

UCSF

UC San Francisco Previously Published Works

Title

A genome-wide association and admixture mapping study of bronchodilator drug response in African Americans with asthma

Permalink

<https://escholarship.org/uc/item/9zf6q8mf>

Journal

The Pharmacogenomics Journal, 19(3)

ISSN

1470-269X

Authors

Spear, Melissa L

Hu, Donglei

Pino-Yanes, Maria

et al.

Publication Date

2019-06-01

DOI

10.1038/s41397-018-0042-4

Peer reviewed



Published in final edited form as:

Pharmacogenomics J. 2019 June ; 19(3): 249–259. doi:10.1038/s41397-018-0042-4.

A Genome-wide Association and Admixture Mapping Study of Bronchodilator Drug Response in African Americans with Asthma

Melissa L. Spear, BS¹, Donglei Hu, PhD², Maria Pino-Yanes, PhD^{3,4,5}, Scott Huntsman, MS², Celeste Eng, BS², Albert M. Levin, PhD⁶, Victor E. Ortega, MD, PhD⁷, Marquitta J. White, PhD, MS², Meghan E. McGarry, MD, MAS⁸, Neeta Thakur, MD, MPH², Joshua Galanter, MD^{1,2,9}, Angel C.Y. Mak, PhD², Sam S. Oh, PhD, MPH², Elizabeth Ampleford, PhD⁷, Stephen P. Peters, MD, PhD⁷, Adam Davis, MA, MPH¹⁰, Rajesh Kumar, MD, MS¹¹, Harold J. Farber, MD, MSPH¹², Kelley Meade, MD¹³, Pedro C. Avila, MD¹⁴, Denise Serebrisky, MD^{15,16}, Michael A. Lenoir, MD¹⁷, Emerita Brigino-Buenaventura, MD¹⁸, William Rodriguez Cintron, MD¹⁹, Shannon M. Thyne, MD²⁰, Jose R. Rodriguez-Santana, MD²¹, Jean G. Ford, MD²², Rocio Chapela, MD²³, Andrés Moreno Estrada²⁴, Karla Sandoval²⁴, Max A. Seibold, PhD²⁵, Cheryl A. Winkler, PhD²⁶, Eugene R. Bleecker, MD²⁷, Deborah A. Myers, PhD²⁷, L. Keoki Williams, MD, MPH^{28,29}, Ryan D. Hernandez, PhD^{1,30,31}, Dara G. Torgerson, PhD², and Esteban G. Burchard, MD, MPH^{1,2}

¹Department of Bioengineering and Therapeutic Sciences, University of California, San Francisco, San Francisco, CA, 94158

²Department of Medicine, University of California, San Francisco, San Francisco, CA, 94158

³Research Unit, Hospital Universitario N.S. de Candelaria, Universidad de La Laguna, Tenerife, Spain, 38010

⁴CIBER de Enfermedades Respiratorias, Instituto de Salud Carlos III, Madrid, Spain, 28029

⁵Genomics and Health Group, Department of Biochemistry, Microbiology, Cell Biology and Genetics, Universidad de La Laguna (ULL), La Laguna (Tenerife), Spain, 38010

⁶Department of Public Health Sciences, Henry Ford Health System, Detroit, MI

⁷Department of Internal Medicine, Wake Forest Baptist Medical Center, Winston Salem, NC. 27157

⁸Department of Pediatrics, University of California, San Francisco, San Francisco, CA, 94143

⁹Department of Epidemiology and Biostatistics, University of California, San Francisco, San Francisco, CA, 94158

Users may view, print, copy, and download text and data-mine the content in such documents, for the purposes of academic research, subject always to the full Conditions of use:http://www.nature.com/authors/editorial_policies/license.html#terms

Corresponding author: Esteban G. Burchard, MD, MPH, Department of Bioengineering & Therapeutic Sciences and Medicine, University of California, San Francisco, 1550 4th St. RM584B, San Francisco, CA 94158, Telephone: (415) 514-9677, Fax: 415-514-4365, esteban.burchard@ucsf.edu.

Conflict of Interest:

The authors declare no competing conflicts of interest.

10. UCSF Benioff Children's Hospital Oakland, Center for Community Health and Engagement
11. Ann & Robert H. Lurie Children's Hospital of Chicago, Pediatrics, Chicago, Illinois, 60614
12. Department of Pediatrics, Section of Pulmonology, Baylor College of Medicine and Texas Children's Hospital, Houston, TX, 77030
13. UCSF Benioff Children's Hospital Oakland, Oakland, California
14. Division of Allergy-Immunology, Northwestern University Feinberg School of Medicine, Chicago, Illinois, 60611
15. Pediatric Pulmonary Division, Jacobi Medical Center, Bronx, NY, 10461
16. Albert Einstein College of Medicine, Pediatrics, Bronx, New York
17. Bay Area Pediatrics, Oakland, CA, 94609
18. Department of Allergy & Immunology, Kaiser Permanente-Vallejo Medical Center, Vallejo, CA, 94589
19. Veterans Caribbean Health System, San Juan, Puerto Rico, 00921
20. Department of Pediatrics, David Geffen School of Medicine at ULCA, Olive View-UCLA Medical Center, Sylmar, CA, 91342
21. Centro de Neumologia Pediatrica, San Juan, Puerto Rico, 00917
22. Columbia University, New York, NY, 21205
23. Instituto Nacional de Enfermedades Respiratorias, Mexico City, MX, 14080
24. National Laboratory of Genomics for Biodiversity (LANGEBIO), CINVESTAV, Irapuato, Guanajuato, MX, 36821
25. Department of Pediatrics, National Jewish Health, Denver, CO, 80206
26. Basic Research Laboratory, National Cancer Institute, Leidos Biomedical Research, Frederick National Laboratory, Frederick, MD, 21701
27. Department of Medicine, The University of Arizona, Tucson, Arizona, 85724
28. Center for Health Policy and Health Services Research, Henry Ford Health System, Detroit, MI, 48202
29. Department of Internal Medicine, Henry Ford Health System, Detroit, MI, 48202
30. California Institute for Quantitative Biosciences (QB3), University of California, San Francisco, CA, 94158
31. Institute for Human Genetics, University of California, San Francisco, CA, 94158

Abstract

Short-acting β_2 -adrenergic receptor agonists (SABAs) are the most commonly prescribed asthma medications worldwide. Response to SABAs is measured as bronchodilator drug response (BDR), which varies among racial/ethnic groups in the U.S.^{1, 2}. However, the genetic variation that contributes to BDR is largely undefined in African Americans with asthma³. To identify genetic

variants that may contribute to differences in BDR in African Americans with asthma, we performed a genome-wide association study (GWAS) of BDR in 949 African American children with asthma, genotyped with the Axiom World Array 4 (Affymetrix, Santa Clara, CA) followed by imputation using 1000 Genomes phase III genotypes. We used linear regression models adjusting for age, sex, body mass index (BMI) and genetic ancestry to test for an association between BDR and genotype at single nucleotide polymorphisms (SNPs). To increase power and distinguish between shared vs. population-specific associations with BDR in children with asthma, we performed a meta-analysis across 949 African Americans and 1,830 Latinos (Total=2,779). Lastly, we performed genome-wide admixture mapping to identify regions whereby local African or European ancestry is associated with BDR in African Americans. We identified a population-specific association with an intergenic SNP on chromosome 9q21 that was significantly associated with BDR (rs73650726, $p=7.69 \times 10^{-9}$). A trans-ethnic meta-analysis across African Americans and Latinos identified three additional SNPs within the intron of *PRKG1* that were significantly associated with BDR (rs7903366, rs7070958, and rs7081864, $p=5 \times 10^{-8}$). Our results failed to replicate in three additional populations of 416 Latinos and 1,615 African Americans. Our findings indicate that both population specific and shared genetic variation contributes to differences in BDR in minority children with asthma, and that the genetic underpinnings of BDR may differ between racial/ethnic groups.

INTRODUCTION

Albuterol, a short-acting β_2 -adrenergic receptor agonist (SABA), is the most commonly prescribed asthma medication worldwide^{4, 5}. SABAs cause rapid smooth muscle relaxation of the airways. Bronchodilator drug response (BDR) is a measure of a patient's clinical response to SABA treatment and is quantitatively assessed as a change in forced expiratory volume in one second (FEV₁) after administration of a SABA. BDR is a complex trait involving interactions among inflammatory cells⁶, airway epithelium⁷, smooth muscle cells⁸, and the autonomic nervous system⁹. Variation in BDR is likely influenced by both population-specific and shared environmental and genetic factors¹⁰⁻¹². In the United States (U.S.), BDR in children with asthma differs significantly between racial/ethnic groups^{2, 10}. Specifically, African Americans have lower BDR compared to European populations even after controlling for asthma severity¹³. Compared to European Americans, African Americans suffer increased asthma morbidity and mortality^{2, 11, 14} and decreased BDR likely contributes to these disparities in disease progression and outcomes. The extensive use of albuterol as a first-line therapy for asthma, coupled with the decreased drug response (BDR) and increased disease burden in African Americans underscores the importance of identifying genetic factors that influence BDR in African American children with asthma. Once identified, these factors may lead to the generation of novel therapies and targeted interventions that will serve to improve patient care and asthma outcomes in an overburdened and under-studied population.

To date, knowledge of genetic variation that contributes to BDR in African Americans is limited to a single genome-wide association study (GWAS) in 328 individuals³. Previous GWAS and candidate gene studies performed in populations of predominantly European ancestry with asthma have identified several BDR candidate genes^{12, 15-24}. A recent study in

Latinos with asthma replicated a number of these findings, and also identified novel population-specific associations with BDR¹⁰. Genetic effects identified in one population are not always generalizable across populations and several population-specific asthma-risk variants have been discovered in African-descent populations (e.g., African Americans and Latinos)^{25–27}. Additionally, previous studies have shown that the varying degrees of African and European ancestry present in the African American population can be leveraged, through a technique known as admixture mapping, to identify the missing heritability of complex traits²⁸. Admixture mapping is a genome-wide approach that uses the variable allele frequencies of multiple SNPs between different ancestral populations to test for an association between local ancestry and phenotype²⁸. The likelihood of population-specific effects, the limited number and scale of prior studies of BDR performed in African Americans, and ability to perform admixture mapping analysis highlights the possibility of gaining novel information through evaluating the impact of common genetic factors on BDR in African American children with asthma.

In this study, we performed a GWAS and admixture mapping study of bronchodilator drug response in 949 African American children with asthma from the Study of African Americans, Asthma, Genes & Environments (SAGE I and II)²⁹. To increase power and distinguish between population-specific vs. shared associations, we also performed a trans-ethnic meta-analysis across our SAGE I and SAGE II participants and 1,830 Latinos from GALA II (Genes-environments and Admixture in Latino Americans) studies²⁶, respectively (total N=2,779). We further attempted replication of our population-specific and trans-ethnic meta-analysis results in 416 Latinos from the Genetics of Asthma in Latino Americans study (GALA I)^{11, 30}, 1,325 African Americans from the Study of Asthma Phenotypes and Pharmacogenomic Interactions by Race-Ethnicity (SAPPHIRE)^{30, 31} and 290 African Americans from the Severe Asthma Research Program (SARP)^{32, 33}.

METHODS

Study subjects from the Study of African Americans, Asthma, Genes & Environments

The Study of African Americans, Asthma, Genes & Environments (SAGE) is an ongoing case-control study of asthma in children and adolescents recruited from the San Francisco Bay Area in California²⁹. Subjects were eligible if they were 8–21 years of age and self-identified all four grandparents as African American. Exclusion criteria included: (1) 10 or more pack-years of smoking; (2) any smoking within 1 year of recruitment date; (3) pregnancy in the third trimester; or (4) history of one of the following conditions: sickle cell disease, cystic fibrosis, sarcoidosis, cerebral palsy, or history of heart or chest surgery. Asthma was defined by physician diagnosis, asthma medication use and reported symptoms of coughing, wheezing, or shortness of breath in the 2 years preceding enrollment. Detailed clinical measurements were recorded for each individual whom DNA was collected from. In addition, trained interviewers administered questionnaires to obtain baseline demographic data, as well as information on general health, asthma status, social, and environmental exposures. Pulmonary function testing was conducted with a KoKo® PFT Spirometer (nSpire Health Inc., Louisville, CO) according to American Thoracic Society recommendations³⁴, to obtain forced expiratory volume in one second (FEV₁) in addition to

other standard measurements of airway obstruction. Subjects with asthma were instructed to withhold their bronchodilator medications for at least 8 hours before testing. After completing baseline spirometry, subjects were given albuterol administered through a metered-dose inhaler (90 mcg/puff) with a spacer, and spirometry was repeated after 15 minutes to obtain post-bronchodilator measurements. The dose of albuterol was different in early stages of SAGE recruitment (2001–2005: SAGE I) than in more recent participants (2006–present: SAGE II). In SAGE I, post-bronchodilator FEV₁ values were measured after providing the participants 2 puffs of albuterol (180 µg) if they were younger than 16 years of age and 4 puffs of albuterol (360 µg) if they were 16 years of age or older. In SAGE II, two doses of albuterol were delivered. For the first dose, 4 puffs of albuterol (360 µg) were provided independently of the age of the participant. For the second dose, two puffs (180 µg) for children < 16 years old were administered and 4 puffs for subjects older 16 years.

Body mass index (BMI) was calculated for each participant using weight and height measures and converted to a categorical scale of underweight, normal, overweight, and obese according to the Centers for Disease Control and Prevention. For participants under 20 years old, standardized sex- and age-specific growth charts were used to calculate BMI percentiles (<http://www.cdc.gov/nccdphp/dnpao/growthcharts/resources/sas.htm>) and categorize their BMI as: underweight (BMI percentile < 5th), normal (5th BMI < 85th), overweight (85th BMI < 95th), and obese (BMI ≥ 95th). For participants 20 years and older, BMI categories (http://www.cdc.gov/healthyweight/assessing/bmi/adult_bmi/index.html - interpreted Adults) were defined as: underweight (BMI < 18.5), normal (18.5 BMI < 25), overweight (25 BMI < 30) and obese (BMI ≥ 30). Further information about SAGE can be found in the Supplementary Text Supplementary Table 1.

Institutional review boards approved the study and all subjects/parents provided written assent/consent, respectively.

Genotyping and quality control (SAGE)

A total of 1,821 samples (1,011 asthma cases and 810 controls) were genotyped with the Axiom® World Array 4 (Affymetrix, Santa Clara, CA) at ~800,000 SNPs. Quality control was performed by removing SNPs that failed manufacturer's quality control, had genotyping call rates below 95%, and/or had a deviation from Hardy-Weinberg equilibrium ($p < 10^{-6}$) within controls. 772,135 genotyped SNPs passed quality control. Samples were filtered based on discrepancy between genetic sex and reported gender and cryptic relatedness (PI_HAT > 0.3). We excluded 3 subjects who were outliers for BDR (BDR of > 60, or < -10). After sample quality control we included 759 SAGE II and 190 SAGE I asthma cases, for a total of 949 individuals with both genome-wide SNP data and measurements of bronchodilator drug response in the current study (Table 1). Phasing of genotyped SNPs was performed using SHAPE-IT³⁵, and genotype imputation was performed using IMPUTE2^{36, 37} using all populations from 1000 Genomes Project Phase III³⁸ as a reference. Following imputation, a total of 9,573,507 genotyped and imputed (info score > 0.3) SNPs with a MAF > 0.05 were analyzed for SAGE II and 9,605,653 were analyzed for SAGE I.

Study subjects from the Genes-environments & Admixture in Latino Americans study (GALA II)

A total of 1,830 Latino children with asthma genotyped with the Axiom LAT1 array (World Array 4, Affymetrix) were included in our analysis (Table 1). Asthma cases were defined in a similar manner as SAGE with detailed clinical measurements recorded for each individual whom DNA was collected from. Additionally, each individual underwent spirometry with BDR calculated as the percentage change in FEV₁ after 2 doses of albuterol (post-FEV₁) compared with baseline values before administration of albuterol (pre-FEV₁). Post-bronchodilator FEV₁ values were measured after providing the participants 2 doses of albuterol, with a 15-minute waiting period after each dose. A total of 6 (if <16 years of age) to 8 (if ≥16 years of age) puffs of albuterol were administered. A total of 408 patients from the Centro de Neumologia Pediátrica in Puerto Rico were recruited based on having a BDR of at least 8%; of these, 121 patients were recruited based on having a BDR of at least 12%. Further details about GALA II are described in the Supplementary Text, Supplementary Table 1 and in depth elsewhere¹⁰. Imputation procedures identical to those described above for SAGE I and SAGE II were implemented, resulting in a total of 7,498,942 genotyped and imputed (info score >0.3) SNPs with a MAF>0.05.

Study subjects from the Genetics of Asthma in Latino Americans study (GALA I)

Our replication phase included 247 Mexican and 169 Puerto Rican asthma cases genotyped with the Genome-Wide Human SNP Array 6.0 (Affymetrix). Subjects were included in the study if they were between the ages of 8–40 with physician diagnosed mild to moderate-severe asthma and had experienced two or more symptoms during the two years preceding time of recruitment (including wheezing, coughing and/or shortness of breath.). BDR was measured in a similar way to GALA II, but with a lower dosage of albuterol. Specifically, post-FEV₁ values were measured after only a single dose of albuterol (compared with 2 doses in GALA II). Two (if <16 years of age) to 4 (if ≥16 years of age) total puffs of albuterol were administered (compared with 4 [if <16 years of age] and 6 [if ≥16 years of age] in GALA II). Further details of the study are described in the Supplementary Text, Supplementary Table 1 and elsewhere^{11, 30}.

Study subjects from the Study of Asthma Phenotypes and Pharmacogenomic Interactions by Race-Ethnicity (SAPPHIRE)

For additional replication, we included 1,325 Africans Americans with asthma from SAPPHIRE³ genotyped with the Genome-Wide Human SNP Array 6.0 (Affymetrix). Subjects met the following criteria: age 12–56 years, had a diagnosis of asthma (based on both patient report and documentation in the medical record), did not have a prior diagnosis of chronic obstructive pulmonary disease or congestive heart failure (CHF), a baseline FEV₁ between 40–90% predicted, >12% baseline bronchodilator reversibility, no smoking in the preceding year or <10 pack-year smoking history total, no oral or inhaled corticosteroid use in the 4 weeks preceding screening, and not pregnant at the time of enrollment and not intending to get pregnant during the study period. Spirometry testing was performed using a KoKo® PFT Spirometer, (nSpire Health Inc., Louisville, CO) following 2005 ATS/ERS spirometry recommendations³⁴. Patients with asthma who were using inhaled

bronchodilators were asked to withhold these medications for the 12 hours prior to spirometry tests. To assess BDR a 360 µg dose (i.e., 4 puffs) of inhaled albuterol sulfate hydrofluoroalkane (HFA) (GlaxoSmithKline, Research Triangle Park, NC) from a standard metered dose inhaler (MDI) using an AeroChamber Plus Flow-Vu® spacer (Monahan Medical Corp., Plattsburgh, NY) was administered to patients. Pulmonary function was reassessed 15 minutes after administering albuterol. BDR was measured as the change in forced expiratory volume at one second (FEV₁) between the baseline (pre-bronchodilator) measure and post-bronchodilator FEV₁. Estimates of local ancestry were obtained using RFMix³⁹.

Study subjects from the Severe Asthma Research Program (SARP)

We included 290 African Americans with mild to severe asthma from SARP genotyped with the Illumina 1Mv1 platform²⁵. SARP is a comprehensively characterized cohort with a range of asthma severities from mild to severe, but was enriched for severe disease defined by the American Thoracic Society (ATS) criteria for refractory asthma. Subjects met the definition of severe persistent asthma^{32, 33, 40}. A physician's diagnosis of asthma was confirmed by evidence of methacholine bronchial hyperresponsiveness or bronchodilator reversibility and documented asthma symptoms. Baseline pre-bronchodilator spirometry was performed after withholding long and short-acting bronchodilators. Post-bronchodilator FEV₁ measurements were performed by increasing doses of albuterol of 200µg (two inhalations) up to a maximum dose of 800µg (eight inhalations).

Assessment of genetic ancestry

Genotypes from two populations were used to represent the ancestral haplotypes of African Americans for estimating local ancestry: HapMap European (CEU) and HapMap Africans (YRI). For Latinos, genotypes from 71 Native Americans were used as an additional ancestral population⁴¹. These 71 individuals included: 14 Zapotec, 2 Mixe, and 11 Mixtec from the southern State of Oaxaca⁴² and 44 Nahua individuals from Central Mexico⁴³. Global ancestry was estimated using ADMIXTURE⁴⁴ in a supervised analysis assuming two ancestral populations for African Americans and three ancestral populations for Latinos. Local ancestry was estimated using the program LAMP-LD⁴² in the GALA and SAGE studies and with RFMix in SAPHIRE³⁹.

Genotype association testing

All statistical analyses were conducted using R (version 2.15.3). For SAGE individuals, we used standard linear regression to test for an association between BDR and allele dosage at each individual SNP, adjusting for age, sex, BMI category, and both global and local African ancestry. A GWAS of BDR in GALA II has been previously published¹⁰. However, since this previous work did not include adjustment for BMI, we re-ran the GWAS using a new reference imputation panel and further adjusted by BMI in the present study⁴⁵. For GALA II individuals, we adjusted for age, sex, BMI category, ethnicity, global Native American and African ancestry, and local ancestry. All analyses were performed using imputed genotypes from 1000 Genomes phase III. Using the fixed-effects model implemented in METAL⁴⁶, we performed a meta-analysis of common variants (MAF ≥ 5%) across African Americans (SAGE I and SAGE II) and Latinos (GALA II). We selected variants that were common

(MAF \geq 5%) within each individual study and then took the intersection of SNPs for the meta-analysis.

Admixture mapping

We used local ancestry estimates generated across the genome to perform admixture mapping in African Americans. Linear regression models adjusted for age, sex, BMI category, and global African ancestry were used to identify significant associations between local ancestry estimates and BDR. The threshold for genome-wide significance was calculated using the empirical autoregression framework with the package *coda* in R to estimate the total number of ancestral blocks^{47, 48}. The Bonferroni threshold was calculated as $\alpha=2.4 \times 10^{-4}$ based on 245 ancestral blocks. For African Americans, admixture mapping was performed separately in SAGE I and SAGE II and combined in a meta-analysis using METAL⁴⁶. An admixture mapping study of BDR in GALA II has been previously published¹⁰, but did not include adjustment for BMI. In the current study, we re-ran the admixture mapping study further adjusting by BMI⁴⁵ to be consistent with the SAGE I and SAGE II analyses. For GALA II Latinos, linear regression models adjusted for age, sex, ethnicity, BMI category, global Native American and African ancestry were used to identify significant associations between local ancestry estimates and BDR. We further combined the African ancestry results of SAGE I, SAGE II and GALA II in a meta-analysis using METAL⁴⁶.

Replication in GALA I, SAPPHIRE, and SARP

We attempted replication of significant population-specific (SAGE I and SAGE II) and cosmopolitan (SAGE I, SAGE II, GALA II) associations with BDR in the GALA I, SAPPHIRE, and SARP studies. Replication in GALA I was performed using genotype imputation (i.e., *in silico* replication), followed by an examination at a locus-wide level for SNPs within \pm 50 kb. We imputed 100 kb regions around each SNP using the program IMPUTE2 for Mexican and Puerto Rican participants run separately using 1000 Genomes phase III haplotypes as a reference. Linear regression was used to test for an association between allele dosage and BDR separately in Mexicans and Puerto Ricans, adjusting for age, sex, BMI category, global and local ancestry. Replication in SAPPHIRE was performed using linear regression to test for an association between allele dosage and BDR in African Americans while adjusting for age, sex, BMI category, and global and local African ancestry. Replication in SARP was performed using linear regression to test for an association between allele dosage and BDR in African Americans while adjusting for age, sex, BMI, and global African ancestry. For GALA I and SAPPHIRE replication, statistical significance at the SNP level was evaluated at $p < 0.05$, and at the locus-wide level was established using a conservative Bonferroni correction adjusting by the number of SNPs within \pm 50 kb of the original candidate SNP. For SARP replication, statistical significance was evaluated at $p < 0.05$ at the SNP level only.

RESULTS

GWAS results

After filtering variants with a MAF $\geq 5\%$ and with imputation quality score (info score) ≥ 0.3 , we tested for an association of BDR at a total of 9,190,349 SNPs in 949 African Americans with asthma ($\lambda = 1.006$). We identified a single genome-wide significantly associated SNP within an intergenic region on chromosome 9 (rs73650726, imputation quality score=0.86) (Figures 1, 2, Supplementary Figure 1A, Table 2). At this variant, additional copies of the A1 allele (A), was associated with decreased drug response ($\beta = -3.8$, $p = 7.69 \times 10^{-9}$) (Table 2 & Supplementary Figure 2, Table 2). The SNP rs73650726 is common in African Americans but rare in Latinos, with a minor allele frequency of 8% in both SAGE studies, but at a frequency of 1% in GALA II. This is consistent with allele frequencies observed in the 1000 Genomes Project, where the variant is common in African populations (8%), rare in Latino populations (1–2%), and absent in European and Asian populations (Figure 3)⁴⁹.

In order to increase power and identify associations shared between populations we performed a trans-ethnic meta-analysis across African American and Latino participants from SAGE I, SAGE II, and GALA II. Following quality control and filtering on variants common in each study (MAF $\geq 5\%$), we took the overlap between the three studies and performed a meta-analysis on 6,570,864 SNPs. We identified genome-wide significant associations at three SNPs located on chromosome 10 within the intron of *PRKGI*: rs7903366 ($\beta = 1.23$, $p = 3.94 \times 10^{-8}$), rs7070958 ($\beta = -1.24$, $p = 4.09 \times 10^{-8}$), and rs7081864 ($\beta = 1.23$, $p = 4.94 \times 10^{-8}$) (imputation quality scores > 0.98 , Figures 4 & 5, Table 2, Supplementary Figures 1B & 2, Table 2). All three SNPs are in linkage disequilibrium and are eQTLs for *PRKGI* in lung tissue from the Genotype-Tissue Expression (GTEx) database (Table 3)⁵⁰, with the minor allele associated with decreased expression.

Replication of African American population-specific (rs73650726) and shared (rs7903366, rs7070958, rs7081864) variants was attempted in three independent Latino (GALA I) and African American (SAPPHIRE and SARP) studies. The African American population-specific association between rs73650726 and BDR, identified in the SAGE studies, was in the same direction in GALA I Puerto Ricans ($\beta = -6.22$) and the SAPPHIRE cohort of African Americans ($\beta = -0.65$), but in the SARP African American cohort the association was in the opposite direction ($\beta = 6.12$, $p = 0.04$) (Supplementary Table 3). In addition, none of the SNPs within 50 kb of the four original SNPs were significantly associated with BDR following Bonferroni correction (Supplementary Table 4). Lastly, we evaluated previously identified candidate SNPs from prior candidate gene and GWAS with BDR in patients with asthma. After accounting for fifteen comparisons, no SNPs met the statistical significance threshold ($p < 3.33 \times 10^{-3}$) (Supplementary Table 5); only rs9551086 in *SPATA13* had a p-value below 0.05 ($p = 0.02$).

Admixture mapping results

We tested for an association of BDR with local genetic ancestry inferred at 478,441 SNPs in 949 African Americans with asthma (190 from SAGE I and 759 from SAGE II)

(Supplementary Figures 3 & 4). A meta-analysis across both studies yielded no significant associations with ancestry ($p < 2.4 \times 10^{-4}$) (Supplementary Figure 5). The most significant peak was located on chromosome 8p11, where African ancestry was associated with higher BDR ($\beta = 1.49$, $p = 6.34 \times 10^{-4}$) (Supplementary Table 6). A meta-analysis across SAGE I, SAGE II and GALA II yielded results consistent with previous findings in the original admixture mapping study of GALA II (see ¹⁰) (Supplementary Figure 6).

DISCUSSION

We performed a genome-wide association study for bronchodilator drug response in African Americans and identified a population-specific association between BDR and rs73650726, located on chromosome 9. Specifically, we discovered that the G (A2) allele of rs73650726 was associated with increased BDR and is more common in African Americans compared to European populations (Figure 3). The variant rs73650726, located on chromosome 9, does not map to any gene, but SNPs in high linkage disequilibrium ($r^2 = 0.8$) with this marker are located in enhancer histone marks in lung tissues [36].

Our results demonstrate that population-specific genetic variation contributes to variation in BDR in African American children with asthma. We further combined our results in a meta-analysis for BDR in African Americans and Latinos and identified multiple intronic variants in *PRKGI* that were associated with BDR in both populations. Overall, our results demonstrate that population-specific and shared genetic factors contribute to variation in BDR among African American children with asthma.

Three of our significantly associated variants fell within the intronic region of an annotated gene, Protein Kinase, CGMP-Dependent, Type I (*PRKGI*). *PRKGI* encodes for a cyclic GMP-dependent protein kinase, which phosphorylates proteins involved in regulating platelet activation and adhesion⁵¹, gene expression^{52, 53}, vascular smooth muscle cell contraction⁵⁴, and feedback of the nitric-oxide (NO) signaling pathway⁵⁵. Notably, the NO pathway is a key pathway in modulating vasodilation in response to beta-agonists such as albuterol via β_2 -adrenergic receptors⁵⁶, making *PRKGI* a highly plausible BDR candidate gene. The three SNPs are in high linkage disequilibrium ($r^2 = 0.8$) with variants known to be functional⁵⁷, and are all associated with the expression of *PRKGI* in the lung – a tissue highly relevant to BDR. From the GTEx project database, the reference allele for all three SNPs was associated with decreased expression of the gene in lung tissue⁵⁰. Thus, additional studies are required to identify the causal underlying variation at this locus, such as direct sequencing of this locus, and how the expression of *PRKGI* may be related to differences in BDR.

We sought to replicate our study findings and candidate SNPs previously found to be associated with BDR. The African American population-specific SNP, rs73650726, replicated in the opposite direction in the SARP cohort which could be due to differences in study design (Supplementary Figure 7). In candidate gene studies of BDR, the gly16arg variant in the Beta-2 adrenergic receptor gene (*ADRB2*) has consistently replicated opposite effects on BDR depending on whether medication exposure was acute or chronic^{12, 58–60}. The SARP and SAGE studies administered different albuterol doses, had differences in

medication withholding periods, and SARP individuals were more likely to be treated with long-acting β_2 -adrenergic receptor agonists (LABAs) over extended periods for severe disease. Additional factors that may have impacted replication include the presence of population specific differences in genetic contributions to BDR, lack of power due to small populations sizes, and/or varying patterns of linkage disequilibrium between populations. Furthermore, we were limited in sample size in GALA I²⁵ to evaluate associations at low frequency variants, and note that SAPPHIRE is comprised of mainly adults³¹ in comparison to SAGE and GALA II, which are comprised of mainly children.

In conclusion, we identified two novel loci with biological plausibility whereby genetic variation is associated with differential response to albuterol, the most commonly prescribed asthma medication. One of these loci contains variation associated with BDR that is common to African Americans, a population that has historically been understudied in genetic studies^{61–63}. Further genetic studies in African Americans are essential for identifying a more comprehensive set of genetic variants that contribute to differences in BDR, which in turn will lead to a better understanding of the pharmacogenetic response to asthma therapies. This will provide the foundation for genetic risk profiling and precision medicine, identifying novel genes and pathways associated with BDR, and the development of novel asthma therapies.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments:

This work was supported in part by the Sandler Family Foundation, the American Asthma Foundation, the RWJF Amos Medical Faculty Development Program, National Institutes of Health 1R01HL117004, R01HL128439, National Institute of Health and Environmental Health Sciences R01 ES015794, R21ES24844, National Institute on Minority Health and Health Disparities 1P60 MD006902, U54MD009523, 1R01MD010443, and the National Institutes of Health National Heart, Lung, and Blood Institute K08 HL118128, U10 HL109164, HL69116, R01 HL69167, HL69170, HL69174, and U10 HL098103. This project has been funded part with federal funds from the National Cancer Institute, National Institutes of Health, under contract HHSN26120080001E. The content of this publication does not necessarily reflect the views or policies of the Department of Health and Human Services, nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. Government. This Research was supported in part by the Intramural Research Program of the NIH, National Cancer Institute, Center for Cancer Research. MLS was supported in part by a National Science Foundation Graduate Research Fellowship under Grant No. 1144247. MP-Y was funded by the Ramón y Cajal Program (RYC-2015–17205) by the Spanish Ministry of Economy and Competitiveness. MP-Y was also supported by award number AC15/00015 by Instituto de Salud Carlos III thorough AES and EC within AAL framework, and the SysPharmPedia grant awarded from the ERACoSysMed 1st Joint Transnational Call from the European Union under the Horizon 2020; DGT was supported in part by the California Institute for Quantitative Biosciences (QB3). JMG was supported in part by NIH Training Grant T32 (5T32GM007546) and career development awards from the NHLBI K23 (5K23HL111636) and NCATS KL2 (5KL2TR000143) as well as the Hewett Fellowship; NT was supported in part by a institutional training grant from the NIGMS (T32GM007546) and career development awards from the NHLBI (5K12HL119997 and K23- HL125551–01A1), Parker B. Francis Fellowship Program, and the American Thoracic Society; RK was supported with a career development award from the NHLBI (5K23HL093023); HJF was supported in part by the GCRC (RR00188); PCA was supported in part by the Ernest S. Bazley Grant. LKW received grant support from the Fund for Henry Ford Hospital, the American Asthma Foundation, and the following NIH institutes: NHLBI (R01HL118267, R01HL079055), NIAID (R01AI079139), and NIDDK (R01DK064695). Study accession numbers in dbGaP are phs000921.v2.p1 and phs001180.v1.p1.

The authors acknowledge the patients, families, recruiters, health care providers, and community clinics for their participation in SAGE and GALA II. In particular, we thank study coordinator Sandra Salazar and the recruiters who obtained the data: Duanny Alva, MD; Gaby Ayala-Rodriguez; Lisa Caine; Elizabeth Castellanos; Jaime Colon;

Denise DeJesus; Blanca Lopez; Brenda Lopez, MD; Louis Martos; Vivian Medina; Juana Olivo; Mario Peralta; Esther Pomares, MD; Jihan Qurashi; Johanna Rodriguez; Shahdad Saeedi; Dean Soto; and Ana Taveras.

The contents of this publication are solely the responsibility of the authors and do not necessarily represent the official views of the NIH.

References

1. Burchard EG, Ziv E, Coyle N, Gomez SL, Tang H, Karter AJ, et al. The importance of race and ethnic background in biomedical research and clinical practice. *The New England journal of medicine* 2003; 348(12): 1170–1175. [PubMed: 12646676]
2. Naqvi M, Thyne S, Choudhry S, Tsai HJ, Navarro D, Castro RA, et al. Ethnic-specific differences in bronchodilator responsiveness among African Americans, Puerto Ricans, and Mexicans with asthma. *The Journal of asthma : official journal of the Association for the Care of Asthma* 2007; 44(8): 639–648. [PubMed: 17943575]
3. Padhukasahasram B, Yang JJ, Levin AM, Yang M, Burchard EG, Kumar R, et al. Gene-based association identifies SPATA13-AS1 as a pharmacogenomic predictor of inhaled short-acting beta-agonist response in multiple population groups. *Pharmacogenomics J* 2014.
4. Palmer LJ, Silverman ES, Weiss ST, Drazen JM. Pharmacogenetics of Asthma. *American Journal of Respiratory and Critical Care Medicine* 2002; 165(7): 861–866. [PubMed: 11934710]
5. Nelson HS. Beta-adrenergic bronchodilators. *The New England journal of medicine* 1995; 333(8): 499–506. [PubMed: 7623883]
6. Loza MJ, Penn RB. Regulation of T cells in airway disease by beta-agonist. *Frontiers in bioscience* 2010; 2: 969–979.
7. Salathe M Effects of beta-agonists on airway epithelial cells. *The Journal of allergy and clinical immunology* 2002; 110(6 Suppl): S275–281. [PubMed: 12464936]
8. Shore SA, Moore PE. Regulation of beta-adrenergic responses in airway smooth muscle. *Respiratory physiology & neurobiology* 2003; 137(2–3): 179–195. [PubMed: 14516725]
9. Jartti T. Asthma, asthma medication and autonomic nervous system dysfunction. *Clinical physiology* 2001; 21(2): 260–269. [PubMed: 11318835]
10. Drake KA, Torgerson DG, Gignoux CR, Galanter JM, Roth LA, Huntsman S, et al. A genome-wide association study of bronchodilator response in Latinos implicates rare variants. *J Allergy Clin Immunol* 2014; 133(2): 370–378. [PubMed: 23992748]
11. Burchard EG, Avila PC, Nazario S, Casal J, Torres A, Rodriguez-Santana JR, et al. Lower bronchodilator responsiveness in Puerto Rican than in Mexican subjects with asthma. *Am J Respir Crit Care Med* 2004; 169(3): 386–392. [PubMed: 14617512]
12. Choudhry S, Ung N, Avila PC, Ziv E, Nazario S, Casal J, et al. Pharmacogenetic differences in response to albuterol between Puerto Ricans and Mexicans with asthma. *Am J Respir Crit Care Med* 2005; 171(6): 563–570. [PubMed: 15557128]
13. Blake K, Madabushi R, Derendorf H, Lima J. Population pharmacodynamic model of bronchodilator response to inhaled albuterol in children and adults with asthma. *Chest* 2008; 134(5): 981–989. [PubMed: 18583517]
14. Gorina Y. QuickStats:asthma*death rates, by race and age group - United States, 2007–2009 *In* (MMWR) *MaMWR* (ed) Centers for Disease Control and Prevention 2012.
15. Martinez FD, Graves PE, Baldini M, Solomon S, Erickson R. Association between genetic polymorphisms of the beta2-adrenoceptor and response to albuterol in children with and without a history of wheezing. *The Journal of clinical investigation* 1997; 100(12): 3184–3188. [PubMed: 9399966]
16. Silverman EK, Kwiatkowski DJ, Sylvia JS, Lazarus R, Drazen JM, Lange C, et al. Family-based association analysis of beta2-adrenergic receptor polymorphisms in the childhood asthma management program. *The Journal of allergy and clinical immunology* 2003; 112(5): 870–876. [PubMed: 14610472]
17. Poon AH, Tantisira KG, Litonjua AA, Lazarus R, Xu J, Lasky-Su J, et al. Association of corticotropin-releasing hormone receptor-2 genetic variants with acute bronchodilator response in asthma. *Pharmacogenetics and genomics* 2008; 18(5): 373–382. [PubMed: 18408560]

18. Tantisira KG, Small KM, Litonjua AA, Weiss ST, Liggett SB. Molecular properties and pharmacogenetics of a polymorphism of adenylyl cyclase type 9 in asthma: interaction between beta-agonist and corticosteroid pathways. *Human molecular genetics* 2005; 14(12): 1671–1677. [PubMed: 15879435]
19. Litonjua AA, Lasky-Su J, Schneiter K, Tantisira KG, Lazarus R, Klanderman B, et al. ARG1 is a novel bronchodilator response gene: screening and replication in four asthma cohorts. *Am J Respir Crit Care Med* 2008; 178(7): 688–694. [PubMed: 18617639]
20. Duan QL, Du R, Lasky-Su J, Klanderman BJ, Partch AB, Peters SP, et al. A polymorphism in the thyroid hormone receptor gene is associated with bronchodilator response in asthmatics. *The pharmacogenomics journal* 2013; 13(2): 130–136. [PubMed: 22212731]
21. Reihnsaus E, Innis M, MacIntyre N, Liggett SB. Mutations in the gene encoding for the beta 2-adrenergic receptor in normal and asthmatic subjects. *American journal of respiratory cell and molecular biology* 1993; 8(3): 334–339. [PubMed: 8383511]
22. Duan QL, Lasky-Su J, Himes BE, Qiu W, Litonjua AA, Damask A, et al. A genome-wide association study of bronchodilator response in asthmatics. *The pharmacogenomics journal* 2014; 14(1): 41–47. [PubMed: 23508266]
23. Israel E, Lasky-Su J, Markezich A, Damask A, Szeffler SJ, Schuemann B, et al. Genome-wide association study of short-acting beta2-agonists. A novel genome-wide significant locus on chromosome 2 near ASB3. *Am J Respir Crit Care Med* 2015; 191(5): 530–537. [PubMed: 25562107]
24. Himes BE, Jiang X, Hu R, Wu AC, Lasky-Su JA, Klanderman BJ, et al. Genome-wide association analysis in asthma subjects identifies SPATS2L as a novel bronchodilator response gene. *PLoS genetics* 2012; 8(7): e1002824. [PubMed: 22792082]
25. Torgerson DG, Ampleford EJ, Chiu GY, Gauderman WJ, Gignoux CR, Graves PE, et al. Meta-analysis of genome-wide association studies of asthma in ethnically diverse North American populations. *Nature genetics* 2011; 43(9): 887–892. [PubMed: 21804549]
26. Galanter JM, Gignoux CR, Torgerson DG, Roth LA, Eng C, Oh SS, et al. Genome-wide association study and admixture mapping identify different asthma-associated loci in Latinos: the Genes-environments & Admixture in Latino Americans study. *J Allergy Clin Immunol* 2014; 134(2): 295–305. [PubMed: 24406073]
27. White MJ, Risse-Adams O, Goddard P, Contreras MG, Adams J, Hu D, et al. Novel genetic risk factors for asthma in African American children: Precision Medicine and the SAGE II Study. *Immunogenetics* 2016; 68(6–7): 391–400. [PubMed: 27142222]
28. Winkler CA, Nelson GW, Smith MW. Admixture mapping comes of age. *Annu Rev Genomics Hum Genet* 2010; 11: 65–89. [PubMed: 20594047]
29. Nishimura KK, Galanter JM, Roth LA, Oh SS, Thakur N, Nguyen EA, et al. Early-life air pollution and asthma risk in minority children. The GALA II and SAGE II studies. *Am J Respir Crit Care Med* 2013; 188(3): 309–318. [PubMed: 23750510]
30. Torgerson DG, Gignoux CR, Galanter JM, Drake KA, Roth LA, Eng C, et al. Case-control admixture mapping in Latino populations enriches for known asthma-associated genes. *The Journal of allergy and clinical immunology* 2012; 130(1): 76–82 e12. [PubMed: 22502797]
31. Gould W, Peterson EL, Karungi G, Zoratti A, Gaggin J, Toma G, et al. Factors predicting inhaled corticosteroid responsiveness in African American patients with asthma. *The Journal of allergy and clinical immunology* 2010; 126(6): 1131–1138. [PubMed: 20864153]
32. Moore WC, Bleeker ER, Curran-Everett D, Erzurum SC, Ameredes BT, Bacharier L, et al. Characterization of the severe asthma phenotype by the National Heart, Lung, and Blood Institute’s Severe Asthma Research Program. *J Allergy Clin Immunol* 2007; 119(2): 405–413. [PubMed: 17291857]
33. Moore WC, Meyers DA, Wenzel SE, Teague WG, Li H, Li X, et al. Identification of asthma phenotypes using cluster analysis in the Severe Asthma Research Program. *Am J Respir Crit Care Med* 2010; 181(4): 315–323. [PubMed: 19892860]
34. Standardization of Spirometry, 1994 Update. American Thoracic Society. *Am J Respir Crit Care Med* 1995; 152(3): 1107–1136. [PubMed: 7663792]

35. Delaneau O, Zagury JF. Haplotype inference. *Methods Mol Biol* 2012; 888: 177–196. [PubMed: 22665282]
36. Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS genetics* 2009; 5(6): e1000529. [PubMed: 19543373]
37. Howie B, Marchini J, Stephens M. Genotype imputation with thousands of genomes. *G3* 2011; 1(6): 457–470. [PubMed: 22384356]
38. Genomes Project C, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, et al. A global reference for human genetic variation. *Nature* 2015; 526(7571): 68–74. [PubMed: 26432245]
39. Maples BK, Gravel S, Kenny EE, Bustamante CD. RFMix: a discriminative modeling approach for rapid and robust local-ancestry inference. *Am J Hum Genet* 2013; 93(2): 278–288. [PubMed: 23910464]
40. Proceedings of the ATS workshop on refractory asthma: current understanding, recommendations, and unanswered questions. American Thoracic Society. *Am J Respir Crit Care Med* 2000; 162(6): 2341–2351. [PubMed: 11112161]
41. Pino-Yanes M, Thakur N, Gignoux CR, Galanter JM, Roth LA, Eng C, et al. Genetic ancestry influences asthma susceptibility and lung function among Latinos. *J Allergy Clin Immunol* 2015; 135(1): 228–235. [PubMed: 25301036]
42. Baran Y, Pasaniuc B, Sankararaman S, Torgerson DG, Gignoux C, Eng C, et al. Fast and accurate inference of local ancestry in Latino populations. *Bioinformatics* 2012; 28(10): 1359–1367. [PubMed: 22495753]
43. Kumar R, Nguyen EA, Roth LA, Oh SS, Gignoux CR, Huntsman S, et al. Factors associated with degree of atopy in Latino children in a nationwide pediatric sample: the Genes-environments and Admixture in Latino Asthmatics (GALA II) study. *J Allergy Clin Immunol* 2013; 132(4): 896–905 e891. [PubMed: 23684070]
44. Alexander DH, Novembre J, Lange K. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res* 2009; 19(9): 1655–1664. [PubMed: 19648217]
45. McGarry ME, Castellanos E, Thakur N, Oh SS, Eng C, Davis A, et al. Obesity and bronchodilator response in black and Hispanic children and adolescents with asthma. *Chest* 2015; 147(6): 1591–1598. [PubMed: 25742612]
46. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 2010; 26(17): 2190–2191. [PubMed: 20616382]
47. PLUMMER M BN, COWLES K, VINES K CODA: Convergence Diagnosis and Output Analysis for MCMC. *R News* 2012;(6): 7–11.
48. Sobota RS, Shriner D, Kodaman N, Goodloe R, Zheng W, Gao YT, et al. Addressing population-specific multiple testing burdens in genetic association studies. *Ann Hum Genet* 2015; 79(2): 136–147. [PubMed: 25644736]
49. Marcus JH, Novembre J. Visualizing the geography of genetic variants. *Bioinformatics* 2016.
50. Consortium GT. The Genotype-Tissue Expression (GTEx) project. *Nature genetics* 2013; 45(6): 580–585. [PubMed: 23715323]
51. Li Z, Xi X, Gu M, Feil R, Ye RD, Eigenthaler M, et al. A stimulatory role for cGMP-dependent protein kinase in platelet activation. *Cell* 2003; 112(1): 77–86. [PubMed: 12526795]
52. Tamura N, Itoh H, Ogawa Y, Nakagawa O, Harada M, Chun TH, et al. cDNA cloning and gene expression of human type Ialpha cGMP-dependent protein kinase. *Hypertension* 1996; 27(3 Pt 2): 552–557. [PubMed: 8613202]
53. Orstavik S, Natarajan V, Tasken K, Jahnsen T, Sandberg M. Characterization of the human gene encoding the type I alpha and type I beta cGMP-dependent protein kinase (PRKG1). *Genomics* 1997; 42(2): 311–318. [PubMed: 9192852]
54. Burgoyne JR, Madhani M, Cuello F, Charles RL, Brennan JP, Schroder E, et al. Cysteine redox sensor in PKGIa enables oxidant-induced activation. *Science* 2007; 317(5843): 1393–1397. [PubMed: 17717153]
55. Pfeifer A, Klatt P, Massberg S, Ny L, Sausbier M, Hirneiss C, et al. Defective smooth muscle regulation in cGMP kinase I-deficient mice. *EMBO J* 1998; 17(11): 3045–3051. [PubMed: 9606187]

56. Dawes M, Chowienczyk PJ, Ritter JM. Effects of inhibition of the L-arginine/nitric oxide pathway on vasodilation caused by beta-adrenergic agonists in human forearm. *Circulation* 1997; 95(9): 2293–2297. [PubMed: 9142007]
57. Boyle AP, Hong EL, Hariharan M, Cheng Y, Schaub MA, Kasowski M, et al. Annotation of functional variation in personal genomes using RegulomeDB. *Genome research* 2012; 22(9): 1790–1797. [PubMed: 22955989]
58. Lima JJ, Thomason DB, Mohamed MH, Eberle LV, Self TH, Johnson JA. Impact of genetic polymorphisms of the beta2-adrenergic receptor on albuterol bronchodilator pharmacodynamics. *Clin Pharmacol Ther* 1999; 65(5): 519–525. [PubMed: 10340917]
59. Taylor DR, Drazen JM, Herbison GP, Yandava CN, Hancox RJ, Town GI. Asthma exacerbations during long term beta agonist use: influence of beta(2) adrenoceptor polymorphism. *Thorax* 2000; 55(9): 762–767. [PubMed: 10950895]
60. Israel E, Drazen JM, Liggett SB, Boushey HA, Cherniack RM, Chinchilli VM, et al. The effect of polymorphisms of the beta(2)-adrenergic receptor on the response to regular use of albuterol in asthma. *Am J Respir Crit Care Med* 2000; 162(1): 75–80. [PubMed: 10903223]
61. Bustamante CD, Burchard EG, De la Vega FM. Genomics for the world. *Nature* 2011; 475(7355): 163–165. [PubMed: 21753830]
62. Popejoy AB, Fullerton SM. Genomics is failing on diversity. *Nature* 2016; 538(7624): 161–164. [PubMed: 27734877]
63. Editors PM, Rid A, Johansson MA, Leung G, Valantine H, Burchard EG, et al. Towards Equity in Health: Researchers Take Stock. *PLoS Med* 2016; 13(11): e1002186. [PubMed: 27898673]

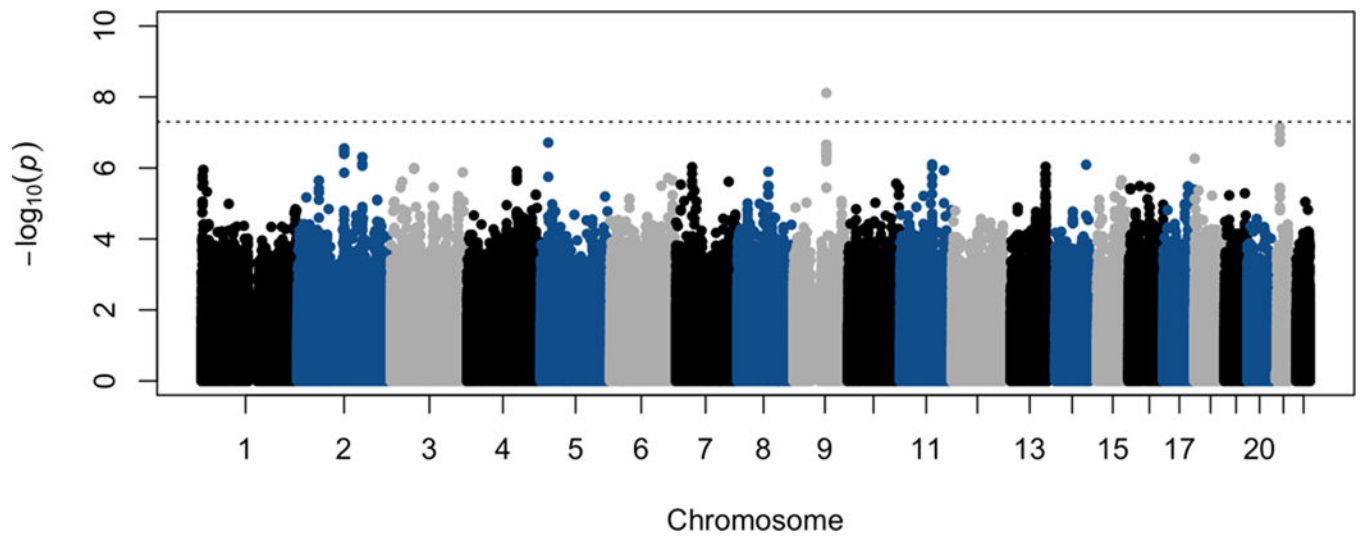


Figure 1: Meta-analysis of genome-wide association studies with BDR in African Americans. Association testing for BDR was performed using linear regression including age, sex, BMI category, local and global ancestry as covariates separately in SAGE I and II and combined in a meta-analysis. Dotted line indicates the genome-wide significance threshold of 5×10^{-8} .

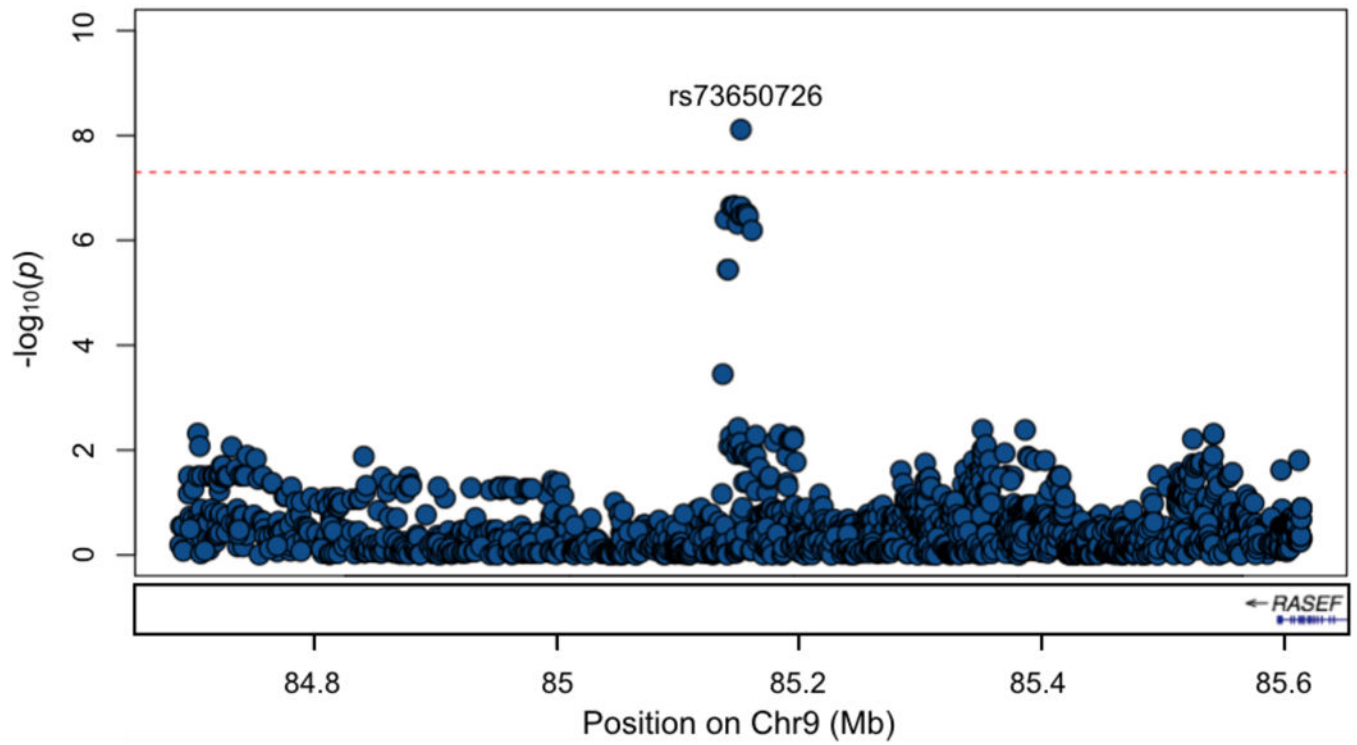


Figure 2: LocusZoom plot of chr9:84653000–85653000.

Region includes genotyped and imputed variants from 1000 Genomes phase 3. Blue = variants common in SAGE I and II. Dotted line indicates the genome-wide significance threshold of 5×10^{-8} .

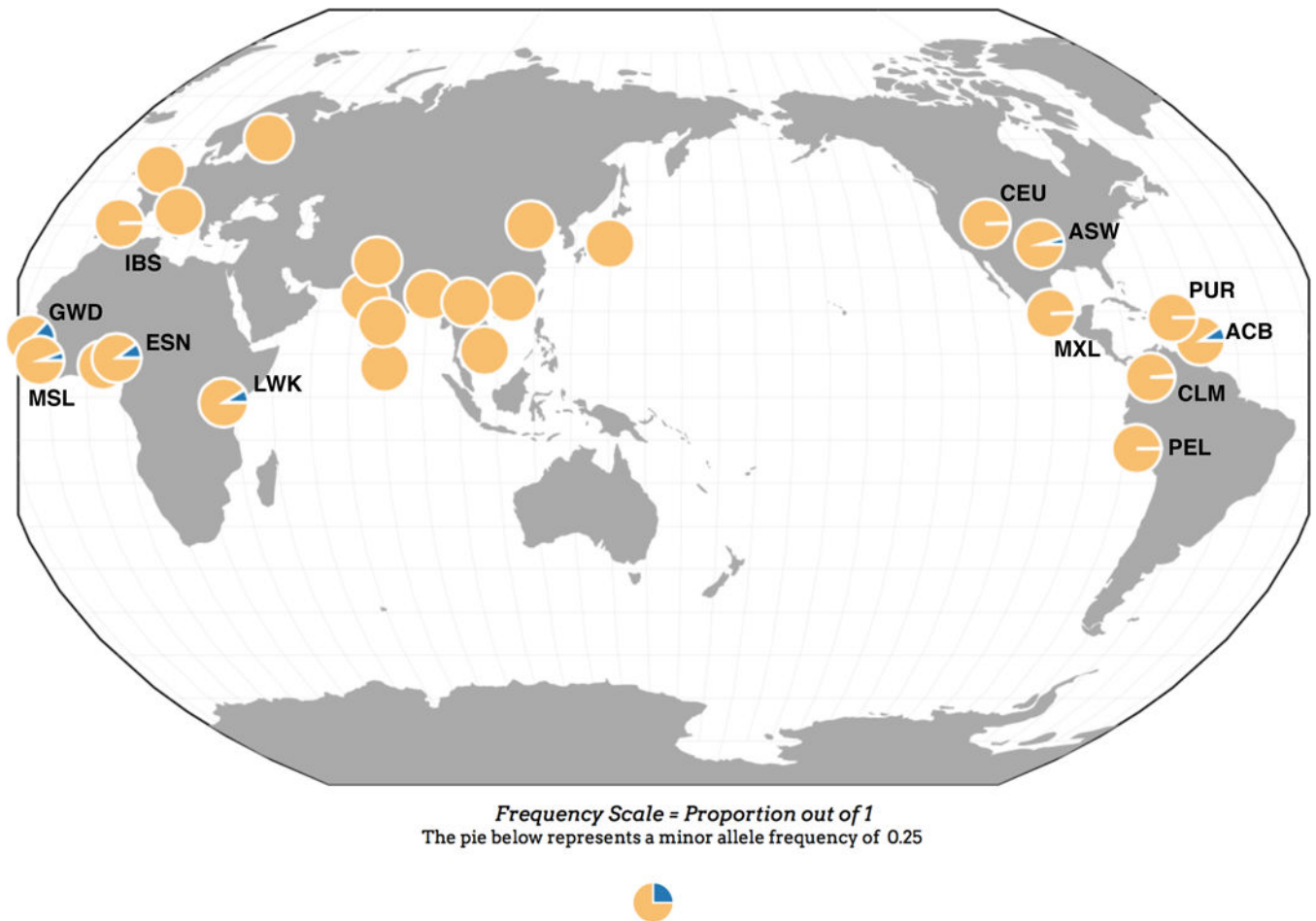


Figure 3: Geographic distribution of allele frequencies of rs73650726.

Each pie chart refers to a population from the 1000 Genomes Project phase 3. Yellow= Major allele (A), blue = minor allele (G). rs73650726 is common only in populations with African ancestry.

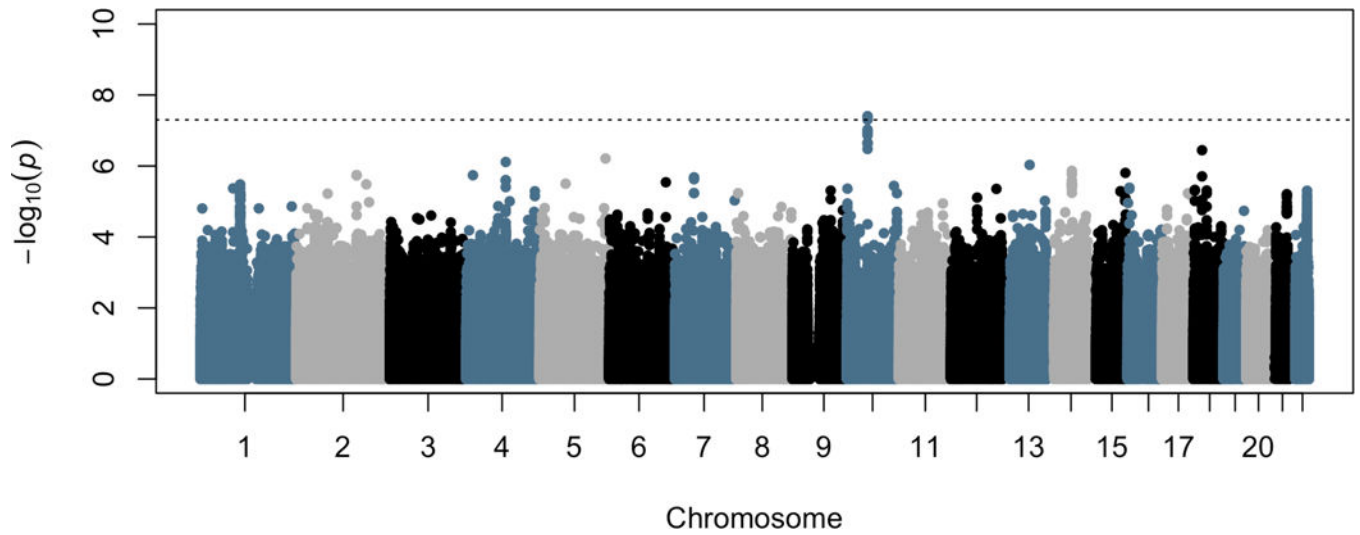


Figure 4: Meta-analysis of genome-wide association studies with BDR in African Americans and Latinos.

Association testing for BDR was performed using linear regression including age, sex, BMI category, local and global ancestry as covariates; including ethnicity for GALA II. Dotted line indicates the genome-wide significance threshold of 5×10^{-8} .

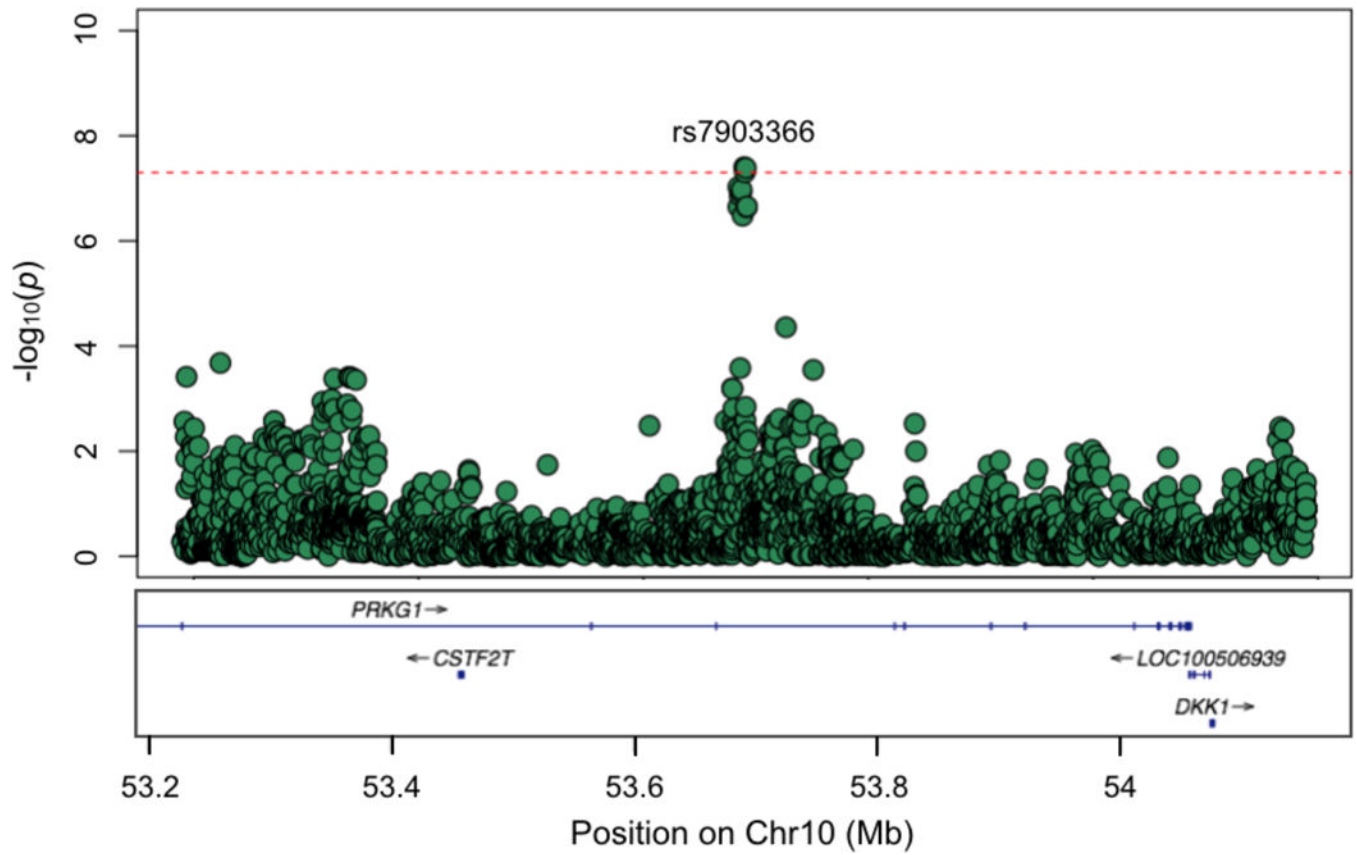


Figure 5: LocusZoom plot of chr10:53200000–54200000.

Region includes genotyped and imputed variants from 1000 Genomes phase 3. Green = variants common in SAGE I, SAGE II and GALA II. Dotted line indicates the genome-wide significance threshold of 5×10^{-8} .

Table 1:
Descriptive statistics of SAGE I, SAGE II, & GALA II asthma cases.

Values shown are the means, with the standard deviation in parentheses.

	SAGE I	SAGE II	GALA II
Total (N)	190	759	1830
Age (year)	18 (9.3)	14 (3.6)	13 (3.2)
<18 years (%)	64%	86%	93%
Sex (%Male)	41%	52%	55%
Race/Ethnicity	African American	African American	Latino
Global African Ancestry	0.81 (0.13)	0.72 (0.12)	0.15 (0.13)
Global Native American Ancestry	-	-	0.30 (0.25)
BMI			
<20 years	25 (7.3) (N=132)	25 (7.2) (N=722)	23 (6.5) (N=1782)
>20 years	31 (7.8) (N=58)	29 (7.0) (N=37)	30 (6.6) (N=48)
Pulmonary Function			
Pre-FEV1 % Predicted	92 (16)	99 (14)	91 (16)
Pre-FVC % Predicted	100 (17)	104 (13)	95 (16)
BDR (%)	9 (9.1)	9.5 (6.9)	11 (8.2)

Table 2:
Genome-wide significant associations identified through a meta-analysis within African Americans (SAGE I and II), and within African Americans and Latinos (SAGE I, SAGE II, and GALA II).

Under 'Direction' the first symbol refers to SAGE I, second to SAGE II, and third to GALA II. 0 = absent/rare in study

African Americans (SAGE I and II):								
Chr	SNP	Position (hg19)	A1	A2	Effect (A1)	StdErr	Pvalue	Direction
9q21	rs73650726	85152666	A	G	-3.8	0.66	7.69×10^{-9}	--0
African Americans + Latinos (SAGE I, SAGE II, GALA II):								
Chr	SNP	Position (hg19)	A1	A2	Effect (A1)	StdErr	Pvalue	Direction
10q21	rs7903366	53689774	T	C	1.23	0.22	3.94×10^{-8}	+++
10q21	rs7070958	53691116	A	G	-1.24	0.23	4.09×10^{-8}	---
10q21	rs7081864	53690331	A	G	1.23	0.22	4.94×10^{-8}	+++

Table 3:
Correlation between the expression of PRKG1 in the lung and minor alleles at three intronic SNPs associated with BDR (cis-eQTLs).

Data is from the GTEx database.

SNP	Ref Allele	Pvalue	Effect (Ref Allele)	T-Statistic	StdErr	Tissue	Gene
rs7903366	C	0.00051	-0.12	-3.5	0.034	Lung	<i>PRKG1</i>
rs7070958	A	0.00046	-0.12	-3.6	0.034	Lung	<i>PRKG1</i>
rs7081864	G	0.00052	-0.12	-3.5	0.034	Lung	<i>PRKG1</i>

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript