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Authors

Zhang, William R
Scherzer, Rebecca
Estrella, Michelle M
et al.

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Tenofovir disoproxil fumarate initiation and changes in urinary biomarker concentrations among HIV-infected men and women

William R. ZHANG¹, Rebecca SCHERZER¹, Michelle M. ESTRELLA¹, Simon B. ASCHER^{1,2}, Anthony MUIRU¹, Vasantha JOTWANI¹, Carl GRUNFELD¹, Chirag R. PARIKH³, Deborah GUSTAFSON⁴, Seble KASSAYE⁵, Anjali SHARMA⁶, Mardge COHEN⁷, Phyllis C. TIEN⁸, Derek K. NG⁹, Frank J. PALELLA Jr.¹⁰, Mallory D. WITT¹¹, Ken HO¹², and Michael G. SHLIPAK¹

¹Kidney Health Research Collaborative, Department of Medicine, San Francisco Veterans Affairs Health Care System, and University of California, San Francisco, CA

²Department of Medicine, University of California, Los Angeles, CA

³Program of Applied Translational Research, Department of Medicine, Section of Nephrology, Yale University, New Haven, CT and Department of Veterans Affairs Medical Center, West Haven, CT

⁴Department of Medicine, SUNY Downstate Medical Center, Brooklyn, NY

⁵Georgetown University Medical Center, Washington, DC

⁶Department of Medicine, Albert Einstein College of Medicine, Bronx, NY

⁷Department of Medicine, Stroger Hospital and Rush University, Chicago, IL

⁸Department of Medicine, Division of Infectious Disease, University of California, San Francisco and Department of Veterans Affairs Health Care System, San Francisco, CA

⁹Department of Epidemiology, Johns Hopkins Bloomberg School of Public Health, Baltimore, MD

¹⁰Department of Medicine, Division of Infectious Disease, Northwestern University Feinberg School of Medicine, Chicago, IL

¹¹Division of HIV Medicine, Department of Medicine, Los Angeles Biomedical Research Institute at Harbor-UCLA Medical Center, Torrance, California.

¹²Division of Infectious Diseases, Department of Medicine, University of Pittsburgh, Pittsburgh, PA

Abstract

Objectives: Urinary biomarkers of kidney injury may have potential to identify subclinical injury attributable to tenofovir disoproxil fumarate (TDF) toxicity.

Design: This observational study included 198 HIV-infected participants from the Multicenter AIDS Cohort Study and the Women's Interagency HIV Study, who initiated TDF between 2009 and 2015 and had urine samples collected at baseline before and after TDF initiation.

Methods: We used linear mixed effects models controlling for urine creatinine and time on TDF to evaluate the effects of TDF initiation on changes in fourteen urinary biomarkers.

Results: Within 1 year after TDF initiation, concentrations of trefoil factor 3 (+78%; 95% CI: +38%, +129%), alpha-1 microglobulin (α 1m) (+32%; 95% CI: +13%, 55%), clusterin (+21%; 95% CI: +6%, +38%), uromodulin (+19%; 95% CI: +4, +36%), and kidney injury molecule-1 (KIM-1) (+13%; 95% CI: +1%, +26%) significantly increased, whereas interleukin-18 (IL-18) significantly decreased (−13%, 95% CI: −7%, −25%). Subsequent to the first year of TDF use, biomarker concentrations stabilized, and these changes were not statistically significant. When stratifying by baseline viremia (HIV-1 RNA < vs. ≥ 80 copies/mL), concentration changes for most biomarkers during the first year of TDF use were greater among aviremic versus viremic participants, with significant differences in α 1m (+80% vs. +22%), KIM-1 (+43% vs. +10%), beta-2 microglobulin (+83% vs. −10%), YKL-40 (+33% vs. −5%), and IL-18 (+20% vs. −27%).

Conclusions: TDF initiation was associated with substantial changes in urinary biomarkers of kidney injury within the first year of use, particularly among aviremic participants. A urinary biomarker panel may be a clinically useful tool to detect and monitor the heterogeneous effects of TDF on the kidney.

Keywords

tenofovir disoproxil fumarate (TDF); nephrotoxicity; HIV; biomarkers; kidney

Introduction

Tenofovir disoproxil fumarate (TDF) is widely utilized as part of first-line antiretroviral therapy (ART) for HIV treatment regimens worldwide.^[1] While early trials reported a favorable safety profile, TDF is now recognized to be associated with increased risk of acute and chronic kidney disease.^[2] However, serum creatinine and urine protein, the clinical standards for assessing kidney disease in HIV-infected persons,^[3] are crude and insensitive measures of kidney damage that are not specific to etiology. This presents significant challenges to effective surveillance of TDF-associated nephrotoxicity, which is often detected late in the disease course when injury may be irreversible and cannot be distinguished from the numerous other kidney disease risk factors in the HIV-infected population.^[4, 5]

Studies in the general population and in HIV-infected persons have shown that urinary biomarkers of kidney tubule injury can be useful for detecting early stages of kidney disease and predicting the onset of CKD and its complications. Because TDF toxicity is known to predominantly involve the proximal renal tubules,^[6, 7] biomarkers of tubular pathology have the potential to detect subclinical kidney injury attributable to TDF^[8] and would be more sensitive and specific than current indirect markers of proximal tubular dysfunction, such as fractional excretion of phosphate, uric acid, and glycosuria.^[9] To date, however, only a limited number of biomarkers of kidney injury have been investigated in HIV-infected TDF-users, which have largely been in cross-sectional studies.^[10-18]

Using stored urine samples from HIV-infected participants in the Multicenter AIDS Cohort Study (MACS) and Women Interagency HIV Study (WIHS) collected before and after TDF initiation, we conducted a longitudinal, observational study of new users of TDF. Our objective was to evaluate the association of TDF initiation with a panel of fourteen urinary biomarkers that is anatomically representative of the nephron and mechanistically diverse. Specifically, this panel included biomarkers of glomerular injury (albumin-creatinine ratio, osteopontin), proximal tubular injury (trefoil factor 3, clusterin, kidney injury molecule-1, neutrophil gelatinase-associated lipocalin, interleukin-18), tubular fibrosis and repair (monocyte chemoattractant protein-1, epidermal growth factor, chitinase-3-like protein-1), proximal tubular dysfunction (cystatin C, α 1-microglobulin, β 2-microglobulin), and loop of Henle function (uromodulin). We hypothesized that TDF initiation would be associated with higher concentrations of kidney injury biomarkers, particularly those associated with proximal tubular pathology.

Materials and Methods

Study Design and Population

This study included 198 HIV-infected, initially TDF-naïve participants from MACS (n=87) and WIHS (n=111), who had a urine sample collected before TDF initiation (“baseline”) and at least one sample collected after TDF initiation while continuing to use TDF (Figure 1). Participants contributed a median of 3 samples (interquartile range [IQR]: 2-4). The MACS is an ongoing, prospective cohort study established in 1984 to describe the natural history of HIV infection among men who have sex with men.^[19] Participants were enrolled from four sites in the United States: Baltimore, MD; Chicago, IL; Los Angeles, CA; and Pittsburgh, PA. The WIHS is a multicenter, prospective cohort study established in 1993 to investigate the progression of HIV in women with and at risk for HIV in the United States.^[20, 21] Women were enrolled from eleven sites in the United States: Bronx, NY; Brooklyn, NY; Chicago, IL; Los Angeles, CA; San Francisco, CA; and Washington, DC (enrolled between 1994-1995, 2001-2002, and 2011-2012); and Atlanta, GA; Birmingham, AL; Jackson, MS, Chapel Hill, NC; and Miami, FL (enrolled between 2013-2015). The institutional review boards of participating institutions approved the study protocol, which was adherent to the Declaration of Helsinki.

Exposure

ART use was ascertained for all participants at each semi-annual study visit. For each participant included in this analysis, a “pre-TDF” baseline sample collected between 2009 and 2015 was identified, and “on TDF” samples were identified at subsequent study visits. Median time from pre-TDF urine collection to TDF initiation was 0.99 years (IQR: 0.89-1.18), median time of TDF exposure at the first post-TDF initiation urine collection was 1.0 years (IQR: 0.9-1.4), and median follow-up time from the baseline sample was 2.9 years (range 0.9-4.7).

Urinary Biomarker Outcomes

The outcomes were changes in concentrations of fourteen urinary biomarkers: trefoil factor 3 (TFF3), alpha-1 microglobulin (α 1m), clusterin, uromodulin (UMOD), kidney injury

molecule-1 (KIM-1), beta-2 microglobulin (β 2M), albumin-creatinine ratio (ACR), neutrophil gelatinase-associated lipocalin (NGAL), anti-chitinase-3-like protein 1 (YKL-40), monocyte chemoattractant protein-1 (MCP-1), cystatin C (CysC), osteopontin (OPN), epidermal growth factor (EGF), and interleukin-18 (IL-18). This biomarker panel included all novel biomarkers approved by the Food and Drug Administration (FDA), European Medicines Agency (EMA), and the Japanese Pharmaceutical and Medical Devices Agency (PMDA) for use in pre-clinical trials of drug-induced kidney toxicity.^[22-24]

Urinary biomarkers were measured at the University of Vermont Laboratory for Clinical Biochemistry Research. Most biomarkers were measured using multiplex Meso Scale Discovery immunoassay kits (Meso Scale Diagnostics, LLC, Gaithersburg, MD), except for α 1m, which was measured using the BN II Nephelometer assay (Siemens, Newark, DE). Urine creatinine, which was used to account for urine sample tonicity, was measured using a Cobas c311 clinical analyzer (Roche Diagnostics, Indianapolis, IN). Details regarding assay ranges, sensitivities, and coefficients of variation are shown in Supplementary Table 1. All urine specimens were in continuous storage without previous freeze-thaw until measurement. Laboratory personnel were blinded to clinical information about the participants, and specimens were evaluated in random order.

Statistical Analyses

We first compared within-participant characteristics before initiating TDF to those at the first post-TDF initiation urine measurement using Wilcoxon signed-rank test and McNemar's test for continuous and categorical variables, respectively. Diabetes mellitus was defined as a fasting glucose ≥ 126 mg/dL, hemoglobin A1c $\geq 6.5\%$, or self-reported history of diabetes and diabetes medication use; and hepatitis C virus (HCV) infection was defined by either detectable HCV RNA and/or positive HCV antibody result. eGFR was calculated using the 2009 CKD Epidemiology Collaboration (CKD-EPI) creatinine equation.²³

To examine the association of TDF initiation with changes in biomarker concentrations, we constructed separate linear mixed models for each biomarker, adjusting for urine creatinine and time on TDF. We used random intercepts and slopes for time on TDF and a linear spline for time on TDF with inflection point at 1 year. Given their right-skewed distributions, biomarker concentrations were log-transformed, and results were back transformed to produce estimated annual percentage changes. The detectable limit of the α 1m assay was 0.5 mg/dL, and approximately 15% of urine α 1m values were undetectable. Thus, we used a left-censored linear mixed model to estimate the change in α 1m concentration for each participant and to impute values that were below the limit of detection. We then examined Spearman correlations among first-year biomarker changes and presented the results in a heat map for ease of comprehension.

Next, we compared biomarker concentration changes by baseline plasma viremia (detectable vs. undetectable). Undetectable viral load was defined as HIV-1 RNA < 80 copies/mL due to the assays in use during the study period. Given that viral load has been shown to be associated with kidney function decline,^[25] we hypothesized that there would be differing impacts of TDF in aviremic participants (direct effects of TDF toxicity) vs. viremic participants (effects of both TDF toxicity and benefits of viral suppression of baseline

viremia). Models were constructed separately for each biomarker, controlling for baseline viral load status, time on TDF, interaction by baseline viral load status, and urine creatinine. We estimated absolute biomarker concentrations at baseline and at 1 year after TDF initiation by baseline viremia status, using marginal means from linear mixed models, since samples were not collected at exact yearly intervals. Finally, we compared biomarker concentration changes by both ritonavir (RTV) and atazanavir (ATZ) usage (users vs. non-users, time-updated at each visit), as concurrent use of these medications has been associated with greater kidney function decline^[26-28]; and by African American race, given the increased risk of kidney disease in this population.^[29]

Although this study involved several biomarkers, we did not include formal adjustments for multiple comparisons as we hypothesized that TDF-associated biomarker changes would show a biologically coherent pattern. In particular, we hypothesized that the nephrotoxic effects of TDF would be reflected by increases in concentrations of all biomarkers with the exceptions of EGF^[30] and UMOD,^[31, 32] which have been shown to be protective. This biologically coherent pattern dictates that results should be mutually reinforcing, rather than a series of independent tests; therefore, formal multiple comparisons adjustments, such as the Bonferroni method, would not be appropriate.^[33]

Results

Among the 198 HIV-infected participants in this study, the median age at baseline (pre-TDF) was 48 years, 56% were female, and approximately two-thirds were African American. The prevalence of kidney disease risk factors, including hypertension, diabetes, and Hepatitis C virus infection, did not significantly differ between the pre-TDF and first post-TDF time points (Table 1). After initiating TDF for a median of 1.0 year (IQR: 0.9-1.4), participants had significantly higher CD4 counts and serum albumin levels and a significantly greater proportion were aviremic. In addition, eGFR significantly decreased during the first year of TDF use. We initially compared participants included in our study based on the inclusion criteria of new TDF users with stored urine samples vs. participants who did not meet these criteria. Included participants were older with a smaller proportion having undetectable viral load and using antiretroviral treatments prior to their initiation of TDF. Baseline eGFR and CD4 counts were similar between included and excluded participants.

We first estimated the annual percent change in concentrations of the fourteen biomarkers after TDF initiation (Figure 2, Supplementary Table 2). During the first year after TDF initiation, we observed large changes in several urine biomarkers: TFF3 (+78%, 95% CI: +38%, +129%), α 1m (+32%; 95% CI: +13%, +55%), clusterin (+21%; 95% CI: +6%, +38%), UMOD (+19%; 95% CI: +4%, +36%), and KIM-1 (+13%; 95% CI: +1%, +26%) concentrations significantly increased; β 2M, ACR, NGAL, YKL-40, MCP-1, Cys C, OPN, and EGF concentrations showed smaller, non-statistically significant changes; and IL-18 (-13%; 95% CI: -7%, -25%) concentrations significantly decreased. These biomarker changes coincided with significant rises in serum creatinine (+10%; 95% CI: +7%, +13%) and declines in eGFR (-10%; 95% CI: -7%, -13%) within the first year. Subsequent to the first year of TDF use, annual changes in biomarker concentrations were substantially smaller and did not reach statistical significance. While the reduced number of urine samples after 1

year may have limited the power to detect differences, we found that changes in serum creatinine and eGFR after 1 year were also much smaller and did not reach statistical significance.

To distinguish the direct effects of TDF from those influenced by HIV viral suppression, we estimated first-year biomarker concentration changes for each biomarker, stratified by baseline viral load (undetectable vs. detectable) (Figure 3, Supplementary Table 3). For multiple biomarkers, viremic participants on average had attenuated increases or even decreases in biomarker concentrations, compared with aviremic participants. In particular, tests for interaction demonstrated statistically significant differences between viremic and aviremic participants for $\alpha 1m$, KIM-1, $\beta 2M$, YKL-40, and IL-18. Notably, IL-18 concentrations significantly decreased from baseline among viremic participants (-27% ; 95% CI: -20% , -33%) and significantly increased from baseline among aviremic participants ($+20\%$; 95% CI: $+3\%$, $+39\%$). In addition, the magnitude of changes in biomarker concentrations from baseline was greater in the stratified analyses compared to the non-stratified analyses. In contrast, first-year eGFR declines did not differ by baseline viremia.

We next compared absolute biomarker concentrations at baseline and 1 year after TDF initiation, stratified by baseline viral load status. On average, most biomarker concentrations at baseline were higher among viremic participants, with statistically significant differences only for $\beta 2M$, YKL-40, and IL-18 (Supplementary Table 4). After 1 year of TDF use, the absolute concentrations of most biomarkers, including $\beta 2M$, YKL-40, and IL-18, converged between the subgroups by baseline viremia (Figure 4).

Finally, we evaluated for additional effect modifiers of the association of TDF initiation and first-year biomarker changes. We evaluated the impact of concurrent use of RTV boosting or co-administration of ATZ on the associations of TDF with biomarker changes. For RTV boosting, we observed significant interactions with larger elevations in IL-18 ($+62.3\%$ vs. -20.3% ; p for interaction <0.0001) and OPN ($+31.5\%$ vs. -8.0% ; $p=0.017$) among RTV users ($N=50$), compared with non-users. In contrast, ATZ users ($N=27$) had significant relative decreases of TFF3 (-45.3% vs. $+93.5\%$; $p=0.037$) and UMOD (-32.0% vs. $+18.3\%$; $p=0.025$), compared with non-users. When stratifying the first-year changes by race, African Americans ($N=126$) had significantly lower elevations or declines in several biomarkers, compared with non-African Americans: $\alpha 1m$ ($+17.8\%$ vs. $+72.1\%$; $p=0.026$), $\beta 2M$ (-20.4% vs. $+120.5\%$; $p<0.0001$), and ACR (-12.2% vs. $+27.8\%$; $p=0.0015$).

Finally, we evaluated the inter-relationships among the biomarker changes after TDF initiation. Overall, the first-year biomarker changes were only modestly inter-correlated ($r<0.5$), with the exception of TFF3 and $\beta 2M$ ($r=0.6$), and the vast majority of biomarker change pairs were weakly inter-correlated ($r \leq 0.3$) (Supplementary Figure 1).

Discussion

In this observational study of longitudinally well-characterized HIV-infected men and women, we evaluated the evolution of kidney injury biomarkers among HIV-infected

persons who initiated TDF. Overall, we found that TDF initiation was associated with substantial and distinct changes in biomarker concentrations within the first year that on average coincided with rising serum creatinine levels. To our knowledge, this is the first study to use a comprehensive panel of urinary biomarkers to characterize TDF-associated nephrotoxicity longitudinally. Our findings demonstrate the diverse impact of TDF on the kidney and suggest that TDF toxicity may not be adequately captured by our current clinical measures, nor by any single biomarker. Rather, a biomarker signature may be able to identify subclinical injury before the onset of overt, irreversible disease and to distinguish kidney injury attributable to TDF from the many other potential causes of rising creatinine levels in HIV-infected persons.^[34]

Consistent with the literature and our hypothesis, we found that TDF initiation was predominantly associated with changes in biomarker concentrations that represent proximal tubular pathology (i.e., TFF3,^[24, 35] clusterin,^[24, 36] KIM-1,^[37, 38] IL-18,^[39] α 1m,^[18, 40] and β 2M^[24, 41]). In prior cross-sectional analyses of HIV-infected men in MACS, we found that each year of TDF exposure was independently associated with higher concentrations of α 1m (7.6%), IL-18 (2.7%), KIM-1 (2.5%), and procollagen type III N-terminal propeptide (PIIINP) (2.0%).^[17, 18] Compared to these prior cross-sectional findings, effect sizes were as much as 10-fold larger for several biomarkers in this current longitudinal study. For example, participants on average had 78% increases in TFF3, 32% increases in α 1m, and 13% increases in KIM-1 concentrations 1 year after TDF initiation.

Interestingly, we found a distinct decrease in IL-18 concentrations in the first year following TDF initiation, in contrast to the annual increase of IL-18 concentrations associated with TDF exposure in prior cross-sectional analyses. This discrepancy may be explained by the effect modification of biomarker changes by baseline viral suppression status, which was most striking in IL-18: initially viremic participants, who were the majority (71%) of the cohort, had significantly decreased IL-18 concentrations, while aviremic participants had significantly increased IL-18 concentrations. This divergence when stratifying by baseline viral suppression status is consistent with the strong association of HIV viral load with urinary IL-18 concentrations.^[34] In the prior cross-sectional study, a large portion of HIV-infected participants were on ARVs for several years and were likely predominantly aviremic, which may explain the increases in IL-18 concentrations associated with TDF exposure.

In addition, stratification by viral load revealed associations of TDF initiation with changes of varying magnitudes in biomarkers of various nephron sites and functions. For example, in aviremic participants, TDF initiation was associated with significant increases in UMOD, a marker of Loop of Henle function,^[31, 32] and YKL-40, a marker of renal fibrosis and repair.^[42] Overall, aviremic participants had the largest biomarker changes, including over 80% increases in TFF3, α 1m, and β 2M after 1 year of TDF use. In contrast, baseline viremic participants consistently had attenuated biomarker increases or even declines, compared with aviremic participants. While the observed effect modification by viral load distinguished the relative differences in TDF toxicity, absolute biomarker concentrations in these two groups converged after 1 year of TDF use. One possible explanation for these findings is that the benefits of viral suppression induced by TDF-containing ART regimens offset or even

outweighed the harms of TDF toxicity on the kidneys of viremic individuals. Notably, eGFR did not differ between the baseline viremia strata, demonstrating the potential of these biomarkers to distinguish etiologies of injury beyond that of current clinical paradigms.

We also observed that TDF initiation was associated with biomarker changes within the first year of initiation that neither significantly progressed nor recovered over a median of 3 years of TDF use. These findings are consistent with the ASSERT trial, which examined longitudinal urinary biomarker changes among 385 HIV-infected participants randomized to a TDF-containing ART regimen or an abacavir-containing regimen.^[43, 44] Participants who received TDF-containing regimens had substantially higher increases of retinol binding protein (RBP) and β 2M, markers of proximal tubular dysfunction, compared to users of abacavir-containing regimens. These relative biomarker concentration increases were noted at 6 months and remained stable through 2 years of follow-up. The observed kinetics of biomarker changes and kidney function decline within the first year following TDF initiation are also concordant with reported TDF-associated eGFR_{SCr} declines that subsequently plateau at 6 months to 1 year.^[45, 46] In another study using this cohort of new TDF users, we have demonstrated that 6 of these 14 biomarkers, including β 2M, KIM-1, clusterin, UMOD, cystatin C, and IL-18, were independently associated with eGFR decline during follow-up.^[47]

The recent introduction of tenofovir alafenamide fumarate (TAF), a less nephrotoxic alternative to TDF, has eased some concerns regarding tenofovir-associated nephrotoxicity. However, TAF has not completely replaced TDF nor the need for novel kidney function diagnostic methods. Only TDF-containing regimens are currently approved for pre-exposure prophylaxis, and use of TDF remains widespread for the treatment of HIV infection in resource-challenged areas.^[1] It is projected that in 2020, TDF will hold 80% of the market share of first-line treatment regimens in low-to-middle income countries that disproportionately bear the burden of HIV infection.^[48] Furthermore, the long-term kidney effects of TAF remain unknown and are critical to understand, as antiretroviral regimens are often life-long medications. While a recent meta-analysis of six trials reported that TAF use was associated with significantly lower declines in eGFR_{SCr} and lower β 2M and RBP concentrations compared to TDF, TAF was still associated with modest kidney injury and change in eGFR_{SCr}.^[49] In fact, a recent case report described the first documented instance of TAF-associated dysmorphic mitochondria on kidney biopsy, a histologic hallmark of tenofovir toxicity.^[50] The use of novel diagnostics could potentially be useful and applicable to both TDF- and TAF-users to identify incident ART-related toxicities with long-term use.

Strengths of this study include the longitudinal, new-user design that captured incident TDF use and simulated the design of a single-arm, clinical trial.^[51] This also allowed for a pre- vs. post-TDF initiation comparison in which individuals served as their own controls, reducing the bias of inter-individual variability in biomarker concentrations and time-invariant characteristics. In addition, we used a large, curated panel of urinary biomarkers supported by the literature^[24, 52-55] and data from well-characterized cohorts of HIV-infected participants.

We also acknowledge several limitations. Although MACS and WIHS are national, multi-center cohorts, there may be important differences between study participants and the general HIV-infected population, including overall health and medical compliance. While we were unable to directly measure TDF adherence, viral load status served as a proxy and self-reported adherence was generally high for this once daily medication. Phosphaturia and hypophosphatemia have been traditionally considered the earliest clinical markers of renal tubular dysfunction but were not measured in our participants; however, prior studies have demonstrated that biomarkers representing tubular pathology may precede alterations in phosphate excretion in the setting of TDF use.^[56] In addition, individuals who started TDF during the time period of our study but did not have urine samples were excluded, which may limit the generalizability of our findings. Our results may also have been attenuated by biomarker degradation in stored, frozen urine samples, particularly β 2M, which has poor stability in acidic solutions.^[57, 58] Because our study design only allowed for biomarker concentration measurements approximately annually, we may have also missed important changes in biomarker concentrations between these measurements, particularly those within a few months after initiation. Given the decrease in sample size in urine measurements following the first post-TDF initiation measurement, there may have been reduced power to detect subsequent changes. Finally, in our interpretation of the results, we considered both directions and magnitudes of estimates, instead of relying on p-values alone. However, we found that biomarker changes were only moderately inter-correlated, and several had highly significant p-values that would have remained significant even if adjustments for multiple comparisons were appropriate and performed.

Our study has several implications for clinical care and future studies. With validation in other diverse cohorts, uniform use of kidney injury biomarkers could be integrated into clinical decision-making as a novel diagnostic and surveillance strategy. For example, detection of a distinct signature of biomarker changes may help distinguish TDF-associated nephrotoxicity from the other causes of kidney disease among HIV-infected individuals that are currently indistinguishable based upon increases in serum creatinine. Recent studies in animal models have demonstrated that several of these biomarkers outperformed traditional measures of kidney function in detecting the onset and progression of histologically-confirmed TDF-associated tubular injury.^[59] Future studies exploring this methodology should include comparisons of the long-term renal impact of TDF and TAF to inform risk stratification and judicious allocation of the newer formulation of tenofovir. To distinguish tenofovir toxicity from other etiologies of kidney injury, future studies should also characterize longitudinal biomarker profiles associated with other kidney disease risk factors among HIV-infected persons. By extension, urinary biomarker panels may better detect and characterize drug-induced nephrotoxicity and kidney disease in general.

In conclusion, we have demonstrated that TDF initiation is associated with a distinct profile of urinary biomarker concentration changes among HIV-infected men and women. If these findings are validated in future studies, a multi-biomarker panel may be an effective tool to detect and monitor TDF toxicity.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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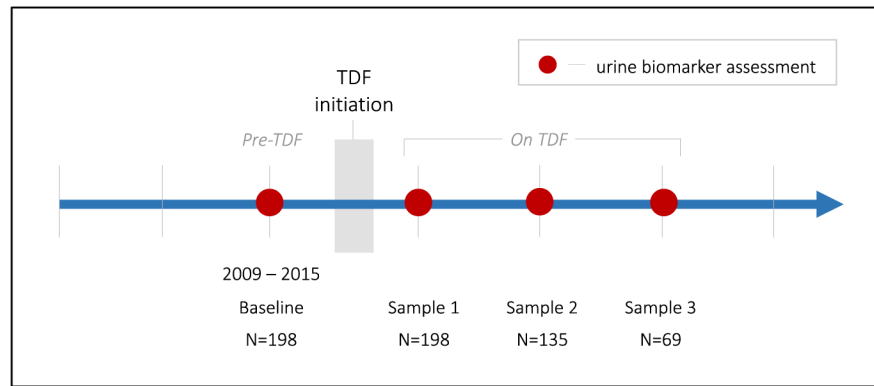


Figure 1. Study design schematic of new TDF-users

Study design schematic of simulated single-arm clinical trial of TDF initiation among HIV-infected men and women. Gray box denotes time of initiation of TDF-containing regimen. Red dots represent urine collections, which occurred at Women Interagency HIV Study (WIHS) and Multicenter AIDS Cohort Study (MACS) annual visits.

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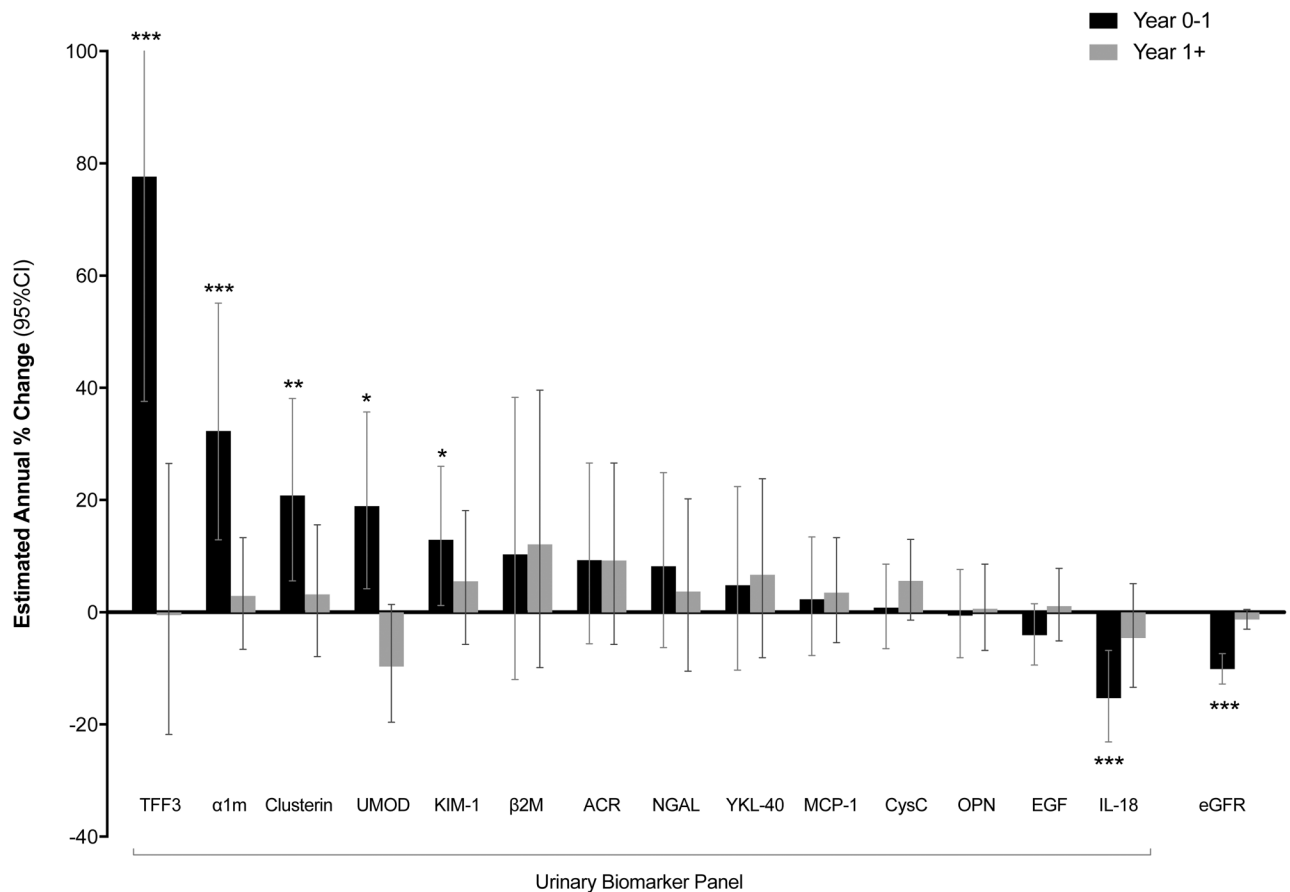


Figure 2. Association of TDF initiation with changes in urinary biomarker concentrations

The black bars (“year 0-1”) denote the relative percent changes in concentrations of each biomarker that occurred during the first year of TDF use. The white bars (“year 1+”) denote the annual percent changes in concentrations of each biomarker that occurred after year 1. Error bars denote the 95% confidence intervals (CIs). Estimated annual percent changes were calculated from separate linear mixed models using all 198 participants, controlling for time on TDF (using linear spline with cutpoint at year 1) and urine creatinine. The y-axis is truncated at 100%. The 95% CI upper bound for change of TFF3 is truncated and extends to 129.1%. * $p < 0.05$. ** $p < 0.01$. *** $p < 0.001$. Numeric values of percent changes for each biomarker, 95% CI, and p-values are presented in Supplementary Table 2. Full names for each biomarker are as follows: trefoil factor 3 (TFF3), α1-microglobulin (α1m), clusterin, uromodulin (UMOD), kidney injury molecule-1 (KIM-1), β2-microglobulin (β2M), albumin-creatinine ratio (ACR), neutrophil gelatinase-associated lipocalin (NGAL), anti-chitinase-3-like protein 1 (YKL-40), monocyte chemoattractant protein-1 (MCP-1), cystatin C (CysC), osteoponin (OPN), epidermal growth factor (EGF), and interleukin-18 (IL-18). eGFR = estimated glomerular filtration rate.

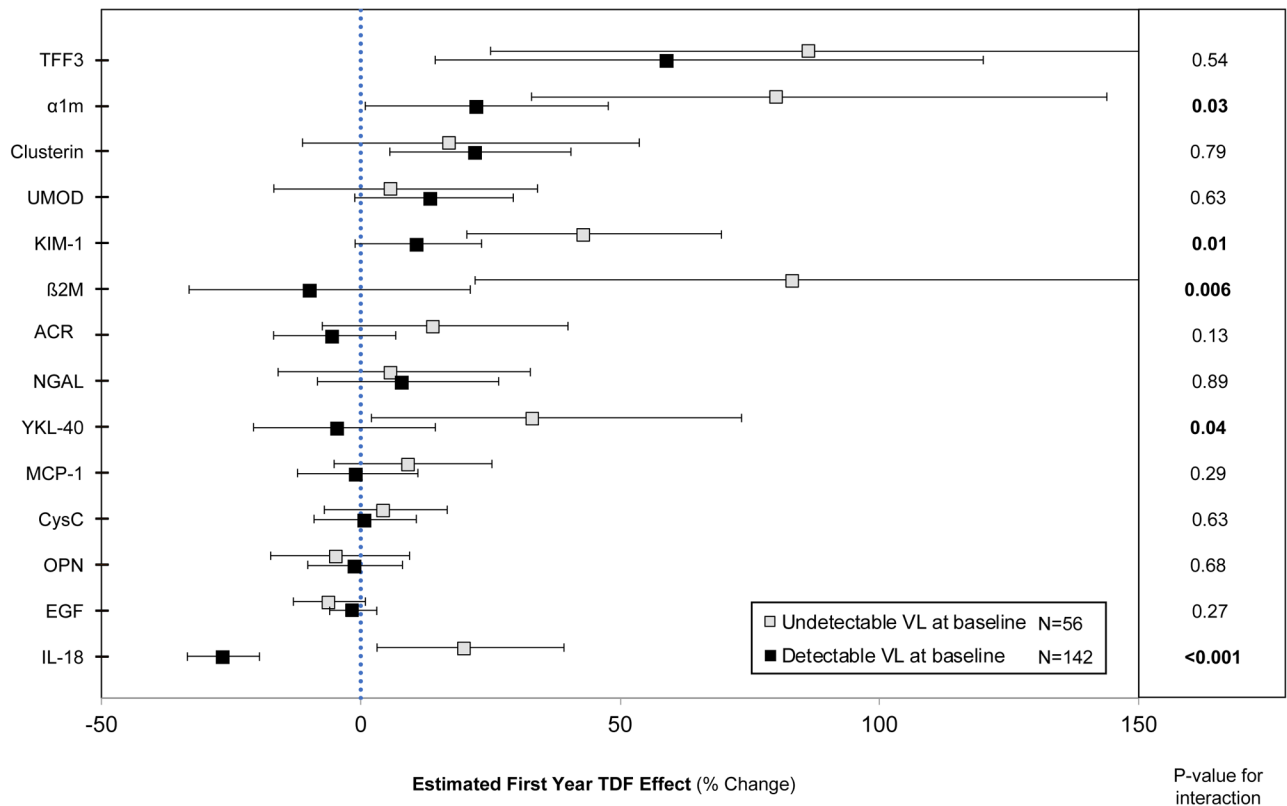


Figure 3. Association of TDF initiation with relative first-year changes in urine biomarker concentrations, stratified by baseline HIV RNA detectable status

The unfilled boxes denote participants with undetectable baseline viral load (VL) (HIV RNA < 80 copies/ mL), N=56. The filled boxes denote participants with detectable baseline viral load (HIV RNA ≥ 80 copies/ mL), N=142. Estimates denote the relative percent changes in concentrations of each biomarker during the first year of TDF use. P-values are calculated from tests of time by viral load interaction for each marker. Error bars denote the 95% confidence intervals (CIs). The x-axis is truncated at 150%. The 95% CI upper bounds for changes in TFF3 and β2M in participants with baseline undetectable viral load are truncated and extend to 177.6% and 174.7%, respectively. Estimates were calculated from separate linear mixed models, controlling for baseline viral load status (HIV RNA detectable vs. undetectable), time on TDF (using linear spline with cutpoint at year 1), interaction by baseline viral load, and urine creatinine. Numeric values of percent changes for each biomarker, 95% CI, and p-values for interaction are presented in Supplementary Table 3. Full names for each biomarker are as follows: trefoil factor 3 (TFF3), α1-microglobulin (α1m), clusterin, uromodulin (UMOD), kidney injury molecule-1 (KIM-1), β2-microglobulin (β2M), albumin-creatinine ratio (ACR), neutrophil gelatinase-associated lipocalin (NGAL), anti-chitinase-3-like protein 1 (YKL-40), monocyte chemoattractant protein-1 (MCP-1), cystatin C (CysC), osteoponin (OPN), epidermal growth factor (EGF), and interleukin-18 (IL-18). eGFR = estimated glomerular filtration rate.

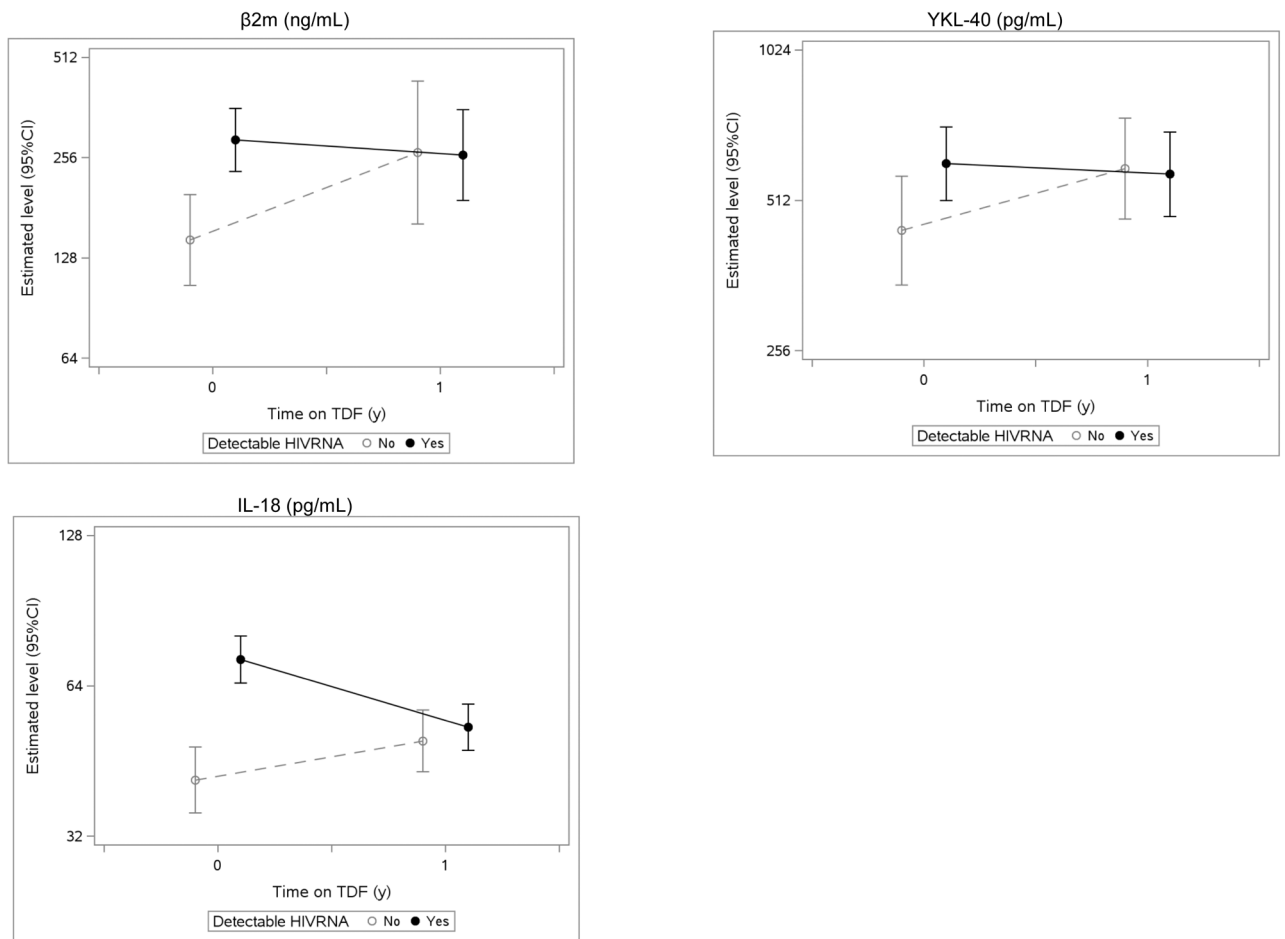


Figure 4. Estimated absolute biomarker concentrations before and after TDF initiation, stratified by baseline HIV RNA detectable status

Estimated absolute biomarker concentrations at baseline (year 0) and year 1 are depicted for the three biomarker concentrations that significantly differed at baseline in viral load-stratified analyses. At baseline, β 2M, YKL-40, and IL-18 levels were higher in participants with undetectable baseline viral loads (HIV RNA < 80 copies/ mL) relative to those with detectable viral loads (HIV RNA \geq 80 copies/ mL). Marginal mean concentrations were calculated from separate linear mixed models, controlling for baseline viral load status (HIV RNA detectable vs. undetectable), time on TDF (using linear spline with cutpoint at year 1), interaction by baseline viral load, and urine creatinine. Full names for biomarkers above are as follows: β 2-microglobulin (β 2M), anti-chitinase-3-like protein 1 (YKL-40), and interleukin-18 (IL-18).

Table 1.

Summary of demographic and clinical characteristics of HIV-infected participants, before and after TDF initiation

	Pre-TDF (n=198)	On TDF (~ year 1)* (n=198)	P-value
Age, years	48 (41, 54)	49 (42, 56)	—
Female		111 (56%)	—
Race			
African American		126 (64%)	
White		59 (30%)	—
Other		13 (7%)	
Smoking			
Current	73 (37%)	70 (35%)	0.83
Past	62 (31%)	66 (33%)	
Never	62 (31%)	62 (31%)	
Diabetes mellitus	32 (17%)	32 (17%)	1.00
Systolic BP, mmHg	126 (114, 137)	122 (113, 135)	0.18
Diastolic BP, mmHg	77 (71, 86)	77 (71, 85)	0.47
Antihypertensive use	70 (35%)	77 (39%)	0.14
History of CVD	13 (7%)	16 (8%)	0.08
Hepatitis C virus-infected	33 (17%)	34 (17%)	0.32
Serum creatinine, mg/dL	0.85 (0.72, 0.95)	0.91 (0.79, 1.0)	<0.001
eGFR, ml/min/1.73m ²	103 (88, 116)	95 (79, 111)	<0.001
LDL, mg/dL	101 (79, 121)	97 (71, 121)	0.29
HDL, mg/dL	46 (38, 57)	47 (38, 57)	0.35
TG, mg/dL	113 (79, 172)	114 (80, 167)	0.95
Serum albumin, g/dL	4.2 (3.8, 4.4)	4.3 (4.0, 4.5)	0.01
Current CD4, cells/mm ³	483 (338, 682)	587 (416, 743)	0.003
Nadir CD4, cells/mm ³	347 (223, 471)	340 (215, 458)	0.68
HIV RNA < 80 copies/mL	56 (29%)	162 (82%)	<0.001
ART use	79 (40%)	198 (100%)	<0.001
BMI, kg/m ²	27 (23, 32)	28 (24, 33)	0.51
Waist Circ., cm	94 (83, 104)	97 (85, 107)	0.19

Data are presented as Median (IQR) or numbers (percent). P-values testing within-subject changes from baseline from Wilcoxon signed-rank test or McNemar's test.

* On TDF represents study visit corresponding to biomarker measurement closest to time at which participant had reached 1 year of TDF exposure. BP = blood pressure; eGFR = estimated glomerular filtration rate (CKD-EPI); LDL = low-density lipoprotein; HDL = high-density lipoprotein; TG = triglycerides; CVD = cardiovascular disease; BMI = body mass index; circ = circumference; ART = antiretroviral therapy.