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### PM<sub>2.5</sub> Concentrations in a Cannabis Store with On-Site Consumption

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#### INTRODUCTION

Recently, California and other states have legalized the use of cannabis in stores, giving people who cannot consume cannabis in their homes a safe and legal place to consume it. However, on-site consumption may expose customers and workers to particulate air pollution. Consumption methods that use temperatures below combustion to aerosolize cannabis are a way to reduce exposure to toxicants (Gieringer et al. 2004). In vaporization of cannabis flower, an aerosol is formed by passing heated air through finely-ground, dried flower. Cannabis concentrates can be consumed by dabbing, where a small amount of concentrate is applied to a heated surface to create an aerosol. Like smoking, vaporizing and dabbing create aerosols that contain particles 2.5 micrometers in diameter and smaller (PM<sub>2.5</sub>) (Jaques et al. 2018) that can penetrate deep into the lung. To assess the effects of on-site consumption of cannabis on PM<sub>2.5</sub> concentrations, we measured PM<sub>2.5</sub> in the retail and consumption space of a cannabis store (a dispensary), where smoking was banned but vaporizing and dabbing were permitted.

#### METHODS

PM<sub>2.5</sub> concentrations were measured continuously, using two, co-located laser photometers (Model AM510, TSI Inc., Shoreview MN), placed 80-100 cm above the floor, for five weeks in 2019. Room occupancy was not monitored. In week 1, instruments were located 30-122 cm from the sources (vaporizers and dab rigs). During week 2 and weeks 3-5, they were 6-9 and 2-4 meters from the nearest sources, respectively. Photometers were operated with impactors to exclude particles over 2.5 µm in diameter. The photometers were zeroed once a day and calibrated gravimetrically using a controlled cigarette smoke generation system (Schick et al.

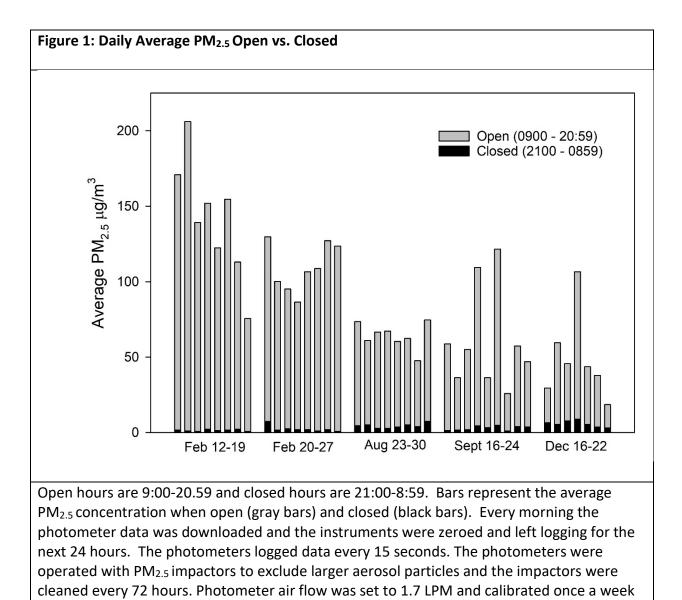
2012) before and after each experiment. Gravimetric data from 20 cigarette smoke experiments, when plotted against the matching photometric data and forced through zero, yielded a calibration factor of 0.31 (R<sup>2</sup> = 0.84), which was was applied to the dispensary photometric data. Cannabis PM<sub>2.5</sub> samples were also collected in the dispensary on filters (EMFAB, Pall Corporation, Cortland, NY) for one week (12/19), and a preliminary photometer calibration factor was calculated as above. PM<sub>2.5</sub> concentrations in outdoor air were estimated using data from an US EPA monitoring station located 2.5 km (1.5 m) from the dispensary in an area with similar ambient pollution sources.

#### RESULTS

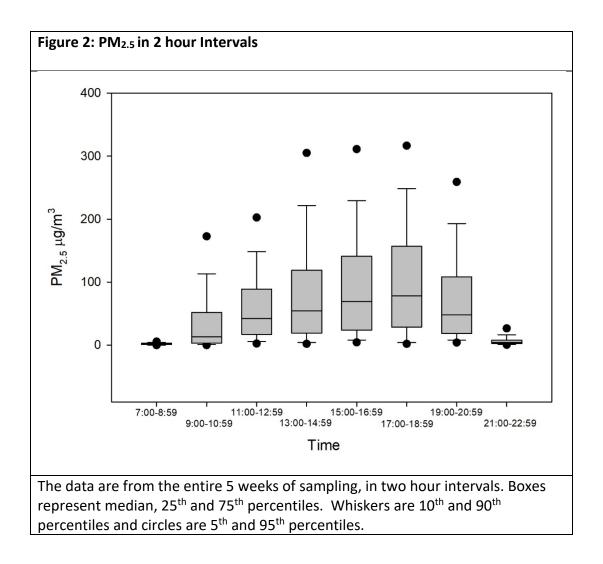
The retail and consumption space was a single room of approximately 400 m<sup>3</sup>. Cannabis consumption occurred at three tables in one corner of the room, with sales counters located in the opposite corner. The room was served by building HVAC and by four window air conditioners that did not admit fresh air. The air conditioners had dust filters and we were unable to examine filtration in the building HVAC system. The dispensary provided electrically-heated cannabis flower vaporizers and dab rigs for use. Smoking (combustion) of cannabis and tobacco were not permitted.

We monitored PM<sub>2.5</sub> in the dispensary for 38 days and 16 hours. During business hours, the average PM<sub>2.5</sub> concentration was 84  $\mu$ g/m<sup>3</sup>, with a standard deviation of ± 124  $\mu$ g/m<sup>3</sup> (Figure 1), an interquartile range of 16-111  $\mu$ g/m<sup>3</sup> and a median of 47  $\mu$ g/m<sup>3</sup>. When the business was closed, the average PM<sub>2.5</sub> concentration was 3 ± 7  $\mu$ g/m<sup>3</sup>, the IQR of 1-4  $\mu$ g/m<sup>3</sup> and the median was 2  $\mu$ g/m<sup>3</sup>. When examined in two-hour intervals, the median PM<sub>2.5</sub>

concentration was highest between 5:00 and 7:00 PM, at 76  $\mu$ g/m<sup>3</sup> (Figure 2). The average PM<sub>2.5</sub> concentration outdoors was 6 ± 4  $\mu$ g/m<sup>3</sup> during business hours and 6 ± 5  $\mu$ g/m<sup>3</sup> when the business was closed. The dispensary gravimetric data yielded a photometer calibration factor of 0.57 (R<sup>2</sup> = 0.43).



with a soap bubble spirometer (Gilibrator-1, Sensidyne, LP. St Petersburg, FL).



### DISCUSSION

Our data show a clear association between the consumption of cannabis and elevated PM<sub>2.5</sub> concentrations in the dispensary. The average PM<sub>2.5</sub> concentration when the business was open was 28 times higher than when the business was closed, the median concentration was 23.5 times higher and peak daily particle concentrations corresponded with the busiest hours. The PM<sub>2.5</sub> concentrations in this cannabis dispensary are similar to those observed in indoor spaces where smoking is permitted (California Air Resources Board 2005). These findings are some of the first field measurements of PM<sub>2.5</sub> emissions from cannabis flower vaporizers and

dabbing of cannabis concentrates. In a space with similar ventilation and consumption activity, it is likely that dabbing and vaporizing would create lower PM<sub>2.5</sub> concentrations than smoking, because smoking decomposes the cannabis more completely, creating more sidestream smoke.

### Limitations

Most of our data are from TSI Sidepak laser photometers, which are factory-calibrated to NIST standard A1 test dust (ISO 12103-1). To deliver accurate measurements of any other aerosol, a specific calibration factor is required. As of this writing, there are no published calibration factors for aerosols created by vaporizing cannabis flower or dabbing cannabis concentrates and little is known of their properties. The gravimetric data from the dispensary yielded a calibration factor of 0.57, but variation was high (R<sup>2</sup> = 0.41) because there were only seven daylong samples. We therefore used the well-validated calibration factor for secondhand cigarette smoke (0.31) (Hyland et al. 2008) to adjust our data. It is unlikely to yield inflated values and if the true calibration factor is higher, that does not affect our finding that on-site consumption was associated with strong and consistent increases in PM<sub>2.5</sub>.

### CONCLUSION

Our data demonstrate that consumption of cannabis products indoors increased  $PM_{2.5}$  concentrations. Psychoactive effects through passive exposure are unlikely (Herrmann et al. 2015). However, exposure to  $PM_{2.5}$  can cause changes in cardiovascular function that increase the risk of myocardial infarction and death (Brook et al. 2010). In healthy nonsmokers, even 30 minutes of exposure to cigarette smoke, at concentrations below 200  $\mu$ g/m<sup>3</sup> PM<sub>2.5</sub>,

decreased endothelial function, a well-validated predictor of increased risk of cardiovascular disease (Yeboah et al. 2009, Frey et al. 2012). It is possible that the aerosols from vaporizers and dabbing are less toxic than standard combustion aerosols. However, even brief increases in ambient  $PM_{2.5}$  from mixed sources are associated with increases in myocardial infarction and total mortality (Brook et al. 2010) and these effects are detectable even at PM2.5 increases of 10 µg/m<sup>3</sup> (Di et al. 2017). It is likely that the  $PM_{2.5}$  concentrations we observed are high enough to cause health problems for some individuals. Further research on the toxicity of cannabis smoke and vaporizer and dabbing aerosols is necessary.

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#### REFERENCES

Brook, R. D., S. Rajagopalan, C. A. Pope, 3rd, J. R. Brook, A. Bhatnagar, A. V. Diez-Roux, F. Holguin, Y. Hong, R. V. Luepker, M. A. Mittleman, A. Peters, D. Siscovick, S. C. Smith, Jr., L. Whitsel, J. D. Kaufman, E. American Heart Association Council on, C. o. t. K. i. C. D. Prevention, P. A. Council on Nutrition and Metabolism (2010). "Particulate matter air pollution and cardiovascular disease: An update to the scientific statement from the american heart association." <u>Circulation</u> **121**(21): 2331-2378.

California Air Resources Board (2005). Proposed identification of environmental tobacco smoke as a toxic air contaminant. <u>Toxi air contaminants</u>. California Environmental Protection Agency, A. R. Board and Office of Environmental Health Hazard Assessment. Oakland, California: 805.

Di, Q., L. Dai, Y. Wang, A. Zanobetti, C. Choirat, J. D. Schwartz and F. Dominici (2017). "Association of short-term exposure to air pollution with mortality in older adults." <u>JAMA</u> **318**(24): 2446-2456. Frey, P. F., P. Ganz, P. Y. Hsue, N. L. Benowitz, S. A. Glantz, J. R. Balmes and S. F. Schick (2012). "The exposure-dependent effects of aged secondhand smoke on endothelial function." <u>J Am Coll Cardiol</u> **59**(21): 1908-1913.

Gieringer, D. H., J. St. Laurent and S. Goodrich (2004). "Cannabis vaporizer combines efficient delivery of thc with effective suppression of pyrolytic compounds." Journal of Cannabis Therapeutics **4**(1): 7-27.

Herrmann, E. S., E. J. Cone, J. M. Mitchell, G. E. Bigelow, C. LoDico, R. Flegel and R. Vandrey (2015). "Non-smoker exposure to secondhand cannabis smoke ii: Effect of room ventilation on the physiological, subjective, and behavioral/cognitive effects." <u>Drug Alcohol Depend</u> **151**: 194-202.

Hyland, A., M. J. Travers, C. Dresler, C. Higbee and K. M. Cummings (2008). "A 32-country comparison of tobacco smoke derived particle levels in indoor public places." <u>Tob Control</u> **17**(3): 159-165.

Jaques, P., M. Zalay, A. Huang, K. Jee and S. F. Schick (2018). Measuring aerosol particle emissions from cannabis vaporization and dabbing. <u>Indoor Air 2018: The 15th conference of the international society for indoor air quality and climate</u>. Philadelphia, PA, International Society for Indoor Air Quality and Climate.

Schick, S. F., K. F. Farraro, J. Fang, S. Nasir, J. S. Kim, D. Lucas, H. Wong, J. Balmes, D. K. Giles and B. Jenkins (2012). "An apparatus for generating aged cigarette smoke for controlled human exposures studies." <u>Aerosol Sci Technol</u> **46**: 1246-1255.

Yeboah, J., A. R. Folsom, G. L. Burke, C. Johnson, J. F. Polak, W. Post, J. A. Lima, J. R. Crouse and D. M. Herrington (2009). "Predictive value of brachial flow-mediated dilation for incident cardiovascular events in a population-based study: The multi-ethnic study of atherosclerosis." <u>Circulation</u> **120**(6): 502-509.