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Journal

Microbiology Resource Announcements, 5(19)

ISSN

2576-098X

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Publication Date

2017-05-11

DOI

10.1128/genomea.00341-17

Peer reviewed



Implication of Sialidases in *Salmonella* Infection: Genome Release of Sialidase Knockout Strains from *Salmonella enterica* Serovar Typhimurium LT2

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ABSTRACT Sialidases, which are widely distributed in nature, cleave the α -ketosidic bond of terminal sialic acid residue. These emerging virulence factors degrade the host glycan. We report here the release of seven sialidase and one sialic acid transporter deletion in *Salmonella enterica* serovar Typhimurium strain LT2, which are important in cellular invasion during infection.

Sialidases are widely distributed among microbes and are one of the least characterized and ill-defined glycosyl hydrolases. Sialidases have been associated with several diseases. Sialidases play a critical role in microbiology by mediating metabolism, adherence, and infection, and they are important regulators of alternate complement pathway activation, red blood cell destruction, cell growth, cell adhesion, and tumor metastasis in mammalian systems (1–5). Recently, the importance of sialidases in infection and commensalism has come to light, opening the potential to use newly measured genomic diversity as a means to investigate infection mechanisms. Though antibiotics are available for treatment of bacterial infections, inhibitors of all sialidases and new drug targets may be medically useful where sialidase activity has been correlated with severe infection pathology.

The presence of sialidases is highly correlated with the progress and severity of the disease, and the most probable role of sialidases is for successful attachment and colonization. Microbes use sialidases to reveal the cell surface that holds sialic acid-containing cell membrane receptors during infection. Sialidases play an important role in infection by altering the host glycan structure to gain access of the host epithelial cells by binding to terminal sialic acid receptors to initiate glycan degradation (6). The two sialidases ($\Delta nanH$ and $\Delta STM1252$) from *Salmonella enterica* serovar Typhimurium LT2 have the same domains and function as sialidases, but they are structurally very different, indicating domain shuffling and lack of structural conservation; therefore, this difference led to different invasion phenotypes during the *in vitro* infection of differentiated colonic epithelial cells (Caco-2) (6).

The 100K Pathogen Genome Project (<http://www.100genomes.org>) is a large-scale sequencing consortium that offers the use of new next-generation sequencing methods to provide cutting-edge methods for pathogen detection and control in the food supply. This project is focused on producing genomes of pathogenic isolates from the environment, plants, animals, and humans worldwide, providing new insights into the genetic diversity of *Salmonella* spp. and other foodborne pathogens. These seven sialidase and one sialic acid transporter mutant strain were constructed in the Weimer Laboratory (UC Davis, Davis, CA, USA) (6) as described by Datsenko and Wanner (7). Cultures were grown on 1.5% Luria–Bertani agar (Difco, Franklin Lakes, NJ, USA), with 10 μ g/mL of chloramphenicol at 37°C, and then lysed (8). Genomic DNA was extracted (9), checked for quality (10), and

Received 21 March 2017 Accepted 27 March 2017 Published 11 May 2017

Citation Arabyan N, Weis AM, Huang BC, Weimer BC. 2017. Implication of sialidases in *Salmonella* infection: genome release of sialidase knockout strains from *Salmonella enterica* serovar Typhimurium LT2. Genome Announc 5:e00341-17. <https://doi.org/10.1128/genomeA.00341-17>.

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TABLE 1 *S. Typhimurium* LT2 sialidase and sialic acid transporter deletion mutants

GenBank accession no.	SRA accession no.	Isolate name	Gene deleted	No. of contigs	Coverage (×)	Total genome size (bp)	No. of coding sequences
MWQQ00000000	SRR5279339	BCW_7500	$\Delta nanT$	65	143	4,895,101	4,810
MWVC00000000	SRR5288771	BCW_7514	$\Delta invA::\Delta nanH$	240	15	4,892,397	4,898
MWQR00000000	SRR5279338	BCW_7515	$\Delta invA::\Delta STM1252$	74	130	4,892,686	4,819
MWQS00000000	SRR5279337	BCW_7516	$\Delta melA::\Delta nanH$	72	142	4,870,638	4,785
MWQT00000000	SRR5279336	BCW_7517	$\Delta melA::\Delta STM1252$	59	302	4,895,400	4,805
MWQU00000000	SRR5279335	BCW_7518	$\Delta nanH::\Delta STM1252$	54	106	4,893,964	4,803
MWQV00000000	SRR3622954	BCW_8441	$\Delta STM1252$	57	165	4,894,714	4,806
MWQW00000000	SRR3622955	BCW_8442	$\Delta nanH$	60	139	4,894,435	4,815

fragmented (11). The 350- to 500-bp libraries (12, 13) were indexed (96 genomes/lane) and sequenced (Illumina HiSeq 3000; 150-bp paired-end) (14–16) at the UC Davis DNA Technologies Core. Paired-end reads were *de novo* assembled using CLC Workbench version 6 with default parameters. Here, the 100K Pathogen Genome Project has assembled seven genomes of single and double sialidases and one sialic acid transporter deletion strain of *S. Typhimurium* LT2.

Accession number(s). All sequences are publicly available and can be found at the 100K Pathogen Genome Project (NCBI PRJNA186441) in the Sequence Read Archive (<http://www.ncbi.nlm.nih.gov/sra>); genome assemblies can be found in NCBI GenBank (Table 1).

ACKNOWLEDGMENTS

B.C.W. gratefully acknowledges funding for this project from Mars, Inc., NIH (1R01HD065122-01A1 and U24-DK097154), and Agilent Technologies (Thought Leader Award).

REFERENCES

- Walther T, Karamanska R, Chan RW, Chan MC, Jia N, Air G, Hopton C, Wong MP, Dell A, Malik Peiris JS, Haslam SM, Nicholls JM. 2013. Glycomic analysis of human respiratory tract tissues and correlation with influenza virus infection. *PLoS Pathog* 9:e1003223. <https://doi.org/10.1371/journal.ppat.1003223>.
- Imai M, Kawaoka Y. 2012. The role of receptor binding specificity in interspecies transmission of influenza viruses. *Curr Opin Virol* 2:160–167. <https://doi.org/10.1016/j.coviro.2012.03.003>.
- Varki A, Gagneux P. 2012. Multifarious roles of sialic acids in immunity. *Ann N Y Acad Sci* 1253:16–36. <https://doi.org/10.1111/j.1749-6632.2012.06517.x>.
- Chan RW, Karamanska R, Van Poucke S, Van Reeth K, Chan IW, Chan MC, Dell A, Peiris JS, Haslam SM, Guan Y, Nicholls JM. 2013. Infection of swine *ex vivo* tissues with avian viruses including H7N9 and correlation with glycomic analysis. *Influenza Other Respir Viruses* 7:1269–1282. <https://doi.org/10.1111/irv.12144>.
- de Graaf M, Fouchier RA. 2014. Role of receptor binding specificity in influenza A virus transmission and pathogenesis. *EMBO J* 33:823–841. <https://doi.org/10.1002/embj.201387442>.
- Arabyan N, Park D, Foutouhi S, Weis AM, Huang BC, Williams CC, Desai P, Shah J, Jeannotte R, Kong N, Lebrilla CB, Weimer BC. 2016. *Salmonella* degrades the host glycocalyx leading to altered infection and glycan remodeling. *Sci Rep* 6:29525. <https://doi.org/10.1038/srep29525>.
- Datsenko KA, Wanner BL. 2000. One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proc Natl Acad Sci U S A* 97:6640–6645. <https://doi.org/10.1073/pnas.120163297>.
- Jeannotte R, Lee E, Kong N, Ng W, Kelly L, Weimer BC. 2014. High-throughput analysis of foodborne bacterial genomic DNA using Agilent 2200 TapeStation and genomic DNA ScreenTape system. Agilent Technologies Application Note. Agilent Technologies, Inc., Santa Clara, CA. <https://doi.org/10.13140/RG.2.1.3354.6961>.
- Kong N, Ng W, Lee V, Kelly L, Weimer BC. 2013. Production and analysis of high molecular weight genomic DNA for NGS pipelines using Agilent DNA extraction kit (p/n 200600). Agilent Technologies Application Note. Agilent Technologies, Inc., Santa Clara, CA. <https://doi.org/10.13140/RG.2.1.2961.4807>.
- Kong N, Ng W, Cai L, Leonardo A, Kelly L, Weimer BC. 2014. Integrating the DNA integrity number (DIN) to assess genomic DNA (gDNA) quality control using the Agilent 2200 TapeStation system. Agilent Technologies Application Note. Agilent Technologies, Inc., Santa Clara, CA. <https://doi.org/10.13140/RG.2.1.3616.8409>.
- Jeannotte R, Lee E, Arabyan N, Kong N, Thao K, Huang BH, Kelly L, Weimer BC. 2014. Optimization of Covaris settings for shearing bacterial genomic DNA by focused ultrasonication and analysis using Agilent 2200 TapeStation. Agilent Technologies Application Note. Agilent Technologies, Inc., Santa Clara, CA. <https://doi.org/10.13140/RG.2.1.2830.4082>.
- Kong N, Ng W, Foutouhi A, Huang BH, Kelly L, Weimer BC. 2014. Quality control of high-throughput library construction pipeline for KAPA HTP library using an Agilent 2200 TapeStation. Agilent Technologies Application Note. Agilent Technologies, Inc., Santa Clara, CA. <https://doi.org/10.13140/RG.2.1.4927.5604>.
- Kong N, Thao K, Huang C, Appel M, Lappin S, Knapp L, Kelly L, Weimer BC. 2014. Automated library construction using KAPA library preparation kits on the Agilent NGS workstation yields high-quality libraries for whole-genome sequencing on the Illumina platform. Agilent Technologies Application Note. Agilent Technologies, Inc., Santa Clara, CA. <https://doi.org/10.13140/RG.2.1.2306.1203>.
- Lüdeke CH, Kong N, Weimer BC, Fischer M, Jones JL. 2015. Complete genome sequences of a clinical isolate and an environmental isolate of *Vibrio parahaemolyticus*. *Genome Announc* 3(2):e00216-15. <https://doi.org/10.1128/genomeA.00216-15>.
- Weis AM, Clothier KA, Huang BC, Kong N, Weimer BC. 2016. Draft genome sequences of *Campylobacter jejuni* strains that cause abortion in livestock. *Genome Announc* 4(6):e01324-16. <https://doi.org/10.1128/genomeA.01324-16>.
- Weis AM, Huang BC, Storey DB, Kong N, Chen P, Arabyan N, Gilpin B, Mason C, Townsend AK, Smith WA, Byrne BA, Taff CC, Weimer BC. 2017. Large-scale release of *Campylobacter* draft genomes: resources for food safety and public health from the 100K pathogen genome project. *Genome Announc* 5(1):e00925-16. <https://doi.org/10.1128/genomeA.00925-16>.