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# The Heritability of Behaviors Associated With the Host Gut Microbiota

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Manterola M, Palominos MF and Calixto A (2021) The Heritability of Behaviors Associated With the Host Gut Microbiota. Front. Immunol. 12:658551. doi: 10.3389/fimmu.2021.658551 What defines whether the interaction between environment and organism creates a genetic memory able to be transferred to subsequent generations? Bacteria and the products of their metabolism are the most ubiquitous biotic environments to which every living organism is exposed. Both microbiota and host establish a framework where environmental and genetic factors are integrated to produce adaptive life traits, some of which can be inherited. Thus, the interplay between host and microbe is a powerful model to study how phenotypic plasticity is inherited. Communication between host and microbe can occur through diverse molecules such as small RNAs (sRNAs) and the RNA interference machinery, which have emerged as mediators and carriers of heritable environmentally induced responses. Notwithstanding, it is still unclear how the organism integrates sRNA signaling between different tissues to orchestrate a systemic bacterially induced response that can be inherited. Here we discuss current evidence of heritability produced by the intestinal microbiota from several species. Neurons and gut are the sensing systems involved in transmitting changes through transcriptional and post-transcriptional modifications to the gonads. Germ cells express inflammatory receptors, and their development and function are regulated by host and bacterial metabolites and sRNAs thus suggesting that the dynamic interplay between host and microbe underlies the host's capacity to transmit heritable behaviors. We discuss how the host detects changes in the microbiota that can modulate germ cells genomic functions. We also explore the nature of the interactions that leave permanent or long-term memory in the host and propose mechanisms by which the microbiota can regulate the development and epigenetic reprogramming of germ cells, thus influencing the inheritance of the host. We highlight the vast contribution of the bacterivore nematode C. elegans and its commensal and pathogenic bacteria to the understanding on how behavioral adaptations can be inter and transgenerational inherited.

Keywords: transgenerational inheritance, germ cells, small RNAs, host-bacteria interactions, microbiota, behaviors

## INTRODUCTION

Microbes present in the environment are the most ubiquitous stimuli to which all organisms are exposed. Microbial ancestors likely shaped the evolution of eukaryotes, establishing mutual adaptation mechanisms for long-term survival and benefit (1). Microorganism and multicellular hosts communicate by a plethora of mechanisms that depend on the nature of their relationship, whether it is of commensalism, mutualism, or pathogenic (2). The outcome of a microbe-host relationship depends on their mutual interaction and cannot be established *a priori* based on the separate parts alone (3).

The recognition of the immediate environment is a key step in the establishment of short and long-term adaptive responses by all organisms. The duration, intensity, and nature of environmental stimuli are the parameters that influence the execution of behavior and its heritability. Brief homeostatic inputs are dealt with as part of the physiological response and likely do not leave traces in organisms' genetic and cellular memory. Conversely, exposure to persistent threats such as viral or bacterial pathogens generate heritable responses to infection in future generations (4–8).

## SENSING THE BIOTIC ENVIRONMENT AND THE CREATION OF A CHEMICAL MEMORY

The bacterivore nematode *Caenorhabditis elegans* is an exceptional model organism to study host microbiota relationships, and importantly for this review, the interactions that create a multigenerational memory. Nematodes show preference over their bacterial food, display olfactory associative learning (9), and develop transgenerationally inherited strategies to survive pathogenesis (5–8).

Microbes contribute essential metabolites and produce toxins that generate disease. Therefore, it becomes essential for the animal to discriminate between pathogenic versus nonpathogenic chemical environments to display appropriate behaviors. Specific olfactory neurons sense soluble and volatile chemicals from the biotic environment (10, 11) to mediate attraction or aversion to pathogenic and commensal bacteria (9, 12-14). Interestingly, a number of potentially pathogenic bacteria are attractive at first since they also produce attractive odorants (15) or contain metabolites of high nutritional value such as vitamin B12 (16). This poses a hard choice for the animal, having to prioritize escaping damage over the potential benefits of good nutrition such as the acceleration of development. Thus, the correct integration of different sensory neuronal circuits allows for the command of behaviors that likely prioritize survival. This learned behaviors are maximally effective if they can be transmitted to the following generations.

*C. elegans* avoids pathogens by activating different G proteincoupled receptors (GPCR) in specialized chemosensory neurons that detect secreted bacterial molecules and orchestrate adaptive behaviors like pathogen avoidance (9, 13, 17–19). Additionally, sensory neurons are able not only to coordinate behavior but also to orchestrate immune responses: The activation of the same GPCR, OCTR-1, in the chemo and thermosensory ASI neuron triggered avoidance, but in the ASH polymodal nociceptor caused a cascade that activates neuropeptidergic immune pathways (20). Sensory neurons thus play a key role in many diverse and complex processes related with pathogen responses, from avoidance and immunity to learning and memory formation.

The gastro-intestinal tract is home to microbes of an immense variety of genus (21, 22) and enteric sensory neurons located next to the intestine (23) can detect changes in the composition of microbe's metabolic byproducts (24-26). Diverse host's physiological responses have shown to be modulated by microbial metabolites, like short-chain fatty acids, polysaccharides, bile acids, and others (27), in a bidirectional communication process between host and microbe that is evolutionary conserved across animals (28). Chemical information about the intestinal luminal environment is transmitted to the CNS through the vagus nerve in mammals (29), and through the recurrent nerve in insects, like Drosophila (30). In C. elegans, sensory neurons detect microbial metabolites such as bacterial autoinducers [homoserine lactones (12), (S)-3-hydroxytridecan-4-one (15), and other virulence factors like phenazine and pyochelin (19). to coordinate behaviors like avoidance (12, 13, 19) and attraction (15). However, it is still unknown how the worm senses intestinal bacterial metabolites. Mammalian intestinal cells respond to bacterial short-chain fatty such as acetate and butyrate through the activation of GPCRs (31) thus suggesting that both neurons and intestinal cells may detect chemical changes in the intestinal lumen. But, how might environmental experiences, decoded by sensory neurons distant from the germline, reprogram the gamete's genetic memory?

In nematodes, the AWC and AWB chemosensory neurons coordinate sensing of pathogens and aversive olfactory learning (17). Recently, these sensory neurons have also been implicated in promoting intestinal p38 MAPK immune responses to pathogens (32). From the intestine, environmental information can be transgenerationally transmitted to the germ cells through inter-tissue regulation of the expression of ASH-2 and RBR-2 histone-modifiers (33). Taken together these raises the possibility that the transmission process involves communication between the brain, the gut and the germline. To add to this idea, the ASI ciliated chemosensory neuron involved in dauer formation (34, 35) and bacterial metabolite detection (19), controls transgenerational pathogen avoidance (6) and germline development (36, 37). These sensory neurons control germline development by coupling proliferation/differentiation (37) and apoptosis of the germline stem cell pool, with sensing of the environment's quality by the activation of the DAF-7/TGF- $\beta$  pathway in parental neurons and DAF-3/DAF-12 in the progeny (36). This suggests that neurons that are continually sensing and detecting changes in the external and internal environment may directly regulate such a delicate process like reproduction. However, how neurons, intestinal, and germ cells coordinate the information extracted from the environment to transmit a long-term behavioral memory associated with changes

in the repertoire of local microbe byproducts remain mostly unknown.

The length of parental exposure to pathogens modulates the behavior of future generations when exposed to those same stimuli. For example, the progeny of nematodes exposed to highly virulent pathogens for short periods of time are attracted to the bacteria, while progeny of parents exposed for more extended periods of time inherit pathogenic avoidance, intergenerationally (8). Is it known that the molecular effectors mediating the inheritance of this behavior are dependent on the endo-siRNAs pathway; however, it is not known yet the neuronal circuits involved in its coordination. In a related paradigm, the progeny of nematodes exposed for two generations to mildly virulent pathogens like P. aeruginosa PAO1 or Salmonella enterica (serovar Typhimurium) enter diapause forming a stress-resistant larva called dauer that does not feed. This response is not observed in the first generation, which implies that the molecular processes that underlie dauer formation under pathogenesis need to build up to a threshold. This depends on the accumulation of molecular damage signals that turn on defensive pathways like FOXO/DAF-16 (5, 38) and RNAidependent molecules (5, 7). Interestingly, a change in the pathogen's virulence or the animal immune status dramatically affects diapause entry (5). This shows that the process of inherited memory formation depends on the duration of the threat but also to the pathogen's virulence determinants. We propose that the inheritance of a pathogenic memory requires sensory inputs (e.g., olfactory cues and bacterially-derived metabolites), to be integrated with internal signaling factors. These factors include host immunity molecules, and the products of the local host-microbe interaction. Importantly, the memory of pathogenic encounters can be inherited by subsequent generations via mechanisms that involve RNA interference pathways (5, 6, 8). This supported the idea that these survival strategies against pathogens maintained for multiple generations in absence of the threat are inherited through mechanisms based on RNA molecules.

#### SMALL RNAS MEDIATE TRANSGENERATIONAL INHERITANCE TRIGGERED BY ENVIRONMENTAL CHANGES

sRNAs can effectively bridge soma and germline throughout generations. sRNAs are mediators of heritable epigenetic changes in the offspring (39) and are able to sustain intergenerational (40) and transgenerational (41) transmission. Among the heritable sRNAs are short-interference RNAs (siRNAs), microRNA (miRNA), PIWI-interacting RNA (piRNA), RNAs-derived from transfer RNAs (tsRNAs), ribosomal RNAs (rRNA), and circular RNA (circRNA). In germ cells, these sRNAs are key transcriptional and post-translational regulators and bind to Argonaute proteins, and Piwi-interacting RNAs (42, 43). In addition to mRNA silencing, RNA mediated alterations occur at chromatin level (43) and are known as RNA-triggered chromatin modifications. Endogenous siRNAs direct chromatin remodeling by inducing the methylation of specific genomic regions and promoting heterochromatin formation (44). In *C. elegans* and *Drosophila*, siRNAs trigger RNA interference (RNAi) pathways in germ cells that are amplified in the offspring for several generations (39, 45–47). Amplification of siRNAs is also essential for targeting specific genomic loci and inducing histone modifications, creating a footprint that is transgenerationally maintained for at least two generations (43, 48).

piRNAs are sRNAs that are highly expressed in germ cells and are involved in the maintenance of the genome and the initiation of multigenerational epigenetic inheritance. piRNAs expression have been also identified in somatic tissues such as follicle cells, hippocampus, kidney and liver (49) as well as in cancer cells (50, 51). piRNA induce heritable epigenetic modifications to silence specific loci in the genome (52-54) and mediate heritable responses to heat stress, toxicants, high-fat diet (54-59) and avoidance to pathogenic bacteria (6). miRNAs are essential regulators of female and male gametogenesis (60, 61), and embryonic development (62). Alterations in sperm miRNA content due to, for example, stress and metabolic changes, are intergenerationally inherited (63-65). In C. elegans miRNAs are highly sensitive to pathogenic exposure and mediate the initiation of transgenerational behaviors like pathogen induced-diapause (7). tsRNAs are enriched in male germ cells and known to inhibit translation, regulate transcription and chromatin modifications (66). tsRNAs can be methylated and have shown to mediate intergenerational transmission of metabolic disorders (67). circRNAs regulate transcription and splicing, modulate translation and post-translational modifications, and act as microRNA (miRNA) sponges (68, 69). circRNAs have been found in seminal plasma, suggesting that they can also mediate a soma-germ cell communication pathway (70). All the above suggests that small RNAs are sensible communicators of environmental variations such as changes in microbiota content. But, can the gut microbiota itself produce sRNAs that reach host tissues beyond the intestine? Or do they produce intestinal effects that propagate systemically? If they can reach the host tissues, do they work as initiators, executers or are they the memory required for behavioral outputs?

Gut microbiota influences siRNA expression in the intestine and brain, affecting host transcriptional reprogramming and raising the possibility that microbe-derived RNAs or byproducts communicate with somatic cells (71–79). In mice, several reports have showed that microbiota influences transcription of miRNAs in the amygdala and prefrontal cortex (73). Among these miRNAs, miR-183-5p and miR-182-5p (73) are of special interest as they are involved in stress- and fear-related responses (80, 81). Further to this, stress was previously shown to produce the accumulation of miRNA membrane vesicles in the epididymis and testes, thus transmitting an intergenerational response to stress in the offspring (82).

In *C. elegans*, neuron-specific synthesis of sRNAs regulates the amplification of endogenous siRNAs in germ cells, thus changing germline gene expression for multiple generations (83, 84). This mechanism opens the possibility that an olfactory stimulus elicited by bacteria changes the repertoire of endogenous neuronal sRNAs, which can later promote epigenetic changes in germ cells that will affect the offspring's behavior and fitness. To date, two sRNAs from bacteria have been identified that influence inherited behaviors in *C. elegans*. P11 from *P. aeruginosa* PA14 influences avoidance behavior (85), while the quorum sensing RNA RsmY from *P. aeruginosa* PAO1 is required for diapause formation under pathogenesis (86). How sRNAs from bacteria might be reaching the host soma and germline is discussed later.

#### TRANSPORT OF INTERSPECIES sRNAs BETWEEN HOST TISSUES

Can bacterial molecules such as RNAs use the host transport machinery to reach host tissues and eventually the germline? Much needs to be learned about the import of bacterial metabolites into host cells, and their selectivity. However, several mechanisms have been described that can accommodate such a travel.

There are dedicated mechanisms to export and uptake RNA molecules from other cells and the extracellular space. In C. elegans, selective receptors and RNA transporters participate in the systemic movement of RNA molecules (SID proteins). SID-1 is a conserved channel gated by double stranded RNA (dsRNA), ref (87), which allows the passage of dsRNA between C. elegans cells and also in heterologous systems (88). Its human homolog -SIDT1- is known to mediate bidirectional, sequence-specific and target-specific small RNA transfer between human cells (89). Other SIDs like SID-3, a conserved tyrosine kinase is required for import of dsRNA in worms (90). SID-5 is an endosomeassociated protein (91) with a role in embryonic parental RNAi in C. elegans (92). A relevant transporter in bacteria-worm communication is SID-2, which localizes to the intestinal lumen and potentially serves as a gate between host enterocytes and intestinal microbes. This protein is required for the import of ingested dsRNA, through an endocytosis-mediated and energy dependent mechanism (93). SID-2 shares functional similarities with Toll-like receptor 3, which in humans localizes to endosomes and recognizes dsRNA and sRNA (94). Defensive mechanisms such as pathogen-induced diapause formation (PIDF) require intact SID-dependent pathways for dsRNA import (5), opening up the possibility that the bacterial sRNA delivery system into the host uses a SID-dependent entry. It is however unknown how are bacterial RNAs found in the intestine. Are they secreted by selective bacterial exocytic mechanisms? Alternatively, can they be contained in membrane vesicles or exposed as cytosolic contents after the explosive stress-induced lysis of bacteria?

Bacterial Outer Membrane Vesicles (OMVs) are able to carry RNA among other cargoes (95–99). Increasing research has revealed the importance of OMVs RNA-cargo in bacteria-host interactions (99–101). Other types of bacterial vesicles are membrane vesicles produced by explosive lysis of both Grampositive and Gram-negative bacteria (102, 103). This, a seemingly stochastic process, generates MV containing specific cargoes. This raises the possibility that bacterial vesicles use the host endocytic pathway to deliver cargoes to specific tissues (104). To induce intergenerational silencing in *C. elegans*, injected dsRNA can be transported to the oocyte from the worms' body cavity within intracellular vesicles that depend on the LDL receptor superfamily homolog RME-2 (92). The existing molecular machinery regulating the inheritance of dsRNA through RME-2 (92, 105, 106) and the capacity of the bacteria to generate OMVs with RNA cargoes suggests that bacterial RNAs present in the intestinal lumen may enter the host cells by two separate but synergic mechanisms: endocytosis-dependent and endocytosis-independent pathways (like SIDs RNA transporters).

It has been shown that RNA can be transported from brain to the germline, and from there inherited to the embryos (107). This demonstrates that sRNAs are central players in soma to germline communication pathways. Within this interaction pathways, sRNAs can be transported through exosomal and non-exosomal transport via blood and follicular fluid to oocytes (108, 109), and via seminal and epididymal luminal fluid to sperms, breast milk, saliva, and cerebrospinal fluid (74, 110). sRNAs can also be loaded onto sperms by extracellular vesicles present in the outer membrane of epididymal cells (111, 112). Several of them are also transferred through RNA transporters. That is, in C. elegans, the RNA transporter SID-1 actively participates in transporting extracellular dsRNA into oocytes and in the inheritance of siRNAs (92). Therefore, RNA delivery is an active mechanism that allows germ cells to acquire information from somatic cells. We expect that future work will shed light onto questions regarding the origin, transport and systemic internalization of bacterial sRNA in the host.

#### EPIGENETIC INHERITANCE OF ENVIRONMENTAL EXPERIENCES: CAN BACTERIA MODIFY THE GERMLINE?

Any new environmental experience that affects individuals has the potential to modify the epigenome. When modifications in the chromatin state and in the RNA content occur in germ cells, the epigenetic memory could be transmitted across subsequent generations. A change in the epigenetic profile of germ cells is therefore a mandatory step to inheritance of environmental adaptations (111). Direct evidence shows that DNA methylation, histone modifications, histone variants, and non-coding RNAs transferred by germ cells can promote phenotypical adaptations to the offspring for several generations in the absence of the stimulus that initiated the response (111, 113, 114). But, can bacterial molecules cause such changes? Until our knowledge, there is no evidence showing a direct effect of gut bacteria over the germ cells. However, by considering that it is very hard to demonstrate this experimentally, we cannot exclude this possibility.

Although the mechanisms that link gut microbiota and the epigenetic modifications in germ cells are largely unknown, the

influence of bacteria in the host epigenome has been unequivocally demonstrated (115). Microbial signals including ncRNAs, metabolites and inflammatory molecules interact with histone writers such as DNA methyltransferases (DNMTs) and histone deacetylases (HDAC) or regulate the availability of writer's substrates such as SAM, the primary methyl donor for DNMT (115). Interestingly, at least histone modifications and sRNAs have been shown to be related to a potential gut bacterianeural- germ cell cross talk, highlighting the importance of gut microbiota in the inheritance of phenotypical traits (116, 117).

Recently, a link between gut microbiome, the epigenome of germ cells and the offspring's behavior has been suggested in *C. elegans.* Intestinal distention produced by bacteria induces acetylation in histone 4 in germ cells, particularly H4K8ac (116). This mark could be sufficient to inherit pathogen avoidance in the offspring, underlying the still unknown mechanism of crosstalk between sensorial neurons, brain, gut and germline to control transgenerational phenotypes. Furthermore, it raises the possibility that mechanical changes in the intestinal epithelium may also drive the production of host-derived metabolites and sRNAs, thus allowing inter-tissue and intergenerational transmission of information.

Histone Post-Translational Modifications (PTM), as well as histone-writers and reader proteins are critical regulators of the gene expression programs in both the germline and the zygote. From C. elegans to the mouse, several PTMs in sperms and oocytes including mono and di-methylations of histone H3 at lysine 4 (H3K4me1/me2) and trimethylation of histone 3 at lysine 9 (H3K9me3), are retained at genomic regulatory elements (118), imprinted regions and miRNA clusters (119, 120). Disruptions in the histone retention regions in sperms produced by environmental stimuli can transgenerationally persist up to the third generation (121-123). Although the evidence of bacterialinduced acetylation in germline epigenome needs to be further supported, the possibility that gut microbiota influence PTM in germ line provides a hint in how bacteria produce environmental changes that are sensed and inherited to the progeny. Whether gut bacteria directly affect PTMs in germ cells or it is a secondary effect after a mechanical or chemical stimulus needs to be further clarified.

#### BACTERIAL METABOLITES AND THEIR EFFECT ON BEHAVIOR

Bacteria are true metabolite factories, producing a large number of neurotransmitters, vitamins among other essential nutrients. It is clear that microbial products affect life history traits of the individuals carrying them. However, can bacterial metabolites such as amino acids, fatty acids and other, influence the progenies inter or transgenerationally? Naturally, whether it is directly or indirectly, the transformation leading to inheritance is stored in the germline. While the contribution of sRNAs in inheritance has been widely documented, transmission of life history traits by metabolites is less clear. Bacterial metabolites orchestrate epigenetic pathways: Inositol phosphate (IP3) influences histone acetylation in the intestine by regulating histone deacetylase 3 (HDAC3) activity (124). Other bacterial metabolites, like folate, had shown to regulate the proliferation of the germline and the fertility of *C. elegans* (125). Although the presence of bacterial genetic material or metabolites has not been directly observed in tissues other than the intestine, it is formally possible that they can reach the germline. Bacterial metabolites and sRNAs stimulate the soma-to germ line communication axis as well as immune responses that could modulate the epigenetic programing and even immune priming in germ cells (111, 126–130). The interactions between histone modifications, sRNAs and transmissible chromatin domains provide a framework in which metabolites may produce transgenerational epigenetic effects in gametes.

The absence of intestinal microbiota is well known to cause impairment in the development of the brain (131) and the enteric nervous system (132, 133). Bacteria can synthesize neurotransmitters like GABA, dopamine, serotonin, melatonin, histamine and acetylcholine (29, 134, 135), which posses' receptors in most animals. Germ-free mice showed reduced enteric contractibility (132) thus suggesting that maybe the lack of a bacterially produced neurotransmitter underlies the observed developmental abnormalities. It was in the late 80's that Minuk (136) suggested for first time that human mood can be modulated by bacterial metabolites: the loss of consciousness seen in patient with total septis or liver disease can be related with an increase in GABA production from pathogenic bacteria colonizing the bowel (136). Recently this idea has been retaken and defined as 'psychobiotics', probiotics with 'mind-altering' capacities (137). This new concept leads to think that bacterial metabolites can make their effect directly onto enteric neurons or intestinal epithelial cells to induce the release of a range of host modulatory molecules that can regulates diverse processes like immunity and behavior. Likewise, the intestinal bacterial metabolome of the honeybee specifically regulates the bee's nursing behavior and the host' circulating metabolites in the hemolymph/blood (138). How changes in the physiology provoked by altering the bacterial metabolome profile are integrated and transmitted to the progeny of complex hosts, like mammals, are yet remain to be elucidated.

## CYTOKINE SIGNALING AS A MECHANISM TO INFLUENCE TRANSGENERATIONAL INHERITANCE IN GERM CELLS

The intestinal biota and its metabolites not only modulate the brain but also the immune system (27, 139). This interaction is a dynamic process essential in maintaining homeostasis, which involves the recognition and tolerance against commensal microbiota, or the battle against pathogens. However, it is still unknown how changes in the immune system, produced by variations in the intestinal microbiota composition, may impact transgenerational memory formation. Research in *C. elegans* has recently shed light on the intergenerational maintenance of

microbe-induced immune transcriptional changes (6, 7). Thus, and similarly than the inheritance of the avoidance behavior, parental encounters with bacteria can lead to primed immune genes in germ cells that allow robust transcriptional responses against future infections in the offspring.

Cytokines are circulating factors produced by immune cells creating a communication pathway between the immune system and the brain, modifying brain function and behavior (140, 141). Furthermore, cytokines modulate mammalian germ cell development and differentiation, in both female and male gonad function. In females, macrophage secretion of TNF- $\alpha$ and interferon- $\gamma$  (IFN) stimulate the development and degeneration of the corpus luteum and, in so doing; regulate ovarian function (142). In the testis, Sertoli and Leydig cells produce numerous cytokines (like TNFα, IL1, IL6, and IL18), which act as a paracrine signal that regulates germ cell development and function (143-147). In macrophages, IFN induces chromatin remodeling and inherited transcriptional immune memory (148), and modifications of histone marks like H4ac, H3K9ac, and H3K4me3 at IFN-activated promoters (149). Although these data support the idea that regulation of the host's immune system controls some aspects of transgenerational inheritance, how microbial-induced cytokine production in the gut may modulate epigenetic changes in germ cells is still mostly unexplored. Future research will be invaluable in resolving how microbial-induced cytokine production influences germ cells and its impact on the host inheritance capacity.

## DISCUSSION

Adaptation to changing environments is essential for any living organism to survive and reproduce. Sensing environmental cues produces a memory that is inherited across several generations, granting short and long-term adaptive responses in all organisms. Inheritance of parental experiences enable behavioral responses in the offspring that can lead to faster recognition of pathogenic and non-pathogenic stimuli, even though individuals may have never been exposed before. Bacteria create a holobiont system with the host that regulates metabolism, immune responses and behavioral outcomes that influence transgenerationally-inherited strategies to survive pathogenesis. Bacteria and gut microbiota regulate different epigenetic mechanisms in both somatic and germ cells in the host modulating the genetic and epigenetic memory that is inherited across generations. To this end, bacteria modulate the gut-brain-germ cell axis to orchestrate and stimulate different pathways in which sRNAs carry most of the transmissible information. Different sRNAs are delivered to germ cells inducing chromatin changes that create particular genomic accessibility regions, which in turn regulate the gene expression program in the offspring (Figure 1). sRNAs are also amplified in germ cells by a RNAi machinery, allowing the transmission of the information throughout several generations. Although the mechanisms by which transgenerational changes are produced in germ cells are still poorly understood, histone modifications as well as endogenous and somatic sRNAs are involved in creating a transmissible memory in animals. Gut bacteria can also directly influence germ cells through bacteria-produced sRNAs and metabolites. Likewise, microbial-induced cytokines could induce epigenetic changes in germ cells that impact the phenotypical traits inherited by the host. Thus, gut bacteria influence the creation of an epigenetic memory in germ cells through indirect mechanisms *via* the gut-brain axis and sRNAs, but may also directly stimulate gametes through bacteria- derived sRNAs and metabolites.

The role of bacteria in the inheritance of behavior and memory for an organism is of great significance in understanding how phenotypic plasticity, adaptability and transgenerational inheritance occur in the species. Nevertheless, the precise mechanisms by which bacteria influence neuronal responses and reprogram the germline epigenome need to be unraveled. Understanding the mechanisms will allow us to better understand how bacteria coordinate the diversity of inherited behavioral outputs in the organisms and their adaptive responses.

#### **CONCLUDING REMARKS**

The dynamic relationship between host and microbe forms a higher-order organism known as the *holobiont* (150, 151). The continuous interaction between them influences each other at metabolic, physiological, and at genetic levels (**Figure 2**).

As mentioned before, we hypothesized that for memories to be transmitted multigenerationally a quantitative threshold must be reached. Once the threshold is exceeded, the information must be "stamped" on the germ cells in order to be heritable. Epigenetics emerged as the main mechanisms from where information about environmental changes (e.g. toxicants presence, high fat diet, osmotic and thermal stress, caloric restriction, among others) is transmitted from parents to subsequent generations (152, 153). In *C. elegans* small RNAs produced by the host (4, 7, 36, 83, 84, 154–156) and bacteria (85, 86, 157) are primary effectors for driving environmental changes are sensed and then transmitted to the progeny breaching the germline to soma barrier is still not fully understood.

In summary, in nematodes, priming or sensitivity of the progeny to a specific bacteria or bacterial byproduct is shaped by the previous generations experiences in a mechanism first decoded by sensory neurons, integrated by other neurons and intestinal cells, and then transmitted to the germ cells before fertilization. Additional studies are required to determine how bacterial host composition and microbe-derived byproducts are sensed by the neurons, integrated and precisely transmitted to the germline. However, neuronally produced small RNAs are essential to induce transgenerational gene silencing (83) and transgenerational behavioral memories (84) thus suggesting that the molecular signal accumulating over time is likely an RNA molecule. Future investigation will shed light on how the length and the virulence of the pathogen modulate the abundance of derived host and bacterial RNA molecules and the diversity of observed (and yet still undiscovered) inherited behavioral outputs in C. elegans (5, 8). However, in the

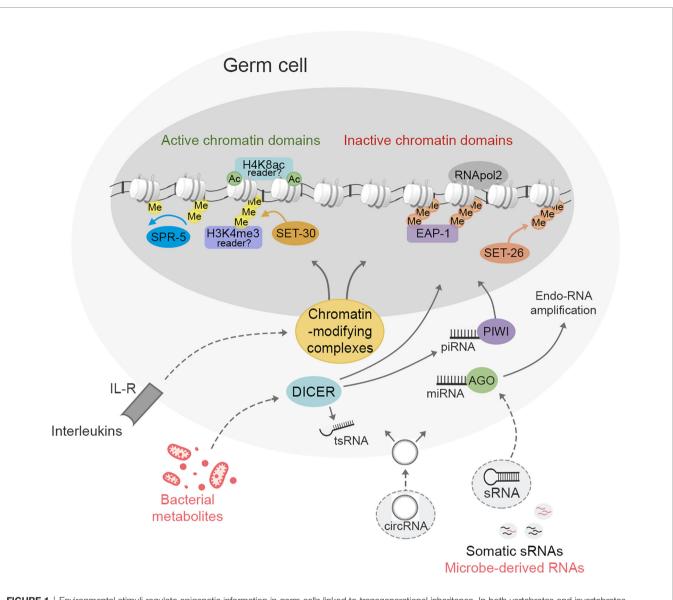
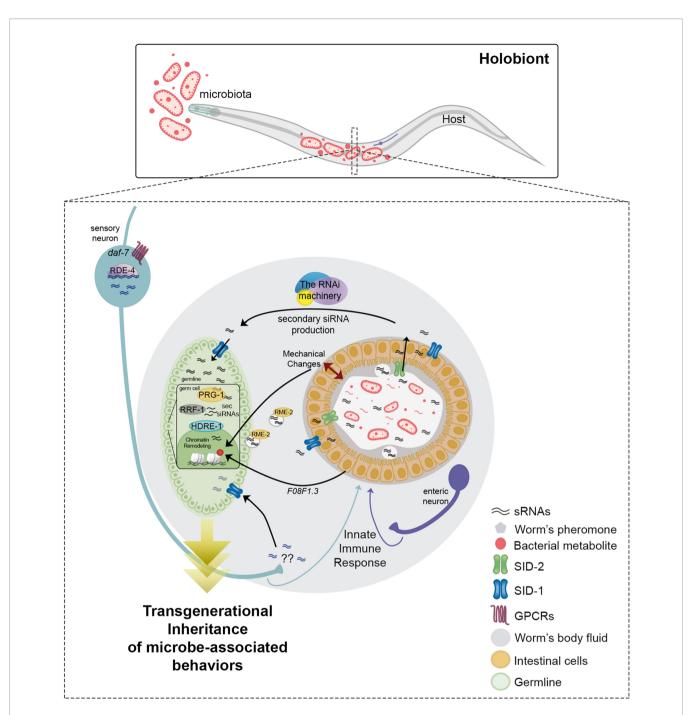


FIGURE 1 | Environmental stimuli regulate epigenetic information in germ cells linked to transgenerational inheritance. In both vertebrates and invertebrates epigenetic changes in the germline lead to intergenerational or transgenerational inheritance. In *C. elegans*, these changes involve chromatin remodeling through activation of the H3K4 tri-methyltransferase SET-30, H3K4 demethylase SPR-5, H3K9 tri-methyltransferase SET-26 and the reader of H3K9me3, EAP-1. The remaining chromatin remodeling mechanisms and the mammalian orthologous involved in the establishment of the active and inactive chromatin domains inherited to the offspring are still poorly known. sRNAs and their processing machinery are central players in germ cells to inherit phenotypical traits across generations. piRNAs and PWI proteins, miRNAs and AGO proteins, circRNAs and tsRNAs present in germ cells mediate transgenerational traits. sRNAs exert their effects via chromatin remodeling or through RNA amplification. The sRNA endonuclease DICER mediates the metabolism and amplification of sRNAs and the cleavage of tRNAs to produce tsRNAs. These RNAs are also epigenetic regulators, participate into RNA interference (RNAI) pathway, and directly inherits phenotypical traits across generations. Both chromatin remodeling processes and RNA pathways in germ cells are modulated by acquisition of smatic sRNAs during germ cell differentiation. It is also possible that bacteria-derived RNAs, bacterial metabolites and immune signals may also regulate transgenerational epigenetic marks in germ cells.

contrary, transgenerational epigenetic inheritances in mammalian systems are only beginning to be unraveled. Recent work led by scientists in Australia demonstrates that the progeny of mouse infected with the parasitic protozoan *Toxoplasma gondii* exhibited abnormalities in anxiety, working memory, object recognition, sociability and mating behaviors. Surprisingly, the behavioral abnormalities displayed by the F1 and F2 were correlated with alterations in the RNA profile of previously infected F0 male gametes (158), and surprisingly, RNA extracted from infected sperm recapitulates the behavioral effects of the paternal infection. Thus, and as the French biologist Jacques Monod once said "What is true for *E. coli* is true for the elephant", what is true for the worm it is also true for mouse and humans: we are just beginning to understand the importance of RNAs as key molecules in the evolution of the organisms as *holobionts*, a network of continuous dynamic interactions between and across the diverse biological levels of complexity that forms it.



**FIGURE 2** | Small RNAs regulate transgenerational inheritance of behaviors triggered by previous microbial experience in *C. elegans*. Neuron to-gut to-germline signaling is essential to induce the transmission of the memory of previous microbial encounters. Sensory neurons perceive bacterial metabolites through GPCRs, modulating the DAF-7/TFG-β signaling pathway. Neuronal RDE-4 is necessary for sRNA production involved in transgenerational control of behavior. Neuronal sRNAs (blue) are transferred into germ cells through the dsRNA transporter SID-1 to induce transgenerational silencing. It is still unknown how these neuronally-derived sRNAs are secreted into the worms' body. Sensory neurons (light blue) and neurons adjacent to the intestine (enteric; purple) control intestinal immunity against pathogens. Whether intestinal responses triggered by bacteria drive sRNAs changes is not known. Bacterial (pink) and host (black) sRNAs are transported from the lumen to the intestine through SID-2 and systemically through SID-1. RME-2 mediates the endocytosis of sRNAs from the worms' cavity to oocytes. The RNAi machinery amplifies sRNAs causing transgenerational silencing. In the germline, the Piwi protein PRG-1, the RNA helicase RRF-1, and the nuclear Argonaute HDRE-1 are essential for driving transgenerational changes by epigenetic modulation. The transmission of epigenetic information between intestine and germ cells is mediated by the transcriptional regulation of F08F1.3. Mechanical changes in the intestine triggered by bacterial colonization may induce intergenerational histone acetylation in the germline.

## **AUTHOR CONTRIBUTIONS**

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#### REFERENCES

- Abdel-Haq R, Schlachetzki JCM, Glass CK, Mazmanian SK. Microbiome-Microglia Connections Via the Gut-Brain Axis. *J Exp Med* (2019) 216(1):41– 59. doi: 10.1084/jem.20180794
- Abu Elhija M, Lunenfeld E, Eldar-Geva T, Huleihel M. Over-Expression of IL-18, ICE and IL-18 R in Testicular Tissue From Sexually Immature as Compared to Mature Mice. *Eur Cytokine Netw* (2008) 19(1):15–24. doi: 10.1684/ecn.2008.0116
- Agorastos A, Bozikas VP. Gut Microbiome and Adaptive Immunity in Schizophrenia. *Psychiatriki* (2019) 30(3):189–92. doi: 10.22365/ jpsych.2019.303.189
- Aho VTE, Pereira PAB, Voutilainen S, Paulin L, Pekkonen E, Auvinen P, et al. Gut Microbiota in Parkinson's Disease: Temporal Stability and Relations to Disease Progression. *EBioMedicine* (2019) 44:691–707. doi: 10.1016/j.ebiom.2019.05.064
- Alegado RA, King N. Bacterial Influences on Animal Origins. Cold Spring Harb Perspect Biol (2014) 6(11):a016162. doi: 10.1101/cshperspect.a016162
- Aravin AA, Hannon GJ. Small RNA Silencing Pathways in Germ and Stem Cells. Cold Spring Harb Symp Quant Biol (2008) 73:283–90. doi: 10.1101/sqb.2008.73.058
- Baj A, Moro E, Bistoletti M, Orlandi V, Crema F, Giaroni C. Glutamatergic Signaling Along The Microbiota-Gut-Brain Axis. *Int J Mol Sci* (2019) 20(6). doi: 10.3390/ijms20061482
- Bargmann CI, Hartwieg E, Horvitz HR. Odorant-Selective Genes and Neurons Mediate Olfaction in C. Elegans. *Cell* (1993) 74(3):515–27. doi: 10.1016/0092-8674(93)80053-h
- Bargmann CI, Horvitz HR. Control of Larval Development by Chemosensory Neurons in Caenorhabditis Elegans. Science (1991) 251(4998):1243–6. doi: 10.1126/science.2006412
- Beale E, Li G, Tan MW, Rumbaugh KP. Caenorhabditis Elegans Senses Bacterial Autoinducers. *Appl Environ Microbiol* (2006) 72(7):5135–7. doi: 10.1128/AEM.00611-06
- Belicard T, Jareosettasin P, Sarkies P. The piRNA Pathway Responds to Environmental Signals to Establish Intergenerational Adaptation to Stress. *BMC Biol* (2018) 16(1):103. doi: 10.1186/s12915-018-0571-y
- Ben Maamar M, Beck D, Nilsson E, McCarrey JR, Skinner MK. Developmental Origins of Transgenerational Sperm Histone Retention Following Ancestral Exposures. *Dev Biol* (2020) 465(1):31–45. doi: 10.1016/j.ydbio.2020.06.008
- Ben Maamar M, Sadler-Riggleman I, Beck D, McBirney M, Nilsson E, Klukovich R, et al. Alterations in Sperm DNA Methylation, non-Coding RNA Expression, and Histone Retention Mediate Vinclozolin-Induced Epigenetic Transgenerational Inheritance of Disease. *Environ Epigenet* (2018) 4(2):dvy010. doi: 10.1093/eep/dvy010
- Ben Maamar M, Sadler-Riggleman I, Beck D, Skinner MK. Epigenetic Transgenerational Inheritance of Altered Sperm Histone Retention Sites. *Sci Rep* (2018) 8(1):5308. doi: 10.1038/s41598-018-23612-y
- Brennecke J, Malone CD, Aravin AA, Sachidanandam R, Stark A, Hannon GJ. An Epigenetic Role for Maternally Inherited piRNAs in Transposon Silencing. Science (2008) 322(5906):1387–92. doi: 10.1126/science.1165171
- Buckley BA, Burkhart KB, Gu SG, Spracklin G, Kershner A, Fritz H, et al. A Nuclear Argonaute Promotes Multigenerational Epigenetic Inheritance and Germline Immortality. *Nature* (2012) 489(7416):447–51. doi: 10.1038/ nature11352

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- Camp JG, Kanther M, Semova I, Rawls JF. Patterns and Scales in Gastrointestinal Microbial Ecology. *Gastroenterology* (2009) 136(6):1989– 2002. doi: 10.1053/j.gastro.2009.02.075
- Cao X, Kajino-Sakamoto R, Doss A, Aballay A. Distinct Roles of Sensory Neurons in Mediating Pathogen Avoidance and Neuropeptide-Dependent Immune Regulation. *Cell Rep* (2017) 21(6):1442–51. doi: 10.1016/j.celrep.2017.10.050
- Cappucci U, Noro F, Casale AM, Fanti L, Berloco M, Alagia AA, et al. The Hsp70 Chaperone is a Major Player in Stress-Induced Transposable Element Activation. Proc Natl Acad Sci USA (2019) 116(36):17943–50. doi: 10.1073/ pnas.1903936116
- Casadevall A, Pirofski LA. The Damage-Response Framework of Microbial Pathogenesis. Nat Rev Microbiol (2003) 1:17–24. doi: 10.1038/nrmicro732
- Celluzzi A, Masotti A. How Our Other Genome Controls Our Epi-Genome. Trends Microbiol (2016) 24(10):777–87. doi: 10.1016/j.tim.2016.05.005
- Chaudhari SN, Mukherjee M, Vagasi AS, Bi G, Rahman MM, Nguyen CQ, et al. Bacterial Folates Provide an Exogenous Signal for C. Elegans Germline Stem Cell Proliferation. *Dev Cell* (2016) 38(1):33–46. doi: 10.1016/j. devcel.2016.06.013
- Chen JJ, Zeng BH, Li WW, Zhou CJ, Fan SH, Cheng K, et al. Effects of Gut Microbiota on the microRNA and mRNA Expression in the Hippocampus of Mice. *Behav Brain Res* (2017) 322(Pt A):34–41. doi: 10.1016/j.bbr.2017.01.021
- Choi HI, Kim M, Jeon J, Han JK, Kim KS. Overexpression of MicA Induces Production of OmpC-enriched Outer Membrane Vesicles That Protect Against Salmonella Challenge. *Biochem Biophys Res Commun* (2017) 490 (3):991–6. doi: 10.1016/j.bbrc.2017.06.152
- Chu C, Zhang YL, Yu L, Sharma S, Fei ZL, Drevet JR. Epididymal Small non-Coding RNA Studies: Progress Over the Past Decade. Andrology (2019) 7 (5):681–9. doi: 10.1111/andr.12639
- Collins J, Borojevic R, Verdu EF, Huizinga JD, Ratcliffe EM. Intestinal Microbiota Influence the Early Postnatal Development of the Enteric Nervous System. *Neurogastroenterol Motil* (2014) 26(1):98–107. doi: 10.1111/nmo.12236
- Dalfó D, Michaelson D, Hubbard EJ. Sensory Regulation of the C. Elegans Germline Through TGF-β-Dependent Signaling in the Niche. *Curr Biol* (2012) 22(8):712–9. doi: 10.1016/j.cub.2012.02.064
- Davis FM, Tsoi LC, Wasikowski R, denDekker A, Joshi A, Wilke C. Epigenetic Regulation of the PGE2 Pathway Modulates Macrophage Phenotype in Normal and Pathologic Wound Repair. *JCI Insight* (2020) 5 (17):e138443. doi: 10.1172/jci.insight.138443
- Daxinger L, Whitelaw E. Understanding Transgenerational Epigenetic Inheritance Via the Gametes in Mammals. *Nat Rev Genet* (2012) 13 (3):153-62. doi: 10.1038/nrg3188
- 30. de Castro Barbosa T, Ingerslev LR, Alm PS, Versteyhe S, Massart J, Rasmussen M, et al. High-Fat Diet Reprograms the Epigenome of Rat Spermatozoa and Transgenerationally Affects Metabolism of the Offspring. *Mol Metab* (2016) 5(3):184–97. doi: 10.1016/j.molmet.2015.12.002
- De Vadder F, Kovatcheva-Datchary P, Goncalves D, Vinera J, Zitoun C, Duchampt A, et al. Microbiota-Generated Metabolites Promote Metabolic Benefits Via Gut-Brain Neural Circuits. *Cell* (2014) 156(1-2):84–96. doi: 10.1016/j.cell.2013.12.016
- Desselberger U. The Mammalian Intestinal Microbiome: Composition, Interaction With the Immune System, Significance for Vaccine Efficacy, and Potential for Disease Therapy. *Pathogens* (2018) 7(3):57. doi: 10.3390/ pathogens7030057

- Devanapally S, Ravikumar S, Jose AM. Double-Stranded RNA Made in C. Elegans Neurons can Enter the Germline and Cause Transgenerational Gene Silencing. *Proc Natl Acad Sci USA* (2015) 112(7):2133–8. doi: 10.1073/ pnas.1423333112
- Dierking K, Pita L. Receptors Mediating Host-Microbiota Communication in the Metaorganism: The Invertebrate Perspective. *Front Immunol* (2020) 11:1251. doi: 10.3389/fimmu.2020.01251
- Dong TT, Yu Q, Qing XR, Ma XL, Dong WW, Shi J, et al. Potential Confounding Factors in Measurement of Specific Cell-Free Seminal mRNAs and microRNAs Derived From Human Reproductive Organs. *Andrology* (2016) 4(6):1010–9. doi: 10.1111/andr.12238
- 36. Dong WW, Li HM, Qing XR, Huang DH, Li HG. Identification and Characterization of Human Testis Derived Circular RNAs and Their Existence in Seminal Plasma. Sci Rep (2016) 6:39080. doi: 10.1038/srep39080
- Dupont C, Kappeler L, Saget S, Grandjean V, Levy R. Role of miRNA in the Transmission of Metabolic Diseases Associated With Paternal Diet-Induced Obesity. Front Genet (2019) 10:337. doi: 10.3389/fgene.2019.00337
- Elhassan MO, Christie J, Duxbury MS. Homo Sapiens Systemic RNA Interference-Defective-1 Transmembrane Family Member 1 (SIDT1) Protein Mediates Contact-Dependent Small RNA Transfer and microRNA-21-driven Chemoresistance. J Biol Chem (2012) 287(8):5267– 77. doi: 10.1074/jbc.M111.318865
- Engel P, Moran NA. The Gut Microbiota of Insects Diversity in Structure and Function. FEMS Microbiol Rev (2013) 37(5):699–735. doi: 10.1111/ 1574-6976.12025
- Evans EA, Kawli T, Tan MW. Pseudomonas Aeruginosa Suppresses Host Immunity by Activating the DAF-2 Insulin-Like Signaling Pathway in Caenorhabditis Elegans. *PloS Pathog* (2008) 4(10):e1000175. doi: 10.1371/ journal.ppat.1000175
- Fast I, Rosenkranz D. Temperature-Dependent Small RNA Expression in Drosophila Melanogaster. RNA Biol (2018) 15(3):308–13. doi: 10.1080/ 15476286.2018.1429881
- Feinberg EH, Hunter CP. Transport of dsRNA Into Cells by the Transmembrane Protein SID-1. Science (2003) 301(5639):1545-7. doi: 10.1126/science.1087117
- 43. Foong JPP, Hung LY, Poon S, Savidge TC, Bornstein JC. Early Life Interaction Between the Microbiota and the Enteric Nervous System. Am J Physiol Gastrointest Liver Physiol (2020) 319(5):G541–8. doi: 10.1152/ ajpgi.00288.2020
- Forsythe P, Kunze WA. Voices From Within: Gut Microbes and the CNS. Cell Mol Life Sci (2013) 70(1):55–69. doi: 10.1007/s00018-012-1028-z
- 45. Foster KJ, Cheesman HK, Liu P, Peterson ND, Anderson SM, Pukkila-Worley R. Innate Immunity in the C. elegans Intestine Is Programmed by a Neuronal Regulator of AWC Olfactory Neuron Development. *Cell Rep* (2020) 31(1):107478. doi: 10.1016/j.celrep.2020.03.042
- 46. Fullston T, Ohlsson-Teague EM, Print CG, Sandeman LY, Lane M. Sperm Microrna Content is Altered in a Mouse Model of Male Obesity, But the Same Suite of Micrornas Are Not Altered in Offspring's Sperm. *PloS One* (2016) 11(11):e0166076. doi: 10.1371/journal.pone.0166076
- Furness JB. The Enteric Nervous System and Neurogastroenterology. Nat Rev Gastroenterol Hepatol (2012) 9(5):286–94. doi: 10.1038/nrgastro.2012.32
- Gabaldon C, Legue M, Palominos MF, Verdugo L, Gutzwiller F, Calixto A. Intergenerational Pathogen-Induced Diapause in Caenorhabditis Elegans Is Modulated by Mir-243. *mBio* (2020) 11(5):e01950-20. doi: 10.1128/ mBio.01950-20
- Gracida X, Eckmann CR. Fertility and Germline Stem Cell Maintenance Under Different Diets Requires nhr-114/HNF4 in C. Elegans. *Curr Biol* (2013a) 23(7):607–13. doi: 10.1016/j.cub.2013.02.034
- Gracida X, Eckmann CR. Mind the Gut: Dietary Impact on Germline Stem Cells and Fertility. *Commun Integr Biol* (2013b) 6:e26004. doi: 10.4161/ cib.26004
- Grant B, Hirsh D. Receptor-Mediated Endocytosis in the Caenorhabditis Elegans Oocyte. *Mol Biol Cell* (1999) 10(12):4311–26. doi: 10.1091/ mbc.10.12.4311
- Greer EL, Beese-Sims SE, Brookes E, Spadafora R, Zhu Y, Rothbart SB, et al. A Histone Methylation Network Regulates Transgenerational Epigenetic Memory in C. Elegans. *Cell Rep* (2014) 7(1):113–26. doi: 10.1016/ j.celrep.2014.02.044

- Griggs EM, Young EJ, Rumbaugh G, Miller CA. MicroRNA-182 Regulates Amygdala-Dependent Memory Formation. J Neurosci (2013) 33(4):1734–40. doi: 10.1523/JNEUROSCI.2873-12.2013
- Grishok A. Biology and Mechanisms of Short Rnas in Caenorhabditis Elegans. Adv Genet (2013) 83:1–69. doi: 10.1016/B978-0-12-407675-4.00001-8
- Gu SG, Pak J, Guang S, Maniar JM, Kennedy S, Fire A. Amplification of siRNA in Caenorhabditis Elegans Generates a Transgenerational Sequence-Targeted Histone H3 Lysine 9 Methylation Footprint. *Nat Genet* (2012) 44 (2):157–64. doi: 10.1038/ng.1039
- Guo B, Li D, Du L, Zhu X. piRNAs: Biogenesis and Their Potential Roles in Cancer. Cancer Metastasis Rev (2020) 39(2):567–75. doi: 10.1007/s10555-020-09863-0
- 57. Ha HI, Hendricks M, Shen Y, Gabel CV, Fang-Yen C, Qin Y, et al. Functional Organization of a Neural Network for Aversive Olfactory Learning in Caenorhabditis Elegans. *Neuron* (2010) 68(6):1173–86. doi: 10.1016/j.neuron.2010.11.025
- Hammoud SS, Nix DA, Zhang H, Purwar J, Carrell DT, Cairns BR. Distinctive Chromatin in Human Sperm Packages Genes for Embryo Development. *Nature* (2009) 460(7254):473–8. doi: 10.1038/nature08162
- Hedger MP, Meinhardt A. Cytokines and the Immune-Testicular Axis. J Reprod Immunol (2003) 58(1):1–26. doi: 10.1016/s0165-0378(02)00060-8
- Hegde M, Guruprasad KP, Ramachandra L, Satyamoorthy K, Joshi MB. Interleukin-6-mediated Epigenetic Control of the VEGFR2 Gene Induces Disorganized Angiogenesis in Human Breast Tumors. J Biol Chem (2020) 295(34):12086–98. doi: 10.1074/jbc.RA120.012590
- Hinas A, Wright AJ, Hunter CP. SID-5 is an Endosome-Associated Protein Required for Efficient Systemic RNAi in C. Elegans. *Curr Biol* (2012) 22 (20):1938–43. doi: 10.1016/j.cub.2012.08.020
- Hoban AE, Stilling RM, M Moloney G, Moloney RD, Shanahan F, Dinan TG, et al. Microbial Regulation of microRNA Expression in the Amygdala and Prefrontal Cortex. *Microbiome* (2017) 5(1):102. doi: 10.1186/s40168-017-0321-3
- Hong C, Lalsiamthara J, Ren J, Sang Y, Aballay A. Microbial Colonization Induces Histone Acetylation Critical for Inherited Gut-Germline-Neural Signaling. *PloS Biol* (2021) 19(3):e3001169. doi: 10.1371/journal.pbio.3001169
- Houri-Ze'evi L, Rechavi O. Plastic Germline Reprogramming of Heritable Small RNAs Enables Maintenance or Erasure of Epigenetic Memories. *RNA Biol* (2016) 13(12):1212–7. doi: 10.1080/15476286.2016.1229732
- Houri-Zeevi L, Korem Kohanim Y, Antonova O, Rechavi O. Three Rules Explain Transgenerational Small RNA Inheritance in C. Elegans. *Cell* (2020) 182(5):1186–97.e12. doi: 10.1016/j.cell.2020.07.022
- Iyer LM, Aravind L, Coon SL, Klein DC, Koonin EV. Evolution of Cell-Cell Signaling in Animals: did Late Horizontal Gene Transfer From Bacteria Have a Role. *Trends Genet* (2004) 20(7):292–9. doi: 10.1016/j.tig.2004.05.007
- Jin X, Pokala N, Bargmann CI. Distinct Circuits for the Formation and Retrieval of an Imprinted Olfactory Memory. *Cell* (2016) 164(4):632–43. doi: 10.1016/j.cell.2016.01.007
- Jose AM, Kim YA, Leal-Ekman S, Hunter CP. Conserved Tyrosine Kinase Promotes the Import of Silencing RNA Into Caenorhabditis Elegans Cells. *Proc Natl Acad Sci USA* (2012) 109(36):14520–5. doi: 10.1073/ pnas.1201153109
- Kaletsky R, Moore RS, Vrla GD, Parsons LR, Gitai Z, Murphy CT. C. Elegans Interprets Bacterial non-Coding RNAs to Learn Pathogenic Avoidance. *Nature* (2020) 586(7829):445–51. doi: 10.1038/s41586-020-2699-5
- Kamada R, Yang W, Zhang Y, Patel MC, Yang Y, Ouda R, et al. Interferon Stimulation Creates Chromatin Marks and Establishes Transcriptional Memory. *Proc Natl Acad Sci USA* (2018) 115(39):E9162–71. doi: 10.1073/ pnas.1720930115
- Kaneshiro KR, Rechtsteiner A, Strome S. Sperm-Inherited H3K27me3 Impacts Offspring Transcription and Development in C. Elegans. *Nat Commun* (2019) 10(1):1271. doi: 10.1038/s41467-019-09141-w
- Kaparakis M, Turnbull L, Carneiro L, Firth S, Coleman HA, Parkington HC, et al. Bacterial Membrane Vesicles Deliver Peptidoglycan to NOD1 in Epithelial Cells. *Cell Microbiol* (2010) 12(3):372–85. doi: 10.1111/j.1462-5822.2009.01404.x
- Kim CH. Immune Regulation by Microbiome Metabolites. *Immunology* (2018) 154(2):220–9. doi: 10.1111/imm.12930

- 74. Knutson AK, Egelhofer T, Rechtsteiner A, Strome S. Germ Granules Prevent Accumulation of Somatic Transcripts in the Adult Caenorhabditis Elegans Germline. *Genetics* (2017) 206(1):163–78. doi: 10.1534/genetics. 116.198549
- Koeppen K, Hampton TH, Jarek M, Scharfe M, Gerber SA, Mielcarz DW, et al. A Novel Mechanism of Host-Pathogen Interaction Through sRNA in Bacterial Outer Membrane Vesicles. *PloS Pathog* (2016) 12(6):e1005672. doi: 10.1371/journal.ppat.1005672
- Komsky A, Huleihel M, Ganaiem M, Kasterstein E, Komorovsky D, Bern O, et al. Presence of IL-18 in Testicular Tissue of Fertile and Infertile Men. *Andrologia* (2012) 44(1):1–8. doi: 10.1111/j.1439-0272.2010.01090.x
- Kronfol Z, Remick DG. Cytokines and the Brain: Implications for Clinical Psychiatry. Am J Psychiatry (2000) 157(5):683–94. doi: 10.1176/ appi.ajp.157.5.683
- Kudlow BA, Zhang L, Han M. Systematic Analysis of Tissue-Restricted miRISCs Reveals a Broad Role for microRNAs in Suppressing Basal Activity of the C. Elegans Pathogen Response. *Mol Cell* (2012) 46(4):530–41. doi: 10.1016/j.molcel.2012.03.011
- La Spina FA, Romanato M, Brugo-Olmedo S, De Vincentiis S, Julianelli V, Rivera RM, et al. Heterogeneous Distribution of Histone Methylation in Mature Human Sperm. J Assist Reprod Genet (2014) 31(1):45–9. doi: 10.1007/s10815-013-0137-4
- Lacal I, Ventura R. Epigenetic Inheritance: Concepts, Mechanisms and Perspectives. Front Mol Neurosci (2018) 11:292. doi: 10.3389/fnmol.2018.00292
- Lee HC, Gu W, Shirayama M, Youngman E, Conte D Jr, Mello CC. C. Elegans piRNAs Mediate the Genome-Wide Surveillance of Germline Transcripts. *Cell* (2012) 150(1):78–87. doi: 10.1016/j.cell.2012.06.016
- Chan JC, Morgan CP, Adrian Leu N, Shetty A, Cisse YM, Nugent BM, et al. Reproductive Tract Extracellular Vesicles are Sufficient to Transmit Intergenerational Stress and Program Neurodevelopment. *Nat Commun* (2020) 11(1):1499. doi: 10.1038/s41467-020-15305-w
- Legüe ME, Aguila B, Pollak B, Calixto A. Bacterial small RNAs and host epigenetic effectors of a transgenerational memory of pathogens in *C. elegans. bioRxiv* [Preprint] (2021). doi: 10.1101/2021.03.26.437277
- Lev I, Toker IA, Mor Y, Nitzan A, Weintraub G, Antonova O, et al. Germ Granules Govern Small RNA Inheritance. *Curr Biol* (2019) 29(17):2880– 91.e4. doi: 10.1016/j.cub.2019.07.054
- Levy M, Blacher E, Elinav E. Microbiome, Metabolites and Host Immunity. *Curr Opin Microbiol* (2017) 35:8–15. doi: 10.1016/j.mib.2016.10.003
- Li X, Yang L, Chen LL. The Biogenesis, Functions, and Challenges of Circular Rnas. *Mol Cell* (2018) 71(3):428–42. doi: 10.1016/j.molcel.2018. 06.034
- Lindmark B, Rompikuntal PK, Vaitkevicius K, Song T, Mizunoe Y, Uhlin BE, et al. Outer Membrane Vesicle-Mediated Release of Cytolethal Distending Toxin (CDT) From Campylobacter Jejuni. *BMC Microbiol* (2009) 9:220. doi: 10.1186/1471-2180-9-220
- Liu H, Wang X, Wang HD, Wu J, Ren J, Meng L, et al. Escherichia coli noncoding RNAs can affect gene expression and physiology of Caenorhabditis elegans. *Nat Commun* (2012) 3:1073. doi: 10.1038/ncomms2071
- Luczynski P, Whelan SO, O'Sullivan C, Clarke G, Shanahan F, Dinan TG, et al. Adult Microbiota-Deficient Mice Have Distinct Dendritic Morphological Changes: Differential Effects in the Amygdala and Hippocampus. *Eur J Neurosci* (2016) 44(9):2654–66. doi: 10.1111/ejn.13291
- Machtinger R, Laurent LC, Baccarelli AA. Extracellular Vesicles: Roles in Gamete Maturation, Fertilization and Embryo Implantation. *Hum Reprod Update* (2016) 22(2):182–93. doi: 10.1093/humupd/dmv055
- Macia L, Tan J, Vieira AT, Leach K, Stanley D, Luong S, et al. Metabolitesensing receptors GPR43 and GPR109A facilitate dietary fibre-induced gut homeostasis through regulation of the inflammasome. *Nat Commun* (2015) 6:6734. doi: 10.1038/ncomms7734
- Maciel E, Mansuy IM. Extracellular Vesicles and Their miRNA Cargo: A Means of Communication Between Soma and Germline in the Mammalian Reproductive System. *Chimia (Aarau)* (2019) 73(6):356–61. doi: 10.2533/ chimia.2019.356
- Mao H, Zhu C, Zong D, Weng C, Yang X, Huang H, et al. The Nrde Pathway Mediates Small-RNA-Directed Histone H3 Lysine 27 Trimethylation in Caenorhabditis elegans. *Curr Biol* (2015) 25(18):2398–403. doi: 10.1016/ j.cub.2015.07.051

- Marré J, Traver EC, Jose AM. Extracellular RNA is transported from one generation to the next in Caenorhabditis elegans. *Proc Natl Acad Sci USA* (2016) 113(44):12496–501. doi: 10.1073/pnas.1608959113
- Martinez RM, Liang L, Racowsky C, Dioni L, Mansur A, Adir M, et al. Extracellular microRNAs profile in human follicular fluid and IVF outcomes. Sci Rep (2018) 8(1):17036. doi: 10.1038/s41598-018-35379-3
- Masuzzo A, Montanari M, Kurz L, Royet J. How Bacteria Impact Host Nervous System and Behaviors: Lessons From Flies and Worms. *Trends Neurosci* (2020) 43(12):998–1010. doi: 10.1016/j.tins.2020.09.007
- Matsumoto M, Takeda Y, Tatematsu M, Seya T. Toll-Like Receptor 3 Signal in Dendritic Cells Benefits Cancer Immunotherapy. *Front Immunol.* (2017) 8:1897. doi: 10.3389/fimmu.2017.01897
- McEwan DL, Weisman AS, Hunter CP. Uptake of Extracellular Double-Stranded RNA by SID-2. *Mol Cell* (2012) 47(5):746–54. doi: 10.1016/ j.molcel.2012.07.014
- Meerson A, Cacheaux L, Goosens KA, Sapolsky RM, Soreq H, Kaufer D. Changes in Brain MicroRNAs Contribute to Cholinergic Stress Reactions. *J Mol Neurosci* (2010) 40(1-2):47–55. doi: 10.1007/s12031-009-9252-1
- 100. Meisel JD, Panda O, Mahanti P, Schroeder FC, Kim DH. Chemosensation of Bacterial Secondary Metabolites Modulates Neuroendocrine Signaling and Behavior of C. Elegans. *Cell* (2014) 159(2):267–80. doi: 10.1016/ j.cell.2014.09.011
- 101. Minuk GY. Gamma-Aminobutyric Acid (GABA) Production by Eight Common Bacterial Pathogens. Scand J Infect Dis (1986) 18(5):465–7. doi: 10.3109/00365548609032366
- Moloney GM, Dinan TG, Clarke G, Cryan JF. Microbial Regulation of microRNA Expression in the Brain-Gut Axis. *Curr Opin Pharmacol* (2019) 48:120–6. doi: 10.1016/j.coph.2019.08.005
- 103. Moore RS, Kaletsky R, Murphy CT. Piwi/PRG-1 Argonaute and TGF-β Mediate Transgenerational Learned Pathogenic Avoidance. *Cell* (2019) 177 (7):1827–41.e12. doi: 10.1016/j.cell.2019.05.024
- 104. Morgado L, Preite V, Oplaat C, Anava S, Ferreira de Carvalho J, Rechavi O, et al. Small RNAs Reflect Grandparental Environments in Apomictic Dandelion. *Mol Biol Evol* (2017) 34(8):2035–40. doi: 10.1093/molbev/ msx150
- 105. Mossad O, Erny D. The Microbiota-Microglia Axis in Central Nervous System Disorders. *Brain Pathol* (2020) 30(6):1159–77. doi: 10.1111/ bpa.12908
- 106. Nast R, Choepak T, Lüder CGK. Epigenetic Control of IFN-γHost Responses During Infection With Toxoplasma Gondii. Front Immunol (2020) 11:581241. doi: 10.3389/fimmu.2020.581241
- 107. Niu Q, Huang X, Zhang L, Xu J, Yang D, Wei K, et al. A Trojan horse mechanism of bacterial pathogenesis against nematodes. *Proc Natl Acad Sci* USA (2010) 107(38):16631–6. doi: 10.1073/pnas.1007276107
- 108. Noferesti SS, Sohel MM, Hoelker M, Salilew-Wondim D, Tholen E, Looft C, et al. Controlled ovarian hyperstimulation induced changes in the expression of circulatory miRNA in bovine follicular fluid and blood plasma. *J Ovarian Res* (2015) 8:81. doi: 10.1186/s13048-015-0208-5
- 109. Nono M, Kishimoto S, Sato-Carlton A, Carlton PM, Nishida E, Uno M. Intestineto-Germline Transmission of Epigenetic Information Intergenerationally Ensures Systemic Stress Resistance in C. Elegans. *Cell Rep* (2020) 30(10):3207–17.e4. doi: 10.1016/j.celrep.2020.02.050
- 110. O'Brien EA, Ensbey KS, Day BW, Baldock PA, Barry G. Direct Evidence for Transport of RNA From the Mouse Brain to the Germline and Offspring. BMC Biol (2020) 18(1):45. doi: 10.1186/s12915-020-00780-w
- 111. Palominos MF, Verdugo L, Gabaldon C, Pollak B, Ortíz-Severín J, Varas MA, et al. Transgenerational Diapause as an Avoidance Strategy against Bacterial Pathogens in Caenorhabditis elegans. *mBio* (2017) 8(5):e01234-17. doi: 10.1128/mBio.01234-17
- Pate JL, Landis Keyes P. Immune Cells in the Corpus Luteum: Friends or Foes. *Reproduction* (2001) 122(5):665–76. doi: 10.1530/rep.0.1220665
- 113. Pereira AG, Gracida X, Kagias K, Zhang Y. C. Elegans Aversive Olfactory Learning Generates Diverse Intergenerational Effects. J Neurogenet (2020) 34:1–11. doi: 10.1080/01677063.2020.1819265
- 114. Perera BPU, Tsai ZT, Colwell ML, Jones TR, Goodrich JM, Wang K, et al. Somatic expression of piRNA and associated machinery in the mouse identifies short, tissue-specific piRNA. *Epigenetics* (2019) 14(5):504–21. doi: 10.1080/15592294.2019.1600389

- 115. Perez MF, Shamalnasab M, Mata-Cabana A, Della Valle S, Olmedo M, Francesconi M, et al. Neuronal perception of the social environment intergenerationally controls germline development and generation time in *C. elegans. bioRxiv* [Preprint] (2020). doi: 10.1101/2020.09.03.279158
- 116. Posner R, Toker IA, Antonova O, Star E, Anava S, Azmon E, et al. Neuronal Small RNAs Control Behavior Transgenerationally. *Cell* (2019) 177(7):1814– 26.e15. doi: 10.1016/j.cell.2019.04.029
- 117. Pradel E, Zhang Y, Pujol N, Matsuyama T, Bargmann CI, Ewbank JJ. Detection and avoidance of a natural product from the pathogenic bacterium Serratia marcescens by Caenorhabditis elegans. *Proc Natl Acad Sci USA* (2007) 104(7):2295–300. doi: 10.1073/pnas.0610281104
- 118. Prihatno SA, Adi YK, Budipitojo T. Immunolocalization of IL-6 and IL-10 in the Testicular Tissue of Testicular Dysfunction Rat Treated With Secretome. J Adv Vet Anim Res (2020) 7(3):514–20. doi: 10.5455/ javar.2020.g448
- 119. Qamar AY, Mahiddine FY, Bang S, Fang X, Shin ST, Kim MJ, et al. Extracellular Vesicle Mediated Crosstalk Between the Gametes, Conceptus, and Female Reproductive Tract. *Front Vet Sci* (2020) 7:589117. doi: 10.3389/ fvets.2020.589117
- 120. Qin Y, Wade PA. Crosstalk Between the Microbiome and Epigenome: Messages From Bugs. J Biochem (2018) 163(2):105–12. doi: 10.1093/jb/ mvx080
- 121. Rechavi O, Houri-Ze'evi L, Anava S, Goh WSS, Kerk SY, Hannon GJ, et al. Starvation-induced Transgenerational Inheritance of Small RNAs in C. Elegans. *Cell* (2014) 158(2):277–87. doi: 10.1016/j.cell.2014.06.020
- Rechavi O, Minevich G, Hobert O. Transgenerational Inheritance of an Acquired Small RNA-based Antiviral Response in C. Elegans. *Cell* (2011) 147 (6):1248–56. doi: 10.1016/j.cell.2011.10.042
- 123. Renelli M, Matias V, Lo RY, Beveridge TJ. DNA-Containing Membrane Vesicles of Pseudomonas Aeruginosa PAO1 and Their Genetic Transformation Potential. *Microbiology* (2004) 150(Pt 7):2161-9. doi: 10.1099/mic.0.26841-0
- 124. Rodgers AB, Morgan CP, Bronson SL, Revello S, Bale TL. Paternal Stress Exposure Alters Sperm microRNA Content and Reprograms Offspring HPA Stress Axis Regulation. J Neurosci (2013) 33(21):9003–12. doi: 10.1523/ JNEUROSCI.0914-13.2013
- 125. Rosenberg E, Koren O, Reshef L, Efrony R, Zilber-Rosenberg I. The Role of Microorganisms in Coral Health, Disease and Evolution. Nat Rev Microbiol (2007) 5:355–62. doi: 10.1038/nrmicro1635
- 126. Russell SJ, LaMarre J. Transposons and the PIWI pathway: genome defense in gametes and embryos. *Reproduction* (2018) 156(4):R111–24. doi: 10.1530/ REP-18-0218
- Schackwitz WS, Inoue T, Thomas JH. Chemosensory Neurons Function in Parallel to Mediate a Pheromone Response in C. Elegans. *Neuron* (1996) 17 (4):719–28. doi: 10.1016/s0896-6273(00)80203-2
- Schoofs A, Hückesfeld S, Surendran S, Pankratz MJ. Serotonergic Pathways in the Drosophila Larval Enteric Nervous System. J Insect Physiol (2014) 69:118–25. doi: 10.1016/j.jinsphys.2014.05.022
- 129. Sharma A. Transgenerational Epigenetic Inheritance: Focus on Soma to Germline Information Transfer. *Prog Biophys Mol Biol* (2013) 113(3):439– 46. doi: 10.1016/j.pbiomolbio.2012.12.003
- Shih JD, Hunter CP. SID-1 is a dsRNA-selective dsRNA-gated Channel. RNA (2011) 17(6):1057–65. doi: 10.1261/rna.2596511
- 131. Skinner MK, Ben Maamar M, Sadler-Riggleman I, Beck D, Nilsson E, McBirney M, et al. Alterations in sperm DNA methylation, non-coding RNA and histone retention associate with DDT-induced epigenetic transgenerational inheritance of disease. *Epigenet Chromatin* (2018) 11 (1):8. doi: 10.1186/s13072-018-0178-0
- Skinner MK, Guerrero-Bosagna C. Environmental Signals and Transgenerational Epigenetics. *Epigenomics* (2009) 1:111–7. doi: 10.2217/epi.09.11
- 133. Söder O, Sultana T, Jonsson C, Wahlgren A, Petersen C, Holst M. The Interleukin-1 System in the Testis. Andrologia (2000) 32(1):52–5.
- 134. Spencer NJ, Hu H. Enteric Nervous System: Sensory Transduction, Neural Circuits and Gastrointestinal Motility. Nat Rev Gastroenterol Hepatol (2020) 17(6):338–51. doi: 10.1038/s41575-020-0271-2
- 135. Stilling RM, Dinan TG, Cryan JF. Microbial Genes, Brain & Behaviour -Epigenetic Regulation of the Gut-Brain Axis. *Genes Brain Behav* (2014) 13 (1):69–86. doi: 10.1111/gbb.12109

- 136. Tesfaye D, Gebremedhn S, Salilew-Wondim D, Hailay T, Hoelker M, Grosse-Brinkhaus C, et al. MicroRNAs: tiny molecules with a significant role in mammalian follicular and oocyte development. *Reproduction* (2018) 155(3): R121–35. doi: 10.1530/REP-17-0428
- 137. Tosar JP, García-Silva MR, Cayota A. Circulating SNORD57 Rather Than piR-54265 is a Promising Biomarker for Colorectal Cancer: Common Pitfalls in the Study of Somatic piRNAs in Cancer. *RNA* (2021) 27(4):403–10. doi: 10.1261/rna.078444.120
- 138. Toyofuku M, Cárcamo-Oyarce G, Yamamoto T, Eisenstein F, Hsiao CC, Kurosawa M, et al. Prophage-triggered membrane vesicle formation through peptidoglycan damage in Bacillus subtilis. *Nat Commun* (2017) 8(1):481. doi: 10.1038/s41467-017-00492-w
- 139. Troemel ER, Kimmel BE, Bargmann CI. Reprogramming Chemotaxis Responses: Sensory Neurons Define Olfactory Preferences in C. Elegans. *Cell* (1997) 91(2):161–9. doi: 10.1016/s0092-8674(00)80399-2
- 140. Tsatsaronis JA, Franch-Arroyo S, Resch U, Charpentier E. Extracellular Vesicle Rna: A Universal Mediator of Microbial Communication. *Trends Microbiol* (2018) 26(5):401–10. doi: 10.1016/j.tim.2018.02.009
- 141. Turnbull L, Toyofuku M, Hynen AL, Kurosawa M, Pessi G, Petty NK, et al. Explosive Cell Lysis as a Mechanism for the Biogenesis of Bacterial Membrane Vesicles and Biofilms. *Nat Commun* (2016) 7:11220. doi: 10.1038/ncomms11220
- 142. Tyebji S, Hannan AJ, Tonkin CJ. Pathogenic Infection in Male Mice Changes Sperm Small Rna Profiles and Transgenerationally Alters Offspring Behavior. *Cell Rep* (2020) 31(4):107573. doi: 10.1016/j.celrep. 2020.107573
- 143. Urrutia A, García-Angulo VA, Fuentes A, Caneo M, Legüe M, Urquiza S, et al. Bacterially produced metabolites protect C. elegans neurons from degeneration. *PloS Biol* (2020) 18(3):e3000638. doi: 10.1371/journal.pbio.3000638
- 144. Valadi H, Ekström K, Bossios A, Sjöstrand M, Lee JJ, Lötvall JO. Exosome-Mediated Transfer of mRNAs and microRNAs is a Novel Mechanism of Genetic Exchange Between Cells. *Nat Cell Biol* (2007) 9(6):654–9. doi: 10.1038/ncb1596
- 145. van Otterdijk SD, Michels KB. Transgenerational Epigenetic Inheritance in Mammals: How Good is the Evidence? FASEB J (2016) 30(7):2457–65. doi: 10.1096/fj.201500083
- 146. Vanaja SK, Russo AJ, Behl B, Banerjee I, Yankova M, Deshmukh SD, et al. Bacterial Outer Membrane Vesicles Mediate Cytosolic Localization of LPS and Caspase-11 Activation. *Cell* (2016) 165(5):1106–19. doi: 10.1016/ j.cell.2016.04.015
- Verdel A, Vavasseur A, Le Gorrec M, Touat-Todeschini L. Common Themes in siRNA-mediated Epigenetic Silencing Pathways. *Int J Dev Biol* (2009) 53 (2-3):245–57. doi: 10.1387/ijdb.082691av
- 148. Wall R, Cryan JF, Ross RP, Fitzgerald GF, Dinan TG, Stanton C. Bacterial Neuroactive Compounds Produced by Psychobiotics. *Adv Exp Med Biol* (2014) 817:221–39. doi: 10.1007/978-1-4939-0897-4\_10
- Wang E, Hunter CP. Sid-1 Functions in Multiple Roles To Support Parental Rnai in. *Genetics* (2017) 207(2):547–57. doi: 10.1534/genetics.117.300067
- 150. Watkins LR, Maier SF, Goehler LE. Cytokine-to-Brain Communication: A Review & Analysis of Alternative Mechanisms. *Life Sci* (1995) 57(11):1011– 26. doi: 10.1016/0024-3205(95)02047-m
- 151. Watson E, MacNeil LT, Ritter AD, Yilmaz LS, Rosebrock AP, Caudy AA, et al. Interspecies Systems Biology Uncovers Metabolites Affecting C. Elegans Gene Expression and Life History Traits. *Cell* (2014) 156(4):759–70. doi: 10.1016/j.cell.2014.01.047
- 152. Werner KM, Perez LJ, Ghosh R, Semmelhack MF, Bassler BL. Caenorhabditis Elegans Recognizes a Bacterial Quorum-Sensing Signal Molecule Through the AWCON Neuron. J Biol Chem (2014) 289 (38):26566-73. doi: 10.1074/jbc.M114.573832
- 153. Wu SE, Hashimoto-Hill S, Woo V, Eshleman EM, Whitt J, Engleman L, et al. Microbiota-derived metabolite promotes HDAC3 activity in the gut. *Nature* (2020) 586(7827):108–12. doi: 10.1038/s41586-020-2604-2
- 154. Xie Y, Yao L, Yu X, Ruan Y, Li Z, Guo J. Action Mechanisms and Research Methods of tRNA-derived Small Rnas. *Signal Transduct Target Ther* (2020) 5 (1):109. doi: 10.1038/s41392-020-00217-4
- 155. Yan W. Potential Roles of Noncoding RNAs in Environmental Epigenetic Transgenerational Inheritance. *Mol Cell Endocrinol* (2014) 398(1-2):24–30. doi: 10.1016/j.mce.2014.09.008

- 156. Zhang Y, Lu H, Bargmann CI. Pathogenic Bacteria Induce Aversive Olfactory Learning in Caenorhabditis Elegans. *Nature* (2005) 438 (7065):179–84. doi: 10.1038/nature04216
- 157. Zhang Y, Zhang X, Shi J, Tuorto F, Li X, Liu Y, et al. Dnmt2 mediates intergenerational transmission of paternally acquired metabolic disorders through sperm small non-coding RNAs. *Nat Cell Biol* (2018) 20(5):535–40. doi: 10.1038/s41556-018-0087-2
- Zhang Z, Mu X, Cao Q, Shi Y, Hu X, Zheng H. Honeybee gut microbiota modulates host behaviors and neurological processes. *bioRxiv* [Preprint] (2020). doi: 10.1101/2020.12.19.423587

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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