

Spontaneous and Selective Formation of HSNO, a Crucial Intermediate Linking H₂S and Nitroso Chemistries

Matthew Nava,[†] Marie-Aline Martin-Drumel,[‡] Christopher A. Lopez,[¶] Kyle N. Crabtree,^{‡,||}
Caroline C. Womack,^{†,⊥} Thanh L. Nguyen,[¶] Sven Thorwirth,[§] Christopher C. Cummins,[†] John
F. Stanton,[¶] and Michael C. McCarthy^{*,‡}

[†]Department of Chemistry, Massachusetts Institute of Technology, Cambridge MA 02139, USA

[‡]Harvard-Smithsonian Center for Astrophysics, Cambridge, MA 02138, USA and School of Engineering
and Applied Sciences, Harvard University, Cambridge, MA 02138, USA

[¶]Institute for Theoretical Chemistry, Department of Chemistry, University of Texas, Austin, TX 78712,
USA

[§]I. Physikalisches Institut, Universität zu Köln, 50937 Köln, Germany

^{||}Present address: Department of Chemistry, University of California-Davis, Davis, CA 95616, USA

[⊥]Present address: Chemical Sciences Division, NOAA ESRL, Boulder, CO 80305, USA

Received August 14, 2016; E-mail: mmccarthy@cfa.harvard.edu

Abstract: Thionitrous acid (HSNO), a potential key intermediate in biological signaling pathways, has been proposed to link NO and H₂S biochemistries, but its existence and stability in vivo remains controversial. We establish that HSNO is spontaneously formed in high concentration when NO and H₂S gases are mixed at room temperature in the presence of metallic surfaces. Our measurements reveal that HSNO is formed by the reaction $\text{H}_2\text{S} + \text{N}_2\text{O}_3 \rightarrow \text{HSNO} + \text{HNO}_2$, where N_2O_3 is a product of NO disproportionation. These studies also suggest that further reaction of HSNO with H₂S may form HNO and HSSH. The length of the S–N bond has been derived to high precision, and is found to be unusually long: 1.84 Å – the longest S–N bond reported to date for an R-SNO compound. The present structural and, particularly, reactivity investigations of this elusive molecule provide a firm foundation to better understand its potential physiological chemistry and propensity to undergo S–N bond cleavage in vivo.

The intriguing similarities between the biological profiles of NO and H₂S, two important gasotransmitters acting notably as blood pressure mediators, have raised questions about the possibility of ‘cross-talk’ between the two species.¹ Such an interaction could explain the surprisingly beneficial effects of these signaling molecules as it may allow H₂S to modulate the availability of NO within the body.² The vast majority of NO in vivo is bound as S-nitrosothiols (RSNO), a family of molecules well known for their crucial role in response to oxygen deprivation^{3–5} as well as for being important biological reservoir for NO.^{6,7} RSNOs are typically formed through the reaction of nitrosylating agents with thiols (RSH),⁸ Once formed, RSNOs can also act as nitrosylating agents thus highlighting their role as NO shuttles at the cellular level.

The simplest RSNO, thionitrous acid (HSNO), has been proposed to be the product of the reaction between NO and H₂S;⁹ other possible species have been implicated in this ‘cross-talk’, including nitrosopersulfide (SSNO[−]) and *N*-nitrosohydroxylamine-*N*-sulfonate.¹⁰ Unlike larger RSNOs, HSNO is thought to be able to freely diffuse through cell membranes² suggesting that it has the ability to transnitrosate proteins removed from the sites where NO is synthesized, and therefore could play a key role in cellular re-

dox regulation.¹¹ Understanding the role of small molecules such as NO and HNO in biological signal transduction has resulted in important advances in medicine.^{12,13} To shed light on the role of HSNO in vivo, sensors are currently being developed to image this elusive molecule. However, the biochemistry (formation, transport, decomposition) of HSNO remains poorly understood and the structure of the molecule has not been determined to date. HSNO has been spectroscopically characterized by infrared measurements in low temperature argon matrices.^{14,15} The molecule was produced from photolysis of HNSO, a low-lying stable structural isomer of HSNO ($\Delta H \geq 10$ kcal/mol,¹⁶ see Supplementary Information). Recently, HSNO has been detected under physiologically relevant conditions through protonation of [SNO][−] or treatment of S-nitrosoglutathione with sodium sulfide by infrared,¹⁵ NMR and mass spectroscopy.² Additionally, coordinated HSNO has been transiently observed by mass spectroscopy¹⁷ and very recently a crystal structure was obtained of an RSNO coordinated to a copper complex.¹⁸ However, these measurements provided only limited insight into the structure and subsequent reactivity of HSNO.

The length of the S–N bond in HSNO is used to infer the stability of this molecule under physiological conditions, ostensibly because a long, weak S–N bond would allow HSNO to undergo facile homolysis to the NO and SH radicals,¹⁹ thus rationalizing its ability to serve as a NO carrier. Theoretical studies of this bond length, remain controversial because, as with larger RSNOs, the length and strength of this central bond appear to be highly dependent on the method and basis set used.²⁰ Calculated S–N bond lengths in HSNO range from as short as 1.78 Å to as long as 1.93 Å.^{21,22} From theoretical calculations, however, it is well-established that HSNO has two conformers very close in energy (the *trans* is 0.8 kcal/mol more stable than the *cis*) but separated by a relatively high torsional barrier (6.7 kcal/mol, see figure S1), a finding which is qualitatively consistent with a partial double bond character of the S–N bond.²³ In contrast, the NO donor character of this class of molecules is likely to arise from the weak strength of the S–N bond (25–31 kcal/mol), an indication of a partial ionic character.^{20,24} To explain these paradoxical bonding properties, theoretical models of the electronic structure of RSNOs include contributions from conventional (R–S–N=O), zwitter-

terionic ($R-S^+=N-O^-$), and ionic (RS^-/NO^+) resonance structures.^{21,25}

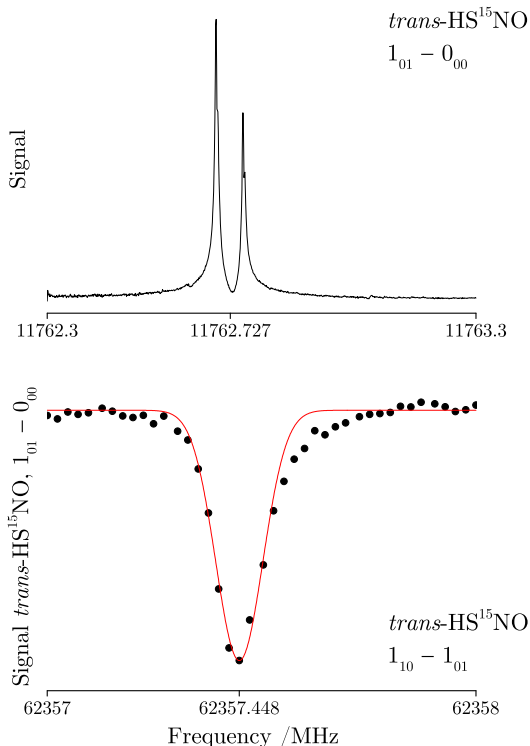


Figure 1. (Top) FTMW spectrum of the two Doppler components of the $1_{01} - 0_{00}$ transition of *trans*-HS¹⁵NO (integration time: ~ 1 min). (Bottom) DR spectrum of the $1_{10} - 0_{01}$ transition of *trans*-HS¹⁵NO, obtained while monitoring the signal of the $1_{01} - 0_{00}$ transition (integration time: ~ 5 min).

In a serendipitous discovery, we found that when a mixture of H₂S and NO heavily diluted in an inert buffer gas (Ne) is expanded into a vacuum chamber, intense rotational lines of a previously unknown molecule, now unambiguously identified as HSNO, are observed by Fourier-transform microwave (FTMW) spectroscopy, a highly sensitive and selective technique which has been used to detect many molecules, both familiar and exotic.^{26,27} Rotational transitions (Figure 1) have been measured for both the *cis* and *trans* conformers of the main isotopologue of HSNO and four singly-substituted isotopic species (DSNO, H³⁴SNO, HS¹⁵NO, HSN¹⁸O). From these data, highly accurate molecular parameters (rotational, centrifugal distortion, and hyperfine constants) have been derived (Table S1). A precise geometrical structure has been obtained for both conformers from a combination of isotopic substitution and vibrational corrections calculated quantum-chemically (see Figure 2 and Tables S2-S3). Most significantly, this analysis yields a S–N bond length of 1.834 ± 0.002 Å for *cis*-HSNO and 1.852 ± 0.002 Å for *trans*-HSNO, both of which are very long in comparison to other RSNOs where the S–N bond is typically 1.75 Å (see figures S2 and S3, and references therein). The S–N bond length in HSNO is in fact the longest experimentally reported for an RSNO compound and has significant consequences on the stability of HSNO with respect to homolytic or heterolytic cleavage, particularly *in vivo*. With respect to calculated S–N bond lengths of HSNO, the precise values reported here should serve as a useful benchmark for future quantum chemical studies. These values are in good agreement with high level quantum chemical calculations [CCSD(T)] from the literature^{21,22} and undertaken in this

study (Table S2). Natural resonance theory analysis performed in this work also indicates that there is only a single intrinsic resonance structure for HSNO, H–S–N=O. Additional experimental details, theoretical calculations, measured frequencies, rotational constants, and structural parameters (experimental and calculated) can be found in the supplementary material.

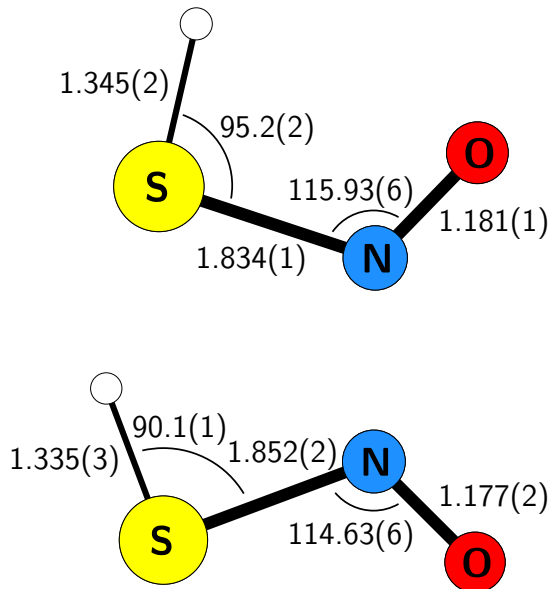
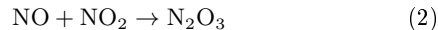
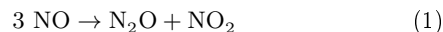


Figure 2. Structure of *cis*- (top panel) and *trans*-HSNO (bottom panel) determined in this study (semi-experimental equilibrium structure, see Table S1). Bond lengths are in Å and angles in degrees.

Our study also shows that although both *trans* and *cis* forms of HSNO are readily produced, *trans*-HSNO is roughly 5 times more abundant than the *cis* form. Since the conformational change $cis\text{-HSNO} \rightleftharpoons trans\text{-HSNO}$ faces a high barrier of 6.7 kcal/mol (see Figure S1), isomerization in the gas phase is rather unlikely once either isomer is formed.

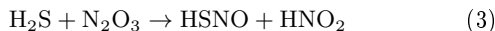
Since the reaction between H₂S and NO to yield HSNO is calculated to be highly endothermic ($\Delta G > 60$ kcal/mol, see Supplementary Information), another formation pathway must be operative under our experimental conditions. We have thus undertaken experiments and calculations to understand the formation chemistry of HSNO and its energetics.

Figure 3 shows how the abundance of several NO-bearing species changes upon addition of H₂S. NO is known to slowly disproportionate on metallic surfaces^{28–30} into N₂O₃ and N₂O ($\Delta G_{(1)+(2)} = -24.4$ kcal/mol³¹) via the reactions:



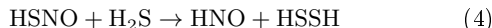
which results in the presence of these species in our experiment. N₂O₃ also appears to be a competent nitrosylating agent *in vitro* since its reaction with RSH has been demonstrated to occur at a rate exceeding that of its hydrolysis,^{19,32} and has been shown to be produced *in vivo*.³⁵ From Fig. 3, it is clear that the production of HSNO is strongly correlated with a decrease in the concentration of N₂O₃ and a commensurate increase in HNO₂, while the concentration of N₂O remains relatively constant. Under the range of the experimental conditions studied here, up to 0.25% of NO is converted into *trans*-HSNO, an abundance presumably limited by the availability of N₂O₃.

Energy calculations performed in this study conclude that N_2O_3 can react exothermically with H_2S ($\Delta H = -7.5$ kcal/mol at 298 K) to form HSNO via the reaction:



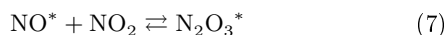
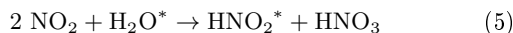
Reaction 3 is analogous to the well-known nitrosation of thiols by N_2O_3 in aqueous solution ($\text{RSH} + \text{N}_2\text{O}_3 \rightarrow \text{RSNO} + \text{NO}_2^- + \text{H}^+$).³² However, this reaction is kinetically improbable under our experimental conditions due to a high activation energy ($\Delta H^\ddagger = 15.9$ kcal/mol at 298 K, see Figure S4). Instead, a surface reaction is almost certainly yielding HSNO. Crucial confirmation that reaction (3) occurs in our experiment is the nearly perfect anti-correlation between the abundances of HNO_2 and N_2O_3 upon addition of H_2S (Figure 3). Furthermore, the maximum concentration of HSNO is reached when the N_2O_3 concentration is close to zero, ostensibly because the steady-state supply of N_2O_3 has been exhausted. Finally, although the production of *trans*-HSNO is remarkably high, it is an order of magnitude smaller than HNO_2 , a possible indication of the chemical instability of HSNO once formed. Weak signal attributed to *trans*-HSNO, the ground state isomer of *trans*-HSNO (see Supplementary Information), has been detected, although an order of magnitude weaker than lines of its metastable isomer. None of the other structural isomers of the [H,S,N,O] system were observed, suggesting that structural reorganization is relatively unimportant in the disappearance of HSNO.

The reactivity of HSNO can be inferred from figure 3, which illustrates that the addition of H_2S beyond the amount resulting in the maximum concentration of HSNO also induces its decay. This results implies that HSNO is unstable with respect to H_2S which would be consistent with the reaction:



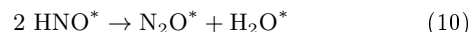
Support for this reaction in solution is by the treatment of S-nitrosoglutathione (RSNO) with an excess of glutathione (RSH) which is known to result in the observation of nitrous oxide, a known decomposition product of HNO.³³ Furthermore, it has been demonstrated that colocalized H_2S and NO result in the generation of HNO which triggers a neuroendocrine signalling cascade.³⁴ Unfortunately, neither product, HNO or HSSH, can be monitored in our apparatus (rotational transitions for the aforementioned molecules cannot be probed in the accessible spectral region) and thus this reaction cannot be directly confirmed.

Addition of a single drop of H_2^{18}O to the gas manifold containing NO resulted in ^{18}O incorporation into N_2O_3 and HNO_2 , but not N_2O (see Table S4), which is consistent with the reactions:



where asterisks indicate possible ^{18}O labeled positions. Once H_2S was mixed with this gas mixture, HSN^{18}O and, interestingly, N_2^{18}O were detected in significant amounts. The latter suggests that N_2^{18}O was formed through decomposition of HN^{18}O , which would be derived from the decomposition

of HSN^{18}O :



A detailed scheme of reactions (5–10) is presented in the supplementary material. The observation of N_2^{18}O is thus an indirect confirmation of reaction (4).

To determine the role of metallic surfaces in the formation of HSNO, high surface area stainless steel mesh has been inserted in the nozzle source, directly before the gas is expanded into the vacuum chamber, resulting in an enhancement in the production of HSNO (by more than a factor of two). In contrast, placement of this mesh farther upstream in the gas line resulted in significant reduction of the HSNO signal (to near depletion), indicating that the mesh, or metallic surfaces, are crucial for forming HSNO, but also that HSNO is only able to diffuse roughly 1-5 centimeters at near atmospheric pressure before most of the molecules have decomposed. Realizing that HSNO has a limited lifetime under our experimental conditions, a new nozzle assembly was designed such that the two gas streams, one containing NO (and N_2O_3) and the other H_2S , could be mixed just prior to expansion. This modified valve design combined with the stainless steel mesh resulted in a 10-fold signal enhancement of HSNO in comparison to the original experimental setup (up to a concentration of 3% relative to NO, see Figure S5). As a side note, under these experimental conditions *trans*-HSNO was not detected which suggests the isomerization from HSNO to HNSO is a slow process.

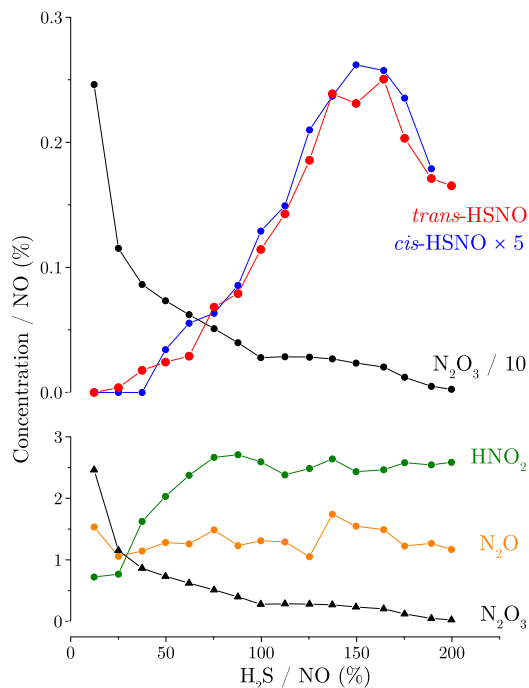
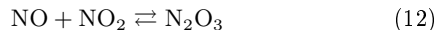
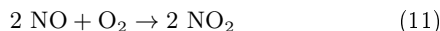


Figure 3. Change in the concentration of several NO containing molecules upon addition of H_2S to the NO gas stream.

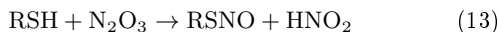
Since N_2O_3 appears to be the limiting reagent in HSNO formation, means were taken to increase its concentration prior to mixing with H_2S . The use of a 1:1 mixture of NO and NO_2 resulted in a five-fold enhancement in the amount of N_2O_3 , by analogy with reaction (7), which correspondingly increased the amount of HSNO. More efficiently, ad-

dition of O₂ to the NO line in a 1:7 ratio yielded a 10-fold enhancement of N₂O₃:



Under these conditions, up to 30% of the NO was converted to HSNO. Compared to the initial experimental conditions, this configuration resulted in a signal enhancement of two orders of magnitude for HSNO (see Figure S5).

A preliminary experimental investigation of the CH₃SH + NO system has also been undertaken. Under experimental conditions similar to the present study, CH₃SNO is formed in high abundance, an indication that the chemical reactions studied here can be generalized to larger RSNO compounds:



A direct confirmation of reaction (4) could be obtained by studying this system for R = CH₃ since the possible products CH₃NO, CH₃SSH, and CH₃SSCH₃ can be measured in our apparatus.

In summary, we find that when dilute samples of H₂S and NO are mixed at room temperature, HSNO is spontaneously formed in high abundance by metallic surface reactions. Precise geometrical structures of both *cis* and *trans* conformers, derived from a combination of highly sensitive microwave spectroscopy and quantum chemical calculations, yield a S–N bond of unusual length, 1.834 Å and 1.852 Å respectively, at least 0.1 Å longer than a typical S–N bond. The extreme length of this bond coupled with its low bond dissociation energy (30.4 kcal/mol at 298 K, see SI) may have important consequences for the proclivity of the molecule to undergo S–N cleavage, either homolytic or heterolytic. Additional experiments and calculations described in this work implicate N₂O₃ as the key reaction partner with H₂S. **It should however be noted that NO and H₂S may also react in vivo under anaerobic conditions (without the intermedicity of N₂O₃) and through different reaction pathways generate SSNO[−] via HSNO,¹⁰ highlighting the complexity of vasodilation.** We show that, once formed, HSNO can undergo further reaction with H₂S most likely producing HNO and HSSH. The present work may also provide a simple and general scheme by which larger RSNOs can be studied in the gas phase. **Although our experimental conditions are markedly different from physiological ones, results from this study may have important biochemical consequences as they show that the production of HSNO from NO and H₂S starting materials is a viable process.** Furthermore, this work implicates that the ‘cross-talk’ observed between H₂S and NO is mediated by HSNO and manifested as HNO.

Acknowledgement The authors are thankful to Ivana Ivanović-Burmazović for an insightful and helpful discussion on the biochemistry of HSNO. The experimental work is supported by NASA grant NNX13AE59G. This material is based upon work supported by the National Science Foundation under CHE-1362118. M. A. M.-D. and K. N. C. were supported by a CfA Postdoctoral Fellowship from the Smithsonian Astrophysical Observatory, and C. C. W. by the Camille and Henry Dreyfus Foundation Postdoctoral Program in Environmental Chemistry. S. T. gratefully acknowledges support by the Deutsche Forschungsgemeinschaft (DFG) through Grant TH 1301/3-2. J. F. S. and T. L. N. were supported by the US Department of Energy, Office of Science, Basic Energy Science [award number DE-

FG02-07ER1588].

Supporting Information Available: Provided are full details of experimental procedures together with results from quantum chemical calculations. This material is available free of charge via the Internet at <http://pubs.acs.org/>.

References

- (1) Moore, P. K.; Bhatia, M.; Mochhala, S. *Trends Pharmacol. Sci.* **2003**, *24*, 609–611.
- (2) Filipovic, M. R.; Miljkovic, J. L.; Nauser, T.; Royzen, M.; Klos, S.; Shubina, T.; Koppenol, W. H.; Lippard, S. J.; Ivanovic-Burmazovic, I. *J. Am. Chem. Soc.* **2012**, *134*, 12016–12027.
- (3) Lipton, A. J.; Johnson, M. A.; Macdonald, T.; Lieberman, M. W.; Gozal, D.; Gaston, B. *Nature* **2001**, *413*, 171–174.
- (4) Lipton, S. A. *Nature* **2001**, *413*, 118–121.
- (5) Berger, P. J.; Skuza, E. M.; Brodecky, V.; Wilkinson, M. H. *Nature* **2002**, *419*, 686–686.
- (6) van Faassen, E.; Vanin, A. F. In *Radicals for Life*; Faassen, E. V., Vanin, A. F., Eds.; Elsevier: Amsterdam, 2007; pp 173 – 199.
- (7) Yang, Y.; Loscalzo, J. In *Radicals for Life*; Faassen, E. V., Vanin, A. F., Eds.; Elsevier: Amsterdam, 2007; pp 201 – 221.
- (8) Singh, R. J.; Hogg, N.; Joseph, J.; Kalyanaraman, B. *J. Bio. Chem.* **1996**, *271*, 18596–18603.
- (9) Whiteman, M.; Li, L.; Kostetski, I.; Chu, S. H.; Siau, J. L.; Bhatia, M.; Moore, P. K. *Biochem. Biophys. Res. Commun.* **2006**, *343*, 303–310.
- (10) Cortese-Krott, M. M. et al. *Proc. Natl. Acad. Sci. U. S. A.* **2015**, *112*, E4651–E4660.
- (11) Filipovic, M. R.; Eberhardt, M.; Prokopovic, V.; Mijuskovic, A.; Orescanin-Dusic, Z.; Reeh, P.; Ivanovic-Burmazovic, I. *J. Med. Chem.* **2013**, *56*, 1499–1508.
- (12) Wrobel, A. T.; Johnstone, T. C.; Liang, A. D.; Lippard, S. J.; Rivera-Fuentes, P. *J. Am. Chem. Soc.* **2014**, *136*, 4697–4705, PMID: 24564324.
- (13) Zhou, X.; Lee, S.; Xu, Z.; Yoon, J. *Chem. Rev.* **2015**, *115*, 7944–8000, PMID: 25651137.
- (14) Müller, R.; Nonella, M.; Russegger, P.; Huber, J. *Chem. Phys.* **1984**, *87*, 351–361.
- (15) Nonella, M.; Huber, J. R.; Ha, T. K. *J. Phys. Chem.* **1987**, *91*, 5203–5209.
- (16) Bharatam, P. V.; Amita; Kaur, D.; Kumar, P. S. *Int. J. Quantum Chem.* **2006**, *106*, 1237–1249.
- (17) Miljkovic, J. L.; Kenkel, I.; IvanoviÄĀ-BurmazoviÄĀ, I.; Filipovic, M. R. *Angew. Chem. Int. Ed.* **2013**, *52*, 12061–12064.
- (18) Zhang, S.; Melzer, M. M.; Sen, S. N.; Çelebi Ölçüm, N.; Warren, T. H. *Nat. Chem.* **2016**, *8*, 663–669.
- (19) Cortese-Krott, M. M.; Fernandez, B. O.; Kelm, M.; Butler, A. R.; Feelisch, M. *Nitric Oxide* **2015**, *46*, 14 – 24.
- (20) Baciu, C.; Gauld, J. W. *J. Phys. Chem. A* **2003**, *107*, 9946–9952.
- (21) Timerghazin, Q. K.; Peshherbe, G. H.; English, A. M. *Phys. Chem. Chem. Phys.* **2008**, *10*, 1532–1539.
- (22) Ivanova, L. V.; Anton, B. J.; Timerghazin, Q. K. *Phys. Chem. Chem. Phys.* **2014**, *16*, 8476–8486.
- (23) Bartberger, M. D.; Houk, K. N.; Powell, S. C.; Mannion, J. D.; Lo, K. Y.; Stamler, J. S.; Toone, E. J. *J. Am. Chem. Soc.* **2000**, *122*, 5889–5890.
- (24) Bartberger, M. D.; Mannion, J. D.; Powell, S. C.; Stamler, J. S.; Houk, K. N.; Toone, E. J. *J. Am. Chem. Soc.* **2001**, *123*, 8868–8869.
- (25) Timerghazin, Q. K.; English, A. M.; Peshherbe, G. H. *Chem. Phys. Lett.* **2008**, *454*, 24–29.
- (26) Crabtree, K. N.; Talipov, M. R.; Martinez, J.; Oscar; O’Connor, G. D.; Khursan, S. L.; McCarthy, M. C. *Science* **2013**, *342*, 1354–1357.
- (27) Womack, C. C.; Martin-Drumel, M.-A.; Brown, G. G.; Field, R. W.; McCarthy, M. C. *Sci. Adv.* **2015**, *1*, e1400105.
- (28) Melia, T. P. *J. Inorg. Nucl. Chem.* **1965**, *27*, 95–&.
- (29) Lim, M. D.; Lorkovic, I. M.; Ford, P. C. *Methods Enzymol.* **2005**, *396*, 3–17.
- (30) Glendening, E. D. *J. Chem. Phys.* **2007**, *127*, 164307.
- (31) Wagman, D. D.; Evans, W. H.; Parker, V. B.; Schumm, R. H.; Halow, I.; Bailey, S. M.; Churney, K. L.; Nuttall, R. L. *J. Phys. Chem. Ref. Data* **1982**, *11*.
- (32) Keshive, M.; Singh, S.; Wishnok, J. S.; Tannenbaum, S. R.; Deen, W. M. *Chem. Res. Toxicol.* **1996**, *9*, 988–993.
- (33) Wong, P. S. Y.; Hyun, J.; Fukuto, J. M.; Shirota, F. N.; DeMaster, E. G.; Shoeman, D. W.; Nagasawa, H. T. *Biochemistry* **1998**, *37*, 5362–5371.
- (34) Eberhardt, M. et al. *Nat Commun* **2014**, *5*, –.
- (35) Basu, S. et al. *Nat. Chem. Biol.* **2007**, *3*, 785–794.

Graphical TOC Entry

