# Impact of the Linked Surface Water-Soil Water-Groundwater System on Transport of *E. coli* in the Subsurface

•			iors						
•	<u>A</u>	<u>Auth</u>	ors and affiliations						
•	[	Dipa	inkar Dwivedi						
•	Binayak P. Mohanty								
•	E	Bruc	e J. Lesikar						
•									
		C							
		C							
•									
		C							
•		0							
		c c							
1.	1								
1. 2.		2.							
2. 3.		 3.							
0.	Article								
				•	327 Down	loads			
					• <u>6</u> Citatio	ons			

## Abstract

*Escherichia coli* (*E. coli*) contamination of groundwater (GW) and surface water (SW) occurs significantly through the subsurface from onsite wastewater treatment systems (OWTSs). However, *E. coli* transport in the subsurface remains inadequately characterized at the field scale, especially within the vadose zone. Therefore, the aim of this research is to investigate the impact of groundwater fluctuations (e.g., recharging, discharging conditions) and variable conditions in the vadose zone (e.g., pulses of *E. coli* flux) by characterizing *E. coli* fate and transport in a linked surface water-soil water-groundwater system (SW-SoW-GW). In particular, this study characterizes the impact of flow regimes on *E. coli* transport in the subsurface and evaluates the sensitivity of parameters that control the transport of *E. coli* in the SW-SoW-GW system. This study was conducted in Lake Granbury, which is an important water supply in north-central Texas providing water for over 250,000 people. Results showed that there was less removal of *E. coli* during groundwater recharge events as compared to GW discharge events. Also, groundwater and surface water systems largely control *E. coli* transport in the subsurface; however, temporal variability of *E. coli* can be explained by linking the SW-SoW-GW system. Moreover, sensitivity analysis revealed that saturated water content of the soil, total retention rate coefficient, and hydraulic conductivity are important parameters for *E. coli* transport in the subsurface.

# Keywords

*E. coli* transport Seasonal variability Septic tanks Surface water and groundwater interaction

## 1 Introduction

Water resources are prone to microbial contamination in rural areas, where a large number of onsite wastewater treatment systems (OWTSs) are present. Onsite/decentralized wastewater treatment systems serve approximately 25 % of US households and almost 40 % new developments (USEPA 2005; Dwivedi et al. 2008; Lowe and Siegrist 2008). The most common OWTS involves a septic tank unit followed by dispersal to a subsurface soil infiltration unit. Onsite systems are one of the many known contributors of pathogens and nutrients to groundwater (GW) (USEPA 2002). *Escherichia coli* (*E. coli*) is a microbe of fecal origin. Therefore, water contamination by various strains of *E. coli* is becoming common in rural areas in the USA (Bradford et al. 2006; Dwivedi et al. 2013).

The level of *E. coli* contamination to GW depends on multiple factors such as precipitation pattern, thickness and composition of the vadose zone (Williams et al. <u>1998</u>), and subsurface heterogeneity (Spalding and Exner <u>1993</u>; Dwivedi and Mohanty <u>2016</u>). Physical processes that control the fate and transport of *E. coli* have been studied extensively (Haznedaroglu et al. 2008). Smith et al. (1985) demonstrated that the extent of E. coli transport largely depends on soil structure. The authors further suggested that flow through soil macropores, which bypasses the retentive capacities of the soil matrix, is a common phenomenon. Gagliardi and Karns (2000) discussed the impact of soil type and method of pathogen delivery on *E. coli* transport in the subsurface. Another study conducted in a karstic aguifer described that the residence time of *E. coli* in the subsurface, which is correlated with the pore velocity, is crucial for GW contamination (Personne et al. 1998). Furthermore, there is evidence in the literature that the survival and transport of *E. coli* in the subsurface is controlled by various factors, such temperature, rainfall, soil type, porosity, and soil water (SoW) content (Federle et al. 1986). Moreover, hydrological interactions between surface water (SW) bodies, SoW, and GW are of fundamental concern to the

migration of contaminants (*E. coli*) in a linked surface water-soil watergroundwater (SW-SoW-GW) system (e.g., McMahon et al. <u>1995</u>; Bethune et al. <u>1996</u>). However, despite the importance of SW-SoW-GW interaction, the characterization of the impact of GW flow patterns, due to dynamic boundary conditions, on fate and transport of *E. coli* near surface water bodies is still lacking. Therefore, the goal of this study is to investigate how the interaction of surface water, soil water, and GW affect *E. coli* transport in the subsurface.

To clearly distinguish the impact of these interactions, we model E. *coli* transport in the subsurface—first with representing groundwater and surface water system only and then with linked SW-SoW-GW system, in other words by explicitly representing vadose zone processes in the model. To investigate and capture the impact of surface water-groundwater interactions, we chose MODFLOW because of its capability to simulate lakegroundwater interactions or fluctuating lake levels (Anderson 1991). However, the influence of the vadose zone is oversimplified in MODFLOW, as recharge and concentration boundary conditions are calculated outside the model without appropriate consideration of the vadose zone (Twarakavi et al. 2008). On that account, to understand the transport of E. coli in the subsurface under dynamic boundary conditions for a linked SW-SoW-GW system, we use HYDRUS and MODFLOW/MT3DMS models, in which HYDRUS is used to produce *E. coli* flux at the water table that becomes a boundary condition for GW flow and transport. The aim of this research is to investigate *E. coli* transport in the vadose zone under rechargingdischarging GW that involves variations in GW table. Besides, this study tests the robustness of model parameters (by using calibrated MODFLOW/MT3DMS parameters without further modification) and characterizes parameter sensitivity to identify dominant controls on the fate and transport of *E. coli* in the saturated zone and SW-SoW-GW system.

# 2 Study Site

### 2.1 Site Description

This study is conducted at a water-quality monitoring station (Segment ID: 1205; latitude 32° 27′ 40″ and longitude 97° 42′ 53″) in the Lake Granbury area, which is monitored by the Brazos River Authority. Lake Granbury is a man-made lake of 35 km<sup>2</sup> within the Middle Brazos-Palo Pinto watershed (USGS Cataloging Unit: 12060201). The watershed is a part of the Brazos River basin and is located in north-central Texas, in Hood County. Summers are long and hot while winters are short and mild. Average temperatures range from a low of 1.1 °C in January to a high of 35.5 °C in July. The average precipitation in the watershed is 0.76 m yearly. The watershed has a diversified land use from urban to agriculture; about 31 to 40 % of the land is considered prime farmland. The landscape is described by rolling hills

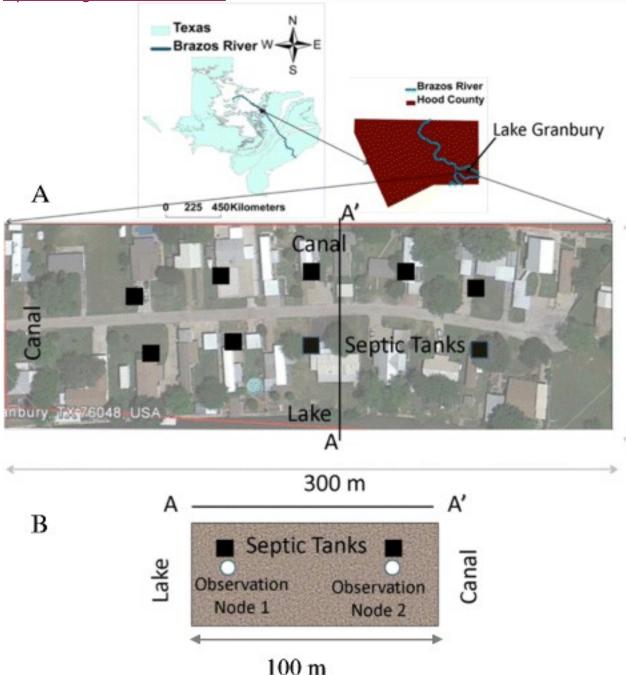
of pasture and cropland surrounded by deciduous forest. Lake Granbury is an important source of water supply in this area, providing water for over 250,000 people in more than 15 cities. Recent studies by the Brazos River Authority (BRA), Texas Water Resources Institute (TWRI), and Espey Consultants, Inc. have concluded that some of the Lake Granbury's coves especially shallow bodies of water are contaminated with *E. coli*.

Several studies have been conducted to assess potential *E. coli* sources in lake or canals in the Lake Granbury Watershed (Riebschleager et al. 2012; Dwivedi et al. 2013). Potential E. coliloads can stem from several sources, including livestock (e.g., cattle, pets), malfunctioning septic systems or onsite wastewater treatment systems (OWTSs), surface runoff, and wastewater treatment plants (WWTPs). The nearby WWTP is located in downstream of the study site and thus is not a source. In addition, pets are primarily kept in homes, and pet waste is disposed directly to solid waste management (to landfill mostly via garbage collection). The other livestock is infrequently sighted. Previous studies in the watershed have demonstrated that point source loadings (e.g., OWTSs) are constant, whereas nonpoint source loadings are associated with high flows due to runoff events (Riebschleager et al. 2012). As runoff begins, E. *coli* concentration increases in the lake and canal, which decreases at a faster rate than the river discharge, eventually approaching background levels (Pachepsky and Shelton 2011). Therefore, we applied *E. coli* loads from the malfunctioning septic systems as the primary source of *E. coli* in this system. Moreover, in the Lake Granbury area, these septic systems are located in a zone, where shallow groundwater and surface water interact. Thus, shallow groundwater in the region can be easily contaminated by the sewage and can further contaminate the lake and canal.

### 2.2 Data Description

The site was characterized for soil texture and shallow subsurface hydrologic conditions (e.g., groundwater level). Seventeen soil cores were carefully excavated from 0.15 to 1.0 m depth at several locations along the *A*-*A*' transect (Fig. <u>1</u>). Saturated hydraulic conductivity, bulk density, and soil texture details were obtained by laboratory experiments conducted on these undisturbed soil cores. In particular, saturated hydraulic conductivity for each core was measured based on the constant-head permeameter method (Klute and Dirksen <u>1986</u>; Mallants et al. <u>1997</u>). In this method, a hydraulic head on top of the cores is imposed, and the resulting flux is measured at the free-water outlet. Subsequently, soil texture analysis was performed using the hydrometer method (Milford <u>1997</u>; Lesikar et al. <u>2005</u>). For soil type, particle-size data were used to classify the soil texture from the USDA textural triangle (Davis and Bennett <u>1927</u>; Twarakavi et al. <u>2010</u>). The soil texture varied from silt clay loam to sandy clay loam.

#### <u>Open image in new window</u>



#### Fig. 1

Plan view of the model domain (m) for **a** MODFLOW/MT3DMS and **b** crosssectional view of the model domain (m) for HYDRUS-2D along the A-A' transect (map courtesy: Google Earth)

Precipitation data were acquired from the National Climatic Data Center (NCDC) (<u>http://www.ncdc.noaa.gov/</u>), and evapotranspiration data from TexasET Network (<u>http://texaset.tamu.edu/index.php</u>). The Brazos River Authority and the Texas Commission on Environmental Quality (TCEQ)

collected the water quality data in Lake Granbury under Clean River Programs (CRP) on Environmental Quality. Concentration data of *E. coli*(CFU/100 mL) in Lake Granbury were available from July 2002 to August 2010, using grab samples collected monthly at 0.3 m below the surface. The *E. coli* concentration data was measured using the Colilert (Defined Substrate Technology) method (Eaton et al. <u>1988</u>). The *E. coli* observations used in this study meet all quality assurance requirements (e.g., holding time) of the CRP and the Brazos River Authority Environmental Services. However, we acknowledge the uncertainty in the observed *E. coli* data due to lack of replicates.

# 3 Methodology

For this study, we modeled *E. coli* transport in the subsurface using MODFLOW/MT3DMS to simulate groundwater and surface water interactions and then with loosely coupled HYDRUS and MODFLOW in a 2-D system to simulate SW-SoW-GW interactions. In the subsequent sections, we provide a description of the conceptual model for the fate and transport of *E. coli* in the subsurface and modeling framework in the Lake Granbury area using the saturated zone (MODFLOW/MT3DMS) and unsaturated zone (HYDRUS-2D) models. We also describe the sensitivity analysis that was performed to identify parameters that control the transport of *E. coli* in the SW-SoW-GW system. For a comprehensive analysis of temporal variability in *E. coli*, we analyzed two different hydrologic scenarios that are possible in the Lake Granbury area:

1.

1.

2.

The first scenario (scenario 1) involves the GW level to be lower than the water level in the lake and the canal.

2.

The second scenario (scenario 2) involves the GW level to be higher than the water level in the lake and the canal.

The GW table fluctuates in response to climatic conditions (precipitation and water loss due to transpiration through capillarity), and this fluctuation imposes a dynamic boundary condition in the linked surface water-soil water-groundwater system (Lake Granbury Area). These scenarios allowed us to investigate the possible effects of different hydrologic conditions and resulting temporal variability in the transport of *E. coli* in the subsurface.

## 3.1 E. coli Transport in Porous Media

The advection-dispersion-sorption (ADS) equation is used to describe the fate and transport of *E. coli* in porous media (Matthess et al. <u>1988</u>; Murphy and Ginn <u>2000</u>; Powelson and Mills <u>2001</u>; Pang et al. <u>2004</u>; Foppen et al. <u>2005</u>; Foppen and Schijven <u>2006</u>). The deposition profile of *E. coli* in porous media is modeled as colloid particles (Bradford et al. <u>2006</u>; Foppen and

Schijven 2006). The dynamics of colloid deposition and blocking can be described by expressing colloid transport equation in terms of particle concentration instead of mass concentration (Wan and Wilson 1994; Johnson and Elimelech 1995; Sun et al. 2001) along with terms for straining, attachment, detachment, and die-off (Cunningham et al. 1991; Bergendahl and Grasso 2000; Bhattacharjee et al. 2002; Tufenkji et al. 2004; Torkzaban et al. 2006; Tufenkji 2007)

 $\partial \square \partial \square = \nabla . (\mathbf{D} \nabla \square) - \nabla . (\mathbf{v} \square) - \square \square \square P \partial \square \partial \square - \square \square \partial C \partial t = \nabla . (D \nabla C) - \nabla . (\mathbf{v} C) - f \pi a p 2 \partial S \partial t - k i C$ 

(1)

(2)

where *C* is the number concentration (or concentration interchangeably in this study) of suspended bacteria in the aqueous phase  $[L^{-3}]$ , **D** is the hydrodynamic dispersion coefficient  $[L^2T^{-1}]$ , **v** is the pore water flow velocity  $[LT^{-1}]$ , *f* is the specific surface area of the porous medium  $[L^{-1}]$ , *a*, is the diameter of the bacteria [L], *S* is a dimensionless quantity that describes the fractional surface coverage and is defined as the ratio of the total cross-sectional area of deposited bacteria and interstitial surface area of the porous medium (Sun et al. 2001), *k*, is the die-off rate coefficient of bacteria in the water  $[T^{-1}]$ , and *t* is time [T].

For bacteria retained by the solid matrix, several authors (Matthess et al. <u>1988</u>; Harvey and Garabedian <u>1991</u>; Lindqvist and Bengtsson <u>1991</u>; Lindqvist et al. 1994; Powelson and Mills 2001; Pang et al. 2004) assume some percentage available for instantaneous sorption ( $S_1$ ) and some percentage of bacteria available for kinetic or rate-limited sorption ( $S_2$ ). On the contrary, some (Tan et al. <u>1994</u>; McCaulou et al. <u>1995</u>; Hendry et al. <u>1999</u>) assume only kinetic sorption ( $S_2$ ). The instantaneous sorption can be modeled using the partition coefficient ( $K_{d}$ ) [MM<sup>-1</sup>]; the kinetic sorption can be modeled using the first-order kinetics parameterized by the attachment  $(k_{i})$  [T<sup>-1</sup>] and detachment  $(k_{i})$  [T<sup>-1</sup>] coefficients. Some studies have demonstrated that the kinetic sorption is limited by the blocking (i.e., the occlusion of collector surface) (Ryan and Elimelech 1996), and the effect of the blocking behavior, a dimensionless function (B) of kinetically sorbed E. *coli* (i.e.,  $B(S_2)$ ) can be accounted for by employing the Langmuirian dynamics (Johnson and Elimelech <u>1995</u>; Foppen and Schijven <u>2006</u>). However, apart from the instantaneous and kinetic sorption models, others (Bradford et al. 2003; Foppen et al. 2005; Bradford and Bettahar 2005; Foppen and Schijven 2006) assume a kinetic fraction also available for straining ( $S_3$ ), which is modeled using the first-order kinetics parameterized using the straining rate coefficient ( $k_{str}$ ) [T<sup>-1</sup>]. Taking all these processes into account, the mass balance equation for retained bacteria can be expressed as  $\partial \square \partial \square = \partial \square 1 \partial \square + \partial \square 2 \partial \square + \partial \square 3 \partial \square - \square is \square \partial S \partial t = \partial S 1 \partial t + \partial S 2 \partial t + \partial S 3 \partial t - kisS$ 

 $\partial \square \partial \square = \square \square 2p \{ \square d \partial \square \partial \square + (\square a \square (\square 2) + \square str) \square \} - \square r \square 2 - \square is \square \partial S \partial t = \pi ap 2 \{ Kd \partial C \partial t + (kaB(S2) + kstr)C \} - krS2 - kisS$ 

where  $k_{is}$  [T<sup>-1</sup>] is the inactivation or die-off rate coefficient of bacteria on the solid matrix.

Several studies have noted that equilibrium adsorption and blocking have little impact on *E. coli* transport in the subsurface (Butler et al. <u>1954</u>; Bouwer et al. <u>1974</u>). Additionally, various field studies reported that detachment is insignificant at the field scale (Sinton et al. <u>1997</u>, <u>2000</u>; Foppen and Schijven <u>2006</u>). Given the little apparent impact of detachment, inactivation of sorbed cells becomes irrelevant, and Eq. (<u>2</u>) can be reduced to:  $\partial \square \partial \square = \square \square 2p(\square a + \square str) \square \partial S \partial t = \pi ap2(ka + kstr)C$ 

 $\partial \square \partial \square = \nabla . (\mathbf{D} \nabla \square) - \nabla . (\mathbf{v} \square) - \square (\square a + \square str + \square i) \square \partial C \partial t = \nabla . (D \nabla C) - \nabla . (vC) - f(ka + kstr + ki)C$ 

 $\partial \square \partial \square = \nabla . (\mathbf{D} \nabla \square) - \nabla . (\mathbf{v} \square) - \square total \square \partial C \partial t = \nabla . (\mathbf{D} \nabla C) - \nabla . (\mathbf{v} C) - ktotal C$ 

(6)

(4)

(5)

where  $k_{\text{total}}$  [T<sup>-1</sup>] is the total retention rate coefficient, which includes the effect of favorable and unfavorable attachment, as well as straining and dieoff or inactivation. Although it is desirable to include the effects of abiotic and biotic factors on the total retention of *E. coli*, we used the total retention rate coefficient without distinguishing the impact of these factors, as the distinction between the underlying processes is not possible with the limited data. For this reason, we perform sensitivity analysis and test the reliability of the available model parameters (see Section <u>3.4</u>).

3.2 Physical Domain Setup for the Saturated Zone

The MODFLOW/MT3DMS model domain is described in Fig. <u>1a</u>. The flow and transport processes were simulated using 1 m × 1 m uniform grids. The model included one layer, for a total modeling depth of 10 m. Model topography was imported from surveyed elevation data. The top of the model is assigned as a recharge boundary. The initial time step was chosen as  $8.64 \times 10^3$  s, while the minimum and maximum time steps of  $8.64 \times 10^2$  and  $4.32 \times 10^4$  s were employed, respectively. The lateral limits are defined by physical and hydraulic boundaries (Fig. <u>1</u>).

The flow and transport parameters used for MODFLOW/MT3DMS models are listed in Table <u>1</u>. The total retention rate coefficient varies in a wide range. For sandy grain size and *E. coli* (1–3 µm), the total retention coefficient is in the order of  $6 \times 10^{-4}$ /h (or 2.16/s) (Foppen and Schijven <u>2006</u>). Similarly, Pang et al. (<u>2004</u>, <u>2006</u>) also showed that the removal rate of *E. coli*for silt loam, sandy loam, and sand is of the order of  $6 \times 10^{-4}$ /h (or 2.16/s). In this study, we assumed the total retention rate coefficient, which also includes the effect of favorable and unfavorable attachment, as well as straining and dieoff or inactivation, as  $6 \times 10^{-4}$ /h (or 2.16/s).

(3)

## Table 1

Initial input parameters for E. coli transport in SW-SoW-GW

	Value	Reference
HYDRUS parameters		
Lateral dispersivity (m)	1	Gelhar et al. ( <u>1992</u> )
Vertical dispersivity (m)	0.1	Gelhar et al. ( <u>1992</u> )
Lateral hydraulic conductivity (m/s)	3.5 × 10-◎	Laboratory-measured value
Vertical hydraulic conductivity (m/s)	3.5 × 10-∘	Laboratory-measured value
Total retention coefficient (1/s)	$1 \times 10^{-4}$	Foppen et al. ( <u>2005</u> ), Pang et al. ( <u>2006</u> )
<i>E. coli</i> concentration in septic tanks (CFU/100 mL)	1 × 10°	Riebschleager et al. ( <u>2012</u> )
Effluent loading (m <sup>3</sup> /s)	1.05 × 10 <sup>-5</sup>	Based on Texas Administrative Code
MODFLOW/MT3DMS parameters		
Total retention coefficient (1/s)	$1 \times 10^{-4}$	Foppen et al. ( <u>2005</u> ), Pang et al. ( <u>2006</u> )
Effective porosity	0.3	Mace et al. ( <u>2000</u> )
Diffusion coefficient (m²/s)	2.7 × 10 <sup>-8</sup>	Foppen et al. ( <u>2007</u> )

Visual MODFLOW and MT3DMS models were run under quasi-steady state and then calibrated to observed *E. coli* concentrations using the parameter estimation (PEST) software. This calibration for estimating *E. coli* concentration was done for a period of 49 months—July 2002 to July 2006 at the observation point located by the lake. Model calibration consisted of modifying the flow and transport parameters to minimize the normalized root mean squared (NRMSE) error between estimated and observed *E. coli* concentrations. The model was then run for a period of 49 months—August 2006 to August 2010, without further calibration. These calibrated model parameters were applied to the linked SW-SoW-GW system without further modification.

3.3 Modeling Framework in the Unsaturated Zone and Linked SW-SoW-GW System

*E. coli* transport in the subsurface was explored in the Lake Granbury area using the physically based HYDRUS-2D hydrologic simulation solver (Šimůnek et al. 2006). HYDRUS-2D allows the user to analyze fate and transport of *E. coli* through saturated, variably saturated regions with complex geometries, and composed of nonuniform soils. In the Lake Granbury area, there are shallow canals, which are hydrologically connected to the lake (Fig. <u>1</u>).

The modeling domain is described in Fig. <u>1b</u>. Two-dimensional model domain is 100 m (*X*) by 2 m (*Z*). The model domain is uniformly discretized with 1 m horizontal (*X*) resolution and 0.25 m vertical (*Z*) resolution using unstructured finite element mesh in HYDRUS. An initial time step of 8.64 × 10<sup>3</sup> s day and minimum and maximum time steps of 8.64 × 10<sup>2</sup> and 4.32 × 10<sup>4</sup> s were employed (similar to MODFLOW/MT3DMS). The flow and transport simulations for *E. coli* were performed for a period of 700 days. This simulation period was chosen based on the time required for *E. coli* to reach the peak concentration at observation nodes (Fig. <u>1</u>). Flow and transport parameters used in the HYDRUS-2D model are listed in Table <u>1</u>. The boundary conditions are listed in Table <u>2</u>.

### Table 2

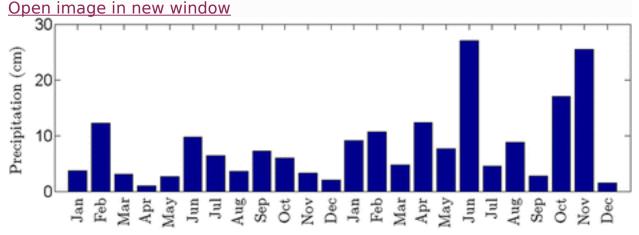
Boundary conditions (BC) for the HYDRUS and MODFLOW/MT3DMS models

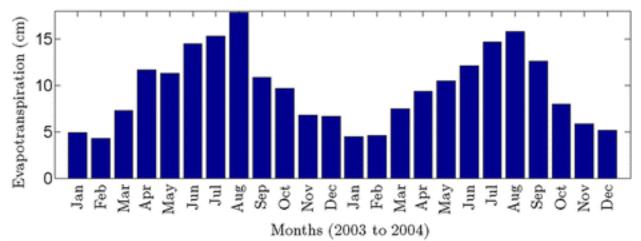
	Boundary conditions
HYDRUS	
Hydrologic	Atmospheric BC
upper BC	$\label{eq:constraint} \begin{array}{l} \square(\square) = 0 \text{ for } h(\square,\square) < 0 \square(\square,\square,\square) \text{ for } h(\square,\square) \ge 0 q(t) = 0 \text{ for } h(t,z) < 0 q(t,x,z) \\ \mbox{ ) for } h(t,z) \ge 0 \end{array}$
Hydrologic	Deep drainage
lower BC	$\begin{array}{l} \square(\square) = 0 \text{ for } h(\square,\square) < 0 \square(\square,\square,\square) \text{ for } h(\square,\square) \ge 0 q(t) = 0 \text{ for } h(t,z) < 0 q(t,x,z) \text{ for } h(t,z) \ge 0 \end{array}$
Hydrologic	Variable head
lateral BCs	h = h(t)

	Boundary conditions				
Solute upper BC	No flux				
be	$q_{c} = 0$				
Septic tanks BC	Constant flux				
	$q_{\rm c} = q_{\rm co}$				
Solute lateral and	Flux type				
lower BCs	$q = q_{c}(t)$				
MODFLOW/MT3D	10DFLOW/MT3DMS				
Hydrologic	Atmospheric BC				
upper BC	$ [([])=0 \text{ for } h([],[])<0 [([],[],[]) \text{ for } h([],[]) \ge 0 q(t)=0 \text{ for } h(t,z)<0 q(t,x,z) \text{ for } h(t,z) \ge 0 $				
Hydrologic lower BC	Deep drainage				
IOWEI DC	$ \Box(\Box) = 0 \text{ for } h(\Box,\Box) < 0 \Box(\Box,\Box,\Box) \text{ for } h(\Box,\Box) \ge 0 q(t) = 0 \text{ for } h(t,z) < 0 q(t,x,z) \text{ for } h(t,z) \ge 0 $				
Lateral BCs	Head-dependent flux boundary				
	$q(t) = -K(h-h_{ref})$				
Solute upper	Flux type				
BC	$q_{c} = q_{c}(t)$				
Solute lower BC	Flux type				
DC	$q_{\rm c} = q_{\rm c}(t)$				
Solute lateral BCs	Flux type				
	$q_{\rm c} = q_{\rm c}(t)$				

Baseline simulations (scenarios 1 and 2) with no background *E. coli* concentration, as initial conditions, and forcing data (Fig. <u>2</u>) were run for 2 years in HYDRUS. The simulation reached a steady state in this time frame for two scenarios individually. *E. coli* flux at the water table was calculated using HYDRUS. For the linked SW-SoW-GW system, *E. coli* flux served as the solute (contaminant) boundary condition for GW flow and transport

(MODFLOW/MT3DMS). The calibrated model parameters from MODFLOW/MT3DMS were used in this loosely coupled simulation so as to test the robustness of these model parameters. As suggested above, we considered multiple scenarios that can alter the *E. coli* concentration at the boundary (interface of the saturated and unsaturated zones). Modeling results (*E. coli* estimation) were tested against observations at the monitoring stations.





### Fig. 2

Precipitation and evapotranspiration in the Lake Granbury area for 2003 and 2004 demonstrate that June to September have high evapotranspiration and lower precipitation as compared to October to May (data: Texas Water Development Board)

### 3.4 Calibration and Sensitivity Analysis

The aim of the sensitivity analysis was to identify model parameters that impact *E. coli* flow and transport in the subsurface as well as to find critical values of parameters that may lead to a reasonable agreement between estimated and observed *E. coli* values. We used a parameter estimation package called PEST, which is model independent. *E. coli* concentrations were available at the water quality monitoring station in Lake Granbury, and therefore, PEST was used for both calibrating the MODFLOW/MT3DMS and performing the sensitivity analysis.

We also analyzed sensitive parameters for HYDRUS-2D, which was used for 2-D flow and solute transport representations in the unsaturated zone. The sensitivity analysis was carried out by individually varying one-factor-at-a-time (OFAT). The OFAT approach has been deemed appropriate for evaluating the sensitivity of different parameters, and this method is computationally frugal that changes one parameter at a time from reference parameter sets, and computes the difference in the outputs (Arora et al. 2012). The OFAT approach involved perturbing each factor individually within a reasonable interval ( $\pm$ 30 %) and keeping the rest of the factors constant at their baseline values.

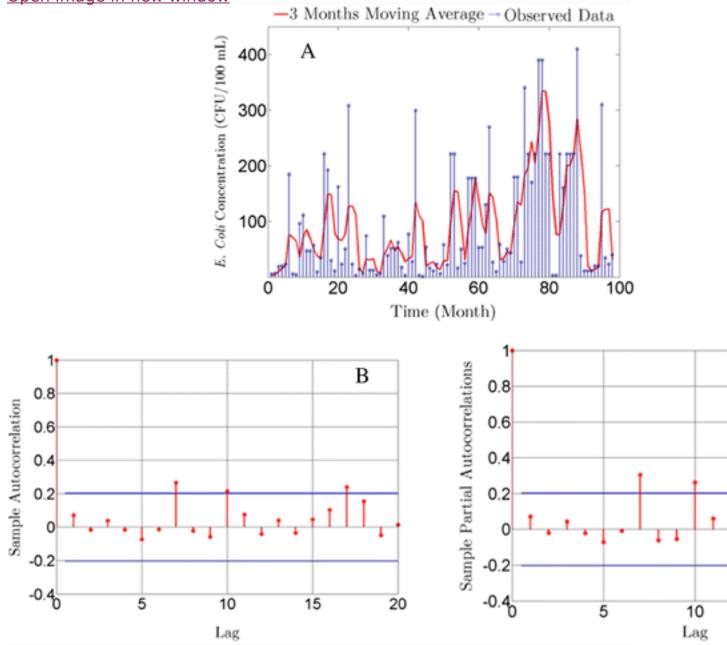
# 4 Results and Discussion

## 4.1 E. coli Transport in the Subsurface

The base case simulation was designed to consider groundwater and surface water system (MODFLOW/MT3DMS) only so as to gain insight into the interpretation of processes controlling *E. coli* transport in the saturated zone. Simulation results (not shown here) were able to capture the average *E. coli* concentrations but failed to capture the variability in time. Simulation results showed that groundwater and surface water systems largely control *E. coli*transport in the subsurface. The normalized mean squared error (NMSE) value was >0.95 from the simulated and observed *E. coli* concentrations. It is worth mentioning that the lower the NMSE, the better the model performance. An NMSE of 0 indicates that the model predictions are perfect.

To gain insights into the temporal variability of *E. coli* concentrations, we used autocorrelation function (ACF) and partial autocorrelation function (PACF) to describe the seasonal occurrence of *E. coli* in the lake. The ACF describes the similarity between *E. coli* observations as a function of the time lag between them. The higher ACF values show repetitive patterns, such as seasonality. The PACF computes the partial correlation with its lagged values in a time series. Figure <u>3</u> demonstrates ACF and PACF of measured *E. coli* in the lake. Repeating patterns of 7 and 10 months are observed. This is because climatic seasonality has an effect on *E. coli*transport. Seasonality also has an impact on GW table that fluctuates in response to climatic conditions. For example, in Lake Granbury site, winter (October to May) precipitation is often higher than summer (June to September) precipitation (Harmel et al. <u>2003</u>), and so GW storage is not fully recharged in summer. This is also apparent in Fig. <u>2</u> where higher

for the years 2003 and 2004. We will evaluate the role of seasonal groundwater fluctuations on *E. coli* fate and transport (Section <u>4.2</u>). Open image in new window

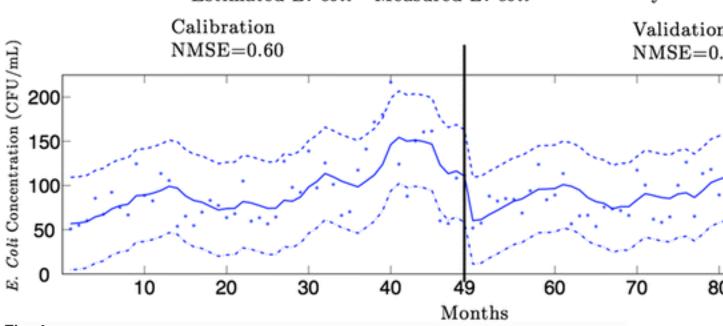


#### Fig. 3

The **a** 3-month moving average of *E. coli* concentrations (July 2002 to August 2010) in the monitoring station shows the seasonality in *E. coli* concentrations. The **b**autocorrelation (ACF) and **c** partial autocorrelation (PACF) functions of *E. coli*concentrations in the monitoring station describe the seasonality in the observed *E. coli*concentrations. The ACF and PACF demonstrate a repeating pattern of 7 and 10 months Because the presence of *E. coli* in the lake is a continuous phenomenon, it is appropriate to test our model performance against smoothed out *E. coli* data. Based on the ACF and PACF analysis, a 3-month moving average (frequency is 3 months after winter months) of *E. coli*concentrations was selected. The NMSE value was 0.77 from the observed and MODFLOW/MT3DMS-simulated *E. coli* concentrations. Simulation results indicated that groundwater and surface water systems affect *E. coli* transport in the subsurface; however, temporal variability of *E. coli* cannot be captured alone by groundwater and surface water integration.

Another set of simulation was carried out to consider SW-SoW-GW system (MODFLOW/MT3DMS and HYDRUS) so as to gain insight into the interpretation of processes controlling the *E. coli* transport in the subsurface. As suggested above, *E. coli* flux at the water table was calculated using HYDRUS and this flux served as the solute (contaminant) boundary condition for GW flow and transport (MODFLOW/MT3DMS). Simulated (using loosely coupled HYDRUS and MODFLOW/MT3DMS) and observed E. *coli* concentrations are shown in Fig. <u>4</u>. Results show a reasonably good agreement between estimated and observed *E. coli*concentrations in the lake for calibration and validation periods. The NMSE value is 0.60 (calibration) to 0.55 (validation) from the simulated and observed E. *coli* concentrations. The uncertainty bands in Fig. 4 reflect the 95 % confidence interval of the model output. More than 85 % observed E. *coli* values fall within the confidence limits of  $\pm \sigma$  (1 standard deviation). Nevertheless, 15 % *E. coli* values, which are not captured within  $\pm \sigma$  interval, still fall within the 95 % confidence limits. Additionally, the mismatch between observed and estimated concentrations of *E. coli* can be a result of various sources of uncertainty. A key source of uncertainty—which is relatively difficult to incorporate—is related to the background concentrations of *E. coli* in soils and sediments (Pachepsky et al. 2006). Therefore, it is believed that as more data become available, these uncertainties will decline.

#### Open image in new window — Estimated E. coli · Measured E. coli - Uncertainty Band



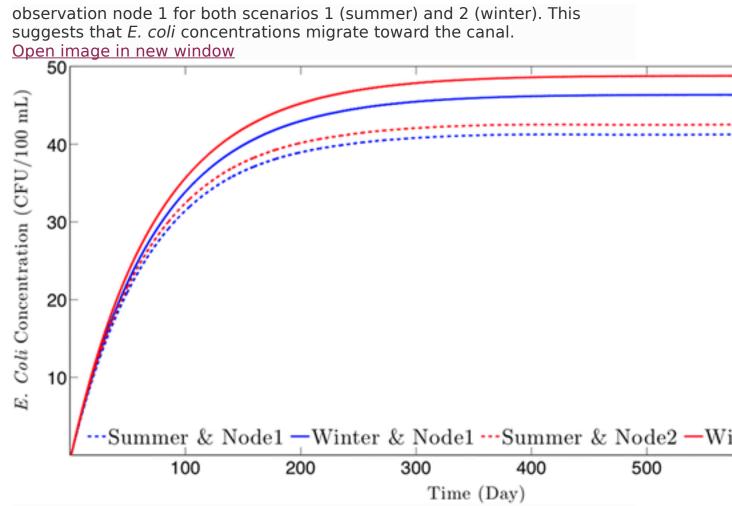
### Fig. 4

Estimated *E. coli* concentrations from the calibrated MODFLOW/MT3DMS model and observed *E. coli* concentrations in the lake show that the observed *E. coli* values fall within the uncertainty band of 95 % confidence interval. Months from July 2002 to July 2006 (months 0–48 on the *X*-axis) and months from August 2006 to August 2010 (months 49–98) were used for calibration and validation, respectively

### 4.2 Effect of Seasonality on E. coli Transport

To better understand the impact of seasonality on *E. coli* transport in the subsurface, we consider two scenarios that incorporate GW table fluctuations in response to climatic conditions. Scenario 1 represents the hydrologic conditions in summer, where the water table is lower than the lake. Likewise, scenario 2 represents the hydrologic conditions in the winter, where the GW level is considered to be higher than the water level in the lake and canal. Consequently, GW gains water from the lake during summertime, and GW loses water to the lake during wintertime.

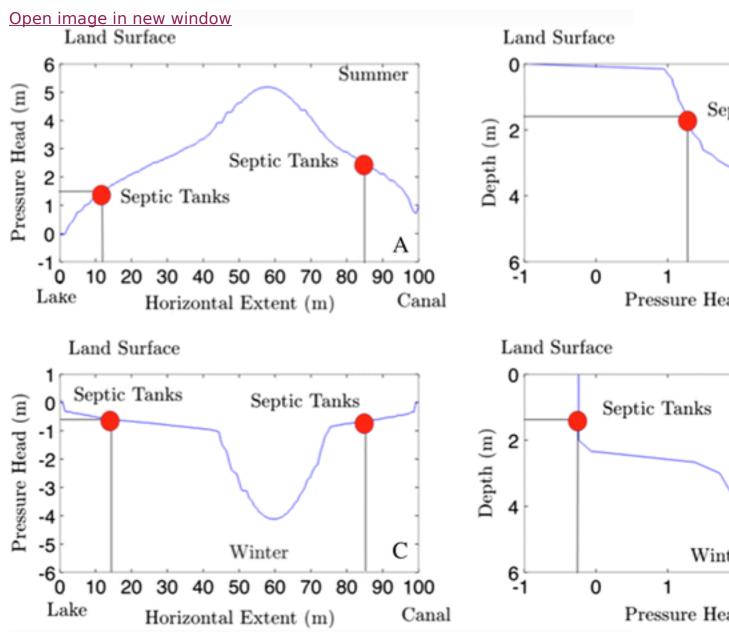
Figure <u>5</u> describes the characteristics of the *E. coli* breakthrough curves providing information on the peak concentrations of *E. coli* at observation nodes 1 and 2 (Fig. <u>1</u>) for scenarios 1 (summer) and 2 (winter). It is evident from Fig. <u>5</u> that the peak concentrations of *E. coli* are smaller in the case of summer months as compared to winter months for both observation nodes 1 and 2. These results indicate less removal of *E. coli* in winter as compared to summer. In addition, Fig. <u>5</u> also demonstrates that higher peak concentrations of *E. coli* were obtained at observation node 2 than



#### Fig. 5

The breakthrough curves of *E. coli* concentration at observation nodes 1 and 2 demonstrate the impact of scenarios 1 and 2 on E. coli transport in the subsurface. Scenario 1 represents summer, and scenario 2 represents winter The E. coli concentration profiles in vertical and horizontal directions were examined to investigate the important characteristics of E. coli transport in the subsurface. Figure 6 shows the change in pressure head profiles during different scenarios (winter and summer). Figure 7 represents the concentration profiles of *E. coli* in the horizontal (1.5 m below land surface) and vertical directions along transect A-A' for scenarios 1 (summer) and 2 (winter). The lake and canal are located at 0 and 100 m, respectively, on the X-axis. Septic tanks are located at 15 and 85 m on the X-axis (Fig. 6). Figure 6a, c again demonstrates higher concentrations near the lake as well as the canal during winter times (scenario 2) as compared to summer times (scenario 1). As GW loses water during winter times (Fig. 5), there is advective transport toward the lake and canal from GW. Since the major transport modes of *E. coli* in the subsurface are through advective transport (Jamieson et al. 2003), higher E. coli concentrations were obtained at observation nodes 1 and 2 during winter times. In addition, wintertime

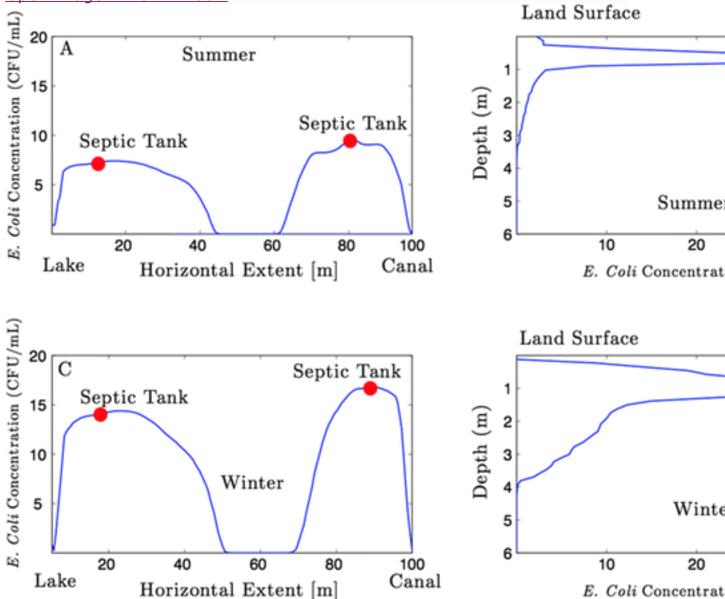
(scenario 2) showed small fluctuations in *E. coli* concentrations near septic tanks (Fig. <u>7</u>). These fluctuations may be attributed to infiltrating water because there is higher precipitation in the Lake Granbury area in the winter months. Figure <u>7</u> also depicts that *E. coli* concentration drops quickly toward the center of the modeling domain (42 to 58 m on the *X*-axis). This phenomenon—higher *E. coli* concentration near the lake and canal, and *E. coli* concentration dropping to zero toward the center of the modeling domain—shows that a GW divide exists in the modeling domain. The GW divide, defined by a hypothetical line on either side of which GW moves in opposite directions, occurs because of the shallow water table, which is strongly influenced by surface water flow (Anderson and Munter <u>1981</u>). The GW divide prevents *E. coli* movement through advective transport toward the center of the modeling domain.



#### Fig. 6

The horizontal and vertical profiles (across septic tanks near the lake) of pressure head demonstrate the impacts of scenario 1 (summer) **a** (horizontal profile) and **b** (vertical profile); and scenario 2 (winter) **c** (horizontal profile) and **d** (vertical profile) on the extent of the saturated and unsaturated zones. Note that 5 m pressure head at 60 m lateral extent in **a** signifies pressure head obtained at depth below 5 m from the land surface (as shown in **b**)

Open image in new window



#### Fig. 7

The horizontal (1.5 m below land surface) and vertical profiles (across septic tanks near the lake) of *E. coli* concentration demonstrate the impacts of scenario 1 (summer)—**a**(horizontal profile) and **b** (vertical profile); and scenario 2 (winter)—**c** (horizontal profile) and **d** (vertical profile) on *E. coli* transport in the subsurface

Figure <u>7b.</u> d represents the concentration profile of *E. coli* in the vertical direction (by the septic tanks) for scenarios 1 (summer) and 2 (winter). Figure <u>7b.</u> d manifests little upward retention of *E. coli* for both the scenarios. It is evident from this figure that the soil matrix within a depth of 0.5 m contains 90 % of *E. coli* from the source (septic tanks), which is consistent with previous studies (DeFlaun et al. <u>1997</u>; Zhang et al. <u>2001</u>; Jiang et

al. <u>2007</u>). In a column study, Tong et al. (<u>2005</u>) also reported that microbial deposition, under a variety of flow conditions and environmentally relevant ionic strength, decreases with distance from the source; however, deposition profiles have also been observed to exhibit a peak at some distance down gradient exhibiting sensitivity to system conditions.

Figure <u>7a–d</u> further demonstrates that *E. coli* concentration gradually changes for scenario 2 (winter) than for scenario 1 (summer), wherein *E. coli* concentration drops quickly. In other words, scenario 2 (winter) displayed increased retained concentrations relative to scenario 1 (summer), possibly reflecting longer *E. coli* residence time for scenario 2 and, hence, greater attachment. These observations are also consistent with previous studies. For example, Tong et al. (2005) noted longer residence time for microbes in a sand column due to greater attachment caused by stagnant flow. Therefore, the gradual change in *E. coli* concentration for scenario 2 than for scenario 1 reflects that stagnant flow led to more attachment and longer residence times near the septic tanks toward the lake or canal. Hence, we conclude that these results demonstrate the importance of the flow regime and seasonal variability (climatic) on *E. coli*transport in the subsurface.

### 4.3 Sensitivity Analysis

It was found that the total retention rate coefficient was the most sensitive parameter followed by the saturated hydraulic conductivity. Similarly, in the unsaturated zone, the OFAT approach was implemented altering each parameter by  $\pm 30$  % while keeping the rest of the parameters constant at their assigned values. It was found that *E. coli* concentration was largely affected by three parameters: saturated water content, total retention coefficient, and dispersivity.

Hydraulic conductivity is an important parameter for *E. coli* transport in the subsurface, and hydraulic conductivity is a function of soil water content in the unsaturated zone (van Genuchten <u>1980</u>). Therefore, we notice that saturated water content and hydraulic conductivity are important parameters in the unsaturated zone and GW, respectively. The total retention rate coefficient is an important parameter for both the unsaturated zone and GW. These findings are congruous with previous studies found in the literature. For instance, a similar study, under variably saturated conditions, conducted by Long and Or (<u>2007</u>) reported diffusion as an important parameter for microbial deposition, which, in turn, is controlled by the soil water content. In addition, Or et al. (<u>2007</u>) demonstrated advective transport in the subsurface as the major mechanism for *E. coli* transport. Another study conducted by Powelson and Mills (<u>2001</u>) also demonstrated

the impact of saturated water content and the retention rate on microbial transport and deposition.

# 5 Application of the Study Findings to Minimize Contamination of Water Resources

This study emphasizes the importance of flow paths and seasonal variability on *E. coli* transport in the subsurface. Our findings suggest that recharging and discharging hydrologic conditions strongly impact *E. coli* transport; these findings have significance for other environments with E. coli contamination. In real-world scenarios, the risk of contamination to water resources from *E. coli* can be minimized by correctly estimating the setback distance. Our study identifies the most sensitive parameters (e.g., hydraulic conductivity) with which a range of the setback distance under different conditions (e.g., soil type, temperature) can easily be estimated. For example, to objectively quantify the setback distance from septic tanks, *E. coli* inactivation or die-off rate is a key factor under variably saturated conditions. E. coli inactivation depends on various biotic and abiotic factors. The abiotic factors such as temperature, pore velocity, and oxygen show large variability in times ranging from hours, days, months, to seasons (Logan et al. <u>1995;</u> Saiers <u>2005</u>; Foppen and Schijven <u>2006</u>). Other abiotic factors, such as grain size, porosity, and soil type, show large spatial variability from pore to field scales (Tufenkji et al. 2004; Tufenkji 2007). We must note that *E. coli* movement in the subsurface under variably saturated conditions may range from submeter to a few meters. Data on these factors may not be available in different field conditions at present; however, we can estimate a range of the setback distance by perturbing the most sensitive parameters (e.g., saturated water content of the soil, total retention rate coefficient, hydraulic conductivity). Additionally, optimal monitoring locations can also be designed to enable informed decisions for safeguarding water resources across the world.

# 6 Limitations and Scope of the Study

To appropriately describe the fate and transport of *E. coli* at the field scale, it is important to determine the level of complexity needed in the modeling framework. Currently, it is not possible to incorporate the effect of various biotic (grazing, competition posed by protozoa) and abiotic (e.g., temperature, pore velocity, oxygen) factors in the modeling framework at the field scale due to unavailability of observations, difficulty in measurements, and inherent stochasticity in these factors. For keeping up with the goal of this study, which was to evaluate *E. coli* transport in the subsurface zone under recharging-discharging GW, *E. coli* inactivation or die-off was simplified and represented by the total retention rate coefficient.

We believe that our conceptual model for describing the *E. coli* transport in SW-SoW-GW represents the adequate complexity and is, therefore, robust to this simplification. However, it is desirable to include more complexity in the modeling framework in future studies to minimize uncertainty in *E. coli* estimation, which may be possible through the availability of more data in the future.

# 7 Conclusions

A major goal of this research was to investigate E. coli transport in the subsurface by considering saturated zone processes only and then by linking the surface water-soil water-groundwater system. Two hydrologic scenarios based on expected water level variations in the lake and canal were implemented to investigate seasonal controls on *E. coli* transport in the subsurface in the Lake Granbury area. The loosely coupled HYDRUS and MODFLOW/MT3DMS models were able to produce a reasonably good agreement between the estimated and observed *E. coli* concentrations in the lake. Results indicate that although groundwater and surface water system largely control E. coli transport in the subsurface, temporal variability of *E. coli* can be described by linking the SW-SoW-GW system. Results further show slightly less removal of *E. coli* in recharging GW months as compared to discharging GW months. It was found that 90 % E. coli are retained in the soil matrix within a depth of 0.5 m from the source (septic tanks). Water content, total retention rate coefficient, and hydraulic conductivity are important parameters for describing *E. coli* transport in the subsurface. The flow paths in the Lake Granbury area indicate that the canal is more at risk of impairment than the lake. However, if this condition persists, in time, the main body of the lake will also be at risk. These results are useful to decision makers and environmental managers to design targeted monitoring programs by incorporating seasonality and supporting real-time decision-making.

## Notes

Acknowledgments

This research was supported by EPA 319(h) grant for TMDL in Texas streams and partly supported by the National Institute of Environmental Health Sciences (grant 5R01ES015634), Texas Water Resources Institute, and Texas A&M support a/c 02-130003. The content is solely the responsibility of the authors and does not necessarily represent the official views of the funding agencies.

## References

- 1. Anderson, J. M. (1991). The effects of climate change on decomposition processes in grassland and coniferous forests. *Ecological Applications*, *1*, 326–347.<u>CrossRefGoogle Scholar</u>
- Anderson, M. P., & Munter, J. A. (1981). Seasonal reversals of groundwater flow around lakes and the relevance to stagnation points and lake budgets. *Water Resources Research*, 17, 1139.<u>CrossRefGoogle</u> <u>Scholar</u>
- 3. Arora, B., Mohanty, B.P., and McGuire, J.T. (2012). Uncertainty in dual permeability model parameters for structured soils. *Water Resources Research, 48*.<u>Google Scholar</u>
- Bergendahl, J., & Grasso, D. (2000). Prediction of colloid detachment in a model porous media: hydrodynamics. *Chemical Engineering Science*, 55(9), 1523–1532. Available at <u>http://www.sciencedirect.com/science/article/pii/S0009250999004224</u> (verified 23 December 2014).<u>CrossRefGoogle Scholar</u>
- Bethune, D. N., Farvolden, R. N., Ryan, M. C., & Guzman, A. L. (1996). Industrial contamination of a municipal water-supply lake by induced reversal of ground-water flow, Managua, Nicaragua. *Ground Water, 34*, 699–708. Available at <Go to ISI>://WOS:A1996UV86600020.<u>CrossRefGoogle Scholar</u>
- 6. Bhattacharjee, S., Ryan, J. N., & Elimelech, M. (2002). Virus transport in physically and geochemically heterogeneous subsurface porous media. *Journal of Contaminant Hydrology*, *57*, 161–187.<u>CrossRefGoogle</u> <u>Scholar</u>
- Bouwer, H., Lange, J. C., & Riggs, M. S. (1974). High-rate land treatment I: infiltration and hydraulic aspects of the Flushing Meadows project. *Journal of the Water Pollution Control Federation*, 46, 834–843. Available at http://www.jstor.org/discover/10.2307/25038728? sid=21105777816983&uid=3739256&uid=4&uid=3739560&uid=2 (verified 3 February 2015).<u>Google Scholar</u>
- Bradford, S. A., & Bettahar, M. (2005). Straining, attachment, and detachment of oocysts in saturated porous media. *Journal of Environmental Quality*, 34(2), 469. Available at <u>https://dl.sciencesocieties.org/publications/jeq/abstracts/34/2/0469</u> (v erified 4 January 2015).<u>CrossRefGoogle Scholar</u>
- Bradford, S. A., Simunek, J., Bettahar, M., Van Genuchten, M. T., & Yates, S. R. (2003). Modeling colloid attachment, straining, and exclusion in saturated porous media. *Environmental Science & Technology*, 37, 2242–2250.<u>CrossRefGoogle Scholar</u>

- Bradford, S.A., Simunek, J., and Walker, S.L. (2006). Transport and straining of E. coli O157:H7 in saturated porous media. *Water Resources Research*, 42.Google Scholar
- 11. Butler, R. G., Orlob, G. T., & McGauhey, P. H. (1954). Underground movement of bacterial and chemical pollutants. *Journal of American Water Works Association, 46*, 97–111.<u>Google Scholar</u>
- Cunningham, A., Characklis, W.G., Abedeen, F., Crawford, D. (1991). Influence of biofilm accumulation on porous media hydrodynamics. *Environmental Science & Technology*, 25, 1305–1311. Available at http://pubs.acs.org/doi/abs/ <u>10.1021/es00019a013</u>.
- 13. Davis, R. O. E., & Bennett, H. H. (1927). *Grouping of soils on the basis of mechanical analysis*. Washington: U.S. Department of Agriculture.<u>Google Scholar</u>
- DeFlaun, M. F., Murray, C. J., Holben, W., Scheibe, T., Mills, A., Ginn, T., Griffin, T., Majer, E., & Wilson, J. L. (1997). Preliminary observations on bacterial transport in a coastal plain aquifer. *FEMS Microbiology Reviews*, 20, 473-487.<u>CrossRefGoogle Scholar</u>
- 15. Dwivedi, D., and Mohanty, B.P. (2016). Hot spots and persistence of nitrate in aquifers across scales. *Entropy*, *18*(1).<u>Google Scholar</u>
- Dwivedi, D., Mohanty, B.P., and Lesikar, B.J. (2008). E. coli fate and transport below subsurface septic tanks in Lake Granbury area. *In* ASA-CSSA-SSSA International Annual Meeting, Houston, TX, 5–9 Oct.<u>Google</u> <u>Scholar</u>
- Dwivedi, D., Mohanty, B. P., & Lesikar, B. J. (2013). Estimating Escherichia coli loads in streams based on various physical, chemical, and biological factors. *Water Resources Research*, 49, 2896– 2906.<u>CrossRefGoogle Scholar</u>
- Eaton, A. D., Clesceri, L. S., Greenberg, A. E., & Franson, M. A. H. (1988). Standard methods for the examination of water and wastewater. Washington: American Public Health Association.<u>Google Scholar</u>
- 19. Federle, T. W., Dobbins, D. C., Thorntonmanning, J. R., & Jones, D. D. (1986). Microbial biomass, activity, and community structure in subsurface soils. *Ground Water, 24*, 365–374. CrossRefGoogle Scholar
- 20. Foppen, J. W. A., & Schijven, J. F. (2006). Evaluation of data from the literature on the transport and survival of Escherichia coli and thermotolerant coliforms in aquifers under saturated conditions. *Water Research*, 40(3), 401–426. Available

at <u>http://www.ncbi.nlm.nih.gov/pubmed/16434075</u> (verified 9 December 2014).<u>CrossRefGoogle Scholar</u>

- Foppen, J. W. A., Mporokoso, A., & Schijven, J. F. (2005). Determining straining of Escherichia coli from breakthrough curves. *Journal of Contaminant Hydrology*, 76(3-4), 191–210. Available at <u>http://www.sciencedirect.com/science/article/pii/S0169772204001512</u> (verified 23 December 2014).<u>CrossRefGoogle Scholar</u>
- 22. Foppen, J. W., van Herwerden, M., & Schijven, J. (2007). Measuring and modelling straining of Escherichia coli in saturated porous media. *Journal of Contaminant Hydrology*, 93, 236–254.<u>CrossRefGoogle</u><u>Scholar</u>
- Gagliardi, J. V., & Karns, J. S. (2000). Leaching of Escherichia coli 0157 :H7 in diverse soils under various agricultural management practices. *Applied and Environmental Microbiology*, 66, 877–883. Available at ISI:000085604800001.<u>CrossRefGoogle Scholar</u>
- Gelhar, L. W., Welty, C., & Rehfeldt, K. R. (1992). A critical review of data on field-scale dispersion in aquifers. *Water Resources Research, 28*, 1955–1974.<u>CrossRefGoogle Scholar</u>
- 25. Harmel, R. D., King, K. W., Richardson, C. W., & Williams, J. R. (2003). Long-term precipitation analyses for the central Texas Blackland Prairie. *Transactions of ASAE, 46*(5), 1381.<u>CrossRefGoogle Scholar</u>
- Harvey, R., & Garabedian, S. (1991). Use of colloid filtration theory in modeling movement of bacteria through a contaminated sandy aquifer. *Environmental Science & Technology, 25*, 178–185. doi: <u>10.1021/es00013a021</u>.<u>CrossRefGoogle Scholar</u>
- 27. Haznedaroglu, B. Z., Bolster, C. H., & Walker, S. L. (2008). The role of starvation on Escherichia coli adhesion and transport in saturated porous media. *Water Research*, *42*, 1547–1554. <u>CrossRefGoogle Scholar</u>
- Hendry, M. J., Lawrence, J. R., & Maloszewski, P. (1999). Effects of velocity on the transport of two bacteria through saturated sand. *Ground Water*, 37, 103–112. Available at <Go to ISI>://000078117900017.<u>CrossRefGoogle Scholar</u>
- Jamieson, R. C., Gordon, R. J., Tattrie, S. C., & Stratton, G. W. (2003). Sources and persistence of fecal coliform bacteria in a rural watershed. Water Quality Research Journal of Canada, 38, 33–47. Google Scholar

- Jiang, G., Noonan, M. J., Buchan, G. D., & Smith, N. (2007). Transport of Escherichia coli through variably saturated sand columns and modeling approaches. *Journal of Contaminant Hydrology*, 93, 2–20.<u>CrossRefGoogle</u> <u>Scholar</u>
- Johnson, P. R., & Elimelech, M. (1995). Dynamics of colloid deposition in porous media: blocking based on random sequential adsorption. *Langmuir*, 11, 801–812. doi: <u>10.1021/la00003a023</u>.<u>CrossRefGoogle Scholar</u>
- Klute, A., & Dirksen, C. (1986). Hydraulic conductivity and diffusivity. Laboratory methods. p. 687–734. *In* Methods of soil analysis—part 1. Physical and mineralogical methods.<u>Google Scholar</u>
- 33. Lesikar, B., Hallmark, C., Melton, R., & Harris, B. (2005). *On-site wastewater treatment systems: soil particle analysis procedure*. Texas Cooperative Extension, Texas A&M University System (p. 21).<u>Google</u> <u>Scholar</u>
- Lindqvist, R., & Bengtsson, G. (1991). Dispersal dynamics of groundwater bacteria. *Microbial Ecology*, 21, 49–72.<u>CrossRefGoogle</u> <u>Scholar</u>
- 35. Lindqvist, R., Cho, J. S., & Enfield, C. G. (1994). A kinetic model for cell density dependent bacterial transport in porous media. *Water Resources Research, 30*, 3291–3299. Available at <Go to ISI>://WOS:A1994PU14200007.CrossRefGoogle Scholar
- Logan, B. E., Jewett, D. G., Arnold, R. G., Bouwer, E. J., & O'Melia, C. R. (1995). Clarification of clean-bed filtration models. *Journal of Environmental Engineering*, 121(12), 869–873. doi: <u>10.1061/</u> (ASCE)0733-9372(1995)121:12(869) (verified 4 January 2015).<u>CrossRefGoogle Scholar</u>
- 37. Long, T., & Or, D. (2007). Microbial growth on partially saturated rough surfaces: simulations in idealized roughness networks. *Water Resources Research*, 43.Google Scholar
- Lowe, K. S., & Siegrist, R. L. (2008). Controlled field experiment for performance evaluation of septic tank effluent treatment during soil infiltration. *Journal of Environmental Engineering*, 134, 93– 101.<u>CrossRefGoogle Scholar</u>
- 39. Mace, R.E., C.A. H., A. R., and Way, S. C. (2000). Groundwater availability of the Trinity Aquifer, Hill Country Area, Texas—numerical simulations through 2050.<u>Google Scholar</u>

- 40. Mallants, D., Mohanty, B. P., Vervoort, A., & Feyen, J. (1997). Spatial analysis of saturated hydraulic conductivity in a soil with macropores. *Soil Technology*, *10*, 115–131.<u>CrossRefGoogle Scholar</u>
- Matthess, G., Pekdeger, A., & Schroeter, J. (1988). Persistence and transport of bacteria and viruses in groundwater—a conceptual evaluation. *Journal of Contaminant Hydrology*, 2(2), 171–188. Available at <u>http://www.sciencedirect.com/science/article/pii/016977228890006X</u> ( verified 3 January 2015).<u>CrossRefGoogle Scholar</u>
- 42. McCaulou, D. R., Bales, R. C., & Arnold, R. G. (1995). Effect of temperature-controlled motility on transport of bacteria and microspheres through saturated sediment. *Water Resources Research*, 31, 271.<u>CrossRefGoogle Scholar</u>
- McMahon, P. B., Tindall, J. A., Collins, J. A., Lull, K. J., & Nuttle, J. R. (1995). Hydrologic and geochemical effects on oxygen uptake in bottom sediments of an effluent-dominated river. *Water Resources Research*, 31, 2561–2569.<u>CrossRefGoogle Scholar</u>
- 44. Milford, M. H. (1997). *Introduction to soils and soil science laboratory exercises*. Dubuque: Kendall/Hunt Publishing Company.<u>Google Scholar</u>
- 45. Murphy, E. M., & Ginn, T. R. (2000). Modeling microbial processes in porous media. *Hydrogeology Journal*, *8*, 142–158. CrossRefGoogle Scholar
- Or, D., Smets, B. F., Wraith, J. M., Dechesne, A., & Friedman, S. P. (2007). Physical constraints affecting bacterial habitats and activity in unsaturated porous media—a review. *Advances in Water Resources*, 30, 1505–1527.<u>CrossRefGoogle Scholar</u>
- 47. Pachepsky, Y. A., & Shelton, D. R. (2011). Escherichia coli and fecal coliforms in freshwater and estuarine sediments. *Critical Reviews in Environmental Science and Technology*, *41*, 1067–1110.<u>CrossRefGoogle</u><u>Scholar</u>
- Pachepsky, Y. A., Sadeghi, A. M., Bradford, S. A., Shelton, D. R., Guber, A. K., & Dao, T. (2006). Transport and fate of manure-borne pathogens: modeling perspective. *Agricultural Water Management*, 86, 81– 92.<u>CrossRefGoogle Scholar</u>
- Pang, L., Close, M., Goltz, M., Sinton, L., Davies, H., Hall, C., & Stanton, G. (2004). Estimation of septic tank setback distances based on transport of E. coli and F-RNA phages. *Environment International, 29*, 907–921.<u>CrossRefGoogle Scholar</u>

- 50. Pang, L., Nokes, C., Šimůnek, J., Kikkert, H., & Hector, R. (2006). Modeling the impact of clustered septic tank systems on groundwater quality. *Vadose Zone Journal, 5*, 599.<u>CrossRefGoogle Scholar</u>
- Personne, J. C., Poty, F., Vaute, L., & Drogue, C. (1998). Survival, transport and dissemination of Escherichia coli and enterococcci in a fissured environment. Study of a flood in a karstic aquifer. *Journal of Applied Microbiology*, 84(3), 431–438. doi: <u>10.1046/j.1365-</u> <u>2672.1998.00366.x</u> (verified 14 January 2015).<u>CrossRefGoogle Scholar</u>
- Powelson, D. K., & Mills, A. L. (2001). Transport of in sand columns with constant and changing water contents. *Journal of Environmental Quality*, 30(1), 238. Available at <u>https://dl.sciencesocieties.org/publications/jeq/abstracts/30/1/238</u> (ver ified 14 January 2015).<u>CrossRefGoogle Scholar</u>
- Riebschleager, K. J., Karthikeyan, R., Srinivasan, R., & McKee, K. (2012). Estimating potential E. coli sources in a watershed using spatially explicit modeling techniques. *JAWRA Journal of the American Water Resources Association*, 48(4), 745–761. doi: <u>10.1111/j.1752-</u> <u>1688.2012.00649.x</u> (verified 2 December 2014).<u>CrossRefGoogle Scholar</u>
- Ryan, J. N., & Elimelech, M. (1996). Colloid mobilization and transport in groundwater. *Colloids and Surfaces A: Physicochemical and Engineering Aspects, 107*(95), 1–56. Available at <u>http://linkinghub.elsevier.com/retrieve/pii/092777579503384X</u>.CrossR <u>efGoogle Scholar</u>
- 55. Saiers, J.E. (2005). Correction to "Ionic-strength effects on colloid transport and interfacial reactions in partially saturated porous media." *Water Resources Research, 41*.<u>Google Scholar</u>
- 56. Šimůnek, J., van Genuchten, M. Th., & Šejna, M. (2006). The HYDRUS software package for simulating two- and three-dimensional movement of water, heat, and multiple solutes in variably-saturated media (version 1.0, edited, PC Progress, Prague, Czech Republic.).<u>Google Scholar</u>
- Sinton, L. W., Finlay, R. K., Pang, L., & Scott, D. M. (1997). Transport of bacteria and bacteriophages in irrigated effluent into and through an alluvial gravel aquifer. *Water, Air, and Soil Pollution, 98*(1-2), 17–42. doi: <u>10.1023/A%3A1026492110757</u> (verified 12 December 2014).<u>CrossRefGoogle Scholar</u>
- Sinton, L. W., Noonan, M. J., Finlay, R. K., Pang, L., & Close, M. E. (2000). Transport and attenuation of bacteria and bacteriophages in an alluvial gravel aquifer. *New Zealand Journal of Marine and Freshwater Research*, 34(1), 175–186.

doi: <u>10.1080/00288330.2000.9516924</u> (verified 12 December 2014).<u>CrossRefGoogle Scholar</u>

- 59. Smith, M. S., Thomas, G. W., White, R. E., & Ritonga, D. (1985). Transport of Escherichia coli through intact and disturbed soil columns. *Journal of Environmental Quality, 14*, 87.<u>CrossRefGoogle</u> <u>Scholar</u>
- 60. Spalding, R. F., & Exner, M. E. (1993). Occurrence of nitrate in groundwater—a review. *Journal of Environmental Quality, 22*, 392.<u>CrossRefGoogle Scholar</u>
- 61. Sun, N., Elimelech, M., Sun, N. Z., & Ryan, J. N. (2001). A novel twodimensional model for colloid transport in physically and geochemically heterogeneous porous media. *Journal of Contaminant Hydrology, 49*, 173–199.<u>CrossRefGoogle Scholar</u>
- 62. Tan, Y., Gannon, J. T., Baveye, P., & Alexander, M. (1994). Transport of bacteria in an aquifer sand: experiments and model simulations. *Water Resources Research*, *30*, 3243. CrossRefGoogle Scholar
- Tong, M., Camesano, T. A., & Johnson, W. P. (2005). Spatial variation in deposition rate coefficients of an adhesion-deficient bacterial strain in quartz sand. *Environmental Science & Technology, 39*, 3679– 3687.<u>CrossRefGoogle Scholar</u>
- 64. Torkzaban, S., Hassanizadeh, S. M., Schijven, J. F., de Bruin, H. A. M., & de Roda Husman, A. M. (2006). Virus transport in saturated and unsaturated sand columns. *Vadose Zone Journal*, *5*(3), 877. Available at <u>https://www.soils.org/publications/vzj/abstracts/5/3/877?</u> access=0&view=pdf (verified 13 December 2014).<u>CrossRefGoogle</u>Scholar
- 65. Tufenkji, N. (2007). Modeling microbial transport in porous media: traditional approaches and recent developments. *Advances in Water Resources, 30*, 1455–1469.<u>CrossRefGoogle Scholar</u>
- Tufenkji, N., Miller, G. F., Ryan, J. N., Harvey, R. W., & Elimelech, M. (2004). Transport of cryptosporidium oocysts in porous media role of straining and physicochemical. *Environmental Science & Technology*, 38(22), 5932–5938.<u>CrossRefGoogle Scholar</u>
- Twarakavi, N. K. C., Šimůnek, J., & Seo, S. (2008). Evaluating interactions between groundwater and vadose zone using the HYDRUSbased flow package for MODFLOW. *Vadose Zone Journal*, 7, 757.<u>CrossRefGoogle Scholar</u>

- 68. Twarakavi, N.K.C., Šimůnek, J., and Schaap, M.G. (2010). Can texturebased classification optimally classify soils with respect to soil hydraulics? *Water Resources Research*, *46*.<u>Google Scholar</u>
- 69. USEPA. (2002). Onsite wastewater treatment systems manual. EPA/625/R-00/008.<u>Google Scholar</u>
- USEPA. (2005). <u>http://www.epa.gov/owm/septic/pubs/onsite\_handbook.pdf</u>.
- 71. van Genuchten, M. T. (1980). A closed-form equation for predicting the hydraulic conductivity of unsaturated soils. *Soil Science Society of America Journal, 44*, 892.<u>CrossRefGoogle Scholar</u>
- 72. Wan, J., & Wilson, J. L. (1994). Visualization of the role of the gas-water interface on the fate and transport of colloids in porous media. *Water Resources Research, 30*, 11.<u>CrossRefGoogle Scholar</u>
- 73. Williams, A. E., Johnson, J. A., Lund, L. J., & Kabala, Z. J. (1998). Spatial and temporal variations in nitrate contamination of a rural aquifer, California. *Journal of Environmental Quality, 27*, 1147.<u>CrossRefGoogle</u> <u>Scholar</u>
- 74. Zhang, P., Johnson, W. P., Scheibe, T. D., Choi, K. H., Dobbs, F. C., & Mailloux, B. J. (2001). Extended tailing of bacteria following breakthrough at the Narrow Channel focus area, Oyster, Virginia. Water Resources Research, 37, 2687–2698. CrossRefGoogle Scholar