UCSF UC San Francisco Previously Published Works

Title

Biomaterials in fetal surgery

Permalink

https://escholarship.org/uc/item/9p58x0vp

Journal Biomaterials Science, 7(8)

ISSN

2047-4830

Authors

Winkler, Sally M Harrison, Michael R Messersmith, Phillip B

Publication Date

2019-08-01

DOI

10.1039/c9bm00177h

Peer reviewed



HHS Public Access

Author manuscript *Biomater Sci.* Author manuscript; available in PMC 2021 June 25.

Published in final edited form as:

Biomater Sci. 2019 August 01; 7(8): 3092–3109. doi:10.1039/c9bm00177h.

BIOMATERIALS IN FETAL SURGERY

Sally M. Winkler^{1,2}, Michael R. Harrison³, Phillip B. Messersmith^{*,1,4,5}

¹.Department of Bioengineering, University of California, Berkeley, CA, USA

² University of California, Berkeley–University of California, San Francisco Graduate Program in Bioengineering, Berkeley, CA, USA

³ Division of Pediatric Surgery, UCSF Benioff Children's Hospital, San Francisco, CA, USA

⁴.Department of Materials Science and Engineering, University of California, Berkeley, CA, USA

⁵ Materials Sciences Division, Lawrence Berkeley National Laboratory, Berkeley, CA, USA

Abstract

Fetal surgery and fetal therapy involve surgical interventions on the fetus *in utero* to correct or ameliorate congenital abnormalities and give a developing fetus the best chance at a healthy life. Historical use of biomaterials in fetal surgery has been limited, and most biomaterials used in fetal surgeries today were originally developed for adult or pediatric patients. However, as the field of fetal surgery moves from open surgeries to minimally invasive procedures, many opportunities exist for innovative biomaterials engineers to create materials designed specifically for the unique challenges and opportunities of maternal-fetal surgery. Here, we review biomaterials currently used in clinical fetal surgery as well as promising biomaterials in development for eventual clinical translation. We also highlight unmet challenges in fetal surgery that could particularly benefit from novel biomaterials, including fetal membrane sealing and minimally invasive myelomeningocele defect repair. Finally, we conclude with a discussion of the underdeveloped fetal immune system and opportunities for exploitation with novel immunomodulating biomaterials.

Graphical Abstract



^{*}Corresponding Author. Phillip B. Messersmith, PhD, University of California-Berkeley, 210 Hearst Memorial Mining Building, Berkeley, CA 94720-1760, Phone (510)643-9631, Fax (510)643-5792, philm@berkeley.edu. Contributions

S.M.W. wrote the manuscript with input from P.B.M. and M.R.H.

Declarations of interest: none.

Keywords

fetal surgery; biomaterials; congenital diaphragmatic hernia; fetal membranes; myelomeningocele; pediatric surgery

Introduction

Since the first successful surgery on a human fetus in the 1980s [4], fetal surgery has evolved from high-risk open surgeries, in which the uterus is opened, and the fetus partially exposed and operated on, to fetoscopic, minimally invasive procedures in which instruments or needles are inserted through small incisions in the mother's abdomen. The history, state of the art, and future potential of minimally invasive fetal surgery were expertly reviewed by Graves and colleagues [5]. While the number of fetal surgeries has increased, and the surgical techniques used in fetal surgery have advanced in the past 30 years, the materials used in these procedures have not seen such progress. For the most part, the materials and devices utilized in fetal surgeries have been modified from existing devices already in use in adults or neonates. The development of biomaterials tailored for fetal surgery represents a significant opportunity for biomaterials engineers to address an unmet clinical need. Furthermore, the immune privilege and relatively short duration of pregnancy present unique materials requirements and opportunities for fetal treatment compared to biomaterials used in adult patients.

For a patient to be a candidate for fetal surgery, the potential benefits of the surgery must outweigh the risk to the fetus and mother. Given the current limitations of fetal surgery, especially the risk of membrane rupture and subsequent preterm birth, surgical procedures have been limited to those conditions for which no intervention would mean fetal or perinatal death or loss of limb or organ function. For fetal surgery to be considered in a specific case, the fetus must be affected enough to merit intervention, but not so severely affected that the intervention would not improve chances of survival. Modalities for determining fetal health status include ultrasound, amniocentesis and other genetic diagnostics, fetal MRI, and fetal echocardiogram. Criteria to stage the severity of a condition, such as lung area to head circumference ratio measurements in congenital diaphragmatic hernia, should be established before considering performing fetal surgeries to address that condition. Additionally, ample pre-clinical evidence of an intervention's success in animal models is necessary prior to deploying new strategies on human patients; animal models of fetal surgery were recently reviewed by Kabagambe and colleagues [6]. Patientspecific inclusion and exclusion criteria vary depending on the type of surgery being performed, and a representative set of criteria first established in 1982 [1] and still utilized today [2] is shown in Box 1.

Maternal-fetal surgery also raises important ethical issues. These include establishment of clinical equipoise in clinical trials, determining "patienthood" in the context of maternal-fetal surgery, resisting the "urge to intervene," and ensuring accurate representation of surgery's risks and benefits to patients, especially in the context of potential fetal or neonatal palliative care [7–12]. Clinicians and engineers should take care that, whenever possible,

potential fetal treatments are evaluated in the context of a randomized trial with multispecialty clinical expertise. Due to small patient populations (made smaller by the fact that mothers may choose conservative management or pregnancy termination rather than enroll in a trial), many of the most successful randomized controlled trials are conducted as multicenter or even multi-national trials. Examples of such multi-center trials include Management of Myelomeningocele Study (MOMS), which compared outcomes from fetal vs. neonatal repair of myelomeningocele (spina bifida) neural tube defects [3]; the percutaneous vesicoamniotic shunting versus conservative management for fetal lower urinary tract obstruction (PLUTO) trial, which compared *in utero* bladder shunting to conservative management for urinary tract obstructions in utero [13, 14]; and a study from the EuroFoetus consortium that compared laser ablation of placental anastomoses versus serial amnioreduction for the treatment of twin-twin transfusion syndrome (TTTS) [15].

The use of biomaterials in fetal surgery has begun to improve clinical outcomes for human patients (Figure 1), for example the use of balloons for tracheal occlusion in congenital diaphragmatic hernia (Figure 1a) [16] and bladder shunts to drain lower urinary tract obstructions *in utero* (Figure 1b) [17]. Other biomaterials solutions are being actively explored in large animal models with promising results, such as patches to prevent damage to abnormally exposed neural tissue in cases of spina bifida (myelomeningocele, Figure 1c) [18]. Yet other materials are being developed and investigated in laboratory settings, such as novel fetal membrane sealant materials (Figure 1d) [19]. Efforts to design and test materials specifically for fetal surgery will not only spur materials innovation in an underexplored area of medicine but also provide the best chance at a healthy life for fetuses and infants with otherwise grim prognoses.

Risk in fetal surgery

Risks associated with fetal surgery can be significantly reduced by decreasing the invasiveness of the procedure [20]. Recent trends in fetal surgery have transitioned away from open fetal surgery, in which the fetus is delivered in a partial Cesarean section (without disruption of placental blood supply), an intervention is performed, and the fetus is returned to the uterus for the duration of pregnancy [3]. In Fetoscopic surgery (Fetendo), small ports are placed in the amniotic space, an endoscope projects the image on a screen, and the surgeon manipulates small diameter instruments under endoscopic guidance to accomplish the surgery. Most Fetendo procedures are done percutaneously, but some require maternal laparotomy. Finally, in image guided fetal surgery, small instruments or needles deliver therapy or perform minor interventions under ultrasound guidance. Such minimally invasive interventions are often termed fetal therapy, and include fetal blood or stem cell transfusion [21], injection or aspiration of amniotic fluid [15], and some laser ablation procedures [22]. Instruments and devices used in fetal surgery and fetal therapy were expertly reviewed by Klaritsch and colleagues [23]. Most fetal procedures are performed in the first or second trimester of pregnancy, and many before the limit of viability (about 25 of a full 40 weeks of gestation) [3]. In its current form, fetal surgery still presents a significant risk to the fetus; thus, it is only performed on those fetuses at-risk for fetal or neonatal demise or loss of limb or organ function, but healthy enough that they will likely benefit from intervention. Major risks are discussed below.

Maternal morbidity in fetal surgery—Maternal morbidity is generally low in cases of fetal surgery and the underlying conditions that require fetal surgery. No maternal deaths following open fetal surgery have been reported [7, 24], but potential serious maternal sequalae of fetal surgery include placental abruption, premature rupture of membranes, premature birth, chorioamnionitis (infection of the membranes), loss of ability to carry future pregnancies, and sepsis [5]. Following open fetal surgery, delivery of the fetus in any future pregnancy must be via Caesarian section [25].

Fetal membrane rupture and preterm birth—In any fetal surgery, as well in fetal therapies and some diagnostic procedures like amniocentesis [26], the fetal membranes (amniotic sac) must be breached. The fetal membranes (FM) are the amnion and chorion, two closely associated membrane tissues that line the uterus and enclose the fetus and amniotic fluid during gestation (Figure 1d). They are largely avascular and do not heal after rupture or surgical cut or puncture [27-30], though recent evidence suggests collagen remodeling may contribute to a small degree of re-sealing [31]. This breaching of the FM is what makes fetal surgery so risky, as it can lead to premature preterm rupture of membranes (PPROM) and preterm birth and its associated sequelae [32, 33]. Today, most fetal surgeries are performed via a minimally invasive approach; nonetheless iatrogenic (surgically caused) PPROM (iPPROM) occurs in about 30% of fetoscopic fetal surgery cases, but this rate varies from 11–50% depending on the intervention or practitioner [20, 34, 35]. iPPROM during the intervention is rare; iPPROM usually happens in the days or weeks following surgery, up to the 37th week, after which time the pregnancy is considered term [20]. Most fetal surgeries are performed during the second trimester, making the risk of iPPROM particularly high as fetuses are unable to survive outside the uterus prior to the limit of viability (25 weeks) [3], and delivery before 32 weeks' gestation carries significant fetal and neonatal risk including perinatal death [36]. iPPROM has accordingly been deemed the "Achilles heel" of fetal surgery; several materials strategies for reducing iPPROM have been investigated (detailed in Membrane Sealing section, below), but no viable solution has been widely adopted. A solution to reduce iPPROM incidence would drastically decrease the overall risk of all fetal surgeries and make fetal surgery a viable option for more patients [20, 351.

Materials and devices in fetal surgery

Materials solutions for preventing fetal membrane rupture—To access the fetus, fetal surgeons necessarily must puncture the fetal membranes, and this puncture site can later rupture, leading to preterm birth. Over the past few decades, rates of neonatal survival and survival without impairment following preterm birth have remained steady, and the limit of viability relatively unchanged, indicating that the limits of post-natal intervention may have been reached [36]. For fetuses that undergo fetal surgery, there is still an unmet clinical need to keep the fetuses in the uterus until at least 37 of 40 weeks' gestation and to reduce the instances of membrane rupture and preterm birth.

Some attempted strategies for fetal membrane repair following fetal surgery (see Table 1) include mixtures of maternal platelets and fibrin cryoprecipitate with and without dry collagen/gelatin plugs ("amniopatch") [26, 37–39], synthetic polymer sealants [40, 41], laser

welding [42], scaffold-type plugs manufactured directly from decellularized amnion [28, 43–45], and tissue engineering approaches [46]. These have had limited success, and no clear pathway to a clinically viable solution has emerged after more than a decade of research. These strategies rely on depositing a material at or near the defect site after the membranes have been punctured surgically. However, recent research on benchtop models of the fetal membranes suggests that applying an adhesive sealant material to the space between the fetal membranes and the uterus prior to surgical membrane puncture can help stabilize membranes during surgery, decrease the size of surgical membrane defects, maintain a watertight seal of the membranes during and after surgery, and decrease the probability of catastrophic membrane rupture [5, 47, 48]. Future development of adhesives for fetal membrane sealing could potentially use a seal-then-puncture membranes strategy ("presealing") with great success. We contend that an ideal material solution for membrane sealing may have some of the following properties: have similar mechanical properties to the fetal membranes, be fluid impenetrable, be nonimmunogenic and not cause an adverse tissue response, maintain adhesive and/or mechanical properties for an appropriate timescale to extend pregnancy (e.g., 4 weeks or up to 24 weeks), stabilize the membranes during and after surgery, be resistant to biofouling, or encourage cellular regrowth when applicable.

Some sealants and adhesives initially designed to seal tissues elsewhere in the body have been studied in benchtop, animal, or clinical models of fetal surgery [19, 49]; many such materials are summarized in Table 1, along with other materials designed specifically for fetal membrane sealing. Thus far, no material has become widely accepted as a clinical solution for fetal membrane sealing, and there exists an opportunity for biomaterials engineers to design or identify a material capable of strong, robust adhesion in the wet and biologically sensitive amniotic space.

Patches in Fetal Surgery

Myelomeningocele: One of the biggest success stories for fetal surgery is in the treatment of severe spina bifida, or myelomeningocele (MMC). Spina bifida affects about 3.5 per 10,000 live births in the US [65], and 25–40% of MMC-affected fetuses are aborted [66, 67]. Briefly, in this condition, the skin and vertebrae do not fully form around the lower portions of the spinal cord. Children with myelomeningocele often have limited lower limb function, develop Arnold-Chiari II malformations (hindbrain herniation), and accumulate excess cerebrospinal fluid in their brains (hydrocephalus), which often requires repeated shunting to drain the fluid throughout the patient's lifetime [3]. The development and progression of MMC follows the two-hit hypothesis where the first "hit" is the failure of the neural tube to become fully enclosed and the second "hit" is the damage to spinal tissue that occurs during gestation due to degradation by enzymes in the amniotic fluid [68, 69]. Fetuses with myelomeningocele can sometimes move their lower limbs during early gestation, but they are often born with total or partial loss of lower limb function because enzymes in the amniotic fluid degrade the spinal cord tissue [70]. It was hypothesized that repairing the defect *in utero* would protect it from such degradation. A large, multi-site clinical trial, the Management of Myelomeningocele Study (MOMS), demonstrated the benefits of open fetal surgery for myelomeningocele repair compared to traditional postnatal repair: increased use of lower limbs and decreased need for cerebrospinal fluid (CSF) shunting due to

hydrocephalus. Risks of the repair surgery included membrane rupture leading to preterm birth and its associated complications [3]. In the MOMS trial, most cases were performed via an open surgical access technique, and the fetus's skin surrounding the spinal cord defect was stretched to cover the exposed neural tissue and sutured in place. This trial, as well as the preceding animal trials and human case studies, was excellently reviewed by Adzick [68] and the state of the field of *in utero* repair of MMC defects in the post-MOMS era was reviewed by Moldenhauer and Adizick [71].

Researchers have also investigated endoscopic and other minimally invasive approaches to tissue closure of MMC defects [72, 73]; tissue engineering approaches towards minimally invasive MMC repair were reviewed by Watanabe, et al. [18]. Recent animal data suggest the potential for the use of materials to aid in closure of MMC defects and to isolate exposed neural tissue from AF enzymes and from surrounding tissues to prevent spinal cord tethering and its long-term sequelae [18, 74–82]. Table 2 describes some of this preliminary work. Materials utilized as scaffolds and/or defect coverings for in utero MMC defect repair in animal models include collagen- or gelatin-based scaffolds, small intestinal submucosa, and polymeric materials including silicone, high density poly ethylene, and polypropylene [18]. Covering MMC defects with a biomaterial could drastically reduce the FM defect size necessary to perform MMC closures compared to open surgery. In one example of biomaterials use for MMC defect coverage, Watanabe and colleagues used an ovine model of *in utero* myelomeningocele [83]. Fetal lambs with surgically-created MMC defects were treated with gelatin or gelatin-collagen sponges laced with bFGF (basic fibroblast growth factor) secured around the defect site with Dermabond cyanoacrylate adhesive with or without a gelatin sheet atop the sponge. Though all sheets detached from the defect site, sponges remained, and compared with sham-operated control animals, treated animals had less hindbrain herniation and more neural tube coverage. Animals treated with bFGF-laced sponges had more granulation and epithelial tissue covering the neural tube compared to non-bFGF controls. This work suggests the potential for eventual clinical translation, though more studies are required to assess the toxicity of the materials used, perfect minimally invasive surgical technique, improve or maintain a water-tight seal, and investigate the potential for spinal cord tethering at the defect site.

Gastroschisis and omphalocele: Gastroschisis and omphalocele are abdominal wall defect conditions that are promising targets for fetal surgery. In gastroschisis, muscles of the fetal abdomen do not fully close around the internal organs, and the abdominal organs are exposed to the amniotic fluid [89]. Omphalocele is a similar condition, except that the organs are surrounded by a thin sac and not exposed directly to amniotic fluid. These conditions are diagnosed and staged *in utero* using ultrasound imaging, with more severe cases presenting with a larger defect and more organs developing outside the abdominal cavity. Most patients have good outcomes following post-natal intervention. However, in 10–20% of fetal gastroschisis patients, prolapsed organs experience long-term damage. Experiments in fetal sheep demonstrate that intestinal damage at birth is likely due to restricted blood flow and enzymatic degradation of the tissues by the amniotic fluid [90–92]. Gastroschisis cases with severe intestinal evisceration are also at risk for oligohydramnios (insufficient amniotic fluid) [89]. Animal models of gastroschisis have been established in

fetal chickens [93], rats [94], rabbits [95], and sheep [84, 90, 96]. Biomaterial patches developed for myelomeningocele may also be adapted for use treating gastroschisis or omphalocele. However, minimally invasive large animal models must first be developed to confirm the efficacy and improve the safety of procedures to deploy biomaterials tissue patches to cover gastroschisis or omphalocele defects in utero before they are attempted in human patients [97].

Materials considerations for tissue patch materials: Several considerations must be made in the development of biomaterials for direct application to internal fetal tissues, such as in the repair of myelomeningocele or gastroschisis defects or, potentially, sealing of FM defects. Materials should be deliverable via a minimally invasive surgical approach, for example a liquid adhesive that cures *in situ* or a patch that can be rolled up to fit into a 4 mm trocar. As these materials will be in direct contact with internal fetal organs, cytocompatibility is an extremely important consideration. Materials should be able to accommodate the fetus's growth throughout pregnancy and should isolate the exposed tissue from the surrounding amniotic fluid in a fluid-impenetrable manner. Attention should be paid to the post-natal and long-term role of the materials implanted in *utero*, and decisions about whether to design removable, degradable, or permanent materials should be application- and tissue-specific. Tatu and Lin [98] present a set of materials characterization experiments that should be considered when developing new fetal patch materials or choosing existing materials to use for these applications.

Occlusion and ablation in fetal surgery and fetal therapy

Congenital diaphragmatic hernia: Congenital diaphragmatic hernia (CDH) occurs in about 1 in 3000 live births when the diaphragm fails to form properly during development, allowing the liver, intestines, stomach, and/or other abdominal organs to invade the lung cavity. Thus, development of one or both lungs is restricted. Most infants with severe CDH defects undergo corrective surgery after birth, but in the most severe cases, prenatal treatment is considered to increase the lung volume of affected neonates at birth. Fetal lung area to head circumference ratio (LHR) is used to stage the severity of CDH, with severely affected fetuses having lower LHRs [99]. Saxena expertly reviewed the materials used for post-natal repair of large congenital diaphragmatic hernia defects [100]; none of the materials used were developed specifically for CDH repair, a trend also seen in fetal surgery and in the medical device industry in general. Jeanty, et al., reviewed non-surgical strategies with promise to address CDH in utero, including stem cell and pharmacologic methods [101], and Eastwood reviewed strategies that have been pursued in *in utero* animal models for reducing pulmonary hypoplasia resulting from CDH [102]. Over the years, several different types of biomaterials have been investigated for use addressing CDH in utero (Figure 2) including polymer fabrics [103], balloons [104], metal clips [99], hydrogels [105], and tissue engineered materials [106]. The first successful in utero CDH repair was reported in 1990 [103]; a Gore-Tex (PTFE fabric) patch was used to repair the diaphragm and another to cover the abdomen (Figure 2a). However, in utero repair of fetal CDH defects did not prove superior to the standard of care, post-natal surgery and monitoring [103, 107].

Eventually, *in utero* hernia repair was replaced with *in utero* tracheal occlusion. Occluding the trachea allows fluid to build up in the lungs, and the lungs expand, pushing the abdominal organs out of lung cavity. Initially, open surgery was performed to clamp Silasticcoated titanium clips around the trachea to occlude it (Figure 2b). This was replaced by the less invasive Fetendo approach, in which a maternal laparotomy is performed to expose the uterus [99, 108, 109]. Then, endoscopic tools were used to place titanium clips around the trachea. In 2005, Deprest and colleagues published the first successful FETO balloon trial, in which an inter-tracheal silicone balloon is deployed laparoscopically and inflated to approximately 2 cm long and 0.5 mm in diameter to occlude the trachea (Figure 2c), eliminating the need for a maternal laparotomy [16, 104]. The timing and removal of tracheal occlusions is an ongoing area of research; titanium clips are removed via a neck incision at birth via an EXIT procedure, whereas balloons can be punctured transcutaneously *in utero* or punctured and removed at birth (vaginal or EXIT). Recently, *in situ* gelating hydrogels have been pursued as an alternative to balloon occlusion. Muensterer and colleagues tested fibrin glue (Tisseel), porcine gelatin, bovine collagen, cyanoacrylate, perfluorocarbon gel, and recombinant thrombin for their abilities to plug tracheal lumens ex vivo and found that fibrin glue performed best [110]. Fibrin glue was further studied in a fetal rabbit model of tracheal occlusion and increased lung mass (beneficial) and airway resistance (detrimental) [110]. Similarly, in another study of fibrin glue on a fetal rabbit tracheal occlusion model, lung performance measures were not improved with tracheal occlusion [105]. Nevertheless, given the minimally invasive potential of this intervention, opportunities exist for further investigation and development of hydrogel sealants for intrauterine tracheal occlusion. Recent animal data suggest that the use of a hydrogel sealant to secure the balloon in the trachea and prevent balloon dislodgement [111] could be a promising strategy. Additional strategies that incorporate biomaterials are currently under development in animal and preclinical studies. In one example, researchers are looking to repair congenital diaphragmatic hernias postnatally with autologous tendon tissue seeded with circulating cells from the amniotic fluid [106].

Monochorionic twin conditions: In twin pregnancies, several circulation-related abnormalities sometimes indicate the use of fetal surgery. In a monochorionic twin pregnancy (about 70% of monozygotic, or identical, twin pregnancies), the fetuses share a placenta, but each has their own amniotic sac [112]. Vessels of the shared placenta sometimes anastomose abnormally, leading to twin-twin transfusion syndrome (TTTS) in 8–10% of monochorionic pregnancies [112]. Blood from one twin (donor) crosses into the other twin (recipient). In TTTS, one twin's heart pumps blood to both twins and causes delayed organ development in the donor twin and polyhydramnios and fetal hydrops (accumulation of excess fluid in fetal organs) in the recipient twin. Untreated, 70–80% of TTTS twins will die, and survivors may have severe organ damage [113]. The standard of care for TTTS is radiofrequency ablation of the vessels connecting the two twins. A 2004 multinational randomized controlled trial of 142 pregnant women with TTTS demonstrated that this method is more effective than serial amnioreduction of the polyhydramniotic sac [15]. Twins treated with laser coagulation (via a 3.3-mm cannula and a neodymium:yttrium– aluminum–garnet or diode laser under fetoscopic guidance) had significantly higher survival

rates and lower rates of neurologic complications. However, survival of at least 1 twin to 6 months was still only 76% in the treatment group.

In Twin Reversed Arterial Perfusion (TRAP), which affects around 3% of monochorionic twin pregnancies, one twin is structurally normal, but, due to aberrant blood vessel formation in the placenta, the other twin lacks a heart and head. Untreated, 50% of normal "pump" twins, whose heart pumps blood both to themselves as well as to their acardiac twin, die *in utero* or as neonates [114]. To treat TRAP, a 1 mm diameter needle is inserted through the maternal abdomen and into the abdomen of the acardiac twin. Radiofrequency ablation is performed through needle to heat and coagulate the vessels of the acardiac twin. This serves to stop blood flow between the twins without exposing the pump twin to the potentially harmful byproducts of the dying acardiac twin. Initial reports suggest a pump twin survival rate of around 90% [115].

Selective intrauterine growth restriction (SIUGR) is another twin abnormality in which unequal sharing of placental blood between monochorionic twins can result in an extreme difference in weight between the twins. In the most severe cases, one twin is drastically underdeveloped and poses a risk to the healthy twin because intrauterine death of the smaller twin could lead to neurological impairment of the healthy twin. In these cases, selective termination of the underdeveloped twin in a way that does not damage the healthy twin is considered [116]. When termination of the non-thriving twin is indicated and desired, similar care is needed to prevent harm to the healthy twin. Radiofrequency ablation is also used in this case, as is bipolar cord coagulation. Bipolar cord coagulation is more invasive (>3mm FM incision) and involves using ultrasound guidance to clamp the umbilical cord of the unhealthy twin and ablate it with radiofrequecy [117, 118].

Shunting in fetal surgery

Fetal Urinary Tract Obstruction: Lower urinary tract obstruction (LUTO) occurs in approximately 1–5 of 10,000 live births when the lower urinary tract fails to develop properly, and urine swells the fetal bladder. This can lead to fluid accumulation in the kidneys or other parts of the renal system (eg. hydronephrosis), renal failure, and oligohydramnios, and is associated with high rates of premature birth and/or perinatal death due to pulmonary hypoplasia (underdeveloped lungs). LUTO affects lung development because amniotic fluid is largely composed of fetal urine; when urinary outflow is obstructed, insufficient amniotic fluid hinders lung maturation. Postnatal repair to address urinary blockage remains the standard of care for this condition, but since severely affected infants often die of pulmonary hypoplasia soon after birth, fetal surgery to place a shunt to allow fluid to flow from the bladder to amniotic fluid during gestation (vesicoamniotic shunting) is an active area of research [119]. Current state of the art for fetal interventions for urinary tract obstructions were recently reviewed by Brock and Clayton [17]. After many case studies in human patients suggested that shunting may improve perinatal outcomes in fetuses with a poor outlook [120–122], a randomized controlled trial was conducted to study LUTO shunting in male fetuses, the percutaneous vesicoamniotic shunting versus conservative management for fetal lower urinary tract obstruction (PLUTO) trial [123, 124]. Though the PLUTO trial was terminated early due to low recruitment, the infants treated

with *in utero* shunting had lower rates of perinatal lung failure-related death. However, shunt treatment did not appear to have a drastic benefit on renal function; two of seven treated and zero of three untreated infants alive at 2 years had normal renal function, respectively. Issues including membrane rupture, shunt dislodging, and shunt blockage were cited as contributing to fetal or infant demise in the treatment group. Further work to develop shunts specifically for fetal bladder shunting is needed.

Unfortunately, little attention has been paid to the materials of the shunts themselves, in urinary tract obstruction, as well as in shunting to address fetal hydrocephalus and drain fetal lungs (see below). In fact, the identity and physicochemical properties of materials used for shunting are often not mentioned in the medical literature. In a report of all 73 fetal obstructive uropathy cases reported to International Fetal Surgery Registry between 1982 and 1985, for example, shunt architecture and materials were not reported [120]. In the PLUTO prenatal urinary tract obstruction clinical trial, a pigtail catheter was inserted with a King's College/Rocket introducer (n = 5) or a Harrison shunting set (n = 10). Double pigtail shunts are commonly used in fetal surgery as they are designed to stay in place and are unlikely to interfere with fetal development. However, a recent analysis revealed that half of shunts used to correct fetal urinary tract obstructions become dislodged [125]. And while shunts have multiple holes on each side, shunt failure due to clogging is common [120, 126, 127]. Future research should focus on the development of shunts with an internal antibiofouling surface coating to reduce biomolecule fouling and clogging of the shunt and valves, and alternative shunt architectures, such as the double basket catheter [128], to ensure the shunt stays in place for the duration of pregnancy.

More than half of fetal lower urinary tract obstructions are caused by posterior urethral valves (PUV) that occlude or block urine flow; this occurs in 1 of 8000 live births [129]. To repair urine flow and reduce fluid buildup in kidneys, some researchers have begun to use fetal cystoscopy and laser ablation to remove the aberrant valves [130]. In a series of 40 cases of diagnostic fetal cystoscopy to enable visualization of PUV formation, 23 fetuses received laser ablation to correct PUV, 14 fetuses survived to birth, and 12 survived with intact renal function. However, in addition to the risks associated with all fetal surgeries (maternal morbidity, preterm birth, spontaneous membrane rupture, etc.), laser fulguration was also associated with the development of urological fistulas (4 fetuses). Fistulae development was found to be associated (P < 0.01) with the materials and instruments used in the procedure, including catheter sheath shape and laser type, power, and energy [131]. Other opportunities for valve disruption for this indication include micro scissors, balloon disruption, or guidewires [131]. At least one case of successful in utero urethral stenting with a 0.9mm stent has been reported in fetus with fetal cystoscopy-confirmed urethral stenosis [132]; the stent material was not reported. In a retrospective analysis comparing fetal cystoscopy and vesicoamniotic shunting in the treatment of severe LUTO cases, Ruano and colleagues found that while both therapies increase the 6-month survival rate, only fetal cystoscopy improves renal function in PUV patients [133]. A randomized controlled trial to compare the efficacy of fetal cystoscopy and vesicoamniotic shunting is planned (trial ID: NCT01552824).

Fetal pleural and pericardial fluid: Similar double-pigtail shunts are also sometimes used *in utero* to drain fluid from the lungs and chest cavity into the amniotic space. Pleural effusion (PE) and macrocystic congenital cystic adenomatoid malformation (CCAM) are rare conditions in which fluid builds up in the pleural sac surrounding the lungs and in cystic lung tissue, respectively [134]. Untreated, severe fetal pleural effusion can have a mortality rate between 57–100% [135]. In a retrospective of 48 cases of fetal hydrothorax (PE of lymphatic fluid) in the Netherlands from 2001–16, overall fetal survival through the neonatal period following *in utero* thoracoamniotic shunting was 75% [136]. A retrospective study from Children's Hospital of Philadelphia (1998–2001) found postnatal survival of treated fetuses to be 67% (6 of 9 fetuses) for PE and 70% (7 of 10 fetuses) for CCAM [134]. These survival rates and rates of adequate lung function following fetal thoracoamniotic shunt placement are similar to those first reported in 1988 – 75% survival of 8 treated fetuses [135]. Many fetuses with fluid accumulation in the chest will not need fetal surgery, but the procedure is considered when the fetus develops severe fetal hydrops and/or the fluid accumulation constricts surrounding organs.

Fetal Hydrocephalus: Severe cerebrospinal fluid (CSF) buildup in the ventricles of the fetal brain (ventriculomegaly) can delay the development of other brain structures and often requires post-natal placement of a ventricoperitoneal shunt to drain excess CSF to the abodmen. These shunts are prone to infection and clogging, and serial shunt replacement throughout the child's lifetime is often required. Some cases of fetal vesicoamniotic shunting (between brain ventricles and AF) have been reported in human patients [120, 137, 138], though overall prognoses remain grim. One factor that contributes to poor outcomes in these fetuses is that most cases of hydrocephalus are accompanied by co-morbidities including neural tube defects, oligo- or polyhydramnios, and other congenital abnormalities [138]. A 2014 report of 222 cases in Poland of fetal hydrocephalus repair conducted between 1992-2012 used Orbis-Sigma and Accu-Flow valves and Cook's shunts to drain fluid from the ventricles into the amniotic sac. In this study, 44% of neonates were pretern, and only 12.5% had normal mental development at age 3 [139]. The study demonstrated that fetal shunting decreased ventricular size, but as this was not a randomized trial, it cannot be fully established that this treatment is better than the standard of care. Other case studies show similarly inconclusive results [126], and a retrospective case study suggests that fetal shunting results in higher rates of severe neurological impairment [140]. A voluntary moratorium against in utero shunting for fetal hydrocephalus has since been imposed until more information about the natural progression of fetal hydrocephalus could be established. In many of these early studies, patient selection was poor as it was difficult to identify which fetuses may benefit most from intervention, however improved fetal diagnostic methods may allow for advances fetal surgery for hydrocephalus in the future [140].

Immune tolerance and exploitation in fetal surgery and fetal therapy

Biomaterial interaction with the innate and adaptive immune systems has long been an area of research in adult patients, but less is known about the response of the fetal immune system to implanted biomaterials. The fetal immune system develops throughout gestation and continues to develop after birth; preterm infants are likely to be born with more immature immune systems, making them especially susceptible to bacterial or viral

infections [141]. A certain degree of fetal immune tolerance or immaturity is necessary to accommodate the presence of maternal alloantigens in the fetal circulation [141, 142]. However, recent evidence in mouse models suggests that fetal interventions, including fetal surgery, increase trafficking of maternal T cells to the uterus and increase maternal T cell recognition of the fetus. This trafficking could contribute to adverse outcomes like preterm birth and immune-mediated fetal demise [143, 144]. Nonetheless, the fetus's lack of a complete immune system could present a unique opportunity for biomaterials development. For example, the complement activation system is incomplete; circulating complement factors in newborns are 10–80% lower than in adults [141]. To our knowledge no study has set out to specifically address questions of long-term biomaterial interactions in the fetus. However, in studies in which materials were implanted in human or animal fetuses, little to no evidence of negative immune response (inflammation, fibrous capsule formation, foreign body response, etc.) was detected, though analysis of tissue-material interactions are underreported in the clinical fetal surgery literature [3, 13, 59, 83]. Though further investigation is needed, it seems that some immune responses elicited by implanted biomaterials are less pronounced in fetuses, possibly creating a more permissive environment for biomaterials in the fetal patient.

Researchers have begun to take advantage of the immature fetal immune system to develop stem cell treatments for alpha thassalemia major (ATM, Hemoglobin Bart's) and other inherited genetic conditions that are incompatible with life and detectable *in utero* [21, 145– 147]. Fetal stem cell and genetic therapies, and initial animal and clinical data thereof, were reviewed by Witt and colleagues [148]. Without intrauterine treatment, fetuses with ATM are severely anemic and will die *in utero* or during the neonatal period or, rarely, survive with major neurological impairments. ATM also presents with significant maternal morbidities including pre-eclampsia and hypertension. Intrauterine blood transfusions to reduce fetal anemia have led to improved outcomes in severely affected patients [149, 150]; transfused fetuses who survive to infancy generally have a good outlook but are reliant on lifelong blood transfusions, medications, and/or bone marrow stem cell transplantations from matched donors [151]. An emerging strategy to combat ATM (and other inherited genetic diseases incompatible with life) is *in utero* hematopoietic cell transplantation (IUHCTx). By introducing donor stem cells before immune maturity, donor specific tolerance could be induced to improve outcomes in affected fetuses [145]. However, only limited cases have been reported in human patients, and maternal rejection of donor cells is an issue [145, 152]. One promising area of investigation is the use of maternal cells as donor cells in fetal transplantation because fetal cells are already de-sensitized to the antigens of the mother [143, 153]. The first news report of a fetus with ATM surviving to birth after serial *in utero* blood transfusions and a bone marrow stem cell transplant from maternal cells was released in May 2018 from the UCSF Fetal Treatment Center [154]; the clinical trial, from which this is the first reported case, is ongoing (ClinicalTrials.gov ID NCT02986698) [152]. Additionally, MacKenzie and colleagues published a consensus statement about the future of fetal stem cell transplantation and gene therapy [155]. Beyond ATM, other candidate conditions include sickle cell anemia and osteogenesis imperfecta. Future work in IUHCTx could utilize novel materials for delivery of stem cells or other therapies to the fetus as cell engraftment remains low.

Early fetal surgeons observed that fetuses exhibit gestational age-dependent scarless healing following fetal surgery [156]. The mechanism underlying this scarless healing has been an active area of investigation [157] and piqued interest in the potential of fetal surgery to improve infant outcomes relative to post-natal (scar-inducing) intervention. This regenerative-type healing could also be used to the advantage of engineers designing biomaterials for the fetal milieu.

Conclusion

Fetal surgery is a growing and promising field of medicine that has the potential to drastically improve or save lives of children with debilitating or terminal diagnoses. In this review, we have presented the progress of several biomaterials solutions for fetal surgery and have suggested potential avenues for further exploration. As the field continues to transition from open surgeries to minimally invasive procedures, biomaterials are poised to become more widely used; for example, in the prenatal repair of myelomeningocele defects, it could be far easier to insert a biomaterial patch through a cannula than to do a full surgical repair of MMC defect in utero. Similarly, successful in utero gastroschisis repair may also rely on the development of an appropriate biomaterial patch. Perhaps biomaterials can have the greatest impact on fetal surgery and fetal therapy through the development of adhesives to prevent fetal membrane rupture following fetal surgery. The risk of iatrogenic membrane rupture, the "Achilles heel" of fetal surgery, is still the riskiest part of most fetal surgeries; a robust method to prevent membrane rupture (and thus subsequent preterm birth) would make fetal surgery accessible to more families by decreasing the overall risk of the procedure, tipping the balance on the risk-benefit analysis. Fetal blood transplantation and stem cell therapy remain an ongoing area of clinical and basic science research; in the future, biomaterials strategies may be useful to improve engraftment or delivery of these cells. As prenatal diagnostic technologies improve, clinicians will be better able to identify patients well-suited for fetal surgery; this trend has already started and demand for fetal surgery centers at major pediatric hospitals is growing. Today, over 30 hospitals have fetal therapy programs registered with NAFTANet (North American Fetal Therapy Network), and other fetal treatment centers exist internationally outside the NAFTANet system. As recently as the 1980s, fetal surgery was accessible to only 10s of patients a year; today it is the standard of care for thousands of patients per year in the United States. Moving forward, targeted biomaterial development will enable fetal surgery to help even more families deliver healthy, thriving children.

Acknowledgements

The authors thank Vamsi K. Aribindi, M.D., for helpful feedback on the manuscript and Colin Fahrion for illustrations. This work was supported by the National Science Foundation (Graduate Research Fellowship DGE 1752814 to SMW) and the National Institutes of Health (1R01EB022031–01).

Glossary

Clinical equipoise

The assumption that it is unknown which of two or more treatment options is better. In the context of clinical trials, it is ethical to establish clinical equipoise between treatment groups.

EXIT procedure

Ex Utero Intrapartum Treatment, a type of Caesarian section in which the baby is kept attached to the umbilical cord to receive oxygenated blood from the placenta until breathing or breathing support can be established. Used in cases where fetal airway obstruction is known or suspected.

Fetal surgery vs. fetal therapy

Fetal therapy usually refers to procedures that have limited number of instruments entering the uterus, for example fetal blood transfusions, while fetal surgery often is used to refer to more complicated invasive procedures like shunt placement or open surgery.

Iatrogenic

Used to describe symptoms or conditions that are caused by medical intervention or treatment. For example, iatrogenic membrane rupture is membrane rupture that results from *in utero* interventions.

Laparotomy

Surgical incision into the abdominal cavity, for example to expose the uterus for fetal surgery.

Amnioreduction

Insertion of a needle to aspirate amniotic fluid from the uterus to reduce amniotic fluid volumes in cases of polyhydramnios.

Oligohydramnios

Insufficient amniotic fluid present during gestation. This can hinder lung maturation and lead to perinatal morbidity.

Polyhydramnios

Excess amniotic fluid present during gestation. This can lead to poor perinatal outcomes.

References

- [1]. Harrison MR, Filly RA, Golbus MS, Berkowitz RL, Callen PW, Canty TG, Catz C, Clewell WH, Depp R, Edwards MS, Fletcher JC, Frigoletto FD, Garrett WJ, Johnson ML, Jonsen A, De Lorimier AA, Liley WA, Mahoney MJ, Manning FD, Meier PR, Michejda M, Nakayama DK, Nelson L, Newkirk JB, Pringle K, Rodeck C, Rosen MA, Schulman JD, Fetal treatment 1982, N Engl J Med 307(26) (1982) 1651–2. [PubMed: 7144864]
- [2]. Deprest JA, Devlieger R, Srisupundit K, Beck V, Sandaite I, Rusconi S, Claus F, Naulaers G, Van de Velde M, Brady P, Devriendt K, Vermeesch J, Toelen J, Carlon M, Debyser Z, De Catte L, Lewi L, Fetal surgery is a clinical reality, Semin Fetal Neonat M 15(1) (2010) 58–67.
- [3]. Adzick NS, Thom EA, Spong CY, Brock JW, Burrows PK, Johnson MP, Howell LJ, Farrell JA, Dabrowiak ME, Sutton LN, Gupta N, Tulipan NB, D'Alton ME, Farmer DL, Investigators M, A Randomized Trial of Prenatal versus Postnatal Repair of Myelomeningocele, New Engl J Med 364(11) (2011) 993–1004. [PubMed: 21306277]

- [4]. Golbus MS, Harrison MR, Filly RA, Callen PW, Katz M, In utero treatment of urinary tract obstruction, Am J Obstet Gynecol 142(4) (1982) 383–8. [PubMed: 7199255]
- [5]. Graves CE, Harrison MR, Padilla BE, Minimally Invasive Fetal Surgery, Clinics in Perinatology 44(4) (2017) 729. [PubMed: 29127956]
- [6]. Kabagambe SK, Lee CJ, Goodman LF, Chen YJ, Vanover MA, Farmer DL, Lessons from the Barn to the Operating Suite: A Comprehensive Review of Animal Models for Fetal Surgery, Annu Rev Anim Biosci 6 (2018) 99–119. [PubMed: 29237141]
- [7]. Antiel RM, Ethical challenges in the new world of maternal-fetal surgery, Semin Perinatol 40(4) (2016) 227–33. [PubMed: 26804036]
- [8]. Munson D, The intersection of fetal palliative care and fetal surgery: Addressing mortality and quality of life, Seminars in Perinatology 41(2) (2017) 101–105. [PubMed: 28108023]
- [9]. Kitagawa H, Pringle KC, Fetal surgery: a critical review, Pediatr Surg Int 33(4) (2017) 421–433. [PubMed: 28058487]
- [10]. Chervenak FA, McCullough LB, The ethics of maternal-fetal surgery, Semin Fetal Neonat M 23(1) (2018) 64–67.
- [11]. Antiel RM, Flake AW, Collura CA, Johnson MP, Rintoul NE, Lantos JD, Curlin FA, Tilburt JC, Brown SD, Feudtner C, Weighing the Social and Ethical Considerations of Maternal-Fetal Surgery, Pediatrics 140(6) (2017) e20170608.
- [12]. Antiel RM, Flake AW, Responsible surgical innovation and research in maternal-fetal surgery, Semin Fetal Neonat M 22(6) (2017) 423–427.
- [13]. Morris R, Kilby M, The PLUTO trial: percutaneous shunting in lower urinary tract obstruction, American Journal of Obstetrics and Gynecology 206(1) (2012) S14–S14.
- [14]. Pluto Collaborative Study G, Kilby M, Khan K, Morris K, Daniels J, Gray R, Magill L, Martin B, Thompson P, Alfirevic Z, Kenny S, Bower S, Sturgiss S, Anumba D, Mason G, Tydeman G, Soothill P, Brackley K, Loughna P, Cameron A, Kumar S, Bullen P, PLUTO trial protocol: percutaneous shunting for lower urinary tract obstruction randomised controlled trial, BJOG 114(7) (2007) 904–5, e1–4. [PubMed: 17567421]
- [15]. Senat MV, Deprest J, Boulvain M, Paupe A, Winer N, Ville Y, Endoscopic laser surgery versus serial amnioreduction for severe twin-to-twin transfusion syndrome, N Engl J Med 351(2) (2004) 136–44. [PubMed: 15238624]
- [16]. Deprest J, Jani J, Gratacos E, Vandecruys H, Naulaers G, Delgado J, Greenough A, Nicolaides K, Grp FT, Fetal intervention for congenital diaphragmatic hernia: The European experience, Seminars in Perinatology 29(2) (2005) 94–103. [PubMed: 16050527]
- [17]. Clayton DB, Brock JW, Current State of Fetal Intervention for Lower Urinary Tract Obstruction, Curr Urol Rep 19(1) (2018) 12. [PubMed: 29468448]
- [18]. Watanabe M, Kim AG, Flake AW, Tissue engineering strategies for fetal myelomeningocele repair in animal models, Fetal Diagn Ther 37(3) (2015) 197–205. [PubMed: 25060746]
- [19]. Bilic G, Brubaker C, Messersmith PB, Mallik AS, Quinn TM, Haller C, Done E, Gucciardo L, Zeisberger SM, Zimmermann R, Deprest J, Zisch AH, Injectable candidate sealants for fetal membrane repair: bonding and toxicity in vitro, American Journal of Obstetrics and Gynecology 202(1) (2010) 85.e1–85.e9. [PubMed: 20096254]
- [20]. Beck V, Lewi P, Gucciardo L, Devlieger R, Preterm prelabor rupture of membranes and fetal survival after minimally invasive fetal surgery: a systematic review of the literature, Fetal Diagn Ther 31(1) (2012) 1–9. [PubMed: 22104520]
- [21]. Merianos D, Heaton T, Flake AW, In Utero Hematopoietic Stem Cell Transplantation: Progress toward Clinical Application, Biology of Blood and Marrow Transplantation 14(7) (2008) 729– 740. [PubMed: 18541191]
- [22]. Papanna R, Molina S, Moise KY, Moise KJ, Johnson A, Chorioamnion plugging and the risk of preterm premature rupture of membranes after laser surgery in twin-twin transfusion syndrome, Ultrasound Obst Gyn 35(3) (2010) 337–343.
- [23]. Klaritsch P, Albert K, Van Mieghem T, Gucciardo L, Done E, Bynens B, Deprest J, Instrumental requirements for minimal invasive fetal surgery, Bjog-Int J Obstet Gy 116(2) (2009) 188–197.

- [24]. Golombeck K, Ball RH, Lee H, Farrell JA, Farmer DL, Jacobs VR, Rosen MA, Filly RA, Harrison MR, Maternal morbidity after maternal-fetal surgery, Am J Obstet Gynecol 194(3) (2006) 834–9. [PubMed: 16522421]
- [25]. Wilson RD, Lemerand K, Johnson MP, Flake AW, Bebbington M, Hedrick HL, Adzick NS, Reproductive outcomes in subsequent pregnancies after a pregnancy complicated by open maternal-fetal surgery (1996–2007), American Journal of Obstetrics and Gynecology 203(3) (2010).
- [26]. Young BK, Roman AS, MacKenzie AP, Stephenson CD, Minior V, Rebarber A, Timor-Tritsch I, The closure of iatrogenic membrane defects after amniocentesis and endoscopic intrauterine procedures, Fetal Diagn Ther 19(3) (2004) 296–300. [PubMed: 15067244]
- [27]. Devlieger R, Gratacos E, Wu J, Verbist L, Pijnenborg R, Deprest JA, An organ-culture for in vitro evaluation of fetal membrane healing capacity, Eur J Obstet Gynecol Reprod Biol 92(1) (2000) 145–50. [PubMed: 10986449]
- [28]. Devlieger R, Millar LK, Bryant-Greenwood G, Lewi L, Deprest JA, Fetal membrane healing after spontaneous and iatrogenic membrane rupture: A review of current evidence, American Journal of Obstetrics and Gynecology 195(6) (2006) 1512–1520. [PubMed: 16681986]
- [29]. Gratacos E, Sanin-Blair J, Lewi L, Toran N, Verbist G, Cabero L, A histological study of fetoscopic membrane defects to document membrane healing, Placenta 27 (2006) 452–6. [PubMed: 15953634]
- [30]. Moore RM, Mansour JM, Redline RW, Mercer BM, Moore JJ, The physiology of fetal membrane rupture: insight gained from the determination of physical properties, Placenta 27(11–12) (2006) 1037–51. [PubMed: 16516962]
- [31]. Carvalho NS, Moron AF, Menon R, Cavalheiro S, Barbosa MM, Milani HJ, Ishigai MM, Histological evidence of reparative activity in chorioamniotic membrane following open fetal surgery for myelomeningocele, Experimental and Therapeutic Medicine 14(4) (2017) 3732– 3736. [PubMed: 29042971]
- [32]. Mercer BM, Preterm premature rupture of the membranes, Obstet Gynecol 101(1) (2003) 178– 93. [PubMed: 12517665]
- [33]. Saigal S, Doyle LW, Preterm birth 3 An overview of mortality and sequelae of preterm birth from infancy to adulthood, Lancet 371(9608) (2008) 261–269. [PubMed: 18207020]
- [34]. Rüegg L, Hüsler M, Krähenmann F, Natalucci G, Zimmermann R, Ochsenbein-Kölble N, Outcome after fetoscopic laser coagulation in twin–twin transfusion syndrome – is the survival rate of at least one child at 6 months of age dependent on preoperative cervical length and preterm prelabour rupture of fetal membranes?, The Journal of Maternal-Fetal & Neonatal Medicine (2018) 1–9.
- [35]. Bryant-Greenwood G, Millar LK, Human fetal membranes: Their preterm premature rupture, Biology of Reproduction 63 (2000) 1575–79.
- [36]. Field DJ, Dorling JS, Manktelow BN, Draper ES, Survival of extremely premature babies in a geographically defined population: prospective cohort study of 1994–9 compared with 2000–5, BMJ 336(7655) (2008) 1221–3. [PubMed: 18469017]
- [37]. Chang J, Tracy TF, Carr SR, Sorrells DL, Luks FI, Port insertion and removal techniques to minimize premature rupture of the membranes in endoscopic fetal surgery, Journal of Pediatric Surgery 41(5) (2006) 905–909. [PubMed: 16677880]
- [38]. Quintero R, New horizons in the treatment of preterm premature rupture of membranes, Clin Perinatol 28 (2001) 861–75. [PubMed: 11817194]
- [39]. Quintero R, Treatment of previable premature ruptured membranes, Clin Perinatol 30 (2003) 573–89. [PubMed: 14533897]
- [40]. Breathnach F, Daly S, Griffin E, Gleeson N, Intracervical application of synthetic hydrogel sealant for preterm prelabor rupture of membranes: a case report, Journal of Perinatal Medicine 33(5) (2005) 458–460. [PubMed: 16250124]
- [41]. Mann LK, Papanna R, Moise KJ Jr., Byrd RH, Popek EJ, Kaur S, Tseng SCG, Stewart RJ, Fetal membrane patch and biomimetic adhesive coacervates as a sealant for fetoscopic defects, Acta Biomaterialia 8(6) (2012) 2160–2165. [PubMed: 22373817]

- [42]. Petratos PB, Baergen RN, Bleustein CB, Felsen D, Poppas DP, Ex vivo evaluation of human fetal membrane closure, Lasers in Surgery and Medicine 30(1) (2002) 48–53. [PubMed: 11857604]
- [43]. Mallik A, Fichter M, Rieder S, Bilic G, Stergioula S, Henke J, Fetoscopic closure of punctured fetal membranes with acellular human amnion plugs in a rabbit model, Obstet Gynecol 110 (2007) 1121–9. [PubMed: 17978128]
- [44]. Ochsenbein-Kolble N, Jani J, Lewi L, Verbist G, Vercruysse L, Portmann-Lanz B, Marquardt M, Zimmermann R, Deprest J, Enhancing sealing of fetal membrane defects using tissue engineered native amniotic-scaffolds in the rabbit model, American Journal of Obstetrics and Gynecology 196(3) (2007) 263–265. [PubMed: 17346548]
- [45]. Zisch A, Zimmermann R, Bioengineering of foetal membrane repair, Swiss Med Wkly 138 (2008) 596–601. [PubMed: 18941945]
- [46]. Kivelio A, Ochsenbein-Koelble N, Zimmermann R, Ehrbar M, Engineered cell instructive matrices for fetal membrane healing, Acta Biomater 15 (2015) 1–10. [PubMed: 25536031]
- [47]. Carnaghan HK, Harrison MR, Presealing of the chorioamniotic membranes prior to fetoscopic surgery: preliminary study with unfertilized chicken egg models, Eur J Obstet Gynecol Reprod Biol 144 Suppl 1 (2009) S142–5. [PubMed: 19304365]
- [48]. Cortes RA, Wagner AJ, Lee H, Clifton MS, Grethel E, Yang SH, Ball R, Harrison MR, Preemptive placement of a presealant for amniotic access, American Journal of Obstetrics and Gynecology 193(3) (2005) 1197–1203. [PubMed: 16157137]
- [49]. Engels AC, Van Calster B, Richter J, DeKoninck P, Lewi L, De Catte L, Devlieger R, Deprest JA, Collagen plug sealing of iatrogenic fetal membrane defects after fetoscopic surgery for congenital diaphragmatic hernia, Ultrasound Obstet Gynecol 43(1) (2014) 54–9. [PubMed: 23801588]
- [50]. Quintero RA, Morales WJ, Allen M, Bornick PW, Arroyo J, LeParc G, Treatment of iatrogenic previable premature rupture of membranes with intra-amniotic injection of platelets and cryoprecipitate (amniopatch): Preliminary experience, American Journal of Obstetrics and Gynecology 181(3) (1999) 744–749. [PubMed: 10486493]
- [51]. O'Brien JM, Milligan DA, Barton JR, Gelatin sponge embolization. a method for the management of iatrogenic preterm premature rupture of the membranes, Fetal Diagn Ther 17(1) (2002) 8–10. [PubMed: 11803208]
- [52]. Richter J, Henry A, Ryan G, DeKoninck P, Lewi L, Deprest J, Amniopatch procedure after previable iatrogenic rupture of the membranes: a two-center review, Prenat. Diagn. 33(4) (2013) 391–396. [PubMed: 23512492]
- [53]. Burke SA, Ritter-Jones M, Lee BP, Messersmith PB, Thermal gelation and tissue adhesion of biomimetic hydrogels, Biomed Mater 2(4) (2007) 203–10. [PubMed: 18458476]
- [54]. Haller CM, Buerzle W, Kivelio A, Perrini M, Brubaker CE, Gubeli RJ, Mallik AS, Weber W, Messersmith PB, Mazza E, Ochsenbein-Koelble N, Zimmermann R, Ehrbar M, Mussel-mimetic tissue adhesive for fetal membrane repair: an ex vivo evaluation, Acta Biomater 8(12) (2012) 4365–70. [PubMed: 22885681]
- [55]. Devaud YR, Zuger S, Zimmermann R, Ehrbar M, Ochsenbein-Kolble N, Minimally Invasive Surgical Device for Precise Application of Bioadhesives to Prevent iPPROM, Fetal Diagnosis and Therapy 45(2) (2019) 102–110. [PubMed: 29920508]
- [56]. Azadani AN, Matthews PB, Ge L, Shen Y, Jhun CS, Guy TS, Tseng EE, Mechanical Properties of Surgical Glues Used in Aortic Root Replacement, Ann Thorac Surg 87(4) (2009) 1154–1160. [PubMed: 19324142]
- [57]. Bures M, Hoffler HK, Friedel G, Kyriss T, Boedeker E, Langer F, Zardo P, Zhang RY, Albuminglutaraldehyde glue for repair of superficial lung defect: an in vitro experiment, J Cardiothorac Surg 11(1) (2016) 63. [PubMed: 27072534]
- [58]. Spotnitz WD, Burks S, Hemostats, sealants, and adhesives: components of the surgical toolbox, Transfusion 48(7) (2008) 1502–1516. [PubMed: 18422855]
- [59]. Kivelio A, Dekoninck P, Perrini M, Brubaker CE, Messersmith PB, Mazza E, Deprest J, Zimmermann R, Ehrbar M, Ochsenbein-Koelble N, Mussel mimetic tissue adhesive for fetal membrane repair: initial in vivo investigation in rabbits, Eur J Obstet Gynecol Reprod Biol 171(2) (2013) 240–5. [PubMed: 24075447]

- [60]. Papanna R, Mann LK, Tseng SC, Stewart RJ, Kaur SS, Swindle MM, Kyriakides TR, Tatevian N, Moise KJ Jr., Cryopreserved human amniotic membrane and a bioinspired underwater adhesive to seal and promote healing of iatrogenic fetal membrane defect sites, Placenta 36(8) (2015) 888– 94. [PubMed: 26059341]
- [61]. Roman S, Bullock AJ, Anumba DO, MacNeil S, Development of an implantable synthetic membrane for the treatment of preterm premature rupture of fetal membranes, J Biomater Appl 30(7) (2016) 995–1003. [PubMed: 26491057]
- [62]. Pensabene V, Patel PP, Williams P, Cooper TL, Kirkbride KC, Giorgio TD, Tulipan NB, Repairing Fetal Membranes with a Self-adhesive Ultrathin Polymeric Film: Evaluation in Midgestational Rabbit Model, Ann Biomed Eng 43(8) (2015) 1978–1988. [PubMed: 25549772]
- [63]. Mendez-Figueroa H, Papanna R, Berry DL, Moise KJ, Precipitated egg white as a sealant for iatrogenic preterm premature rupture of the membranes, American Journal of Obstetrics and Gynecology 202(2) (2010) 191.e1–6. [PubMed: 19942207]
- [64]. Shao Y, Taniguchi K, Gurdziel K, Townshend RF, Xue X, Yong KM, Sang J, Spence JR, Gumucio DL, Fu J, Self-organized amniogenesis by human pluripotent stem cells in a biomimetic implantation-like niche, Nat Mater 16(4) (2017) 419–425. [PubMed: 27941807]
- [65]. Parker SE, Mai CT, Canfield MA, Rickard R, Wang Y, Meyer RE, Anderson P, Mason CA, Collins JS, Kirby RS, Correa A, National N Birth Defects Prevention, Updated National Birth Prevalence estimates for selected birth defects in the United States, 2004–2006, Birth Defects Res A Clin Mol Teratol 88(12) (2010) 1008–16. [PubMed: 20878909]
- [66]. Velie EM, Shaw GM, Impact of prenatal diagnosis and elective termination on prevalence and risk estimates of neural tube defects in California, 1989–1991, Am J Epidemiol 144(5) (1996) 473–479. [PubMed: 8781462]
- [67]. Lary JM, Edmonds LD, Prevalence of Spina Bifida at Birth -- United States, 1983–1990: a Comparison of Two Surveillance Systems, MMWR CDC Surveill Summ 42(2) (1996) 15–26.
- [68]. Adzick NS, Fetal myelomeningocele: Natural history, pathophysiology, and in-utero intervention, Semin Fetal Neonat M 15(1) (2010) 9–14.
- [69]. Heffez DS, Aryanpur J, Hutchins GM, Freeman JM, The paralysis associated with myelomeningocele: clinical and experimental data implicating a preventable spinal cord injury, Neurosurgery 26(6) (1990) 987–92. [PubMed: 2362676]
- [70]. Bruner JP, Tulipan N, Paschall RL, Boehm FH, Walsh WF, Silva SR, Hernanz-Schulman M, Lowe LH, Reed GW, Fetal surgery for myelomeningocele and the incidence of shunt-dependent hydrocephalus, JAMA 282(19) (1999) 1819–25. [PubMed: 10573272]
- [71]. Moldenhauer JS, Adzick NS, Fetal surgery for myelomeningocele: After the Management of Myelomeningocele Study (MOMS), Semin Fetal Neonat M 22(6) (2017) 360–366.
- [72]. Bruner JP, Richards WO, Tulipan NB, Arney TL, Endoscopic coverage of fetal myelomeningocele in utero, Am J Obstet Gynecol 180(1 Pt 1) (1999) 153–8. [PubMed: 9914596]
- [73]. Bruner JP, Tulipan NE, Richards WO, Endoscopic coverage of fetal open myelomeningocele in utero, Am J Obstet Gynecol 176(1 Pt 1) (1997) 256–7.
- [74]. Watanabe M, Jo J, Radu A, Kaneko M, Tabata Y, Flake AW, A tissue engineering approach for prenatal closure of myelomeningocele with gelatin sponges incorporating basic fibroblast growth factor, Tissue Eng Part A 16(5) (2010) 1645–55. [PubMed: 19954327]
- [75]. Watanabe M, Li H, Roybal J, Santore M, Radu A, Jo J, Kaneko M, Tabata Y, Flake A, A tissue engineering approach for prenatal closure of myelomeningocele: comparison of gelatin sponge and microsphere scaffolds and bioactive protein coatings, Tissue Eng Part A 17(7–8) (2011) 1099–110. [PubMed: 21128864]
- [76]. Eggink AJ, Roelofs LA, Feitz WF, Wijnen RM, Mullaart RA, Grotenhuis JA, van Kuppevelt TH, Lammens MM, Crevels AJ, Hanssen A, van den Berg PP, In utero repair of an experimental neural tube defect in a chronic sheep model using biomatrices, Fetal Diagn Ther 20(5) (2005) 335–40. [PubMed: 16113549]
- [77]. Eggink AJ, Roelofs LA, Lammens MM, Feitz WF, Wijnen RM, Mullaart RA, van Moerkerk HT, van Kuppevelt TH, Crevels AJ, Hanssen A, Lotgering FK, van den Berg PP, Histological evaluation of acute covering of an experimental neural tube defect with biomatrices in fetal sheep, Fetal Diagn Ther 21(2) (2006) 210–6. [PubMed: 16491005]

- [78]. Eggink AJ, Roelofs LA, Feitz WF, Wijnen RM, Lammens MM, Mullaart RA, van Moerkerk HT, van Kuppevelt TH, Crevels AJ, Verrijp K, Lotgering FK, van den Berg PP, Delayed intrauterine repair of an experimental spina bifida with a collagen biomatrix, Pediatr Neurosurg 44(1) (2008) 29–35. [PubMed: 18097188]
- [79]. Fontecha CG, Peiro JL, Aguirre M, Soldado F, Anor S, Fresno L, Martinez-Ibanez V, Inert patch with bioadhesive for gentle fetal surgery of myelomeningocele in a sheep model, Eur J Obstet Gynecol Reprod Biol 146(2) (2009) 174–9. [PubMed: 19615808]
- [80]. Fontecha CG, Peiro JL, Sevilla JJ, Aguirre M, Soldado F, Fresno L, Fonseca C, Chacaltana A, Martinez V, Fetoscopic coverage of experimental myelomeningocele in sheep using a patch with surgical sealant, Eur J Obstet Gynecol Reprod Biol 156(2) (2011) 171–6. [PubMed: 21353374]
- [81]. Peiro JL, Fontecha CG, Ruano R, Esteves M, Fonseca C, Marotta M, Haeri S, Belfort MA, Single-Access Fetal Endoscopy (SAFE) for myelomeningocele in sheep model I: amniotic carbon dioxide gas approach, Surg Endosc 27(10) (2013) 3835–40. [PubMed: 23670742]
- [82]. Joyeux L, De Bie F, Danzer E, Van Mieghem T, Flake AW, Deprest J, Safety and efficacy of fetal surgery techniques to close a spina bifida defect in the fetal lamb model: A systematic review, Prenat. Diagn. 38(4) (2018) 231–242. [PubMed: 29388237]
- [83]. Watanabe M, Li HY, Kim AG, Weilerstein A, Radu A, Davey M, Loukogeorgakis S, Sanchez MD, Sumita K, Morimoto N, Yamamoto M, Tabata Y, Flake AW, Complete tissue coverage achieved by scaffold-based tissue engineering in the fetal sheep model of Myelomeningocele, Biomaterials 76 (2016) 133–143. [PubMed: 26520044]
- [84]. Stephenson JT, Pichakron KO, Vu L, Jancelewicz T, Jamshidi R, Grayson JK, Nobuhara KK, In utero repair of gastroschisis in the sheep (Ovis aries) model, Journal of Pediatric Surgery 45(1) (2010) 65–69. [PubMed: 20105581]
- [85]. Sanchez e Oliveira Rde C, Valente PR, Abou-Jamra RC, Araujo A, Saldiva PH, Pedreira DA, Biosynthetic cellulose induces the formation of a neoduramater following pre-natal correction of meningomyelocele in fetal sheep, Acta Cir Bras 22(3) (2007) 174–81. [PubMed: 17546289]
- [86]. Papanna R, Moise KJ Jr., Mann LK, Fletcher S, Schniederjan R, Bhattacharjee MB, Stewart RJ, Kaur S, Prabhu SP, Tseng SC, Cryopreserved human umbilical cord patch for in-utero spina bifida repair, Ultrasound Obstet Gynecol 47(2) (2016) 168–76. [PubMed: 26489897]
- [87]. Papanna R, Fletcher S, Moise KJ Jr., Mann LK, Tseng SC, Cryopreserved Human Umbilical Cord for In Utero Myeloschisis Repair, Obstet Gynecol 128(2) (2016) 325–30. [PubMed: 27400004]
- [88]. Chen YJ, Chung KR, Pivetti C, Lankford L, Kabagambe SK, Vanover M, Becker J, Lee C, Tsang J, Wang AJ, Farmer DL, Fetal surgical repair with placenta-derived mesenchymal stromal cell engineered patch in a rodent model of myelomeningocele, Journal of Pediatric Surgery 53(1) (2018) 183–188.
- [89]. Holland AJA, Walker K, Badawi N, Gastroschisis: an update, Pediatr Surg Int 26(9) (2010) 871– 878. [PubMed: 20686898]
- [90]. Langer JC, Longaker MT, Crombleholme TM, Bond SJ, Finkbeiner WE, Rudolph CA, Verrier ED, Harrison MR, Etiology of Intestinal Damage in Gastroschisis 1. Effects of Amniotic-Fluid Exposure and Bowel Constriction in a Fetal Lamb Model, Journal of Pediatric Surgery 24(10) (1989) 992–997. [PubMed: 2530329]
- [91]. Langer JC, Bell JG, Castillo RO, Crombleholme TM, Longaker MT, Duncan BW, Bradley SM, Finkbeiner WE, Verrier ED, Harrison MR, Etiology of intestinal damage in gastroschisis, II. Timing and reversibility of histological changes, mucosal function, and contractility, Journal of Pediatric Surgery 25(11) (1990) 1122–1126. [PubMed: 2148773]
- [92]. Srinathan SK, Langer JC, Blennerhassett MG, Harrison MR, Pelletier GJ, Lagunoff D, Etiology of intestinal damage in gastroschisis. III: Morphometric analysis of the smooth muscle and submucosa, J Pediatr Surg 30(3) (1995) 379–83.
- [93]. Klück P, Tibboel D, van der Kamp AW, Molenaar JC, The effect of fetal urine on the development of the bowel in gastroschisis, Journal of pediatric surgery 18(1) (1983) 47–50.[PubMed: 6834226]

- [94]. Correia-Pinto J, Tavares ML, Baptista MJ, Estevão-Costa J, Flake AW, Leite-Moreira AF, A new fetal rat model of gastroschisis: Development and early characterization, Journal of Pediatric Surgery 36(1) (2001) 213–216. [PubMed: 11150468]
- [95]. Phillips JD, Kelly RE Jr., Fonkalsrud EW, Mirzayan A, Kim CS, An improved model of experimental gastroschisis in fetal rabbits, J Pediatr Surg 26(7) (1991) 784–7. [PubMed: 1832713]
- [96]. Roelofs LAJ, Geutjes PJ, Hulsbergen-van de Kaa CA, Eggink AJ, van Kuppevelt TH, Daamen WF, Crevels AJ, van den Berg PP, Feitz WFJ, Wijnen RMH, Prenatal coverage of experimental gastroschisis with a collagen scaffold to protect the bowel, Journal of Pediatric Surgery 48(3) (2013) 516–524. [PubMed: 23480905]
- [97]. Bergholz R, Krebs T, Wenke K, Andreas T, Tiemann B, Paetzel J, Jacobsen B, Fahje R, Schmitz C, Mann O, Roth B, Appl B, Hecher K, Fetoscopic management of gastroschisis in a lamb model, Surgical Endoscopy and Other Interventional Techniques 26(5) (2012) 1412–1416. [PubMed: 22179441]
- [98]. Tatu R, Lin CY, Characterization of Biomaterial Patches as Fetal Surgery Implants, Front Nanobiomed Res 10 (2018) 29–47.
- [99]. Harrison MR, Mychaliska GB, Albanese CT, Jennings RW, Farrell JA, Hawgood S, Sandberg P, Levine AH, Lobo E, Filly RA, Correction of congenital diaphragmatic hernia in utero IX: Fetuses with poor prognosis (liver herniation and low lung-to-head ratio) can be saved by fetoscopic temporary tracheal occlusion, Journal of Pediatric Surgery 33(7) (1998) 1017–1022. [PubMed: 9694087]
- [100]. Saxena AK, Surgical perspectives regarding application of biomaterials for the management of large congenital diaphragmatic hernia defects, Pediatr Surg Int 34(5) (2018) 475–489. [PubMed: 29610961]
- [101]. Jeanty C, Kunisaki SM, MacKenzie TC, Novel non-surgical prenatal approaches to treating congenital diaphragmatic hernia, Semin Fetal Neonat M 19(6) (2014) 349–356.
- [102]. Eastwood MP, Russo FM, Toelen J, Deprest J, Medical interventions to reverse pulmonary hypoplasia in the animal model of congenital diaphragmatic hernia: A systematic review, Pediatr Pulm 50(8) (2015) 820–838.
- [103]. Harrison MR, Langer JC, Adzick NS, Golbus MS, Filly RA, Anderson RL, Rosen MA, Callen PW, Goldstein RB, Delorimier AA, Correction of Congenital Diaphragmatic-Hernia in Utero 5. Initial Clinical-Experience, Journal of Pediatric Surgery 25(1) (1990) 47–57. [PubMed: 2405147]
- [104]. Deprest J, Gratacos E, Nicolaides KH, Grp FT, Fetoscopic tracheal occlusion (FETO) for severe congenital diaphragmatic hernia: evolution of a technique and preliminary results, Ultrasound Obst Gyn 24(2) (2004) 121–126.
- [105]. Elattal R, Rich BS, Harmon CM, Muensterer OJ, Pulmonary alveolar and vascular morphometry after gel plug occlusion of the trachea in a fetal rabbit model of CDH, Int J Surg 11(7) (2013) 558–561. [PubMed: 23721663]
- [106]. Fauza DO, Marler JJ, Koka R, Forse RA, Mayer JE, Vacanti JP, Fetal tissue engineering: diaphragmatic replacement, J Pediatr Surg 36(1) (2001) 146–51. [PubMed: 11150454]
- [107]. Harrison MR, Adzick NS, Flake AW, Jennings RW, Estes JM, Macgilivray TE, Chueh JT, Goldberg JD, Filly RA, Goldstein RB, Rosen MA, Cauldwell C, Levine AH, Howell LJ, Correction of Congenital Diaphragmatic-Hernia in-Utero 6. Hard-Earned Lessons, Journal of Pediatric Surgery 28(10) (1993) 1411–1418. [PubMed: 8263712]
- [108]. Harrison MR, Sydorak RM, Farrell JA, Kitterman JA, Filly RA, Albanese CT, Fetoscopic temporary tracheal occlusion for congenital diaphragmatic hernia: Prelude to a randomized, controlled trial, Journal of Pediatric Surgery 38(7) (2003) 1012–1020. [PubMed: 12861529]
- [109]. VanderWall KJ, Bruch SW, Meuli M, Kohl T, Szabo Z, Adzick NS, Harrison MR, Fetal endoscopic ('Fetendo') tracheal clip, Journal of Pediatric Surgery 31(8) (1996) 1101–1103. [PubMed: 8863243]
- [110]. Muensterer OJ, Nicola T, Farmer S, Harmon CM, Ambalavanan N, Temporary fetal tracheal occlusion using a gel plug in a rabbit model of congenital diaphragmatic hernia, Journal of Pediatric Surgery 47(6) (2012) 1063–1066. [PubMed: 22703770]

- [111]. Chang R, Komura M, Andreoli S, Jennings R, Wilson J, Fauza D, Rapidly polymerizing hydrogel prevents balloon dislodgement in a model of fetal tracheal occlusion, J Pediatr Surg 39(4) (2004) 557–60. [PubMed: 15065027]
- [112]. Lewi L, Gucciardo L, Van Mieghem T, de Koninck P, Beck V, Medek H, Van Schoubroeck D, Devlieger R, De Catte L, Deprest J, Monochorionic diamniotic twin pregnancies: natural history and risk stratification, Fetal Diagn Ther 27(3) (2010) 121–33. [PubMed: 20413975]
- [113]. Chalouhi GE, Essaoui M, Stirnemann J, Quibel T, Deloison B, Salomon L, Ville Y, Laser therapy for twin-to-twin transfusion syndrome (TTTS), Prenat. Diagn. 31(7) (2011) 637–646. [PubMed: 21660997]
- [114]. van Gemert MJC, van den Wijngaard JPHM, Vandenbussche FPHA, Twin reversed arterial perfusion sequence is more common than generally accepted, Birth Defects Res A 103(7) (2015) 641–643.
- [115]. Lee H, Wagner AJ, Sy E, Ball R, Feldstein VA, Goldstein RB, Farmer DL, Efficacy of radiofrequency ablation for twin-reversed arterial perfusion sequence, Am J Obstet Gynecol 196(5) (2007) 459 e1–4. [PubMed: 17466701]
- [116]. Russell Z, Quintero RA, Kontopoulos EV, Intrauterine growth restriction in monochorionic twins, Semin Fetal Neonat M 12(6) (2007) 439–449.
- [117]. Robyr R, Yamamoto M, Ville Y, Selective feticide in complicated monochorionic twin pregnancies using ultrasound-guided bipolar cord coagulation, Bjog-Int J Obstet Gy 112(10) (2005) 1344–1348.
- [118]. Lanna MM, Rustico MA, Dell'Avanzo M, Schena V, Faiola S, Consonni D, Righini A, Scelsa B, Ferrazzi EM, Bipolar cord coagulation for selective feticide in complicated monochorionic twin pregnancies: 118 consecutive cases at a single center, Ultrasound Obstet Gynecol 39(4) (2012) 407–13. [PubMed: 22173905]
- [119]. Morris RK, Malin GL, Quinlan-Jones E, Middleton LJ, Diwakar L, Hemming K, Burke D, Daniels J, Denny E, Barton P, Roberts TE, Khan KS, Deeks JJ, Kilby MD, Grp PC, Fetal bladder obstruction and its treatment, Health Technol Asses 17(59) (2013) 1.
- [120]. Manning FA, Harrison MR, Rodeck C, Catheter Shunts for Fetal Hydronephrosis and Hydrocephalus - Report of the International Fetal Surgery Registry, New Engl J Med 315(5) (1986) 336–340. [PubMed: 3724830]
- [121]. Elder JS, Duckett JW, Snyder HM, Intervention for Fetal Obstructive Uropathy Has It Been Effective, Lancet 2(8566) (1987) 1007–1010. [PubMed: 2889913]
- [122]. Clark TJ, Martin WL, Divakaran TG, Whittle MJ, Kilby MD, Khan KS, Prenatal bladder drainage in the management of fetal lower urinary tract obstruction: A systematic review and meta-analysis, Obstetrics and Gynecology 102(2) (2003) 367–382. [PubMed: 12907115]
- [123]. Morris RK, Malin GL, Quinlan-Jones E, Middleton LJ, Hemming K, Burke D, Daniels JP, Khan KS, Deeks J, Kilby MD, Shunti PV, Percutaneous vesicoamniotic shunting versus conservative management for fetal lower urinary tract obstruction (PLUTO): a randomised trial, Lancet 382(9903) (2013) 1496–1506. [PubMed: 23953766]
- [124]. Morris RK, Malin GL, Quinlan-Jones E, Middleton LJ, Diwakar L, Hemming K, Burke D, Daniels J, Denny E, Barton P, Roberts TE, Khan KS, Deeks JJ, Kilby MD, The Percutaneous shunting in Lower Urinary Tract Obstruction (PLUTO) study and randomised controlled trial: evaluation of the effectiveness, cost-effectiveness and acceptability of percutaneous vesicoamniotic shunting for lower urinary tract obstruction, Health Technol Assess 17(59) (2013) 1–232.
- [125]. Kurtz MP, Koh CJ, Jamail GA, Sangi-Haghpeykar H, Shamshirsaz AA, Espinoza J, Cass DL, Olutoye OO, Olutoye OA, Braun MC, Roth DR, Belfort MA, Ruano R, Factors associated with fetal shunt dislodgement in lower urinary tract obstruction, Prenat. Diagn. 36(8) (2016) 720–725. [PubMed: 27247093]
- [126]. Bruner JP, Davis G, Tulipan N, Intrauterine shunt for obstructive hydrocephalus--still not ready, Fetal Diagn Ther 21(6) (2006) 532–9. [PubMed: 16969010]
- [127]. Johnson ML, Pretorius D, Clewell WH, Meier PR, Manchester D, Fetal hydrocephalus: diagnosis and management, Semin Perinatol 7(2) (1983) 83–9. [PubMed: 6635672]

- [128]. Jeong BD, Won HS, Lee MY, Perinatal Outcomes of Fetal Lower Urinary Tract Obstruction After Vesicoamniotic Shunting Using a Double-Basket Catheter, J Ultrasound Med 37(9) (2018) 2147–2156. [PubMed: 29498072]
- [129]. Stanasel I, Gonzales E, Posterior Urethral Valves, Curr Bladder Dysfunct Rep 10(3) (2015) 250–255.
- [130]. Quintero RA, Shukla AR, Homsy YL, Bukkapatnam R, Successful in utero endoscopic ablation of posterior urethral valves: a new dimension in fetal urology, Urology 55(5) (2000) 774.
- [131]. Sananes N, Favre R, Koh CJ, Zaloszyc A, Braun MC, Roth DR, Moog R, Becmeur F, Belfort MA, Ruano R, Urological fistulas after fetal cystoscopic laser ablation of posterior urethral valves: surgical technical aspects, Ultrasound Obstet Gynecol 45(2) (2015) 183–9. [PubMed: 24817027]
- [132]. Ruano R, Yoshizaki CT, Giron AM, Srougi M, Zugaib M, Cystoscopic placement of transurethral stent in a fetus with urethral stenosis, Ultrasound Obstet Gynecol 44(2) (2014) 238– 40. [PubMed: 24375864]
- [133]. Ruano R, Sananes N, Sangi-Haghpeykar H, Hernandez-Ruano S, Moog R, Becmeur F, Zaloszyc A, Giron AM, Morin B, Favre R, Fetal intervention for severe lower urinary tract obstruction: a multicenter case-control study comparing fetal cystoscopy with vesicoamniotic shunting, Ultrasound Obstet Gynecol 45(4) (2015) 452–8. [PubMed: 25157756]
- [134]. Wilson RD, Baxter JK, Johnson MP, King M, Kasperski S, Crombleholme TM, Flake AW, Hedrick HL, Howell LJ, Adzick NS, Thoracoamniotic shunts: fetal treatment of pleural effusions and congenital cystic adenomatoid malformations, Fetal Diagn Ther 19(5) (2004) 413–20. [PubMed: 15305098]
- [135]. Rodeck CH, Fisk NM, Fraser DI, Nicolini U, Long-term in utero drainage of fetal hydrothorax, N Engl J Med 319(17) (1988) 1135–8. [PubMed: 3173443]
- [136]. Witlox R, Klumper F, Te Pas AB, van Zwet EW, Oepkes D, Lopriore E, Neonatal management and outcome after thoracoamniotic shunt placement for fetal hydrothorax, Arch Dis Child Fetal Neonatal Ed 103(3) (2018) F245–F249. [PubMed: 28780497]
- [137]. Clewell WH, Johnson ML, Meier PR, Newkirk JB, Zide SL, Hendee RW, Bowes WA Jr., Hecht F, O'Keeffe D, Henry GP, Shikes RH, A surgical approach to the treatment of fetal hydrocephalus, N Engl J Med 306(22) (1982) 1320–5. [PubMed: 7070456]
- [138]. Pretorius DH, Davis K, Manco-Johnson ML, Manchester D, Meier PR, Clewell WH, Clinical course of fetal hydrocephalus: 40 cases, AJR Am J Roentgenol 144(4) (1985) 827–31. [PubMed: 3883714]
- [139]. Szaflik K, Czaj M, Polis L, Wojtera J, Szmanski W, Krzeszowski W, Polis B, Litwinska M, Mikolajczyk W, Janiak K, Maroszynska I, Gulczynska E, Fetal therapy--evaluation of ventriculoamniotic shunts in the treatment of hydrocephalus, Ginekol Pol 85(12) (2014) 16–22. [PubMed: 25669060]
- [140]. von Koch CS, Gupta N, Sutton LN, Sun PP, In utero surgery for hydrocephalus, Childs Nerv Syst 19(7–8) (2003) 574–86. [PubMed: 12955423]
- [141]. Simon AK, Hollander GA, McMichael A, Evolution of the immune system in humans from infancy to old age, Proc Biol Sci 282(1821) (2015) 20143085.
- [142]. Anderson JM, Rodriguez A, Chang DT, Foreign body reaction to biomaterials, Semin Immunol 20(2) (2008) 86–100. [PubMed: 18162407]
- [143]. MacKenzie TC, Fetal Surgical conditions and the unraveling of maternal-fetal tolerance, J Pediatr Surg 51(2) (2016) 197–9. [PubMed: 26653947]
- [144]. Wegorzewska M, Nijagal A, Wong CM, Le T, Lescano N, Tang Q, MacKenzie TC, Fetal intervention increases maternal T cell awareness of the foreign conceptus and can lead to immune-mediated fetal demise, J Immunol 192(4) (2014) 1938–45. [PubMed: 24415782]
- [145]. Derderian SC, Jeanty C, Walters MC, Vichinsky E, MacKenzie TC, In utero hematopoietic cell transplantation for hemoglobinopathies, Front Pharmacol 5 (2014) 278. [PubMed: 25628564]
- [146]. Flake AW, In utero stem cell transplantation, Best Practice & Research Clinical Obstetrics & Gynaecology 18(6) (2004) 941–958. [PubMed: 15582548]
- [147]. Roybal JL, Santore MT, Flake AW, Stem cell and genetic therapies for the fetus, Seminars in Fetal and Neonatal Medicine 15(1) (2010) 46–51. [PubMed: 19540822]

- [148]. Witt R, MacKenzie TC, Peranteau WH, Fetal stem cell and gene therapy, Semin Fetal Neonat M 22(6) (2017) 410–414.
- [149]. Weisz B, Rosenbaum O, Chayen B, Peltz R, Feldman B, Lipitz S, Outcome of severely anaemic fetuses treated by intrauterine transfusions, Arch Dis Child Fetal Neonatal Ed 94(3) (2009) F201–4. [PubMed: 19000998]
- [150]. Vichinsky EP, Alpha thalassemia major--new mutations, intrauterine management, and outcomes, Hematology Am Soc Hematol Educ Program (2009) 35–41. [PubMed: 20008180]
- [151]. Elsaid MY, Capitini CM, Diamond CA, Porte M, Otto M, DeSantes KB, Successful matched unrelated donor stem cell transplant in Hemoglobin Bart's disease, Bone Marrow Transplant 51(11) (2016) 1522–1523. [PubMed: 27295273]
- [152]. Kreger EM, Singer ST, Witt RG, Sweeters N, Lianoglou B, Lal A, Mackenzie TC, Vichinsky E, Favorable outcomes after in utero transfusion in fetuses with alpha thalassemia major: a case series and review of the literature, Prenat. Diagn. 36(13) (2016) 1242–1249. [PubMed: 27862048]
- [153]. Witt R, Nguyen Q-HL, MacKenzie T, In Utero Hematopoietic Cell Transplantation: Past Clinical Experience and Future Clinical Trials, Current Stem Cell Reports 4(1) (2018) 74–80.
- [154]. Grady D, Five Blood Transfusions, One Bone Marrow Transplant All Before Birth, The New York Times (2018) https://www.nytimes.com/2018/05/25/health/fetal-bone-marrowtransplant.html.
- [155]. MacKenzie TC, David AL, Flake AW, Almeida-Porada G, Consensus statement from the first international conference for in utero stem cell transplantation and gene therapy, Front Pharmacol 6 (2015) 15. [PubMed: 25713535]
- [156]. Larson BJ, Longaker MT, Lorenz HP, Scarless Fetal Wound Healing: A Basic Science Review, Plastic and reconstructive surgery 126(4) (2010) 1172–1180. [PubMed: 20885241]
- [157]. Chen SY, Han B, Zhu YT, Mahabole M, Huang J, Beebe DC, Tseng SCG, HC-HA/PTX3 Purified from Amniotic Membrane Promotes BMP Signaling in Limbal Niche Cells to Maintain Quiescence of Limbal Epithelial Progenitor/Stem Cells, Stem Cells 33(11) (2015) 3341–3355. [PubMed: 26148958]

Box 1.

Criteria for fetal surgery, adapted from Harrison, et al. [1], Deprest et al. [2], and Adzick, et al [3].

- The disease must be diagnosable *in utero* and have no effective post-natal therapy.

- *In utero* disease staging criteria must be established to ensure that only severely affected fetuses undergo fetal surgery but also that surgery is not performed on fetuses too severely affected to benefit from intervention.

- Mothers and fetuses should be free of co-morbidities including fetal karyotype abnormalities.

- There must be proof in form of a clinical trial, or animal study evidence supporting a reasonable cause to assume (in case of pre-trial case studies), that the benefits of the therapy outweigh the risks of the procedure.

- Timing of the procedure must be optimized to provide maximum benefit to the fetus while decreasing the risk of preterm labor before the gestational limit of viability.

- Surgery and delivery should be performed at a specialty center with established ethical protocols, informed consent of parents, and an experienced multi-specialty team.

Winkler et al.



Figure 1.

Examples of biomaterials in fetal surgery. (**A**) In cases of congenital diaphragmatic hernia, *in utero* occlusion of the trachea with a silicone balloon allows lungs to fill with fluid and expand, pushing abdominal organs out of the pleural cavity. (**B**) *In utero* shunting can be used to drain fluid from swollen organs into the amniotic sac. For example, in cases of lower urinary obstruction, double-pigtail shunts can be inserted to drain the bladder. (**C**) Biomaterial patches can prevent amniotic fluid enzymes from degrading abnormally exposed tissues *in utero*, for example, in covering exposed neural tissue in myelomeningocele. (**D**) Materials to seal the fetal membranes (chorion and amnion) and reduce the risk of membrane rupture following fetal surgery are currently in development. These injectable hydrogels can seal between the uterus and membranes (called presealing) or inside the amniotic sac (as shown).



Figure 2.

Biomaterials methods to address congenital diaphragmatic hernia *in utero*. (A) In early fetal surgeries, Gore-Tex patches were used to repair the diaphragm defect and to patch the abdomen. (B) In later attempts, fetal tracheal occlusion was achieved by clamping the fetal trachea with a metal clip to occlude it and allow lung volume to expand. (C) Currently, tracheal occlusion is achieved by inserting a silicone balloon into the fetal trachea and inflating it with saline to occlude the trachea.

Table 1:

Selected materials approaches to sealing fetal membranes after surgical puncture

Material	Status	Notes	Selected Literature	Material performance
Maternal platelets and fibrin cryoprecipitate, "amniopatch"	Human cases; mixed results	Used following fetal surgery <i>after iPPROM</i> but before onset of preterm labor. Slurry of platelets and cryoprecipitate injected into amniotic fluid, through FM at site distal to initial intervention hole. In some cases, transvaginal fluid leakage ceased, but FM seal at defect site hard to confirm. One series reported fetal or neonatal death of 1 fetus in 11 of 21 cases [38]. Some intrauterine fetal deaths attributed to platelet overactivation.	Quintero et al., 1999 [50]: $n = 7$ cases. 3 healthy infants. Quintero, 2001 [38]: $n = 21$. 11 pregnancies with 1 healthy infant. O'Brien et al., 2002 [51]: $n = 1$ case, amniopatch + gelatin sponge. Healthy infant Young et al., 2004 [26]: $n = 8$ cases. 6 with no evidence of AF leakage from puncture site. Richter , et al. 2013 [52]: $n = 24$. 58% amniopatch success rate; 55% survival to discharge.	Formation of sealing plug at defect site difficult to document; mechanical properties akin to blood clot, not robust. Amniopatch method utilized only after onset of iPPROM; does not decrease incidence of membrane rupture.
Collagen or gelatin plug. For example decellularized amnion [41] or porcine-skin derived gelatin	Some in-human use; no improvement relative to the standard of care (no treatment)	As laparoscopic instruments are removed from the amniotic cavity, a small gelatin or collagen plug is left behind in the membrane defect, like a tampon.	Chang, et al., 2006 [37]: n = 27 TTTS laser coagulation cases. PPROM rate 4.2% attributed to "meticulous technique and atraumatic insertion and removal of ports." Engels, et al., 2014 [49]: n = 54 with plug; n = 87 without plug. No evidence that collagen reduces risk of PPROM after minimally invasive CDH repair. Papana et al., 2010 [22]: n = 79 TTTS laser coagulation cases. PPROM rate = 34%.	Plug rapidly swells with amniotic fluid to occlude the defect site but does not form a water-tight seal.
Commercially avail	able surgical glues			
Fibrin glues (e.g., Tisseal)	Benchtop evaluations and animal studies; some in-human use for other applications	Several groups have attempted to study the performance of commercially available surgical glues to seal the FM following surgical puncture. While some have been evaluated in animal or human trials and show promise, others were eliminated during phases of benchtop testing due to their poor biological or mechanical properties.	 Bilic, et al., 2009 [19]: Compared biological and adhesive properties of surgical glues for FM sealing. Burke, et al., 2007 [53]: Compared adhesive properties of fibrin and mussel-inspired tissue adhesives. Haller, et al., 2012 [54]: Evaluated FM tissue sealing properties of glues using burst device. Devaud, et al., 2019 [55]: Comparative studies using novel delivery device to apply sealants to <i>ex vivo</i> human fetal membranes in benchtop uterine models. 	Poor adhesion to wet tissues.
Cyanoacrylate (Dermabond, Histoacryl)			Bilic, et al., 2009 [19]. Devaud, et al., 2019 [55].	Dermabond: poor cytocompatibility, not for application to wet wounds (per manufacturer). Histoacryl: cytocompatible, adheres to tissues.
BioGlue (2- component bovine serum albumin/ glutaraldehyde glue)			Azandani, et al., 2009 [56]: Comparison of mechanical properties of vascular glues. Bures, et al., 2015 [57]: <i>In vitro</i> evaluation of BioGlue for sealing lung defects	Mismatch of mechanical properties relative to FM. When cured, BioGlue elastic modulus is 3.1 ± 1.6 MPa [56]
SprayGel (2- component multi-			Bilic , et al., 2009 [19]	Poor adhesion to wet tissues

Material	Status	Notes	Selected Literature	Material performance
arm PEG modified				
with NHS ester and lysine)				
CoSeal (2- component multi- arm PEG with NHS esters and thiols)			Spotnitz & Burks , 2008 [58]	Swelling up to 400%; cannot be used in confined spaces such as the FM-uterus interface. Skin sensitization issues in animals.
Duraseal (2- component solution of PEG ester and trilysine amine)			Spotnitz & Burks, 2008 [58]	Swelling <i>in vivo</i> up to 50%; potential for wound-site infections.
Adhesives designed	specifically for fetal	membrane sealing		
Catechol-PEG (cPEG)	Promising animal data	Multi-arm PEG modified with mussel-inspired catechol groups	 Bilic, et al., 2009 [19]. Haller, et al., 2012 [54]: cPEG improved fetal survival in a rabbit model of fetal membrane sealing. Kivelio, et al., 2013 [59]. Devaud, et al., 2019 [55]. 	Catechol-mediated wet adhesion superior to other injectable glues. Biocompatibility and material cohesion could be improved
Nanosilica coacervate glue + decellularized amnion (DAm)	Studied in porcine FM sealing model	Nanosilica coacervate glue was used to adhere sheets of DAm to seal swine FM defects, but no significant difference was found between treatment and control groups.	Mann, et al., 2012 [41]: Benchtop evaluation of coacervate glue. Papanna, et al, 2015 [60]: Mini- swine FM sealing study; inconclusive as swine membranes healed spontaneously.	Further work needed to identify the properties of this adhesive system and validate in a non-self- healing animal fetal membrane model
Tissue engineering	and other approache	28		
Laser welding	Not suitable for FM sealing	Laser welding with albumin solders was attempted <i>in vitro</i> to seal a FM defect. This method was not effective in sealing membrane defects	Petratos , et al., 2002 [42].	Laser welding produces worse adhesion than either sutures or polymeric sealants.
Membrane- mimetic sheets	Preliminary benchtop studies, some animal studies	Non-adhesive sheets are sutured, glued, or placed in or on membrane defects. Cell infiltration and sealing ability is assessed.	Roman, et al., 2016 [61]: Electrospum polymer bi-layer with elastic properties similar to the fetal membranes. Phase: Benchtop testing. Ochsenbein-Kölble, et al., 2006 [44]: Comparing decellularized amnion sheets to polyesterurethane sheets in rabbit model of FM puncture. Pensabene, et al., 2015 [62]: Ultrathin poly-L-lactic acid film adheres to uterus and exposed FM following puncture in rabbit model. Adhesion mechanism unclear.	Most materials are non- adhesive, but approach is promising for both iatrogenic and spontaneous PPROM
Precipitated egg white	Bench top studies	Precipitated egg whites were assessed for ability to plug fluid leaks human FM in a benchtop model	Mendez-Figueroa , et al., 2010 [63].	Translation potential unclear as unvalidated in animal work.
Tissue engineering <i>de novo</i> FM from hPSCs	Preliminary / basic science	Shao, et al., 2017 [64]: Protocol to cells. In the future, this could be ex	differentiate human pluripotent sten spanded to create implantable FM tis	n cells (hPSCs) into amnion sues for FM repair.

Table 2:

Materials approaches for defect repair in MMC and gastroschisis.

Material	Status	Selected Literature	Material performance
Surrounding tissue stretched to cover defect	Standard of care in fetal surgery to correct MMC; used postnatally to cover gastroschisis & omphalocele defects	 Adzick et al., 2011 [3]: Repairing MMC defect in utero via open fetal surgery superior to post-natal repair in large human randomized controlled trial. Stephenson, et al., 2010 [84]: Gastrochisis repair successful in 2/2 fetal sheep via open surgery. 	Surrounding tissue can successfully be used to cover MMC or gastroschisis defects, but <i>in utero</i> , this approach likely necessitates an open surgery approach. Also, in some gastroschisis and omphalocele cases, surrounding tissue is not large enough to stretch across the defect.
Gelatin/collagen sponges laced with bFGF and adhered to defect with cyanoacrylate adhesive	Successful rat and sheep studies	 Watanabe, et al., 2010 [74]: Successful repair of MMC defect in fetal rat model using gelatin sheet/gelatin sponge combination, adhered to tissue with cyanoacrylate and laced with bFGF. Epithelial and vascular cell ingrowth into sponges. Watanabe, et al., 2016 [83]: Successful repair of MMC defect in ovine model using gelatin/collagen sponges laced with bFGF and with or without gelatin sheet covering. Histology revealed epithelial layers covering the defect as well as neovascularization. 	Promising. Further work needed to fully characterize materials properties of sponge, including mechanical properties and cellular response to biomaterial, and to investigate if these findings could be reproduced using laparoscopic surgery. Materials did not cause inflammation of MMC defect site.
Biocellulose film	Sheep studies	 Sanchez e Oliveira, et al., 2007 [85]: In sheep model, biocellulose films were placed atop exposed spinal tissue before skin was closed around the defect <i>in utero</i>. Film used to prevent cord tethering sometimes associated with MMC repair. Papanna, et al., 2016 (1) [86]: In sheep model, biocellulose films were attached with sandcastle worm-inspired sealants [41] that were cured with 532 nm laser light at 200 mW for 10 s/cm². Films dislodged, and no defect repair was seen. 	Biocellulose films are still candidate materials, but attachment method is important. This sandcastle-worm inspired adhesive seems unsuitable for this application.
Cryopreserved human umbilical cord (HUC) + sutures	Sheep studies promising and ongoing; early human case reports successful	 Papanna, et al., 2016 (1) [86]: In sheep model, HUC was used to patch MMC defect <i>in utero</i>. Repair was excellent, including almost full skin coverage and layered tissue regeneration. Papanna, et al., 2016 (2) [87]: Case report in 2 human patients. Promising results. Hindbrain herniation was reversed and minimal cord tethering was found at birth. 	Promising. Further studies to validate the method and compare to <i>in utero</i> repair without patches are needed.
Placenta-derived mesenchymal stromal cells seeded onto porcine small intestine submucosa-derived ECM	Fetal rat study yielded promising results	Chen et al., 2018 [88]: In rat MMC model, decreased spinal cord deformity and apoptosis seen in placental mesenchymal stromal cell-seeded ECM compared to ECM-only scaffold repair in fetal rats.	Promising. More work needed in larger animal models.