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The RNA Polymerase II Core Promoter in *Drosophila*

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Running Head: *Drosophila* Pol II Core Promoter

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ABSTRACT

Transcription by RNA polymerase II initiates at the core promoter, which is sometimes referred to as the 'gateway to transcription'. Here we describe the properties of the RNA polymerase II core promoter in *Drosophila*. The core promoter is at a strategic position in the expression of genes, as it is the site of convergence of the signals that lead to transcriptional activation. Importantly, core promoters are diverse in terms of their structure and function. They are composed of various combinations of sequence motifs such as the TATA box, initiator (Inr), and downstream core promoter element (DPE). Different types of core promoters are transcribed via distinct mechanisms. Moreover, some transcriptional enhancers exhibit specificity for particular types of core promoters. These findings indicate that the core promoter is a central component of the transcriptional apparatus that regulates gene expression.

A critically important yet often overlooked component of the gene regulatory apparatus is the RNA polymerase II (Pol II) core promoter (for reviews, see: Smale and Kadonaga 2003; Goodrich and Tjian 2010; Kadonaga 2012; Lenhard et al. 2012; Danino et al. 2015; Roy and Singer 2015; Vo ngoc et al. 2017a; Haberle and Stark 2018). The core promoter is generally considered to be the stretch of DNA, which typically ranges from about -40 nt to +40 nt relative to the transcription start site (TSS), that is sufficient to direct the accurate initiation of transcription. The core promoter is thus at a strategic position in the activation of a gene, as it is the site of convergence of the regulatory signals that lead to the initiation of transcription. For this reason, the core promoter is sometimes referred to as the 'gateway to transcription'.

In a simple transcription unit, the core promoter acts in conjunction with an enhancer and a proximal promoter region (Figure 1). The core promoter directs the initiation of transcription by Pol II, and this signal is modulated by the sequence-specific DNA-binding transcription factors (ssTFs) that bind to the enhancer and proximal promoter. A core promoter by itself exhibits little transcription activity.

The Pol II core promoter is a diverse regulatory unit. There are different types of core promoters that function by different mechanisms and have distinct biological properties. For example, some transcriptional enhancers exhibit specificity for particular types of core promoters. The basis for this phenomenon is the diversity of the DNA sequence motifs that comprise core promoters. Some of the known core promoter sequence elements in *Drosophila* are shown in Figure 2. These motifs include the TATA box, Initiator (Inr), and downstream core promoter element (DPE). It is common, for instance, to find core promoters that contain a TATA box, an Inr, and a DPE, or an Inr and a DPE (Chen et al. 2014). Moreover, somewhat strikingly, the core promoter arrangements at specific genes are conserved in different *Drosophila* species (Chen et al. 2014). There are no universal core promoter elements. In addition, some core promoters lack any of the known sequence motifs. It is thus likely that additional core promoter elements remain to be discovered. Because of the diversity of function of the core promoter, it is important to have a fundamental understanding of its composition and function for the

successful design, execution, and interpretation of experiments involving the regulation of gene expression.

In this review, we will describe our current understanding of the core promoter in *Drosophila*, an organism in which many important advances in our knowledge of the Pol II core promoter have been made. At the onset, it is useful to mention that basal transcription in *Drosophila* is somewhat simple and straightforward, at least in comparison to transcription in other animals. In *Drosophila*, a substantial fraction of the core promoters are driven by the TATA box and/or DPE motifs (Figure 2) (Kutach and Kadonaga 2000; Chen et al. 2014). There are extremely low levels of 5-methylcytosine in *Drosophila* (Capuano et al. 2014). Consistent with this property, *Drosophila* essentially lack CpG methylation and CpG islands. In addition, *Drosophila* promoters are generally amenable to biochemical analysis, as there are excellent in vitro transcription systems (see, for example, Soeller et al. 1988). Importantly, there are countless genetic tools in *Drosophila* for the study of transcription in cells as well as in the entire organism. In short, it is a truly a delight to do research on the core promoter and the basal transcription process in *Drosophila*.

Brief Overview of the Basal Transcription Initiation Factors

To study the core promoter, it is important to have a basic understanding of the factors that are involved in the initiation of Pol II transcription. Pol II by itself does not recognize the core promoter. Instead, auxiliary factors, which are known as the basal (or general) transcription factors, recognize core promoter sequence motifs, recruit Pol II to the promoter, and initiate the transcription process (for review, see Sainsbury et al. 2015). The best understood basal transcription factors are those that mediate transcription from TATA box-dependent core promoters. These basal factors, in the approximate order in which they assemble along with Pol II onto the promoter, are as follows: TFIID (Transcription Factor for RNA polymerase II D) and TFIIA, TFIIIB, Pol II and TFIIIF, TFIIIE, and TFIIH. The resulting assemblage of factors is

termed the transcription preinitiation complex, or PIC. In the presence of ribonucleoside 5'-triphosphates, transcription initiates rapidly (in seconds) from the PIC.

For the purpose of studying the core promoter, it is important to remember that TFIID is a key core promoter recognition factor. TFIID comprises the TATA box-binding protein (TBP) along with about 13-14 TBP-associated factors (TAFs), some of which have alternate forms (for reviews, see: Goodrich and Tjian 2010; Fuller 2016). TBP as well as some TAFs bind to specific core promoter motifs. In addition, TFIIA significantly affects the interaction of TFIID with the core promoter region (see, for example, Cianfrocco et al. 2013). The basal transcription factor TFIIB, which is a single polypeptide, also binds with some sequence specificity to a subset of promoters. Thus, particular note should be made of TFIID (including its TBP and TAF subunits), TFIIA, and TFIIB. There are also TBP-related factors (TRFs) that are discussed below.

The Transcription Start Site (TSS)

Identification of TSSs

The successful analysis of a promoter region is critically dependent upon the correct identification of its TSS or TSSs. The accuracy in the mapping of TSSs is continuing to improve as new techniques are being developed. At the present time, it appears that the best methods for the mapping of TSSs in cells involve the determination of the 5' ends of capped nascent transcripts (see, for example, Nechaev et al. 2010; Kruesi et al. 2013; Kwak et al. 2013; Lam et al. 2013). The characterization of nascent transcripts ensures that minimal processing of the transcripts has occurred in cells and in the processing of the samples. In the study of any particular promoter, it is important to examine the existing TSS data carefully and, if possible, perform additional experiments that would provide independent verification of the TSS or TSSs.

Focused versus dispersed transcription initiation

There are different patterns of transcription initiation, which are sometimes referred to as "promoter shape". One mode of transcription initiation, termed "focused" (also known as "narrow peak", "peaked", "sharp peak", and "single peak"), occurs when transcription initiates from either a single site or a narrow (~5 nt or less) cluster of sites. Transcription from focused promoters probably derives from a single positioned PIC. An alternate mode of transcription initiation, termed "dispersed" (also known as "broad" or "weak"), occurs when there are multiple TSSs that are spread out over a region that might be as large as 50 to 100 nt. The multiple TSSs from dispersed promoters likely derive from multiple distinct PICs at different locations. In addition, many promoters have the combined features of both focused and dispersed promoters – that is, they have a distinct major TSS along with dispersed minor TSSs. Focused, dispersed, and combined transcription patterns are observed in *Drosophila* (Rach et al. 2009, 2011; Ni et al. 2010; Hoskins et al. 2011).

Focused promoters often contain core promoter motifs such as the TATA box, Inr, motif ten element (MTE), and DPE, and are frequently associated with regulated genes, whereas dispersed promoters are associated with Ohler core promoter motifs 1, 6, 7 (Ohler et al. 2002) and DNA replication-related element (DRE) sequences, and are commonly found in ubiquitously expressed genes (Rach et al. 2009, 2011; Ni et al. 2010; Hoskins et al. 2011). In addition, focused promoters are more evolutionarily constrained than dispersed promoters (Schor et al. 2017). From a teleological standpoint, it is reasonable that regulated promoters would be most facilely turned on or off at a single site and that focused transcription would be less essential at constitutive promoters at which there is sustained gene expression under different conditions.

It is also interesting to note that the comparison of promoter shape and chromatin structure showed that dispersed promoters are generally associated with periodic arrays of nucleosomes, whereas focused promoters tend to lack an organized chromatin structure (Figure 3A) (Rach et al. 2011). Further analysis revealed, however, that focused promoters with a TCT motif are associated with positioned arrays of nucleosomes (Figure 3B) (Rach et al. 2011).

These findings thus indicate that periodic arrays of nucleosomes in the promoter region are not a special characteristic of dispersed transcription, but rather appear to be associated with housekeeping genes rather than regulated genes. It is possible, for instance, that the chromatin structure in the promoter regions of regulated genes is maintained in a disrupted state that allows for more facile gene activation.

Core Promoter Sequence Motifs

In this section, we will describe core promoter sequence motifs in *Drosophila*. In this regard, it is important to mention a key study in which Ohler et al. (2002) identified 1941 TSS clusters in *Drosophila* and used MEME to determine the 10 most over-represented sequence motifs in the region from -60 to +40 relative to the +1 TSS. This analysis yielded the DRE (DNA replication-related element; motif 2), TATA box (motif 3), Inr (motif 4), E-box (motif 5), DPE (motif 9), and MTE (motif ten element; Lim et al. 2004) as well as other sequences, such as those that are currently known as Ohler motifs 1, 6, and 7. Individual motifs will be described in greater detail below.

The TATA box

The TATA box is an ancient core promoter motif that is present in Archaea as well as in eukaryotes. It was discovered in the DNA sequence analysis of the *Drosophila* histone genes (Goldberg 1979). The TATA box is recognized by the TATA box-binding protein (TBP) subunit of the TFIID complex. In addition to conventional TATA boxes (for example, TATAAA at around -30 relative to the +1 TSS), it is likely that TATA-like sequences also function as TATA box elements due at least in part to the ability of TBP to accommodate variation in the TATA sequence (see, for example, Patikoglou et al. 1999). In this regard, it is notable that a TATA-like sequence termed DMv5 is commonly found in *Drosophila* (FitzGerald et al. 2006; Chen et al. 2014).

The Initiator (Inr)

The Inr is probably the most commonly occurring core promoter motif in bilateria (bilaterally symmetric animals). It was originally described by Corden et al. (1980), and its functions were incisively articulated by Smale and Baltimore (1989). The Inr interacts with the TAF1 and TAF2 subunits of the TFIID complex (Chakley and Verrijzer 1999; Louder et al. 2016). As a core promoter motif, the Inr does not have strong core promoter activity by itself (Smale and Baltimore 1989); rather, it functions with other core promoter elements such as the TATA box, MTE, and DPE.

The *Drosophila* Inr was first described by Hultmark et al. (1986) (consensus in non-heat shock genes: ATCA+1KTY), and Inr sequences that are bound by *Drosophila* TFIID (DYA+1KTG) were determined by Purnell et al. (1994). The current consensus for the *Drosophila* Inr is TCA+1GTY (Ohler et al. 2002; FitzGerald et al. 2006). This sequence is more distinct than the human Inr consensus of BBCCA+1BW (Vo ngoc et al. 2017b). In addition, an Inr-like sequence termed DMv4 is commonly found in *Drosophila* core promoters (FitzGerald et al. 2006; Chen et al. 2014).

The TCT motif (polypyrimidine initiator)

The TCT motif (polypyrimidine initiator) is a rare but biologically important core promoter motif in bilateria. The TCT motif is found in almost all ribosomal protein gene promoters as well as in the promoters of some other genes that encode factors that are involved in translation (Perry 2005; Parry et al. 2010). In *Drosophila*, the TCT motif is present in only about 120 core promoters, including the ribosomal protein gene promoters (Parry et al. 2010); thus, it is a rare motif. The TCT consensus sequence in *Drosophila* is YYC+1TTTTYY, which is similar to the TCT consensus in humans (YC+1TYTTY; Parry et al. 2010). Because of its scarcity, the TCT motif is frequently not identified in the sequence analysis of promoter regions. Nevertheless, it is likely to be important for the coordination of the expression of the ribosomal protein genes. Thus, in the case of the TCT motif, its rarity is a reflection of its specificity of function.

Although the TCT motif has a resemblance to the Inr, the two elements are functionally distinct. Somewhat strikingly, a single T to A nucleotide substitution (TC+1T to TCA+1) can convert a TCT motif into an Inr (Parry et al. 2010). Moreover, as discussed below, transcription that is driven by the TCT motif in *Drosophila* is mediated by TBP-related factor 2 (TRF2) instead of TBP (Wang et al. 2014). These findings indicate that there is a specialized transcription system involving TRF2 and the TCT motif for the expression of the ribosomal protein genes in *Drosophila*. This TCT-based system complements the RNA polymerase I and RNA polymerase III transcription systems for the synthesis of the components of the ribosome.

The downstream core promoter element (DPE)

The DPE was discovered in the analysis of TATA-less promoters in *Drosophila* (Burke and Kadonaga 1996). The consensus for the *Drosophila* DPE is RGWYV from +28 to +32 relative to the +1 TSS (Vo ngoc et al. 2017a). It is estimated that about 40% of *Drosophila* promoters contain a DPE motif (Kutach and Kadonaga 2000). A variant of the DPE is "motif 9" in Ohler et al. (2002). The motif 9 sequence has some of the combined features of the MTE and DPE, and is sometimes referred to as the "Ohler DPE".

The DPE and Inr elements function cooperatively for the binding of TFIID as well as for transcriptional activity (Burke and Kadonaga 1996). The mutation of either element results in a loss of TFIID binding and promoter activity. There is a precise spacing requirement between the DPE and Inr, as a single nucleotide increase or decrease in the Inr to DPE spacing can result in a several-fold decrease in transcriptional activity (Burke and Kadonaga 1997; Kutach and Kadonaga 2000). Photocrosslinking experiments with purified TFIID indicated that the DPE is in close proximity to the TAF6 and TAF9 subunits of TFIID (Burke and Kadonaga 1997); however, the cryo-EM structure of a PIC showed that the TAF1 and TAF2 subunits are in contact with the DPE (Louder et al. 2016). The basis for this difference is not known but could be due to alternate conformations of promoter-bound TFIID or possibly to the presence of multiple TFIID complexes at the promoter in the photocrosslinking experiments.

The DPE is also present in humans and is recognized by the human basal transcriptional machinery (Burke and Kadonaga 1997; Juven-Gershon et al. 2006), but it is rarely found in human promoters (discussed in Vo ngoc et al. 2017a). The apparent scarcity of the DPE in humans may be due to our current lack of knowledge of the human DPE consensus.

One interesting feature of the DPE is that it is frequently found in the promoters of *Drosophila* long interspersed nuclear elements (LINEs; non-LTR retrotransposons), which are transcribed by Pol II. LINEs lack an LTR and must therefore have internal promoter sequences that are downstream of the +1 TSS. Thus, the DPE is well suited for the transcription of these elements, and many *Drosophila* LINEs, such as *jockey*, *Doc*, *297*, *G*, and *I*, contain DPE-dependent promoters (Burke and Kadonaga 1996; Kutach and Kadonaga 2000).

The motif ten element (MTE)

The MTE is another core promoter element that was discovered in *Drosophila*. The MTE region was identified as an over-represented promoter sequence (motif 10) by Ohler et al. (2002), and the function of the MTE as a core promoter element was determined by Lim et al. (2004). Like the DPE, the MTE functions cooperatively with the Inr for TFIID binding and transcriptional activity (Lim et al. 2004). In addition, there is synergy between the MTE and the DPE (Lim et al. 2004; Theisen et al. 2010). Photocrosslinking data with purified TFIID showed that TAF6 and TAF9 are in close proximity to the MTE (Theisen et al. 2010). The photocrosslinking results with the MTE are similar to those seen with the DPE (Burke and Kadonaga 1997), but differ from the structural studies of TFIID in the PIC (Louder et al. 2016). The differences remain to be clarified.

An analysis of the downstream region of *Drosophila* core promoters suggested that the MTE through the DPE might be considered to be single functional unit with multiple contact points with TFIID (Theisen et al. 2010). In this study, two important DNA segments in the MTE region were found to be from +18 to +22 and from +27 to +29 relative to the +1 TSS. A comprehensive analysis might reveal a unified view of the downstream core promoter region. In

this regard, it is notable that G+24 contributes to DPE-dependent transcription (Kutach and Kadonaga 2000). In addition, as mentioned above, the "Ohler DPE" has some of combined features of the MTE and DPE.

The TFIIB and TFIIA recognition elements

The TFIIB recognition elements (BREs) are sites at which the basal transcription factor TFIIB contacts the promoter DNA immediately upstream (BRE^u) or downstream (BRE^d) of the TATA box (Lagrange et al. 1998; Deng and Roberts 2005; Tsai and Sigler 2000). This interaction involves the binding of TFIIB to TATA box-bound TBP, and hence, the functions of the BREs are dependent upon the presence of a TATA box.

Like TBP, TFIIB is an ancient transcription factor that is present in Archaea and eukaryotes. In humans, the BRE^u consensus is SSRCGCC (Lagrange et al. 1998), and the BRE^d consensus is RTDKKKK (Deng and Roberts 2005). The frequency of occurrence of the BRE motifs has been estimated to be about 10 to 35% (Gershenzon and Ioshikhes 2005; Deng and Roberts 2005). It should be noted, however, that there have not been more recent revisions of the human BRE consensus sequences. In addition, the BRE consensus sequences in *Drosophila* have not yet been determined.

The functions of the BREs remain to be clarified, as they have been observed to increase as well as to decrease transcriptional activity in different contexts (Lagrange et al. 1998; Evans et al. 2001; Deng and Roberts 2005). The BRE^u motif was also found to inhibit the ability of the *Drosophila* Caudal protein (an enhancer-binding factor) to activate transcription from TATA-dependent promoters (Juven-Gershon et al. 2008). Thus, it can be seen that further analysis of the BRE motifs is needed to determine their roles in the regulation of transcription.

In addition to the BREs, there is a TFIIA-recognition element, termed the IIARE, which was identified in the analysis of the binding of TFIIA to promoter DNA in TFIIA-TBP-DNA complexes (Wang et al. 2017). The IIARE is located immediately upstream of the TATA box, and its consensus sequence is GKGVSRTKKT. In TATA-containing promoters, the IIARE enhances

transcription, whereas in the absence of the TATA box, the IIARE represses transcription. The IIARE has not yet been studied in *Drosophila*.

Sequence-specific Transcription Factors and Transcription Initiation

It is important to consider the role of sequence-specific DNA-binding transcription factors (ssTFs) in the initiation of transcription, particularly at promoters that lack core promoter motifs such as the TATA box and DPE. In *Drosophila*, ssTF binding sites such as the DRE (DNA replication-related element; Ohler motif 2), E-box (Ohler motif 5), and Ohler motif 1 are frequently observed from about -60 to -1 relative to the +1 TSS (Ohler et al. 2002; FitzGerald et al. 2006). The DRE is bound by the DRE-binding factor (DREF; Hirose et al. 1993, 1996), the E-box is bound by helix-loop-helix (HLH) proteins such as Myc family transcription factors (Massari and Murre 2000), and Ohler motif 1 is bound by motif 1 binding protein (M1BP; Li and Gilmour 2013). DREF is present in *Drosophila* and humans (in which it is known as ZBED1 or hDREF) but not in yeast, HLH proteins that bind to the E-box are in yeast, *Drosophila*, and humans, and M1BP is in *Drosophila* but not in yeast or humans. Other ssTFs such as the GAGA factor (also known as GAF) and Zelda are also commonly found in the proximal promoter region (about -250 to -40 relative to the +1 TSS) of *Drosophila* genes.

It is likely that most of the ssTFs in the proximal promoter region activate transcription, as is well established (see, for example, Kadonaga 2004). In promoters that lack core promoter motifs or contain only an Inr or a TCT motif (which cannot direct efficient transcription by themselves; Smale and Baltimore 1989), it is possible that proximal promoter-bound ssTFs drive transcription initiation in conjunction with an Inr, a TCT motif, or an Inr-like element that might be as minimal as a CA+1 or TA+1 dinucleotide (Figure 4). It has been shown that an ssTF can activate transcription in lieu of a TATA box (Smale et al. 1990; Emami et al. 1995). It is thus reasonable to postulate that proximal promoter-bound ssTFs mediate the initiation of transcription at promoters lacking TATA box and DPE motifs. This hypothesis is relevant to the function of many promoters and should be investigated in greater depth.

TBP-related Factors (TRFs)

TBP (TATA box-binding protein) is an ancient protein that is present in Archaea and eukaryotes. At TATA-containing promoters, TBP has a critical role in the recruitment of the basal transcription machinery for assembly into the PIC. Most unicellular organisms contain a single TBP protein, but bilateria additionally have TBP-related factors (TRFs) with activities that are distinct from those of TBP.

The first identified TRF, which was initially termed "TRF" and is now known as TRF1, was discovered in *Drosophila* (Crowley et al. 1993). TRF1 appears to exist only in insects. A second TRF, termed TRF2 (also known as TBPL1, TLP, TRP, and TLF), is found in bilateria (reviewed in Goodrich and Tjian 2010; Akhtar and Veenstra 2011; Duttke et al. 2014). A third TRF, denoted TRF3 (also known as TBPL2 and TBP2), is present in vertebrates, and is the TRF that is most closely related to TBP (Persengiev et al. 2003; for reviews, see: Goodrich and Tjian 2010; Akhtar and Veenstra 2011; Vo ngoc et al. 2017a). In *Drosophila*, two additional TRFs, termed TRF4 and TRF5, have been identified (Kurshakova et al. 2018). Analysis of TRF4 revealed that it lacks a nuclear localization signal and appears to be cytoplasmic. Here, we will focus on the roles of *Drosophila* TBP, TRF1, and TRF2 in transcription (Figure 5).

TRF1

TRF1 is an insect-specific TRF that binds to the TATA box as well as to a TC-rich cluster and substitutes for TBP in the transcription of some Pol II promoters in vitro (Hansen et al. 1997; Holmes and Tjian 2000). In addition, TRF1 associates with the RNA polymerase III (Pol III) transcription factor termed BRF1 (also known as BRF), and has been found to be required for the transcription of several *Drosophila* Pol III promoters in vitro and to colocalize with BRF1 at Pol III genes in *Drosophila* cells and polytene chromosomes (Takada et al. 2000; Isogai et al. 2007a; Verma et al. 2013). Thus, TRF1 functions in transcription by Pol II and Pol III. It is interesting to note that the combined functions of TBP and TRF1 in *Drosophila* are similar to

those of TBP in organisms (such as the yeast *Saccharomyces cerevisiae*) that lack TRFs (Figure 5). Hence, the emergence of TRF1 generally resulted in the subdivision or sharing of the original functions of TBP between TBP and TRF1.

TRF2

TRF2 is present in bilateria (Duttke et al. 2014) and is the TRF that is the least closely related to TBP. Unlike TBP, TRF1, and TRF3, TRF2 does not exhibit any detectable DNA-binding activity (see, for example, Dantonel et al. 1999; Rabenstein et al. 1999; Wang et al. 2014; Zehavi et al. 2015). TRF2 does, however, interact with the TFIIA and TFIIB basal transcription factors (Rabenstein et al. 1999; Teichmann et al. 1999). Thus, TRF2 is similar to TBP except that it does not bind to the TATA box.

The emergence of TRF2 may have facilitated the evolution of TATA-less transcription systems in bilateria. Consistent with this idea, *Drosophila* TRF2 is mainly associated with TATA-less promoters (Isogai et al. 2007b). In addition, TRF2, but not TBP, has been found to be essential for transcription from *Drosophila* TATA-less promoters that contain either a TCT or a DPE core promoter motif (Hsu et al. 2008; Kedmi et al. 2014; Wang et al. 2014). Moreover, TRF2, but not TBP, is required for transcription of the TATA-less (and TCT- and DPE-deficient) histone H1 genes in *Drosophila* (Isogai et al. 2007b). It is further notable that the TCT and DPE core promoter motifs are found only in bilateria. Importantly, the TRF2-driven TATA-less transcription systems have added new gene regulatory functions (Figure 5), and the biological functions of TRF2-regulated genes suggests that TRF2 may have had an important role in the evolution of bilateria (Duttke et al. 2014).

TRF2 is essential in many bilateria. For instance, the loss of TRF2 is embryonic lethal in *Drosophila* (Kopytova et al. 2006), *Caenorhabditis elegans* (Dantonel et al. 2000; Kaltenbach et al. 2000), zebrafish (*Danio rerio*; Müller et al. 2001), and *Xenopus* (Veenstra et al. 2000). In contrast, TRF2-deficient mice are viable but have a defect in spermiogenesis (Martianov et al.

2001; Zhang et al. 2001; Zhou et al. 2013). It is possible that mice have another factor that can compensate for the absence of TRF2. This question remains to be resolved.

Because TRF2 does not bind to DNA, it is probably recruited to core promoters via interactions with other factors. In *Drosophila*, the ssTFs DREF and M1BP have been found to bind to TRF2. DREF recruits TRF2 to some promoters via its interaction with DREF binding sites (*i.e.*, DREs) (Hochheimer et al. 2002). M1BP recruits TRF2 to a majority of the TCT motif-containing ribosomal protein (RP) gene promoters via its binding to motif 1 sequences (Baumann and Gilmour 2017). M1BP does not, however, generally recruit TRF2 to promoters, as less than 3% of M1BP ChIP-exo peaks correlate with TRF2 ChIP-exo peaks at non-RP gene promoters. As mentioned above, E-box sequences are also commonly found in core promoter regions (Ohler et al. 2002; FitzGerald et al. 2006), and it is possible that some E-box-binding proteins recruit TRF2 to core promoters. It will be interesting to determine how promoter-associated TRF2 functions to mediate the initiation of transcription.

Enhancer-Core Promoter Specificity

Transcriptional enhancers are critical determinants of the spatial and temporal regulation of gene expression. In the *Drosophila* genome, it has been estimated that there are 50,000 to 100,000 enhancers (Kvon et al. 2014), and it is important to have the appropriate functional connections between each of these tens of thousands of enhancers and their cognate promoters. One means by which these connections are properly established is by enhancer-core promoter specificity (Figure 6).

The phenomenon of enhancer-core promoter specificity has been well studied in *Drosophila*. For instance, when the *Drosophila* AE-1 and IAB5 enhancers were each placed in a competitive situation in which they could activate transcription from either a TATA-dependent core promoter or a DPE-dependent core promoter, they exhibited a distinct preference for the TATA-dependent core promoter, even though they could, in the absence of competition, activate the DPE-dependent core promoter (Ohtsuki et al. 1998). In another set of experiments,

DPE-specific as well as TATA-specific enhancers were identified on the basis of their ability to activate transcription from a TATA-dependent versus a DPE-dependent core promoter (Butler and Kadonaga 2001). More recently, enhancer-core promoter specificity, such as with "developmental core promoters" (with TATA, Inr, MTE, DPE motifs) versus "housekeeping core promoters" (with TCT, DRE, E-box, and Ohler motifs 1, 6, 7), was seen on a genome-wide level (Arnold et al. 2013; Zabidi et al. 2015).

It will be important to understand the molecular basis of enhancer-core promoter specificity. In this regard, it was found that the *Drosophila* Caudal protein, a sequence-specific transcription factor (ssTF) and a master developmental regulator, is a DPE-specific transcription factor that functions preferentially with DPE-dependent core promoters relative to TATA-dependent core promoters (Juven-Gershon et al. 2008; Shir-Shapira et al. 2015). In addition, the transcriptional cofactors NC2 (negative cofactor 2, also known as Dr1-Drap1) and Mot1 were observed to activate DPE-dependent transcription and to inhibit TATA-dependent transcription (Willy et al. 2000; Hsu et al. 2008). Thus, ssTFs such as Caudal and cofactors such as NC2 and Mot1 may function to mediate enhancer-core promoter specificity. We are, however, only at the early stages of understanding the factors and mechanisms that are involved in this phenomenon.

Promoter-Proximal Pausing by RNA Polymerase II

At many genes, Pol II pauses at about 30 to 50 nt downstream of the TSS. This phenomenon was discovered in the analysis of the *Drosophila hsp70* promoter (Rougvie and Lis 1988). Promoter-proximal pausing involves factors such as NELF (negative elongation factor) and DSIF (DRB sensitivity-inducing factor), and is released by P-TEFb (positive transcription elongation factor b), which is a kinase that phosphorylates NELF, DSIF, and the C-terminal domain of the largest subunit of Pol II (for review, see Adelman and Lis 2012). Recent studies have shown that the paused Pol II is not a static entity, but is rather subject to rapid turnover

(Krebs et al. 2017; Erickson et al. 2018). Thus, the paused Pol II is frequently regenerated via new initiation events at the core promoter.

It is interesting to consider whether the core promoter might have a role in the promoter-proximal pausing and premature termination of Pol II. Promoter elements that have been found to be associated with Pol II pausing include GAGA factor binding sites, Ohler motif 1, the Inr, and a sequence termed the pause button (PB) (Hendrix et al. 2008; Lee et al. 2008; Li and Gilmour 2013; Duarte et al. 2016). Notably, the PB is frequently located in the vicinity of the DPE, and its consensus sequence (KCGRWCG) has a resemblance to that of the DPE (Hendrix et al. 2008). It is not known, however, whether the DPE and PB are functionally related.

Practical Applications of Synthetic Core Promoters

It is possible to increase gene expression by using synthetic core promoters with optimized sequences. For instance, super core promoters (SCPs) were designed with highly active versions of the TATA box, Inr, MTE, and DPE motifs (Juven-Gershon et al. 2006). In addition to studies involving gene expression, SCPs have been employed in biochemical and biophysical analyses of TFIID and the PIC (see, for example: Cianfrocco et al. 2013; Louder et al. 2016). There are also CpG-deficient SCPs that are resistant to repression by CpG methylation (Theisen et al. 2013). For studies in *Drosophila*, the *Drosophila* synthetic core promoter (DSCP; Pfeiffer et al. 2008) has been used in the analysis of enhancer activity (see, for example: Pfeiffer et al. 2008; Arnold et al. 2013; Zabidi et al. 2013). The DSCP contains both TATA box and DPE motifs, and is thus able to function with both TATA-specific enhancers and DPE-specific enhancers.

For any particular gene, the replacement of the core promoter region (approximately -40 to +40 relative to the +1 TSS, as in Figure 1) with an SCP sequence would likely result in an increase in gene expression. The core promoter region can be identified by mapping of the +1 TSS (in *Drosophila*, see: <http://labs.biology.ucsd.edu/Kadonaga/drosophila.tss.data/>) and by the possible presence of position-specific core promoter motifs such as the TATA box, Inr, MTE,

and DPE. For instance, if there is a distinct +1 TSS that correlates with a consensus Inr motif and perhaps a correctly positioned TATA-like sequence or DPE, then there is a good likelihood that it is a core promoter. In addition, when possible, core promoters can be confirmed and characterized by in vitro transcription analysis.

Perspectives

The RNA polymerase II core promoter is a diverse regulatory element. Here, we have described some of DNA sequence motifs and transcription factors that function in the core promoter region. It is our hope that this knowledge will be useful for studies of the regulation of gene activity in *Drosophila* and other bilateria.

In the future, we hope to gain new insights into the many remaining unsolved issues relating to the core promoter. From the perspective of the basal transcription process, it would be important to determine the factors and mechanisms that mediate TATA-less transcription, as the majority of promoters in *Drosophila* as well as in humans lack a TATA box or a TATA-like sequence. A related matter is the function of ssTFs that bind near the TSS of TATA-less promoters. It is possible, for instance, that ssTFs bound to their recognition sites act in a manner that is analogous to TBP bound to the TATA box (Figure 4). The activation of TATA-less transcription by ssTFs might also be related to the synthesis of species such as enhancer RNAs (eRNAs). More generally, as discussed here and elsewhere (Vo ngoc et al. 2017a), it would likely be useful to expand the concept of the core promoter to incorporate the functions of ssTFs and chromatin structure and signals.

From the perspective of gene regulation, it will be essential to elucidate the mechanism of enhancer-core promoter specificity. Such studies should reveal new insights not only into enhancer-promoter communication, but also into the factors that mediate transcriptional activation by enhancers. At the level of gene networks, it would be important to understand how core promoter motifs control groups of genes, such as in the regulation of ribosomal protein gene expression by the TCT motif, which is present in almost all ribosomal protein gene

promoters in *Drosophila* and humans (Parry et al. 2010). With respect to gene networks, it is also notable that nearly all of the *Drosophila* Hox genes have TATA-less, DPE-driven promoters (Juven-Gershon et al. 2008). Thus, core promoter motifs have roles in broader biological processes.

In conclusion, the core promoter is a central component of the transcriptional apparatus. It is a downstream target of signals that lead to gene activation, and it is upstream of events that occur subsequent to transcription initiation. It can thus be seen that our understanding of the mechanisms of gene regulation are critically dependent upon our knowledge of the multifarious functions of this 'gateway to transcription'.

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Figure Legends

Figure 1 The core promoter is a component of the transcriptional control region of a gene. The core promoter typically comprises the region from -40 to $+40$ relative to the $+1$ transcription start site (TSS), and contains the information that directs the initiation of transcription by RNA polymerase II. The core promoter functions with the proximal promoter and enhancer regions, both of which contain sites for the binding of sequence-specific DNA-binding transcription factors. The proximal promoter region spans from about -40 to -250 relative to the TSS. Many genes contain multiple enhancers that are located at variable distances both upstream and downstream of the TSS.

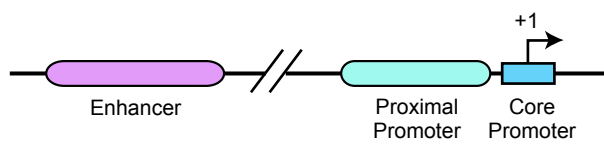
Figure 2 Schematic diagram of some RNA polymerase II core promoter motif arrangements that are observed in *Drosophila*. The specific motifs are discussed in the text. In this figure, "Inr" refers to the consensus Inr as well as to an Inr-like sequence termed DMv4, and "TATA" refers to the consensus TATA box as well as to a TATA-like sequence termed DMv5. DMv4 and DMv5 are commonly observed in *Drosophila* core promoters (FitzGerald et al. 2006; Chen et al. 2014). This drawing is roughly to scale.

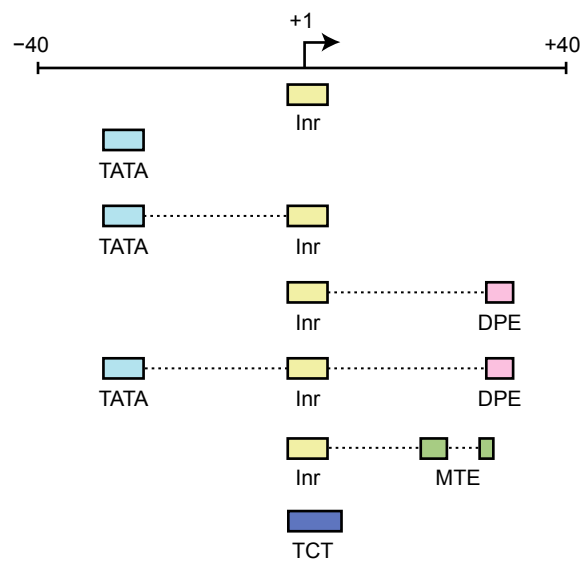
Figure 3 Analysis of chromatin structure at different types of core promoters suggests that promoters of regulated genes generally lack periodic arrays of nucleosomes. The diagrams are based on data from Rach et al. (2011). (A) Comparison of chromatin structure at focused versus dispersed promoters. (B) Comparison of focused promoters that contain TATA, Inr, MTE, or DPE motifs versus focused promoters that contain the TCT motif.

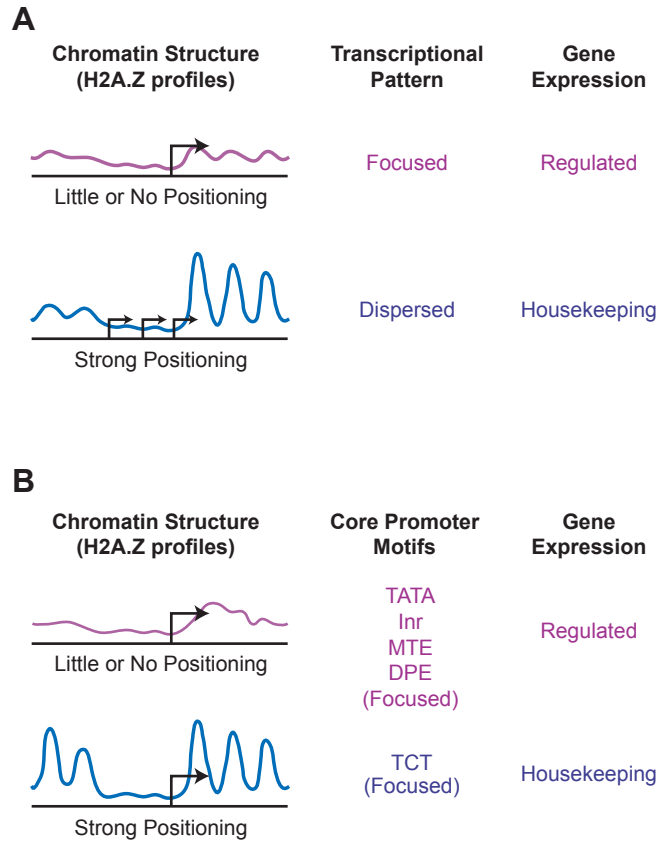
Figure 4 Postulated model for the role of sequence-specific DNA-binding transcription factors (ssTFs) in the initiation of transcription from promoters that lack core promoter motifs such as the TATA box and DPE. This model is discussed in the text.

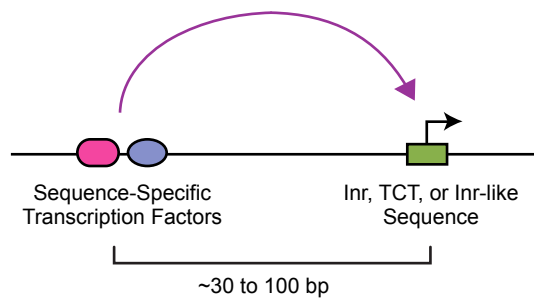
Figure 5 Summary of our current knowledge of transcriptional programs that are carried out by TBP, TRF1, and TRF2 in *Drosophila*. The combined functions of *Drosophila* TBP and TRF1 are approximately the same as the functions of TBP in organisms, such as *Saccharomyces cerevisiae*, that contain only TBP. In contrast, TRF2 mediates new functions that are not present in organisms that lack TRFs.

Figure 6 Schematic diagram of enhancer-core promoter specificity, in which enhancers establish functional interactions with their cognate core promoters.









TBP	TRF1	TRF2
<p>RNA Pol I RNA Pol II – TATA RNA Pol III – U6 snRNA Other Programs?</p>	<p>RNA Pol III – tRNA RNA pol II – TC box RNA pol II – TATA? Other Programs?</p>	<p>RNA Pol II – TCT RNA Pol II – DPE RNA Pol II – H1 Other Programs?</p>
<p><i>Functions that are mostly similar to those in organisms with only TBP (no TRFs)</i></p>		<p><i>New functions with TRF2</i></p>

