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# Draft genome sequence of multidrug-resistant *Citrobacter portucalensis* BAU\_133-2 strain isolated from a domestic duck (*Anas platyrhynchos domesticus*) in Bangladesh

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**ABSTRACT** We announce the genome sequence of the *Citrobacter portucalensis* BAU\_133-2 strain isolated from a domestic duck. Our assembled genome contained a length of 4.8 Mb, 110.0× genome coverage, 51.91% of an average GC content, 1 plasmid, 1 CRISPR array, 8 prophages, 27 antibiotic resistance genes, and 75 virulence factor genes.

**KEYWORDS** *Citrobacter portucalensis*, duck, WGS, antimicrobial resistance, virulence determinants, metabolic functional features, Bangladesh

The widespread and improper utilization of antimicrobial agents has led to the rise of antimicrobial resistance in bacteria, giving rise to numerous multidrug-resistant (MDR) strains, which have now become a significant worldwide public health issue (1, 2). Ducks possess the capability to carry antimicrobial-resistant and MDR pathogens that may be transmitted to humans through their interactions with them (1, 3). *C. portucalensis* shows the potential to emerge as a critically important bacteria of global significance for both public health and One Health (4). Previously, antimicrobial-resistant *C. portucalensis* was isolated from humans, animals, and environments throughout the globe (5–8).

All the protocols and methodologies associated with the current study were approved by the Bangladesh Agricultural University's ethics committee, "Animal Welfare and Experimentation Ethical Committee" [AWEEC/BAU/2020(10)], Mymensingh, Bangladesh. Between January 2020 and January 2022, cloacal swab samples of diseased wild domestic ducks (Anas platyrhynchos domesticus) were taken from the Kishoreganj District (24.4260° N, 90.9821° E) of Bangladesh and transported to the laboratory. These samples were incubated overnight in nutrient broth (HiMedia, India) at 37°C. The incubated samples were streaked on xylose-lysine deoxycholate agar (HiMedia, India) media, and the resulting pure colonies underwent staining and biochemical tests to isolate C. portucalensis (9). The MDR properties of the isolates were ascertained through the disk diffusion method (10) and the CLSI guidelines (11). The C. portucalensis BAU\_133-2 isolate, showing phenotypic resistance to gentamicin, ciprofloxacin, cephalexin, azithromycin, tetracycline, ampicillin, cotrimoxazole, imipenem, and cefotaxime, was selected in this study. This isolate was cultured in nutrient broth (HiMedia, India) at 37°C overnight, and its complete genomic DNA was obtained from the broth culture using the Qiagen DNA mini kit (QIAGEN, Hilden, Germany). The extracted DNA from the isolate was then subjected to enzymatic fragmentation using the NEBNextdsDNA Fragmentase kit (NEB, MA, USA) according to the manufacturer's instructions. The fragmented DNA was subjected to size selection using SPRI beads, which helped in isolating DNA fragments of the desired size range for sequencing

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(12). Subsequently, a sequencing library was generated using the Nextera DNA Flex library prep kit (Illumina, San Diego, CA, USA). The library was then sequenced on the Illumina NextSeq2000 platform using 2  $\times$  150 paired-end reads. The genome assembly was performed using the Unicycler.v0.4.9 (13), and the raw paired-end reads (n = 7,587,670) were trimmed using Trimmomatic.v0.39 (14) (with parameters leading:20, slidingwindow:4:20:20, trailing:20, minlen = 36), with the aim of eliminating Illumina adapters, recognized Illumina irregularities, and phiX reads from the data set and the quality was evaluated using FastQC.v0.11.7 (15). Following that, the genome was annotated using PGAP.v3.0 (16). The antibiotic resistance genes (ARGs) were identified using CARD.v3.2.7 with RGI.v6.0.2 (17) and ResFinder.v4.1 (18); virulence factor genes (VFGs) by VFDB with VFanalyzer (19); the sequence type by MLST.v2.0 (20); metabolic functional features from RAST.v2.0 (21); and CRISPR array and prophages by CRISPRimmunity (22) within the assembled genome. Default parameters were used except where otherwise noted.

The draft assembly of the *C. portucalensis* BAU\_133-2 genome comprised 36 contigs, a G + C content of 51.91%, 4 contig L50, and the N50 value was 532,725 bp. The genome size of *C. portucalensis* BAU\_133-2 was 4,881,935 bp, with a genome coverage of 110.0×. This genome had a total of 4,574 genes, 4,493 CDSs, 71 transfer RNA genes, 4 ribosomal RNA genes, and 58 pseudogenes. This assembled genome identified one CRISPR array (with three signature genes, i.e., *cas3, csa3*, and *DEDDh*), eight prophages, and one plasmid [IncFIB(pB171)]. According to MLST analysis, this genome corresponds to sequence type ST211. The strain carried 27 predicted ARGs, 75 predicted VFGs, 383 subsystems with 34% coverage, and 2,118 genes.

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#### AUTHOR CONTRIBUTIONS

Tarana Ahmed, Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Writing – original draft | Md. Saiful Islam, Conceptualization, Data curation, Formal analysis, Methodology, Software, Writing – original draft, Writing – review and editing | Mohammad Nuruzzaman, Conceptualization, Methodology, Resources | Mohammad Sadekuzzaman, Formal analysis | S. M. Lutful Kabir, Supervision, Validation, Writing – review and editing | Md. Tanvir Rahman, Supervision, Validation, Writing – review and editing | Md. Shahidur Rahman Khan, Resources, Supervision, Validation, Writing – review and editing

#### DATA AVAILABILITY

The research on *C. portucalensis* BAU\_133-2, conducted using the WGS shotgun approach, was submitted to NCBI/GenBank, and it was assigned the accession number JAPQVZ000000000. The pertinent data, including the original readings, were stored with BioProject accession number PRJNA907481, BioSample accession number SAMN31981961, and SRA accession number SRR24872939. The specific version mentioned in this document is labeled as JAPQVZ00000000.

#### REFERENCES

- Ahmed T, Islam MS, Nuruzzaman M, Sadekuzzaman M, Kabir SML, Rahman MT, Khan MSR. 2023. Draft genome sequence of the multidrugresistant *Citrobacter freundii* 132-2 strain isolated from a domestic duck in Bangladesh. Microbiol Resour Announc 12:e0037823. https://doi.org/ 10.1128/mra.00378-23
- Islam MS, Rahman AT, Hassan J, Rahman MT. 2023. Extended-spectrum beta-lactamase in *Escherichia coli* isolated from humans, animals, and environments in Bangladesh: a one health perspective systematic review and meta-analysis. One Health 16:100526. https://doi.org/10. 1016/j.onehlt.2023.100526
- Ahmed T, Islam MS, Haider N, Elton L, Hasan B, Nuruzzaman M, Rahman MT, Kabir SML, Khan MSR. 2023. Phenotypic and genotypic characteristics of antimicrobial resistance in *Citrobacter freundii* isolated from domestic ducks (*Anas platyrhynchos domesticus*) in Bangladesh. Antibiotics (Basel) 12:769. https://doi.org/10.3390/antibiotics12040769
- Sellera FP, Fernandes MR, Fuga B, Fontana H, Vásquez-Ponce F, Goldberg DW, Monte DF, Rodrigues L, Cardenas-Arias AR, Lopes R, Cardoso B, Costa DGC, Esposito F, Lincopan N. 2022. Phylogeographical landscape of *Citrobacter portucalensis* carrying clinically relevant resistomes. Microbiol Spectr 10:e0150621. https://doi.org/10.1128/spectrum.01506-21
- Cao X, Xie H, Huang D, Zhou W, Liu Y, Shen H, Zhou K. 2021. Detection of a clinical carbapenem-resistant *Citrobacter portucalensis* strain and the dissemination of *C. portucalensis* in clinical settings. J Glob Antimicrob Resist 27:79–81. https://doi.org/10.1016/j.jgar.2021.04.027
- Hasan MS, Sultana M, Hossain MA. 2019. Complete genome arrangement revealed the emergence of a poultry origin superbug *Citrobacter portucalensis* strain NR-12. J Glob Antimicrob Resist 18:126–129. https:// doi.org/10.1016/j.jgar.2019.05.031
- Tsypin LM, Saunders SH, Bar-On Y, Leadbetter JR, Newman DK. 2020. Draft genome sequence of the redox-active enteric bacterium *Citrobacter portucalensis* strain MBL. Microbiol Resour Announc 9:e00695-20. https://doi.org/10.1128/MRA.00695-20
- Igbinosa EO, Rathje J, Habermann D, Brinks E, Cho G-S, Franz C. 2018. Draft genome sequence of multidrug-resistant strain *Citrobacter portucalensis* MBTC-1222, isolated from uziza (piper guineense) leaves in Nigeria. Genome Announc 6:e00123-18. https://doi.org/10.1128/ genomeA.00123-18
- Ribeiro TG, Gonçalves BR, da Silva MS, Novais Â, Machado E, Carriço JA, Peixe L. 2017. *Citrobacter Portucalensis* sp. nov., isolated from an aquatic sample. Int J Syst Evol Microbiol 67:3513–3517. https://doi.org/10.1099/ ijsem.0.002154
- Bauer AW, Kirby WMM, Sherris JC, Turck M. 1966. Antibiotic susceptibility testing by a standardized single disk method. Am J Clin Pathol 45:493– 496.

- 11. M100-S32. 2022. Performance standards for antimicrobial susceptibility testing. Clinical and Laboratory Standards Institute, Wayne, PA, USA.
- Liu D, Li Q, Luo J, Huang Q, Zhang Y. 2023. An SPRI beads-based DNA purification strategy for flexibility and cost-effectiveness. BMC Genomics 24:125. https://doi.org/10.1186/s12864-023-09211-w
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/ 10.1093/bioinformatics/btu170
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. Available from: http://www.bioinformatics.babraham.ac. uk/projects/fastqc/
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44:6614– 6624. https://doi.org/10.1093/nar/gkw569
- 17. Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W, Nguyen A-L, Cheng AA, Liu S, Min SY, Miroshnichenko A, Tran H-K, Werfalli RE, Nasir JA, Oloni M, Speicher DJ, Florescu A, Singh B, Faltyn M, Hernandez-Koutoucheva A, Sharma AN, Bordeleau E, Pawlowski AC, Zubyk HL, Dooley D, Griffiths E, Maguire F, Winsor GL, Beiko RG, Brinkman FSL, Hsiao WWL, Domselaar GV, McArthur AG. 2020. CARD 2020: antibiotic resistome surveillance with the comprehensive antibiotic resistance database. Nucleic Acids Res 48:D517–D525. https:// doi.org/10.1093/nar/gkz935
- Florensa AF, Kaas RS, Clausen P, Aytan-Aktug D, Aarestrup FM. 2022. ResFinder - an open online resource for identification of antimicrobial resistance genes in next-generation sequencing data and prediction of phenotypes from genotypes. Microb Genom 8:000748. https://doi.org/ 10.1099/mgen.0.000748
- Liu B, Zheng D, Zhou S, Chen L, Yang J. 2022. VFDB 2022: a general classification scheme for bacterial virulence factors. Nucleic Acids Res 50:D912–D917. https://doi.org/10.1093/nar/gkab1107
- Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total-genome-sequenced bacteria. J Clin Microbiol 50:1355–1361. https://doi.org/10.1128/JCM.06094-11
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75

22. Zhou F, Yu X, Gan R, Ren K, Chen C, Ren C, Cui M, Liu Y, Gao Y, Wang S, Yin M, Huang T, Huang Z, Zhang F. 2023. CRISPRimmunity: an interactive web server for CRISPR-associated important molecular events and modulators used in geNome edIting tool identifYing. Nucleic Acids Res 51:W93–W107. https://doi.org/10.1093/nar/gkad425