UC Davis UC Davis Previously Published Works

Title

Draft Genome Sequences of Salmonella enterica Serovar Typhimurium LT2 with Deleted Chitinases That Are Emerging Virulence Factors

Permalink <https://escholarship.org/uc/item/8x2426sm>

Journal Microbiology Resource Announcements, 5(31)

ISSN 2576-098X

Authors

Arabyan, Narine Huang, Bihua C Weimer, Bart C

Publication Date

2017-08-03

DOI

10.1128/genomea.00659-17

Peer reviewed

PROKARYOTES

Draft Genome Sequences of Salmonella enterica Serovar Typhimurium LT2 with Deleted Chitinases That Are Emerging Virulence Factors

Narine Arabyan,a,b Bihua C. Huang,a,b [Bart C. Weimera](http://orcid.org/0000-0002-7471-1978),b

Department of Population Health and Reproduction, School of Veterinary Medicine, University of California, Davis, California, USA^a; 100K Pathogen Genome Project, University of California, Davis, California, USA^b

ABSTRACT Chitinases are glycosyl hydrolases that catalyze the hydrolysis of the β -1,4 linkages in complex carbohydrates and those that contain GlcNAc. These enzymes are considered emerging virulence factors during infection because the host glycan changes. This is the release of four single chitinase deletion mutants in Salmonella enterica serovar Typhimurium LT2.

Chitinases are glycosyl hydrolases (GHs) that belong to the GH18 and GH19 families $(1-6)$ $(1-6)$ $(1-6)$. GH enzymes play a significant role in virulence by altering the host glycan structure during infection and gaining access to the host epithelial cells, which results in the microbe binding to terminal monosaccharides to initiate glycan degradation on the host epithelial cell [\(7,](#page-2-3) [8\)](#page-2-4). Chitinases are emerging virulence factors because they recognize host GlcNAc-containing glycans in mucin and other N-glycosylated proteins in the host membrane, which enable host association as well as glycan digestion, to gain access to the cell membrane to initiate invasion [\(9,](#page-2-5) [10\)](#page-2-6). Glycans with GlcNAc molecules with a β -1,4-glycosidic bond [\(11\)](#page-2-7) are found on intestinal epithelial cells (IECs) and are hydrolyzed during association [\(1,](#page-2-0) [10\)](#page-2-6). This provides Salmonella spp. a method to degrade the glycan and digest the glycocalyx to establish intracellular infections.

Deletion of chitinase genes in Listeria monocytogenes led to a reduction in bacterial counts in the liver and spleen of infected mice [\(12\)](#page-2-8). An adherent-invasive Escherichia coli (AIEC) LF82 deletion of the chiA gene significantly reduced the adhesion to IECs compared to that of the wild type [\(13\)](#page-2-9). Furthermore, AIEC LF82 interacted with an N-glycosylated chitin-binding protein (CHI3L1) on the host cell to mediate close interaction between the host membrane and bacterial cell, which is regulated in animal models of colitis and in human inflammatory bowel diseases (IBDs) [\(14\)](#page-2-10). Microarray analysis showed that SL0018 (chiA) gene in the Salmonella SL1344 strain was strongly induced during the infection of murine macrophage cells [\(15,](#page-2-11) [16\)](#page-2-12). These data indicate that chitinases relandscape the host glycan to promote the attachment of bacteria to the host cells through the interaction with mucin or N-glycosylated glycans during association. The genus Salmonella contains four chitinases that were derived from bacteria and phages. Park et al. [\(8\)](#page-2-4) also demonstrated that Salmonella initiates glycan relandscaping during infection via host gene expression changes and microbe grazing to degrade the glycan, making these enzymes important for infection.

The 100K Pathogen Genome Project [\(http://www.100kgenomes.org\)](http://www.100kgenomes.org) is a large-scale sequencing consortium that offers the use of new next-generation sequencing methods to provide cutting-edge methods for pathogen detection and control in the food supply. This project is focused on sequencing genomes of bacteria from the environment, plants, animals, and humans worldwide, providing new insights into the genetic diversity of pathogens and the microbiome. Four chitinase deletions $(\Delta STMOO18,$

Received 23 May 2017 **Accepted** 25 May 2017 **Published** 3 August 2017

Citation Arabyan N, Huang BC, Weimer BC. 2017. Draft genome sequences of Salmonella enterica serovar Typhimurium LT2 with deleted chitinases that are emerging virulence factors. Genome Announc 5:e00659-17. [https://doi](https://doi.org/10.1128/genomeA.00659-17) [.org/10.1128/genomeA.00659-17.](https://doi.org/10.1128/genomeA.00659-17)

Copyright © 2017 Arabyan et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0](http://creativecommons.org/licenses/by/4.0/) [International license.](http://creativecommons.org/licenses/by/4.0/)

Address correspondence to Bart C. Weimer, [bcweimer@ucdavis.edu.](mailto:bcweimer@ucdavis.edu)

GenBank	SRA	Isolate	Gene		No. of	Coverage	Total genome	No. of coding
accession no.	accession no.	name	deleted	Enzyme activity	contias	(x)	size (bp)	sequences
MZNO00000000	SRR5288763	BCW 8404	Δ STM0018	Exochitinase	61	139	4,893,048	4,810
MXBA00000000	SRR5288762	BCW 8406	$\triangle STM0233$	Endochitinase	63	162	4,894,557	4,808
MXBB00000000	SRR5288761	BCW 8409	Δ STM0907	Prophage chitinase	61	188	4,895,634	4,808
MZYL00000000	SRR5288732	BCW 8417	Δ STM1869A	Putative chitinase	63	177	4,895,461	4,811

TABLE 1 Salmonella enterica serovar Typhimurium LT2 chitinase deletion mutants

 Δ STM0233, Δ STM0907, and Δ STM1869A) were constructed in the Weimer laboratory (University of California, Davis) [\(7\)](#page-2-3) as described by Datsenko and Wanner [\(17\)](#page-2-13). Cultures were prepared for sequencing as described previously [\(18](#page-2-14)[–](#page-3-0)[25\)](#page-3-1). Genome sequences were de novo assembled using CLC Workbench version 6.5.1 with default parameters [\(18\)](#page-2-14).

Accession number(s). All sequences are publicly available and can be found at the 100K Pathogen Genome Project BioProject (NCBI PRJNA186441) in the Sequence Read Archive [\(http://www.ncbi.nlm.nih.gov/sra\)](http://www.ncbi.nlm.nih.gov/sra). NCBI GenBank accession numbers for the genome assemblies are listed in [Table 1.](#page-2-15)

ACKNOWLEDGMENTS

B.C.W. is grateful for the funding contributed by the NIH (1R01HD065122-01A1; NIH-U24-DK097154) and an Agilent Technologies Thought Leader Award.

REFERENCES

- 1. Frederiksen RF, Paspaliari DK, Larsen T, Storgaard BG, Larsen MH, Ingmer H, Palcic MM, Leisner JJ. 2013. Bacterial chitinases and chitin-binding proteins as virulence factors. Microbiology 159:833– 847. [https://doi.org/](https://doi.org/10.1099/mic.0.051839-0) [10.1099/mic.0.051839-0.](https://doi.org/10.1099/mic.0.051839-0)
- 2. Davies G, Henrissat B. 1995. Structures and mechanisms of glycosyl hydrolases. Structure 3:853– 859. [https://doi.org/10.1016/S0969](https://doi.org/10.1016/S0969-2126(01)00220-9) [-2126\(01\)00220-9.](https://doi.org/10.1016/S0969-2126(01)00220-9)
- 3. Henrissat B. 1991. A classification of glycosyl hydrolases based on amino acid sequence similarities. Biochem J 280:309 –316. [https://doi.org/10](https://doi.org/10.1042/bj2800309) [.1042/bj2800309.](https://doi.org/10.1042/bj2800309)
- 4. Henrissat B, Bairoch A. 1993. New families in the classification of glycosyl hydrolases based on amino acid sequence similarities. Biochem J 293: 781–788. [https://doi.org/10.1042/bj2930781.](https://doi.org/10.1042/bj2930781)
- 5. Henrissat B, Bairoch A. 1996. Updating the sequence-based classification of glycosyl hydrolases. Biochem J 316:695– 696. [https://doi.org/10.1042/](https://doi.org/10.1042/bj3160695) [bj3160695.](https://doi.org/10.1042/bj3160695)
- 6. Henrissat B, Davies G. 1997. Structural and sequence-based classification of glycoside hydrolases. Curr Opin Struct Biol 7:637– 644. [https://doi.org/](https://doi.org/10.1016/S0959-440X(97)80072-3) [10.1016/S0959-440X\(97\)80072-3.](https://doi.org/10.1016/S0959-440X(97)80072-3)
- 7. Arabyan N, Park D, Foutouhi S, Weis AM, Huang BC, Williams CC, Desai P, Shah J, Jeannotte R, Kong N, Lebrilla CB, Weimer BC. 2016. Salmonella degrades the host glycocalyx leading to altered infection and glycan remodeling. Sci Rep 6:29525. [https://doi.org/10.1038/srep29525.](https://doi.org/10.1038/srep29525)
- 8. Park D, Arabyan N, Williams CC, Song T, Mitra A, Weimer BC, Maverakis E, Lebrilla CB. 2016. Salmonella typhimurium enzymatically landscapes the host intestinal epithelial cell (IEC) surface glycome to increase invasion. Mol Cell Proteomics 15:3653–3664. [https://doi.org/10.1074/mcp.M116.063206.](https://doi.org/10.1074/mcp.M116.063206)
- 9. Jacobs H, Mink SN, Duke K, Bose D, Cheng ZQ, Howlett S, Ferrier GR, Light RB. 2005. Characterization of membrane N-glycan binding sites of lysozyme for cardiac depression in sepsis. Intensive Care Med 31: 129 –137. [https://doi.org/10.1007/s00134-004-2487-y.](https://doi.org/10.1007/s00134-004-2487-y)
- 10. Tran HT, Barnich N, Mizoguchi E. 2011. Potential role of chitinases and chitin-binding proteins in host-microbial interactions during the development of intestinal inflammation. Histol Histopathol 26:1453–1464. [https://doi.org/10.14670/HH-26.1453.](https://doi.org/10.14670/HH-26.1453)
- 11. Stanley P, Schachter H, Taniguchi N. 2009. N-Glycans, chap. 8. In Varki A, Cummings RD, Esko JD, Freeze HH, Stanley P, Bertozzi CR, Hart GW, Etzler ME (ed.), Essentials of glycobiology, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- 12. Chaudhuri S, Bruno JC, Alonzo F III, Xayarath B, Cianciotto NP, Freitag NE. 2010. Contribution of chitinases to Listeria monocytogenes pathogenesis.

Appl Environ Microbiol 76:7302–7305. [https://doi.org/10.1128/AEM](https://doi.org/10.1128/AEM.01338-10) [.01338-10.](https://doi.org/10.1128/AEM.01338-10)

- 13. Low D, Tran HT, Lee IA, Dreux N, Kamba A, Reinecker HC, Darfeuille-Michaud A, Barnich N, Mizoguchi E. 2013. Chitin-binding domains of Escherichia coli ChiA mediate interactions with intestinal epithelial cells in mice with colitis. Gastroenterology 145:602– 612. [https://doi.org/10](https://doi.org/10.1053/j.gastro.2013.05.017) [.1053/j.gastro.2013.05.017.](https://doi.org/10.1053/j.gastro.2013.05.017)
- 14. Mizoguchi E. 2006. Chitinase 3-like-1 exacerbates intestinal inflammation by enhancing bacterial adhesion and invasion in colonic epithelial cells. Gastroenterology 130:398 –411. [https://doi.org/10.1053/j.gastro.2005.12.007.](https://doi.org/10.1053/j.gastro.2005.12.007)
- 15. Eriksson S, Lucchini S, Thompson A, Rhen M, Hinton JC. 2003. Unravelling the biology of macrophage infection by gene expression profiling of intracellular Salmonella enterica. Mol Microbiol 47:103–118. [https://doi](https://doi.org/10.1046/j.1365-2958.2003.03313.x) [.org/10.1046/j.1365-2958.2003.03313.x.](https://doi.org/10.1046/j.1365-2958.2003.03313.x)
- 16. Larsen T, Petersen BO, Storgaard BG, Duus JØ, Palcic MM, Leisner JJ. 2011. Characterization of a novel Salmonella Typhimurium chitinase which hydrolyzes chitin, chitooligosaccharides and an N-acetyllactosamine conjugate. Glycobiology 21:426 –436. [https://doi.org/10.1093/glycob/cwq174.](https://doi.org/10.1093/glycob/cwq174)
- 17. Datsenko KA, Wanner BL. 2000. One-step inactivation of chromosomal genes in Escherichia coli K-12 using PCR products. Proc Natl Acad Sci U S A 97:6640 – 6645. [https://doi.org/10.1073/pnas.120163297.](https://doi.org/10.1073/pnas.120163297)
- 18. Arabyan N, Weis AM, Huang BC, Weimer BC. 2017. Implication of sialidases in Salmonella infection: genome release of sialidase knockout strains from Salmonella enterica serovar Typhimurium LT2. Genome Announc 5(19):e00341-17. [https://doi.org/10.1128/genomeA.00341-17.](https://doi.org/10.1128/genomeA.00341-17)
- 19. Jeannotte R, Lee E, Kong N, Ng W, Kelly L, Weimer BC. 2014. Highthroughput analysis of foodborne bacterial genomic DNA using Agilent 2200 TapeStation and genomic DNA ScreenTape system. Agilent Technologies Application Note. [https://doi.org/10.13140/RG.2.1.3354.6961.](https://doi.org/10.13140/RG.2.1.3354.6961)
- 20. Kong N, Ng W, Lee V, Kelly L, Weimer BC. 2013. Production and analysis of high molecular weight genomic DNA for NGS pipelines using Agilent DNA extraction kit (p/n 200600). Agilent Technologies Application Note. [https://doi.org/10.13140/RG.2.1.2961.4807.](https://doi.org/10.13140/RG.2.1.2961.4807)
- 21. Weis AM, Huang BC, Storey DB, Kong N, Chen P, Arabyan N, Gilpin B, Mason C, Townsend AK, Smith WA, Byrne BA, Taff CC, Weimer BC. 2017. Large-scale release of campylobacter draft genomes: resources for food safety and public health from the 100K pathogen genome project. Genome Announc 5(1):e00925-00916. [https://doi.org/10.1128/genomeA.00925-16.](https://doi.org/10.1128/genomeA.00925-16)
- 22. Kong N, Ng W, Foutouhi A, Huang BH, Kelly L, Weimer BC. 2014. Quality control of high-throughput library construction pipeline for KAPA HTP library using an Agilent 2200 TapeStation. Agilent Technologies Application Note. [https://doi.org/10.13140/RG.2.1.4927.5604.](https://doi.org/10.13140/RG.2.1.4927.5604)
- 23. Kong N, Thao K, Huang C, Appel M, Lappin S, Knapp L, Kelly L, Weimer BC. 2014. Automated library construction using KAPA library preparation kits on the Agilent NGS workstation yields high-quality libraries for whole-genome sequencing on the Illumina platform. Agilent Technologies Application Note. [https://doi.org/10.13140/RG.2.1.2306](https://doi.org/10.13140/RG.2.1.2306.1203) [.1203.](https://doi.org/10.13140/RG.2.1.2306.1203)
- 24. Lüdeke CH, Kong N, Weimer BC, Fischer M, Jones JL. 2015. Complete

genome sequences of a clinical isolate and an environmental isolate of Vibrio parahaemolyticus. Genome Announc 3(2):e00216-00215. [https://](https://doi.org/10.1128/genomeA.00216-15) [doi.org/10.1128/genomeA.00216-15.](https://doi.org/10.1128/genomeA.00216-15)

25. Weis AM, Clothier KA, Huang BC, Kong N, Weimer BC. 2016. Draft genome sequences of Campylobacter jejuni strains that cause abortion in livestock. Genome Announc 4(6):e01324-16. [https://doi.org/10.1128/](https://doi.org/10.1128/genomeA.01324-16) [genomeA.01324-16.](https://doi.org/10.1128/genomeA.01324-16)