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# Population Pharmacokinetics of Fluconazole in Premature Infants with Birth Weights Less than 750 Grams

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Fluconazole is an effective agent for prophylaxis of invasive candidiasis in premature infants. The objective of this study was to characterize the population pharmacokinetics (PK) and dosing requirements of fluconazole in infants with birth weights of <750 g. As part of a randomized clinical trial, infants born at <750 g birth weight received intravenous (i.v.) or oral fluconazole at 6 mg/kg of body weight twice weekly. Fluconazole plasma concentrations from samples obtained by either scheduled or scavenged sampling were measured using a liquid chromatography-tandem mass spectrometry assay. Population PK analysis was conducted using NONMEM 7.2. Population PK parameters were allometrically scaled by body weight. Covariates were evaluated by univariable screening followed by multivariable assessment. Fluconazole exposures were simulated in premature infants using the final PK model. A population PK model was developed from 141 infants using 604 plasma samples. Plasma fluconazole PK were best described by a one-compartment model with first-order elimination. Only serum creatinine was an independent predictor for clearance in the final model. The typical population parameter estimate for oral bioavailability in the final model was 99.5%. Scavenged samples did not bias the parameter estimates and were as informative as scheduled samples. Simulations indicated that the study dose maintained fluconazole troughs of >2,000 ng/ml in 80% of simulated infants at week 1 and 59% at week 4 of treatment. Developmental changes in fluconazole clearance are best predicted by serum creatinine in this population. A twice-weekly dose of 6 mg/kg achieves appropriate levels for prevention of invasive candidiasis in extremely premature infants.

nvasive candidiasis is a common cause of death and neurodevelopmental impairment in extremely premature infants (1). Fluconazole, a triazole antifungal drug that exhibits fungistatic activity against a variety of *Candida* species, is an effective agent as a prophylaxis for treatment of invasive candidiasis in this population (2–4). Fluconazole exhibits pharmacokinetic (PK) characteristics that make it an attractive candidate for prevention of *Candida* infections. It has a long half-life allowing infrequent administration, is minimally (12%) bound to plasma proteins, penetrates the cerebrospinal fluid, and achieves saliva and lung concentrations that are 1.3 and 1.2 times the plasma concentrations, respectively, thereby providing higher concentrations at key areas of colonization (5–7). Additionally, in adults, fluconazole has very high (>90%) oral bioavailability (8, 9).

Previous PK studies in infants suggested increasing fluconazole clearance (CL) over the first postnatal weeks (10–13). However, the PK of fluconazole in infants of <750 g birth weight have not been extensively evaluated, and it is this population that has the highest risk for invasive candidiasis where prophylaxis has the most potential for benefit (14). Therefore, the objective of this study was to characterize the population PK and dosing requirements of fluconazole in infants of <750 g birth weight.

## MATERIALS AND METHODS

**Study design.** This PK study was associated with a multicenter, randomized, placebo-controlled trial that evaluated the efficacy and safety of fluconazole in preventing death or invasive candidiasis in premature infants weighing <750 g at birth (4). Inclusion criteria required that infants were <120 h old at the time of randomization and of <750 g birth weight and that informed consent was received from a legally authorized representative. Participants were excluded from the trial if they had significant liver dysfunction (aspartate aminotransferase [AST] and alanine aminotransferase [ALT], >250 U/liter), renal dysfunction (serum creatinine [SCR], >2 mg/dl), a diagnosis of invasive candidiasis or congenital *Candida* infection, or a history of hypersensitivity to any azole antifungal. Participants randomized to fluconazole therapy received 6 mg/kg of body weight twice weekly (Tuesdays and Fridays) for up to 42 days of treatment. The fluconazole dose was administered either by an intravenous (i.v.) infusion given over approximately 60 min or orally (in infants that were receiving enteral medications). Clinical data were collected and included demo-

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graphic information (gestational age [GA], postnatal age [PNA], birth weight, current weight, race, sex, and ethnicity), laboratory values collected within the 72 h prior to the first fluconazole dose and during treatment (serum creatinine [SCR], total bilirubin, alanine transaminase [ALT], and albumin [ALB]), details of concomitant medications of interest (all antimicrobials and vasopressors), intubation status, mode of delivery (Cesarean section or vaginal), and microbiological cultures from sterile sites. The study was approved by the Institutional Review Boards at each center, and informed consent was obtained from a legally authorized representative.

PK sample collection. Participants were randomized to 1 of 8 sampling schemes with a maximum of 3 timed blood samples. Each infant had two PK samples drawn after a single dose taken around the time of administration of dose 3, 5, 7, or 9 and one sample taken around the time of administration of the final dose. Samples obtained from infants receiving placebo were used for other purposes. One additional plasma PK sample was requested from infants who developed invasive candidiasis. Up to 10 scavenged samples left over from laboratory blood samples obtained per routine care were also collected for PK evaluation. Sample collection consisted of whole blood (200  $\mu$ l) collected in EDTA tubes. Timed blood samples were processed within 6 h of collection, and plasma was stored at  $-80^{\circ}$ C until analysis for fluconazole concentration determination.

**Bioanalytical assay.** Plasma samples were analyzed for fluconazole concentrations using a validated liquid chromatography method with tandem mass spectrometric detection (LC-MS/MS). The lower limit of quantitation for fluconazole was 10 ng/ml. The precision determined at each concentration level did not exceed 8.5% of the coefficient of variation, including the lower limit of quantification.

Population PK analysis. Concentration-time data were analyzed with nonlinear mixed-effect modeling using NONMEM version 7.2 (Icon; Ellicott City, MD, USA). Plasma samples were excluded from the analysis if they (i) were below the limit of quantitation (<10 ng/ml), (ii) were collected >120 h after the last recorded fluconazole dose, (iii) demonstrated increased fluconazole concentrations (compared to prior samples) without a recorded dose, or (iv) were extreme outliers (>10-fold difference between observed and predicted concentrations). A 1-compartment model (ADVAN2, TRANS2 subroutine) and a first-order conditional estimation method (FOCE with interaction) were used to describe the fluconazole concentration data. Plasma concentrations following intravenous and oral administration were simultaneously analyzed, allowing estimation of absolute oral bioavailability. Parameters of the model were the absorption rate constant  $(k_a)$ , volume of distribution (V), clearance (CL), and bioavailability (F1). Diagnostic plots were executed in PLT Tools 5.1.0 (PLTSoft; San Francisco, CA), SAS 9.3 (Cary NC), and R Project 3.0.1 (downloaded from a website of the University of California, Los Angeles, Los Angeles, CA). The bootstrap procedure was performed using WINGS for NONMEM version 7.2 (Auckland, NZ), and 1,000 bootstrap sample data sets were generated. Consistent with prior PK analyses of fluconazole, a 1-compartment model with first-order absorption was selected for development. Population PK parameters were scaled by body size prior to evaluation of potential covariates. Clearance was scaled by allometric weight (WT<sup>0.75</sup>), and volume of distribution was scaled by weight (WT<sup>1.0</sup>). The initial model used a combined proportional error and additive residual error value. Diagnostic plots were used to assess the appropriateness of this structure for the base model.

Once the base model was identified, covariates were investigated for their potential influence on PK parameters, CL, and volume (V). Continuous covariates were evaluated by normalization to median values and included PNA, GA, postmenstrual age (PMA), SCR, and ALB. Categorical covariates included race and ethnicity, intubation status, and mode of delivery (Cesarean section or vaginal). Missing covariate values were imputed using the closest value available for that participant and either a carry-forward approach or a backfill approach, depending on which date was closest. The investigation of the relationship between potential covariates and PK parameters proceeded by developing the base population

TABLE 1 Patient demographic and clinical data at first PK evaluation<sup>a</sup>

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Parameter	Value
PNA (days)	23 (3-47)
GA (wks)	24.7 (22.6–28.7)
PMA (wks)	28.3 (23.7–35.1)
Wt (g)	710 (345–2,680)
Serum creatinine (mg/dl)	0.7 (0.1–3.6)
Albumin (g/dl)	2.5 (1.0-4.7)
Male	40
Intubation status	81
Delivery by Cesarean section	67
Race	
American Indian or Alaska Native	5
Asian	1
Black or African American	53
White	40

<sup>*a*</sup> *n* = 141. Values presented as median (range), except for sex, intubation status, delivery by Cesarean section, and race, which are presented as percentages.

PK model and *post hoc* generation of the Bayesian estimates of individual PK parameters. Individual subject etas ( $\eta$ ), representing deviation from the typical population parameter values, were generated. Graphical assessment of the relationships between PK parameters and potential covariates was performed by plotting etas versus potential clinically relevant covariates. Covariates with an evident graphical relationship to  $\eta$ CL and  $\eta$ *V* were evaluated for inclusion in the final model. A forward-addition, backward-elimination approach to covariate selection was used when two or more covariates were found to be significant for CL or *V*. The threshold for the significance of a single covariate was reduction of the objective function value (OFV) by >7.88.

**Model evaluation.** Model evaluation included successful minimization, goodness-of-fit plots, precision-of-parameter estimates, bootstrap procedures, and visual predictive checks. The precision of the final population PK model parameter estimates was evaluated using nonparametric bootstrapping (1,000 replicates) to generate the 95% confidence intervals for parameter estimates. The final model was used to perform Monte Carlo simulations in 14,100 virtual subjects with demographic and laboratory characteristics simulated from the same distribution as the study population. The simulated trough fluconazole concentrations were determined during an 8-week course of fluconazole and predose concentrations compared to a minimum target of 2,000 ng/ml. This target was selected based upon typical drug MICs for *Candida* species in infants.

#### RESULTS

Study population and PK samples. A total of 141 premature infants weighing <750 g at birth who received i.v. or oral fluconazole were included (Table 1), and fluconazole concentrations were determined from 619 plasma samples obtained from the infants. Of these, 15 samples (2.4%) were excluded, resulting in 604 plasma samples available for population PK modeling. Samples were excluded for the following reasons: concentration below the limit of quantitation (n = 7); collection >120 h after the last dose, indicating an incomplete dosing history (n = 3); outlier concentrations based on individual predictions (IPRED) and population predictions (PRED) (n = 3); and increased concentrations compared to prior samples without a documented dose (n = 2). The majority of PK samples (n = 368, 61%) were from scavenged samples. The PK collection time after dose was most frequently the first 6 h postdose, with an overall median (range) of 30 h (0 to 115 h). The medians (ranges) of the fluconazole concentrations



**FIG 1** Goodness-of-fit plots for the base model. (A) Predicted versus observed concentrations (Conc). (B) Individual (Indiv) predicted versus observed concentrations. (C) Conditional weighted residuals (CWRES) versus population predictions. (D) Conditional weighted residuals versus time after dose. For panels A and B, the line of identity is included as a reference. For panels C and D, a solid line at y = 0 is included as a reference.

were 4,144 ng/ml (491 to 14,050 ng/ml) from scavenged samples and 6,154 ng/ml (320 to 13,167 ng/ml) from timed samples.

**Population PK model building.** A 1-compartment model with first-order absorption and with combined proportional error plus additive residual error adequately described the concentration versus time data (Fig. 1). Due to limited numbers of early samples after oral administration in the data set, between-subject variability (BSV) was not estimated for the absorption rate constant ( $k_a$ ). Data from the base fluconazole population PK model of observed and conditionally weighted residuals versus population and individual-predicted concentrations were evenly scattered at the line of unity with no obvious biases.

Graphical plots of GA, PNA, PMA, SCR, and mode of delivery were suggestive of a relationship with  $\eta$ CL, which was consistent with the significant changes in OFV that resulted during the uni-

variable screen assessment of these covariates for CL (Table 2). Plots of PMA and SCR were suggestive of a relationship with  $\eta V$ , and both of these covariates met the OFV reduction criteria. All of the potential covariates identified in the univariate screen were incorporated into a combined model for backward elimination assessment. Although PMA had the second greatest impact on OFV during the univariable screen, it is a function of PNA and GA, both of which were also identified as potential covariates. The multivariable process started with the elemental components of PMA (PNA and GA) rather than PMA itself. Sequential removal was performed in the reverse order of the magnitude of OFV change seen with the covariate in the univariable screening process. Attempts to remove GA (CL), PNA (CL), and SCR (CL) each resulted in increases of >10 in the OFV, and the parameters were thus deemed significant independent covariates. Finally, a model

**TABLE 2** Summary of key univariable population PK model buildingprocess<sup>a</sup>

			Change in OFV		
Model description	Population model	OFV	from base model		
CL (base model)	$\mathrm{CL} = \theta_{\mathrm{CL}} * (\mathrm{WT})^{0.75}$	9,624			
PNA	$CL = \theta_{CL} * (WT)^{0.75 *}$ $(PNA/25)^{\theta CL-PNA}$	9,492	-132		
GA	$CL = \theta_{CL} * (WT)^{0.75} *$ $(GA/25)^{\theta CL-GA}$	9,599	-25		
PMA	$CL = \theta_{CL} * (WT)^{0.75} *$ $(PMA/28)^{\theta CL-PMA}$	9,450	-174		
SCR	$CL = \theta_{CL} * (WT)^{0.75} *$ $(SCR/0.8)^{\theta CL-SCR}$	9,405	-219		
CSCT	$CL = \theta_{CL} * (WT)^{0.75} * \\ \theta CL - CSCT^{CSCT}$	9,617	-7		
V (base model)	$V = \theta_V^* (WT)^{1.0}$	9,624			
PMA	$V = \theta_V^* (WT)^{1.0 *} (PMA/28)^{\theta V-PMA}$	9,620	-4		
SCR	$V = \theta_V * (WT)^{1.0} * (SCR/0.8)^{\theta V-SCR}$	9,600	-24		

<sup>a</sup> OFV, objective function value; CSCT, delivery by Cesarean section.

using SCR (CL) and PMA (CL) as a function of GA and PNA was assessed and performed better than the model with SCR (CL), GA (CL), and PNA (CL) with an OFV reduction of 37.3 despite having one fewer covariate. The model-estimated absolute oral bioavailability of fluconazole was 100% in the final model, which is in agreement with prior adult data.

No significant relationships were observed between fluconazole CL or V and sex, race, ethnicity, intubation, or mode of infant delivery. The typical population PK parameter estimates in the final model were as follows: CL (liters/h/kg<sup>0.75</sup>) = 0.0127 \* (SCR/ 0.8)<sup>-0.41</sup> \* (PMA/28)<sup>2.05</sup>; V (liters/kg) = 1.00;  $k_a$  (1/h) = 0.96; F1 = 100% (where SCR is in milligrams per deciliter and PMA is in weeks). BSV was estimated as 23% for CL, 13% for V, and 25% for F1. The final PK parameters are displayed in Table 3. The model was evaluated using a 1,000-set bootstrap analysis with the program WINGS for NONMEM; 98.7% of bootstrap data sets converged to  $\geq$ 3 significant digits. The medians of bootstrap

TABLE 3 F	inal PK	model	parameters'
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fixed-effect parameter estimates were within 1% of population estimates from the original data set for all parameters (Table 3). The visual predictive check indicated that the model adequately described the data, with 3.1% of observed values >95th percentile and 4.1% <5th percentile (Fig. 2).

The potential influence of the collection type on the final model was assessed by fitting a reduced data set that contained only timed samples to the final model structure. The parameter estimates using this reduced data set were within 10% of those estimated with the full data set, indicating that the scavenged samples did not bias the PK parameter estimates and were as informative as the scheduled samples. Further, the visual predictive check demonstrated that the model adequately captured fluconazole concentrations irrespective of the sample type (Fig. 2). While the residual error was somewhat smaller when limiting the analysis to scheduled samples, of particular interest, the CL estimated from timed samples was altered by only 6% compared to that estimated from the full data set (0.0119 versus 0.0127 liters/h/kg<sup>0.75</sup>).

Using Monte Carlo simulations, the fluconazole exposure from doses of 6 mg/kg twice weekly was assessed. The trough fluconazole concentrations were determined during an 8-week course of fluconazole, and predose concentrations were compared to a minimum target of 2,000 ng/ml. This threshold was exceeded in 80% of simulated infants at week 1 and in 59% of simulated infants at week 4 of fluconazole prophylaxis. This is consistent with 95.7% of the first measured fluconazole concentrations being >2,000 ng/ml and with 89.9% of the overall fluconazole concentrations being >2,000 ng/ml (Fig. 3).

### DISCUSSION

The present study evaluated the population PK of fluconazole in 141 premature infants of <750 g birth weight receiving twiceweekly fluconazole for 42 days for candidiasis prophylaxis. This represents a particularly difficult study population for collecting PK samples, and the use of scavenged samples more than doubled the data set size, leading to improved parameter estimates. In this population, fluconazole CL is low and is associated with a long half-life, which allows infrequent administration. Oral bioavailability appears to be high, as has been demonstrated in older populations, and fluconazole may therefore be given at the same dose

Parameter	Symbol	Point estimate	SEE	Bootstrap CI		
				2.5%	Median	97.5%
V (liters/kg)	$\theta_V$	1.00	0.0378	0.93	1.00	1.08
CL (liters/h/kg <sup>0.75</sup> )	$\theta_{\rm CL}$	0.0127	0.00033	0.0120	0.0127	0.0133
F1 (%)	$\theta_{F1}$	1.00	0.065	0.86	1.00	1.13
$k_{\rm a}$ (1/h)	$\theta_{KA}$	0.96	0.25	0.52	0.96	1.81
SCR-CL	$\theta_{SCR}$	-0.410	0.0498	-0.53	-0.41	-0.32
PMA-CL	$\theta_{\rm PMA}$	2.05	0.35	1.23	2.05	2.62
Interindividual variance (CV%)						
V	$\omega_V^2$	13	61	1	13	18
CL	$\omega^{2}_{CL}$	23	29	15	22	27
F1	$\omega_{F1}^2$	31	73	1	22	50
Residual variance (CV%)	$\sigma^2$	46	27	37	46	51
Additive value (ng/ml)	$\sigma^2$	505	329	5	495	858

<sup>a</sup> CI, confidence interval; CV%, percent coefficient of variation; SEE, standard error of estimate.



FIG 2 Visual predictive check of the final model for scheduled samples (A) and scavenged samples (B), displaying time after first dose.

in young infants and administered either as an oral suspension or intravenously. The population PK model identified SCR as the best predictor of developmental changes in fluconazole CL. Given fluconazole's renal CL in adults, this is not surprising. Although SCR levels decrease with age and are significantly correlated with measures of maturation, PMA was also independently associated with fluconazole CL. The individual impacts of GA and PNA on CL were modest and not of a magnitude to justify dose stratification in this population during the first 30 days of life. Overall, CL and V were in the range expected based on prior neonatal fluconazole PK studies. In the study by Wade et al., the typical CL was 0.015 liters/h/kg<sup>0.75</sup> for a typical infant (GA, 26 weeks; PNA, 2 days) (10). This is similar to our typical CL value of 0.0127 liters/ h/kg<sup>0.75</sup> at 28 weeks PMA. Wade et al. also found that GA, PNA, and SCR were significant covariates for fluconazole CL in infants (10).

This study employed scavenged PK sampling, which is a minimal-risk approach that uses leftover blood collected in the course of routine clinical care that would otherwise be discarded (15). The scavenged samples were as informative as the scheduled samples and did not bias parameter estimates. Scavenged PK sampling is particularly effective in populations that are difficult to study, such as infants, and has been successfully used in population PK studies of anti-infection drugs (16, 17). In addition, scavenged sampling is useful for drugs with long half-lives where traditional sampling schemes may not capture the full PK profile.

Effective prophylaxis dosing in adults suggests that fluconazole concentrations of >2,000 ng/ml would be beneficial. Further, fluconazole MICs for Candida species in infants typically range from 250 to 4,000 ng/ml (18-20). In this study, 90% of measured fluconazole concentrations were >2,000 ng/ml, and the Monte Carlo simulations predict that trough concentrations would be maintained at >2,000 ng/ml in a high proportion of participants receiving 6 mg/kg twice weekly for the first few weeks of life. Maturation of renal function and other developmental processes would result in lower fluconazole concentrations at later PNA. Another commonly used fluconazole regimen for the prophylaxis of invasive candidiasis is 3 mg/kg given twice weekly (2, 3). Although the lower fluconazole regimen of 3 mg/kg twice weekly may be effective for *Candida* species with an MIC of  $\leq 2,000$  ng/ ml, our results suggest that the higher 6 mg/kg twice-weekly regimen would be needed to optimize fluconazole exposure for the first few weeks of life, when the fluconazole MIC range for Can*dida* species is >2,000 ng/ml.

This study had several limitations. Only 2 infants with PK data had invasive candidiasis, which precluded any meaningful exploration regarding fluconazole exposure and outcomes. Additionally, on the basis of typical MICs of *Candida* species in infants, we



FIG 3 Monte Carlo simulations of fluconazole given at 6 mg/kg intravenously (IV) twice weekly using the final population PK model displaying simulated fluconazole concentrations (left) and proportion of fluconazole troughs of >2,000 ng/ml (right). Min, minimum.

chose a pharmacodynamic target of 2,000 ng/ml by which to evaluate fluconazole dosing regimens. Alternative fluconazole dosing strategies may be appropriate in instances where the MIC differs significantly from 2,000 ng/ml. Additionally, MIC-based approaches to guide antifungal drug selection and dosing do not take into consideration the complexity of host-drug-microbe interactions, which may limit the power of correlation with patient outcomes. Finally, since only about 20% of the PK samples were collected following oral administration and since there was limited sampling available during the absorption phase, no BSV was estimated for  $k_a$ .

In summary, the population PK of fluconazole were successfully characterized in premature infants of <750 g birth weight using sparse sampling that included scavenged samples. Much of the variability in fluconazole CL was explained by SCR and, to a lesser extent, by PMA. SCR was confounded with PNA and GA, and SCR explained most of the BSV in CL, limiting the impact of age on CL. The 6 mg/kg twice-weekly dosage given by either i.v. or oral administration maintained trough fluconazole concentrations of >2,000 ng/ml in the vast majority of infants for the first few weeks of life. While the clinical outcomes of this trial did not show a survival benefit (4), the fluconazole prophylaxis dosage used appears to be appropriate for maintaining fluconazole trough concentrations above 2,000 ng/ml in extremely premature infants.

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