# UC San Diego UC San Diego Previously Published Works

# Title

Monitoring Neutrophil-Expressed Cell Surface Esophageal Cancer Related Gene-4 after Severe Burn Injury

**Permalink** https://escholarship.org/uc/item/9qm713j2

**Journal** Surgical Infections, 16(6)

**ISSN** 1096-2964

# **Authors**

Costantini, Todd W Coimbra, Raul Lopez, Nicole E <u>et al.</u>

Publication Date

2015-12-01

# DOI

10.1089/sur.2014.209

Peer reviewed

# Monitoring Neutrophil-Expressed Cell Surface Esophageal Cancer Related Gene-4 after Severe Burn Injury

Todd W. Costantini, Raul Coimbra, Nicole E. Lopez, Jeanne G. Lee, Bruce Potenza, Alan Smith, Andrew Baird, and Brian P. Eliceiri

## Abstract

**Background:** We identified recently esophageal cancer related gene-4 (*ECRG4*) as a candidate cytokine that is expressed on the surface of quiescent polymorphonuclear leukocytes (PMNs) and shed in response to ex vivo treatment with lipopolysaccharide. To investigate the potential biologic relevance of changes in cell surface *ECRG4* in human samples, we performed a pilot study to examine a population of burn patients in whom blood could be analyzed prospectively. We hypothesized that cutaneous burn injury would alter cell surface expression of *ECRG4* on PMNs.

*Methods:* Patients admitted with more than 20% total burn surface area (TBSA) (n = 10) had blood collected at the time of admission and weekly thereafter. For comparison, blood was obtained from a control group of healthy human volunteers (n = 4). We used flow cytometry to measure changes in *ECRG4*<sup>+</sup> PMNs from patients during recovery from injury. Esophageal cancer related gene-4 expression at each time point was compared with the patient's clinical status based on a Multiple Organ Dysfunction (MOD) score.

**Results:** Esophageal cancer related gene-4 was detected on the PMN surface of cells collected from healthy volunteers, however, within 48 h of admission after burn injury (n=10 patients), the number of PMNs with cell surface *ECRG4* was decreased. Esophageal cancer related gene-4 expression in PMNs was re-established over the course of patient recovery, unless complications occurred. In this case, the decrease in cell surface *ECRG4*<sup>+</sup> PMNs preceded the clinical diagnosis of infectious complications and was reflected by increased organ injury scores. *Conclusion:* From a small sample set, we were able to determine that PMN cell surface *ECRG4* expression was decreased after burn injury and returned to baseline during recovery from injury. Although larger studies are needed to define the role of *ECRG4* in human PMNs further, this report is the first assessment of cell surface *ECRG4* protein in a patient population to support analogous findings in animal studies.

**S** EVERE BURN AND TRAUMA can elicit an overwhelming inflammatory response that is characterized by dysfunctional innate immunity and multiple organ failure [1–3]. Gene profiling of circulating leukocytes after such injuries suggests that there is an induction of a genomic storm that activates and/or suppresses pathways regulating innate immunity and adaptive immunity respectively [4,5]. Whereas all phases of the injury response influence the final clinical outcome, none is more relevant than the role played by the inflammatory "set point" at the time of injury. Accordingly, it is critical to define how these inflammatory set points are established and how they affect the injury response because they pre-define the capacity, duration, and intensity of the

injury response and might account for differences in outcomes observed in individuals exposed to identical injuries.

We have focused on the potential for esophageal cancer related gene-4 (*ECRG4*) to serve as a sentinel factor on immune cells that may regulate the immune set point based on its expression on the surface of leukocytes, its release from the cell surface after inflammatory stimuli, and its epigenetic regulation that may explain differences in Ecrg4 expression between individuals [6–9]. Esophageal cancer related gene-4 encodes a 148 amino acid precursor protein that is tethered to the cell surface but shed on activation [6,10]. We have demonstrated previously that *ECRG4* is present on the surface of polymorphonuclear leukocytes (PMNs) and, on ex

Division of Trauma, Surgical Critical Care, and Burn, Department of Surgery, University of California San Diego Health Sciences, San Diego, California.

Presented at the Thirty-second Annual Meeting of the Surgical Infection Society, Dallas, Texas, April 18-21, 2012.

vivo activation by agonists such as lipopolysaccharide (LPS), *ECRG4* is released and can be detected by immunoblotting and proteomic analyses of biologic fluids including conditioned media, cerebrospinal fluid, and serum [6,7,10–12]. We have also demonstrated that *ECRG4* interacts with the human innate immunity receptor complex (TLR4-CD14-MD2) supporting further an important role in responding to inflammatory insults such as severe injury [13].

We sought to advance these findings by determining whether leukocytes from burn-injured patients exhibit analogous losses of cell surface *ECRG4*. If so, we reasoned that *ECRG4* could serve as potential as a surrogate marker of injury responsiveness. This study describes the results of a serial analysis of cell surface *ECRG4* on PMNs collected from patients over their 3- to 10-week recovery from severe burn injury. Esophageal cancer related gene-4 was monitored in clinical peripheral blood samples by flow cytometry and *ECRG4*<sup>+</sup> PMNs found sensitive to burn injury.

#### **Patients and Methods**

#### Ethics statement

The University of California San Diego Institutional Review Board approved study participants, protocols, and consent forms. Study participants provided written informed consent to participate in this study.

### Patients

Blood samples were collected from 10 male burn patients with a greater than 20% total body surface area (TBSA) burn admitted to the University of California San Diego Burn Center between June 2011 and March 2012. All patients were treated based on modern burn patient care protocols with aggressive fluid resuscitation, early enteral feeds, and ventilator support when indicated. Blood was collected at the time of admission and weekly thereafter. For comparison, blood was obtained from a control group comprising healthy male volunteers 20–60 years old (n=4). Demographic data, burn injury location, burn depth, mechanism of injury, presence of inhalation injury, hospital length of stay, morbidity, and mortality were recorded. Clinical status and the presence of organ failure was assessed using the Multiple Organ Dysfunction (MOD) score [14]. Clinical and laboratory data for each patient were obtained prospectively.

### Leukocyte preparation and staining

Primary staining was performed with affinity-purified anti-*ECRG4* (Genway, San Diego, CA) and compared with isotypematched control antibodies as described previously [6]. Leukocytes were prepared from heparinized blood after red blood cell lysis and fixed in Cytofix (Becton Dickson, San Jose, CA). Flow cytometry was performed with a FACSCalibur and data analysis performed with FloJo (Treestar, Ashland, OR). Analysis of *ECRG4*<sup>+</sup> staining was gated on PMNs. Cell surface *ECRG4* expression was verified with CD16<sup>+</sup> surface staining of parallel cell populations using a second anti-*ECRG4* antibody isotype (Phoenix Pharmaceuticals, Burlingame, CA).

#### Data set mining of gene chip assays

In a recent study from the Inflammation and Host Response to Injury Program [4], a database of gene expression in human burn patients was made available publicly (www.ncbi.nlm .nih.gov/geo accession number GSE19743). The database was searched for c2orf 40, the human gene encoding *ECRG4* and its expression in the peripheral blood of burn patients compared with expression in healthy volunteers.

## Statistics

Statistical analysis was conducted using Microsoft Excel (Microsoft, Redmond, WA). Comparison of *ECRG4* expression from serial analysis after injury was performed using analysis of variance (ANOVA). Bivariate correlations were analyzed using simple least squares regression, and relations between parameters that did not show a linear relation were evaluated using polynomial regression.

#### Results

# Neutrophil ECRG4 expression decreases post-burn injury

Based on our recent report that membrane-anchored *ECRG4* is expressed on human leukocytes and is shed after ex vivo activation [6], we enrolled severely burned patients (more than 20% TBSA) presenting to the UCSD Burn Center and performed a serial analyses of cell surface *ECRG4* concentrations on PMNs using flow cytometry. Demographic data and the severity of burns in this patient population are presented in Table 1. The patients were all males ranging in age from 21 to 57 years (mean, 37.8) and presenting with

	Age	Gender	%TBSA	% Full thickness burn	Mechanism of injury	Inhalation injury	LOS	Mortality
BP 1	34	М	60	60	Flame	Yes	93	No
BP 2	21	Μ	65	65	Flame	Yes	114	No
BP 3	38	Μ	34	0	Flame	Yes	25	No
BP 4	29	Μ	35	8	Flame	No	17	No
BP 5	50	Μ	40	5	Flame	Yes	82	No
BP 6	24	Μ	45	25	Flame	Yes	84	No
BP 7	50	Μ	20	0	Scald	No	48	No
BP 8	26	Μ	70	40	Flame	Yes	39	No
BP 9	51	Μ	15	0	Flame	Yes	31	No
BP 10	57	М	50	20	Flame	No	65	No

TABLE 1. BURN PATIENT ENROLLMENT DATA

TBSA=total body surface area; LOS=length of stay; BP=burn patient.



**FIG. 1.** Neutrophil esophageal cancer related gene-4 (*ECRG4*) expression decreases after burn injury. (**A**) Flow cytometry of peripheral blood from normal, healthy control groups to gate polymorphonuclear cell (PMN) population. (**B**) Specificity of *ECRG4* antibody by flow cytometry. Histograms of the gated PMN populations illustrated in (**A**) display cell surface  $ECRG4^+$  PMNs (gray line) compared with isotype antibody control (black line). (**C**) Effect of severe burn injury on expression of *ECRG4* on PMN cell surface. (**D**) Flow cytometry at three different times after severe burn. Analyses of mean fluorescent intensity (MFI) of each peak reflects the decreasing loss of cell surface *ECRG4* on PMNs after burn injury.

burns of 34% to 90% TBSA that required up to 114 days of hospitalization.

Flow cytometry was used to measure the expression of *ECRG4* on the surface of PMNs (Fig. 1). Comparison of *ECRG4* expression with isotype control groups (Fig. 1B) and with healthy volunteer control groups (Fig. 1C) indicated that monitoring of cell surface *ECRG4* on PMNs could reveal changes in the activation state of PMNs in the burn injured patient population

[6]. We observed a time-dependent loss of cell surface *ECRG4* on PMNs over two to seven days post-burn injury (Fig. 1D).

## Neutrophil ECRG4 expression return to baseline during recovery from injury

We performed a serial analysis of neutrophil *ECRG4* expression from burn patients during their recovery from injury



**FIG. 2.** Serial analysis of esophageal cancer related gene-4 (*ECRG4*<sup>+</sup>) polymorphonuclear cells (PMNs) during recovery from burn injury. (**A**) Serial flow cytometric analysis of PMN *ECRG4* expression from patients admitted after a greater than 20% total body surface area (TBSA) burn injury (n=7). \*p<0.05 versus week 3 using analysis of variance (ANOVA). (**B**) Esophageal cancer related gene-4 expression measured from a separate cohort of burn patients (n=3) using flow cytometric analysis gating on CD16<sup>+</sup> PMNs and stained with a second anti-*ECRG4* antibody isotype (Phoenix Pharmaceuticals, Burlingame, CA). (**C**) Data mining a publicly available database for leukocyte *ECRG4* expression in a cohort of patients with greater than 20% TBSA burn [5] revealed a significant reduction in *ECRG4* gene expression among severely burned patients (p<0.05).

where clinical status was measured by calculating MOD score at the time of blood sample collection. Neutrophil *ECRG4* expression was similar to healthy control groups when measured at the time of admission. This was followed by a decrease in PMN *ECRG4* expression of up to 60% within two to three weeks post-burn injury (Fig. 2A). We also found that PMN *ECRG4* expression returned to baseline concentrations after recovery from injury, with *ECRG4* concentrations similar to healthy volunteer *ECRG4* expression by six weeks post-injury. To confirm our findings from fluorescenceactivated cell sorting (FACS) analysis of *ECRG4* expression, we performed a serial analysis from a second cohort of burninjured patients using an analysis gating on CD16+ PMNs and stained with a second anti-*ECRG4* antibody isotype (Fig. 2B).

## Leukocyte ECRG4 expression is decreased in burn-injured patients enrolled in the inflammation and host response to injury database

To determine whether our study of *ECRG4* expression after burn injury was supported by independent gene ex-

pression data, we examined gene expression findings in the Inflammation and Host Response to Injury database [5], which includes leukocytes from volunteers (n=63) and burn patients (n=67) with more than 20% TBSA burn. Whereas these databases did not distinguish gene expression in specific leukocyte sub-populations (i.e., PMNs versus monocytes), they nevertheless revealed a statistically significant reduction (p<0.05) in *ECRG4* expression in leukocytes of severely burned patients (Fig. 2C). Because circulating PMNs and monocytes are the primary source of cell surface *ECRG4* [6], these data support a role for *ECRG4* expression in the inflammatory cell response to burn injury.

# Changes in neutrophil ECRG4 expression are associated with changes in clinical status during recovery from injury

Whereas clinical scores such as the MOD score and other organ dysfunction scores are known to be of somewhat limited predictive value for the complications that arise in severe burn patients, this MOD score was a useful tool to correlate



**FIG. 3.** Changes in neutrophil esophageal cancer related gene-4 (*ECRG4*) expression are associated with changes in clinical status. Representative time-courses demonstrating changes in polymorphonuclear cell (PMN) PMN *ECRG4* from individual burn patients correlated with Multiple Organ Dysfunction (MOD) score in brackets. Regression analysis was performed on patients with a length of stay of at least six weeks to compare the concentrations of Ecrg4<sup>+</sup> PMNs versus MOD score. (A) BP1 ( $R^2$ =0.78); (B) BP2 ( $R^2$ =0.62); and (C) BP5 ( $R^2$ =0.25). BP=burn patient.

#### PMN ECRG4 EXPRESSION AFTER INJURY

with *ECRG4*<sup>+</sup> PMNs. Using this combination of measuring *ECRG4*<sup>+</sup> PMN concentrations and MOD score, we analyzed the relation between PMN ECRG4 expression and organ injury in a patients with at least five weekly samples collected after admission after burn injury (Fig. 3). To address directly the possibility that the concentrations of  $ECRG4^+$  PMNs could be associated with changes in clinical status, we performed regression analyses of the concentrations of ECRG4<sup>+</sup> PMNs versus MOD score. The comparison demonstrated that there is an association between ECRG4<sup>+</sup> PMNs and the MOD score when there are at least six weekly samples collected over the length of stay (range of  $R^2$  values = 0.25–0.78; Fig. 3A-3C). These findings support the possibility that ECRG4 may serve a sentinel function on the surface of PMNs with expression concentrations changing based on the clinical status of the patients.

#### Discussion

The kinetics of cytokine and cell surface marker expression post-injury have been studied widely in human leukocytes after injury [15–18]. Yet the predictive power of global genetic changes has been disappointing [4,5]. In an effort to address this issue, we used a more targeted approach for gene analyses [6]. First, we mined gene databases [19] for open reading frames that encode proteins that are restricted to the human secretome because we reasoned that if these genes are expressed in leukocytes [6] and regulated by trauma-burn [4,5] they might be involved in maintaining set points of leukocyte homeostasis. In addition, their inclusion in secretome databases [19] would select genes that encode proteins found outside the cell and hence more likely involved in cell-cell signaling. Similarly, their inclusion in the neuropeptidome [20,21] would select genes that encode ligandcandidates and hence more likely to be involved in cell-cell communication. Finally, we reasoned that the presence of CpG islands in their promoter would point to epigenetic regulation by DNA methylation and hence predict that gene expression is affected by both environment [22,23] and aging [24], both of which may alter outcome after trauma-burn. One of the few genes to meet all these criteria is c2orf40, a human open reading frame that encodes the candidate tumor ECRG4.

In this clinical pilot study we analyzed *ECRG4*, a candidate sentinel factor expressed on the surface of PMNs that is shed on injury from the cell surface. In a series of experiments using peripheral blood from 10 burn-injured patients we demonstrated *ECRG4* expression on the surface of PMNs of normal volunteers and after severe burn injury. The decrease of surface *ECRG4* protein that was detected by flow cytometry was consistent with our previous in vitro study demonstrating evidence that *ECRG4* is expressed highly on normal PMNs and decreased on activation/injury [6,25]. Using regression analyses, we studied the association between *ECRG4* expression and MOD score as the basis to correlate gene expression with patient clinical status, and observed that expression of PMN *ECRG4* is sensitive to injury and organ dysfunction.

We and others have recently shown that ECRG4 gene expression can be influenced by epigenetic mechanisms [6,22,23], therefore, it is interesting to speculate that differences in the initial expression of ECRG4 and the inflamma-

tion response in general have an underlying epigenetic mechanism [26]. If so, the partial recovery of  $ECRG4^+$  PMN concentrations during hospitalization in some patients after burn injury might trace back to the epigenetic regulation of ECRG4 gene expression. This is an active topic of investigation that we are pursuing by measuring ECRG4 promoter hypermethylation in patient samples.

Burn and trauma injury share many, albeit not all, inflammatory responses. Accordingly it is noteworthy that a recently published gene micro-array database from trauma patients has been mined for *ECRG4* gene expression in leukocytes and similar to burn, *ECRG4* was also decreased after severe injury [6]. As in the case of burn patients, the decrease in *ECRG4* gene expression is likely a combination of decreased surface *ECRG4* protein expression as well as reduced expression of mRNA.

In the studies reported here, the sample size of the patient cohort limits our ability to draw definitive conclusions but the findings are nevertheless consistent with genomic databases, in vitro studies, and the proposed function of cell surface ECRG4 in gauging the response to injury, presumably by modulating the inflammatory response. Larger studies are ongoing to determine whether ECRG4 can be useful in analyzing the clinical trajectory of patients with severe injury or provide insight into why individuals with similar burntrauma injuries can have significantly different clinical outcomes. In conclusion, our findings support the hypothesis that ECRG4 is candidate sentinel factor that is shed from the surface of PMNs after injury with expression improving to baseline concentrations after recovery from injury. These findings provide further insight into the neutrophil inflammatory response to burn injury.

#### Acknowledgments

The authors would like to thank Lindsey Prescher, MD, for her assistance in analyzing clinical data and Ann-Marie Hageny for her technical assistance with this project.

Funded by the National Institutes of Health P20 Exploratory Center grant for Wound Healing Research (P20GM078421), American Burn Association/Department of Defense (W81XWH-10-1-0527).

#### Author Disclosure Statement

No competing financial interests exist.

#### References

- 1. Paterson HM, Murphy TJ, Purcell EJ, et al. Injury primes the innate immune system for enhanced toll-like receptor reactivity. J Immunol 2003;171:1473–1483.
- Schwacha MG, Chaudry IH. The cellular basis of post-burn immunosuppression: Macrophages and mediators. Int J Mol Med 2002;10:239–243.
- 3. Saffle JR, Sullivan JJ, Tuohig GM, Larson CM. Multiple organ failure in patients with thermal injury. Crit Care Med 1993;21:1673–1683.
- Xiao W, Mindrinos MN, Seok J, et al. A genomic storm in critically injured humans. J Exp Med 2011;208:2581–2590.
- Zhou B, Xu W, Herndon D, et al. Analysis of factorial time-course microarrays with application to a clinical study of burn injury. Proc Natl Acad Sci USA 2010;107:9923– 9928.

- 6. Baird A, Coimbra R, Dang X, et al. Cell surface localization and release of the candidate tumor suppressor Ecrg4 from polymorphonuclear cells and monocytes activate macrophages. J Leukoc Biol 2012;91:773–781.
- 7. Podvin S, Gonzalez AM, Miller MC, et al. Esophageal cancer related gene-4 is a choroid plexus-derived injury response gene: Evidence for a biphasic response in early and late brain injury. PloS One 2011;6:e24609.
- Shaterian A, Kao S, Chen L, et al. The candidate tumor suppressor gene Ecrg4 as a wound terminating factor in cutaneous injury. Arch Dermatol Res 2013;305:141–149.
- Mori Y, Ishiguro H, Kuwabara Y, et al. Expression of ECRG4 is an independent prognostic factor for poor survival in patients with esophageal squamous cell carcinoma. Oncol Rep 2007;18:981–985.
- Dang X, Podvin S, Coimbra R, et al. Cell-specific processing and release of the hormone-like precursor and candidate tumor suppressor gene product, Ecrg4. Cell Tissue Res 2012;348:505–514.
- 11. Kurabi A, Pak K, Dang X, et al. Ecrg4 attenuates the inflammatory proliferative response of mucosal epithelial cells to infection. PloS One 2013;8:e61394.
- Kao S, Shaterian A, Cauvi DM, et al. Pulmonary preconditioning, injury, and inflammation modulate expression of the candidate tumor suppressor gene ECRG4 in lung. Exp Lung Res 2015;41:162–172.
- Podvin S, Dang X, Meads M, et al. Esophageal cancerrelated gene-4 (ECRG4) interactions with the innate immunity receptor complex. Inflamm Res 2015;64:107–118.
- 14. Buckley TA, Gomersall CD, Ramsay SJ. Validation of the multiple organ dysfunction (MOD) score in critically ill medical and surgical patients. Intensive Care Med 2003;29: 2216–2222.
- Jeschke MG, Finnerty CC, Herndon DN, et al. Severe injury is associated with insulin resistance, endoplasmic reticulum stress response, and unfolded protein response. Ann Surg 2012;255:370–378.
- Kobayashi M, Jeschke MG, Asai A, et al. Propranolol as a modulator of M2b monocytes in severely burned patients. J Leukoc Biol 2011;89:797–803.
- Winfield RD, Delano MJ, Cuenca AG, et al. Obese patients show a depressed cytokine profile following severe blunt injury. Shock 2012;37:253–256.

- Minei JP, Cuschieri J, Sperry J, et al. The changing pattern and implications of multiple organ failure after blunt injury with hemorrhagic shock. Crit Care Med 2012;40:1129– 1135.
- Clark HF, Gurney AL, Abaya E, et al. The secreted protein discovery initiative (SPDI), a large-scale effort to identify novel human secreted and transmembrane proteins: A bioinformatics assessment. Genome Res 2003;13:2265–2270.
- Delfino KR, Southey BR, Sweedler JV, Rodriguez-Zas SL. Genome-wide census and expression profiling of chicken neuropeptide and prohormone convertase genes. Neuropeptides 2010;44:31–44.
- 21. Tadross JA, Patterson M, Suzuki K, et al. Augurin stimulates the hypothalamo-pituitary-adrenal axis via the release of corticotrophin-releasing factor in rats. Br J Pharmacol 2010;159:1663–1671.
- 22. Gotze S, Feldhaus V, Traska T, et al. ECRG4 is a candidate tumor suppressor gene frequently hypermethylated in colorectal carcinoma and glioma. BMC Cancer 2009;9:447.
- 23. Wang YB, Ba CF. Promoter methylation of esophageal cancer-related gene 4 in gastric cancer tissue and its clinical significance. Hepatogastroenterology 2012;59:1696–1698.
- 24. Kujuro Y, Suzuki N, Kondo T. Esophageal cancer-related gene 4 is a secreted inducer of cell senescence expressed by aged CNS precursor cells. Proc Natl Acad Sci USA 2010;107:8259–8264.
- 25. Baird A, Lee J, Podvin S, et al. Esophageal cancer-related gene 4 at the interface of injury, inflammation, infection, and malignancy. Gastrointest Cancer 2014;2014:131–142.
- Adcock IM, Tsaprouni L, Bhavsar P, Ito K. Epigenetic regulation of airway inflammation. Curr Opin Immunol 2007;19:694–700.

Address correspondence to: Dr. Todd W. Costantini Division of Trauma, Surgical Critical Care, Burns and Acute Care Surgery Department of Surgery University of California San Diego Health Sciences 200 West Arbor Dr., #8896 San Diego, CA 92103

E-mail: tcostantini@ucsd.edu