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Consumption of Fructose and High Fructose Corn Syrup Increase Postprandial Triglycerides, LDL-Cholesterol, and Apolipoprotein-B in Young Men and Women

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Context: The American Heart Association Nutrition Committee recommends women and men consume no more than 100 and 150 kcal of added sugar per day, respectively, whereas the Dietary Guidelines for Americans, 2010, suggests a maximal added sugar intake of 25% or less of total energy.

Objective: To address this discrepancy, we compared the effects of consuming glucose, fructose, or high-fructose corn syrup (HFCS) at 25% of energy requirements (E) on risk factors for cardiovascular disease.

Participants, Design and Setting, and Intervention: Forty-eight adults (aged 18–40 yr; body mass index 18–35 kg/m²) resided at the Clinical Research Center for 3.5 d of baseline testing while consuming energy-balanced diets containing 55% E complex carbohydrate. For 12 outpatient days, they consumed usual ad libitum diets along with three servings per day of glucose, fructose, or HFCS-sweetened beverages (n = 16/group), which provided 25% E requirements. Subjects then consumed energy-balanced diets containing 25% E sugar-sweetened beverages/30% E complex carbohydrate during 3.5 d of inpatient intervention testing.

Main Outcome Measures: Twenty-four-hour triglyceride area under the curve, fasting plasma low-density lipoprotein (LDL), and apolipoprotein B (apoB) concentrations were measured.

Results: Twenty-four-hour triglyceride area under the curve was increased compared with baseline during consumption of fructose (+4.7 ± 1.2 mmol/liter × 24 h, P = 0.0032) and HFCS (+1.8 ± 1.4 mmol/liter × 24 h, P = 0.035) but not glucose (−1.9 ± 0.9 mmol/liter × 24 h, P = 0.14). Fasting LDL and apoB concentrations were increased during consumption of fructose (LDL: +0.29 ± 0.082 mmol/liter, P = 0.0023; apoB: +0.093 ± 0.022 g/liter, P = 0.0005) and HFCS (LDL: +0.42 ± 0.11 mmol/liter, P < 0.0001; apoB: +0.12 ± 0.031 g/liter, P < 0.0001) but not glucose (LDL: +0.012 ± 0.071 mmol/liter, P = 0.86; apoB: +0.0097 ± 0.019 g/liter, P = 0.90).

Conclusions: Consumption of HFCS-sweetened beverages for 2 wk at 25% E increased risk factors for cardiovascular disease comparably with fructose and more than glucose in young adults. (J Clin Endocrinol Metab 96: E1596–E1605, 2011)
In epidemiological studies, consumption of sugar and/or sugar-sweetened beverages has been linked to the presence of unfavorable lipid levels (1–5), insulin resistance (6, 7), fatty liver (8, 9), type 2 diabetes (10–12), cardiovascular disease (13), and metabolic syndrome (14). We have recently reported that consumption of fructose-sweetened beverages at 25% of energy requirements (E) increased visceral adipose deposition and de novo lipogenesis, produced dyslipidemia, and decreased glucose tolerance/insulin sensitivity in older, overweight/obese men and women, whereas consumption of glucose-sweetened beverages did not (15). Because the commonly consumed sugars, sucrose and high-fructose corn syrup (HFCS), are composed of 50–55% fructose, these results provide a potential mechanistic explanation for the associations between sugar consumption and metabolic disease. However, the adverse metabolic effects of fructose consumption observed in the older, overweight/obese population (15) may not occur in a younger, leaner population.

Authors of three recent reviews have concluded that long-term sugar intakes as high as 25–50% E have no adverse effects with respect to components of metabolic syndrome (16) and that fructose consumption up to 140 g/d does not result in biologically relevant increases of fasting or postprandial triglycerides (TG) in healthy, normal-weight (17), or overweight or obese (18) humans. These reviews (16, 17) are cited in the Report of the Dietary Guidelines Advisory Committee on the Dietary Guidelines for Americans 2010, released June of 2010, in which a maximal intake level of 25% or less of total energy from added sugars is suggested (19). However, in August of 2009, the American Heart Association Nutrition Committee recommended that women consume no more than 100 kcal/d and men consume no more than 150 kcal/d of added sugar (20). This equates to differences between the two guidelines of 400 kcal/d for women consuming 2000 kcal/d and 525 kcal/d for men consuming 2500 kcal/d. To address this discrepancy, we compared the effects of consuming 25% E as glucose, fructose or HFCS for 2 weeks on risk factors for cardiovascular disease in young adults.

Materials and Methods

The subjects who participated in this study are a subgroup of participants from an ongoing 5-yr National Institutes of Health-funded investigation in which a total of eight experimental groups (n = 25/group) will be studied. The objectives include comparing the metabolic effects of fructose, glucose, and HFCS consumption at 25% E and to compare the metabolic effects of fructose and HFCS consumption at 0, 10, 17.5, and 25% E. The results reported in this paper are from the first 48 subjects to complete the study protocol in the experimental groups consuming 25% E as glucose, fructose, or HFCS (n = 16/group). Participants were recruited through an internet listing (Craigslist.com) and underwent telephone and in-person interviews with medical history, complete blood count, and serum biochemistry panel to assess eligibility. Inclusion criteria included age 18–40 yr and body mass index (BMI) 18–35 kg/m² with a self-report of stable body weight during the prior 6 months. Exclusion criteria included diabetes (fasting glucose >125 mg/dl), evidence of renal or hepatic disease, fasting plasma TG greater than 400 mg/dl, hypertension (>140/90 mm Hg), or surgery for weight loss. Individuals who smoked, habitually ingested more than two alcoholic beverages per day, exercised more than 3.5 h/wk at a level more vigorous than walking, or used thyroid, lipid-lowering, glucose-lowering, antihypertensive, antidepressant, or weight loss medications were also excluded. The University of California, Davis, Institutional Review Board approved the experimental protocol for this study, and subjects provided written informed consent to participate.

For the 5 wk before study, subjects were asked to limit daily consumption of sugar-containing beverages to one 8-oz serving of fruit juice. Fifty-five subjects were enrolled in the experimental groups consuming 25% E as glucose, fructose, or HFCS. Four subjects withdrew due to unwillingness to comply with the study protocol (two in the HFCS group, two before group assignment), and two were withdrawn due to medical conditions not apparent during screening (HFCS and glucose group). The samples from one subject (HFCS group) who completed the study protocol were not analyzed because of illness during the 24-h serial blood collection. The experimental groups were matched for gender (nine men, seven women/group), BMI, fasting TG, cholesterol, high-density lipoprotein (HDL) and insulin concentrations. The subjects and University of California, Davis, Clinical Research Center (CCRC) and technical personnel were blinded to the sugar assignments.

This was a parallel-arm, diet intervention study with three phases: 1) a 3.5-d inpatient baseline period during which subjects resided at the CCRC; 2) a 12-d outpatient intervention period; and 3) a 3.5-d inpatient intervention period at the CCRC. During d 2 and 3 of the baseline and intervention inpatient periods, subjects consumed energy-balanced meals consisting of conventional foods. Daily energy requirements were calculated by the Mifflin equation (21) with adjustment of 1.3 for activity on the days of 24-h serial blood collections, and adjustment of 1.5 for the other days. The baseline diet contained 55% E mainly as complex carbohydrate, 30% fat, and 15% protein. The intervention inpatient meals were as identical as possible to baseline meals, excepting the carbohydrate component consisted of 25% E as glucose-, fructose-, or HFCS-sweetened beverages and 30% E as complex carbohydrate. Sugar-sweetened beverages were provided to subjects as three daily servings consumed with meals and were flavored with an unsweetened drink mix (Kool-Aid; Kraft Foods, Northfield, IL). The timing of inpatient meal service and the energy distribution were: breakfast, 0900 h (25%); lunch, 1300 h (35%); dinner, 1800 h (40%).

During the 12-d outpatient phase of the study, the subjects were provided with and instructed to drink three servings of sugar-sweetened beverage per day (one per meal), to consume their usual diet, and to not consume other sugar-containing beverages, including fruit juice. To monitor compliance, the sugar-sweetened beverages contained a biomarker (riboflavin), which was measured fluorometrically in urine samples collected at the time of beverage pickup. These measurements indicated that the three groups of subjects were comparably compliant.
Twenty-four-hour serial blood collections were conducted on the third day of the baseline (0 wk) and intervention (2 wk) inpatient periods. Three fasting blood samples were collected at 0800, 0830, and 0900 h. Twenty-nine postprandial blood samples were collected at 30- to 60-min intervals from 0930 until 0800 h the next morning. Additional 6-ml samples were collected at the fasting time points, 0800, 0830, and 0900 h and also at 2200, 2300, and 2400 h, the period during which TG concentrations peaked during our previous study (13). The additional plasma from the three fasting samples was pooled, as was that from the three late-evening postprandial samples; multiple aliquots of each pooled sample were stored at −80 C.

Analyses

Primary outcomes include fasting TG, 24-h TG incremental area under the curve (AUC), late-evening postprandial TG concentrations, and fasting LDL, non-HDL-cholesterol (−C), apolipoprotein (apo)B concentrations, and the apoB to apoAI ratio. Secondary outcomes included body weight, fasting HDL, postprandial LDL, non-HDL-C, apoB, remnant lipoprotein-cholesterol (RLP)-C and RLP-TG, and fasting and postprandial small dense LDL-cholesterol (sdLDL-C). Fasting concentrations, 24-h AUC, and postmeal peaks for glucose and insulin, and homeostasis model assessment insulin resistance index (HOMA-IR) are presented in the online supplement, published on The Endocrine Society's Journals Online web site at http://jcem.endojournals.org. Fasting measures were conducted on samples collected or pooled from the 0800, 0830, and 0900 h time points, and postprandial measures were conducted on samples collected or pooled from the 2200, 2300, and 2400 h time points. Lipid and lipoprotein concentrations (total cholesterol, HDL, TG, apoB, apoA1) were determined with a Polychem chemistry analyzer (PolymedCo, Inc., Cortlandt Manor, NY). LDL concentrations were determined by direct homogenous assay using detergents (Denka Seiken, Tokyo, Japan) (22) and sdLDL-C concentrations were quantified using the sdLDL-C-EX"SEIKEN" homogeneous assay kit (Denka Seiken) (23). RLP concentrations were quantified with an immunopreparation assay (24). Glucose was measured with an automated glucose analyzer (YSI, Inc., Yellow Springs, OH), and insulin by RIA (Millipore, St. Charles, MO).

The incremental 24-h area AUC was calculated for TG, glucose, and insulin by the trapezoidal method. Glucose and insulin postmeal peaks were assessed as the mean amplitudes of the three postmeal peaks; specifically the peak postmeal value minus the premeal value was averaged for breakfast, lunch, and dinner for each subject. The absolute change (Δ from 2 wk when 25% E carbohydrate at baseline (0 wk) and at the end of the 2-wk intervention. The 24-h TG profiles during baseline and the end of the 2-wk intervention are shown in Fig. 1, A–C. The 24-h TG AUC (Fig. 2A) was significantly increased compared with baseline (LS means of Δ different from zero) in subjects consuming fructose (+4.7 ± 1.2 mmol/liter × 24 h, P = 0.0032) and HFCS (+1.8 ± 1.4 mmol/liter × 24 h, P = 0.035), whereas it tended to decrease during consumption of glucose (−1.9 ± 0.9 mmol/liter × 24 h, P = 0.14). The consumption of all three sugars resulted in a late-evening TG peak between 2200 and 2400 h that was not apparent when complex carbohydrate was consumed (Fig. 1, A–C). The late-evening peaks (Fig. 2B) were significantly increased compared with baseline during consumption of fructose (+0.59 ± 0.11 mmol/liter, P < 0.0001) and HFCS (+0.46 ± 0.082 mmol/liter, P < 0.0001) but not by glucose (+0.22 ± 0.10 mmol/liter, P = 0.077). All three sugars tended to increase fasting TG, but this was significant only in the group consuming glucose (Fig. 2C). Fasting LDL-C concentrations (Fig. 3A) were increased during consumption of fructose (+0.29 ± 0.082 mmol/liter, P = 0.0023) and HFCS (+0.42 ± 0.11 mmol/liter, P < 0.0001) but not glucose (+0.012 ± 0.071 mmol/liter, P = 0.86).

### TABLE 1. Subjects’ baseline anthropomorphic and metabolic parameters

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Glucose (n = 16)</th>
<th>Fructose (n = 16)</th>
<th>HFCS (n = 16)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yr)</td>
<td>27.0 ± 7.2</td>
<td>28.0 ± 6.8</td>
<td>27.8 ± 7.6</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>76.8 ± 14.1</td>
<td>76.8 ± 10.6</td>
<td>74.3 ± 14.9</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>26.2 ± 3.6</td>
<td>25.4 ± 3.8</td>
<td>24.9 ± 4.8</td>
</tr>
<tr>
<td>Waist circumference (cm)</td>
<td>80.6 ± 10.4</td>
<td>77.8 ± 9.6</td>
<td>78.0 ± 10.8</td>
</tr>
<tr>
<td>Body fat (%)</td>
<td>28.0 ± 9.3</td>
<td>26.7 ± 11.8</td>
<td>25.0 ± 10.1</td>
</tr>
<tr>
<td>TG (mmol/liter)</td>
<td>1.2 ± 0.5</td>
<td>1.2 ± 0.4</td>
<td>1.3 ± 0.6</td>
</tr>
<tr>
<td>Total cholesterol (mmol/liter)</td>
<td>4.5 ± 0.8</td>
<td>3.9 ± 0.8</td>
<td>4.1 ± 0.8</td>
</tr>
<tr>
<td>HDL-C (mmol/liter)</td>
<td>1.2 ± 0.4</td>
<td>1.2 ± 0.4</td>
<td>1.2 ± 0.4</td>
</tr>
<tr>
<td>Insulin (pmol/liter)</td>
<td>97.9 ± 30.4</td>
<td>102.8 ± 86.4</td>
<td>89.1 ± 31.6</td>
</tr>
</tbody>
</table>

*P > 0.05 for differences among groups at baseline for all parameters, PROC MIXED ANOVA. Mean ± SD.

Results

There were no significant differences among the three experimental groups in anthropomorphic (Table 1) or outcome measures at baseline (Tables 2 and 3 and Supplemental Table 1). Body weight (Table 3) and blood pressure (data not shown) were not affected by 2 wk consumption of glucose, fructose, or HFCS.

Primary outcomes: comparing glucose, fructose, and HFCS with complex carbohydrate consumption

Table 2 presents the primary outcomes during consumption of complex carbohydrate at baseline (0 wk) and at the end of the 2-wk sugar interventions. The 24-h TG profiles during baseline and the end of the 2-wk intervention are shown in Fig. 1, A–C. The 24-h TG AUC (Fig. 2A) was significantly increased compared with baseline (LS means of Δ different from zero) in subjects consuming fructose (+4.7 ± 1.2 mmol/liter × 24 h, P = 0.0032) and HFCS (+1.8 ± 1.4 mmol/liter × 24 h, P = 0.035), whereas it tended to decrease during consumption of glucose (−1.9 ± 0.9 mmol/liter × 24 h, P = 0.14). The consumption of all three sugars resulted in a late-evening TG peak between 2200 and 2400 h that was not apparent when complex carbohydrate was consumed (Fig. 1, A–C). The late-evening peaks (Fig. 2B) were significantly increased compared with baseline during consumption of fructose (+0.59 ± 0.11 mmol/liter, P < 0.0001) and HFCS (+0.46 ± 0.082 mmol/liter, P < 0.0001) but not by glucose (+0.22 ± 0.10 mmol/liter, P = 0.077). All three sugars tended to increase fasting TG, but this was significant only in the group consuming glucose (Fig. 2C). Fasting LDL-C concentrations (Fig. 3A) were increased during consumption of fructose (+0.29 ± 0.082 mmol/liter, P = 0.0023) and HFCS (+0.42 ± 0.11 mmol/liter, P < 0.0001) but not glucose (+0.012 ± 0.071 mmol/liter, P = 0.86).
TABLE 2. Primary outcomes during consumption of complex carbohydrates at 0 wk and during consumption of glucose-, fructose-, or HFCS-sweetened beverages at 2 wk

<table>
<thead>
<tr>
<th>Primary outcomes</th>
<th>Glucose</th>
<th>Fructose</th>
<th>HFCS</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0 wk</td>
<td>2 wk</td>
<td>0 wk</td>
</tr>
<tr>
<td>24-h TG AUC (mmol/liter per 24 h)*</td>
<td>5.6 ± 1.1</td>
<td>3.6 ± 1.3b</td>
<td>2.9 ± 1.5</td>
</tr>
<tr>
<td>Late-evening TG (mmol/liter)*</td>
<td>1.3 ± 0.2</td>
<td>1.5 ± 0.2h</td>
<td>1.2 ± 0.1</td>
</tr>
<tr>
<td>Fasting TG (mmol/liter)*</td>
<td>1.2 ± 0.1</td>
<td>1.4 ± 0.2*</td>
<td>1.2 ± 0.1</td>
</tr>
<tr>
<td>Fasting LDL-C (mmol/liter)*</td>
<td>2.6 ± 0.2</td>
<td>2.6 ± 0.2h</td>
<td>2.1 ± 0.2</td>
</tr>
<tr>
<td>Fasting non-HDL-C (mmol/liter)*</td>
<td>3.2 ± 0.2</td>
<td>3.3 ± 0.2h</td>
<td>2.7 ± 0.2</td>
</tr>
<tr>
<td>Fasting apoB (g/liter)*</td>
<td>0.82 ± 0.06</td>
<td>0.83 ± 0.06b</td>
<td>0.65 ± 0.04</td>
</tr>
<tr>
<td>apoB to apoAI ratio*</td>
<td>0.70 ± 0.06</td>
<td>0.70 ± 0.05b</td>
<td>0.54 ± 0.04</td>
</tr>
</tbody>
</table>

P > 0.05 for differences among groups at baseline for all outcomes. Mean ± SEM.

* PROC MIXED two-factor (sugar, gender) analysis with adjustment for BMI, ΔBW (2 wk to 0 wk), and outcome on absolute Δ (2 wk vs. 0 wk).

Δ (2 wk vs. 0 wk) significantly different from *Δ (2 wk vs. 0 wk), Tukey’s multiple comparison test.

PROC MIXED two-factor (sugar, gender) analysis with adjustment for BMI on absolute Δ (2 wk vs. 0 wk).

Δ (2 wk vs. 0 wk) significantly different from *Δ (2 wk vs. 0 wk), Tukey’s multiple comparison test.

* P < 0.01, ** P < 0.05, *** P < 0.0001, **** P < 0.001, LS means of Δ different from zero.

Similarly, fasting non-HDL-C (Fig. 3B), apoB (Fig. 3C), and the apoB to apoAI ratio (Fig. 3D) were all significantly increased in subjects consuming fructose (non-HDL-C: +0.29 ± 0.066 mmol/liter, P = 0.0081; apoB: +0.093 ± 0.022 g/liter, P = 0.0005; apoB to apoAI ratio: +14.6 ± 3.8%, P = 0.0006) and HFCS (non-HDL-C: +0.55 ± 0.14 mmol/liter, P < 0.0001; apoB: +0.12 ± 0.031 g/liter, P < 0.0001; apoB to apoAI ratio: +19.5 ± 4.4%, P < 0.0001) compared with baseline but not in subjects consuming glucose (non-HDL-C: +0.055 ± 0.080 mmol/liter, P = 0.49; apoB: +0.0097 ± 0.019 g/liter, P = 0.90; apoB to apoAI ratio: +1.9 ± 2.5%, P = 0.81).

Primary outcomes: comparing glucose, fructose, and HFCS consumption

The effects of the three sugars were significantly different (PROC MIXED two factor analysis with adjustment for BMI, ΔBW and outcome) for all primary outcomes except fasting TG (see effects of sugar P values in Table 2). The effects of HFCS compared with fructose consumption on all primary outcomes were not significantly different (P > 0.05, Tukey’s). The increases in 24-h TG AUC (P = 0.0068), late evening TG peaks (P = 0.015), fasting apoB (P = 0.037), and the apoB to apoAI ratio (P = 0.028) were larger after fructose consumption compared with glucose consumption. The increases in 24-h TG AUC (P = 0.034), fasting LDL (P = 0.0083), non-HDL-C (P = 0.0055), apoB (P = 0.0056), and apoB to apoAI ratio (P = 0.0034) were larger after HFCS consumption than glucose consumption.

With regard to BMI, although the statistical results presented in Table 1 and Figs. 2 and 3 include adjustment for BMI, Online Supplemental Fig. 1, A–F, presents the changes of the primary outcomes with subjects grouped as normal weight (BMI <25 kg/m²) or overweight/obese (BMI ≥25 kg/m²). The effect of BMI status was significant for the change of the 24-h TG AUC (P = 0.016) and the late-evening TG peaks (P = 0.019) but not for the fasting TG (P = 0.55, data not shown), LDL-C (P = 0.30), non-HDL-C (P = 0.93), apoB (P = 0.62), and apoB to apoAI ratio (P = 0.51). Normal weight and overweight/obese subjects consuming HFCS had comparable absolute (Supplemental Fig. 1) and percent increases of late-evening TG (BMI <25 kg/m²: +46 ± 11%; BMI ≥25 kg/m²: +31 ± 6%), fasting LDL-C (BMI <25 kg/m²: +22 ± 1%; BMI ≥25 kg/m²: +28 ± 1%), non-HDL-C (BMI <25 kg/m²: +36 ± 19%; BMI ≥25 kg/m²: +17 ± 7), apoB (BMI <25 kg/m²: +17 ± 6%; BMI ≥25 kg/m²: +20 ± 8%), and the apoB to apoAI ratio (BMI <25 kg/m²: +22 ± 7%; BMI ≥25 kg/m²: +20 ± 9%).

Secondary outcomes: comparing glucose, fructose, and HFCS with complex carbohydrate consumption

Table 3 presents the secondary outcomes during consumption of complex carbohydrate at baseline and at the end of the 2-wk sugar interventions. Fasting HDL con-
centrations were unaffected by consumption of the three sugar-sweetened beverages. Similar to the responses in the fasting state, subjects consuming fructose and HFCS had increased postprandial concentrations of LDL-C, non-HDL-C, and apoB compared with baseline, whereas subjects consuming glucose did not. Fructose and HFCS consumption increased postprandial concentrations of RLP-C and RLP-TG compared with baseline, whereas consumption of glucose did not. Consumption of all three sugars increased fasting and postprandial sdLDL-C compared with baseline.

**Secondary outcomes: comparing glucose, fructose, and HFCS consumption**

The effects of the three sugars were significantly different (PROC MIXED two factor analysis with adjustment for BMI, ΔBW, andoutcomeb) for postprandial LDL-C, non-HDL-C, apoB, RLP-C, and sdLDL-C (see effects of sugar P values in Table 3). The effects of HFCS compared with fructose consumption on all secondary outcomes were not significantly different (P > 0.05, Tukey’s). The increases in postprandial RLP-C were larger during consumption of fructose compared with glucose (P = 0.044), and HFCS consumption caused larger increases in postprandial LDL (P = 0.0024), non-HDL-C (P = 0.0007), apoB (P = 0.025), and sdLDL-C (P = 0.014) (Tukey’s) than glucose consumption.

For glucose, insulin, and HOMA-IR, the 24-h glucose and insulin profiles during baseline (0 wk) and at the end of the 2-wk intervention are presented in Online Supplemental Figs. 2, A–C, and 3, A–C, respectively. Compared with baseline, the 24-h glucose and insulin 24-h AUC and the postmeal insulin peaks were significantly increased in subjects consuming glucose, significantly decreased in subjects consuming fructose, and were unchanged in subjects consuming HFCS (Online Supplemental Table 1). Postmeal glucose peaks were increased in subjects consuming glucose and HFCS. Fasting glucose concentrations were significantly decreased in subjects consuming glucose, whereas fasting insulin concentrations and HOMA-IR did not change significantly in any group.

**Gender**

Although there were no significant sugar-gender interactions for any of the primary or secondary outcomes, men
exhibited larger increases of fasting TG, non-HDL-C, apoB, and sdLDL-C concentrations and postprandial LDL, non-HDL-C, and sdLDL-C concentrations in response to sugar consumption than women (see effects of gender $P$ values in Tables 2 and 3). However, postprandial TG responses, as assessed by the 24-h TG AUC, late-evening TG peaks, postprandial apoB, and RLP-TG concentrations, were not different between genders. The subjects consuming glucose exhibited the most divergent gender responses, particularly in sdLDL-C. Fasting and postprandial sdLDL-C levels were increased compared with baseline by $+0.22 \pm 0.07$ mmol/liter ($P = 0.0001$) and
+0.24 ± 0.05 mmol/liter \( (P < 0.0001) \), respectively, in men after glucose consumption but were unchanged in women (fasting sdLDL-C: −0.004 ± 0.02 mmol/liter, \( P = 0.61 \); postprandial sdLDL-C: +0.006 ± 0.019 mmol/liter, \( P = 0.69 \)).

**Discussion**

The current study provides evidence that postprandial TG and fasting and postprandial concentrations of LDL, non-HDL-C, apoB, and the apoB to apoA1 ratio, established risk factors for coronary heart disease (25), are significantly increased in response to 2 wk consumption of 25% of energy requirements as glucose-, fructose-, or HFCS-sweetened beverages for 2 wk. \( P < 0.01 \), effect of sugar; two-factor (sugar, gender) PROC MIXED analysis on \( \Delta \) with adjustment for BMI, \( \Delta BW \) (D), and outcome at baseline (A–C). **, \( P < 0.01 \), ***, \( P < 0.001 \), ****, \( P < 0.0001 \), LS means different from zero. A, \( \Delta \) different from B, \( \Delta \), Tukey’s \( (n = 16/group) \). Data are mean ± SEM.

Discussion

The current study provides evidence that postprandial TG and fasting and postprandial concentrations of LDL, non-HDL-C, apoB, and the apoB to apoA1 ratio, established risk factors for coronary heart disease (25), are significantly increased in response to 2 wk consumption of 25% of E as fructose and HFCS, but not glucose, in younger, normal-weight, and overweight subjects. In contrast and as was observed in older subjects (15), fasting TG concentrations were increased in subjects consuming glucose but not in those consuming fructose-containing sugars. The differential effects of fructose and glucose consumption on fasting and postprandial TG responses in subjects from both studies suggest that fasting TG concentrations are not a reliable indicator of the adverse changes in postprandial TG and other lipid/lipoprotein risk factors induced by fructose consumption. There is growing evidence linking increases of postprandial TG concentrations with proatherogenic conditions (26–28). It is important to note that for both the current and previous study (15), the differential effects of fructose and HFCS compared with complex carbohydrate on the 24-h TG profile were most marked in the late evening, approximately 4 and 6 h after dinner. Studies investigating the relationship between this late-evening peak and proatherogenic changes would be of interest, as would investigations into the sources of the TG that contributes to these peaks (de novo lipogenesis, diet, or fatty acids derived from adipose lipolysis).

To our knowledge this is the first study to directly compare the effects of sustained consumption of HFCS with 100% fructose and glucose-sweetened beverages. This comparison is important because it would seem likely that the effects of HFCS-sweetened beverages on circulating lipids and lipoproteins would be less than those of pure fructose-sweetened beverage because they contain 45%
less fructose. And indeed, the postprandial TG and RLP responses exhibited the expected pattern based on the fructose content of the sugars, with increases being greatest in subjects who consumed 145 ± 4 g fructose per day from beverages, lowest in subjects who consumed 144 ± 5 g glucose per day and 0 g fructose per day from beverages and intermediate in subjects who consumed HFCS-sweetened beverages providing 64 ± 2 g glucose per day and 79 ± 3 g fructose per day. However, the changes of fasting and postprandial concentrations of LDL, non-HDL-C, apoB, and the apoB to apoAI ratio in subjects consuming HFCS were significantly larger compared with subjects consuming glucose and tended to be higher compared with subjects consuming pure fructose. More studies are needed to confirm this unexpected pattern and to determine whether it is a result of a synergistic effect of consuming fructose and glucose in combination. Additional studies are also needed to determine whether the substantial increases, seen after just 2 wk, are further aggravated with longer-term consumption of HFCS-sweetened beverages.

Compared with baseline, postmeal glucose and insulin responses (indexed as 24 h AUC and postmeal peaks) were mainly increased during glucose consumption, decreased during fructose consumption, and unchanged during HFCS consumption. This pattern is expected and further supports our data indicating that the adverse effects associated with chronic consumption of sugar-sweetened beverages result from the specific effects of fructose (29), rather than from increased circulating glucose and insulin excursions (i.e., glycemic index) (30–32). Although consumption of fructose increased fasting glucose and insulin concentrations in 2 wk and decreased insulin sensitivity by 17% in 10 wk (15), in the current study, HOMA-IR was unchanged after 2 wk consumption of fructose, HFCS, or glucose. This may be related to the subjects in the current study being younger and leaner (28 ± 7 yr; 25.5 ± 4.0 kg/m²) than the subjects in the previous study (54 ± 8 yr; 29.1 ± 2.9 kg/m²). In a study by Le et al. (33), inclusion of fructose with an energy-balanced diet for 4 wk in young, normal-weight men (24.7 ± 1.3 yr; ∼22 kg/m²) increased fasting glucose levels, but other indices of insulin sensitivity were unaffected. However, it was recently reported that consumption of fructose or glucose (150 g/d) for 4 wk lowered insulin sensitivity and increased HOMA-IR in subjects of similar age and BMI (31 ± 9 yr; 25.9 ± 2.2 kg/m²) (34).

As would be expected based on the evidence that both increasing age and postmenopausal status result in augmented postprandial lipid responses in women (35), more significant gender differences in lipid outcomes were observed in these younger subjects in the current study than in the older subjects previously studied (15). With the exception of postprandial TG, apoB, and RLP-C and RLP-TG, younger men exhibited larger lipoprotein responses after 2 wk of sugar consumption than younger women. The comparable responses in postprandial TG and apoB concentrations and the significantly different fasting TG and apoB responses between the genders suggest that rates of very low-density lipoprotein secretion may be similar between men and women, whereas rates of very low-density lipoprotein clearance are different. This is supported by kinetic studies, which demonstrate that women have higher TG-rich lipoprotein and LDL-apoB fractional catabolic rates than men, whereas production rates are comparable (36, 37).

The greater effect of glucose consumption on sdLDL-C levels in younger men compared with younger women represents the most marked difference between the current and our previous lipid results, which showed older men and women were comparably nonresponsive to consumption of glucose (15). The increase of fasting sdLDL-C concentrations compared with baseline in younger men consuming glucose was unexpected because they did not exhibit increases in fasting LDL and apoB concentrations.

The added sugar component of the typical U.S. diet consists of nearly equal amounts of HFCS and sucrose (38); therefore, it is a limitation of this study that we did not also investigate the effects of sucrose consumption. However, we expect that the effects of sucrose would be comparable with those of HFCS because its composition (50% glucose/50% fructose) is very similar to the composition of the HFCS used for this study (45% glucose/55% fructose). This is supported by results from a cross-over study in which subjects consumed standardized diets containing 5, 18, or 33% of energy as sucrose, each for 6 wk. Compared with the 5% sucrose diet, LDL concentrations increased by 17% on the 18% sucrose diet and by 22% on the 33% sucrose diet (39).

Self-reported intake data suggest that 13% of the U.S. population consumes 25% or more of energy from added sugar (40). Importantly, the current results provide evidence that sugar consumption at this level increases risk factors for cardiovascular disease within 2 wk in young adults, thus providing direct experimental support for the epidemiological evidence linking sugar consumption with dyslipidemia (1–5) and cardiovascular disease (13). The results contradict the conclusions from recent reviews that sugar intakes as high as 25–50% of energy have no adverse long-term effects with respect to components of the metabolic syndrome (16) and that fructose consumption up to 140 g/d does not result in a biologically relevant increase of fasting or postprandial TG in healthy, normal-weight (17) or overweight or obese (18) humans. Additionally
they provide evidence that the maximal upper limit of 25% of total energy requirements from added sugar, suggested by the Dietary Guidelines for Americans 2010 (19), may need to be reevaluated.

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