

UC Irvine

UC Irvine Previously Published Works

Title

Laser coagulation and hemostasis of large diameter blood vessels: effect of shear stress and flow velocity

Permalink

<https://escholarship.org/uc/item/9988747w>

Journal

Scientific Reports, 12(1)

ISSN

2045-2322

Authors

Katta, Nitesh
Santos, Daniel
McElroy, Austin B
et al.

Publication Date

2022

DOI

10.1038/s41598-022-12128-1

Peer reviewed



OPEN

Laser coagulation and hemostasis of large diameter blood vessels: effect of shear stress and flow velocity

Nitesh Katta^{1✉}, Daniel Santos², Austin B. McElroy², Arnold D. Estrada², Glori Das², Mohammad Mohsin³, Moses Donovan⁴ & Thomas E. Milner¹

Photocoagulation of blood vessels offers unambiguous advantages to current radiofrequency approaches considering the high specificity of blood absorption at available laser wavelengths (e.g., 532 nm and 1.064 μm). Successful treatment of pediatric vascular lesions, such as port-wine stains requiring microvascular hemostasis, has been documented. Although laser treatments have been successful in smaller diameter blood vessels, photocoagulation of larger sized vessels is less effective. The hypothesis for this study is that a primary limitation in laser coagulation of large diameter blood vessels (500–1000 μm) originates from shear stress gradients associated with higher flow velocities along with temperature-dependent viscosity changes. Laser (1.07 μm) coagulation of blood vessels was tested in the chicken chorio-allantoic membrane (CAM). A finite element model is developed that includes hypothetical limitations in laser coagulation during irradiation. A protocol to specify laser dosimetry is derived from OCT imaging and angiography observations as well as finite element model results. Laser dosimetry is applied in the CAM model to test the experimental hypothesis that blood shear stress and flow velocity are important parameters for laser coagulation and hemostasis of large diameter blood vessels (500–1000 μm). Our experimental results suggest that shear stress and flow velocity are fundamental in the coagulation of large diameter blood vessels (500–1000 μm). Laser dosimetry is proposed and demonstrated for successful coagulation and hemostasis of large diameter CAM blood vessels.

Laser-based microvascular hemostasis was first demonstrated in 1981 under the principle of selective photothermolysis¹. Subsequently, additional work^{2–7} was carried out to provide laser dosimetry to optimize various parameters (e.g., wavelength, pulse duration, fluence) to achieve hemostasis. Port wine stains (PWSs) are vascular malformations^{7–10} and were one of the first target applications of selective photothermolysis, where a vascular-specific wavelength irradiates blood and is differentially absorbed by hemoglobin in blood.

Conventional surgical procedures still rely on radio frequency (RF) electrocautery for coagulation and hemostasis^{11,12}, where collateral nonspecific damage zones are as large as a few millimeters (much larger than the size of targeted blood vessels). These RF tools operate at higher input energy/power and are utilized for soft-tissue cutting and represent commonly employed tools for most surgical procedures. Laser coagulation of blood vessels, however, has limited use in the operating room (OR) for coagulation/hemostasis during conventional surgery, outside of dermal/oral/dental soft tissue surgical applications^{13–16}. One possible reason for underutilization of lasers in the OR relates to the selection of laser dosimetry that offers good hemostasis while providing relatively comparable tissue removal rates to conventional cautery tools. For example, because the emission wavelength (2.94 μm) of Er:YAG laser radiation coincides with an absolute peak in water absorption¹⁷, tissue ablation is highly energy efficient¹⁸; however, Er:YAG lasers are not used in general surgery (outside of skin cosmetic resurfacing) and are not a viable candidate for vascular coagulation. Alternatively, diode lasers have been shown to perform well for coagulation of blood vessels through hot tip hemostasis¹⁵ approach but suffer from nonspecific thermal damage. Pulsing strategies have been demonstrated *ex vivo* to reduce thermal damage and modeling results have shown laser sealing of blood vessels to have adequate mechanical strength^{19,20}.

¹University of California at Irvine, Irvine, CA, USA. ²The University of Texas at Austin, Austin, TX, USA. ³University of Connecticut, Storrs, CT, USA. ⁴Pennsylvania State University, State College, PA, USA. ✉email: nkatta@uci.edu

Recently, dual-wavelength approaches^{21,22} have been successfully tested by demonstrating high tissue removal rates through pulsed laser irradiation, while coagulation is achieved through continuous (CW) irradiation^{21,23}.

Second, another challenge for laser-based vascular hemostasis during surgery is the presence of large diameter (500–1000 μm) blood vessels. In this manuscript we refer to blood vessels between 500 μm and 1 mm as “large blood vessels”. Histopathological and other imaging studies have shown that PWS dilated blood vessel sizes vary between patients in the range of 200–750 μm ^{24–26}. PWS lesions often consist of slower blood flow rates, with most blood vessels being dilated venules, unlike blood vessels encountered during surgery in the OR. Limitations of selective photothermolysis are also recognized in treating PWS lesions in adults compared to infants who possess smaller diameter blood vessel capillary networks (PWS is a congenital disorder)^{8,10}. Similarly, during surgery, typical target blood vessel networks are more pronounced in terms of size and flow rates. For these cases, vascular specific wavelengths for laser hemostasis during surgery potentially offer many improvements over existing electrocautery tools. Specifically, infrared laser sources at a wavelength of approximately 1 μm allow for fiber delivery; deeper penetration in blood potentially allows for hemostasis in millimeter-diameter blood vessels (in comparison to 100 s of microns in PWS) while still offering high specificity to blood optical absorption; and deep (centimeters) tissue penetration^{27,28}. Hence, there is a need for the development of laser irradiation protocols and dosimetry for vascular-specific hemostasis in vascular networks with larger blood vessels. In this study, a state-of-the-art ytterbium (Yb) fiber laser (1.07 μm) was used to produce hemostasis in millimeter-sized blood vessels in the chicken chorio-allantoic membrane (CAM) model^{29–31}. CAMs allow for quick iterative studies due to a relatively simple preparation protocol and short gestation time and are attractive for hemostasis studies in relatively large diameter blood vessels (millimeters) with high flow rates³². In addition, for the study reported here, due to the absence of an epithelial layer with vascular networks being formed on the membrane, confounding complexities, including the presence of additional chromophores (e.g., melanin), were not present, simplifying the experiments (CAM protocols, benchtop setup, vascular analysis and photothermolysis estimations for laser dosimetry at 1.07 μm can be found in supplementary Sects. 1–3). A finite element model (FEM) was developed in COMSOL software to investigate the limitations of the current theory of photothermolysis of large blood vessels. Experimental results are presented that illustrate the importance of visco-velocity changes that occur during laser irradiation of large diameter blood vessels. Initially, laser dosimetry derived from the conventional theory of selective photothermolysis was applied to CAM vessels, resulting in failed coagulation of larger vessels. By increasing radiant energy input based on FEM, laser dosimetry is proposed and demonstrated for successful coagulation of large blood vessels.

Results and discussion

Shear stress FEM computations at the vessel wall indicate that peak stress increases during and immediately after laser irradiation (Fig. 1A). Computed changes in visco-elastic properties show significant localization of increased shear stress near the walls of larger blood vessels. The observed shear stress increase near the vessel wall can be attributed to two effects: (1) an increase in viscosity with increasing temperature (Fig. 1B) (modeled as an interpolated look-up table with data from prior literature³³) and density changes associated with the coagulum that grows from the central lumen toward the vessel wall, resulting in a higher shear force. Although heat transfer through convective flow is observed (Fig. 1D) in larger vessels as opposed to smaller vessels, Arrhenius computations yielded values of $\Omega = 1$ (corresponding to coagulation) in all cases. The results suggest that relying solely on an Arrhenius damage integral computation to predict a successful hemostatic response while ignoring the mechanical shearing effects of the coagulum, especially at the vessel wall, can introduce erroneous conclusions and artifacts. The importance of mechanical shearing effects at the vessel wall is especially important at higher blood flow velocities in larger diameter vessels.

Considering the dynamic flow-convective properties in larger blood vessels and limitations in explosive vapor bubble formation, an alternative laser-irradiation protocol was developed to increase the initial energy input into larger diameter blood vessels. First, a long duration pulse was introduced to modify the viscosity-dependent flow dynamics in the blood vessel. Long pulse durations have been previously recommended^{34–37} for effective thermolysis when using intense infrared sources, such as those utilized in this study. Considering some of the effects observed in response to long pulse irradiation, including shear stress, velocity and viscosity changes, we hypothesized that subsequent shorter duration laser pulses of reduced energy may allow for successful photocoagulation of large blood vessels. An analog to possibly better understand the laser irradiation protocol proposed here is mechanically reducing the flow/shear stress dynamics by first pinching the vessel (i.e., long pulse laser irradiation) before applying a radiant energy pulse to coagulate the vessel^{38,39}. Instead of mechanically constraining the vessel to change the shear/velocity dynamics for successful hemostasis, a long duration pulse changes these parameters as if the vessel was pinched (Fig. 1) before subsequent shorter pulses perform successful hemostasis. The mechanical strain applied to larger blood vessels while pinching using electrocautery tools often results in tissue welding to the electrodes, resulting in rupture when the tool is pulled away. COMSOL simulations highlighted the limitations of employing long laser pulses that were possible with available fluence rates at pulse durations of approximately 100–200 ms (Fig. 1). In blood vessels with smaller diameters, thermal relaxation was observed (Fig. 1D). In larger blood vessels, heat is primarily carried away from the irradiation site through fast convective blood flow (Fig. 1C,D). FEM suggests that a vascular system exposed to an initial high-energy light pulse is modified and can then better satisfy the conditions of the theory of selective photothermolysis due to the modified flow dynamics.

Limitations of laser coagulation in large vessels. After application of the derived laser irradiation protocol (Fig. 2, green bar plot), OCT angiography images recorded pre- and post- laser irradiation (Fig. 2)

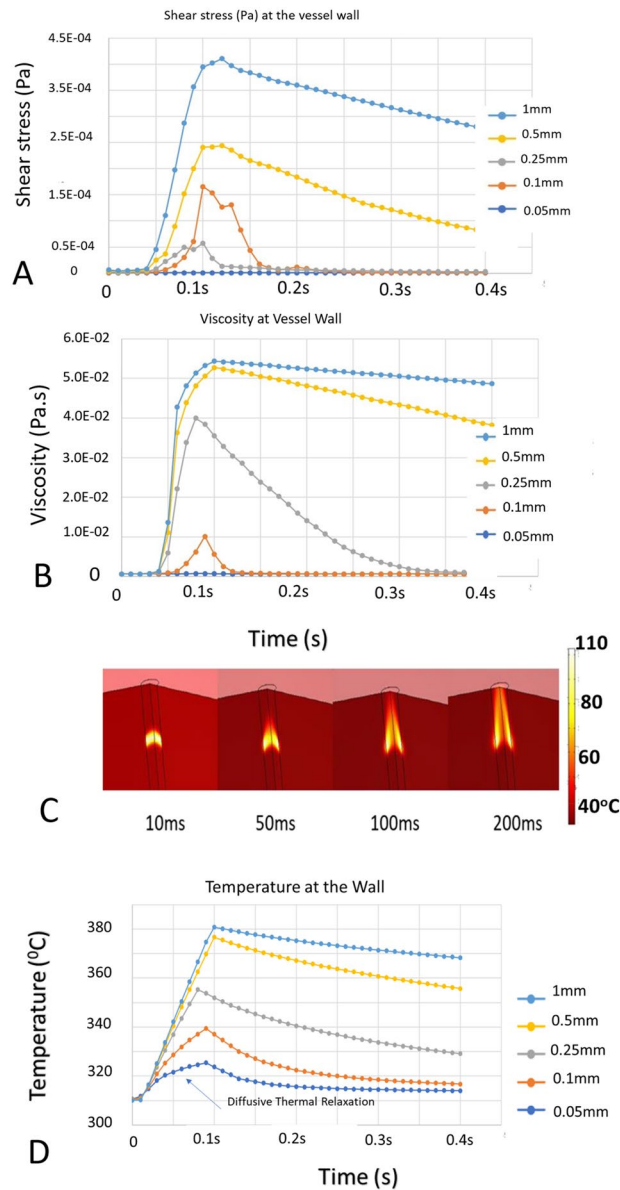


Figure 1. (A) Shear stress (Pa) at the vessel luminal wall for five blood vessel lumen diameters (0.05–1 mm) in response to pulsed laser ($1.07 \mu\text{m}$) irradiation (80 ms, 300 W average power). Shear stress increase is significantly higher post irradiation in the larger blood vessel case suggesting higher chances of coagulum breaking free from the wall as observed experimentally (Fig. 3). Irradiation is simulated as a flat top laser fluence from 0.05 to 0.1 s. (B) Viscosity (Pa.s) at the vessel luminal wall for five blood vessel sizes (0.05–1 mm) in response to pulsed laser ($1.07 \mu\text{m}$) irradiation (80 ms, 300 W average power). (C) Temperature profile within a blood vessel (1 mm diameter lumen) modified by blood flow dynamics (blood flowing from bottom to top). Heat at the walls persists while thermal energy is carried away at the center of the blood vessel. For large blood vessel diameters, heat transfer is dominated by blood flow over conduction. (D) Temperature relaxation observed during irradiation of smaller diameter blood vessels reduces viscosity whereas the viscosity in larger vessels remains elevated for longer times. The effect is consistent with an experimental result (Fig. 3, Panel C), just before the coagulum attached to the vessel wall is dislodged.

showed that larger blood vessels (greater than $200 \mu\text{m}$) were not coagulated (additional examples are provided in supplementary Figs. S5, S7, S8, S9).

We observed that arterioles more readily coagulated (red dotted line in Panel A, Fig. 3), while flow in venules (blue arrow line, Panel D, Fig. 3) frequently persisted after laser irradiation, similar to experimental observations reported previously^{5,40}. A possible explanation may be a constriction of the blood vessels and changes in blood vessel shape, as noted before⁴, where a change in vessel size was observed following laser irradiation. Since arterioles feed into vessels with smaller diameters as opposed to venules that feed into vessels with larger diameters, arterioles have a higher chance of coagulum interacting with the vessel wall downstream, resulting in flow stoppage. As observed in the FEM, shearing of the coagulum near the vessel wall is a likely failure mechanism

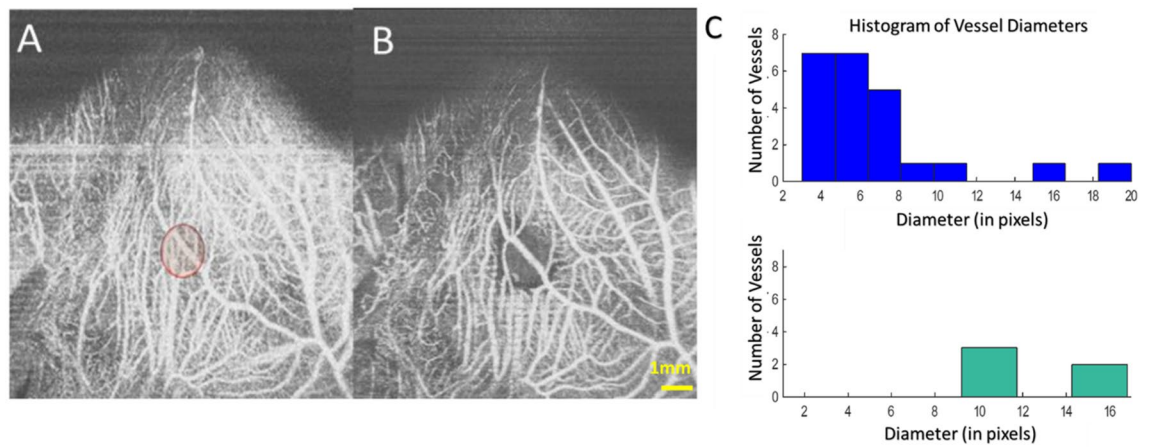


Figure 2. Coagulation of a CAM vasculature showing flow intact in the larger diameter vessels compared to the smaller microvasculature. Histogram analysis of blood vessel size distribution showing hemostasis only occurred in smaller vessels (right). Laser dosimetry: 7.7 J, 25 ms. Spot size 3 mm. Scale Bar is 1 mm. In the 512×512 enface image, each pixel corresponds to approximately $20 \mu\text{m}$.

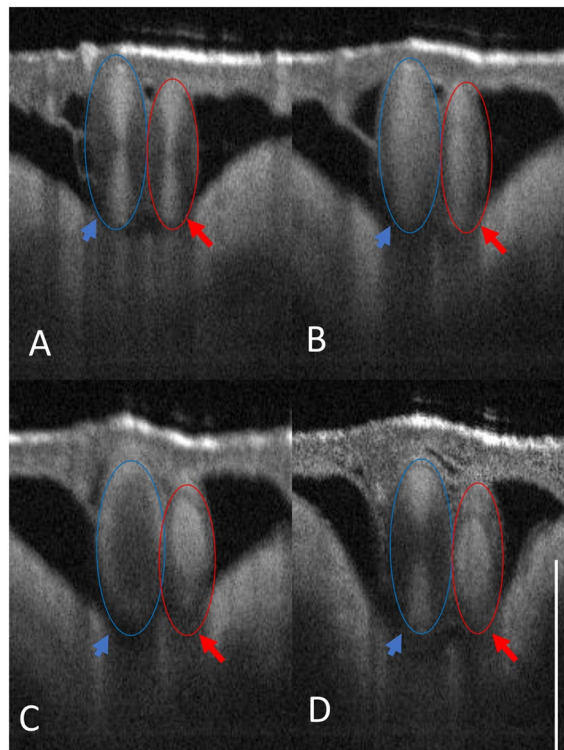


Figure 3. OCT B-scan image of CAM showing an arteriole (red)/venule (blue) pair irradiated at the same fluence rate. (A) Initial OCT B-scan before laser irradiation. (B) Coagulum formation observed in both arterioles and venules. (C) In the arteriole, the coagulum holds, resulting in hemostasis, while in the venule, the coagulum breaks free near the wall. (D) Blood flow is restored in the venule where the coagulum breaks from the vessel wall (Panel C). Please see supplementary Fig. S10 for enface images collected from this experiment. Scale bar $500 \mu\text{m}$. These arteriole/venule pairs were chosen on the CAM model appropriately sized at Day 7 with direct connection to major CAM vessels.

in the coagulation process. Experimentally, shearing of the coagulum (Fig. 3, Panel C, blue arrow) near the wall was observed in cases where laser-induced hemostasis in larger blood vessels failed.

Coagulation of smaller-sized blood vessels followed expectations of the theory of selective photothermolysis, resulting in consistent stoppage of blood flow in vessels smaller than $100\text{--}200 \mu\text{m}$ (depending on blood vessel location with respect to the laser irradiation spot). Reports from other studies also suggest the inefficacy of pulsed

dye lasers for coagulation of larger sized blood vessels and utilization of longer pulse durations to achieve better hemostasis^{35–37,41,42}.

Blood velocity and viscosity limitations. A limitation for laser photothermolysis is the relative speed of blood in larger vessels and associated higher shear stresses. Larger sized blood vessels (200/400 μm) were measured to support blood flow velocities of⁴³ 20/40 mm/s. From the principle of selective photothermolysis, long pulse durations are recommended for blood vessels with sizes greater than^{40,44} 200 μm . For larger blood vessels, the profile of laminar blood flow is parabolic, with a 40 mm/s average flow velocity (corresponding to a transit time of 1 mm per 25 ms). This simple calculation suggests that the pulse duration recommendation derived from photothermolysis can fail within the transit time of convective heat transfer via blood flow.

The Nusselt number provides a measure of the ratio of convective to conductive heat transfer and varies from 0.2 to 1.2 for CAM vessels ranging in size^{45,46} from 0.05 mm to 1 mm. Interestingly, the initial temperature increase following absorption of pulsed laser light causes a viscosity decrease³³ along with a density reduction (coagulum density being lower in solidified form due to expansion of blood cells forming spheroids as observed before^{47,48}). Additionally, a finite time was reported for the formation of spheroids (through transient time in the formation of met-Hb as described before^{47,48}).

Limitations due to shear stresses near the vessel wall. Typical blood flow velocity profiles in a vessel lumen are parabolic, implying low shear stress near the vessel center, in contrast to the vessel wall where velocity gradients are steepest. Shear stress in large vessels is an order of magnitude (at least 10 times) higher than that in the microvasculature⁴³ (< 100 microns) and can supersede the bonding energies coupling the coagulum with surrounding sites of the coagulation/vessel wall. Figure 3 shows two side-by-side cases of successful hemostasis vs unsuccessful hemostasis, where shearing near the wall (Panel C, Fig. 3) was observed, resulting in flow restoration (Panel D).

Limitations from explosive bubble formation and rupture of blood vessels. Competing requirements for laser coagulation of large blood vessels are recognized. On the one hand, too slow heating may result in transit of the coagulum through the irradiation site before bonding with the vessel wall to complete hemostasis. Considering the exponentially decreasing absorption across the blood vessel cross-section, fluence distribution can result in the formation of vapor bubbles at the top of the blood vessel (toward incident laser irradiation)²⁵.

Results from proposed pulse stacking for coagulation in larger blood vessels. Initially, in response to laser irradiation, coagulum formation is observed growing from the center to edges of the vessel lumen (Fig. 4, middle panel). In some cases, where blood flow velocity (assumed to be dependent on the size of the blood vessel) was very high (> 50 mm/s), formation of a different-colored coagulum was observed (possibly met-hemoglobin^{47,48}) that was subsequently displaced from the laser irradiation site by incoming cooler blood. Successful coagulation was observed with the enhanced stacked pulse scheme with a long duration initial pulse (in some cases approximately 5 times longer than that derived from the theory of selective photothermolysis). In some experiments, a rapid hemodynamic response was the result of successful hemostasis (Fig. S13), causing intense motions in the embryo post irradiation. OCT images highlighting hemostasis (Fig. S11) and brightfield photographs (Fig. S6) showed some hemorrhaging of capillaries around the outer rim of the laser irradiation zone. In cases where this initial energy titration was too high, explosive rupture/hemorrhage of the blood vessel was observed. Experimentally, laser dosimetry required for coagulation was computed for each vasculature. By carefully increasing the energy, coagulation success (Fig. 4, other examples in supplementary Figs. S11–13) was confirmed by histogram analysis of the resultant remnant vasculature.

FEM simulations provide a measure of the rate radiant energy that can be input into the vessel before causing explosive bubble formation within the blood vessel lumen and whether peak radiant power needs adjustment. Experimentally, irradiation times of 100–200 ms worked well for blood vessels in the range of 0.5–1 mm. Given differences in CAM vasculature in comparison to most tissues, where the CAM venules (furthest downstream from the heart) carry oxygenated blood and the arterioles (closest upstream to the heart) have deoxygenated blood, Hb has a higher absorption of 1.07 μm wavelength radiation, and the flow direction of venules (smaller into larger cross sections) makes coagulating CAM venules more challenging compared to CAM arterioles (flow into smaller cross-sections increasing likelihood of coagulum sticking to the wall or stopping flow). The CAM scenario is reversed in most tissues where the oxygenation Hb/HbO₂ content in venules/arterioles is juxtaposed. Additionally, given the arterial structure (possessing thicker and more elastic walls than venules), explosive rupture was more commonly observed in initial laser coagulation trials within CAM venules than in CAM arterioles. In addition, coagulation of CAM venules was less successful than CAM arterioles (Fig. 3). These results are consistent with the fact that CAM arterioles were more uniformly heated given the higher penetration depth of 1.07 μm light in deoxygenated blood, and CAM venules experienced increased temperatures toward the incoming radiation direction, causing localized cavitation and rupture (flow in CAM venules was faster than in arterioles^{32,43}). To prevent local cavitation and vessel rupture, the peak power for laser irradiation into CAM venules was reduced (halved) so that to maintain constant energy input, the laser irradiation times of the pulse sequence were proportionally increased (doubled). Frequency of cavitation and vessel rupture was reduced, and more successful coagulation was observed in CAM venules after reducing the peak laser power while keeping the radiant energy fixed.

In addition to major factors, including visco-velocity profiles and shear stress, another source of discrepancy between photocoagulation of smaller and larger blood vessels may be the relative size between the beam spot and blood vessel diameters. Differences in CAM vascular networks of an arteriole versus venule: As discussed

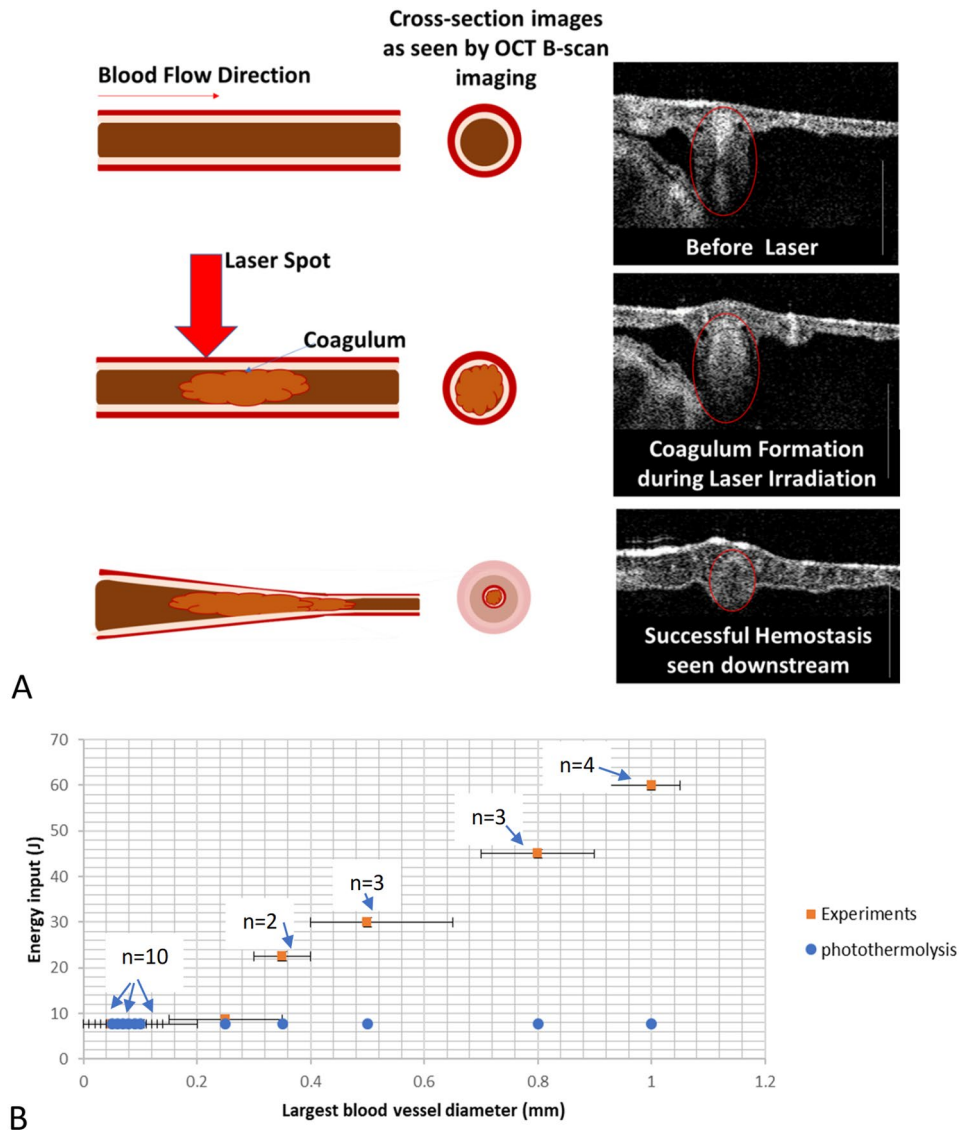


Figure 4. (A) Cross-sectional OCT monitoring of laser ($1.07\ \mu\text{m}$) irradiation of CAM blood vessels. Downstream imaging across a blood vessel shows initial formation of the coagulum during laser irradiation, and after successful hemostasis, the blood vessel collapses downstream with no observable flow. Scale bar is $500\ \mu\text{m}$. (B) Radiant pulse energy ($1.07\ \mu\text{m}$) determination with increasing blood vessel diameter in CAM arterioles for a $3\ \text{mm}$ spot size. The number of trials at each vessel diameter is highlighted in the image with an 'n' number of trials/eggs. Blue circles indicate pulse energy computed from the theory of selective pulsed photothermolysis.

earlier, although optical absorption in arterial vs venous blood is different (venules appear bright red upon visual inspection and carry oxygenated blood in CAMs, while arterioles carry deoxy hemoglobin, which is juxtaposed in most tissues⁴³), arterioles have a higher likelihood of coagulation than venules, as reported in previous CAM studies^{40,49} using $585\ \text{nm}$ irradiation. In the CAM vasculature, blood vessels develop in artery/vein pairs⁴³, as observed in experiments reported above (Fig. 3, corresponding enface Fig. S10). In this case, the suspected arteriole underwent successful hemostasis, while blood flow was observed in the venule post irradiation. Beam Size Limitations: Compared to capillaries and smaller blood vessels, the length to volume ratio exposed to laser irradiation is significantly different for larger blood vessels. Time duration of Arrhenius damage: Arrhenius damage threshold temperatures and corresponding time durations vary in blood vessels compared to surrounding tissue and the collagenous arterial vessel wall⁴¹. This suggests that the time variation of temperature for achieving hemostasis has a complex dependence on these factors given that the condition for success is Arrhenius damage in these tissues (blood, arterial collagen). Successful coagulation of vessels larger than $150\ \mu\text{m}$ is challenging when laser dosimetry is derived from the theory of selective photothermolysis^{37,41,42}.

Laser coagulation of blood vessels with large cross-sections and corresponding high blood flow rates was investigated in this study. A near infrared fiber laser ($1.07\ \mu\text{m}$) was utilized to investigate blood vessel coagulation. The existing literature reports numerous studies of blood vessel coagulation in the treatment of pediatric

vascular lesions, such as port-wine stains for microvascular hemostasis using pulse dye lasers. In addition, infrared lasers have also been successfully utilized in the long pulse regime for performing hemostasis equally well or better than pulsed dye lasers^{34–36,41,42}. Laser (1.07 μm) dosimetry for coagulation of large diameter vessels with high flow rates was investigated using the CAM vascular model and highlights the limitations of our current understanding. Limitations of the theory of selective photothermolysis for hemostasis of large blood vessels with high flow rates were investigated, suggesting a hypothesis that current limitations originate from high shear stress variations in large blood vessels along with temperature-dependent viscosity and velocity changes. FEMs that include fluid flow and thermal diffusion were developed that aid in contextualizing and testing limitations in hemostasis resulting from laser irradiation. Fluence rate parameters were then adjusted (not only accounting for some of the recommendations^{25,44} involving Monte Carlo simulations to avoid explosive vaporization pockets during irradiation) based on experimental observations utilizing OCT angiography (OCTA), resulting in consistent coagulation.

Future work that may improve the methodology is to study coagulum formation and develop an FEM model with a frontier tracking methodology⁵⁰. A visual inspection of the coagulation site during laser irradiation shows an accumulation of the coagulum in the direction of the flow. This effect may be due to the reduced density making the coagulum flow faster relative to surrounding uncoagulated blood causing the frontier of coagulum formation to have a skewed shape in the flow direction, which could be simulated with some of the frontier tracking methods introduced and discussed by Tryggvason et al. Additionally, considering the asymmetry between the leading and lagging fronts of the coagulum and changes in visco-velocity profiles, in future work, a two-beam approach where the leading edge of the blood flow experiences higher power photocoagulation irradiation whereas the lagging edge is irradiated with subthreshold shear velocity modifying radiation may help improve photocoagulation and achieve successful and consistent hemostasis. Additionally, this could be achieved by applying mechanical pressure to reduce the diameter of the vessel, similar to current bipolar clamping devices, to coagulate vessels in the 2–8 mm diameter range^{39,51,52}. Another limitation in coagulation of blood vessels with an incident collimated beam is the concentration of fluence on the side closer to the irradiation, sometimes causing explosive cavitation resulting in rupture of large blood vessels. This limits the average power input into blood vessels. A dynamic focusing methodology may be explored to focus the light according to a depth-resolved OCT image to cause uniform heating accounting for scattering properties at NIR wavelengths, thus improving coagulation success for larger sized (2–5 mm) blood vessels where currently clamping devices have been utilized^{51,52}. To model these effects, a frontier tracking methodology that has been utilized in predicting the performance of molten metal solidification⁵⁰ can provide additional insight into the hemostasis process. This methodology hypothesized and successfully tested the effects of the velocity profile along the solidification phase front along with the effects of shear stress. An FEM incorporating a frontier tracking approach along with the viscosity and flow dynamics incorporated in this report can provide a more accurate description of the coagulation/hemostasis process. Another potential limitation of the FEM model is incorporation of viscosity dependence utilized as a direct relationship between viscosity and temperature directly derived from published experimental results³³. The correlation between temperature and viscosity (Fig. 1B,C) is observed at the vessel wall with reduction in viscosity and temperature due to thermal diffusion. In reality, the viscosity may reach a steady state value and a frontier tracking methodology⁵⁰ may aid in incorporating any potential hysteresis involved in the relationship of viscosity with temperature. Additionally, another potential limitation is that a flat top beam was simulated versus non-flat top (gaussian-like) beam intensity profile utilized in experiments. Finally, given that cautery tools are able to coagulate up to 7 mm diameter blood vessels, further experiments are required to study the possibility of laser coagulation of blood vessels (e.g., up to 7 mm diameter). Finally, if the laser induced coagulum were to fail, the coagulated blood in larger diameter blood vessels (500 μm) could result in complications such as an embolism. Future studies are needed in larger blood vessels (> 3 mm) potentially using a larger animal like a rabbit femoral model.

Materials and methods

The theory of selective photothermolysis, in the context of blood vessel coagulation, suggests that pulses of light with a wavelength targeted for absorption by hemoglobin with appropriate pulse duration and fluence can heat blood vessels to achieve coagulation and hemostasis without causing nonspecific damage to surrounding tissues^{1,6,53}. In the theory of selective photothermolysis, the duration of pulsed laser exposure is governed by the thermal relaxation time of the target vessel. For vascular coagulation, the blood vessel thermal relaxation time varies with the square of the lumen diameter (D), Eq. (1) prescribes a quadratic relationship between vessel thermal relaxation time and blood vessel diameter with an inverse relationship to thermal diffusivity. From prior art^{54–56}, a typical value for thermal diffusivity (α) is 0.15 mm^2/s in relation to Eq. (1). Vessels of larger size (e.g., millimeter sized) diffuse heat slower into surrounding tissues compared to smaller sized structures. In the theory of pulsed photothermolysis, convective heat transfer due to blood flow through the laser irradiation spot is ignored. This assumption is justified by the significantly shorter laser pulse duration compared to the duration of blood flow through the irradiation spot—the blood is viewed as stationary during laser irradiation. Constraint on laser pulse duration in Eq. (1) is designed to ensure substantial confinement of deposited heat during irradiation.

The temperature increase to coagulate target blood vessels is governed by time–temperature first-order rate kinetics given by the Arrhenius thermal damage integral^{25,44,57}. The temperature increase (ΔT) is governed by the absorption coefficient (after accounting for the fluence distribution by scattering and absorption to surrounding tissue regions^{58,59}, Eq. (2)). The temperature increase (ΔT) from the absorption of pulsed laser radiation can be computed using the absorption coefficient in blood (μ_a) as given in Eq. (2).

$$\text{Thermal relaxation time (s)} = \frac{D^2}{16\alpha} \quad (1)$$

$$\Delta T = \mu_a \frac{\Phi}{\rho C} \quad (2)$$

Here, μ_a corresponds to the absorption coefficient of blood at 1.07 μm (approximately 0.3 mm^{-1}), and fluence (Φ , J/cm^2) is computed for a given laser spot size (e.g., 3 mm here). ρ and C are the density and specific heat (product yields $0.004 \text{ J}/\text{mm}^3/\text{K}$). Yb fiber laser pulse durations and peak powers were constrained to limit the increase in blood temperature to greater than 55°C . The peak power limit on the Yb fiber-laser module utilized in the study (see supplementary 2.1) was 3000 W (10% max. duty cycle, 300 W Avg.), the pulse duration was fixed at 50 μs , and the repetition rate was adjusted to account for the 10% duty cycle limitation (2000 Hz). If a large vessel receives too much radiant energy (to cause local temperature $T > 100^\circ\text{C}$) over a short pulse duration, the blood vessel can rupture, as water in the top of the vessel can create a vapor bubble and possibly burst the vessel wall, resulting in hemorrhage. Conversely, if a small diameter blood vessel absorbs radiant energy through a low power pulse over a longer period of time, the vessel will not coagulate as heat will have diffused out of the lumen without hemostasis. In the theory of selective photothermolysis, the required laser dosimetry is determined by the requisite pulse energy and thermal relaxation time (Fig. S4).

An FEM was developed to investigate the observed limitations when coagulating large diameter blood vessels. COMSOL FEM simulation software allows for concurrent mass and heat transfer simulations in materials encompassing these domains. A multiphysics model consisting of individual modules for heat transfer in liquids and mass transfer is proposed to investigate limitations observed in experiments targeting large diameter blood vessels for laser hemostasis.

To investigate the temperature distribution and fluid transfer of light-generated heat in vessels with fast-flowing blood, a 3D FEM was constructed. A COMSOL heat transfer module was utilized to compute the thermal relaxation in a cylindrical blood vessel, with the surrounding media being tissue. The COMSOL Multiphysics simulation incorporated heat transfer (ht) in fluids, flow (spf) and the bioheat transfer model (ht2). Multiphysics simulations were run in transient mode in a 3D coordinate system. Flow was incorporated into the model via the fluid laminar flow module linked with the heat transfer module via COMSOL Multiphysics. The peak value of the parabolic flow distribution was derived from experimentally measured flows (e.g., approx. 40 mm/s peak velocity for 400 μm blood vessels)⁴³. In addition, temperature-dependent viscosity was modeled by parametrically changing the temperature from data reported previously³³. Input to the 3D FEM was absorbed fluence determined from Monte Carlo simulations (see supplementary Sect. 3). Second, to reduce the computation time for a 3D model, a symmetric 2D FEM in COMSOL was used. 3D model incorporating blood flow and thermal diffusion with optical fluence input resulted in long simulation times (> hours) to converge whereas a 2D FEM model allowed for simulation in a few minutes allowing for multiple time scale iterations. Different sized blood vessels and their corresponding parabolic velocity profiles were input into the model from prior reports^{32,43}. For the flow (spf) simulation, dynamic viscosity of blood was allowed to vary with temperature based on experimentally obtained viscosity values³³. According to the experiment and considered model in this study, the dynamic viscosity reduces up to 55°C before increasing as determined through rheology measurements reported previously³³. An Arrhenius kinetic model of protein coagulation was added to the model to include the respective parameters corresponding to blood inside the lumen and tissue outside the lumen.

OCT angiography images were recorded of CAM blood vessels in conjunction with irradiation by a Yb fiber laser (1070 nm) coaligned bench-top system (see supplementary Sect. 2). A histogram of CAM blood vessel diameters was constructed from recorded angiography images to derive a laser irradiation protocol (see supplementary Sect. 2.2) for the CAM vasculature. Additionally, OCT images were recorded during laser coagulation of larger CAM blood vessels in response to the proposed enhanced stacked pulse irradiation scheme. In all CAMs that were imaged and irradiated ($n = 15$), pre- and post- irradiation images were recorded with OCT angiography. All the irradiation experiments were carried out at approximately day 8 of the gestation period and all methods were carried out in accordance with relevant guidelines and regulations (please see supplementary methods section for detailed overview of the CAM experiment protocols).

Received: 24 October 2021; Accepted: 29 April 2022

Published online: 19 May 2022

References

- Anderson, R. R. & Parrish, J. A. Microvasculature can be selectively damaged using dye lasers: A basic theory and experimental evidence in human skin. *Lasers Surg. Med.* **1**, 263–276 (1981).
- Lanigan, S. W. & Taibjee, S. M. Recent advances in laser treatment of port-wine stains. *Br. J. Dermatol.* <https://doi.org/10.1111/j.1365-2133.2004.06163.x> (2004).
- Tunnell, J. W. *et al.* Effects of cryogen spray cooling and high radiant exposures on selective vascular injury during laser irradiation of human skin. *Arch. Dermatol.* **139**, 743 (2003).
- Barton, J. K. *et al.* Photothermal coagulation of blood vessels: A comparison of high-speed optical coherence tomography and numerical modelling. *Phys. Med. Biol.* **46**, 1665–1678 (2001).
- Kimel, S. *et al.* Differential Vascular Response to Laser Photothermolysis. *J. Invest. Dermatol.* **103**, 693–700 (1994).
- Anderson, R. R., Jaenicke, K. F. & Parrish, J. A. Mechanisms of selective vascular changes caused by dye lasers. *Lasers Surg. Med.* **3**, 211–215 (1983).
- Garden, J. M., Polla, L. L. & Tan, O. T. The treatment of port-wine stains by the pulsed dye laser: Analysis of pulse duration and long-term therapy. *Arch. Dermatol.* **124**, 889–896 (1988).

8. Pratt, A. G. Birthmarks in infants. *AMA. Arch. Derm. Syphilol.* **67**, 302–305 (1953).
9. Lanigan, S. W., Cotterill, J. A. Port wine stains and treatment. *Bioengineering of the Skin: Cutaneous Blood Flow and Erythema* **2**, 183 (1994).
10. Burton, B. K., Schulz, C. J., Angle, B. & Burd, L. I. An increased incidence of haemangiomas in infants born following chorionic villus sampling (CVS). *Prenat. Diagn.* **15**, 209–214 (1995).
11. Iacopino, D. G. Hemostasis in brain tumor surgery using the Aquamantys system. *Med. Sci. Monit.* **20**, 538–543 (2014).
12. Hammond, J. S., Muirhead, W., Zaitoun, A. M., Cameron, I. C. & Lobo, D. N. Comparison of liver parenchymal ablation and tissue necrosis in a cadaveric bovine model using the Harmonic Scalpel™, the LigaSure™, the Cavitron Ultrasonic Surgical Aspirator® and the Aquamantys® devices. *HPB* **14**, 828–832 (2012).
13. Yoshida, S. *et al.* A morphological study of the blood vessels associated with periodontal probing depth in human gingival tissue. *Okajimas Folia Anatomica Japonica* **88**, 103–109 (2011).
14. Romanos, G. E. Diode laser soft-tissue surgery: Advancements aimed at consistent cutting, improved clinical outcomes. *Compend. Contin. Educ. Dent.* **34**, 752–7 (2013).
15. Vitruk, P. & Levine, R. Hemostasis and coagulation with ablative soft-tissue dental lasers and hot-tip devices. *Insid. Dent.* **12**, 1–4 (2016).
16. Halak, F. *et al.* Immediate laser-induced hemostasis in anticoagulated rats subjected to oral soft tissue surgery: A double-blind study. *Braz. Oral Res.* **32**, 1–8 (2018).
17. Hale, G. M. & Querry, M. R. Optical constants of water in the 200-nm to 200- μ m wavelength region. *Appl. Opt.* **12**, 555 (1973).
18. Jacques, S. L. Optical properties of biological tissues: A review. *Phys. Med. Biol.* **58**, 37–61 (2013).
19. Giglio, N. C. & Fried, N. M. Computational simulations for infrared laser sealing and cutting of blood vessels. *IEEE J. Sel. Top. Quantum Electron Publ. IEEE Lasers Electro Opt. Soc.* **27**, 1–8 (2021).
20. Giglio, N. C. *et al.* Rapid sealing and cutting of porcine blood vessels, ex vivo, using a high-power, 1470-nm diode laser. *J. Biomed. Opt.* **19**, 38002 (2014).
21. Katta, N. *et al.* Laser brain cancer surgery in a xenograft model guided by optical coherence tomography. *Theranostics* **9**, 3555–3564 (2019).
22. Katta, N., Estrada, A. D., McErloy, A. B. & Milner, T. E. Fiber-laser platform for precision brain surgery. *Biomed. Opt. Express* **13**, 1985–1994 (2022).
23. Katta, N., McElroy, A. B., Estrada, A. D. & Milner, T. E. Optical coherence tomography image-guided smart laser knife for surgery. *Lasers Surg. Med.* **50**, 202–212 (2017).
24. Nelson, J. S., Kelly, K. M., Zhao, Y. & Chen, Z. Imaging blood flow in human port-wine stain in situ and in real time using optical doppler tomography. *Arch. Dermatol.* **137**, 741 (2001).
25. Tunnell, J. W. Selective vascular injury during cutaneous laser therapy. Proceedings of the Second Joint 24th Annual Conference and the Annual Fall Meeting of the Biomedical Engineering Society, *Engineering in Medicine and Biology* (2002).
26. Barsky, S. H., Rosen, S., Geer, D. E. & Noe, J. M. The nature and evolution of port wine stains: A computer-assisted study. *J. Invest. Dermatol.* **74**, 154–157 (1980).
27. Kaufmann, R. & Hibst, R. Pulsed erbium:YAG laser ablation in cutaneous surgery. *Lasers Surg. Med.* **19**, 324–330 (1996).
28. Vogel, A. & Venugopalan, V. Mechanisms of pulsed laser ablation of biological tissues. *Chem. Rev.* **103**, 577 (2003).
29. Tawaza, H. Measurement of respiratory parameters in blood of chicken embryo. *J. Appl. Physiol.* **30**, 17 (1971).
30. Segura, P. N. T. The chicken chorioallantoic membrane model in biology, medicine and bioengineering the chicken chorioallantoic membrane model in biology, medicine and bioengineering. *Angiogenesis* <https://doi.org/10.1007/s10456-014-9440-7> (2014).
31. Honda, N. *et al.* Optical properties of tumor tissues grown on the chorioallantoic membrane of chicken eggs : tumor model to assay of tumor response to photodynamic therapy. *J. Biomed. Opt.* **20**, 125001 (2015).
32. Nadort, A., Kalk, K., Van Leeuwen, T. G. & Faber, D. J. Quantitative blood flow velocity imaging using laser speckle flowmetry. *Sci. Rep.* <https://doi.org/10.1038/srep25258> (2016).
33. Lecomte, B. Y. P., Nooy, D. U. & York, N. The viscosity of blood serum as a function of temperature. *J. Gen. Physiol.* **12**(3), 363–377 (1929).
34. Babilas, P., Schreml, S., Eames, T. & Hohenleutner, U. Split-face comparison of intense pulsed light with short- and long-pulsed dye lasers for the treatment of port-wine stains. *Lasers Surg. Med.* **727**, 720–727 (2010).
35. Date, P. Confocal microscopy study of nerves and blood vessels in untreated and treated port wine stains : Preliminary observations. *Dermatol. Surg.* <https://doi.org/10.1111/j.1524-4725.2004.30259.x> (2004).
36. Drosner, M., Stockmeier, M., Gatty, F. & Hellbru, G. Comparison of intense pulsed light (IPL) and pulsed dye laser (PDL) in port-wine stain treatment. *Med. Laser Appl.* **23**, 133–140 (2008).
37. Parlette, E. C. *et al.* Optimal pulse durations for the treatment of leg telangiectasias with a neodymium YAG Laser. *Lasers Surg. Med. Off. J. Am. Soc. Laser Med. Surg.* **105**, 98–105 (2006).
38. Wallwiener, C. W. *et al.* Bipolar vessel sealing : Instrument contamination and wear have little effect on seal quality and success in a porcine in vitro model. *Langenbeck's Arch. Surg.* <https://doi.org/10.1007/s00423-014-1234-2> (2014).
39. Ikami, T. M., Anibuchi, M. W. & Ikumi, N. M. Bumping phenomenon during continuous coagulation with bipolar forceps. *Neurologia Medico-Chirurgica* **52**, 731–735 (2012).
40. Kimel, S., Svaasand, L. O., Cao, D., Hammer-Wilson, M. J. & Nelson, J. S. Vascular response to laser photothermolysis as a function of pulse duration, vessel type, and diameter: Implications for port wine stain laser therapy. *Lasers Surg. Med.* **30**, 160–169 (2002).
41. Murphy, M. J. & Torstenson, P. A. Thermal relaxation times : An outdated concept in photothermal treatments. *Lasers Med. Sci.* <https://doi.org/10.1007/s10103-013-1445-8> (2014).
42. Lanigan, S. W. Port-wine stains unresponsive to pulsed dye laser : Explanations and solutions. *Br. J. Dermatol.* **139**, 173–177 (1998).
43. Maibier, M. *et al.* Structure and hemodynamics of vascular networks in the chorioallantoic membrane of the chicken. *Am. J. Physiol. Hear. Circ. Physiol.* <https://doi.org/10.1152/ajpheart.00786.2015> (2016).
44. Tunnell, J. W., Wang, L. V. & Anvari, B. Optimum pulse duration and radiant exposure for vascular laser therapy of dark port-wine skin : A theoretical study. *Appl. Opt.* **42**, 1367–1378 (2003).
45. Barozzi, G. S. & Dumas, A. Convective heat transfer coefficients in the circulation. *J. Biomech. Eng.* **113**, 308 (2008).
46. Consiglieri, L., dos Santos, I. & Haemmerich, D. Theoretical analysis of the heat convection coefficient in large vessels and the significance for thermal ablative therapies. *Phys. Med. Biol.* **48**, 4125–4134 (2003).
47. Black, J. F. & Barton, J. K. Chemical and structural changes in blood undergoing laser photocoagulation. *Photochem. Photobiol.* **80**, 89–97 (2004).
48. Black, J. F., Wade, N. & Barton, J. K. Mechanistic comparison of blood undergoing laser photocoagulation at 532 and 1,064 nm. *Lasers Surg. Med. Off. J. Am. Soc. Laser Med. Surg.* **165**, 155–165 (2005).
49. Kimel, S. *et al.* Differential vascular response to laser photothermolysis. *J. Invest. Dermatol.* <https://doi.org/10.1111/1523-1747.ep12398548> (1994).
50. Tryggvason, G. *et al.* A front-tracking method for the computations of multiphase flow. *J. Comput. Phys.* **759**, 708–759 (2001).
51. Harold, K. L. *et al.* Comparison of ultrasonic energy, bipolar thermal energy, and vascular clips for the hemostasis of small-, medium-, and large-sized arteries. *Surg. Endosc. Other Interv. Tech.* **17**, 1228–1230 (2003).
52. Chastagner, M. W., Miller, S. F., Shih, A. J. & Geiger, J. D. Vessel sealing using the bipolar electrosurgical method. In *International Manufacturing Science and Engineering Conference* 673–680 (2009). <https://doi.org/10.1115/msec2007-31166>.

53. Anderson, R. R. *et al.* Selective photothermolysis of cutaneous pigmentation by Q-switched Nd: YAG laser pulses at 1064, 532 and 355 nm. *J. Invest. Dermatol.* **93**, 28–32 (2004).
54. Friebel, M. Determination of optical properties of human blood in the spectral range 250 to 1100 nm using Monte Carlo simulations with hematocrit-dependent effective. *J. Biomed. Opt.* **11**, 1–10 (2006).
55. Friebel, M. Influence of oxygen saturation on the optical scattering properties of human red blood cells in the spectral range 250 to 2000 nm. *J. Biomed. Opt.* **14**, 1–6 (2009).
56. Meinke, M., Müller, G., Helfmann, J. & Friebel, M. Empirical model functions to calculate hematocrit-dependent optical properties of human blood. *Appl. Opt.* **46**, 1742 (2007).
57. Chae, Y., Aguilar, G., Lavernia, E. J. & Wong, B. J. F. Characterization of temperature dependent mechanical behavior of cartilage. *Charact. Temp. Depend. Mech. Behav. Cartil.* **278**, 271–278 (2003).
58. Shafirstein, G., Buckmiller, L. M. & Waner, M. Mathematical modeling of selective photothermolysis to aid the treatment of vascular malformations and hemangioma with pulsed dye laser. *Lasers Med. Sci.* **22**, 111–118. <https://doi.org/10.1007/s10103-006-0427-5> (2007).
59. Dai, T., Pikkula, B. M., Tunnell, J. W., Chang, D. W. & Anvari, B. Thermal response of human skin epidermis to 595-nm laser irradiation at high incident dosages and long pulse durations in conjunction with cryogen spray cooling : An ex-vivo study. *Lasers Surg. Med. Off. J. Am. Soc. Laser Med. Surg.* **24**, 16–24 (2003).

Acknowledgements

The authors would like to thank the summer research BME CuRES Cancer REU program for their support to undergraduate students under the mentorship of Dr. Mia K. Markey and Dr. Laura Suggs. Additionally, the authors would like to thank the Cancer Prevention Research Institute of Texas for providing funding for this research. The authors would like to thank Dr. James W. Tunnell for his insights into this work and suggestions. The authors would also like to thank Dr. Michael R. Gardner for his mentorship to Donovan and Mohsin in conducting experiments. Additionally, Drs. Bharadwaj Muralidharan and Farah Andleeb also helped guide the undergraduate students in their experiments. Other undergraduate students the authors would like to acknowledge: Kevin Choy, Anagata (Annie) Anand and Shivani R Kalavala from the Biomedical Engineering Department at the University of Texas at Austin.

Author contributions

N.K. contributed to the experimental design, COMSOL simulations, laser assembly, testing, and manuscript preparation. D.S. contributed to COMSOL simulations, A.M.B. contributed to software design of the pulsed stack profile along with near real-time OCTA image processing, A.D.E. contributed in experimental design, testing and G.D. contributed to data analysis, M.M. and M.D. contributed to the initial experiments and developing the chorioallantoic membrane protocol and preparation. T.E.M. was the principal investigator of the project, contributed to the experimental design and manuscript preparation and supervised all the other authors.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-022-12128-1>.

Correspondence and requests for materials should be addressed to N.K.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022