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Authors

Groth, Caroline P
Banerjee, Sudipto
Ramachandran, Gurumurthy
et al.

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RESEARCH ARTICLE

Multivariate left-censored Bayesian modeling for predicting exposure using multiple chemical predictors

Caroline P. Groth¹ | Sudipto Banerjee² | Gurumurthy Ramachandran³ |
Mark R. Stenzel⁴ | Patricia A. Stewart⁵

¹Department of Preventive Medicine, Feinberg School of Medicine, Northwestern University, Chicago, IL 60611, U.S.A.

²Department of Biostatistics, University of California, Los Angeles, Los Angeles, CA 90095, U.S.A.

³Department of Environmental Health and Engineering, Bloomberg School of Public Health, Johns Hopkins University, Baltimore, MD 21205, U.S.A.

⁴Exposure Assessments Applications, LLC, Arlington, VA 22207, U.S.A.

⁵Stewart Exposure Assessments, LLC, Arlington, VA 22207, U.S.A.

Correspondence

Sudipto Banerjee, Department of Biostatistics, University of California, Los Angeles, Los Angeles, CA 90095, U.S.A.
Email: sudipto@ucla.edu

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Environmental health exposures to airborne chemicals often originate from chemical mixtures. Environmental health professionals may be interested in assessing exposure to one or more of the chemicals in these mixtures, but often, exposure measurement data are not available, either because measurements were not collected/assessed for all exposure scenarios of interest or because some of the measurements were below the analytical methods' limits of detection (i.e., censored). In some cases, based on chemical laws, two or more components may have linear relationships with one another, whether in single or multiple mixtures. Although bivariate analyses can be used if the correlation is high, correlations are often low. To serve this need, this paper develops a multivariate framework for assessing exposure using relationships of the chemicals present in these mixtures. This framework accounts for censored measurements in all chemicals, allowing us to develop unbiased exposure estimates. We assessed our model's performance against simpler models at a variety of censoring levels and assessed our model's 95% coverage. We applied our model to assess vapor exposure from measurements of three chemicals in crude oil taken on the *Ocean Intervention III* during the *Deepwater Horizon* oil spill response and cleanup.

KEYWORDS

chemical mixtures, correlations, Deepwater Horizon oil spill, exposure assessment

1 | INTRODUCTION

Frequently, vapor emissions from multiple sources contribute to workers' overall airborne exposure. When the sources are mixtures with varying composition, they may share common chemical components that are correlated. In other situations, only a single mixture may be present (either as a single source or from multiple sources, and the temperature of the mixture varies between sources over time, such that these are actually different mixtures), with its components having high or low correlations. For brevity, in the remainder of this paper, this second situation will not be referred to in the text below but should be considered to have the same characteristics as multiple mixtures.

In either instance, some chemicals may have been measured in only some of the groups of interest, resulting in missing data, or measurement results may be below the analytical method's limit of detection (LOD, described as left censored), adding uncertainty about the exposure of interest. A LOD is a threshold below which exposure cannot be quantified with a given analytical method to the degree of accuracy and precision deemed adequate to estimate exposure. Measurements below the LOD cannot be distinguished from each other or quantified. Because exposure assessors aim to provide exposure estimates that are as unbiased as possible, these measurements below the LOD must be considered. Not including measurements below the LOD can result in a biased estimate that overestimates exposure metrics typically used in epidemiology studies, such as the arithmetic average (AM) or geometric mean (GM), which can lead to an underestimation of the disease risk.

Assessing exposures from mixtures with missing data is dependent on the correlation of the components. If there is a high correlation between two components of one or more mixtures, one may use a simple linear regression between the two highly correlated components to estimate exposure when the measurements of one component are missing for some scenarios while accounting for censoring (Groth et al., 2017). If the correlation among components is low, however, the relationships of two or more components in measured scenarios can help us estimate exposure to a third component that is missing information or is highly censored. In particular, in the presence of multiple mixtures, one source may allow for the strong prediction of the chemical of interest using a chemical $X_{(1)}$ where another source would use a chemical predictor $X_{(2)}$. As a result, a multiple linear regression framework with both chemicals as predictors may be useful.

To account for multiple mixtures of common chemicals, this paper extends the work of Groth et al. (2017) to a multiple linear regression framework for the estimation of a chemical of interest while accounting for censoring in multiple predictors (\mathbf{X}) and in the primary chemical response of interest (Y). Due to the multivariate framework (which allows us to derive a multiple linear regression equation for each chemical using the other chemicals in the model as predictors), this model also allows us obtain exposure estimates for both the chemical of interest and the chemical predictors simultaneously.

In the next section, we briefly discuss the chemical and the statistical backgrounds of censored data methods in environmental health. Next, we describe a multivariate model to predict low correlated components of multiple mixtures, as well as a method for comparing it with other models with different numbers of predictors. Then, we discuss the results of several simulations that compare our model with simpler models with fewer chemical covariates and that test our model's coverage, that is, the ability to encompass the true parameter values (the values we set for the data set) in the model's 95% credible intervals (CIs). Finally, we conclude with an example using air measurement data collected from a vessel that participated in the *Deepwater Horizon* oil spill response and cleanup efforts, the *Ocean Intervention III*.

2 | CHEMICAL AND STATISTICAL BACKGROUND

When assessing exposures in a workplace, particularly in an epidemiologic study, measurements may be missing or non-detectable (due to levels below the LOD) for some workers of interest for a particular chemical. These workers, however, may have been measured for other exposures. If there is a relationship between the observed measurements, it may be possible to use that knowledge to inform the exposure of interest of the measured workers. In this paper, we assume that the primary interest is assessing exposure to one chemical, using other exposures experienced at the time.

The bases for linear relationships of air concentrations among the components of mixtures such as oil are the Ideal Gas Law, Raoult's Law, and Henry's Law (Stenzel & Arnold, 2015). The vapor concentration (VC) of a pure chemical in the air above the chemical's liquid surface at a specific liquid temperature is the ratio of the chemical's vapor pressure (VP) divided by atmospheric pressure. With mixtures, the VP of each component in the mixture is lower than if it were the pure chemical. The degree of lowering is proportional to the chemical's molar concentration in the mixture. This lower VP is called the chemical's adjusted VP. Once the adjusted VP is determined, it can be divided by atmospheric pressure to estimate the VC of the chemical in the air above the mixture surface. If the composition of the mixture (in mass percentage) is constant, the relative VCs of the components of the mixture will be constant (Stenzel & Arnold, 2015). This concept then is the basis for estimating unmeasured exposures using correlational analyses (linear relationships).

Workplaces can be very complex and contain multiple mixtures with common components. It is seldom possible or practical to measure a significant portion of exposure scenarios that exist in a workplace; therefore, decisions regarding exposure must be made with data collected on only a fraction of the possible exposure scenarios. Industrial hygienists have simplified this complexity and the lack of measurements by developing exposure groups. The definition of an exposure group depends on the purpose of the exposure assessment, but generally, it is a group of individuals who are exposed to the same inventory of chemicals and perform the same tasks or activities under similar exposure conditions at a comparable frequency and duration, resulting in a similar exposure distribution in each chemical.

In this work, we focus on estimating exposure in the a situation characterized by multiple mixtures containing some common chemicals at different concentrations. Because the components are at different concentrations in the various mixtures, they are not likely to be highly correlated in the air. If the workplace is under “pseudo”-equilibrium conditions, multivariate correlation among the components from the multiple sources is expected to be observed. Therefore, this approach can be used to estimate exposures to the chemicals from these multiple mixtures, either when the mixtures are present in exposure groups that were not measured or when one of the chemicals is more highly censored than the others (Stenzel & Arnold, 2015).

2.1 | Limits of detection

An analytical method's inability to quantify exposures below a measurement's LODs provides uncertainty to exposure estimation. LODs vary based on the duration of the measurement and the chemical's analytical method. In our setting, values of the LOD are known and observable (rather than absent and thus not observable).

Scientifically, it is important to account for these values below LOD. It is possible that exposures below the LOD may be associated with detrimental health effects that cannot be investigated because we do not know the concentration of the exposure below the LOD. By accounting for censored values, we may be better able to investigate if such an association exists. In addition, low exposures may have additive or synergistic effects with other exposures experienced simultaneously. Furthermore, often, exposure-based health effect studies quantify exposure for a low exposed or unexposed reference group to which higher exposed groups are compared. This reference group may be more likely to have measurements below the LOD. Perhaps most importantly, omitting values below the LOD may lead to a bias of the mean estimate. By not including values below the LOD, the mean estimate (AM or GM) is likely to be higher than was actually experienced because the lower values are not included in the calculation of the mean to pull the mean estimate down to its true value. Furthermore, as described by Groth et al. (2017), one may come to fundamentally different statistical conclusions if censored data are ignored compared with when they are included in an analysis.

2.2 | Left-censored statistical methods

Scientists have proposed a variety of different statistical methods to account for left-censored variables. For occupational health studies with left censoring, Huynh et al. (2014) compared several classical approaches including β -substitution, reverse Kaplan–Meier, and maximum likelihood. Of these approaches, the β -substitution method generally had the lowest relative root mean squared error and bias. However, Huynh et al. (2016) suggested that Bayesian models have advantages over classical methods such as β -substitution (Ganser & Hewett, 2010). Huynh et al. (2016) concluded that while the β -substitution method performed similarly to Bayesian methods (for bias and root mean squared error), Bayesian models provided variance estimates (important for uncertainty characterization), and formulas for calculating confidence intervals on the imputed β were not available for the β -substitution method.

Groth et al. (2017) expanded the results of Huynh et al. (2016), developing a method for regression where a predictor, a response, or both may be censored. When analyzing data from the *Deepwater Horizon* oil spill response and cleanup efforts, Groth et al. (2017) found that linear relationships between total hydrocarbons (THC) and xylene (one of the BTEXH chemicals; BTEXH stands for benzene, toluene, ethylbenzene, xylene, and *n*-hexane), as measured using dosimeters worn by workers, were present in many groups of workers on the *Development Driller III* from May 15 to July 15, 2010. As indicated above (Section 2), chemical laws suggest that the relationship between the exposures to THC and each of the BTEXH chemicals should be linear in nature if there is a single source, such as crude oil. The *Development Driller III* was in the immediate area of the leaking wellhead, and crude oil was being continuously released, so it was expected that crude oil was the primary source of exposure and a “pseudo”-equilibrium existed. That is, as the volatiles in the crude oil evaporated, new crude oil surfaced to provide a new supply of volatiles. Using a hierarchical Bayesian structure, Groth et al. (2017) estimated the exposure of multiple groups of workers simultaneously to each of the BTEXH chemicals using THC as the chemical predictor. This method allowed groups with higher censoring levels or lower sample sizes to gain inference from the overall relationship in the data set. This paper focuses on extending this method to allow for more than one chemical predictor with values below the LOD.

2.3 | Multivariate modeling

Another benefit of our modeling structure is that it allows us to assess exposure to multiple chemicals through one model. If it is known that several chemicals could have linear relationships among themselves, we may gain a significant amount of knowledge from modeling all chemicals and their correlations into one modeling framework. The multivariate

framework allows us to model simultaneously correlations between all chemicals included in the model. As a result, multiple linear regression equations for each chemical in the model can be derived, along with estimates of exposure to all chemicals in the model. This structure avoids the need to fit additional models to assess exposure to a chemical already included in the model.

However, using a single multivariate model to develop exposure estimates for all chemicals in the model may not be useful in all scenarios. For example, for one chemical, it may be appropriate to include two particular chemicals as predictors. However, in another model of one of the chemical predictors, other chemicals not in the original model may be useful in predicting exposure. Therefore, in this paper, we assume we are primarily interested in a chemical Y given the predictors (\mathbf{X}). However, if the multivariate model is structured in a way such that the groups of chemicals are good predictors of each other, one could use this model to generate exposure estimates for multiple chemicals simultaneously.

2.4 | Multicollinearity

In this paper, we seek to use multiple predictors that are correlated with one another to estimate exposures. This situation, however, presents a potential problem because our model is parameterized based on conditional relationships and regression expressions. The inclusion of multiple highly correlated predictors in a linear regression setting creates a commonly known problem called *multicollinearity* (also called collinearity). High correlations between at least two predictors can lead to misleading regression estimates and inflated standard errors. Inflated standard errors are a significant problem as they may lead us to conclude that the linear relationships are not significant by inflating slope estimates. Similarly, the inflated standard errors are likely to inflate the uncertainty in the mean posterior distribution. Dormann et al. (2013) found that variables with greater than 0.7 correlation will likely create problems with statistical inference if included as predictors in a model. While 0.7 is the generally accepted threshold, others argue that researchers should try to avoid correlations above 0.5 if possible (Dormann et al., 2013; Kutner, Nachtsheim, & Neter, 2004).

To further understand how multicollinearity may influence our model, we performed a simulation study described in our simulation study section. For more information on multicollinearity, see Dormann et al. (2013) and Kutner et al. (2004).

3 | STATISTICAL METHODS

We will assume that we are primarily interested in assessing exposure estimates to a single chemical Y given several other chemicals (\mathbf{X}). To build our model, we start with a simple linear Bayesian regression framework. Let Y_i be the natural log of the primary chemical of interest and $X_{i(1)}$ be the natural log of the first predictor of interest for observation i with $i = 1, \dots, N$. Later on, we will add multiple chemical covariates, so we denote this first chemical covariate with a subscript (1). Distribution of air measurements in the occupational setting are lognormal. We assume that $X_{i(1)}$ follows a normal distribution with mean $\beta_{0(1)}$ and variance $\sigma_{(1)}^2$. Let the regression coefficient vector $\beta_{(Y|X)}$ be the slope ($\beta_{1,(Y|X)}$) and intercept ($\beta_{0,(Y|X)}$) for $Y|X_{(1)}$. Therefore, we assume that Y_i follows a normal distribution with mean $\beta_{0,(Y|X)} + \beta_{1,(Y|X)}X_{i(1)}$ and conditional variance $\sigma_{(Y|X)}^2$. To this structure, we place priors on the variance components ($\sigma_{(Y|X)}^2, \sigma_{(1)}^2$) and mean components ($\beta_{0(1)}, \beta_{(Y|X)}$). With traditional definitions of inverse gamma ($IG()$) and normal ($N()$) distributions as described by Gelman, Carlin, Stern, and Rubin (2013), we arrive at the following joint distribution:

$$IG\left(\sigma_{Y|X}^2 | a, b\right) \times IG\left(\sigma_{(1)}^2 | c, d\right) \times N\left(\beta_{0(1)} | \theta_\mu, \sigma_\mu^2\right) \times N\left(\beta_{(Y|X)} | \mu_\beta, \mathbf{V}_\beta\right) \\ \times \prod_{i=1}^N N\left(Y_i | \beta_{0,(Y|X)} + \beta_{1,(Y|X)}X_{i(1)}, \sigma_{Y|X}^2\right) \times \prod_{i=1}^N N\left(X_{i(1)} | \beta_{0(1)}, \sigma_{(1)}^2\right). \quad (1)$$

In this framework, we used inverse gamma priors ($IG()$) on the variance components and normal priors on the mean components. Other priors are possible, and the above priors are used to allow for weakly informative priors and conjugate relationships to be used. In the above model, we also specify the values mean θ_μ and variance σ_μ^2 in the prior for $\beta_{0(1)}$. Finally, we specify the values of the mean (μ_β) and variance–covariance matrix (\mathbf{V}_β) for our prior on the regression coefficients $\beta_{(Y|X)}$. Alternatively, one may set hyperpriors on this mean and variance–covariance matrix for $\beta_{(Y|X)}$ to allow these parameters to be estimated. It may be of particular interest to estimate these parameters for more than one exposure group. However, for simplicity, we focus on modeling one group's exposures.

To account for censored observations in both the response and predictor, we adjust the above model's likelihood. Let $\text{LOD}_i(X_{(1)})$ be the LOD on the natural log (ln) scale for the i th observation for chemical predictor $X_{(1)}$ and $\text{LOD}_i(Y)$ be the LOD for the i th observation of Y . These LODs are fixed and known prior to modeling and supplied to the model as data.

Then, we separate the N observations for each variable into censored and observed sets. Define $C_{(1)} = \{i : X_{i(1)} \leq \text{LOD}_i(X_{(1)})\}$ and $C_{(Y)} = \{i : Y_i \leq \text{LOD}_i(Y)\}$ as the censored sets of observations of $X_{i(1)}$ and Y , respectively. Let $O_{(1)}$ and $O_{(Y)}$ be the observed values. Using the same definitions of parameters and variables defined in (1), the resulting joint distribution of all model parameters including the censored values is as follows:

$$\begin{aligned}
& IG\left(\sigma_{Y|X}^2 | a, b\right) \times IG\left(\sigma_{(1)}^2 | c, d\right) \times N\left(\beta_{0,(1)} | \theta_\mu, \sigma_\mu^2\right) \times N\left(\beta_{(Y|X)} | \mu_\beta, \mathbf{V}_\beta\right) \\
& \times \prod_{i \in O_{(Y)}} N\left(Y_i | \left(\beta_{0,(Y|X)} + \beta_{1,(Y|X)} X_{i(1)}\right), \sigma_{Y|X}^2\right) \\
& \times \prod_{i \in C_{(Y)}} \Phi\left(\frac{\text{LOD}_i(Y) - \left(\beta_{0,(Y|X)} + \beta_{1,(Y|X)} X_{i(1)}\right)}{\sigma_{Y|X}}\right) \\
& \times \prod_{i \in O_{(1)}} N\left(X_{i(1)} | \beta_{0,(1)}, \sigma_{(1)}^2\right) \times \prod_{i \in C_{(1)}} \Phi\left(\frac{\text{LOD}_i(X_{(1)}) - \beta_{0,(1)}}{\sigma_{(1)}}\right), \tag{2}
\end{aligned}$$

where $\Phi(u)$ denotes the cumulative density function of a standard normal random variable at u .

We extend the bivariate setting described by Groth et al. (2017) by adding more covariates with potential censoring. Our ultimate goal is to model a chemical response Y that is dependent on all chemical predictors (\mathbf{X}). We will consider p chemical predictors (which are arbitrarily called $X_{(1)}$, $X_{(2)}$, etc.). Let $X_{(k)}$ represent one of the chemical predictors of interest (natural log scale) where $k = 1, \dots, p$. We model each additional $X_{(k)}$, where $k = 2, \dots, p$, as conditional on the set of all $X_{(l)}$, where $l = 1, \dots, k-1$, to generate a multivariate model framework. Due to conditional distributions, this model can also be written using a multivariate normal distribution framework.

To adjust the likelihood for multiple chemical covariates when modeling Y , we divide our likelihood into three likelihood components that are multiplied to form the full likelihood of this multivariate model. The first component is the likelihood of our chemical response Y , which is dependent on all chemical predictors \mathbf{X} . Like in Equations 1 and 2, let $\beta_{(Y|X)}$ and $\sigma_{Y|X}^2$ represent the vector of regression coefficients and the conditional variance, respectively, for the conditional expression of $Y | \mathbf{X}$, where we now have multiple X components. To simplify the notation of the conditional means, let $\mathbf{D}_{i,(Y|X)}$ represent the row of the design matrix for the i th observation for Y . Here, the design matrix would consist of a column of 1s for the intercept and $X_{(1)}, \dots, X_{(k)}$ in columns. Therefore, we can abbreviate the mean expression for $Y_i | \mathbf{X}_i$ ($\beta_{0,(Y|X)} + \beta_{1,(Y|X)} X_{i(1)} + \dots + \beta_{p,(Y|X)} X_{i(p)}$) as $\mathbf{D}_{i,(Y|X)} \beta_{(Y|X)}$. To account for censored Y values, this likelihood of Y will contain a censored set $C_{(Y)}$ (dependent on the $\text{LOD}_i(Y)$) and observed set $O_{(Y)}$ as previously defined in equation (2). The second component is the likelihood of all chemical predictors other than $X_{(1)}$. Here, each chemical $X_{(k)}$ is modeled as conditional on $X_{(1)}$ and other chemical predictors $X_{(l)}$ where $l = 2, \dots, k-1$. Let $\beta_{(k)}$ represent the vector of all regression coefficients for the conditional expression on $X_{(k)}$. Like we did for Y_i , we let $\mathbf{D}_{i,(k)}$ represent the row of the design matrix for the i th observation for $X_{(k)}$. The design matrix for $X_{(k)}$ is a column of 1s followed by columns of each $X_{(l)}$ where $l = 1, \dots, k-1$. This allows us to abbreviate the conditional mean of $X_{i,(k)}$ as $\mathbf{D}_{i,(k)} \beta_{(k)}$. Then, we define the conditional variance of $X_{(k)} | X_{(1)}, \dots, X_{(k-1)}$ as $\sigma_{(k|1, \dots, k-1)}^2$. Again as in (2), we consider a censored set $C_{(k)}$ (with $\text{LOD}_i(X_{(k)})$) and observed set $O_{(k)}$ in the likelihoods of each $X_{(k)}$. Finally, the third component is the likelihood of $X_{(1)}$ that is dependent on its mean $\beta_{(1)}$ and variance $\sigma_{(1)}^2$ as previously defined in Equations 1 and 2. In order to account for censoring, we use the previous definitions of the observed set $O_{(1)}$, the censored set $C_{(1)}$, and $\text{LOD}_i(X_{(1)})$ from (2).

From these likelihood components, we obtain the following joint distribution of all model parameters and censored values:

$$\begin{aligned}
& N\left(\beta_{(1)} | \mu_{\beta,(1)}, \gamma_{\beta,(1)}\right) \times IG\left(\sigma_{(1)}^2 | a_{(1)}, b_{(1)}\right) \times N_{(p+1)}\left(\beta_{(Y|X)} | \mu_{\beta,(Y|X)}, \mathbf{V}_{\beta,(Y|X)}\right) \\
& \times IG\left(\sigma_{(Y|X)}^2 | a_{(Y)}, b_{(Y)}\right) \times \prod_{k=2}^p IG\left(\sigma_{(k|1, \dots, k-1)}^2 | a_{(k)}, b_{(k)}\right) \times \prod_{k=2}^p N_k\left(\beta_{(k)} | \mu_{\beta,(k)}, \mathbf{V}_{\beta,(k)}\right) \\
& \times \prod_{i \in O_{(Y)}} N\left(Y_i | \mathbf{D}_{i,(Y|X)} \beta_{(Y|X)}, \sigma_{(Y|X)}^2\right) \times \prod_{i \in C_{(Y)}} \Phi\left(\frac{\text{LOD}_i(Y) - \mathbf{D}_{i,(Y|X)} \beta_{(Y|X)}}{\sigma_{(Y|X)}}\right) \\
& \times \prod_{k=2}^p \prod_{i \in O_{(k)}} N\left(X_{i,(k)} | \mathbf{D}_{i,(k)} \beta_{(k)}, \sigma_{(k|1, \dots, k-1)}^2\right) \times \prod_{k=2}^p \prod_{i \in C_{(k)}} \Phi\left(\frac{\text{LOD}_i(X_{(k)}) - \mathbf{D}_{i,(k)} \beta_{(k)}}{\sigma_{(k|1, \dots, k-1)}}\right) \\
& \times \prod_{i \in O_{(1)}} N\left(X_{i(1)} | \beta_{(1)}, \sigma_{(1)}^2\right) \times \prod_{i \in C_{(1)}} \Phi\left(\frac{\text{LOD}_i(X_{(1)}) - \beta_{(1)}}{\sigma_{(1)}}\right), \tag{3}
\end{aligned}$$

where $\mu_{\beta_{(1)}}$ and $\gamma_{\beta_{(1)}}$ are the mean and variance, respectively, of $\beta_{(1)}$. We let all variances and conditional variances follow inverse gamma distributions with shape parameters $a_{(1)}, a_{(2)}, \dots, a_{(k)}, a_{(Y)}$ and scale parameters $b_{(1)}, b_{(2)}, \dots, b_{(k)}, b_{(Y)}$. In practice, we usually use the same prior for each variance or conditional variance, although the estimated values of each variance are allowed to be different from each other (i.e., we do not assume that all chemicals have the same variance). We define $\mu_{\beta_{(Y|X)}}$ as the mean vector for regression coefficients $\beta_{(Y|X)}$ (of length $p + 1$) and $\mathbf{V}_{\beta_{(Y|X)}}$ as the $(p + 1) \times (p + 1)$ variance–covariance matrix for $\beta_{(Y|X)}$. Finally, we define each $\mu_{\beta_{(k)}}$ as the mean vector for the regression coefficients $\beta_{(k)}$ and $\mathbf{V}_{\beta_{(k)}}$ as the $k \times k$ variance–covariance matrix of $\beta_{(k)}$. For $\beta_{(k)}$, we will consider $k = 2, \dots, p$, and define a separate variance–covariance matrix and mean vector for each k .

In both our simulation and real data analysis, we set shape and scale parameters for all variance terms ($a_{(1)}, \dots, a_{(p)}, b_{(1)}, \dots, b_{(p)}$, or a, b, c , and d in the bivariate framework of Equations (1) or (2)) to 0.01 within our inverse gamma priors on the variances ($\sigma_{(Y|X)}^2, \sigma_{(k|1, \dots, k-1)}^2$ for $k = 2, \dots, p$ and $\sigma_{(1)}^2$). We assume that all regression coefficients are independent with variances of 100,000 (i.e., $\gamma_{\beta_{(1)}}, \mathbf{V}_{\beta_{(k)}}$ with $k = 2, \dots, p$, and $\mathbf{V}_{\beta_{(YX)}}$ are equal to the identity matrix times 100,000). We also assume that the regression coefficients are sampled from the normal distribution with a mean of 0 (i.e., $\mu_{\beta_{(1)}}, \mu_{\beta_{(k)}}$ for $k = 1, \dots, p$, and $\mu_{\beta_{(Y|X)}}$ are set to vectors of 0) for all regression parameters and mean estimates ($\beta_{(1)}, \beta_{(k)}$ where $k = 2, \dots, p$, and $\beta_{(Y|X)}$). These weekly informative prior settings allow the data to drive the inference.

Many possible extensions of (3) exist. For example, if we were interested in modeling more than one group of workers at the same time and in allowing groups with minimal information (e.g., smaller sample sizes and/or high censoring) to be informed by an overall regression relationship, we may add a hierarchical structure and hyperpriors across each set of β coefficients. Furthermore, researchers may be interested in modeling individual level variability if the subjects have more than one measurement. The flexible nature of the Bayesian regression framework allows us easily to add in random effects for the individual to the above model.

3.1 | Posterior inference

In a noncensored setting, all posterior inference of marginal means and variances extends from conditional means and variance formulas. As previously mentioned, this model with p chemical predictors and primary response Y may be represented through multivariate normal framework. Let μ_Y and $\sigma_{(Y)}^2$ be the marginal mean and variance, respectively, of Y . Similarly, for each $X_{(k)}$ with $k = 1, \dots, p$, let $X_{(k)}$ have marginal mean $\mu_{(k)}$ and marginal variance $\sigma_{(k)}^2$. Then, the set of all \mathbf{X} and Y would be a modeled multivariate normal with a $(p + 1)$ length vector of marginal means ($\mu_{(1)}, \mu_{(2)}, \dots, \mu_{(p)}, \mu_{(Y)}$), and a $(p + 1) \times (p + 1)$ variance–covariance matrix. This symmetric variance–covariance matrix would have marginal variances of all X s followed by the marginal variance of Y ($\sigma_{(1)}^2, \sigma_{(2)}^2, \dots, \sigma_{(p)}^2, \sigma_{(Y)}^2$) on the diagonal. Covariances between each set of variables would be found on the off-diagonals.

From this framework, it is easy to derive relationships between conditional and marginal variables. For example, for the response Y , let $\mu_{(Y|X)}$ be the conditional mean of $Y|\mathbf{X}$. Therefore, the marginal mean ($\mu_{(Y)}$) and variance ($\sigma_{(Y)}^2$) for the response Y are defined by the following:

$$\mu_{(Y)} = \mu_{(Y|X)} - \mathbf{B}^T \Sigma_{(X)}^{-1} (\mathbf{X} - \mu_{(X)}) \quad \sigma_{(Y)}^2 = \sigma_{(Y|X)}^2 + \mathbf{B}^T \Sigma_{(X)}^{-1} \mathbf{B}, \quad (4)$$

where \mathbf{B} is a vector of length p of the covariances of Y with each X , and $\Sigma_{(X)}$ is a $p \times p$ variance–covariance matrix with variances of all X s on the diagonal and covariances on the off-diagonals. It should be noted that we can obtain marginal means and variances for each chemical in the model, not just Y . Therefore, we can obtain a multiple linear regression expression for every chemical included in the model (with all other chemicals as predictors). This allows us to estimate exposure to multiple chemicals simultaneously while accounting for correlations among chemicals.

Posterior inference for the parameters of interest in the censored setting uses an overarching Gibbs sampler as described by Gelfand, Smith, and Lee (1992). In the Bayesian setting, we treat the censored values as parameters in our model. Therefore, within each iteration j ($j = 1, \dots, M$), we can first sample these censored values using the full conditionals (with parameters defined by the previous iteration) of each $X_{(k)}$ and Y . This fills in the censored values, giving us a full data set. Second, using the full data set (including the previously censored $X_{(k)}$ values), we can sample the conditional means and variances using full conditional distributions. We repeat these steps within each iteration until convergence of the parameters is met.

Convergence of this model is immediate (well within 5,000 iterations) as assessed by Gelman–Rubin diagnostics, trace plots, and Monte Carlo standard error. To account for additional uncertainty where higher censoring levels exist, we recommend running the chain long enough to allow the model to fully explore the parameter space. In all cases, we provide

25,000 iterations after 5,000 iterations of burn-in to ensure the convergence of our model. A more formal description of the convergence of our model, including assessments of autocorrelation, is provided in the Supporting Information.

Inference for GMs, AMs, and geometric standard deviations (GSDs) stems from the posterior samples of the marginal means and variances. First, through the conditional formulas, we calculate the marginal means and variances for each iteration. Then, we obtain inference on the GM of each chemical by exponentiating the posterior samples of the marginal means previously found for each chemical. The variances reported in the model are for the natural log of each variable. Once marginal standard deviations are found, we simply exponentiate them to find the GSD. The AMs are derived using the posterior samples of the GSD and GM of each variable and calculated using $AM = GM \times e^{\frac{1}{2}(\log(GSD))^2}$.

The above framework can be easily coded in OpenBUGS (<http://openbugs.net/>) or rjags (<https://cran.r-project.org/package=rjags>; also see Plummer, 2014) programming environments for a specific number of chemicals. A sample RJAGS code for three chemicals is provided in the Supporting Information (Plummer, 2014). The Gibbs sampler for any specific number of chemicals written in R described above is also provided in the Supporting Information.

3.2 | Posterior predictive model comparisons: Widely applicable information criterion

In any model, there may be multiple chemicals to consider as chemical covariates. We need a method to decide if we should add an additional chemical covariate to the model. We want to compare the ability of each model (with different number of chemical covariates) to predict the real data provided to the model. This method must account for censoring in any of the predictors, in the chemical response of interest, or in both the predictions and the response.

Watanabe (2010) developed a criteria known as widely applicable information criterion (WAIC) that is based on the likelihood of each observation (i) over a set of iterations j ($j = 1, \dots, M$) that can be used to compare models fit to the same data sets. The use of the likelihood allows us to also account for censored observations. Other methods such as Gelfand and Ghosh's D -statistics does not allow us to account for censored observations because it relies on a statistics that compares points generated from the model to the true data values (Gelfand & Ghosh, 1998). The true censored measurements are not known in our case, so using this statistics would require us to ignore censored response values.

In the multivariate framework, the full likelihood of each observation consists of three primary components as modeled in (3). Again, the components include the likelihood for of $Y | \mathbf{X}$, the likelihoods for each $X_{i,(k)} | X_{i,(k-1)}, \dots, X_{i,(1)}$, and the likelihood of $X_{i,(1)}$. These likelihood components when multiplied form the full likelihood for a particular observation i . As in (3), a censored Y_i would have likelihood $\Phi\left(\frac{LOD_i(Y) - \mathbf{D}_{i,(Y|\mathbf{X})}\beta_{(Y|\mathbf{X})}}{\sigma_{Y|\mathbf{X}}}\right)$, whereas observed values in set O_Y would have likelihood $N(Y_i | \mathbf{D}_{i,(Y|\mathbf{X})}\beta_{(Y|\mathbf{X})}, \sigma_{Y|\mathbf{X}}^2)$. Similarly, the likelihood of a censored $X_{i,(k)}$ for any $k, k = 2, \dots, p$, would be $\Phi\left(\frac{LOD_i(X_p) - \mathbf{D}_{i,(k)}\beta_{(k)}}{\sigma_{k|k-1, \dots, 1}}\right)$. The likelihood for an observed $X_{i,(k)}$ would be $N(X_{i,(k)} | \mathbf{D}_{i,(k)}\beta_{(k)}, \sigma_{(k|k-1, \dots, 1)}^2)$. Finally, the likelihood for a censored $X_{i,(1)}$ is $\Phi\left(\frac{LOD_i(X_1) - \beta_{0,(1)}}{\sigma_1}\right)$, whereas an observed $X_{i,(1)}$ would have likelihood $N(X_{i,(1)} | \beta_{0,(1)}, \sigma_{(1)}^2)$.

The full likelihood is calculated using parameter estimates at each iteration j , where M is the total iterations sampled after burn-in (the number of posterior samples of each parameter). These parameter estimates include censored \mathbf{X}_i , regression coefficients ($\beta_{0,(1)}, \beta_{0,(2)}$ etc.), and variances ($\sigma_{(1)}^2, \sigma_{(2|1)}^2, \dots, \sigma_{(p|p-1, \dots, 1)}^2, \sigma_{(Y|\mathbf{X})}^2$). A censored chemical measurement $X_{i,(k)}$ may depend on likelihoods of other X_i for observation i , as well as other parameters.

3.2.1 | WAIC of full likelihood

WAIC involves two statistics that rely on the likelihoods, known as $LPPD$, or the log posterior predictive density, and P , the penalty term. The quantity $LPPD$ can be thought of as the goodness-of-fit term. P is the penalty for more model parameters and greater variation in the likelihoods. Let L_{ij} be the full likelihood formed using the three components previously defined for a particular measurement i calculated using parameter estimates at iteration j . Then, $LPPD$ and P are defined as follows:

$$LPPD = \sum_{i=1}^N \log\left(\frac{1}{M} \sum_{j=1}^M L_{ij}\right) \quad P = \sum_{i=1}^N \text{Var}(\log(L_{ij})), \quad (5)$$

where we provide here the penalty term (which we call P_2 to distinguish it with the penalty defined by Watanabe, 2010) recommended by Gelman, Hwang, and Vehtari (2014). The variance is over the M iterations for each observation i . The penalty developed by Watanabe (2010), which we will call P_1 , is defined as $\sum_{i=1}^N 2(\log(\frac{1}{M} \sum_{j=1}^M L_{ij}) - \frac{1}{M} \sum_{j=1}^M \log(L_{ij}))$ but usually is close to that defined by Gelman et al. (2014).

Because we want to maximize the mean likelihood for each observation, we want to maximize $LPPD$. We want to minimize the penalty term P because we want variability in the parameter estimates to be small, leading to lower

variability in the likelihood of each observation. To develop a deviance-like statistics, WAIC is calculated using the expression $WAIC = -2(LPPD - P)$. Lower values of WAIC are preferred (Watanabe, 2010).

In the Supporting Information, we describe an additional type of WAIC, the WAIC of Y . The WAIC of Y may be particularly useful if the primary interest is in modeling the response. See the Supporting Information for more information.

3.2.2 | Model comparisons

Because the components in the likelihood vary slightly between WAIC and $WAIC_Y$ of the full likelihood, we perform two different model comparisons in this paper, one with each WAIC form. WAIC of the full likelihood allows us to identify if the multivariate nature of the model is beneficial for all chemicals included in the model (not just Y), whereas $WAIC_Y$ focuses on if the model is beneficial/preferred when modeling Y only.

In practice, one may want to compare the multivariate model with a bivariate model or analysis of variance (ANOVA) model (here defined as an intercept-only regression model) accounting for censoring (or a combination of models). The calculations of WAIC remain the same, but the likelihood changes for these models. We will assume similar notation for censored and observed values (such as in Equation 2 and 3). For a bivariate model of Y and X , let $\mu_{Y|X_i}$ be the expression for $Y_i | X_i$ (mean expression is $\alpha_0 + \alpha_1 X_i$) where α_0 is an intercept, α_1 is the slope, and X_i is the chemical covariate at observation i . Let $\sigma_{Y|X}^2$ be the variance of $Y | X$ under the bivariate model. Then, the likelihood of a censored Y is $\Phi\left(\frac{LOD_i(Y) - \mu_{Y|X_i}}{\sigma_{Y|X}}\right)$, whereas an observed observation in set O_Y would have likelihood $N(Y_i | \mu_{Y|X_i}, \sigma_{Y|X}^2)$. Then, X would be modeled like the $X_{(1)}$ component above with a mean estimate $\beta_{0,(1)}$ and variance estimate $\sigma_{(1)}^2$. For an ANOVA framework (an intercept-only regression model), μ_Y and σ_Y^2 are the marginal mean and marginal variance of Y , respectively. Then, the likelihood of a censored Y would be $\Phi\left(\frac{LOD_i(Y) - \mu_Y}{\sigma_Y}\right)$, and the observed likelihood of Y would be $N(Y_i | \mu_Y, \sigma_Y^2)$. Modeling other variables as ANOVAs would have similar forms for the likelihood components.

4 | SIMULATION STUDIES

In practice, it is important to assess if additional chemical predictors will add meaningful information. In order to formally test if our multivariate model provides more meaningful information with additional covariates, we performed a series of simulation studies where we compare our multivariate modeling framework with simpler models.

In addition to comparing models, we also tested the multivariate model's 95% coverage and observed the 95% width of various parameters. As part of this simulation, we also investigated how multicollinearity may influence the 95% CI width and coverage rates.

4.1 | Data development

In all simulation studies, we assume that the true model contains three chemicals. The response Y would be best modeled by two chemical covariates. To generate data to fit this modeling scenario, we generated 100 observations (i ; $N = 100$) for each $X_{(1)}$, $X_{(2)}$, and Y . Specifically, we modeled $X_{i,(1)} \stackrel{iid}{\sim} N(2.6, 2.25)$, $X_{i,(2)} \stackrel{iid}{\sim} N(2.75 + 0 * X_{i,(1)}, 0.64)$, and $Y_i \stackrel{iid}{\sim} N(1.25 + 0.35X_{i,(1)} + 0.25X_{i,(2)}, 0.36)$. This particular data set assumes that both $X_{(1)}$ and $X_{(2)}$ contribute significantly to the estimation of Y , but $X_{(1)}$ and $X_{(2)}$ are not correlated to avoid multicollinearity concerns (in our multicollinearity simulation, we change this). We also assume that $X_{(1)}$ contributes slightly more than $X_{(2)}$. The coefficients chosen here were made based on data commonly presented in environmental health studies (variables on natural log scale).

After developing the raw data, we imposed censoring on each variable by censoring all values below particular quantiles of each variable. In this context, censoring values means labeling an indicator as censored and changing the value to missing. We imposed censoring on the raw data set using all possible combinations of 25%, 50%, and 75% censoring for Y , $X_{(1)}$, and $X_{(2)}$. We selected this range of censoring levels to reflect the typical censoring values seen in industrial hygiene. For example, censoring of airborne chemical exposures during the *Deepwater Horizon* oil spill response and cleanup varied from 0 to as much as 100%. We consider each of these data sets with different imposed censoring levels to be a separate modeling scenario.

As indicated earlier, the LOD of a measurement depends on the analytical method and the duration. In practice, measurements are taken over a variety of durations; thus, for a single chemical analyzed by a single method, multiple LODs

may be observed across measurements. We also developed LODs for each chemical by sampling the LODs from a uniform distribution defined by the chemical's censoring status. For a given variable Z at observation i , we determined its censored status. For censored values, we sampled from $Unif(Z_i + 0.1, quantile(Z) + 0.2)$, and for observed values, we sampled from $Unif(min(Z) - 0.1, Z_i - 0.1)$. For censored values, we chose the upper bound to be a value slightly above the quantile of Z (*quantile*: value below which censoring occurs). This allowed LODs to occur roughly in a band below a censoring value, which provides a range of LODs in each chemical (allowing for multiple LODs). Censored values had LODs higher than the original simulated datapoint, whereas observed values had LODs below the real simulated datapoint values. LODs are not used in our model formulation if the values are observed, so this definition of LODs is sufficient for our purposes as long as the LODs are less than the real datapoint values we simulated (and classified as observed).

We also ran a completely noncensored modeling scenario to ensure that our model would be preferred to simpler models under a completely observed data scenario. We maintained the same sampling strategy for LODs here but using only the observed LOD sampling strategy.

4.2 | Model comparison simulation

We have two primary purposes of this modeling framework. First, we are interested if our multivariate model is preferred over simpler model combinations with respect to WAIC for all chemicals included in the model (both Y and all \mathbf{X}). Second, we are interested if the multivariate framework leads to lower $WAIC_Y$ compared with simpler models with fewer covariates. We consider the best model as the model with the lowest WAIC and/or $WAIC_Y$, reflecting that the model is a good fit to the data while not adding unnecessary complexity. To test how well our model meets these two goals, we performed two sets of model comparisons. On our first set of models, we calculated WAIC for the full likelihood including all chemical predictors and responses. On a second set of models, we assessed $WAIC_Y$. The methods and results of the simulation with $WAIC_Y$ are included in the Supporting Information.

4.2.1 | WAIC for full likelihood

To each unique data set, we fit a series of five separate modeling frameworks that each account for censored observations. The first model was the (true) multivariate model as described above (in Section 4.1). We will call this the three-variable model. Frameworks 2–4 involved a bivariate model and an ANOVA (intercept-only regression) that when multiplied allowed us to model all three variables. Framework 2 modeled $X_{(1)}$ and Y using a bivariate model and modeled $X_{(2)}$ using an ANOVA modeling framework. Modeling framework 3 modeled $X_{(2)}$ and Y with a bivariate model and used an ANOVA model for $X_{(1)}$. Modeling framework 4 modeled $X_{(1)}$ and $X_{(2)}$ using a bivariate model and used an ANOVA model for Y . Framework 5 used an ANOVA model for each variable individually and multiplied them to form the full modeling framework. Modeling frameworks 2–5 assume that the models for the different components are independent.

We calculated WAIC for each of these frameworks by calculating the likelihood components (for $X_{(1)}$, $X_{(2)}$, and Y) as described above (Section 3.2).

4.2.2 | Model comparison simulation results

WAIC values for the full likelihood model comparison are shown in Table 1. WAIC values should only be compared with models fit to the same data set, and therefore, WAIC values should only be compared across a given row of Table 1.

To ensure that our model framework was set up correctly, we ran WAIC for the completely observed scenario (percentage censoring of 0). The WAIC values in this scenario indicated that our three-variable model would be preferred as expected and designed. Similarly, results of our model comparison across the scenarios with different percentage censoring levels showed that the three-variable model consistently had the lowest WAIC of the modeling frameworks tested. This result demonstrated that the multivariate model had the best fit given the complexity of the models tested (which matches how we designed the simulation), even under various censoring levels.

As expected, modeling framework 2 (bivariate ($X_{(1)}$, Y) ANOVA($X_{(2)}$)) had the second lowest WAIC across all model scenarios. Modeling frameworks 4 and 5 performed poorly, as expected, because the modeling frameworks failed to account for correlation between either chemical predictor or response Y .

TABLE 1 WAIC statistics for the full likelihood by model type for various data sets with different degrees of censoring in $X_{(1)}$, $X_{(2)}$, and Y

Modeling Framework Percent Censoring			WAIC of Full Likelihood				
			1 Three-Variable Model	2 Bivariate ANOVA	3 Bivariate ANOVA	4 Bivariate ANOVA	5 ANOVAs
$X_{(1)}$	$X_{(2)}$	Y	$(X_{(1)}, X_{(2)}, Y)$	$(X_{(1)}, Y) \times (X_{(2)})$	$(X_{(2)}, Y) \times (X_{(1)})$	$(X_{(1)}, X_{(2)}) \times (Y)$	$(X_{(1)}) \times (X_{(2)}) \times (Y)$
0	0	0	799.3	814.5	865.5	874.3	875.7
25	25	25	762.3	776.2	819.6	828.1	826.3
25	25	50	736.6	749.3	795.0	802.3	800.5
25	25	75	691.4	699.0	735.0	739.5	737.6
25	50	25	737.9	752.3	797.2	803.9	802.4
25	50	50	712.6	725.4	772.5	778.1	776.6
25	50	75	666.1	675.1	711.7	715.3	713.8
25	75	25	688.4	695.3	742.2	747.3	745.5
25	75	50	659.2	668.5	715.9	721.5	719.7
25	75	75	612.0	618.2	655.1	658.7	656.8
50	25	25	715.4	728.0	766.0	774.2	772.6
50	25	50	685.8	695.6	741.3	748.4	746.8
50	25	75	641.8	648.3	681.2	685.6	684.0
50	50	25	691.6	704.1	743.5	749.7	748.7
50	50	50	660.5	671.7	718.8	723.9	722.9
50	50	75	616.5	624.4	658.0	661.0	660.1
50	75	25	641.3	647.2	688.5	693.6	691.8
50	75	50	607.1	614.8	662.3	667.8	666.0
50	75	75	561.9	567.5	601.4	605.0	603.1
75	25	25	619.1	628.7	667.3	675.9	673.8
75	25	50	594.0	601.9	642.5	650.1	648.0
75	25	75	538.1	544.0	582.5	587.2	585.2
75	50	25	593.9	604.8	644.7	651.4	649.9
75	50	50	569.6	578.1	620.0	625.6	624.2
75	50	75	513.4	520.1	559.2	562.8	561.3
75	75	25	540.3	547.8	589.8	594.7	593.0
75	75	50	513.2	521.1	563.5	569.0	567.2
75	75	75	456.9	463.2	502.7	506.1	504.3

Note. WAIC of the full likelihood values are reported using the same model seed. WAIC = widely applicable information criterion; ANOVA = analysis of variance.

4.3 | Coverage and multicollinearity simulation

In order to understand our model's ability to develop parameter estimates that resemble the true parameters we set, we ran a simulation study to estimate the 95% coverage of the regression coefficients for $(\beta_{0,(1)}, \beta_{0,(2)}, \beta_{1,(2)}, \beta_{0,(Y|X)}, \beta_{1,(Y|X)}, \beta_{2,(Y|X)})$ and variances $(\sigma_{(1)}^2, \sigma_{(2|1)}^2, \sigma_{(Y|X)}^2)$. First, we generated 1,000 different data sets for each censored combination (including a scenario with no censored observations), using the characteristics described in Section 4.1 (i.e., $\beta_{0,(1)} = 2.6$, $\beta_{0,(2)} = 2.75$, $\beta_{1,(2)} = 0$, $\beta_{0,(Y|X)} = 1.25$, $\beta_{1,(Y|X)} = 0.35$, $\beta_{2,(Y|X)} = 0.25$, $\sigma_{(1)}^2 = 2.25$, $\sigma_{(2|1)}^2 = 0.64$, and $\sigma_{(Y|X)}^2 = 0.36$). Then, we fit our three-variable multivariate model to each data set and obtained 95% CIs for the regression coefficients and variances. For each of the 1,000 runs, we identified whether the true value of each parameter of interest was contained within the 95% CI. Moreover, 95% coverage was defined as the percentage of these CIs that contained the true parameter value. We repeated this process for each separate data set (with different censored combinations of $X_{(1)}$, $X_{(2)}$, and Y).

4.3.1 | Multicollinearity simulation

To test how our three-variable model responded to the presence of multicollinearity, we repeated the coverage analysis on several different modeling scenarios with different correlations between $X_{(1)}$ and $X_{(2)}$. In the initial simulation study described above, the correlation between $X_{(1)}$ and $X_{(2)}$ was assumed to be 0. In addition to these results, we also ran a series of scenarios using all of the earlier censoring combinations of $X_{(1)}$, $X_{(2)}$, and Y with the correlation between $X_{(1)}$ and $X_{(2)}$ adjusted to 0.25, 0.50, and 0.75. Only the $\beta_{1,(2)}$ was altered to adjust for the different correlations.

For each unique censoring level and correlation level combination, we assessed 95% coverage of the regression coefficients and variances using 1,000 different generated data sets. In addition, we obtained the median posterior 95% CI width of each of the regression coefficients.

4.3.2 | Coverage and multicollinearity simulation results

Coverage probability are provided in Table 2. To show that our model worked correctly, we reported the coverage probability for a scenario with no censored data points (completely observed data). All coverage levels, as expected, were near 95% in all parameters.

Results when censoring was introduced revealed that as censoring levels increased, coverage decreased for the intercepts. As censoring in $X_{(1)}$ increased (holding censoring constant in both $X_{(2)}$ and Y), for example, we saw the coverage of $\beta_{0,(1)}$ decrease. Similarly, as censoring in $X_{(2)}$ increased (holding censoring constant in both $X_{(1)}$ and Y), the coverage for the intercept $\beta_{0,(2)}$ decreased.

Trends in coverage for the regression coefficients associated with Y ($\beta_{(Y|X)}$) are related to the graphical region of the plot of Y and X where the censored points are being estimated. To better understand the influence of the estimated censored values, Figure 1 shows a scatter plot with predictor X and response Y . In this graphic, we assume that the LODs of X and LODs of Y were each at a particular value, represented by the dark solid lines. These lines divide our graphic into four regions, and each region corresponds to a different censored and observed combination of the variables X and Y . For example, the upper right-hand region contains observed X and Y coordinate pairs, whereas the lower left region contains coordinate pairs where both X and Y are censored. From this graphic, it is easy to see that the region where points are estimated will influence the overall regression estimates. For example, if we estimate many censored values for Y where

TABLE 2 Coverage and median 95% credible interval (CI) width for different degrees of censoring in $X_{(1)}$, $X_{(2)}$, and Y when the correlation between $X_{(1)}$ and $X_{(2)}$ is 0.00

Percent Censoring			95% Coverage Probabilities									Median Posterior 95% CI Width					
			Regression Coefficients						Variances			Regression Coefficients					
Y	$X_{(1)}$	$X_{(2)}$	$\beta_{0,(1)}$	$\beta_{0,(2)}$	$\beta_{1,(2)}$	$\beta_{0,(Y X)}$	$\beta_{1,(Y X)}$	$\beta_{2,(Y X)}$	$\sigma^2_{(1)}$	$\sigma^2_{(2 1)}$	$\sigma^2_{(Y X)}$	$\beta_{0,(1)}$	$\beta_{0,(2)}$	$\beta_{1,(2)}$	$\beta_{0,(Y X)}$	$\beta_{1,(Y X)}$	$\beta_{2,(Y X)}$
0	0	0	94.3	95.8	95.3	94.2	94.6	94.3	95.2	94.9	94.0	0.59	0.64	0.22	0.94	0.16	0.29
25	25	25	92.6	95.8	94.9	95.7	95.2	95.4	83.0	90.2	91.7	0.68	0.66	0.22	1.12	0.19	0.33
25	25	50	92.6	89.4	95.5	94.5	95.9	94.6	82.9	47.1	93.0	0.68	0.86	0.28	1.04	0.19	0.31
25	25	75	92.6	40.5	95.5	74.1	95.8	80.0	83.2	1.1	94.6	0.68	1.62	0.47	0.90	0.19	0.27
25	50	25	62.7	95.2	95.2	92.9	87.1	95.9	33.5	89.9	93.8	0.91	0.58	0.20	1.13	0.18	0.34
25	50	50	61.9	86.5	95.2	84.6	86.9	94.5	33.8	48.3	94.0	0.91	0.76	0.25	1.05	0.18	0.32
25	50	75	62.3	27.3	95.9	44.8	86.6	81.2	33.9	1.5	94.9	0.91	1.48	0.42	0.91	0.19	0.28
25	75	25	1.0	95.5	95.2	62.9	40.4	95.1	1.9	91.2	95.8	1.85	0.45	0.18	1.21	0.18	0.37
25	75	50	1.0	78.9	95.0	43.1	39.4	95.6	1.9	50.1	95.0	1.85	0.60	0.22	1.13	0.18	0.35
25	75	75	1.1	7.7	94.1	8.8	41.5	83.2	1.7	1.8	95.3	1.85	1.26	0.36	0.96	0.18	0.31
50	25	25	92.7	95.7	95.2	79.1	82.5	91.8	81.2	89.7	65.5	0.68	0.66	0.22	1.56	0.25	0.43
50	25	50	92.4	89.4	95.6	88.1	82.2	94.8	81.2	46.7	68.9	0.68	0.86	0.28	1.46	0.26	0.40
50	25	75	92.7	39.8	95.1	97.2	84.2	92.6	82.1	1.2	73.5	0.68	1.63	0.47	1.26	0.26	0.35
50	50	25	58.7	94.9	95.6	92.7	96.4	92.5	29.7	89.9	72.4	0.94	0.57	0.20	1.54	0.24	0.43
50	50	50	58.8	85.7	94.3	95.8	97.1	95.7	29.5	47.7	74.8	0.94	0.75	0.25	1.44	0.24	0.40
50	50	75	58.4	25.1	96.3	94.2	96.1	92.3	30.0	1.6	79.7	0.94	1.48	0.42	1.23	0.25	0.35
50	75	25	0.8	95.0	95.1	94.5	84.9	92.4	1.0	90.9	84.6	1.94	0.44	0.17	1.56	0.22	0.46
50	75	50	0.9	77.5	94.9	87.5	84.6	94.2	0.9	50.4	86.2	1.94	0.59	0.22	1.45	0.23	0.43
50	75	75	0.6	4.9	94.8	54.1	85.0	91.7	0.9	1.8	89.2	1.94	1.26	0.36	1.22	0.23	0.37
75	25	25	92.5	95.7	94.9	23.0	40.2	83.8	81.7	89.8	16.8	0.68	0.66	0.22	3.18	0.48	0.73
75	25	50	92.6	89.4	95.1	27.7	40.3	88.3	82.1	47.0	17.9	0.68	0.86	0.28	3.02	0.48	0.68
75	25	75	92.6	39.1	95.1	43.6	43.0	95.5	81.8	1.1	24.3	0.68	1.65	0.47	2.62	0.49	0.58
75	50	25	56.5	95.4	95.7	38.6	59.3	84.3	29.1	90.0	22.2	0.96	0.57	0.20	3.11	0.45	0.74
75	50	50	56.7	85.7	94.8	44.8	59.7	88.6	29.0	46.7	24.0	0.96	0.75	0.25	2.92	0.46	0.69
75	50	75	56.4	24.6	95.8	68.3	61.7	95.6	29.3	1.1	30.5	0.96	1.49	0.43	2.52	0.46	0.58
75	75	25	0.4	95.1	95.0	81.9	95.4	85.6	0.5	90.8	37.5	2.04	0.43	0.17	2.94	0.40	0.77
75	75	50	0.4	77.9	95.3	88.6	95.2	89.9	0.6	50.2	40.2	2.04	0.58	0.22	2.74	0.39	0.72
75	75	75	0.3	4.4	94.6	98.4	95.8	96.2	0.5	1.8	47.7	2.05	1.26	0.36	2.32	0.40	0.61

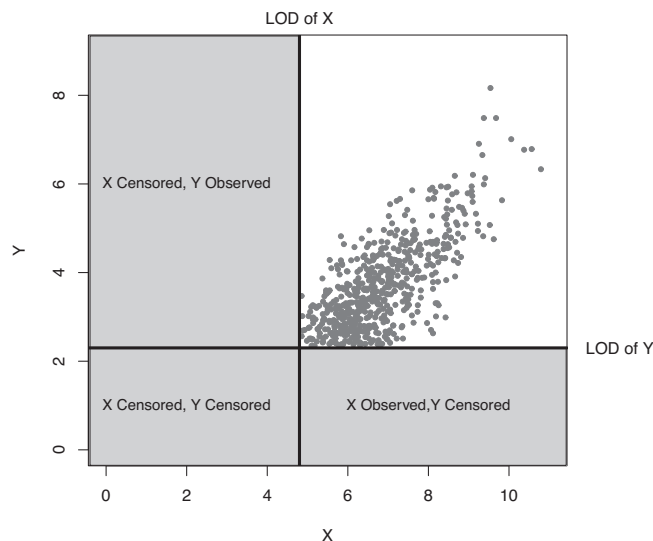


FIGURE 1 Graphical representation of regression with censoring in both X and Y . This graphic demonstrates that knowing the censoring status of both the predictor X and response Y allows us to know the region in which censored data points will be estimated. The locations where points will be estimated are important because it will influence the slope and intercept of the regression line

X is observed (for a range of observed X values), and if we do not estimate many points in other regions, we are likely to see the regression line become flatter.

Coverage of $\beta_{1,(Y|X)}$, the slope coefficient for $X_{(1)}$, depended greatly on the censoring of both $X_{(1)}$ and Y (Table 2). If $X_{(1)}$ and Y had the same censoring levels, coverage of this coefficient was around 95% regardless of the amount of censoring. However, when there were differences in the censoring levels, coverage increasingly dropped the further apart the censoring became. Referring again to Figure 1, if censoring levels were different in X and Y , we would be more likely to estimate points in the region where only $X_{(1)}$ is censored (Y observed) or only Y is censored ($X_{(1)}$ observed) in the coordinate pair. Estimating values in these two regions will be likely pull the regression line (either up or down depending on the region) and therefore lead to regression estimates that differ from the true values. Coverage of $\beta_{2,(Y|X)}$, the slope coefficient for $X_{(2)}$, performed similarly and was dependent on censoring in both $X_{(2)}$ and Y . Because an intercept coefficient changes with any changes in the slope, whenever either of the slopes coverage dropped, we also saw drops in the coverage for $\beta_{0,(Y|X)}$.

The variances also showed reduced coverage at higher censoring levels in the chemical of interest. For example, as censoring increased in $X_{(2)}$, the coverage of $\sigma_{(2|1)}^2$ decreased. This result is expected because we expect greater uncertainty when we have to estimate more censored values. All median posterior estimates of the variances when CIs did not contain the true variance were higher than the true variance (not shown). The conditional variance for $Y|X$ did not decrease as quickly. This result is likely related to information being provided from each X , and therefore, this statistics is partially dependent on censoring in each X .

The coverage for the slope of $X_{(1)}$ in the equation for $X_{(2)}$ ($\beta_{1,(2)}$) remained relatively constant regardless of censoring. The slope we used to generate the original data was a slope of 0. As censoring increased, the CI width became larger (due to increased variances). When the censoring differed, resulting in one region in Figure 1 to have more influence on the slope than the other region, the center of the CI changed and the width of the CI increased, leading us to still capture 0 most of the time. This led to high coverage.

While coverage may suffer greatly at high censoring levels, we do not necessarily expect the estimates from the model to be close to the truth with high levels of censoring. With increased censoring, we fundamentally change the data set and introduce additional parameters (the censored values) we need to estimate. With each censored value, we also introduce an LOD. These LODs greatly limit the values that can be estimated and may lead to changes in the model parameters. Therefore, our CIs will be unable to capture the original true model parameters.

Tables of the 95% CI width and coverage probabilities for different correlation levels between $X_{(1)}$ and $X_{(2)}$ are provided in the Supporting Information. Results indicate that the 95% CI width for the regression coefficients of $\beta_{1,(Y|X)}$ and $\beta_{2,(Y|X)}$ inflated as the correlation increased. While censoring did increase the width of the 95% CI, the increase in the width of the 95% CI from multicollinearity (increasing the correlation between $X_{(1)}$ and $X_{(2)}$) did not appear to be moderated by censoring level. Coverage probabilities were not dramatically different between correlation levels.

TABLE 3 Characteristics of *Ocean Intervention III* measurements ($N = 60$) from July 16 to September 30

Chemical	Count		Percent			Noncensored Correlations		
	Noncensored	Censored	Hexane	Toluene	Xylene	Hexane	Toluene	Xylene
Hexane (Y)	53	12		0.42 ($N = 40$)	0.25 ($N = 44$)			
Toluene	45	25			0.01 ($N = 38$)			
Xylene	48	20						

Note. We report count, percentage censoring, and noncensored correlations between hexane, toluene, and xylene.

5 | PRELIMINARY ANALYSIS: DEEPWATER HORIZON OIL SPILL RESPONSE AND CLEANUP EFFORTS

Approximately 5 million barrels of oil were released into the Gulf of Mexico as a result of the *Deepwater Horizon* oil spill. In the months following the spill, over 50,000 workers were involved in the response and cleanup efforts. Crude oil releases potentially harmful chemicals into the air including the THC group of chemicals, specifically the BTEXH chemicals. In this illustration, crude oil is expected to be the primary source of exposure to these chemicals. As a part of the National Institutes for Environmental Health Science's Gulf Longterm Follow-up Study (GuLF STUDY), researchers are working to quantify several of these airborne chemical exposures for these workers and to investigate if any association exists with detrimental health outcomes in this population (Kwok et al., 2017).

In this data example, we are interested in developing a model to estimate hexane exposure from July 16, 2010 to September 30, 2010 on the *Ocean Intervention III*, a vessel involved in the response and cleanup effort by deploying a remotely operated vehicle. One of the BTEXH chemicals is *n*-hexane (called hexane from now on), which is important to estimate because hexane has been associated with neuropathology (Ritchie et al., 2001). Furthermore, in the GuLF STUDY, hexane was not always measured for many scenarios that were measured for THC and benzene, toluene, ethylbenzene, and xylene (BTEX), so its exposure level needs to be approximated using these other chemicals. Also of interest are exposures for toluene and xylene, as these chemicals have also been associated with detrimental health effects (U.S. Department of Health and Human Services, 2017, 2007). Although crude oil is a single mixture, because of evaporation of the volatile components over time, it is actually several mixtures, thus meeting our criteria of multiple mixtures.

This data set consists of sixty 4- to 18-h air measurements that were collected using passive dosimeters worn by workers who worked outside of the vessel (compared with the living quarters inside). Each sample was analyzed for a variety of chemicals including THC, BTEX, and hexane, in some cases. Censoring in this sample for hexane, toluene, and xylene was generally low, with censoring levels $\leq 25\%$ (Table 3).

We performed two model comparisons using the two forms of WAIC as defined above (Section 3.2). Under model comparison 1, we compared model fit for all three chemicals (toluene, xylene, and hexane), using WAIC for the full likelihood. For model comparison 2, we compared model fit for hexane only using WAIC_Y. Results of model comparison 2 are included in the online Supporting Information.

5.1 | Model comparison 1

First, we assessed how well our model fit the toluene, xylene, and hexane data accounting for complexity. As in the simulation study (Section 4), framework 1 was the multivariate model, and frameworks 2–4 used a bivariate model multiplied by an ANOVA model. In framework 2, we modeled hexane and toluene using a bivariate model and modeled xylene using an ANOVA model. In framework 3, we modeled hexane and xylene using a bivariate model and used an ANOVA model for toluene. In framework 4, we modeled xylene and toluene using a bivariate model and used an ANOVA model for hexane. Framework 5 used separate ANOVAs for hexane, toluene, and xylene and assumed that these chemicals were not correlated with each other (overall once accounting for censoring). Models were compared using WAIC for the full likelihood (which includes likelihood components for toluene, xylene, and hexane). Variability in WAIC was assessed by changing model seeds and doing 100 runs for each model type.

5.2 | General results

Results of our first model comparison are shown in Table 4. These results revealed that the three-variable model (the multivariate model) had the lowest WAIC of the modeling frameworks tested. An assessment of variability of WAIC

TABLE 4 Model comparison for the *Ocean Intervention III* data

Modeling Framework	Model Types	<i>LPPD</i> Estimate	P_2 Estimate	WAIC (5%, 95%)	
1	Three-Variable (Hexane, Toluene, Xylene)	-277.7	9.2	573.9	(573.9, 574.2)
2	Bivariate \times ANOVA (Hexane, Toluene) \times (Xylene)	-280.7	7.0	575.3	(575.0, 575.2)
3	Bivariate \times ANOVA (Hexane, Xylene) \times (Toluene)	-283.7	6.9	581.3	(581.1, 581.3)
4	Bivariate \times ANOVA (Toluene, Xylene) \times (Hexane)	-285.7	6.3	584.0	(583.8, 584.0)
5	ANOVAs (Hexane) \times (Xylene) \times (Toluene)	-286.7	5.2	583.7	(583.6, 583.7)

Note. Comparing widely applicable information criterion (WAIC) with full likelihood values for modeling frameworks 1–5. WAIC estimates are reported for the same model seed of the program. The 5th and 95th percentiles of 100 runs with different model seeds are reported for WAIC. *LPPD* = log posterior predictive density; ANOVA = analysis of variance.

TABLE 5 Exposure estimates and parameter estimates from the three-variable multivariate model for samples taken outside on the *Ocean Intervention III* from July 16, 2010 to September 30, 2010

	Parameter	Median	2.5 Quantile	97.5 Quantile
Geometric Mean (ppb)	Hexane	9.66	7.08	13.03
	Toluene	9.16	5.29	15.34
	Xylene	10.89	7.73	15.01
Geometric Standard Deviations	Hexane	3.21	2.63	4.20
	Toluene	7.16	4.98	11.93
	Xylene	3.48	2.77	4.82
Arithmetic Means (ppb)	Hexane	19.06	13.47	30.51
	Toluene	63.72	29.87	209.35
	Xylene	23.82	16.30	40.59
Correlations	$\rho(\ln(\text{Hexane}), \ln(\text{Toluene}))$	0.41	0.17	0.61
	$\rho(\ln(\text{Hexane}), \ln(\text{Xylene}))$	0.29	0.03	0.52
	$\rho(\ln(\text{Toluene}), \ln(\text{Xylene}))$	0.18	-0.08	0.43
Regression Coefficients	$\beta_{0,(1)}$	2.21	1.67	2.73
	$\beta_{0,(2)}$	2.13	1.61	2.62
	$\beta_{1,(2)}$	0.12	-0.05	0.29
	$\beta_{0,(Y X)}$	1.29	0.62	1.92
	$\beta_{1,(Y X)}$	0.22	0.08	0.37
	$\beta_{2,(Y X)}$	0.21	-0.02	0.44

Note. The median and 95% CI (2.5 and 97.5 quantiles) are reported for each parameter. ppb = parts per billion.

indicated that this difference was significant (at alpha of 0.10) even though the magnitude of the differences in the five models' WAIC was minimal. Modeling frameworks 4 and 5 had the highest WAIC reported of all models tested. This result indicates that modeling frameworks accounting for the correlation between hexane and toluene or between hexane and xylene were preferred to modeling frameworks that did not account for this correlation.

We also saw consistent trends in both *LPPD* and P_2 . *LPPD* was highest for the three-variable model, indicating strong goodness of fit. P_2 also was the highest for the three-variable model because this model contained more model parameters (increased model complexity). However, the improvement in the goodness-of-fit measure *LPPD* outweighed the increased complexity of the three-variable model (hence the lower WAIC).

Next, we examined the estimates from the three-variable model framework because it was preferred in both model comparisons. Table 5 contains the median posterior estimates and 95% CI of the GMs, GSDs, and AMs for hexane, toluene, and xylene. We also report the median posterior estimates and 95% CIs for the correlations among hexane, toluene, and xylene and the regression coefficients defined for hexane as Y (toluene was $X_{(1)}$ and xylene was $X_{(2)}$).

Results of the three-variable model demonstrated that the three-variable model was preferred in the model comparisons. The intercepts for hexane ($\beta_{0,(Y|X)}$), toluene ($\beta_{0,(1)}$), and xylene ($\beta_{0,(2)}$) were all significantly positive. The regression coefficient corresponding to the slope between hexane and toluene ($\beta_{1,(Y|X)}$) was statistically significant

(median posterior estimate: 0.22; 95% CI: (0.08, 0.37)), and the correlation coefficient was moderate with a median posterior estimate of 0.41. In contrast, the slope coefficient ($\beta_{2,(Y|X)}$) of xylene when modeling hexane was insignificant (median posterior estimate: 0.21; 95% CI: (-0.02, 0.44)). The lack of significance in this regression coefficient implied that xylene did not explain a significant amount of variation in hexane when toluene was already a predictor in the model. However, xylene was related to hexane as demonstrated by the significant correlation coefficient (median posterior estimate: 0.29; 95% CI= (0.03, 0.52)). So, while the model containing toluene and xylene as predictors had a slightly better fit than a model containing only toluene as a predictor, the improvement in fit (for all variables and for hexane) was not substantial, leading to a minimal difference in WAIC and WAIC_Y (Table 4; see Supporting Information). Finally, the slope and correlation estimates between toluene and xylene ($\beta_{1,(2)}$) were slightly positive but insignificant.

There was little difference in the median posterior estimates of the GM exposure estimates (in parts per billion or ppb). The GSD estimates were reasonable for hexane and xylene but higher for toluene; however, the 95% CIs were within the range commonly observed for the GuLF STUDY data (1.01–12). Hexane was slightly less variable than xylene. The AM estimates were greatly influenced by the GSDs. The median posterior estimates for the AMs of hexane, toluene, and xylene were 19.06 ppb, 63.72 ppb, and 23.82 ppb, respectively. The increased variability in toluene led to a higher AM estimate for toluene. Because hexane had a lower GSD and GM, a lower AM of hexane was found.

6 | DISCUSSION

This model provides a statistical framework for exposure estimation when measurements are not available (due to not being measured or being censored) for a chemical in one or more mixtures but that measurements are available on other common chemicals. The simulation study and data analysis example both showed that we can use the linear relationships in chemical mixtures to better inform exposure estimates when data are missing or when censoring is present.

Results of our simulation study in Section 4 indicated that we could correctly identify when both particular covariates ($X_{(1)}$ and $X_{(2)}$) may be useful in a model setting under a variety of censoring levels. We were able to correctly identify that both $X_{(1)}$ and $X_{(2)}$ contributed significantly to the model regardless of the censoring levels in the variables in the model.

Results of our coverage simulation demonstrated that coverage generally decreased as censoring became more different in the covariate and response. This result is expected because censored points would be estimated in a region on Figure 1, which will pull the regression line down or up. This change in the regression line will therefore change the slope and intercept estimates. Results also indicated that with increased censoring, variability was generally higher.

Results of the multicollinearity simulation study showed that as the correlation increases between $X_{(1)}$ and $X_{(2)}$, the width of the 95% CIs increases for the slope coefficients in the conditional expression of $Y | X_{(1)}, X_{(2)}$. However, while censoring increased the CI width overall, the inflation in the width of the CI did not appear to be moderated by censoring levels. In the context of environmental exposure assessment, this inflation in the width of the CI could be concerning because it would affect the uncertainty quantification in the estimates. Because the conditional mean expression is used in the calculation of the GMs and AMs, the inflated width of the 95% CI could lead to greater uncertainty on the mean of Y and therefore greater uncertainty in the final exposure estimate. Such an increase in uncertainty may be undesirable, because in an epidemiologic study, the uncertainty would cause an exposure–disease association to be missed due to misclassification. Therefore, it is recommended that correlations between chemical covariates be kept to less than 0.5 to decrease the likelihood of this increase in uncertainty. One may be able to reduce the increase in the width of the 95% CI by introducing shrinkage priors. The impact of shrinkage priors in this setting should be further investigated at different censoring levels.

Results of our analysis of hexane exposure from July 16, 2010 to September 30, 2010 on the *Ocean Intervention III* indicated that the three-variable model was preferred over other simpler models. Both xylene and toluene were moderately correlated with hexane in the final model ($r = 0.2$ – 0.4). Models with only toluene or only xylene as a predictor for hexane (i.e., frameworks 2 and 3; models 2 and 3) did not fit the data quite as well, given the model complexity, as the multivariate model. Together, however, the correlations provided enough information to our model to allow us to develop reasonable estimates for hexane when hexane measurements were missing either because the measurements were not collected or because the measurement results were below the LOD. In addition, we used the model to better inform toluene and xylene estimates using the available but limited hexane data.

This model assumes that there is a clear linear relationship between the logarithms of the response and each predictor. Other relationships were not explored. This model also relies on the assumptions of linear regression including independence of observations, equal variances, and normality of the residuals. Other distributions or violations of these assumptions were not investigated. Departures from these assumptions should be further investigated. We also did not

investigate censoring levels greater than 75%. Furthermore, we did not assess the impact of influential points or outlying observations in this work. Future work similar to that of Bayes, Bazán, and García (2012) should investigate methods for dealing with potential outlying observations.

Additional information and supporting material for this article is available online at the journal's website.

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SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

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