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Loss of Glucocorticoid Receptor Expression Mediates *in Vivo* Dexamethasone Resistance in T-cell Acute Lymphoblastic Leukemia

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Abstract

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Despite decades of clinical use, mechanisms of glucocorticoid resistance are poorly understood. We treated primary murine T lineage acute lymphoblastic leukemias (T-ALLs) with the glucocorticoid dexamethasone (DEX) alone and in combination with the pan-PI3 kinase inhibitor GDC-0941 and observed a robust response to DEX that was modestly enhanced by GDC-0941. Continuous *in vivo* treatment invariably resulted in outgrowth of drug-resistant clones, ~30% of which showed low glucocorticoid receptor (GR) protein expression. A similar proportion of relapsed human T-ALLs also exhibited markedly reduced GR protein levels. *De novo* or pre-existing mutations in the gene encoding GR (*Nr3c1*) occurred in relapsed clones derived from multiple independent parental leukemias. CRISPR/Cas9 gene editing confirmed that loss of GR expression confers DEX resistance. Exposing drug-sensitive T-ALLs to DEX *in vivo* altered transcript levels of multiple genes, and this response was attenuated in relapsed T-ALLs. These data implicate reduced GR protein expression as a frequent cause of glucocorticoid resistance in T-ALL.

Introduction

Glucocorticoids are a cornerstone of therapeutic protocols for T-ALL and other lymphoid cancers and a poor initial response to glucocorticoids remains one of the strongest predictors of relapse in ALL (1). Studies performed in the 1980s revealed low GR protein levels in some diagnostic and relapsed pediatric ALL samples, but uncovered no clear association with treatment response (2–5). In T-ALL cell lines exposed to glucocorticoids *in vitro*, resistance was associated with somatic *NR3C1* mutations resulting in impaired GR activity, reduced receptor expression, and failure to up-regulate GR in response to glucocorticoid exposure (6–12). However, *NR3C1* alterations were found only rarely in relapsed human B and T lineage ALLs (13–17) with the exception of relapsed B-cell precursor ALLs, which harbor frequent copy number alterations resulting in *NR3C1* deletions (18–21). Thus, the molecular mechanisms contributing to glucocorticoid resistance and relapse in ALL remain poorly understood.

Transplanting primary mouse T-ALLs generated via retroviral insertional mutagenesis (RIM) in wild-type (WT) and *Kras^{G12D}* mice and treating them *in vivo* is an unbiased strategy for elucidating resistance mechanisms (22). In a previous study, the pan-PI3 kinase (PI3K) inhibitor GDC-0941 exhibited anti-leukemia activity as a single agent and in combination with the MEK inhibition PD0325901 in this system (REF). These data and a report that identified GR as a substrate for phosphorylation and degradation by the PI3K effector AKT (23) raised the possibility that PI3K inhibition might be synergistic with glucocorticoid treatment in T-ALL. To address this question, we treated 10 independent T-ALLs with the glucocorticoid dexamethasone (DEX) alone and in combination with GDC-0941. These studies revealed frequent down-regulation of GR protein expression in drug-resistant leukemias isolated at relapse, including clones with either pre-existing or acquired *Nr3c1* mutations that followed different evolutionary trajectories. Directly inactivating *NR3C1* induced profound glucocorticoid resistance in drug-sensitive human T-ALL cells, and primary murine leukemias with reduced GR expression showed impaired regulation of glucocorticoid response genes upon *in vivo* DEX exposure. These data support the existence of diverse molecular mechanisms that converge on reduced GR expression as a

frequent cause of glucocorticoid resistance in T-ALL and potentially other lymphoid cancers.

Results

Dexamethasone and GDC-0941 show *in vivo* efficacy in T-ALL

Treating healthy mice with 15 mg/kg/day of DEX alone and in combination with 125 mg/kg/day of GDC-0941 (24) caused lymphopenia and splenic and thymic atrophy without other toxicities (data not shown). T-ALLs generated by performing RIM with MOL4070 (25) in wild-type mice (*Kras*^{WT}) and congenic animals harboring a *Kras* oncogene (*Kras*^{G12D}) (22, 26) contain frequent insertions and/or somatic mutations in known human T-ALL genes (26, 27). Ten independent T-ALLs (n=5 each of *Kras*^{WT} and *Kras*^{G12D}) were transplanted into 14 mice each and recipients were assigned to treatment with vehicle (n=4), DEX (n=5), or DEX/GDC-0941 (n=5) (Fig. 1a). Mice were treated for eight weeks or until disease progression, and bone marrow was obtained at euthanasia.

DEX-treated recipient mice showed a significant survival advantage compared to vehicle-treated controls (median 31 versus 20 days; p<0.0001, Log-rank test) that was modestly enhanced by the addition of GDC-0941 (median 31 versus 40 days; p=0.0805, Log-rank test) (Fig. 1b). Examining the 10 individual leukemias revealed variable sensitivity to DEX and DEX/GDC-0941 (Table S1) that was independent of *Kras*^{G12D} mutation status (Fig. S1a, b). To characterize genetic differences between parental and relapsed T-ALLs, we isolated leukemia cells from 71 mice that progressed after a significant (p<0.005) response to DEX or DEX/GDC-0941 treatment and performed Southern blotting to detect retroviral insertion sites. Twenty-eight relapsed leukemias (39%) exhibited one or more novel integrations compared to the corresponding parental T-ALL (Fig. 1c). Parental and relapsed T-ALLs invariably shared multiple retroviral integrations indicating evolution from a common founder clone, consistent with studies of relapsed human ALLs (13). Moreover, T-ALLs that relapsed after an initial response to DEX acquired intrinsic drug resistance, confirmed by transplantation into secondary recipients and *in vivo* re-treatment with DEX (Fig. 1d and Fig. S1c). Relapsed T-ALLs with and without clonal evolution as visualized by Southern blotting exhibited DEX resistance, suggesting the existence of both insertional and non-insertional mechanisms.

Reduced GR protein expression is frequent in relapsed mouse and human T-ALLs

We performed Western blotting to assess Notch1 and Ras-regulated signaling molecules. These studies did not uncover consistent differences in the levels of cleaved Notch1, PTEN, phosphorylated ERK, phosphorylated Akt or phosphorylated S6 between parental and relapsed leukemias (Fig. S2). However, we observed markedly reduced or absent GR protein expression in multiple independent relapsed T-ALLs compared to the corresponding DEX-sensitive parental leukemia, which was independent of *Kras* mutation status (Fig. 2a). We next performed immunohistochemistry (IHC) to assess GR protein levels in a cohort of 102 diagnostic and relapsed T-ALL patient bone marrow biopsies in order to determine if human leukemias also exhibited loss of receptor expression at relapse (Fig. 2b). Analysis of Western blotting data in our murine leukemias demonstrated high levels of GR in 80% of parental T-

ALLs (n=10) versus 32% of relapsed leukemias (n=65), with the remaining samples exhibiting medium (34%) or low (34%) GR protein expression (Fig. 2c; p-value=0.0109, Fisher's test). Strikingly, blinded IHC analysis of patient samples with comparable disease burden (Table S2) revealed robust GR protein expression in 90% of diagnostic T-ALL samples (n=72), whereas 40% of the relapsed leukemias (n=30) were either weakly positive (27%) or negative (13%) for GR staining (Fig. 2d; p-value=0.0013, Fisher's test). Together, these data demonstrate a significant increase in the proportion of T-ALLs with reduced or absent GR protein expression at relapse in large cohorts of mouse and human T-ALLs.

In order to assess the relationship between GR protein expression and *Nr3c1* transcript levels, we performed quantitative RT-PCR (qPCR) analysis of bone marrow harvested from paired parental and relapsed murine T-ALLs. These studies revealed variably decreased *Nr3c1* mRNA levels at relapse (Fig. 2e), suggesting GR protein levels may reflect reduced *Nr3c1* transcription or result from post-transcriptional or post-translational mechanisms. We next performed CRISPR-mediated gene editing of *Nr3c1* in CCRF-CEM, an established DEX-sensitive human T-ALL cell line (28). CRISPR editing using ribonucleoproteins (RNPs) containing one of two independent sgRNA molecules targeting the *Nr3c1* locus with high efficiency (58.9% for sgGR.1, 64.7% for sgGR.2) resulted in reduced GR protein expression (Fig. 2f) and also rendered CCRF-CEM cells resistant to DEX-induced cell death (Fig. 2g).

Relapsed T-ALLs harbor pre-existing or acquired *Nr3c1* mutations

To identify genomic alterations that might cause DEX resistance, we performed exome sequencing of 25 relapsed murine T-ALLs, including 22 with reduced or absent GR protein expression. This analysis revealed distinct point mutations or small insertions/deletions (indels) within *Nr3c1* coding exons in 18% of relapsed T-ALLs analyzed that emerged after treatment with DEX or DEX/GDC-0941 (n=22, including four samples from different recipients transplanted with *Kras*^{G12D} T-ALL 78A that harbored the same indel). *Nr3c1* mutations were also detected in mice transplanted with *Kras*^{WT} T-ALL 5C and *Kras*^{G12D} T-ALLs 2M and 20M (Fig. 3a and Table S3). Targeted Sanger sequencing of additional drug-treated relapsed clones isolated from recipients of parental T-ALL 78A revealed the same *Nr3c1* L282fs indel at a variant allele frequency (VAF) that correlated with GR protein levels (Fig. 3b and Fig. S3a), and ultra-deep sequencing at approximately 30,000x coverage identified the L282fs indel at 1.1% VAF in parental T-ALL 78A (Fig. S3b). By contrast, the *Nr3c1* mutations in T-ALLs 5C.C3, 2M.C4, and 20M.D1 were each detected at relapse in only one recipient mouse (n=8 relapsed clones analyzed for T-ALL 5C; n=8 for T-ALL 2M; n=9 for T-ALL 20M). Accordingly, targeted sequencing did not identify pre-existing *Nr3c1* mutations Q517X and I513fs in parental T-ALLs 5C and 2M, respectively at a minimum sensitivity of <0.1% VAF. As in human T-ALL (29), somatic *Notch1* mutations represent late genetic events in this model (26, 27). T-ALLs 2M and 20M contain somatic *Notch1* PEST domain mutations that were retained in resistant clones with or without *Nr3c1* mutations, strongly suggesting that the *Nr3c1* mutation was acquired later. Together, these data indicate T-ALL cells with *Nr3c1* mutations are selected by DEX treatment, either as rare pre-existing subclones or after *de novo* acquisition of this alteration, consistent with

data in human ALL demonstrating both selection of pre-existing relapse clones during treatment and development of new mutations that drive drug resistance (13, 16, 30–32).

Loss of *Nr3c1* enhances clonal fitness and confers dexamethasone resistance

To determine if *Nr3c1* mutations identified in relapsed T-ALLs are sufficient to induce DEX resistance, we performed functional studies in both relapsed T-ALL 20M.D1 and CCRF-CEM cells. Relapsed T-ALL 20M.D1 harbors a truncating *Nr3c1* K514X stop-gain mutation at approximately 70% VAF, and exhibited phenotypic resistance upon DEX re-treatment in secondary recipients (Fig. 1d). In addition, *in vivo* re-treatment of relapsed clone 20M.D1 with DEX, but not the control vehicle, strongly selected for outgrowth of *Nr3c1*-mutant T-ALL cells (Fig. 3c) that exhibited profoundly reduced GR protein expression (Fig. 3d). We also performed nucleofection in CCRF-CEM cells with RNPs containing sgRNAs targeting the homologous region of *Nr3c1* harboring the K514X mutation identified in T-ALL 20M.D1, then used limiting dilution to isolate multiple independent single-cell clones that exhibited bi-allelic editing. Whereas three CRISPR-edited clones exhibited loss of GR protein expression and resistance to DEX, three clones isolated after nucleofection with a control RNP containing a scrambled sgRNA showed no change in GR levels and remained drug-sensitive (Fig. 3e, f). Together, these studies indicate the *Nr3c1* K514X stop-gain mutation identified in T-ALL 20M.D1 causes loss of GR protein expression and enhances the fitness of primary leukemia cells exposed to DEX *in vivo*, and also show that targeted disruption of *Nr3c1* is sufficient to induce glucocorticoid resistance *in vitro*.

Relapsed T-ALLs with pre-existing or acquired *Nr3c1* mutations follow distinct evolutionary trajectories

The experimental strategy of transplanting primary T-ALLs into multiple independent recipient mice and treating them *in vivo* (Fig. 1a) provided a unique opportunity to explore the genetic relatedness of individual relapsed clones with either a pre-existing or acquired *Nr3c1* mutation. We analyzed retroviral integration and exome sequencing data from relapsed clones isolated from different recipients transplanted with parental T-ALLs 78A (n=4) or 20M (n=6). Consistent with rare pre-existing T-ALL 78A cells bearing an *Nr3c1* L282fs indel that conferred DEX resistance, Southern blotting revealed an identical pattern of MOL4070 integrations in the parental leukemia and all of the relapsed subclones (data not shown). The majority of mutations identified in T-ALL 78A, including the engineered *Kras*^{G12D} mutation and a *Notch1* indel, were also present at similar VAFs in all four relapsed clones analyzed by exome sequencing (78A.D1, 78A.D4, 78A.C2, 78A.C3) (Fig. 4a). The subclone harboring an *Nr3c1* L282fs indel was positively selected during treatment and emerged in all four recipients analyzed with relatively few additional relapse-specific mutations identified (Fig. 4b and Table S3). The limited genetic diversity of T-ALL 78A relapsed clones is fully consistent with the existence of a pre-existing *Nr3c1* mutant subclone identified in the parental leukemia.

In contrast to T-ALL 78A, the *Nr3c1* K514X mutation was only detected in relapsed T-ALL 20M.D1. Interestingly, leukemia cells isolated from four other recipient mice that relapsed during treatment with DEX or DEX/GDC-0941 exhibited clonal evolution by Southern blot. T-ALLs 20M.D3 and 20M.D4 harbored two novel insertions also present in 20M.C1 (Fig.

1c), indicating evolution from common pre-existing founder clone. Exome sequencing of relapsed T-ALLs with (20M.D4, 20M.C1) or without (20M.C3, 20M.C5, 20M.D1, 20M.D2) novel retroviral integrations (Fig. 1c) revealed multiple mutations present at similar VAFs as in the parental leukemia (Fig. 4c, gray circles). In addition, relapsed clones 20M.D4, 20M.C1, 20M.C3, and 20M.C5 shared mutations that were absent in the parental leukemia (Fig. 4c; blue circles and Table S3), indicating derivation from a common ancestral clone. Within this group, relapsed clones 20M.C1 and 20M.C3 exhibited complete or partial loss of numerous mutations identified in the parental T-ALL (Fig. 4c, yellow circles), and thus arose prior to their acquisition. Additionally, 20M.D4, 20M.C3, and 20M.C5 represent multi-lineage relapses harboring at least two subclones based on relapse-specific and partially shared VAFs lower than shared VAFs. The *Nr3c1* K514X mutation is among the alterations found in a single relapsed clone (20M.D1), with additional unique mutations identified in the other relapsed leukemias analyzed (Fig. 4d and Table S3). Together, these data indicate DEX resistance in T-ALL 20M is characterized by complex clonal dynamics with outgrowth from distinct but related pre-existing subclones that undergo further genetic evolution upon *in vivo* drug exposure. These data are most consistent with the existence of an ancestral drug-tolerant clone that persists during treatment, but must acquire additional somatic mutations to cause relapse.

Relapsed T-ALLs show attenuated transcriptional responses to *in vivo* dexamethasone exposure

We sought to define common transcriptional changes within genetically diverse drug-sensitive T-ALLs following glucocorticoid treatment, which has pleiotropic and cell context-dependent effects (33). Each parental leukemia was first transplanted into a cohort of recipient mice, which received a single dose of DEX (15 mg/kg) or control vehicle when they became moribund. The mice were euthanized four hours later and bone marrow was harvested for RNA sequencing (RNA-seq). We observed modulation of known GR transcriptional targets after short-term DEX exposure in drug-sensitive leukemias. Notably, *Myc* and several of its target genes were consistently down-regulated upon DEX exposure, while primary GR target genes (*Fkbp5*, *Tsc22d3*, *Nr3c1*) and pro-apoptotic genes (*Bcl2l11*, *Txnip*, *Bmf*) were consistently up-regulated (Fig. 5a). We used qPCR to validate a subset of the transcriptional changes identified by RNA-seq, and then assessed whether short-term *in vivo* DEX treatment also modulated expression of GR-responsive genes in relapsed T-ALLs with low GR protein levels (Fig. 5b and Fig. S4a, b). Analysis of T-ALL 20M revealed a blunted transcriptional response to DEX in the relapsed sample harboring the *Nr3c1* K514X mutation (20M.D1) as compared to the corresponding parental leukemia. Interestingly, four additional relapsed samples from T-ALL 20M that express low GR protein levels but lack *Nr3c1* mutations (20M.D2, 20M.D4, 20M.C1, 20M.C5) also showed a blunted transcriptional response to DEX (Fig. 5b). Similarly, DEX treatment of relapsed subclones from T-ALLs 2M and 73A induced a pattern of target gene expression distinct from that observed in the corresponding parental T-ALL (Fig. S4a, b). These data demonstrate a lack of the expected transcriptional response to *in vivo* DEX treatment in relapsed T-ALLs with reduced GR protein levels that either harbor or lack a *Nr3c1* mutation.

Discussion

Delineating how individual drugs contribute to resistance and relapse is challenging in the context of multi-agent chemotherapy protocols used in human ALL. To overcome this limitation, we transplanted and treated genetically heterogeneous primary T-ALLs with DEX as a single agent or in combination with a PI3K inhibitor and performed molecular and functional analyses of relapsed leukemias. Of 65 T-ALLs that relapsed after initially responding to glucocorticoid treatment *in vivo*, 68% showed reduced or absent GR protein expression. Importantly, GR protein levels were also reduced in 40% of relapsed human T-ALL samples, implicating loss of GR expression as a major cause of glucocorticoid resistance. This observation is consistent with clinical data correlating poor response to glucocorticoid treatment during induction with an elevated risk of relapse (1). Although previous studies examining ALL patient samples have generated conflicting data regarding the correlation between GR expression level and glucocorticoid resistance (34–36), functional studies in human T-ALL cell lines have demonstrated that modulation of GR expression level affects glucocorticoid response and resistance *in vitro* (37) and in xenograft models (38). Consistent with these data, our unbiased forward genetic screen identified loss of GR expression as a mechanism of glucocorticoid resistance *in vivo*, which we validated using CRISPR/Cas9 gene editing as a functional approach *in vitro*.

We identified *Nr3c1* mutations in 18% of relapsed murine T-ALLs analyzed, and showed those with reduced or absent GR protein expression with or without an *Nr3c1* mutation were resistant to DEX upon re-transplant and re-treatment. Whereas previous studies including relatively small numbers of relapsed ALLs uncovered *NR3C1* mutations or deletions either rarely (16) or not at all (13, 15), a very recent study in which comprehensive serial sequencing was performed on diagnostic and relapsed samples from 103 Chinese pediatric ALL patients identified *NR3C1* mutations as the most frequent relapse-specific alteration, occurring in 14% of cases (39). Copy number alterations leading to deletion of *NR3C1* have also been identified in 9% of relapsed B-cell precursor ALLs (21), with one study reporting *NR3C1* deletions in 28% of the ETV6/RUNX1-positive subgroup (18). Importantly, *NR3C1* deletions were consistently associated with inferior treatment response and adverse outcome in B-cell precursor ALL (18–21). Non-mutational mechanisms have also been implicated in causing reduced GR expression and glucocorticoid resistance in ALL. Paugh and colleagues identified caspase-induced cleavage of GR due to epigenetic up-regulation of the NALP inflammasome as a glucocorticoid resistance mechanism, and showed this reduction in GR protein levels induced substantial DEX resistance using cell line models (40). More recently, a preliminary report showed T-ALL cells with a mutation in the histone methyl transferase *NSD2* exhibit both reduced GR expression and impaired up-regulation after glucocorticoid treatment, resulting in a lack of GR binding to DNA and a profound reduction in the subsequent transcriptional response (41). Together with our studies of human and mouse leukemias, these emerging data indicate reduced GR expression is common in relapsed T-ALL and identify *Nr3c1/NR3C1* mutations as one mechanism of receptor down-regulation and glucocorticoid resistance.

Recent serial targeted ultra-deep sequencing in a large human ALL cohort of diagnosis/relapse pairs revealed most *NR3C1* mutations detected at relapse are likely acquired during

treatment and arise in patients that relapse later than those harboring a pre-existing drug resistance mutation (39). Similarly, three of the four *Nr3c1* mutations we identified in relapsed mouse T-ALLs were not detected in the corresponding parental leukemia, and present in a single recipient mouse that relapsed later than recipients of the T-ALL harboring a pre-existing *Nr3c1* mutation. Gain-of-function *PRPS1* and *NT5C2* mutations that drive thiopurine resistance in ALL were also not detected at diagnosis by high-depth sequencing (30, 42). A recent study showing *NT5C2* mutations reduce fitness of ALL cells in the absence of drug treatment provided an elegant mechanistic explanation for this observation (32). The ability of *NR3C1* mutations to alter the competitive fitness of leukemia-initiating cells may be context-dependent, particularly given the diverse physiologic roles of glucocorticoid signaling at different stages of lymphoid development (43).

Several sequencing studies comparing diagnostic and relapsed ALL patient samples have identified pre-existing drug-resistant cells that are selected during treatment (13, 16, 31). An alternative mechanism in which drug-tolerant “persister” cells survive treatment and then acquire additional alterations rendering them fully resistant was first described in solid tumor cell lines (44–48), and has been observed in γ -secretase inhibitor resistant T-ALL cell lines in which drug-tolerant cells with an altered chromatin state exhibit sensitivity to inhibition of the epigenetic reader protein BRD4 (49). However, the role of drug-tolerant persister cells in primary cancers and in the context of treatment with conventional chemotherapy agents remains unclear. Our analysis of evolutionary trajectories in murine T-ALLs uncovered remarkable genetic diversity in a relapsed leukemia harboring an acquired *Nr3c1* mutation as compared to the T-ALL with a pre-existing *Nr3c1* mutation. These data support a two-step model of glucocorticoid resistance whereby pre-existing drug-tolerant clones survive induction, but must acquire additional alterations that confer full resistance to DEX and drive relapse. Furthermore, our data indicating the majority of *Nr3c1* mutations were acquired during treatment suggest this resistance mechanism may be more common than selection of a pre-existing resistant clone in ALL. Similarly, deep genomic analysis of relapsed pediatric ALLs supports acquired mutations in genes involved in drug action within a drug-tolerant cell population as a frequent mechanism of resistance, and demonstrated a comparable evolutionary trajectory as we observed in relapsed mouse leukemias (50).

Our data provide a strong rationale for implementing treatment strategies to eradicate drug-tolerant clones that survive induction and ultimately give rise to resistance in patients with sub-optimal responses to front-line therapies. Although the addition of a PI3K inhibitor markedly enhanced survival in some T-ALLs as compared to treatment with DEX alone, this combination therapy was not sufficient to overcome DEX resistance across our cohort of primary murine leukemias. Newer compounds targeting specific PI3K isoforms (51) or inhibitors of downstream effectors (52) may provide more effective suppression of the pathway and enhance survival in combination with glucocorticoids. In addition, recent studies suggest that targeting interleukin 7 receptor signaling (53, 54) and developing methods for antagonizing aberrant JDP2 expression through MCL1 inhibition (55) are potential strategies for overcoming intrinsic glucocorticoid resistance in select patients. As relapsed/refractory T-ALLs with *NR3C1* mutations or loss of GR protein expression are unlikely to respond to glucocorticoids, this knowledge may identify patients who will not benefit from additional glucocorticoid therapy and could thereby avoid potential toxicities.

Similarly, developing clinical tests to identify *NR3C1* mutations and/or low GR protein expression in the leukemic blasts of patients with detectable minimal residual disease during treatment may identify those at high risk for relapse who could benefit from alternative or novel therapies. Intriguingly, restoring glucocorticoid sensitivity may prove feasible with combination therapy in some leukemias that exhibit reduced GR protein expression due to non-mutational mechanisms, potentially through the use of drugs targeting epigenetic modification (56, 57).

Materials and methods

Preclinical trials.

Retroviral insertional mutagenesis was performed using MOL4070LTR in wild-type (*Kras^{WT}*) or *Mx1-Cre, Lox-STOP-Lox (LSL)-Kras^{G12D} (Kras^{G12D})* mice on a C57BL/6 × 129Sv/Jae (F1) strain background as described previously (26). Cryopreserved primary T-ALL cells were transplanted into sublethally irradiated (450 rads) recipient mice for *in vivo* expansion, then (2×10^6) bone marrow cells from these mice were intravenously or retro-orbitally injected into sublethally irradiated 8–12 week old male F1 recipient mice for the preclinical trial. Starting four days post-transplant, GDC-0941 (125 mg/kg/day) or control vehicle (0.5% hydroxypropyl methylcellulose, 0.2% Tween 80) was administered by oral gavage, and DEX (15 mg/kg/day) or control vehicle (phosphate buffered saline) was administered by intraperitoneal injection without blinding at any stage of the study. For the trial cohort, 14 mice were randomly assigned to receive vehicle (n=4), DEX (n=5), or DEX/GDC-0941 (n=5). To confirm drug resistance in relapsed T-ALLs, seven secondary recipients were randomly assigned to receive vehicle (n=3) or DEX (n=4). Mice were euthanized when they appeared moribund and survival was calculated from day of transplant. The time to death for vehicle-treated mice transplanted with individual T-ALLs was highly consistent, allowing determination of significant differences in the survival of drug-treated mice. GraphPad Prism Software was used generate Kaplan-Meier survival curves. Statistical significance was calculated by comparing individual trial arms using a two-tailed Log-rank test. Data analysis from previous studies has verified the statistical power of these cohort sizes to reliably detect significant differences (22, 26, 58–60). All animals assigned to treatment groups were included in the survival analysis. Very rarely (n=3 mice in the original preclinical trial cohort), animals were censored due to a mortality that was not caused by leukemia but rather an unrelated event including complications during drug administration or lack of leukemia engraftment leading to bone marrow failure. Dexamethasone sodium phosphate for injection (NDC 63323-516-10) was purchased through the University of California, San Francisco (UCSF) pharmacy. GDC-0941 was obtained through a Materials Transfer Agreement with Genentech, Inc. All animal studies were approved by the UCSF Institutional Animal Care and Use Committee (AN179357).

Southern blot analysis and MOL4070LTR integration cloning.

Restriction enzyme digestion with HindIII or BamHI of genomic DNA from primary mouse T-ALLs was followed by gel electrophoresis and hybridization with a ³²P-radiolabeled MOLO4070LTR-specific probe as described previously (58). Retroviral junction fragments were cloned and sequenced using a linker-based approach (61).

Western blot analysis and immunohistochemistry.

Bone marrow from moribund T-ALL recipients was subjected to red blood cell lysis, then resuspended in RIPA buffer to generate lysates. Protein concentration was determined using a Pierce BCA Protein Assay Kit (Thermo Fisher Scientific), then equivalent amounts of protein were mixed with Laemmli Sample Buffer (Bio-Rad Laboratories) followed by separation on a 8–20% Criterion TGX Precast Gel (Bio-Rad Laboratories). Immunoblots were assayed using primary antibodies listed in Table S4. IRDye secondary antibodies were used for detection and Western blots were imaged using an Odyssey Imaging System (LI-COR Biosciences). GR expression level for each leukemia was assigned an unblinded score of high (comparable to the corresponding parental T-ALL), medium (reduced as compared to the corresponding parental T-ALL), or low (nearly or completely absent). The level of GR protein expression for each T-ALL was assessed in bone marrow from at least two independent recipient mice with comparable results. Immunohistochemistry was performed on de-identified human bone marrow samples from children and adults of both sexes diagnosed with T-cell acute lymphoblastic leukemia using GR antibody at 1:400. Diagnostic and relapsed samples were unpaired, with the exception of two patients. T-ALL samples subjected to IHC analysis were scored blindly by two hematopathologists. A detailed protocol for immunohistochemistry and image acquisition in human T-ALLs is provided in the Supplemental Materials and Methods. GraphPad Prism Software was used for graphical representation of the data. The two-tailed Fisher's exact test was used to determine the significance of association with respect to GR expression levels in parental/diagnostic versus relapsed mouse and human T-ALLs. De-identified human tissue samples were obtained with Institutional Review Board approval UCSF (10–01080), Brigham and Women's Hospital (2010P002736) and Boston Children's Hospital (P00025328).

CRISPR/Cas9 modification of CCRF-CEM cells.

CCRF-CEM cells were purchased through the UCSF Cell Culture Facility (ATCC, CCL-119) and cultured in RPMI media supplemented with 10% fetal bovine serum and glutamine. All cell lines described in this study were authenticated through the UC Berkeley DNA Sequencing Facility and tested negative for mycoplasma. Cas9 protein was purchased from the University of California, Berkeley. tracrRNA, sgRNAs, and the HDR template were purchased from Dharmacon, and sequences are listed in Table S5. Transfection was performed using Amaxa Cell Line Nucleofector Kit C (Lonza, VACA-1004) in an Amaxa Nucleofector II Device. Transfection efficiencies were determined using TIDE analysis software (62). CRISPR-edited clones were generated using limiting dilution in conjunction with TIDE analysis to confirm editing of all alleles and Sanger sequencing to confirm HDR template incorporation.

***In vitro* dexamethasone dose response assays.**

Unperturbed or CRISPR-edited CCRF-CEM cells were seeded into 96-well round-bottom tissue culture plates in triplicate (50,000 cells/well) and exposed to increasing concentrations of DEX (Sigma; D4902) or vehicle (DMSO). Cells were resuspended in Hoechst viability dye after 72 hours and analyzed on a BD FACSVerse 8 color Flow Cytometer. FlowJo software was used to determine the percentage of live cells at each DEX concentration.

GraphPad Prism Software was used for graphical representation of the data. Each dose response assay was repeated three times with comparable results.

Exome and targeted sequencing.

Genomic DNA was isolated from the bone marrow of moribund T-ALL recipient mice after prolonged *in vivo* drug exposure using a Puregene DNA Isolation Kit (Qiagen), quantified with PicoGreen and quality controlled by Agilent BioAnalyzer, and 264–500 ng of DNA were used to prepare libraries using the KAPA Hyper Prep Kit (Kapa Biosystems KK8504) with 8 PCR cycles. After sample barcoding, 500–2100 ng of library were captured by hybridization using SureSelectXT Mouse All Exon (Agilent #5190–4641) according to the manufacturer's protocol. In total, 8–10 cycles of PCR amplification of the post-capture libraries was undertaken after which samples were sequenced on either HiSeq 4000 or HiSeq 2500 instrumentation in 100 bp paired end format using the HiSeq 3000/4000 SBS Kit or HiSeq Rapid SBS Kit v2 (Illumina). Normal and tumor samples were covered to an average of 86- and 173-fold, respectively. Detailed descriptions of the targeted sequencing approach, exome sequencing analysis, and Sanger sequencing validation methods are described in the Supplementary Materials and Methods. Primers used to validate *Nr3c1* mutations identified by exome sequencing are listed in Table S6. The exome sequencing data presented in Figures 3 and 4 of this manuscript was deposited into the DNA Data Bank for Japan (<https://www.ddbj.nig.ac.jp/dra/index-e.html>) under accession code .

Short-term dexamethasone treatment and RNA-seq.

Moribund T-ALL recipient mice received a single dose of vehicle (phosphate buffered saline) or DEX (15 mg/kg), and were euthanized four hours later. Bone marrow cells from these animals were subjected to red blood cell lysis, then frozen in Trizol. Total RNA was extracted and assessed for quality and quantity using the Eukaryote Total RNA Nano Bioanalyzer chip (Agilent Technologies, Inc) and Qubit fluorometer (Thermo Fisher), respectively. RNA-Seq libraries were prepared on the Sciclone liquid handling workstation (PerkinElmer) from 1 µg of total RNA per sample using the TruSeq Stranded Total RNA kit (Illumina, Inc.) following the manufacturer's recommendations. RNA-Seq libraries were clustered on the cBot and sequenced on HiSeq 2000 and HiSeq 2500 sequencers at 2×100 bp using HiSeq v4 sequencing reagents (Illumina, Inc.). RNA-seq was performed on bone marrow from one vehicle- or DEX-treated mouse per T-ALL indicated. Detailed analysis methods are described in the Supplementary Materials and Methods. False discovery rate and p-values were calculated from RNA-seq data to assess the significance of gene expression changes. RNA-sequencing data presented in Figure 5 of this manuscript was deposited into the National Center for Biotechnology Information Gene Expression Omnibus (NCBI Geo; <https://www.ncbi.nlm.nih.gov/geo/>) under accession code GSE141967.

Quantitative RT-PCR validation of transcriptome analysis.

Bone marrow cells collected after short-term DEX treatment as described above were subjected to red blood cell lysis, then total RNA was isolated using the RNeasy Mini Kit (Qiagen) and used to generate cDNA using Superscript III (Thermo Fisher Scientific). TaqMan Gene Expression Assays (Applied Biosystems) are listed in Table S7, and qPCR

was performed on a QuantStudio 5 Real-Time PCR Instrument (Applied Biosystems). qPCR was performed on bone marrow from one vehicle- or DEX-treated mouse per T-ALL as indicated using technical triplicates. GraphPad Prism Software was used for graphical representation of the data.

Statistical analysis.

The two-tailed Log-rank test was used to perform pairwise comparisons of Kaplan-Meier survival analyses in order to determine significance. The two-tailed Fisher's exact test was used to determine the significance of association with respect to GR expression levels in parental/diagnostic versus relapsed mouse and human T-ALLs. False discovery rate and p-values were calculated to assess the significance of gene expression changes as determined by RNA-sequencing analysis of murine T-ALLs. Standard deviation was calculated using technical triplicates for *in vitro* assessment of DEX sensitivity in human T-ALL cell lines. Standard error of the mean was calculated using technical triplicates for qPCR in primary murine T-ALLs.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

1. Gao J & Liu WJ (2018) Prognostic value of the response to prednisone for children with acute lymphoblastic leukemia: a meta-analysis. *European review for medical and pharmacological sciences* 22(22):7858–7866. [PubMed: 30536331]
2. Pui CH, Dahl GV, Rivera G, Murphy SB, & Costlow ME (1984) The relationship of blast cell glucocorticoid receptor levels to response to single-agent steroid trial and remission response in children with acute lymphoblastic leukemia. *Leukemia research* 8(4):579–585. [PubMed: 6590931]
3. Pui CH, Ochs J, Kalwinsky DK, & Costlow ME (1984) Impact of treatment efficacy on the prognostic value of glucocorticoid receptor levels in childhood acute lymphoblastic leukemia. *Leukemia research* 8(3):345–350. [PubMed: 6379308]
4. Pui CH & Costlow ME (1986) Sequential studies of lymphoblast glucocorticoid receptor levels at diagnosis and relapse in childhood leukemia: an update. *Leukemia research* 10(2):227–229. [PubMed: 3456475]

5. Lauten M, Cario G, Asgedom G, Welte K, & Schrappe M (2003) Protein expression of the glucocorticoid receptor in childhood acute lymphoblastic leukemia. *Haematologica* 88(11):1253–1258. [PubMed: 14607754]
6. Ashraf J & Thompson EB (1993) Identification of the activation-labile gene: a single point mutation in the human glucocorticoid receptor presents as two distinct receptor phenotypes. *Molecular endocrinology* 7(5):631–642. [PubMed: 8316249]
7. Powers JH, Hillmann AG, Tang DC, & Harmon JM (1993) Cloning and expression of mutant glucocorticoid receptors from glucocorticoid-sensitive and -resistant human leukemic cells. *Cancer research* 53(17):4059–4065. [PubMed: 8358735]
8. Strasser-Wozak EM, et al. (1995) Splice site mutation in the glucocorticoid receptor gene causes resistance to glucocorticoid-induced apoptosis in a human acute leukemic cell line. *Cancer research* 55(2):348–353. [PubMed: 7812967]
9. Geley S, et al. (1996) Resistance to glucocorticoid-induced apoptosis in human T-cell acute lymphoblastic leukemia CEM-C1 cells is due to insufficient glucocorticoid receptor expression. *Cancer research* 56(21):5033–5038. [PubMed: 8895760]
10. Hala M, Hartmann BL, Bock G, Geley S, & Kofler R (1996) Glucocorticoid-receptor-gene defects and resistance to glucocorticoid-induced apoptosis in human leukemic cell lines. *International journal of cancer* 68(5):663–668. [PubMed: 8938150]
11. Riml S, Schmidt S, Ausserlechner MJ, Geley S, & Kofler R (2004) Glucocorticoid receptor heterozygosity combined with lack of receptor auto-induction causes glucocorticoid resistance in Jurkat acute lymphoblastic leukemia cells. *Cell death and differentiation* 11 Suppl 1:S65–72. [PubMed: 15017388]
12. Mullighan CG, et al. (2008) Genomic analysis of the clonal origins of relapsed acute lymphoblastic leukemia. *Science* 322(5906):1377–1380. [PubMed: 19039135]
13. Hogan LE, et al. (2011) Integrated genomic analysis of relapsed childhood acute lymphoblastic leukemia reveals therapeutic strategies. *Blood* 118(19):5218–5226. [PubMed: 21921043]
14. Kunz JB, et al. (2015) Pediatric T-cell lymphoblastic leukemia evolves into relapse by clonal selection, acquisition of mutations and promoter hypomethylation. *Haematologica* 100(11):1442–1450. [PubMed: 26294725]
15. Oshima K, et al. (2016) Mutational landscape, clonal evolution patterns, and role of RAS mutations in relapsed acute lymphoblastic leukemia. *Proceedings of the National Academy of Sciences of the United States of America* 113(40):11306–11311. [PubMed: 27655895]
16. Spinella JF, et al. (2018) Mutational dynamics of early and late relapsed childhood ALL: rapid clonal expansion and long-term dormancy. *Blood advances* 2(3):177–188. [PubMed: 29365312]
17. Kuster L, et al. (2011) ETV6/RUNX1-positive relapses evolve from an ancestral clone and frequently acquire deletions of genes implicated in glucocorticoid signaling. *Blood* 117(9):2658–2667. [PubMed: 21224468]
18. Grausenburger R, et al. (2016) Genetic alterations in glucocorticoid signaling pathway components are associated with adverse prognosis in children with relapsed ETV6/RUNX1-positive acute lymphoblastic leukemia. *Leukemia & lymphoma* 57(5):1163–1173. [PubMed: 26327566]
19. Piovan E, et al. (2013) Direct reversal of glucocorticoid resistance by AKT inhibition in acute lymphoblastic leukemia. *Cancer cell* 24(6):766–776. [PubMed: 24291004]
20. Paugh SW, et al. (2015) NALP3 inflammasome upregulation and CASP1 cleavage of the glucocorticoid receptor cause glucocorticoid resistance in leukemia cells. *Nature genetics* 47(6):607–614. [PubMed: 25938942]
21. Li J, et al. (2018) A gain of function mutation in the NSD2 histone methyltransferase drives glucocorticoid resistance of acute lymphoblastic leukemia. *Blood* 132(Suppl 1):653.
22. Wolff L, Koller R, Hu X, & Anver MR (2003) A Moloney murine leukemia virus-based retrovirus with 4070A long terminal repeat sequences induces a high incidence of myeloid as well as lymphoid neoplasms. *J Virol* 77(8):4965–4971. [PubMed: 12663802]
23. Lauchle JO, et al. (2009) Response and resistance to MEK inhibition in leukaemias initiated by hyperactive Ras. *Nature* 461(7262):411–414. [PubMed: 19727076]

24. Dail M, et al. (2010) Mutant Ikzf1, KrasG12D, and Notch1 cooperate in T lineage leukemogenesis and modulate responses to targeted agents. *Proceedings of the National Academy of Sciences of the United States of America* 107(11):5106–5111. [PubMed: 20194733]
25. Dail M, et al. (2014) Loss of oncogenic Notch1 with resistance to a PI3K inhibitor in T-cell leukaemia. *Nature* 513(7519):512–516. [PubMed: 25043004]
26. Burgess MR, et al. (2017) KRAS Allelic Imbalance Enhances Fitness and Modulates MAP Kinase Dependence in Cancer. *Cell* 168(5):817–829 e815. [PubMed: 28215705]
27. Folkes AJ, et al. (2008) The identification of 2-(1H-indazol-4-yl)-6-(4-methanesulfonyl-piperazin-1-ylmethyl)-4-morpholin-4-yl-thieno[3,2-d]pyrimidine (GDC-0941) as a potent, selective, orally bioavailable inhibitor of class I PI3 kinase for the treatment of cancer. *J Med Chem* 51(18):5522–5532. [PubMed: 18754654]
28. Norman MR & Thompson EB (1977) Characterization of a glucocorticoid-sensitive human lymphoid cell line. *Cancer research* 37(10):3785–3791. [PubMed: 269011]
29. Mansour MR, et al. (2007) Notch-1 mutations are secondary events in some patients with T-cell acute lymphoblastic leukemia. *Clinical cancer research : an official journal of the American Association for Cancer Research* 13(23):6964–6969. [PubMed: 18056171]
30. Tzoneva G, et al. (2013) Activating mutations in the NT5C2 nucleotidase gene drive chemotherapy resistance in relapsed ALL. *Nature medicine* 19(3):368–371.
31. Ma X, et al. (2015) Rise and fall of subclones from diagnosis to relapse in pediatric B-acute lymphoblastic leukaemia. *Nature communications* 6:6604.
32. Tzoneva G, et al. (2018) Clonal evolution mechanisms in NT5C2 mutant-relapsed acute lymphoblastic leukaemia. *Nature* 553(7689):511–514. [PubMed: 29342136]
33. Reddy TE, et al. (2009) Genomic determination of the glucocorticoid response reveals unexpected mechanisms of gene regulation. *Genome research* 19(12):2163–2171. [PubMed: 19801529]
34. Li B, et al. (2018) Mutational Landscape and Temporal Evolution during Treatment of Relapsed Acute Lymphoblastic Leukemia. *Blood* 132(Suppl 1):1. [PubMed: 29976776]
35. Li B, et al. (2015) Negative feedback-defective PRPS1 mutants drive thiopurine resistance in relapsed childhood ALL. *Nature medicine* 21(6):563–571.
36. Jamieson CA & Yamamoto KR (2000) Crosstalk pathway for inhibition of glucocorticoid-induced apoptosis by T cell receptor signaling. *Proceedings of the National Academy of Sciences of the United States of America* 97(13):7319–7324. [PubMed: 10860997]
37. Sharma SV, et al. (2010) A chromatin-mediated reversible drug-tolerant state in cancer cell subpopulations. *Cell* 141(1):69–80. [PubMed: 20371346]
38. Ramirez M, et al. (2016) Diverse drug-resistance mechanisms can emerge from drug-tolerant cancer persister cells. *Nature communications* 7:10690.
39. Raha D, et al. (2014) The cancer stem cell marker aldehyde dehydrogenase is required to maintain a drug-tolerant tumor cell subpopulation. *Cancer research* 74(13):3579–3590. [PubMed: 24812274]
40. Hata AN, et al. (2016) Tumor cells can follow distinct evolutionary paths to become resistant to epidermal growth factor receptor inhibition. *Nature medicine* 22(3):262–269.
41. Hangauer MJ, et al. (2017) Drug-tolerant persister cancer cells are vulnerable to GPX4 inhibition. *Nature* 551(7679):247–250. [PubMed: 29088702]
42. Knoechel B, et al. (2014) An epigenetic mechanism of resistance to targeted therapy in T cell acute lymphoblastic leukemia. *Nature genetics* 46(4):364–370. [PubMed: 24584072]
43. Delgado-Martin C, et al. (2017) JAK/STAT pathway inhibition overcomes IL7-induced glucocorticoid resistance in a subset of human T-cell acute lymphoblastic leukemias. *Leukemia* 31(12):2568–2576. [PubMed: 28484265]
44. Mansour MR, et al. (2018) JDP2: An oncogenic bZIP transcription factor in T cell acute lymphoblastic leukemia. *The Journal of experimental medicine* 215(7):1929–1945. [PubMed: 29941549]
45. Mansour MR, et al. (2014) Oncogene regulation. An oncogenic super-enhancer formed through somatic mutation of a noncoding intergenic element. *Science* 346(6215):1373–1377. [PubMed: 25394790]

46. Jing D, et al. (2018) Lymphocyte-Specific Chromatin Accessibility Pre-determines Glucocorticoid Resistance in Acute Lymphoblastic Leukemia. *Cancer cell* 34(6):906–921 e908. [PubMed: 30537513]
47. Uren AG, et al. (2009) A high-throughput splinkerette-PCR method for the isolation and sequencing of retroviral insertion sites. *Nature protocols* 4(5):789–798. [PubMed: 19528954]
48. Brinkman EK, Chen T, Amendola M, & van Steensel B (2014) Easy quantitative assessment of genome editing by sequence trace decomposition. *Nucleic acids research* 42(22):e168. [PubMed: 25300484]
49. Pronier E, et al. (2018) Genetic and epigenetic evolution as a contributor to WT1-mutant leukemogenesis. *Blood* 132(12):1265–1278. [PubMed: 30064973]
50. Iyer G, et al. (2012) Genome sequencing identifies a basis for everolimus sensitivity. *Science* 338(6104):221. [PubMed: 22923433]
51. Al-Ahmadie H, et al. (2014) Synthetic lethality in ATM-deficient RAD50-mutant tumors underlies outlier response to cancer therapy. *Cancer discovery* 4(9):1014–1021. [PubMed: 24934408]
52. Li H & Durbin R (2009) Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25(14):1754–1760. [PubMed: 19451168]
53. DePristo MA, et al. (2011) A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nature genetics* 43(5):491–498. [PubMed: 21478889]
54. Cibulskis K, et al. (2013) Sensitive detection of somatic point mutations in impure and heterogeneous cancer samples. *Nature biotechnology* 31(3):213–219.
55. Ye K, Schulz MH, Long Q, Apweiler R, & Ning Z (2009) Pindel: a pattern growth approach to detect break points of large deletions and medium sized insertions from paired-end short reads. *Bioinformatics* 25(21):2865–2871. [PubMed: 19561018]
56. Robinson JT, et al. (2011) Integrative genomics viewer. *Nature biotechnology* 29(1):24–26.
57. Amarasinghe KC, et al. (2014) Inferring copy number and genotype in tumour exome data. *BMC genomics* 15:732. [PubMed: 25167919]
58. Bolger AM, Lohse M, & Usadel B (2014) Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30(15):2114–2120. [PubMed: 24695404]
59. Langmead B & Salzberg SL (2012) Fast gapped-read alignment with Bowtie 2. *Nature methods* 9(4):357–359. [PubMed: 22388286]
60. Li B & Dewey CN (2011) RSEM: accurate transcript quantification from RNA-Seq data with or without a reference genome. *BMC bioinformatics* 12:323. [PubMed: 21816040]
61. Robinson MD, McCarthy DJ, & Smyth GK (2010) edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics* 26(1):139–140. [PubMed: 19910308]
62. Subramanian A, et al. (2005) Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proceedings of the National Academy of Sciences of the United States of America* 102(43):15545–15550. [PubMed: 16199517]

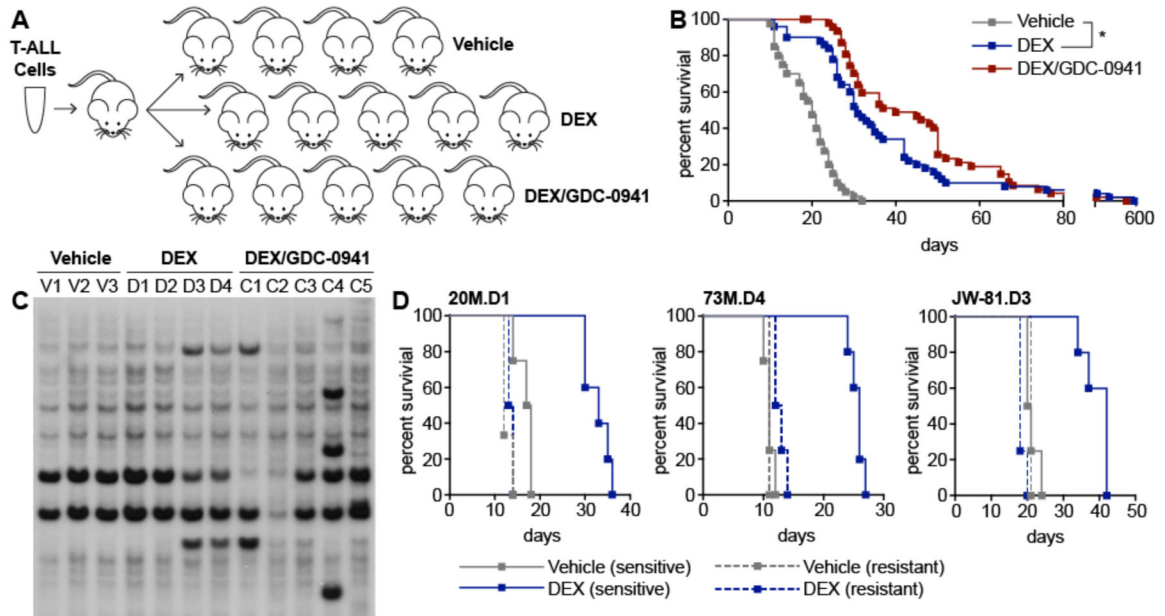


Figure 1. RIM-induced T-ALLs respond to dexamethasone alone or in combination with GDC-0941 and exhibit intrinsic glucocorticoid resistance upon re-treatment.

(A) Primary T-ALLs were first expanded in a single recipient. After euthanasia, 2×10^6 bone marrow cells were harvested and transplanted into 14 recipient mice, which were then randomly assigned to one of the three indicated treatment groups. (B) Kaplan-Meier survival analysis of recipient mice transplanted with 10 independent primary T-ALLs (five *Kras*^{WT} and five *Kras*^{G12D}) and treated with vehicle (n=40), DEX (n=50), or DEX/GDC-0941 (n=50) indicated that DEX significantly extended survival ($p < 0.0001$, Log-rank test), which was modestly but not significantly enhanced by GDC-0941 ($p = 0.0805$, Log-rank test). (C) Southern blot analysis with a probe for the MOL4070 virus of DNA extracted from leukemia cells harvested from individual moribund recipients of T-ALL 20M that were treated with vehicle (V1-V3), DEX (D1-D4), or the DEX/GDC-0941 combination (C1-C5). Novel restriction fragments indicative of retroviral integrations that are not present in the dominant parental clone are visible in relapsed leukemias D3, D4, C1, and C4. (D) Kaplan-Meier survival analysis of resistant T-ALLs (dotted lines) treated with vehicle (n=3) or DEX (n=4) compared to the corresponding drug-sensitive parental T-ALLs (solid lines) treated with vehicle (n=4) or DEX (n=5). Resistant T-ALLs demonstrated significantly shorter survival after DEX treatment versus the corresponding parental leukemias; 20M.D1 ($p = 0.0039$, Log-rank test), 73M.D4 ($p = 0.0027$, Log-rank test), and JW-81.D3 ($p = 0.0025$, Log-rank test).

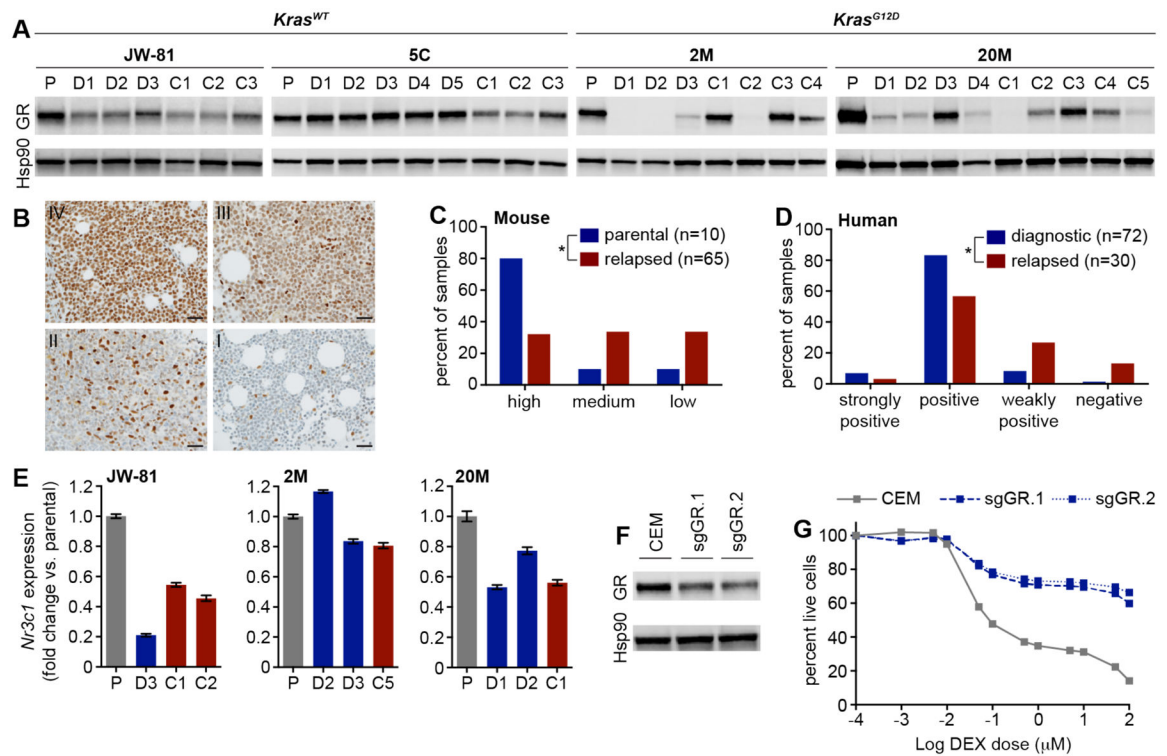


Figure 2. Relapsed mouse and human T-ALLs show reduced glucocorticoid receptor protein expression which confers dexamethasone resistance *in vitro*.

(A) Western blot analysis of bone marrow lysates from *Kras*^{WT} and *Kras*^{G12D} T-ALLs in parental (P), DEX treated (D) and DEX/GDC-0941 combination treated (C) mice at relapse. (B) Human bone marrow from T-ALL patients was subjected to immunohistochemistry analysis of GR protein expression and scored in a blinded fashion independently by two hematopathologists to classify samples as strongly positive (IV), positive (III), weakly positive (II) or negative (I). Scale bar represents 30 microns. Graphical representation of GR expression levels assessed by Western blot in primary murine T-ALLs (C) and by immunohistochemistry in human T-ALL patient samples (D). GR expression in murine T-ALLs was classified as low (examples in panel A include 2M.D1, 2M.C2, 20M.D4, 20M.C5), medium (examples include JW-81.C2, 5C.C2, 2M.C4, 20M.C4), or high (examples include 5C.D1, 2M.C1, 2M.C3, 20M.D3). Asterisk in panel C denotes a significant difference between high and medium/low classifications in parental versus relapsed murine T-ALLs (p-value=0.0109, Fisher's test). Asterisk in panel D denotes a significant difference between strongly positive/positive and weakly positive/negative classifications in diagnostic versus relapsed human T-ALLs (p-value=0.0013, Fisher's test). (E) Gene expression analysis using a TaqMan assay to measure *Nr3c1* transcript levels in three independent parental T-ALLs (P), and corresponding DEX (D) or DEX/GDC-0941 combination (C) treated relapsed T-ALLs. Error bars represent standard error of the mean for technical replicates. (F) Western blot analysis of GR expression in lysates from the CCRF-CEM human T-ALL cell line edited using nucleofection with Cas9 ribonucleoproteins containing two independent sgRNAs targeting the *Nr3c1* gene. (G) Analysis of viability in

the CCRF-CEM cells CRISPR edited at the GR locus and exposed to increasing doses of DEX. Error bars represent standard deviation of technical replicates.

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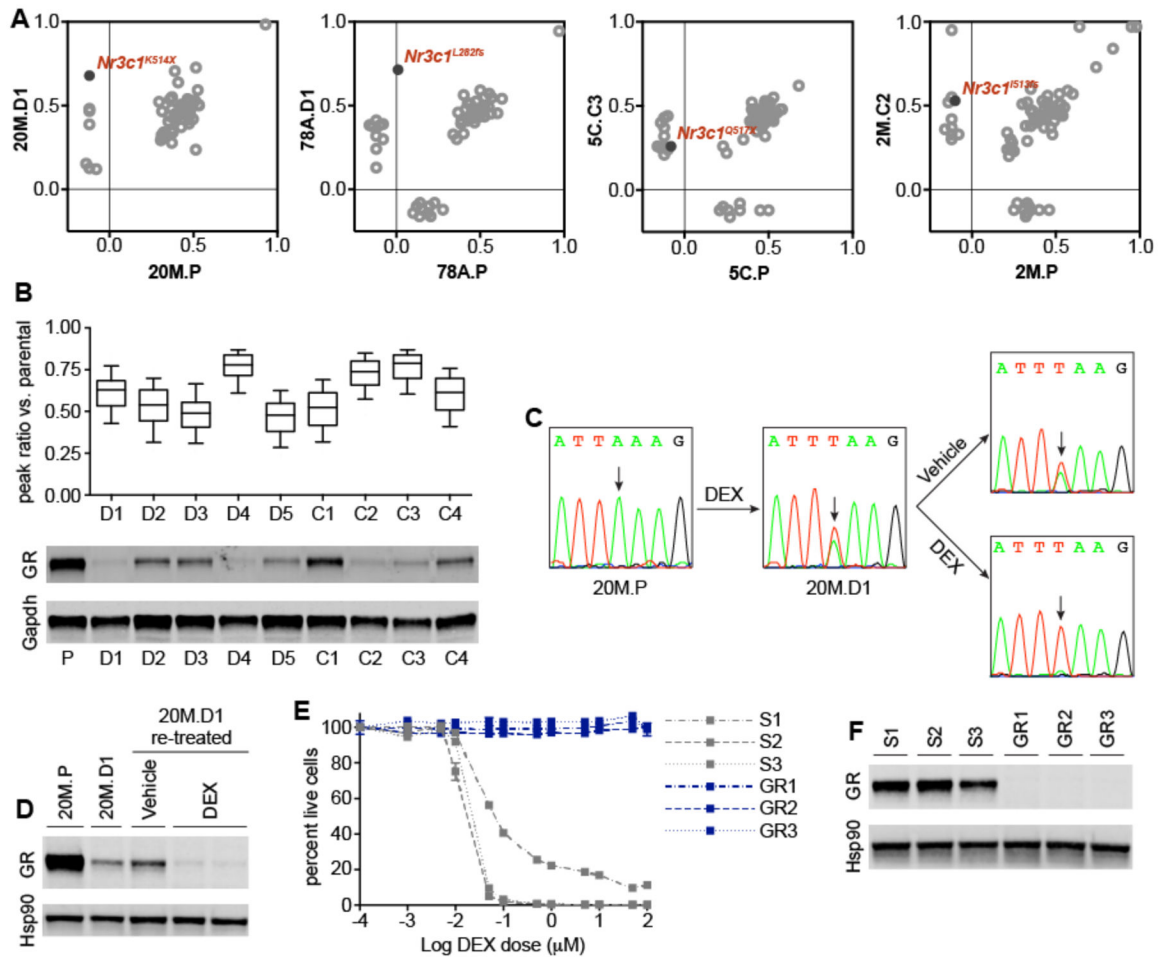


Figure 3. *Nr3c1* mutations in relapsed T-ALLs drive leukemia outgrowth *in vivo* and confer dexamethasone resistance *in vitro*.

(A) Mutant allele frequencies for mutations identified in the parental (P, x-axis) and the corresponding DEX-treated (D) or DEX/GDC-0941 combination-treated (C) relapsed T-ALLs (y-axis) highlighting the specific *Nr3c1* mutations or indels. (B) Box and whiskers plot showing allele frequency of the *Nr3c1* indel in each relapsed sample as compared to parental T-ALL 78A as determined from Sanger sequencing data (top), and corresponding Western blot data showing GR protein expression (bottom). Whiskers represent 10–90 percentile. (C) Sanger sequencing traces showing the site of the *Nr3c1* point mutation (arrow) in parental and relapsed 20M T-ALLs. Note that representation of the mutant allele is stable after transplantation in the absence of treatment, but increases markedly in secondary recipients that received DEX. (D) Western blot showing GR protein expression in the parental (P), relapsed (D) and re-treated 20M T-ALLs shown in panel C. (E) Viability of independent CRISPR-edited CCRF-CEM cell clones nucleofected with a scramble guide (S1, S2, S3) or with a guide targeting the region of exon 4 in which the mutation in T-ALL 20M.D was identified (GR1, GR2, GR3) in response to increasing doses of DEX. Error bars represent standard deviation of technical replicates. (F) Western blot analysis showing GR expression in lysates from the CCRF-CEM cell clones edited with scramble or *Nr3c1*-targeting guides.

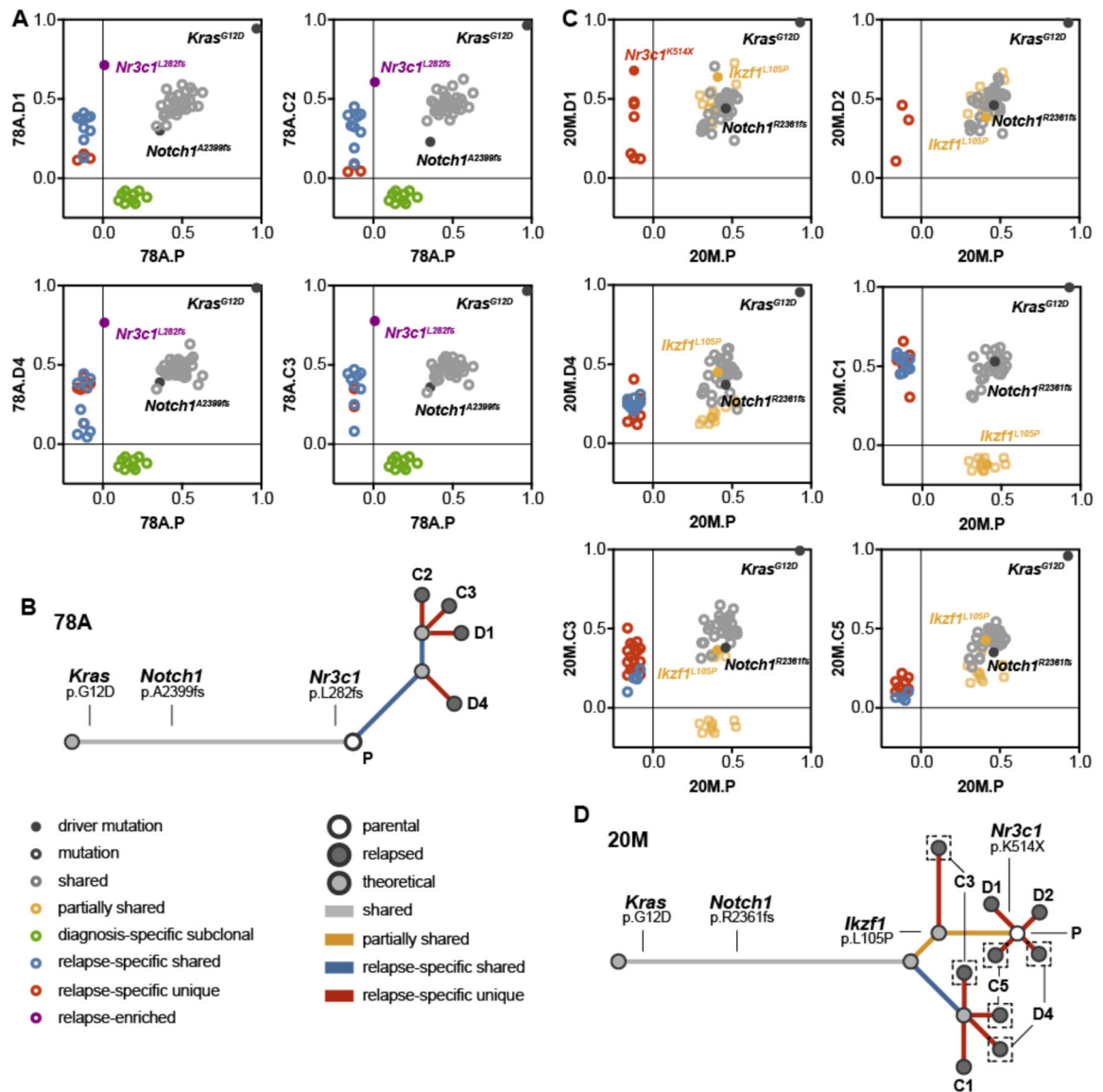


Figure 4. Relapsed T-ALLs with pre-existing or acquired *Nr3c1* mutations exhibit different evolutionary trajectories in response to dexamethasone treatment.

(A) Graphical representation of the mutant allele frequencies of missense mutations and indels shared between the parental T-ALL 78A (x-axis) and four independent relapsed leukemias (y-axis; grey), present only in the parental leukemia (green), shared only with other relapsed leukemias (blue), significantly enriched in the relapsed leukemia (purple), or specific to one of the relapsed leukemias (red). (B) Analysis of the evolutionary relationships among genetic alterations identified by WES in T-ALL 78A indicated a pattern of linear clonal evolution. (C) Graphical representation of the mutant allele frequencies shared between the parental T-ALL 20M (x-axis) and six independent relapsed leukemias (y-axis; grey), present in the parental and a subset of relapsed leukemias (yellow), shared only with other relapsed leukemias (blue), or specific to one of the relapsed leukemias (red). (D) Evolutionary analysis of the genetic alterations identified by WES in T-ALL 20M indicates a branching pattern of clonal dynamics involving multiple ancestral populations. Legends

indicate the type of mutation represented by each dot shown in panels *A* and *B* (left), and each dot and branch in panels *C* and *D*. The same color scheme is used in the evolutionary analysis of panels *A* through *D*.

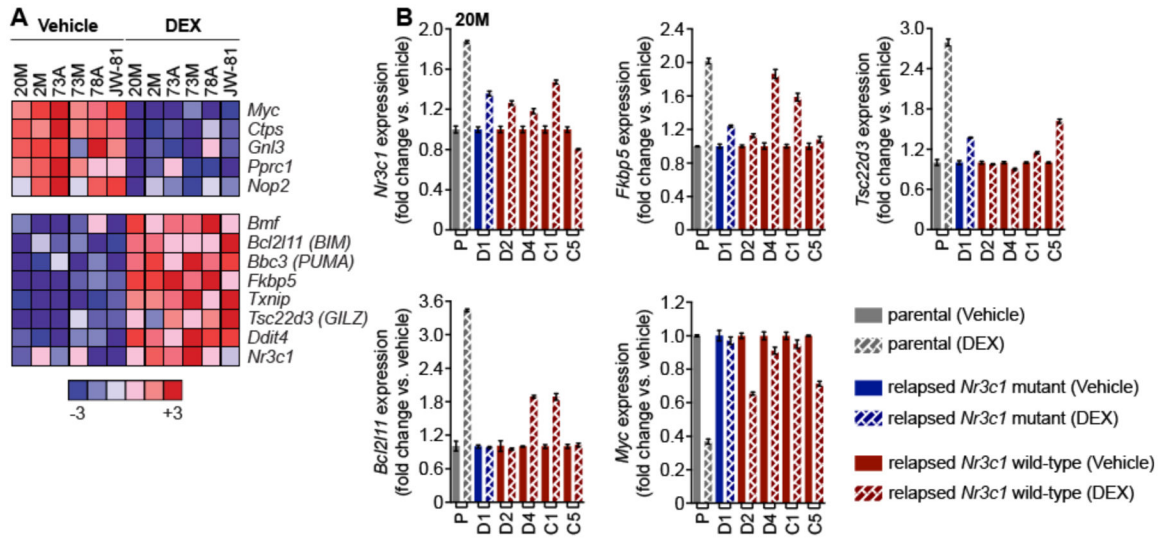


Figure 5. Blunted transcriptional responses of relapsed T-ALLs to dexamethasone treatment *in vivo*.

(A) Heatmap showing fold increased (p-value<0.001, FDR=0.002) or decreased (p-value<0.001, FDR=0.050) expression of selected GR-responsive genes after short-term *in vivo* DEX treatment measured by RNA-sequencing. (B) Gene expression analysis using TaqMan assays for each indicated gene in parental (P, grey bars) and relapsed (D) 20M T-ALLs with low GR protein expression with (blue bars) or without (red bars) an *Nr3c1* mutation showing transcript levels of GC response genes as compared to vehicle-treated controls. Error bars represent standard error of the mean for technical replicates.