# **UCSF**

# **UC San Francisco Previously Published Works**

### **Title**

Maternal immunisation: collaborating with mother nature

### **Permalink**

https://escholarship.org/uc/item/992025x5

## Journal

The Lancet Infectious Diseases, 17(7)

### **ISSN**

1473-3099

#### **Authors**

Marchant, Arnaud Sadarangani, Manish Garand, Mathieu et al.

### **Publication Date**

2017-07-01

#### DOI

10.1016/s1473-3099(17)30229-3

Peer reviewed

# Series

THELANCETID-D-16-00250 [PII\_REPLACE]

Embargo: [add date when known]

[A: We have edited your paper to avoid repetition, enhance readability, reduce length, and achieve consistency with Lancet style]

A

This version saved: 15:55, 27-Feb-17

# Maternal immunisation 1



# Maternal immunisation: a worldwide landscape analysis

# [A: title edited to be more descriptive and explanatory]

Arnaud Marchant\*, Manish Sadarangani\*, Mathieu Garand\*, Nicolas Dauby, Valerie Verhasselt, Lenore Pereira, Gordean Bjornson, Christine E Jones, Scott A Halperin, Kathryn M Edwards, Paul Heath, Peter J Openshaw, David W Scheifele†, Tobias R Kollmann†

Maternal immunisation has the potential to substantially reduce morbidity and mortality from infectious diseases after birth. The success of tetanus, influenza, and pertussis immunisation during pregnancy has led to consideration of additional maternal immunisation strategies to prevent group B streptococcus [A: We avoid using abbreviations when possible, particularly for disease and drug names] and respiratory syncytial virus infections, among others. However, many gaps in knowledge regarding the immunobiology of maternal immunisation prevent the optimal design and application of this successful public health intervention. Therefore, we did an innovative landscape analysis to identify research priorities. Key topics were delineated through review of the published literature, consultation with vaccine developers and regulatory agencies, and a collaborative workshop that gathered experts across several maternal immunisation initiatives—group B streptococcus, respiratory syncytial virus, pertussis, and influenza. Finally, a global online survey prioritised the identified knowledge gaps on the basis of expert opinion about their importance and relevance. Here we present the results of this worldwide landscape analysis and discuss the identified research gaps.

#### Introduction

Failure to improve survival in neonates by 2035 could lead to an estimated 116 million preventable stillbirths or neonatal deaths, 99 million survivors with disability, and millions more with a lifelong increased risk for non-communicable diseases. The underlying causes for the 2.6 million stillbirths per year are largely unknown, but roughly 20% of the 2.9 million annual neonatal deaths are thought to be due to infection. The transfer of antibodies from pregnant women to their offspring is profoundly important for the health and survival of neonates and young infants, particularly because it reduces the risk of severe infections. Unfortunately, not all pregnant women have protective concentrations of antibodies against pathogens that affect their offspring.

The strategy of maternal immunisation to enhance protection of young infants is rapidly gaining support from both the public and health professionals.² Factors contributing to this momentum include the global reduction in neonatal tetanus as a result of maternal immunisation, the benefits of seasonal and pandemic influenza immunisation for both mother and infant, and the positive effect of maternal immunisation on pertussis outbreaks. These factors are also stimulating commercial development of new vaccines against additional threats, such as group B streptococcus and respiratory syncytial virus.

In recognition of the need to enhance the science of maternal immunisation, the Bill & Melinda Gates Foundation commissioned the authors of this Series paper to do a landscape analysis of the immunobiology that underpins successful vaccination during pregnancy. The scope of the analysis included all relevant immunobiological issues in general terms and as applied

to immunisation against pertussis, influenza, group B streptococcus, and respiratory syncytial virus specifically. We aimed to identify differences that might exist between pregnant women in low-income and middle-income countries (LMICs) and those in high-income countries that could affect the success of maternal immunisation programmes. We used an innovative approach to identify and prioritise the current knowledge gaps to inform future studies.

Here we describe the methods and the results of this effort and discuss the identified research gaps in immunobiology of maternal immunisation that can be generalised across pathogens. The two companion papers in this Series [Editor: Add references for the following two papers in this Series here when details are known] discuss research gaps specific to individual pathogens. Other crucially important aspects of maternal immunisation, including safety, public perception, and into existing global immunisation programmes, are outside the scope of this Series, but are discussed in another publication that summarises the outcome of a series of meetings sponsored by the National Institutes of Health.3

# Landscape review process and prioritization of knowledge gaps

We used an innovative multistage review process to best capture the state of knowledge about maternal immunisation. The appendix provides a detailed description of the methods used and the results of the analysis. Briefly, an international team of ten recognised experts did a scoping review of the English literature published since 2000 [A: Please provide month and date here (eg. Jan 1?)]. The experts summarised the state of

#### Lancet Infect Dis 2017

This is the first in a **Series** of three papers about maternal immunisation

\*Joint first authors

†Joint last authors

Institute for Medical Immunology, Université Libre de Bruxelles, Brussels, Belgium (A Marchant XX, N Dauby XX); Oxford Vaccine Group, Department of Paediatrics. University of Oxford, Oxford, UK (M Sadarangani XX): Division of Infectious Diseases, Department of Pediatrics (M Sadarangani, DW Scheifele XX, T R Kollmann XX) and Vaccine **Evaluation Centre** (M Sadarangani, M Garand XX, G Bjornson XX, D W Scheifele, TR Kollmann) University of British Columbia and BC Children's Hospital, Vancouver, BC. Canada: Vaccine and Immunity Theme, Medical Research Council Unit, Fajara, The Gambia (M Garand); Department of Infectious Diseases, Centre Hospitalier Universitaire Saint-Pierre. Brussels, Belgium (N Dauby); EA 6302 Immune Tolerance Team, University Nice Sophia Antipolis, Nice, France (V Verhasselt); [A: Please provide a department name if possible] University of California, San Francisco, CA. USA (L Pereira XX); Paediatric Infectious Diseases Research Group (C E Jones XX) and St George's Vaccine Institute (P Heath XX), Institute of Infection and Immunity, St George's, University of London, London, UK; Canadian Center for Vaccinology, Dalhousie University, IWK [A: Can IWK abbreviation be defined?] Health Centre, and Nova Scotia Health Authority. Halifax, NS, Canada (S A Halperin XX): Vanderbilt Vaccine Research Program, Department of Pediatrics,

1

Vanderbilt University School of Medicine, Nashville, TN, USA (K M Edwards XX); and Respiratory Medicine, National Heart and Lung Institute, Imperial College London, London, UK (P J Openshaw XX)

[A: Please check all author names and affiliations are correct, please provide one degree per author (XX), and indicate whether any authors are full professors]

Correspondence to:

[A: Please provide a title for
Arnaud Marchant eg, Mr/Dr/
Prof] Arnaud Marchant, Institute
for Medical Immunology,
Université Libre de Bruxelles,
Brussels 6041, Belgium
arnaud.marchant@ulb.ac.be

[A: As a rule, we include only one corresponding author. Please amend if you wish]

# Panel: Top 20 knowledge gaps and Likert scores identified by the online survey

#### Immunisation during pregnancy

- Effect of vaccine antigen type on maternal responses (Likert score 4-1)
- Effect of health conditions on maternal immune responses (Likert score 4-2)

### Transplacental transfer of antibodies

- Effect of timing of vaccination during pregnancy on net transfer (Likert score 4-4)
- Effect of antigen type on maternal responses and transferability (Likert score 4-1)
- Effect of complications during pregnancy on antibody transfer (Likert score 4.0)

#### Protection of fetus and newborn infant

- Effect of maternal immunisation regimen on cord titres (Likert score 4·3)
- Effect of maternal immunisation regimen on infant responses (Likert score 4·3)
- Clinical relevance of interference with active immunisation (Likert score 4-3)
- Effect of maternal antibodies on effector and memory B-cell responses of infants (Likert score 4-0)
- Modulation of breastmilk immune components by immunisation (Likert score 4-2)

#### Pertussis vaccination

- Correlates of protection against colonisation, disease, and death (Likert score 4·4)
- Requirement for multiple pertussis antigens, role of pertussis toxin (Likert score 4·2)
- Reactogenicity of repeated doses of tetanus, diphtheria, acellular pertussis vaccine in sequential pregnancies (Likert score 4·0)

#### Group B streptococcal vaccine

- Correlates of protection against colonisation, disease, outcomes (Likert score 4-5)
- Serotype specific immunogenicity, transfer, and protection (Likert score 4·3)
- Effect of serotype on correlates of protection (Likert score 4-0)
- Effect of carrier proteins on responses of infants to vaccination (Likert score 4-0)

#### Respiratory syncytial virus vaccine

- Correlates of protection against infant disease and death (Likert score 4·6)
- Protection against lower respiratory infection and disease (Likert score 4-6)
- Impact of pre-existing immunity on maternal responses (Likert score 4-0)

Likert scores were assigned by use of a 5 point scale. A score of 4 indicates high importance and a score of 5 (the maximum score) indicates very high importance.

1 knowledge pertaining to their assigned area, including assessments of the gaps in understanding about the biology of the immunisation process. The team met at a collaborative workshop in Vancouver (BC, Canada) to 5 share their assessments with 26 additional international experts who commented critically on the presentations. More than 100 knowledge gaps were identified through this process, attesting to the underdevelopment of the underlying science of maternal immunisation. To ensure 10 that deliberation was sufficiently broad and issues affecting translation were addressed, further consultations were held with leaders of maternal vaccine development programmes at three major vaccine companies and with representatives of two major regulatory agencies (the US 15 Food and Drug Administration and the European Medicines Agency) who freely shared their insights into the knowledge gaps and challenges.

To prioritise the identified knowledge gaps, topics deemed most relevant during the collaborative workshop 20 were included in an online survey completed by nearly 200 [A: Can the exact number of experts be provided?] content experts from the global maternal immunisation community. Respondents rated the importance of each knowledge gap; the results were consistent among 25 respondents, including industry representatives, academic researchers, and national immunisation policy makers. The panel [A: Table I has been converted to a Panel, in line with our house style shows the top 20 knowledge gaps; each gap was rated as 4 or more on the 5 point Likert scale 30 (high to very high importance). To prepare this Series, we integrated and summarised the information gathered from each step of the multistage review process [A: correct as edited?].

# 35 General considerations regarding maternal immunisation strategies

When considering the four disease targets for maternal immunisation included in the landscape analysis (pertussis, influenza, group B streptococcus, and 40 respiratory syncytial virus), it is striking that no two vaccine programmes are alike (table), and that different strategies are likely to be needed for each disease, which could make the production of a combined vaccine challenging. To focus on the immunobiology of maternal 45 immunisation, contextual differences such as maternal disease risk, infant disease burden, global epidemiology, and microbial diversity will not be discussed further in this paper.

The common goal among maternal vaccination programmes is temporary protection of the young infant against severe illness and death by ensuring sufficient and timely transfer of protective antibodies from the mother. This passive protection should persist until the infant is no longer at high risk of disease (eg, 3 months of age for group B streptococcus disease) or until protection can be achieved by active infant immunisation (eg, pertussis). Protection of the infant might also be achieved indirectly

by reducing carriage or disease in the mother, which 1 subsequently reduces transmission of pathogens to the infant (eg, group B streptococcus, pertussis). Whether or not protection of the mother against disease is also required is another important factor in determining the 5 timing of maternal immunisation. For example, in the case of influenza immunisation early during pregnancy might be the favoured strategy to protect both the pregnant woman and neonate. Additionally, immunisation before pregnancy might have the benefit of preventing infections 10 that could have harmful effects on a developing fetus. However, understanding of optimal maternal immunisation for any target is limited by the scarcity of defined correlates of protection for young infants. Without a validated measure of protection, it will be difficult to 15 compare results of studies in different settings or to improve vaccines or immunisation regimens by use of serological criteria.

Immunisation during pregnancy relies on the capacity of the pregnant woman to mount appropriate primary or 20 secondary antibody responses, depending on whether the pathogen has been encountered before pregnancy. The notion that pregnancy is associated with the induction of various immunoregulatory mechanisms that are essential for the survival of the fetus suggests 25 that antibody responses to vaccines might be different in pregnant women compared with non-pregnant women. Vaccine responses might be further influenced by complications affecting pregnant women, such as chronic infections. Optimal protection of the young 30 the potential effect on antibody responses to primary infant is considered to rely on the effective transfer of maternal immunity through the placenta and the persistence of this passive immunity for the duration of infant exposure to the particular pathogen. Additional protection might be provided by transfer of immunity via 35 including transitional or marginal zone B cells, remains breastmilk. However, the relative contributions of breastmilk and serum antibodies to infant protection will be difficult to define, but are important to understand, especially for infants born prematurely with restricted transplacental transfer of antibodies. These passively 40 reduced capacity to produce immunoglobulins, their transferred maternal immune factors can further influence active immunity induced in the infant by natural infection or immunisation. Experts at the collaborative workshop identified 68 knowledge gaps related to the effect of pregnancy on vaccine responses, 45 the transfer of maternal immunity to the infant, and infant immunity (appendix). The panel presents the top ten of these knowledge gaps deemed most relevant in the online survey.

#### Effect of pregnancy on vaccine responses Pregnancy and B lymphocytes

Studies indicate that pregnancy influences B cells and antigen-presenting cells; no studies have assessed the potential effect on follicular helper T cells.

Oestrogen and pregnancy reduce B-cell lymphopoiesis in mice.4 Reductions in the number of circulating B cells

	Pertussis	Influenza	Group B streptococcus	Respiratory syncytial virus
Maternal disease risk	+	+++	++	+
Infant mortality	++	+	+++	++
Infant disease frequency	+ (cyclic*)	++	+	+++
Disease seasonality	✓	✓	×	✓
Microbial diversity	+	++	++	+
Licensed vaccine available	✓	✓	×	×
Maternal booster response expected†	✓	Quasi [A1]‡	Not assumed	✓
Passive protection of infant	✓	✓	✓	✓
Maternal to cord antibody ratio	1.1-1.9	0.7–1.0	0.7-0.8	1.0
Antibody half-life (days)	36-40	40-50	30-44	36-79
Infant vaccination	✓	<sup>3</sup> 6 months	×	(✓)§
Correlate of protection	×	Quasi [A1]¶	×	×
Functional immunoassay	×	✓	?	✓
Competing control option	×	×	<b>√</b> **	<b>√</b> ††

 $^* Increased \ disease \ incidence \ usually \ occurs \ every \ 3-4 \ years. \ † Via \ previous \ vaccination \ or \ infection. \ \ddagger Previous \ vaccination \ or \ infection. \ \ddagger Previous \ vaccination \ or \ infection.$ vaccination or infection will lead to partial protection due to virus evolution. §Monoclonal antibody administered to high-risk infants during respiratory syncytial virus season. ¶Correlates of protection based on haemagglutinin inhibition assay or microneutralisation titres have not been validated in young infants and are not based on maternal immunisation. ||Bacterial killing in an opsonophagocytic assay has been suggested as a possible correlate of  $protection.\ ^{**} Intrapartum\ antibiotic\ prophylax is\ has\ reduced\ the\ incidence\ of\ early\ onset\ group\ B\ streptococcus$ neonatal sepsis. ††Monoclonal antibodies administered to high risk infants during respiratory syncytial virus season reduces rates of hospital admission. [A1: Please explain what is meant by Quasi in this context] [A: Please describe in the legend what the symbols +, ++, +++,? in the table mean for clarity]

Table: Targets of maternal immunisation

have likewise have been shown in pregnant women, but immunisation is unknown.<sup>5-7</sup> Some studies<sup>8-10</sup> have shown an effect of pregnancy on memory B-cell subsets, but no consistent evidence has yet emerged. Additionally, the potential effect of pregnancy on other B-cell subsets, to be assessed. In populations living in LMICs, chronic exposure to microbial antigens, such as Plasmodium falciparum, induces high numbers of circulating atypical memory B cells.89 Because these memory cells have a increased numbers could hamper the immune response on subsequent challenge [A: Is this suitable instead of 'recall immunisation'?] in both pregnant and nonpregnant women living in LMICs.

#### Pregnancy and immunoglobulins

Studies assessing the influence of hormones on B-cell See Online for appendix functions support the notion that pregnancy might affect the production of immunoglobulins. Oestrogen increases 50 the production of IgG by human B cells. Additionally, activated human B cells upregulate the expression of the prolactin receptor, and prolactin further decreases the threshold of B-cell activation.12 In mice, oestrogen also upregulates the expression of the activation-induced 55 deaminase—the enzyme that initiates somatic hypermutation and class switch recombination of immunoglobulins.13 By contrast, serum IgG concentrations are

LMICs and high-income countries. 14,15 The mechanism involved is unclear, but could, at least partly, be due to haemodilution. Pregnancy is also associated with modifications in IgG glycosylation.16

IgG are glycoproteins that carry N-glycans at both the Fc and Fab segments, which modulate their effector functions.17 In pregnancy, IgG antibodies have increased sialylation and decreased N-acetylglucosamine bisection of both Fc and Fab fragments, and increased galactosylation 10 pertussis vaccine (Tdap) immunisation in pregnant and of Fc fragments.<sup>16</sup> Although the functional consequences of Fab fragment glycosylation remain unclear, sialylation and galactosylation of Fc fragments have been associated with decreased inflammation and were suggested to be involved in the remission of rheumatoid arthritis 15 streptococcus vaccine was studied in South Africa.39 associated with pregnancy.18,19 The potential implications of the anti-inflammatory properties of maternal IgG on immune homoeostasis and antimicrobial defenses in the fetus and newborn baby have not been determined. Surprisingly, IgGs of different antigen specificity have 20 been extensively studied. Similar antibody responses to different glycosylation profiles and this profile is modified after recent antigen exposure.20 Moreover, IgG glycosylation patterns are different in populations living in high-income countries and LMICs.20 Studies are needed to establish the effect of pregnancy on the glycosylation 25 seasonal influenza vaccine were higher during the third and effector functions of vaccine-induced IgG.

#### Pregnancy and antigen-presenting cells

Pregnancy is associated with changes in the numbers and phenotypes of antigen-presenting cells. The number 30 results on the avidity of antibodies following pertussis of myeloid dendritic cells increases in the first trimester of pregnancy and decreases in the third trimester as pregnancy progresses to reach similar cell counts as in non-pregnant women.<sup>21,22</sup> By contrast, the number of plasmacytoid dendritic cells is reduced during the third 35 of immunisation and the requirement to repeat trimester of pregnancy.23 Myeloid dendritic cells and plasmacytoid dendritic cells were shown to express higher concentrations of toll-like receptors in pregnant women than in non-pregnant women.24 Several differences exist between antigen-presenting cells from 40 pregnant women.33 Pertussis immunisation has been women and men that are induced by sex hormones and could therefore be relevant to pregnancy.25 Modifications of antigen-presenting cells are likely to be important for successful pregnancy, but the potential effect on vaccine responses have not been determined.

## Pregnancy and vaccine response

The effect of pregnancy and sex hormones on B cells and antigen-presenting cells suggests a possible influence on antibody responses to vaccines. This potential is indirectly 50 trimester, suggesting cumulative transfer of antibodies. supported by the observation that the magnitude of antibody responses to many vaccines is often higher in women than men.25 However, most studies of pregnant women that showed potent vaccine immunogenicity women.26-29 A few controlled studies have been done, but only in small populations. Some studies reported similar

lower in pregnant than in non-pregnant women in both 1 responses to seasonal influenza vaccines in pregnant and non-pregnant women, whereas others detected differences in titres or seroconversion rates.<sup>30-34</sup> Factors responsible for the discrepancies between studies might include 5 differences in tested vaccines and participant characteristics. The results of two controlled studies [A: Please provide references for the two studies immediately after their mention] done in high-income countries showed similar antibody responses to tetanus, diphtheria, acellular non-pregnant women, whereas two other studies in LMICs reported no effect of pregnancy on the response to tetanus immunisation.35-38

> In 2016, the immunogenicity of a conjugated group B Although the responses were not compared between pregnant and non-pregnant women, the vaccine was immunogenic in both groups. Whether the gestational stage of pregnancy affects responses to vaccines has not seasonal and pandemic influenza vaccination were observed throughout pregnancy in two studies [A: Please provide references for the two studies immediately after their mention], whereas seroconversion rates with a trimester than during the first and second trimesters [A: comparator correct as added?] in one study. 27,31,40 The effect of pregnancy on the quality of antibody responses to vaccines remains largely uncharacterised. Conflicting immunisation during early pregnancy compared with late in pregnancy have been obtained in small-scale studies. 41,42

The persistence of antibodies after maternal immunisation will influence the optimum timing immunisation during consecutive pregnancies; however, little information about this topic is available. Antibody decay following immunisation with adjuvant pandemic influenza vaccine was similar in pregnant and nonrecommended during the second or early third trimester of pregnancy to achieve sufficiently high titres of antibodies close to delivery. A: To maintain style, the journal reference provided has been added to the 45 reference list—see x1, the references will be renumbered accordingly]. This recommendation was challenged by a 2016 study,43 which showed higher titres of cord-blood antibodies following pertussis immunisation during the second trimester of pregnancy than during the third

Innate immune responses after maternal immunisation have not been explored. One study [A: Please provide a reference for this study directly after first mention] reported similar plasma concentrations of inflammatory did not include a comparison group of non-pregnant 55 cytokines in pregnant and non-pregnant women following seasonal influenza immunisation. This result accords with the similar or even lower reactogenicity

observed in pregnant women following influenza 1 factor that restricts the transfer of maternal immunity immunisation.44,45

#### Influence of maternal factors on vaccine responses

the use of significant for instances of clinical or statistical significance. Is this the case here, or should 'substantial' 'relevant' eg, be used instead?] of maternal age, parity, socioeconomic status, or bodyweight on antibody response to vaccines during pregnancy.46-48 However, 10 parity was associated with reduced antibody responses to Haemophilus influenzae type b conjugate vaccine in The Gambia and with heightened responses to pertussis toxin in Belgium. 49,50 This finding could be particularly important in LMICs, where high-order multiparity is 15 more common than in high-income countries [A: comparator correct as added?]. Some studies suggested a small effect [A: correct as rephrased?] of nutrition on vaccine responses during pregnancy. 51,52

Whether obesity affects immune response to 2 vaccination in pregnancy is poorly understood because obese women (body-mass index >30 kg/m²) are typically excluded from clinical trials. Little information is available about the possible differences in vaccine immunogenicity between LMICs and high-income 25 are transported at different rates across the placenta, countries resulting from health conditions of the mother. One study<sup>35</sup> reported that *P falciparum* parasitaemia had no effect at the time of immunisation on antibody response to tetanus toxoid. However, HIV infection impairs responses to vaccines. In South Africa, pregnant 30 differences might be partly related to the differences in women with HIV have lower seroconversion rates after seasonal influenza vaccination than do uninfected pregnant women, but antibody half-life and vaccine efficacy are similar between the two groups.53,54 HIV infection was also associated with lower immunogenicity 35 of a glycoconjugate group B streptococcus vaccine in pregnant women in South Africa.55 The effect of helminth infection on vaccine responses during pregnancy has not been systematically analysed.<sup>56</sup> [A: Summary paragraphs have been deleted to increase flow and readability]

#### Transfer of maternal immunity through the placenta

#### IgG transfer and preterm birth

the placenta.<sup>57</sup> A 2015 study<sup>58</sup> indicated that other maternal immunoglobulins can be transported to the fetus when complexed with IgG. IgG antibodies are actively transported through the placenta by the neonatal Fc receptor (FcRn), and possibly by additional receptors 5 that have not yet been identified.59,60 The FcRn is expressed by syncytiotrophoblasts covering the surface of the chorionic villi, and transports IgG by transcytosis into the fetal circulation. Although the FcRn is expressed and functional in the placenta from the first trimester, 55 gammaglobulinaemia, but not with placental malaria most of the antibody transfer occurs after 28 weeks' gestation.61,62 Preterm birth is therefore an important

through the placenta and might affect the transport of IgG1 more than IgG2.63-66

Preterm birth occurs in 5-18% of pregnancies globally Most studies reported no significant effect [A: We reserve 5 and is a leading contributor to infant morbidity and mortality. In a 2012 systematic analysis,67 over 60% of all preterm births were estimated to occur in sub-Saharan Africa and south Asia (>9 million of roughly 15 million births per year globally). At 28-33 weeks' gestation, fetal-maternal antibody ratios are typically 0.5-0.6 compared with 1.0 or higher at full term. Thus, transfer of maternal antibody could afford some potential protection even in prematurely born babies if their antibody concentrations were elevated by previous immunisation.66

#### Factors influencing IgG transfer

The rate of IgG transfer through the placenta is influenced by several factors, including IgG subclass, antigen specificity, and chronic maternal infections. IgG subclasses are transcytosed at different rates, with IgG1 being most actively transferred, followed by IgG4, IgG3, and IgG2. 59,68,69 IgG3 allotypes have different affinity for FcRn and this results in differential transfer ratios. 69 It is puzzling that antibodies of different antigen specificities resulting in different maternal to cord-blood antibody ratios.70-72 Reported cord-blood to maternal ratios range from 1.9 for pertussis to 0.7 for group B streptococcus, with influenza ranging between 0.7 and 1.0.26,53,73-75 These IgG subclass proportions, as protein antigens generally induce IgG1 and IgG3 subclasses, whereas polysaccharide antigens induce mainly IgG2 antibodies, but this hypothesis has not been systematically examined. 57,72

Whether or not the structure of maternal IgG influences placental transfer beyond subclasses has not been clearly established. Two studies76,77 have suggested that high avidity antibodies can be transferred preferentially across the placenta. Previously, studies also suggested a 40 preferential transfer of hypergalactosylated IgG, but this theory was not supported by a more recent study that used more advanced technologies, which showed that Fc galactosylation had no effect on IgG transfer.78,79

Chronic maternal infections and hypergamma-IgG is the only antibody that is directly transferred across 45 globulinaemia have a profound effect on maternal antibody transfer.66 Reduced transfer of IgG is observed in women with hypergammaglobulinaemia, a condition that might be associated with the saturation of FcRn.80-82 Hypergammaglobulinaemia and the denudation of syncytiotrophoblasts from chorionic villi could also be involved in the reduced transfer of IgG associated with placental malaria. 66,81 A 2016 study83 in Papua New Guinea indicated an association between reduced transfer of respiratory syncytial virus-specific IgG and hyperitself. Maternal HIV infection also results in a reduction of maternal IgG transfer.82,84-86 Intriguingly, the effect

globulinaemia seems to depend on the subclass and antigen specificity of IgG. In a study85 in South Africa, maternal HIV infection was associated with reduced transfer of naturally acquired group B streptococcus- 5 specific IgG1, but not IgG2. In a study<sup>81</sup> in The Gambia, maternal hypergammaglobulinaemia was found to be associated with impaired transfer of total IgG1 and IgG2, but not IgG3 and IgG4, and with a reduced transfer of IgG against pathogens, but not vaccine antigens. [A: 10 Summary paragraph has been deleted to increase flow and readability.]

### Transfer of maternal immunity through breastfeeding

The importance of breastmilk in postnatal life is highlighted by the strong correlation between breastfeeding and the profound reduction in risks of infection and infection-associated mortality in infancy.87,88 However, only one study [A: Please reference this study at first mention assessed the role of breastfeeding in protection against an infectious pathogen after maternal immunisation. In Bangladesh, exclusive breastfeeding was associated with a decrease in the number of episodes of respiratory illness with fever in children 25 The stimulation of low-level inflammation [A: correct as born to mothers immunised against influenza during pregnancy.89 Prevention of infectious diseases by breastfeeding is thought to be due to the strengthening

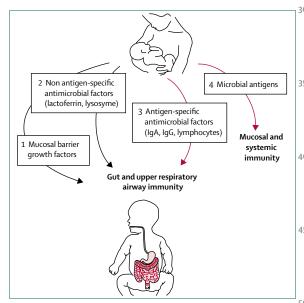


Figure 1: Transfer of maternal immunity through breastfeeding Microbe-non-specific immunity (blue) is promoted by breastmilk through growth factors that improve the function of the epithelial barrier (1) and antimicrobial molecules (2). Microbe-specific immunity (red) is provided by antigen-specific maternal IgA and IgG and lymphocytes (3). Breastmilk also contains antigens and attenuated microbes that can stimulate infant immunity (4). Maternal vaccination could improve prevention of infectious disease in breastfed children by increasing the concentration of antigen-specific antimicrobial factors and microbial antigens in breastmilk.

of chronic maternal infections and hypergamma- 1 of gastrointestinal and respiratory mucosal immunity via improvement of the function of the epithelial barrier through the high content of growth factors in breastmilk, and by transference of antimicrobial factors, such as lactoferrin and lysozyme, and microbial antigen-specific immunity (figure 1). Thus, maternal immunisation might modulate antigen-specific immune factors in breastmilk and promote antigen-specific immune responses in infants.

#### Breastmilk IgA

Breastmilk secretory IgA antibodies are specific for various common intestinal and respiratory pathogens as a result of the selective migration of B cells originating 15 from the mucosal membranes to the mammary gland.90 Therefore, concentrations of secretory IgA should be higher when induced by mucosal immunisation than by systemic immunisation, as observed following HIV immunisation of lactating Rhesus macagues.91 The o antimicrobial properties of secretory IgA depend on the inhibition of pathogen adherence to, and invasion of, mucosal epithelia, the neutralisation of pathogens and toxins, the transfer of antigens across the mucosal barrier, and the stimulation of low-level inflammation.92 edited?] has been mainly described in mice [A: Please provide a reference]. Some studies 90,93,94 in humans have demonstrated the transport of breastmilk IgA into the circulation of breastfed mature and premature newborn o babies. In LMICs, where prematurity and gut mucosal inflammation are common, IgA transport to neonatal circulation might be increased and prolonged and could therefore be particularly beneficial. By contrast, breastmilk IgA could have a negative effect on the 35 response to mucosal vaccines, but this finding remains controversial.95,96

Several studies97 showed increased concentrations of antigen-specific IgA in breastmilk following maternal immunisation against influenza, pertussis, respiratory 40 syncytial virus, Streptococcus pneumoniae, and Neisseria meningitidis. The amount of breastmilk and magnitude of secretory IgA responses against a consensus HIV envelope protein have been associated with the reduced risk of postnatal transmission of HIV in 45 Malawi [A: Please provide a reference]. This observation highlights the need for development of maternal vaccination strategies that increase the concentration of [A: ok as edited?] HIV-1 envelopespecific breastmilk IgA to reduce mother-to-child HIV 50 transmission.98 Importantly, maternal conditions that are known to negatively affect transplacental transfer of IgG do not affect IgA transfer through breastmilk. Prematurity increases the transfer of growth and immune factors, particularly IgA, in colostrum and 55 milk. 99,100 Furthermore, the concentration of total and pathogen-specific IgA in breastmilk is not affected by maternal HIV infection or by malnutrition. 101-04

#### Breastmilk IgG

Breastmilk IgG originates from serum via FcRn transport and from resident B lymphocytes. 105 The total IgG concentration in breastmilk is about 10% of the IgA concentration, but tends to increase with duration 5 of breastfeeding. 100,106,107 Increased concentrations of antigen-specific IgG are detected in breastmilk following immunisation against respiratory syncytial virus and pneumococcus, and following natural infection with Evidence of a protective role of breastmilk IgG was shown in studies of HIV infection, whereby IgG had higher neutralising activity than IgA, mediated antibody-dependent cellular cytotoxicity, and was inversely correlated with the risk of HIV transmission.<sup>109</sup> 15 and readability] Breastmilk IgG was also inversely correlated with human cytomegalovirus load, suggesting a protective role against human cytomegalovirus transmission.110 However, the role of breastmilk IgG in the defense against other pathogens has not been studied.

Experiments in mice suggest that breastmilk IgG can cross the gut barrier through FcRn and can thereby promote the transport of IgG-antigen immune complexes and stimulate immune response to antigens and pathogens. 60,111-14 Whether this process occurs in 25 above the protective concentrations in the infant is a human beings is unknown.

#### **Breastmilk leucocytes**

Breastmilk contains neutrophils, macrophages, and lymphocytes.115 Common infections increase the number 30 duration of protection. However, high concentrations of of total leucocytes in breastmilk, but whether similar changes occur after immunisation is unknown.116 Breastmilk B lymphocytes are IgG-producing memory cells. The antigen specificity of these lymphocytes was demonstrated in the context of HIV infection. <sup>105</sup> Similarly, 35 infant beyond passive protection. HIV-specific CD4 and CD8 T lymphocytes were detected in breastmilk and might contribute to virus control through inflammatory cytokines and cytotoxicity.117,118 Studies93,119,120 suggest that CD4 T cells in breastmilk transient specific cellular immunity.

#### Transfer of microbial antigens through breastmilk

Although pathogens can be detected in breastmilk after maternal infection, transmission to the offspring is not 45 the minimisation of invasive disease severity rather than commonly observed, with notable exceptions, including HIV, human cytomegalovirus, and human T-cell lymphotropic virus 1.<sup>121</sup> The evidence suggests that breastmilk immunity can prevent pathogen transmission. Additionally, studies 102,122 suggest that exposure 50 animal models. 127-129 The observed effectiveness of to pathogens through breastmilk induces immune responses in infants independently of transmission. Exposure to HIV-containing breastmilk is associated with the induction of mucosal IgG and IgA responses and with systemic cell-mediated immune responses in 55 could result in passive and active immunity [A: Correct uninfected infants. Similarly, Vibrio cholera can be transferred through breastmilk and induce either

1 disease or colonisation associated with specific IgG responses in infants. 123 These observations suggest that breastfeeding can promote immunity to pathogens in infants by transmitting pathogens that are attenuated by maternal immune responses or transfer of pathogen antigens.

Studies<sup>124</sup> indicate that a similar process occurs following immunisation of lactating women with the live attenuated rubella vaccine. Studies in mice<sup>125</sup> have group B streptococcus, rotavirus, and HIV.96,108,109 10 shown that the intrinsic adjuvant properties of antigens and the concentration of IgG and amount of vitamin A in breastmilk are crucial factors in the induction of effector immune responses in the offspring. [A: Summary paragraph has been deleted to increase flow

#### Maternal immunisation and infant immunity

Placental transfer of maternal antibodies is expected to protect the infant from disease. However, a specific 20 concentration of antibody (the presumed correlate of protection) has to be reached to provide clinical protection and this concentration needs to be maintained until the infant is no longer at risk, or is protected by active immunisation. How long maternal antibodies persist function of the concentration of the antibody in the newborn baby at birth and the antibody half-life  $(t_{10})$ . Thus, the transplacental transfer and decay kinetics of maternal IgG in the infant are key determinants of the maternal antibodies present at the time of infant vaccination might also interfere with the immune response of the infant to the respective vaccine. Maternal immunisation can have effects on the fetus and newborn

#### Prevention of infection and disease

The distribution of serum antibodies beyond the bloodstream of the neonate or infant is not well defined, might be transferred to human neonates and induce 40 but could restrict what is achievable in terms of mucosal protection. For example, little IgG is detectable in saliva of young infants until the teeth erupt, 126 making sterilising immunity against respiratory pathogens unlikely. A more readily achievable objective would be prevention of portal of entry infection and colonisation, as emphasised by the failure of various preparations of pertussis immunoglobulin to prevent colonisation (and subsequent invasive infection) in human beings and maternal pertussis immunisation in preventing infant disease represents an important advancement.<sup>130</sup> If the benefit of maternal immunisation is largely attributable to minimisation of disease severity such encounters as edited?], with active immunity following attenuated natural infection.131

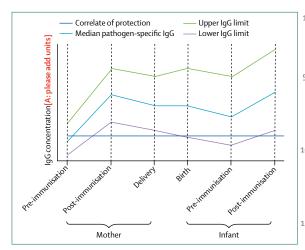


Figure 2: Influence of maternal immunisation on infant IgG before and after vaccination

[A: First two sentences have been removed, as this information can be shown in the figure itself using a key.] In the absence of maternal immunisation, maternal IgG concentrations are low and could be below the correlate of protection. An ideal vaccine would raise this IgG concentration such that even the lower end of the range would be above the correlate of protection, and would remain above the correlate of protection until delivery, which would depend on the initial response to vaccination and the timing between immunisation and delivery. The infant IgG concentration at birth will depend on placental health, gestation, and antibody-specific factors. The concentration of transferred maternal IgG will decrease until the infant receives additional protection via direct immunisation, and the rate of decrease will vary between pathogens and between individuals. Ideally, maternal vaccination would ensure the IgG concentration is above the correlate of protection until infant immunisation, which will depend on the initial IgG concentration at birth and the interval until infant immunisation, creating a so-called window of susceptibility when the IgG concentration falls below the correlate of protection. Following infant immunisation, the IgG concentration will rise again, and the extent of this would be influenced by any interference caused by the presence of maternal IgG.

#### Maternal antibody decay in infants

The  $t_{1/2}$  of IgG differs by subclass and is not a fixed entity, but is directly proportional to the total IgG concentration. This mechanism is called the concentration-catabolism effect, whereby IgG catabolism is accelerated in individuals with increased IgG concentrations and, conversely, reduced 40 haemophilus b conjugate [A: Is this the correct definition in individuals with a low serum IgG concentration. 132 The molecular mechanisms underlying the differences in t<sub>10</sub> of the various IgG subclasses and the concentrationcatabolism effect centre around FcRn. 59,60

Subclass and structural modifications of IgG have a 45 profound effect on the interaction with FcRn, and thus t<sub>1/2</sub>. For example, IgG3 allotypes have different affinity for the FcRn, which results in different t<sub>1/2</sub>.69 Furthermore, aglycosylated human IgG1 has a considerably shorter t<sub>1/2</sub> (62 h) than the glycosylated form (153 h).<sup>132</sup> 50 could be detected in 38·5% of cord-blood specimens, and Glycosylation of maternal antibodies is modified during pregnancy,16,133 but how this relates to t<sub>1/2</sub> in the infant is not known. Moreover, studies suggest that the t<sub>1/2</sub> of IgG in infants varies depending on the antigen specificity of the antibodies and between populations. For example, 55 detection of human influenza haemagglutinin-specific reported t<sub>1/2</sub> in the infant of maternal antibodies specific for pertussis antigens is roughly 30-40 days, for tetanus

1 roughly 50 days, but for group B streptococcus roughly 60 days. 29,134,135 The t<sub>1/2</sub> of maternal antibodies of a given specificity can also vary substantially between populations; however, whether this variability involves diff-5 erences in IgG subclass or other structural differences has not been delineated. 136-38

#### Interference with infant immunisation

The presence of maternal antibodies to a particular o vaccine antigen has been reported to reduce antibody generation following vaccination of the infant with the same antigen.<sup>139-41</sup> a process known as interference. Maternal antibodies not only affect concentrations of antibodies produced by the infant, but can also influence 15 their quality (strength of antigen binding or avidity). 141,142 Priming of T-cell responses to vaccines does not seem to be affected by passive antibodies and this probably contributes to the good response to booster doses. 139,140 The key factors influencing interference are antigen-20 specific maternal antibody titres at the time of infant immunisation, and the antigen content (including dose) of the infant vaccine schedule.

For pertussis, maternally-derived antibodies interfere with antibody responses to whole-cell vaccines in the 25 infant, but less so to acellular vaccines. 37,50,143-47 Whether the improved response to acellular versus whole-cell vaccine among infants with higher antecedent titres of pertussis toxin [A: Correct as defined?] is due to higher antigen load in the acellular product or to the absence of 30 other components of the whole-cell vaccine in the acellular product has not been determined.148 In view of the fact that the current lead candidates for a maternal group B streptococcus vaccine are tetanus toxoid or CRM197 (non-toxic mutant of diphtheria toxin) conjugate 35 polysaccharide vaccines, it is worth noting that infants born to mothers with high titres of anti-tetanus toxoid immunised with Haemophilius influenza type b vaccine conjugated with tetanus toxoid have reduced anti-group B streptococcus responses, but infants immunised with for HbOC?] (CRM197) had no interference. 149-51 Although several mechanisms have been proposed, the molecular and cellular basis of the interference remains incompletely understood. 139,140

#### Influence of maternal immunisation beyond passive immunity

Following influenza vaccination during pregnancy, antihuman influenza haemagglutinin [A: correct as added?] anti-matrix protein IgM antibodies could be detected 40.0%.152 Because IgM does not cross the placenta, this finding suggests an active adaptive B-cell response in the fetus. This hypothesis was further corroborated by the T-cell responses in some newborn babies of immunised women with synthetic peptide-human leucocyte antigen

multimers [A: Please provide a reference]. Similarly, earlier 1 and synthesis. AM, MS, VV, MG, DWS, and TRK drafted the initial studies153,154 of tetanus vaccination during pregnancy reported detection of anti-toxoid IgM in sera of some infants. Because vaccines can have immune modulatory effects in postnatal life beyond initiating antigen-specific 5 adaptive responses (ie, non-specific effects<sup>155</sup>) immunisation during pregnancy could also have non-specific effects not only in the mother, but also in the fetus or newborn baby. To our knowledge, this notion has not been systematically investigated. However, MF59-adjuvanted 1 influenza vaccination during pregnancy led to an altered cytokine production profile in the nasal mucosa of 4-weekold infants from vaccinated versus unvaccinated mothers. 156 The clinical relevance of these unexpected findings (active in-utero immune response and non-specific effects on 15 publication. the newborn baby after maternal immunisation) is unclear. [A: Summary paragraph has been deleted to increase flow and readability

#### Conclusion

The passive transfer of maternal immunity is considered central to antimicrobial defenses in early life (figure 2 [A: Please move this figure citation to earlier in the paper, rather that readers reaching the end of the paper before seeing this informative graph]). The proposed 25 mechanisms centre around active transport of maternal IgG through the placenta providing systemic immunity during the first months after birth until the infant actively acquires immunity through exposure to pathogens or vaccines. The immune components of 30 breastmilk can provide longer-term immunity at the mucosal level and could also contribute to the development of infant immunity at the systemic level.

Although maternal immunisation is an effective strategy to increase antimicrobial immunity in early life, 35 many knowledge gaps remain in the understanding of vaccine responses during pregnancy, the transfer and persistence of maternal immunity in infants, and the interactions between maternal antibodies and the infant immune system. In this landscape analysis, we 40 7 prioritised gaps of particular relevance to the development of new vaccines for pregnant women and to the implementation of maternal immunisation worldwide [A: Don't need to refer to the panel again in this section of concluding remarks]. Addressing these knowledge 45 9 gaps [A: edit OK?] offers the potential to further improve this important public health intervention, and will require immunological studies of existing vaccines administered to pregnant women and the inclusion of immunological endpoints in the clinical studies of 50  $_{11}$ vaccines under development.

AM, DWS, and TRK developed and managed the landscape analysis, and synthesised the information. AM, VV, LP, and TRK led the literature review on the immunobiology of maternal immunisation. MG and GB provided major administrative support and participated in  $^{55}$ the synthesis of the information. AM, MS, ND, VV, LP, CEJ, SAH, KME, PH, PJO, DWS, and TRK contributed to the literature review

manuscript and all authors contributed to the final version of the manuscript.

#### Declaration of interests

AM, DWS, and TRK received funding from the Bill & Melinda Gates Foundation to support this project. AM is a Research Director of the Fonds de la Recherche Scientifque, Belgium. MS was a co-investigator on investigator-initiated research grants from Pfizer outside of the submitted work. VV received funding from the University of Sophia-Antipolis and the Institut National de la Santé et de la Recherche Santé France SAH served on ad-hoc advisory boards for Sanofi Pasteur, GlaxoSmithKline, the Bill & Melinda Gates Foundation, and PATH. TRK is supported in part by a Career Award in the Biomedical Sciences from the Burroughs Wellcome Fund, and a Michael Smith Foundation for Health Research Career Investigator Award. The funders had no role in determining content of the manuscript, writing of the report, or the decision to submit for

#### Acknowledaments

Videos from the collaborative workshop in Vancouver (BC, Canada) are available upon request from corresponding authors. We thank Véronique Flamand, Kinga Smolen, and Fabienne Willems for their help in the landscape analysis: Ajoke Sobanjo-ter Meulen for advice and 20 direction during the project; and Kim Marty and Simonetta Leduc of the Vaccine Evaluation Centre, Vancouver, BC, Canada, for their excellent administrative support [A: Written permission is required for all individuals mentioned by name in the acknowledgments section]

- Lawn JE, Blencowe H, Oza S, et al. Every Newborn: progress, priorities, and potential beyond survival. Lancet 2014: 384: 189-205.
- Laenen J, Roelants M, Devlieger R, Vandermeulen C. Influenza and pertussis vaccination coverage in pregnant women. Vaccine 2015;
- Beigi RH, Fortner KB, Munoz FM, et al. Maternal immunisation: opportunities for scientific advancement. Clin Infect Dis 2014; 59 (suppl 7): S408-14.
- Medina KL, Kincade PW. Pregnancy-related steroids are potential negative regulators of B lymphopoiesis. Proc Natl Acad Sci USA 1994; 91: 5382-86.
- Mahmoud F, Abul H, Omu A, Al-Rayes S, Haines D, Whaley K. Pregnancy-associated changes in peripheral blood lymphocyte subpopulations in normal Kuwaiti women. Gynecol Obstet Invest 2000; 52: 232-36.
- Zimmer JP, Garza C, Butte NF, Goldman AS. Maternal blood B-cell (CD19+) percentages and serum immunoglobulin concentrations correlate with breast-feeding behavior and serum prolactin concentration. Am J Reprod Immunol 1998; 40: 57-62.
- Matthiesen L, Berg G, Ernerudh J, Håkansson L. Lymphocyte subsets and mitogen stimulation of blood lymphocytes in preeclampsia. Am J Reprod Immunol 1989; 41: 192-203.
- Ampomah P, Stevenson L, Ofori MF, Barfod L, Hviid L. Kinetics of B cell responses to Plasmodium falciparum erythrocyte membrane protein 1 in Ghanaian women naturally exposed to malaria parasites. J Immunol 2014; 192: 5236-44.
  - Requena P, Campo JJ, Umbers AJ, et al. Pregnancy and malaria exposure are associated with changes in the B cell pool and in plasma eotaxin levels. J Immunol 2014; 193: 2971-83.
- Dauby N, Kummert C, Lecomte S, et al. Primary human cytomegalovirus infection induces the expansion of virus-specific activated and atypical memory B cells. J Înfect Dis 2014; 210: 1275-85.
- Kanda N, Tamaki K. Estrogen enhances immunoglobulin production by human PBMCs. J Allergy Clin Immunol 1999; 103: 282-88.
- 12 Correale J, Farez MF, Ysrraelit MC. Role of prolactin in B cell regulation in multiple sclerosis. J Neuroimmunol 2014; 269: 76-86.
- Pauklin S, Sernández IV, Bachmann G, Ramiro AR, Petersen-Mahrt SK. Estrogen directly activates AID transcription and function. J Exp Med 2009; 206: 99-111.

- McGregor IA, Rowe DS, Wilson ME, Billewicz WZ. Plasma immunoglobulin concentrations in an African (Gambian) community in relation to season, malaria and other infections and pregnancy. Clin Exp Immunol 1970; 7: 51–74.
- 15 Amino N, Tanizawa O, Miyai K, et al. Changes of serum immunoglobulins IgG, IgA, IgM, and IgE during pregnancy. Obstet Gynecol 1978; 52: 415–20.
- 16 Bondt A, Rombouts Y, Selman MHJ, et al. Immunoglobulin G (IgG) Fab glycosylation analysis using a new mass spectrometric high-throughput profiling method reveals pregnancy-associated changes. Mol Cell Proteomics 2014; 13: 3029–39.
- 17 Pincetic A, Bournazos S, DiLillo DJ, et al. Type I and type II Fc receptors regulate innate and adaptive immunity. Nat Immunol 2014; 15: 707–16.
- 18 Bondt A, Selman MHJ, Deelder AM, et al. Association between galactosylation of immunoglobulin G and improvement of rheumatoid arthritis during pregnancy is independent of sialylation. J Proteome Res 2013; 12: 4522–31.
- 19 Ackerman ME, Crispin M, Yu X, et al. Natural variation in Fc glycosylation of HIV-specific antibodies impacts antiviral activity. J Clin Invest 2013; 123: 2183–92.
- 20 Mahan AE, Jennewein MF, Suscovich T, et al. Antigen-specific antibody glycosylation is regulated via vaccination. PLoS Pathog 2016: 12: e1005456.
- 21 Yoshimura T, Inaba M, Sugiura K, et al. Analyses of dendritic cell subsets in pregnancy. Am J Reprod Immunol 1989; 50: 137–45.
- 22 Della Bella S, Giannelli S, Cozzi V, et al. Incomplete activation of peripheral blood dendritic cells during healthy human pregnancy. Clin Exp Immunol 2011: 164: 180–92.
- 23 Ueda Y, Hagihara M, Okamoto A, et al. Frequencies of dendritic cells (myeloid DC and plasmacytoid DC) and their ratio reduced in pregnant women: comparison with umbilical cord blood and normal healthy adults. *Hum Immunol* 2003; 64: 1144–51.
- 24 Young BC, Stanic AK, Panda B, Rueda BR, Panda A. Longitudinal expression of toll-like receptors on dendritic cells in uncomplicated pregnancy and postpartum. Am J Obstet Gynecol 2014; 210: 445.
- 25 Klein SL, Flanagan KL. Sex differences in immune responses. *Nat Rev Immunol* 2016; **16**: 626–38.
- 26 Healy CM, Rench MA, Baker CJ. Importance of timing of maternal combined tetanus, diphtheria, and acellular pertussis (Idap) immunisation and protection of young infants. Clin Infect Dis 2013; 56: 539–44.
- 27 Sperling RS, Engel SM, Wallenstein S, et al. Immunogenicity of trivalent inactivated influenza vaccination received during pregnancy or postpartum. Obstet Gynecol 2012; 119: 631–39.
- 28 Gupta I, Ratho RK. Immunogenicity and safety of two schedules of hepatitis B vaccination during pregnancy. J Obstet Gynaecol Res 2003; 29: 84–86
- 29 Baker CJ, Rench MA, McInnes P. Immunisation of pregnant women with group B streptococcal type III capsular polysaccharide-tetanus toxoid conjugate vaccine. *Vaccine* 2003; 21: 3468–72.
- 30 Hulka JF. Effectiveness of polyvalent influenze vaccine in pregnancy. Report of a controlled study during an outbreak of Asian influenza. Obstet Gynecol 1964; 23: 830–37.
- 31 Murray DL, Imagawa DT, Okada DM, St Geme JW. Antibody response to monovalent A/New Jersey/8/76 influenza vaccine in pregnant women. J Clin Microbiol 1979; 10: 184–87.
- 32 Schlaudecker EP, McNeal MM, Dodd CN, Ranz JB, Steinhoff MC. Pregnancy modifies the antibody response to trivalent influenza immunisation. J Infect Dis 2012; 206: 1670–73.
- 33 Bischoff AL, F
  ølsgaard NV, Carson CG, et al. Altered response to A(H1N1)pnd09 vaccination in pregnant women: a single blinded randomized controlled trial. PLoS One 2013; 8: e56700.
- 34 Kay AW, Bayless NL, Fukuyama J, et al. Pregnancy does not attenuate 50 the antibody or plasmablast response to inactivated influenza vaccine. J Infact Dis 2015; 212: 861–70.
- 35 Brabin BJ, Nagel J, Hagenaars AM, Ruitenberg E, van Tilborgh AM. The influence of malaria and gestation on the immune response to one and two doses of adsorbed tetanus toxoid in pregnancy. Bull World Health Organ 1984; 62: 919–30.
- 36 Hardegree MC, Barile MF, Pittman M, Schofield FD, Maclennan R, Kelly A. Immunisation against neonatal tetanus in New Guinea. Bull World Health Organ 1970; 43: 439–51.

- Munoz FM, Bond NH, Maccato M, et al. Safety and immunogenicity of tetanus diphtheria and acellular pertussis (Tdap) immunisation during pregnancy in mothers and infants: a randomized clinical trial. JAMA 2014; 311: 1760–69.
- 38 Huygen K, Cabore RN, Maertens K, Van Damme P, Leuridan E. Humoral and cell mediated immune responses to a pertussis containing vaccine in pregnant and nonpregnant women. *Vaccine* 2015: 33: 4117–23.
  - 39 Madhi SA, Cutland CL, Jose L, et al. Safety and immunogenicity of an investigational maternal trivalent group B streptococcus vaccine in healthy women and their infants: a randomised phase 1b/2 trial. *Lancet Infect Dis* 2016; 16: 923–34.
- 10 40 Ohfuji S, Fukushima W, Deguchi M, et al. Immunogenicity of a monovalent 2009 influenza A (H1N1) vaccine among pregnant women: lowered antibody response by prior seasonal vaccination. J Infect Dis 2011; 203: 1301–08.
  - 41 Abu Raya B, Bamberger E, Almog M, Peri R, Srugo I, Kessel A. Immunisation of pregnant women against pertussis: the effect of timing on antibody avidity. *Vaccine* 2015; 33: 1948–52.
- <sup>5</sup> 42 Maertens K, Hoang THT, Caboré RN, Leuridan E. Avidity of maternal pertussis antibodies after vaccination during pregnancy. Vaccine 2015; 33: 5489.
- Eberhardt CS, Blanchard-Rohner G, Lemaître B, et al.
   Maternal immunisation earlier in pregnancy maximizes antibody transfer and expected infant seropositivity against pertussis.
   Clin Infect Dis 2016; 62: 829–36.
  - 44 Christian LM, Porter K, Karlsson E, Schultz-Cherry S, Iams JD. Serum proinflammatory cytokine responses to influenza virus vaccine among women during pregnancy versus non-pregnancy. Am J Reprod Immunol 1989; 70: 45–53.
- 45 Regan AK, Tracey L, Blyth CC, et al. A prospective cohort study comparing the reactogenicity of trivalent influenza vaccine in pregnant and non-pregnant women. *BMC Pregnancy Childbirth* 2015; **15**: 61.
- 46 Gandhi M, Devaraj S, Sangi-Haghpeykar H, Mastrobattista J. The effect of body mass index on post-vaccination maternal and neonatal pertussis antibody levels. J Reprod Immunol 2015; 112: 24 27
- 30 47 Jones CE, Naidoo S, De Beer C, Esser M, Kampmann B, Hesseling AC. Maternal HIV infection and antibody responses against vaccine-preventable diseases in uninfected infants. *JAMA* 2011: 305: 576–84.
  - 48 van den Berg JP, Westerbeek EA, Berbers GA, van Gageldonk PG, van der Klis FR, van Elburg RM. Transplacental transport of IgG antibodies specific for pertussis, diphtheria, tetanus, Haemophilus influenzae type b, and Neisseria meningitidis serogroup C is lower in preterm compared with term infants. Pediatr Infect Dis J 2010; 29: 801–05.
  - 49 Mulholland K, Suara RO, Siber G, et al. Maternal immunisation with *Haemophilus influenzae* type b polysaccharide-tetanus protein conjugate vaccine in The Gambia. *JAMA* 1996; 275: 1182–88.
  - 50 Maertens K, Caboré RN, Huygen K, Hens N, Van Damme P, Leuridan E. Pertussis vaccination during pregnancy in Belgium: results of a prospective controlled cohort study. *Vaccine* 2016; 34: 142–50.
  - 51 Cavalcante RS, Kopelman BI, Costa-Carvalho BT. Placental transfer of *Haemophilus influenzae* type b antibodies in malnourished pregnant women. *Braz J Infect Dis* 2008; 12: 47–51.
- 5 52 Siddiqua TJ, Ahmad SM, Ahsan KB, et al. Vitamin B12 supplementation during pregnancy and postpartum improves B12 status of both mothers and infants but vaccine response in mothers only: a randomized clinical trial in Bangladesh. Eur J Nutr 2016: 55: 281–93.
  - 53 Madhi SA, Cutland CL, Kuwanda L, et al. Influenza vaccination of pregnant women and protection of their infants. N Engl J Med 2014; 371: 918–31.
- 54 Nunes MC, Cutland CL, Dighero B, et al. Kinetics of hemagglutination-inhibiting antibodies following maternal influenza vaccination among mothers with and those without HIV infection and their infants. J Infect Dis 2015; 212: 1976–87.
- 55 Heyderman RS, Madhi SA, French N, et al. Group B streptococcus vaccination in pregnant women with or without HIV in Africa: a non-randomised phase 2, open-label, multicentre trial. *Lancet Infect Dis* 2016; 16: 546–55.

- 56 McSorley HJ, Maizels RM. Helminth infections and host immune regulation. Clin Microbiol Rev 2012; 25: 585–608.
- 57 Simister NE. Placental transport of immunoglobulin G. *Vaccine* 2003; 21: 3365–69.
- 58 Bundhoo A, Paveglio S, Rafti E, Dhongade A, Blumberg RS, Matson AP. Evidence that FcRn mediates the transplacental passage of maternal IgE in the form of IgG anti-IgE/IgE immune complexes. Clin Exp Allergy 2015; 45: 1085–98.
- 59 Stapleton NM, Einarsdóttir HK, Stemerding AM, Vidarsson G. The multiple facets of FcRn in immunity. *Immunol Rev* 2015; 268: 253–68.
- 60 Roopenian DC, Akilesh S. FcRn: the neonatal Fc receptor comes of age. Nat Rev Immunol 2007; 7: 715–25.
- 61 Firan M, Bawdon R, Radu C, et al. The MHC class I-related receptor, FcRn, plays an essential role in the maternofetal transfer of gamma-globulin in humans. *Int Immunol* 2001; 13: 993–1002.
- 62 Malek A, Sager R, Kuhn P, Nicolaides KH, Schneider H. Evolution of maternofetal transport of immunoglobulins during human pregnancy. Am J Reprod Immunol 1996; 36: 248–55.
- 63 Heininger U, Riffelmann M, Leineweber B, Wirsing von Koenig CH. Maternally derived antibodies against Bordetella pertussis antigens pertussis toxin and filamentous hemagglutinin in preterm and full term newborns. Pediatr Infect Dis J 2009; 28: 443–45.
- 64 van den Berg JP, Westerbeek EA, van der Klis FR, Berbers GA, van Elburg RM. Transplacental transport of IgG antibodies to preterm infants: a review of the literature. Early Hum Dev 2011; 87: 67–72.
- 65 van den Berg JP, Westerbeek EAM, Smits GP, van der Klis FRM, Berbers GAM, van Elburg RM. Lower transplacental antibody transport for measles, mumps, rubella and varicella zoster in very preterm infants. PLoS One 2014; 9: e94714.
- 66 Palmeira P, Quinello C, Silveira-Lessa AL, Zago CA, Carneiro-Sampaio M. IgG placental transfer in healthy and pathological pregnancies. Clin Dev Immunol 2012; 2012: 985646.
- 67 Blencowe H, Cousens S, Oestergaard MZ, et al. National, regional, and worldwide estimates of preterm birth rates in the year 2010 with time trends since 1990 for selected countries: a systematic analysis and implications. *Lancet* 2012; 379: 2162–72.
- 68 Garty BZ, Ludomirsky A, Danon YL, Peter JB, Douglas SD. Placental transfer of immunoglobulin G subclasses. Clin Diagn Lab Immunol 1994; 1: 667–69.
- 69 Vidarsson G, Dekkers G, Rispens T. IgG subclasses and allotypes: from structure to effector functions. Front Immunol 2014; 5: 520.
- 70 de Voer RM, van der Klis FR, Nooitgedagt JE, et al. Seroprevalence and placental transportation of maternal antibodies specific for Neisseria meningitidis serogroup C, Haemophilus influenzae type B, diphtheria, tetanus, and pertussis. Clin Infect Dis 2009; 49: 58–64.
- 71 Leineweber B, Grote V, Schaad UB, Heininger U. Transplacentally acquired immunoglobulin G antibodies against measles, mumps, rubella and varicella-zoster virus in preterm and full term newborns. *Pediatr Infect Dis J* 2004; 23: 361–63.
- 72 Munoz FM, Englund JA, Cheesman CC, et al. Maternal immunisation with pneumococcal polysaccharide vaccine in the third trimester of gestation. *Vaccine* 2001; 20: 826–37.
- 73 Lin FY, Weisman LE, Azimi PH, et al. Level of maternal IgG anti-group B streptococcus type III antibody correlated with protection of neonates against early-onset disease caused by this pathogen. J Infect Dis 2004; 190: 928–34.
- 74 Baker CJ, Carey VJ, Rench MA, et al. Maternal antibody at delivery protects neonates from early onset group B streptococcal disease. J Infect Dis 2014; 209: 781–88.
- 75 Steinhoff MC, Omer SB, Roy E, et al. Influenza immunisation in pregnancy—antibody responses in mothers and infants. N Engl J Med 2010; 362: 1644–46.
- 76 Avanzini MA, Pignatti P, Chirico G, Gasparoni A, Jalil F, Hanson LA. Placental transfer favours high avidity IgG antibodies. Acta Paediatr 1998; 87: 180–85.
- 77 Sennhauser FH, Balloch A, Macdonald RA, Shelton MJ, Roberton DM. Maternofetal transfer of IgG anti-Escherichia coli antibodies with enhanced avidity and opsonic activity in very premature neonates. Pediatr Res 1990; 27: 365–71.
- 78 Williams PJ, Arkwright PD, Rudd P, et al. Short communication: selective placental transport of maternal IgG to the fetus. *Placenta* 1995; 16: 749–56.

- 79 Einarsdottir HK, Selman MHJ, Kapur R, et al. Comparison of the Fc glycosylation of fetal and maternal immunoglobulin G. Glycoconj J 2013; 30: 147–57.
- 80 Hartter HK, Oyedele OI, Dietz K, Kreis S, Hoffman JP, Muller CP. Placental transfer and decay of maternally acquired antimeasles antibodies in Nigerian children. *Pediatr Infect J* 2000; 19: 635–41.
- 81 Okoko BJ, Wesumperuma LH, Ota MO, et al. The influence of placental malaria infection and maternal hypergammaglobulinemia on transplacental transfer of antibodies and IgG subclasses in a rural west African population. J Infect Dis 2001; 184: 627–32.
- 82 Cumberland P, Shulman CE, Maple PA, et al. Maternal HIV infection and placental malaria reduce transplacental antibody transfer and tetanus antibody levels in newborns in Kenya. J Infect Dis 2007; 196: 550–57.
- 83 Atwell JE, Thumar B, Robinson LJ, et al. Impact of placental malaria and hypergammaglobulinemia on transplacental transfer of respiratory syncytial virus antibody in Papua New Guinea. J Infect Dis 2016; 213: 423–31.
- Name 15 84 Dangor Z, Kwatra G, Izu A, et al. HIV-1 Is associated with lower group B streptococcus capsular and surface-protein IgG antibody levels and reduced transplacental antibody transfer in pregnant women. J Infect Dis 2015; 212: 453–62.
  - 85 Le Doare K, Taylor S, Allen L, et al. Placental transfer of anti-group B streptococcus immunoglobulin G antibody subclasses from HIV-infected and uninfected women to their uninfected infants. AIDS 2016: 30: 471–75.
  - 86 Abu-Raya B, Smolen KK, Willems F, Kollmann TR, Marchant A. Transfer of maternal antimicrobial immunity to HIV-exposed uninfected newborns. Front Immunol 2016; 7: 338.
  - 87 Bahl R, Frost C, Kirkwood BR, et al. Infant feeding patterns and risks of death and hospitalization in the first half of infancy: multicentre cohort study. Bull World Health Organ 2005; 83: 418–26.
- Note of the State of State
- 89 Schlaudecker EP, Steinhoff MC, Omer SB, et al. IgA and neutralizing antibodies to influenza a virus in human milk: a randomized trial of antenatal influenza immunisation. *PLoS One* 2013; 8: e70867.
- 30 90 Brandtzaeg P. Mucosal immunity: integration between mother and the breast-fed infant. Vaccine 2003; 21: 3382–88.
  - 91 Fouda GG, Amos JD, Wilks AB, et al. Mucosal immunisation of lactating female rhesus monkeys with a transmitted/founder HIV-1 envelope induces strong Env-specific IgA antibody responses in breast milk. J Virol 2013; 87: 6986–99.
- 92 Corthesy B. Multi-faceted functions of secretory IgA at mucosal surfaces. Front Immunol 2013; 4: 185.
- 93 Ogra SS, Weintraub D, Ogra PL. Immunologic aspects of human colostrum and milk. III. Fate and absorption of cellular and soluble components in the gastrointestinal tract of the newborn. J Immunol 1977; 119: 245–48.
- 94 Vukavic T. Intestinal absorption of IgA in the newborn.
   40 J Pediatr Gastroenterol Nutr 1983; 2: 248–51.
- 95 Moon SS, Wang Y, Shane AL, et al. Inhibitory effect of breast milk on infectivity of live oral rotavirus vaccines. *Pediatr Infect Dis J* 2010; 29: 919–23.
- 96 Rongsen-Chandola T, Strand TA, Goyal N, et al. Effect of withholding breastfeeding on the immune response to a live oral rotavirus vaccine in north Indian infants. Vaccine 2014; 32 (suppl 1): A134–39.
- 97 Maertens K, De Schutter S, Braeckman T, et al. Breastfeeding after maternal immunisation during pregnancy: providing immunological protection to the newborn: a review. Vaccine 2014; 32: 1786–92.
- Pollara J, McGuire E, Fouda GG, et al. Association of HIV-1 envelope-specific breast milk IgA responses with reduced risk of postnatal mother-to-child transmission of HIV-1. *J Virol* 2015;
   89: 9952–61.
  - 99 Castellote C, Casillas R, Ramirez-Santana C, et al. Premature delivery influences the immunological composition of colostrum and transitional and mature human milk. J Nutr 2011; 141: 1181–87.
  - 100 Ballard O, Morrow AL. Human milk composition: nutrients and bioactive factors. Pediatr Clin North Am 2013; 60: 49–74.
- 55 101 Shapiro RL, Lockman S, Kim S, et al. Infant morbidity, mortality, and breast milk immunologic profiles among breast-feeding HIV-infected and HIV-uninfected women in Botswana. J Infect Dis 2007; 196: 562–69.

- 102 Moussa S, Jenabian MA, Gody JC, et al. Adaptive HIV-specific B cell-derived humoral immune defenses of the intestinal mucosa in children exposed to HIV via breast-feeding. PLoS One 2013; 8: e63408.
- 103 Brussow H, Barclay D, Sidoti J, et al. Effect of malnutrition on serum and milk antibodies in Zairian women. Clin Diagn Lab Immunol 1996: 3: 37-41.
- 104 Islam SK, Ahmed L, Khan MN, Huque S, Begum A, Yunus AB. Immune components (IgA, IgM, IgG, immune cells) of colostrum of Bangladeshi mothers. Pediatr Int 2006; 48: 543-48.
- 105 Tuaillon E, Valea D, Becquart P, et al. Human milk-derived B cells: a highly activated switched memory cell population primed to secrete antibodies. J Immunol 2009; 182: 7155-62.
- 106 Hurley WL, Theil PK. Perspectives on immunoglobulins in colostrum and milk. Nutrients 2011; 3: 442-74.
- 107 Gao X, McMahon RJ, Woo JG, Davidson BS, Morrow AL, Zhang Q. Temporal changes in milk proteomes reveal developing milk functions. J Proteome Res 2012; 11: 3897-907.
- 108 Edwards MS, Munoz FM, Baker CI, Antibodies to type III group B streptococcal polysaccharide in breast milk. Pediatr Înfect Dis J 2004; 15 **23**: 961–63.
- 109 Mabuka J, Nduati R, Odem-Davis K, Peterson D, Overbaugh J. HIV-specific antibodies capable of ADCC are common in breastmilk and are associated with reduced risk of transmission in women with high viral loads. PLoS Pathog 2012; 8: e1002739.
- 110 Ehlinger EP, Webster EM, Kang HH, et al. Maternal cytomegalovirus-specific immune responses and symptomatic postnatal cytomegalovirus transmission in very low-birth-weight preterm infants. J Infect Dis 2011; 204: 1672-82.
- 111 Yoshida M, Claypool SM, Wagner JS, et al. Human neonatal Fc receptor mediates transport of IgG into luminal secretions for delivery of antigens to mucosal dendritic cells. Immunity 2004;
- 112 Yoshida M, Kobayashi K, Kuo TT, et al. Neonatal Fc receptor for IgG regulates mucosal immune responses to luminal bacteria. I Clin Invest 2006; 116: 2142-51.
- 113 Harris NL, Spoerri I, Schopfer JF, et al. Mechanisms of neonatal mucosal antibody protection. J Immunol 2006; 177: 6256-62.
- 114 Baker K. Oiao SW. Kuo T. et al. Immune and non-immune functions of the (not so) neonatal Fc receptor, FcRn. Semin Immunopathol 2009; 31: 223-36.
- Wirt DP, Adkins LT, Palkowetz KH, Schmalstieg FC, Goldman AS. Activated and memory T lymphocytes in human milk. Cytometry 1992; 13: 282-90.
- 116 Hassiotou F, Geddes DT, Hartmann PE. Cells in human milk: state of the science. J Hum Lact 2013; 29: 171-82.
- 117 Wilks AB, Christian EC, Seaman MS, et al. Robust vaccine-elicited cellular immune responses in breast milk following systemic simian immunodeficiency virus DNA prime and live virus vector boost vaccination of lactating rhesus monkeys. J Immunol 2010; 185: 7097-106
- 118 Mahlokozera T, Kang HH, Goonetilleke N, et al. The magnitude and kinetics of the mucosal HIV-specific CD8+ T lymphocyte response and virus RNA load in breast milk. PLoS One 2011;
- 119 Mohr JA. The possible induction and-or acquisition of cellular hypersensitivity associated with ingestion of colostrum. J Pediatr 1973; 82: 1062-64.
- 120 Schlesinger JJ, Covelli HD. Evidence for transmission of lymphocyte 45 145 Mooi FR, de Greeff SC. The case for maternal vaccination against responses to tuberculin by breast-feeding. Lancet 1977; 2: 529-32.
- 121 Lawrence RM, Lawrence RA. Breast milk and infection. Clin Perinatol 2004; 31: 501-28.
- 122 John-Stewart GC, Mbori-Ngacha D, Payne BL, et al. HV-1-specific cytotoxic T lymphocytes and breast milk HIV-1 transmission. I Infect Dis 2009; 199: 889-98.
- 123 Qureshi K, Molbak K, Sandstrom A, et al. Breast milk reduces the risk of illness in children of mothers with cholera; observations from an epidemic of cholera in Guinea-Bissau. Pediatr Infect Dis I 2006: **25**: 1163–66.
- 124 Alain S, Dommergues MA, Jacquard AC, Caulin E, Launay O. State of the art: could nursing mothers be vaccinated with attenuated live virus vaccine? Vaccine 2012; 30: 4921-26.
- Verhasselt V. Is infant immunisation by breastfeeding possible? Philos Trans R Soc Lond B Biol Sci 2015; 370: 20140139.

- 1 126 Wan AK, Seow WK, Purdie DM, Bird PS, Walsh LJ, Tudehope DI. Immunoglobulins in saliva of preterm and full-term infants. Oral Microbiol Immunol 2003; 18: 72-78.
- Morris D, McDonald JC. Failure of hyperimmune gamma globulin to prevent whooping cough. Arch Dis Child 1957; 32: 236-39.
- 5 128 Kirimanjeswara GS, Mann PB, Harvill ET. Role of antibodies in immunity to bordetella infections. Infect Immun 2003; 71: 1719-24.
  - Warfel JM, Zimmerman LI, Merkel TJ. Acellular pertussis vaccines protect against disease but fail to prevent infection and transmission in a nonhuman primate model. Proc Natl Acad Sci USA 2014; 111: 787-92.
- 130 Dabrera G, Amirthalingam G, Andrews N, et al. A case-control study to estimate the effectiveness of maternal pertussis vaccination in protecting newborn infants in England and Wales, 2012-2013. Clin Infect Dis 2015; 60: 333-37.
  - 131 Zinkernagel RM. Maternal antibodies, childhood infections, and autoimmune diseases. N Engl J Med 2001; 345: 1331-35.
  - Ghetie V, Ward ES. Transcytosis and catabolism of antibody. 132 Immunol Res 2002; 25: 97-113.
  - 133 Gutierrez G, Gentile T, Miranda S, Margni RA. Asymmetric antibodies: a protective arm in pregnancy. Chem Immunol Allergy 2005; 89: 158–68.
  - Sarvas H, Seppälä I, Kurikka S, Siegberg R, Mäkelä O. Half-life of the maternal IgG1 allotype in infants. J Clin Immunol 1993;
- <sup>20</sup> 135 Healy CM, Munoz FM, Rench MA, Halasa NB, Edwards KM, Baker CJ. Prevalence of pertussis antibodies in maternal delivery, cord, and infant serum. J Infect Dis 2004; 190: 335-40.
  - Caceres VM, Strebel PM, Sutter RW. Factors determining prevalence of maternal antibody to measles virus throughout infancy: a review. Clin Infect Dis 2000; 31: 110-19.
- Ochola R, Sande C, Fegan G, et al. The level and duration of RSV-specific maternal IgG in infants in Kilifi Kenya. PLoS One 2009; 4: e8088.
- 138 Chu HY, Steinhoff MC, Magaret A, et al. Respiratory syncytial virus transplacental antibody transfer and kinetics in mother-infant pairs in Bangladesh. J Infect Dis 2014; 210: 1582-89.
- Siegrist CA. Mechanisms by which maternal antibodies influence infant vaccine responses: review of hypotheses and definition of main determinants. Vaccine 2003; 21: 3406-12.
- 140 Niewiesk S. Maternal antibodies: clinical significance, mechanism of interference with immune responses, and possible vaccination strategies. Front Immunol 2014; 5: 446.
- Faucette AN, Unger BL, Gonik B, Chen K. Maternal vaccination: moving the science forward. Hum Reprod Update 2015; 21: 119-35.
- 142 Nair N, Gans H, Lew-Yasukawa L, Long-Wagar AC, Arvin A, Griffin DE. Age-dependent differences in IgG isotype and avidity induced by measles vaccine received during the first year of life. J Infect Dis 2007; 196: 1339-45.
- Van Savage J, Decker MD, Edwards KM, Sell SH, Karzon DT. Natural history of pertussis antibody in the infant and effect on 40 vaccine response. J Infect Dis 1990; 161: 487–92.
  - 144 Englund JA, Anderson EL, Reed GF, et al. The effect of maternal antibody on the serologic response and the incidence of adverse reactions after primary immunisation with acellular and whole-cell pertussis vaccines combined with diphtheria and tetanus toxoids. Pediatrics 1995; **96**: 580–84.
- pertussis. Lancet Infect Dis 2007; 7: 614-24.
  - Ladhani SN, Andrews NJ, Southern J, et al. Antibody responses after primary immunisation in infants born to women receiving a pertussis-containing vaccine during pregnancy: single arm observational study with a historical comparator. Clin Infect Dis 2015; 61: 1637-44
- Hoang HTT, Leuridan E, Maertens K, et al. Pertussis vaccination during pregnancy in Vietnam: results of a randomized controlled trial pertussis vaccination during pregnancy. Vaccine 2016; 34: 151-59.
  - 148 Edwards KM. Pertussis: an important target for maternal immunisation. Vaccine 2003; 21: 3483-86.
- 149 Barington T, Gyhrs A, Kristensen K, Heilmann C. Opposite effects of actively and passively acquired immunity to the carrier on 55 responses of human infants to a Haemophilus influenzae type b conjugate vaccine. Infect Immun 1994; 62: 9-14.

- 150 Englund JA, Glezen WP, Turner C, Harvey J, Thompson C, Siber GR. Transplacental antibody transfer following maternal immunisation with polysaccharide and conjugate Haemophilus influenzae type b vaccines. J Infect Dis 1995; 171: 99–105.
- 151 Kurikka S, Olander RM, Eskola J, Käyhty H. Passively acquired anti-tetanus and anti-haemophilus antibodies and the response to Haemophilus influenzae type b-tetanus toxoid conjugate vaccine in infancy. Pediatr Infect Dis J 1996; 15: 530–35.
- 152 Rastogi D, Wang C, Mao X, Lendor C, Rothman PB, Miller RL. Antigen-specific immune responses to influenza vaccine in utero. J Clin Invest 2007; 117: 1637–46.
- 153 Vanderbeeken Y, Sarfati M, Bose R, Delespesse G. In utero immunisation of the fetus to tetanus by maternal vaccination during pregnancy. Am J Reprod Immunol Microbiol 1985; 8: 39–42.
- 154 Gill TJ 3rd, Repetti CF, Metlay LA, et al. Transplacental immunisation of the human fetus to tetanus by immunisation of the mother. J Clin Invest 1983; 72: 987–96.

- 1 155 Aaby P, Kollmann TR, Benn CS. Nonspecific effects of neonatal and infant vaccination: public-health, immunological and conceptual challenges. Nat Immunol 2014; 15: 895–99.
  - 156 Bischoff AL, Folsgaard NV, Vissing NH, Birch S, Brix S, Bisgaard H. Airway mucosal immune-suppression in neonates of mothers receiving A(H1N1)pnd09 vaccination during pregnancy. Pediatr Infect Dis J 2014; 30: 84–90.
  - x1 Centers for Disease Contol and Prevention. Updated recommendations for use of tetanus toxoid, reduced diphtheria toxoid, and acellular pertussis vaccine (Tdap) in pregnant women— Advisory Committee on Immunization Practices (ACIP). MMWR Morb Mortal Wkly Rep 2013; 62: 131–35. [A: Journal ref added]

35 40

50

55

10

20

25