ, VPA	Spreading seizures	(Eastman et al., 2010)

ASD, anti-seizure drug; SRS, spontaneous recurrent seizures; KA, kainic acid; PILO, pilocarpine; TBI, traumatic brain injury; EZG, ezogabine; LTG, lamotrigine; CBZ, carbamazepine; VPA, valproic acid; LEV, levetiracetam; TPM, topiramate; TGB, tiagabine; PHT, phenytoin; PB, phenobarbital; FBM, felbamate; PGB, pregabalin; DZP, diazepam, CZP, clonazepam; VGB, vigabatrin; GBP, gabapentin; ESM, ethosuximide;

* unpublished ASP data;

[§] effective for only one week

Table 2
Pharmacological properties of *in vitro* models of drug resistance

	Pharmacosensitivity		Comments	References
	Sensitive to ASD	Resistant to ASD		
Human resected brain slice		CBZ	Slices prepared from resected tissue of CBZ resistant patients	(Jandova et al., 2006; Remy et al., 2003)
Brain slice from KA treated rat - low Mg ²⁺	EZG	PHT, TPM, CBZ	Recurrent bursting	(Smith et al., 2007)
Low Mg ²⁺ brain slice	EZG	PHT, TPM, VPA, PB	Recurrent bursting	(Wahab et al., 2010a; Zhang et al., 1995)
Organotypic hippocampal slice culture		PHT, CBZ, VPA, PB, DZP, CZP	Seizure-like events (SLEs)	(Albus et al., 2008; Wahab et al., 2010b)

ASD, anti-seizure drug; KA, kainic acid; EZG, ezogabine; CBZ, carbamazepine; VPA, valproic acid; TPM, topiramate; PHT, phenytoin; PB, phenobarbital; DZP, diazepam, CZP, clonazepam